

1 **Three-spined stickleback armour predicted by body size, minimum**  
2 **winter temperature and pH**

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16 **Abstract**

17           Similar phenotypes evolve under equivalent environmental conditions through parallel  
18 evolution. Because they have repeatedly invaded and adapted to new freshwater environments, the  
19 three-spined stickleback (*Gasterosteus aculeatus*) offers a powerful system for understanding the  
20 agents of selection in nature that drive parallel evolution. Here we examine the ecological and  
21 environmental variables responsible for morphological variation in three-spined stickleback  
22 populations across its European range. We collected fish from 85 populations, encompassing much  
23 of the European latitudinal range of the species and including lowland rivers and lakes, coastal  
24 lagoons, and moorland ponds. We measured biotic and environmental variables at all sites along  
25 with morphological traits for 2,358 individuals. Using an information theory approach, we identified  
26 body size, minimum average winter temperature and pH as primary predictors of stickleback armour  
27 evolution, challenging current hypotheses for stickleback morphological diversification and  
28 demonstrating the fundamental role played by body size and scaling in mediating responses to  
29 selection. Stickleback lateral plate phenotype represents a potentially powerful tool for monitoring  
30 change in climate variables across the northern temperate region.

31

## 32 **Introduction**

33 Similar phenotypes are expected to evolve under the same environmental selection pressures  
34 through parallel evolution (Haldane, 1932; Bolnick *et al.*, 2018). However, identifying the  
35 ecological and abiotic agents of selection that underpin parallel evolution is often difficult. Parallel  
36 evolution is a feature of the three-spined stickleback (*Gasterosteus aculeatus*) complex, which  
37 shows substantial inter-population phenotypic variation in morphology, behaviour, physiology and  
38 life-history traits (Wootton, 1976, 1984; Bell & Foster, 1994). The evolutionary history of the three-  
39 spined stickleback is one of repeated, independent invasions of freshwater habitats by marine  
40 populations. These invasions are characterized by a reduction in anti-predator ‘armour’, comprising  
41 lateral bony plates, pelvic girdle and spines, and dorsal spines, as well as other aspects of their  
42 biology (Bell & Foster, 1994). Their wide distribution, tolerance of a wide range of environmental  
43 conditions, and striking phenotypic variability makes the three-spined stickleback an ideal model  
44 for investigating selective forces underpinning parallel evolution.

45         Freshwater populations of three-spined sticklebacks distributed over large geographical  
46 distances typically converge on a *low* lateral plate morph, with <10 plates on each side of the anterior  
47 portion of the body, along with dorsal spines, pelvic girdle and pelvic spines. The separate elements  
48 of the armour function in concert, with anterior lateral plates bracing the dorsal and pelvic spines,  
49 which thereby resist compression while limiting ingestion by gape-limited vertebrate predators  
50 (Reimchen, 1994). Marine sticklebacks are almost exclusively represented by a *complete* morph,  
51 with >30 lateral plates along the entire body along with robust dorsal and pelvic spines (Reimchen,  
52 1994). Posterior lateral plates appear to function in preventing skin puncture by toothed predators  
53 (Reimchen, 1994). A *partial* morph also occurs, expressing an intermediate but variable number of  
54 lateral plates. This morph is commonly, though not exclusively, encountered in brackish water  
55 (Wootton, 1976). The rate of evolutionary response shown by three-spined sticklebacks in

56 freshwater environments appears rapid, with a reversal in plate morph dominance possible in as few  
57 as 10 generations (e.g. Bell *et al.*, 2004; Kitano *et al.*, 2008).

58 Parallel phenotypic evolution in three-spined sticklebacks is reflected by genomic  
59 parallelism, with 242 genomic regions associated with repeated adaptation to fresh water (Jones *et al.*,  
60 2012). The *ectodysplasin A* (EDA) locus has been specifically implicated in lateral plate morph  
61 evolution, with ancient variants of the EDA locus reused across multiple freshwater populations;  
62 approximately 70% of variation in plate number and size is associated with variation in EDA  
63 (Cresko *et al.*, 2004; Colosimo *et al.*, 2005), though the association between variation in EDA and  
64 lateral plate morph is not universal (Pujolar *et al.*, 2017). The contemporary model for stickleback  
65 morph evolution involves the repeated, independent establishment of freshwater populations from  
66 ancestral, panmictic populations of *complete* lateral plate marine fish following the end of the last  
67 glacial period (*ca.* 11,700 cal years BP) when numerous freshwater environments in the northern  
68 hemisphere became accessible with the retreat of the Laurentide and Scandinavian ice sheets  
69 (Carlson, 2013). Under this model, *complete* morph marine three-spined stickleback populations are  
70 assumed to possess a pool of standing genetic variation, with otherwise rare alleles experiencing  
71 strong selection once fish enter fresh water where they rapidly increase in frequency (Barrett, 2010).

72 Several hypotheses have been proposed to explain the selective advantage of armour loss  
73 associated with colonization of fresh waters by the three-spined stickleback. These include the  
74 effects of temperature, salinity, predation, geographic location, dissolved calcium and phosphorus  
75 availability, proximity to the marine environment, water density, parasitism, competition, water  
76 turbidity and body size (Table 1). However, there are striking deviations from the typical pattern of  
77 lateral plate morph evolution. Notably the *complete* morph occurs widely in fresh waters in eastern  
78 Europe, the east coast of North America and sites in Japan (Wootton, 1976, 2009; Hagen & Moodie,  
79 1982). In contrast, some populations on the west coast of North America, Norway, Scotland, Ireland

80 and Iceland express greatly diminished armour development, typically with <3 lateral plates of  
81 reduced size and thickness, and in some cases with plates and spines entirely absent (Giles, 1983).

82 We addressed the ecological basis to variation in three-spined stickleback lateral plate  
83 number using phenotypic and ecological data collected from across the European range of the  
84 species. Using an information theory approach (Burnham & Anderson, 2002), we compared a series  
85 of *a priori* alternative models of plate loss based on previously published studies (Table 1) to identify  
86 the ecological and abiotic agents of selection that underpin parallel evolution in this species. We  
87 focused on plate number, rather than other skeletal elements, because lateral plates show greater  
88 variation than dorsal spines and pelvic girdle, appears more sensitive to environmental variation and  
89 can be unambiguously quantified.

90

## 91 **Materials and methods**

### 92 **Fish collection and analysis**

93 Field collection of three-spined sticklebacks was undertaken across Europe using dip nets and Gee  
94 minnow traps. A total of 2,358 individuals from 85 locations in England, Estonia, France, Norway,  
95 Poland, Scotland and Turkey were collected (Table S1, Fig. S1). Where they co-occurred with three-  
96 spined sticklebacks, samples of nine-spined sticklebacks (*Pungitius pungitius*) were also collected.  
97 Collected fish were killed with anaesthetic (benzocaine) and fixed in 4% buffered formalin. Samples  
98 were restricted to adult fish >27 mm standard length (SL) to ensure morphological characteristics  
99 were fully developed (Bańbura, 1989). Nine populations with <16 individuals were dropped from  
100 the analysis (Table S1). After fixing, fish were transferred to 70% ethanol for 24 hours, stained with  
101 0.08% alizarin red in 1% KOH for 24 hours, rinsed in water for 24 hours, and stored in 70% ethanol  
102 (Brubaker & Angus, 1984). For each fish, a record was made of SL and the number of lateral plates  
103 on its left flank. The presence of the parasite *Schistocephalus solidus* (Platyhelminthes) was also

104 noted.

105

106 **Environmental data**

107 A water sample was collected at each sample site in one-litre sample bottles from below the water  
108 surface at a depth of approximately 1 m. Dissolved calcium concentration was measured by atomic  
109 absorption spectrometric methods (ISO 7980:1986), phosphorus concentration by ammonium  
110 molybdate spectrometric methods (EN 1189:1996), and pH was measured electrochemically (ISO  
111 10523:2008). An accredited commercial laboratory (ZWiK, Łódź) performed all water chemistry  
112 measurements. Estimates of dissolved calcium and phosphorus were not obtained for all sites, but  
113 in these cases, pH was measured directly in the field using a multi-probe portable water quality  
114 meter (Eutech, PCSTestr 35). Dissolved calcium and pH were correlated among sites (Fig. S2).  
115 Consequently, pH was used as a proxy for dissolved calcium in models to maximise the number of  
116 sites included in the analysis. Habitat variables were also scored at each site, these were; water  
117 turbidity (either high, <0.8 m Secchi depth, or low >0.8 m) and vegetation cover (either high, >5%  
118 of water body area, or low <5%). The presence/absence of predatory fishes was determined by  
119 electrofishing (EFGI 650, BSE Bretschneider Specialelektronik, Germany) at sites in Poland.  
120 Detailed fish faunal lists were available for sites in France and England. In Scotland and Norway,  
121 angling records were available for all sites. All marine sites (North Poland, Estonia, Turkey) were  
122 assumed to be support marine fish predators. The presence/absence of predatory dragonfly  
123 (Odonata) and beetle (Coleoptera) larvae and nine-spined sticklebacks was ascertained by dip  
124 netting in vegetation at the lake/river bank. For every sampling location the elevation above sea  
125 level was estimated from Google Earth using Web-based software ([www.freemaptools.com](http://www.freemaptools.com)).  
126 Average minimum winter temperature data for each site over a 30-year period from 1961–1990 were  
127 obtained from the nearest meteorological station (Table S1).

128

129 **Data analysis**

130 Data were modelled using R (version 3.5.3; R Development Core Team, 2019) with models fitted  
131 in a Bayesian framework using Integrated Nested Laplace Approximation (R-INLA; Rue *et al.*,  
132 2017). Using an information theory approach, 15 alternative models were formulated comprising  
133 variables proposed as having importance as agents of selection for lateral plates in previously  
134 published studies (Table 1). Default priors were put on all model parameters and best-fitting models  
135 were identified using Watanabe-Akaike Information Criterion (WAIC) (Vehtari *et al.*, 2017).

136 Data for lateral plate number were multimodal and zero inflated, with three discrete  
137 distributions for modal plate numbers of 0, 5 (range 1-8) and 30 (28-34) plates (Fig. 1). To  
138 accommodate this data structure, we fitted two models. We compared fish without lateral plates to  
139 those possessing plates using a binary (Bernoulli) process to model zero and positive lateral plate  
140 counts, while data for *low* and *complete* morphs were modelled using a hurdle model. Hurdle models  
141 are partitioned into two parts, with a binary process modelling probability of an event, and a second  
142 process modelling the magnitude of an event (Hilbe, 2014). In the case of plate counts, we modelled  
143 data using a negative binomial distribution to accommodate overdispersion in the data (Smith &  
144 Warren, 2019), fitting models to *low* and *complete* morphs separately. This approach enabled us to  
145 fit models that explained: 1. the probability of possessing lateral plates, 2. the probability of  
146 expressing the *complete* rather than *low* plate morph, 3. the number of plates expressed by the *low*  
147 plate morph, 4. the number of plates expressed by the *complete* plate morph. Data for fish expressing  
148 the *partial* morph were excluded from the analysis, since they occurred at low frequency (6% of  
149 specimens). Including *partial* morph fish as either *complete* or *low* plate morph did not result in a  
150 qualitative change to the outcome of the analysis. Exploratory analyses demonstrated differences  
151 among populations in the number of lateral plates. To accommodate this effect, a random intercept

152 for fish population of origin was included in models to introduce a correlation structure between  
153 observations for different fish from the same population. Similarly, to accommodate potential  
154 dependency in the data due to temperature data for different populations deriving from the same  
155 meteorological station, this factor was included as an additional random term in models.

156 As a follow-up to the main results, three-spined stickleback body size was modelled with a  
157 gamma GLMM, which assumed body size was strictly positive and continuous. The full model  
158 included salinity, turbidity, flow regime (lentic/lotic), vegetation cover, presence of predatory  
159 invertebrates, predatory fishes, nine-spined sticklebacks and *Schistocephalus solidus*, average  
160 minimum temperature, pH, and elevation, with an optimal fixed structure identified with a backward  
161 selection procedure based on WAIC. Population and month of sample collection were random  
162 intercepts in the model. To characterize the relationship between the size of three- and nine-spined  
163 sticklebacks among populations a gamma GLM was fitted to the SL of both species in 33 populations  
164 where they co-occurred.

165

## 166 **Results**

167 The best-fitting models to predict the probability of possessing lateral plates were models M15 and  
168 M06 (Table 2). Both showed a comparably good fit to the data, though M15 showed higher  
169 weighting. Model M15 comprised three explanatory variables, with pH positively associated (Table  
170 3, Fig. 2a) and average minimum temperature negatively associated with the probability of  
171 expressing plates (Table 3, Fig. 2b). There was no statistically important association between the  
172 probability of expressing plates and SL (Table 3). The model predicted that all fish from locations  
173 with pH values <5 would be plateless, while from a pH of 5.0-6.5 there was a switch in the  
174 probability of the *zero* morph from 1 to 0. Above a pH of 6.5 all fish were predicted to possess lateral  
175 plates. Model predictions matched the data, though with a single specimen from Turkey, exhibiting

176 the *zero* morph at a pH exceeding 7.0 (Fig. 2a). Similarly, below an average minimum temperature  
177 of -1 °C, sticklebacks were predicted to express lateral plates (Fig. 2b).

178 The best-fitting hurdle model for the probability of expressing a *complete* rather than *low*  
179 plate morph was again model M15, though with models M13, M14, M04 and M10 showing good  
180 fits to the data, though with lower weighting (Table 2). The probability of expressing the *complete*  
181 morph was positively associated with pH (Fig. 3a, Table 4), negatively with average minimum  
182 temperature (Fig. 3b, Table 4), and positively with SL (Fig. 3c; Table 4). The number of plates  
183 expressed by both the *low* and *complete* morphs was also best predicted by model M15 (Table 2).  
184 In the case of the *low* plate morph, pH (Fig. 4a), average minimum temperature (Fig. 4b) and SL  
185 (Fig. 4c) all proved statistically important in the model (Table 5). In the case of the *complete* plate  
186 morph, only SL was a statistically important predictor of plate number (Fig. 4c; Table 4).

187 The best-fitting model for three-spined stickleback SL, identified by backward selection,  
188 showed a statistically important negative association with the presence of nine-spined sticklebacks  
189 and average minimum temperature, and a positive association with pH (Table 5). There was a  
190 statistically important positive relationship between mean SL of three- and nine-spined sticklebacks  
191 among populations (Table 6; Fig. 5).

192

## 193 **DISCUSSION**

194 The repeated independent evolution of identical traits in similar environments implies natural  
195 selection (Bolnick *et al.*, 2018). We modelled the expression of lateral plate armour in the three-  
196 spined stickleback from a range of habitats across its European range, showing a consistent pattern  
197 within habitats with the trait predicted by pH, average minimum temperature and body size.

198 Two environmental variables were consistent in predicting morph and plate number; pH and  
199 average minimum temperature. Stickleback populations without lateral plates (the *zero* morph),

200 were predicted by pH values <6.5 (Fig. 2a), though with a single specimen from southern Turkey,  
201 at the southern geographic limit of the distribution, showing the *zero* morph at a pH exceeding 7.0.  
202 The *zero* morph is globally rare, and typically associated with low pH sites on the west coast of  
203 Europe and North America, where they experience mild winter temperatures (Hagen & Moodie,  
204 1982; Wootton, 1984). It is also associated with small body size. In the Scottish Hebrides the *zero*  
205 morph has been described as a ‘dwarf’ form, with sexual maturity achieved at <30 mm SL in some  
206 populations (MacColl *et al.*, 2013). The *zero* morph contrasts with the *complete* morph in fresh  
207 water, which is associated with high pH, low winter temperatures and large body size (Fig. 3a-c).

208         While an association between pH and lateral plate morph is clear, the mechanism by which  
209 pH affects lateral plate evolution is opaque. Dissolved calcium and pH are correlated, and we used  
210 pH as a proxy for dissolved calcium in the present study (Table 1, Fig.S2). In the Hebrides, plateless  
211 and spine-deficient sticklebacks are confined to lochs with low dissolved calcium concentrations  
212 and correspondingly low pH (Giles, 1983; Spence *et al.*, 2013; Klepaker *et al.*, 2016). An extreme  
213 reduction in plates may result from calcium limitation; calcium is required for the normal  
214 development and maintenance of the skeletal system of vertebrates. However, an additional  
215 consideration is that both pH and dissolved calcium correlate with productivity in fresh water (Moss,  
216 2010). This means that any apparent effect of dissolved calcium on plate morph may be mediated  
217 by feeding conditions and, therefore, body size. Since body size predicted plate number, and body  
218 size was predicted by pH (Table 5), the relationship between plate morph and pH may reflect a more  
219 general pattern associated with resource availability and body size, rather than direct selection from  
220 low pH or limited calcium availability.

221         An association between stickleback plate morph and average minimum temperature has been  
222 long recognized, though never satisfactorily explained (Heuts, 1947; Hagen & Moodie, 1982;  
223 Wootton, 2009). The *complete* morph is characteristic of sites that experience low winter

224 temperatures, both marine and fresh water, on the eastern and northern fringes of continents  
225 (Wootton, 1976; Hagen & Moodie, 1982). In contrast, the *low* morph is associated with milder  
226 winter temperatures in the south and west of the geographic range of the species, while the *zero*  
227 morph is associated with mild maritime climates at the southern and western periphery of the  
228 distribution. This pattern was captured in our analysis, which demonstrated a negative relationship  
229 between average minimum temperature and the probability of expressing the *complete* morph (Fig.  
230 3c). Thermal regimes reflect latitude and altitude and, as such, geographical gradients are often  
231 tightly linked to metabolism and phenotypic traits. Average minimum temperature predicted  
232 stickleback body size (Table 5), a prediction of Bergmann's rule (Bergmann, 1847). The causal  
233 explanation for Bergmann's rule is unclear but applies across a range of endotherms and ectotherms,  
234 including fishes (Partridge & Coyne, 1997; Belk & Houston, 2002). In the case of ectotherms, higher  
235 resting metabolic rates may incur higher energetic costs that compromise growth. Selection for large  
236 offspring body size at low temperatures may also play a role (Pettersen *et al.*, 2019). Within  
237 generational plasticity and trans-generational epigenetic effects may also contribute to this pattern.  
238 Female sticklebacks that develop at low temperatures tend to produce fewer but larger offspring  
239 (Shama, 2015), and males raised at high winter temperatures achieve a smaller body size (Kim *et*  
240 *al.*, 2016). Hence, like pH, the relationship between plate morph and temperature may be mediated  
241 through body size.

242         Body size reliably predicted lateral plate morph, as well as lateral plate number within  
243 morphs (Fig. 3c,4c). A model identified pH and temperature, as well as co-occurrence with nine-  
244 spined sticklebacks, as predictors of body size (Table 5). The presence of nine-spined sticklebacks  
245 was negatively associated with three-spined stickleback body size, suggesting that competition from  
246 nine-spined sticklebacks might limit three-spined stickleback growth (MacColl *et al.*, 2013).  
247 However, the mean size of three and nine-spined sticklebacks was correlated among populations

248 (Fig. 5), implying body size in both species is a function of local conditions. Overall then, variance  
249 in lateral plate number was explained directly by body size, or by variables with consequences for  
250 body size. An implication is that lateral plate morph in three-spined stickleback is a function of  
251 scaling (*sensu* Schmidt-Nielsen, 1984).

252         Body size is the most important attribute of an organism, imposing limitations on structure  
253 and function (Bartholomew, 1981). As the scale of biological systems change, so anatomical and  
254 physiological attributes must be altered to enable an organism to function (Schmidt-Nielsen 1984).  
255 If three-spined stickleback morph evolution is a product of scaling rules, a consequence is that  
256 selection on armour must experience scaling constraints. Hence while lateral plates may be the  
257 targets of selection, for instance from predators, this selection must occur within limitations imposed  
258 by scaling relationships; a case of Thompsonian evolution (Thompson, 1917) constraining  
259 Darwinian adaptive evolution (Brock, 2000). Under this model of lateral plate evolution, the  
260 *complete* plate morph is predicted to be associated with environments that favour large body size,  
261 such as marine environments and fresh waters experiencing low winter temperatures. The *low* morph  
262 is predicted in environments that limit body size; primarily freshwater environments and sites with  
263 mild winter temperatures, such as on the Mediterranean coast of Europe and Asia Minor where the  
264 *low* morph is ubiquitous. The *zero* morph, without lateral plates and highly reduced or missing dorsal  
265 spines and pelvic girdle and spines, is predicted in resource-poor environments experiencing mild  
266 temperatures, exemplified by acid lochs and peat bogs in the Hebrides (Giles, 1983). Thus, a  
267 conceptual model of stickleback plate evolution would involve a potential role for multiple  
268 environmental variables in shaping lateral plate number, but with their effects mediated through  
269 body size.

270         If body size predicts lateral plate number, this raises the question of what scaling constraints  
271 generate this relationship. One possibility is that variation in lateral plate number has no Darwinian

272 adaptive advantage and solely reflects scaling (Thompson, 1917). Alternatively, natural selection  
273 may drive changes in body size, with plate number reflecting optimal mechanical efficiency at a  
274 given body size (Bonner & Horn, 2000). In aquatic animals scaling effects have consequences for  
275 hydrodynamic resistance to movement (Schmidt-Nielsen, 1984). Because three-spined sticklebacks  
276 are neutrally buoyant, they expend a negligible amount of energy supporting their bodies in water.  
277 However, energy is needed to offset the resistance of swimming through water; i.e. to overcome  
278 drag. Drag comprises two components. Friction drag derives from the force required to overcome  
279 the viscosity of water while pressure drag stems from the requirement to displace water during  
280 movement. Friction drag is a function of fluid density, surface area and speed of the moving object,  
281 and a drag coefficient (Schmidt-Nielsen, 1984). Small fish swim slower than large fish in the ratio,  
282 all things equal, of the square-root of body length, in accordance with Froude's Law (Thompson,  
283 1917; Videler, 1993). Small fish also incur a relatively greater cost of swimming in comparison with  
284 larger fish, a relationship that applies both within and among species (Videler, 1993). In the three-  
285 spined stickleback the lateral plates increase the drag coefficient; lateral plate number correlates  
286 negatively with swimming velocity after correcting for body size (Bergstrom, 2002). Selection can  
287 compensate for lower swimming speed and efficiency in small fish by minimizing the drag  
288 coefficient. In sticklebacks this can be achieved through armour loss, since armour demonstrably  
289 compromises hydrodynamic efficiency (Walker, 1997; Bergstrom, 2002). Thus, sticklebacks that  
290 experience selection for reduced body size will simultaneously experience selection for plate loss to  
291 avoid compromising swimming speed and energy expenditure. A prediction is for a trade-off  
292 between armour development and growth rate in sticklebacks, which has been demonstrated  
293 experimentally (Barrett *et al.*, 2008).

294           Below a threshold body size selection appears to converge on the *low* plate morph, an armour  
295 configuration that provides robust protection from predators, while facilitating rapid and efficient

296 swimming at a smaller body size. It is noteworthy that the most striking examples of rapid  
297 contemporary change in morph frequency, from domination by the *complete* plate morph to *low*  
298 morph following invasion of marine fish to freshwater sites (e.g. Bell *et al.*, 2004), and the reverse  
299 following environmental change (Kitano *et al.*, 2008), can be explained on the basis of selection for  
300 changes in body size. We further propose that in the case of populations reduced to unusually small  
301 body sizes, like those in acid lochs, complete loss of pelvic apparatus, spines and lateral plates  
302 appears to be favoured under selection as a mechanism to further reduce friction drag and thereby  
303 maintain sufficiently rapid escape responses to enable fish to avoid predators. An implication of our  
304 findings is that elevated temperatures will tend to shift selection pressures on stickleback plate  
305 morph, potentially driving plate loss (Des Roches *et al.*, 2019). As such, stickleback lateral plate  
306 phenotype offers a powerful tool for monitoring change in climate variables.

307

## 308 **CONCLUSION**

309 We used an information theory approach to identify body size, average minimum temperature, and  
310 pH as the primary predictors of the parallel evolution of stickleback lateral plate phenotype. Our  
311 results indicate that lateral plate armour development is underpinned by selection for body size,  
312 which has implications for swimming velocity and predator avoidance. Temperature-linked changes  
313 in stickleback plate morph offers a tool for monitoring temperature trends across the northern  
314 temperate region.

315

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321

## 322 **References**

323 Bañbura, J. (1989). Lateral plate number development in the complete morph of the three-spined  
324 stickleback, *Gasterosteus aculeatus* L. *Zool. Script.* **18**, 157-159.

325 Barrett, R.D.H. (2010). Adaptive evolution of lateral plates in three-spined stickleback *Gasterosteus*  
326 *aculeatus*: a case study in functional analysis of natural variation. *J. Fish Biol.* **77**, 311-328.

327 Barrett, R.D., Rogers, S.M. & Schluter, D. (2008). Natural selection on a major armor gene in  
328 threespine stickleback. *Science* **322**, 255-257.

329 Bartholomew, G.A. (1981). A matter of size: an examination of endothermy in insects and terrestrial  
330 vertebrates. In *Insect thermoregulation*: 45-78. Heindrich, B. (Ed.). London: Wiley.

331 Belk, M.C. & Houston, D.D. (2002). Bergmann's rule in ectotherms: A test using freshwater  
332 fishes. *Am. Nat.* **160**, 803-808.

333 Bell, M.A., Aguirre, W.E. & Buck, N.J. (2004). Twelve years of contemporary armor evolution in  
334 a threespine stickleback population. *Evolution* **58**, 814-824.

335 Bell, M.A. & Foster, S.A. (1994). Introduction to the evolutionary biology of the threespine  
336 stickleback. In *The evolutionary biology of the threespine stickleback*: 1-27. Bell, M.A. & Foster,  
337 S.A. (Ed.). Oxford University Press.

338 Bergmann, C. (1847). Über die verhältnisse der warmeökonomie de thiery zu ihrer gross. *Göttinger*  
339 *Studien* **1**, 595-708.

340 Bergstrom, C.A. (2002). Fast-start swimming performance and reduction in lateral plate number in  
341 threespine stickleback. *Can. J. Zool.* **80**, 207-213.

342 Bolnick, D.I., Barrett, R.D., Oke, K.B., Rennison, D.J. & Stuart, Y.E. (2018). (Non)parallel  
343 evolution. *Ann. Rev. Ecol. Evol. Syst.* **49**, 303-330.

344 Bonner, J.T. & Horn, H.S. (2000). Allometry and natural selection. In *Scaling in biology*: 1-24.  
345 Brown, J.H. & West, G.B. (Ed.). Oxford University Press.

346 Brock, J.P. (2000). *The evolution of adaptive systems: The general theory of evolution*. London:  
347 Academic Press.

348 Brubaker, J.M. & Angus, R.A. (1984). A procedure for staining fish with alizarin without causing  
349 exfoliation of scales. *Copeia* **1984**, 989-990.

350 Burnham, K.P. & Anderson, D.R. (2002). *Model selection and multimodel inference*. Berlin:  
351 Springer.

352 Carlson, A.E. (2013). The Younger Dryas climate event. In *The encyclopedia of quaternary science*  
353 *Vol. 3*: 126–134. Elias, S.A. (Ed.). Amsterdam: Elsevier.

354 Colosimo, P.F., Hosemann, K.E., Balabhadra, S., Villarreal, G., Dickson, M., Grimwood, J.  
355 Schmutz, J., Myers, R.M., Schluter, D. & Kingsley, D.M. (2005). Widespread parallel evolution  
356 in sticklebacks by repeated fixation of ectodysplasin alleles. *Science* **307**, 1928-1933.

357 Cresko, W.A., Amores, A., Wilson, C., Murphy, J., Currey, M., Phillips, P., Bell, M.A., Kimmel,  
358 C.B. & Postlethwait, J.H. (2004). Parallel genetic basis for repeated evolution of armor loss in  
359 Alaskan threespine stickleback populations. *Proc. Nat. Acad. Sci.* **101**, 6050-6055.

360 Des Roches, S., Bell, M.A. & Palkovacs, E.P. (2019). Climate-driven habitat change causes  
361 evolution in threespine stickleback. *Glob. Change Biol.* DOI10.1111/gcb.14892.

362 Giles, N. (1983). The possible role of environmental calcium levels during the evolution of  
363 phenotypic diversity in Outer Hebridean populations of the three-spined stickleback,  
364 *Gasterosteus aculeatus*. *J. Zool.* **199**, 535-544.

365 Hagen, D.W. & Moodie, G.E.E. (1982). Polymorphism for plate morphs in *Gasterosteus aculeatus*  
366 on the east coast of Canada and an hypothesis for their global distribution. *Can. J. Zool.* **60**, 1032-  
367 1042.

368 Haldane, J.B.S. (1932). *The causes of evolution*. London: Longman.

369 Heuts, M.J. (1947). Experimental studies on adaptive evolution in *Gasterosteus aculeatus*  
370 L. *Evolution* **1**, 89-102.

371 Hilbe, J.M. (2014). *Modeling count data*. Cambridge University Press.

372 Hoogland, R., Morris, D. & Tinbergen, N. (1956). The spines of sticklebacks (*Gasterosteus* and  
373 *Pygosteus*) as means of defence against predators (*Perca* and *Esox*). *Behaviour* **10**, 205-236.

374 Jones, F.C., Grabherr, M.G., Chan, Y.F., Russell, P., Mauceli, E., Johnson, J., Swofford, R., Pirun,  
375 M., Zody, M.C., White, S. & Birney, E. (2012). The genomic basis of adaptive evolution in  
376 threespine sticklebacks. *Nature* **484**, 55.

377 Klepaker, T., Østbye, K., Spence, R., Warren, M. Przybylski, M. & Smith, C. (2016). Selective  
378 agents in the adaptive radiation of Hebridean sticklebacks. *Evol. Ecol. Res.* **17**, 243-262.

379 Kim, S.Y., Costa, M.M., Esteve-Codina, A. & Velando, A. (2017). Transcriptional mechanisms  
380 underlying life-history responses to climate change in the three-spined stickleback. *Evol. Appl.*  
381 **10**, 718-730.

382 Kitano, J., Bolnick, D.I., Beauchamp, D.A., Mazur, M.M., Mori, S., Nakano, T. & Peichel, C.L.  
383 (2008). Reverse evolution of armor plates in the threespine stickleback. *Curr. Biol.* **18**, 769-774.

384 MacColl, A.D. Nagar, A.E. & de Roij, J. (2013). The evolutionary ecology of dwarfism in three-  
385 spined sticklebacks. *J. Anim. Ecol.* **82**, 642-652.

386 Magalhaes, I.S., D'Agostino, D., Hohenlohe, P.A. & MacColl, A.D. (2016). The ecology of an  
387 adaptive radiation of three-spined stickleback from North Uist, Scotland. *Mol. Ecol.* **25**, 4319-  
388 4336.

389 Morozińska-Gogol, J. (2011). Changes in levels of infection with *Schistocephalus solidus* of the  
390 three-spined stickleback *Gasterosteus aculeatus* (Actinopterygii: Gasterosteidae) from the  
391 Gdynia Marina. *Oceanologia* **53**, 181-187.

392 Moss, B.R. (2010). *Ecology of fresh waters*. Oxford: Wiley-Blackwell.

393 Münzing, J. (1963). The evolution of variation and distributional patterns in European populations  
394 of the three-spined stickleback, *Gasterosteus aculeatus*. *Evolution* **17**, 320-332.

395 Myhre, F. & Klepaker, T. (2009). Body armour and lateral-plate reduction in freshwater three-spined  
396 stickleback *Gasterosteus aculeatus*: adaptations to a different buoyancy regime? *J. Fish Biol.* **75**,  
397 2062-2074.

398 Partridge, L. & Coyne, J.A. (1997). Bergmann's rule in ectotherms: is it adaptive? *Evolution* **51**,  
399 632-635.

400 Pettersen, A.K., White, C.R., Bryson-Richardson, R.J. & Marshall, D.J. (2019). Linking life-history  
401 theory and metabolic theory explains the offspring size-temperature relationship. *Ecol. Lett.* **22**,  
402 518-526.

403 Pujolar, J.M., Ferchaud, A.L., Bekkevold, D. & Hansen, M.M. (2017). Non-parallel divergence  
404 across freshwater and marine three-spined stickleback *Gasterosteus aculeatus* populations. *J.*  
405 *Fish Biol.* **91**, 175-194.

406 R Development Core Team (2019). *R: A language and environment for statistical computing*.  
407 Vienna: R Foundation for Statistical Computing.

408 Raeymaekers, J.A., Van Houdt, J.K., Larmuseau, M.H., Geldof, S. & Volckaert, F.A. (2007).  
409 Divergent selection as revealed by PST and QTL-based FST in three-spined stickleback  
410 (*Gasterosteus aculeatus*) populations along a coastal-inland gradient. *Mol. Ecol.* **16**, 891-905.

411 Reimchen, T.E. 1994. Predators and evolution in threespine stickleback. In *The evolutionary*  
412 *biology of the threespine stickleback*: 240-273. Bell, M.A. & Foster, S.A. (Ed.). Oxford  
413 University Press.

414 Reimchen, T.E., Bergstrom, C. & Nosil, P. (2013). Natural selection and the adaptive radiation of  
415 Haida Gwaii stickleback. *Evol. Ecol. Res.* **15**, 241-269.

416 Rue, H., Riebler, A., Sørbye, S.H., Illian, J.B., Simpson, D.P. & Lindgren, F.K. (2017). Bayesian  
417 computing with INLA: a review. *Ann. Rev. Stat. Appl.* **4**, 395-421.

418 Shama, L.N. (2015). Bet hedging in a warming ocean: predictability of maternal environment shapes  
419 offspring size variation in marine sticklebacks. *Global Change Biol.* **21**, 4387-4400.

420 Schmidt-Nielsen, K. (1984). *Scaling: Why is animal size so important?* Cambridge University Press.

421 Smith, C. & Warren, M. (2019). *GLMs in R for Ecology*. Amazon Publishing.

422 Spence, R., Wootton, R.J., Barber, I., Przybylski, M. & Smith, C. (2013). Ecological causes of  
423 morphological evolution in the three-spined stickleback. *Ecol. Evol.* **3**, 1717-1726.

424 Thompson, D.W. (1917). *On growth and form*. Cambridge University Press.

425 Vehtari, A., Gelman, A. & Gabry, J. (2017). Practical Bayesian model evaluation using leave-one-  
426 out cross-validation and WAIC. *Stat. Comp.* **27**, 1413-1432.

427 Videler, J.J. (1993). *Fish swimming*. London: Chapman and Hall.

428 Walker, J.A. (1997). Ecological morphology of lacustrine threespine stickleback *Gasterosteus*  
429 *aculeatus* L. (Gasterosteidae) body shape. *Biol. J. Linn. Soc.* **61**, 3-50.

430 Wootton, R.J. (1976). *Biology of the sticklebacks*. London: Academic Press.

431 Wootton, R.J. (1984). *A functional biology of sticklebacks*. London: Croom Helm.

432 Wootton, R.J. (2009). The Darwinian stickleback *Gasterosteus aculeatus*: a history of evolutionary  
433 studies. *J. Fish Biol.* **75**, 1919-1942.

434

435 **Figure legends**

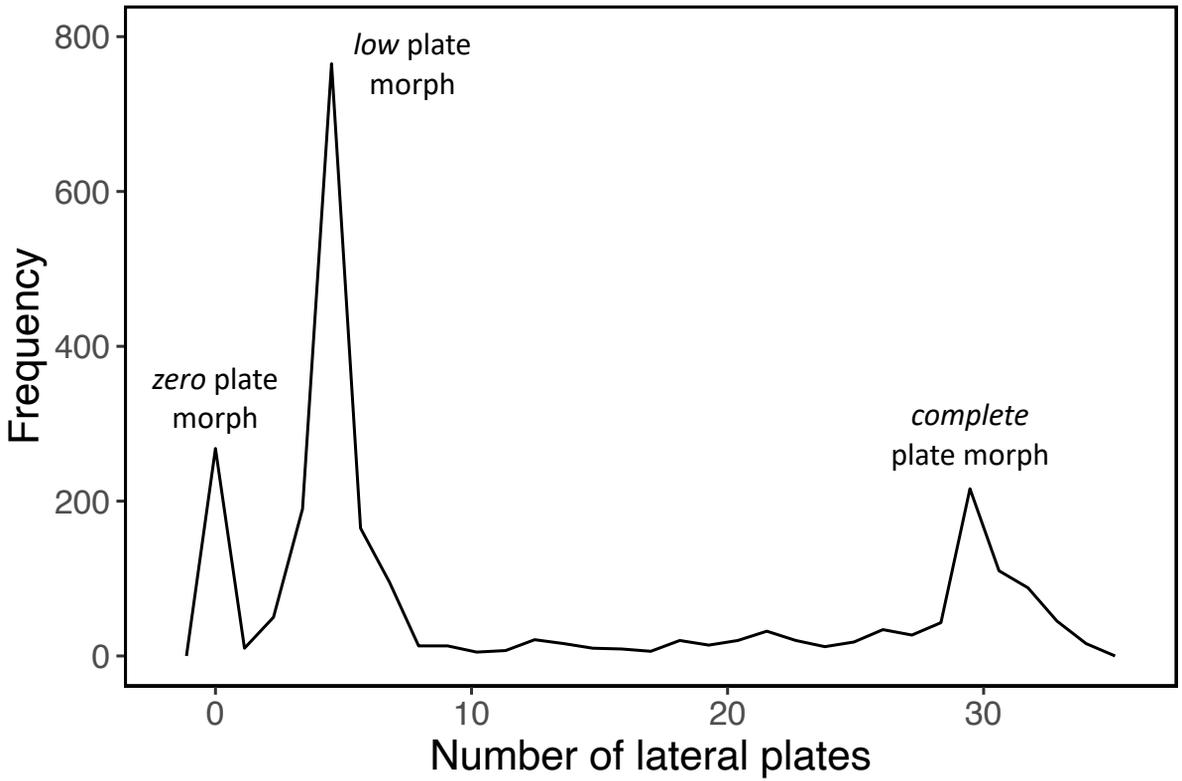
436 **Figure 1** Frequency distribution of three-spined stickleback lateral plate number for 2,358  
437 individuals from 85 European populations.

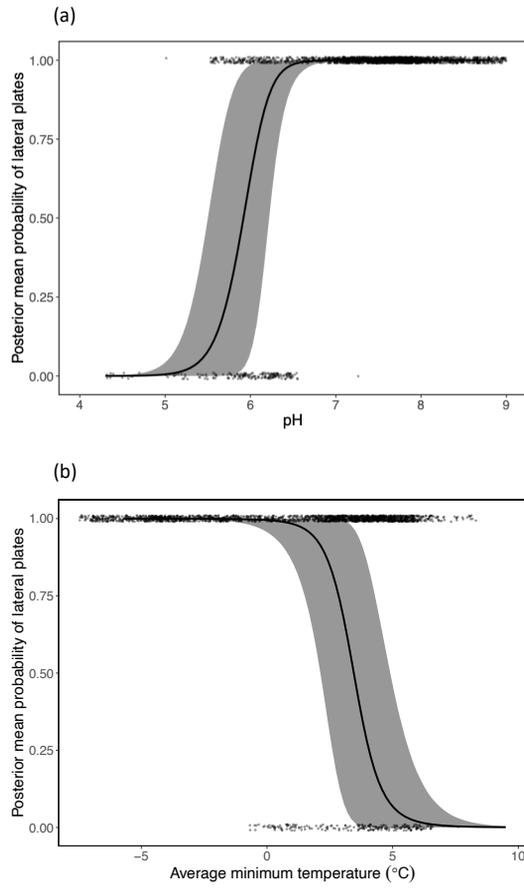
438 **Figure 2.** (a) Posterior mean fitted probability of three-spined sticklebacks possessing lateral plates  
439 as a function of pH; (b) average minimum environmental temperature (°C). Shaded areas are 95%  
440 credible intervals. Black points are observed data for individual fish.

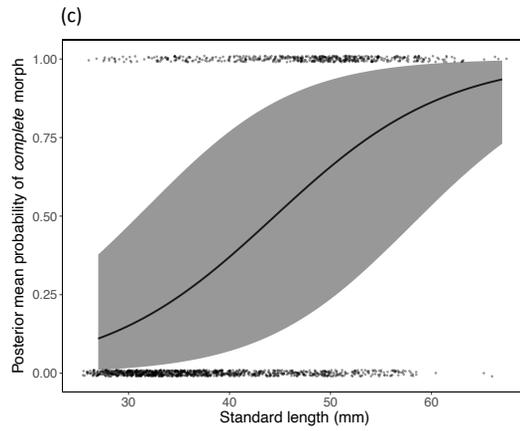
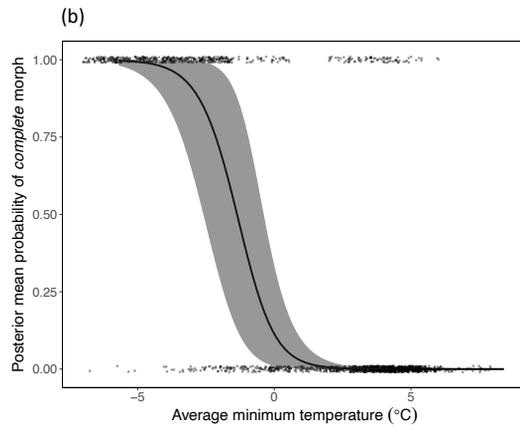
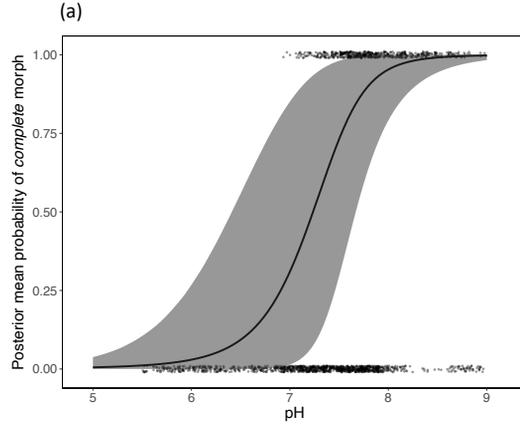
441 **Figure 3.** (a) Posterior mean probability of expressing the *complete* lateral plate morph as a function  
442 of pH; (b) average minimum environmental temperature (°C); (c) standard length (mm). Shaded  
443 areas are 95% credible intervals. Black points are observed data for individual fish.

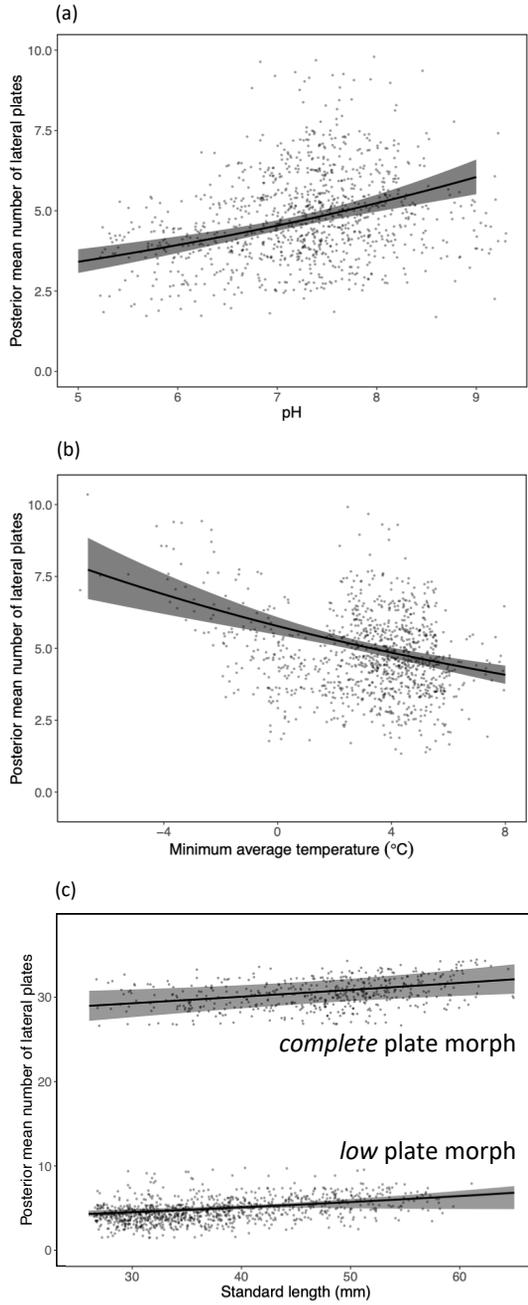
444 **Figure 4.** Posterior mean fitted number of lateral plates for three-spined sticklebacks for: (a) the  
445 *low* plate morph as a function of pH; (b) the *low* plate morph as a function of average minimum  
446 environmental temperature (°C); (c) *low* and *complete* morphs as a function of standard length  
447 (mm). Shaded areas are 95% credible intervals. Black points are observed data for individual fish.

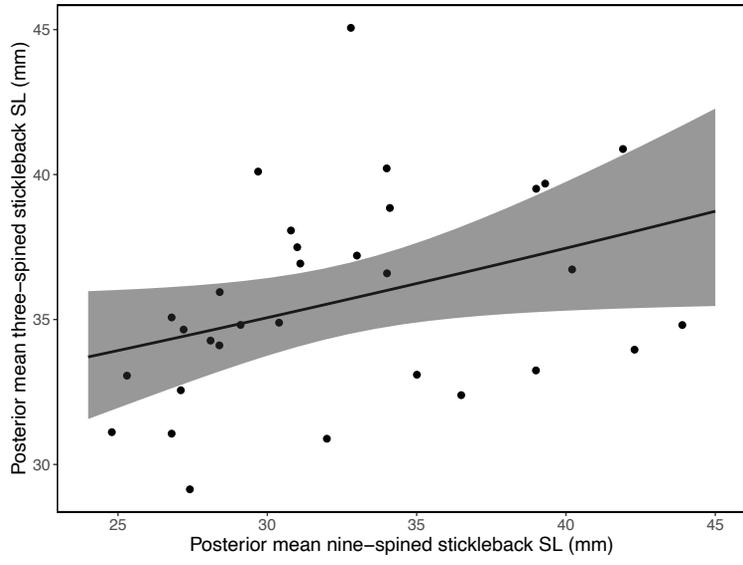
448 **Figure 5.** Posterior mean standard length (mm) of three-spined sticklebacks as a function of nine-  
449 spined stickleback standard length (mm). Shaded areas are 95% credible intervals. Black points  
450 are observed data for different populations.











**Table 1.** *A priori* models for the evolution of lateral plate morph. In some cases multiple authors have proposed and tested the same model. We cite a single source for each model for brevity, giving either the first author to propose the model or the one that most fully articulated it.

model	fitted model	source	model description
M01	temperature + salinity	Heuts (1947)	average minimum temperature, salinity
M02	presence/absence of fish predators	Hoogland <i>et al.</i> (1956)	vertebrate predation
M03	latitude x longitude	Münzig (1963)	geographical location
M04	temperature	Wootton (1976)	average minimum temperature
M05	presence/absence dragonfly larvae	Reimchen (1994)	invertebrate predation
M06	pH	Giles (1983)	dissolved calcium
M07	elevation	Raeymaekers <i>et al.</i> (2007)	coastal-inland gradient
M08	salinity	Myhre & Klepaker (2009)	water density
M09	presence/absence <i>Schistocephalus solidus</i>	Morozińska-Gogol (2011)	parasitism by <i>S. solidus</i>
M10	presence/absence <i>Pungitius pungitius</i>	MacColl <i>et al.</i> (2013)	competition with <i>P. pungitius</i>
M11	turbidity + presence/absence of fish predators	Reimchen <i>et al.</i> (2013)	light spectra and vertebrate predation
M12	pH + presence/absence of fish predators	Spence <i>et al.</i> (2013)	dissolved calcium, vertebrate predation
M13	pH + presence/absence of fish predators + turbidity	Klepaker <i>et al.</i> (2016)	dissolved calcium, vertebrate predation, light spectra
M14	presence/absence of fish predators + <i>P. pungitius</i>	Magalhaes <i>et al.</i> (2016)	vertebrate predation, competition with <i>P. pungitius</i>
M15	temperature + standard length + pH	this study	average minimum temperature, body size, dissolved calcium

**Table 2.** Best-fitting models for expression of lateral plates, expression of either a *low* and *complete* morph, and for the number of plates expressed by *low* and *complete* morphs. WAIC is Watanabe-Akaike Information Criterion score,  $\Delta i$  is delta WAIC,  $\omega$  is WAIC weighting.

Expression of lateral plates				Expression of <i>low</i> or <i>complete</i> morph				Number of <i>low</i> morph plates				Number of <i>complete</i> morph plates			
model	WAIC	$\Delta i$	$\omega$	model	WAIC	$\Delta i$	$\omega$	model	WAIC	$\Delta i$	$\omega$	model	WAIC	$\Delta i$	$\omega$
M15	194	0.0	0.56	M15	160	0.0	0.26	M15	3864	0	1.00	M15	2760	0.0	0.47
M05	196	2.0	0.20	M13	160	0.6	0.19	M01	3883	19.1	0.00	M07	2764	3.8	0.07
M12	198	4.0	0.09	M14	161	0.8	0.17	M08	3887	22.9	0.00	M10	2764	4.1	0.06
M02	199	5.0	0.04	M04	161	1.4	0.13	M05	3888	24.5	0.00	M08	2764	4.2	0.06
M06	199	5.0	0.04	M10	161	1.6	0.12	M12	3888	24.7	0.00	M09	2765	4.8	0.04
M10	199	5.0	0.04	M09	163	3.0	0.06	M13	3890	26.1	0.00	M01	2765	4.8	0.04
M09	200	6.0	0.03	M12	163	3.3	0.05	M04	3892	27.7	0.00	M05	2765	5.1	0.04
M14	202	8.0	0.01	M02	165	5.0	0.02	M11	3892	28.6	0.00	M04	2765	5.1	0.04
M07	220	26.0	0.00	M05	168	8.2	0.00	M03	3893	29.1	0.00	M14	2765	5.1	0.04
M08	220	26.0	0.00	M08	172	12.5	0.00	M02	3894	30.5	0.00	M06	2765	5.2	0.04
M13	224	30.0	0.00	M06	177	16.9	0.00	M07	3894	30.5	0.00	M02	2765	5.2	0.04
M03	245	51.0	0.00	M07	219	59.3	0.00	M14	3895	30.8	0.00	M12	2766	6.1	0.02
M11	266	72.0	0.00	M11	225	65.1	0.00	M06	3895	30.9	0.00	M03	2766	6.1	0.02
M04	468	274.0	0.00	M03	229	69.1	0.00	M10	3895	31.4	0.00	M11	2767	6.9	0.01
M01	8622	8428	0.00	M01	4092	3932	0.00	M09	3896	32	0.00	M13	2768	8.1	0.01

**Table 3.** Posterior mean estimates for the expression of lateral plates in three-spined sticklebacks modelled with a Bernoulli GLMM fitted using INLA. Stickleback population of origin and nearest meteorological station were fitted as random intercepts. CrI are the Bayesian credible intervals. Credible intervals that do not encompass zero indicate statistical importance.

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Parameter	Posterior mean	Lower CrI	Upper CrI
Intercept	-53.43	-108.30	-26.22
SL	0.04	-0.06	0.15
Temperature	-2.89	-6.68	-1.00
pH	10.35	5.26	21.18

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**Table 4.** Posterior mean estimates for the expression of lateral plates by *low* and *complete* morph three-spined sticklebacks for the best-fitting hurdle model using INLA. Stickleback population of origin and nearest meteorological station were fitted as a random intercepts. Frequency data were fitted to a Poisson distribution. CrI are the Bayesian credible intervals. Credible intervals that do not encompass zero indicate statistical importance.

Parameter	Occurrence model			Frequency model ( <i>low</i> )			Frequency model ( <i>complete</i> )		
	Posterior mean	Lower CrI	Upper CrI	Posterior mean	Lower CrI	Upper CrI	Posterior mean	Lower CrI	Upper CrI
Intercept	-4.78	-7.88	-2.77	1.55	1.49	1.63	3.42	3.40	3.44
SL	1.29	0.88	1.72	0.10	0.07	0.15	0.02	0.01	0.04
Temperature	-6.44	-10.20	-4.14	-0.10	-0.21	-0.04	0.01	-0.01	0.02
pH	3.47	1.42	6.37	0.12	0.07	0.23	-0.01	-0.02	0.02

**Table 5.** Posterior mean estimates for three-spined stickleback standard length as a function of the presence of nine-spined sticklebacks, average minimum temperature and pH, modelled using a gamma GLMM. Stickleback population of origin and month of collection were fitted as random intercepts. CrI is the 95% Bayesian credible interval. Credible intervals that do not encompass zero indicate statistical importance.

Model parameter	Posterior mean	Lower CrI	Upper CrI
Intercept	3.67	3.58	3.74
Nine-spined sticklebacks <sub>(present)</sub>	-0.12	-0.19	-0.05
Temperature	-0.13	-0.16	-0.09
pH	0.05	0.03	0.08

**Table 6.** Posterior mean estimates for the SL of three-spined sticklebacks as a function of nine-spined stickleback SL, modelled with a gamma GLM fitted using INLA. CrI are the Bayesian credible intervals. Credible intervals that do not encompass zero indicate statistical importance.

Parameter	Posterior mean	Lower CrI	Upper CrI
Intercept	3.220	2.887	3.552
Nine-spined stickleback SL	0.011	0.001	0.021