

Reducing arsenic toxicity using the interfacial oxygen nanobubble technology in sediment remediation

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Abstract

The worldwide arsenic (As)-bearing eutrophic waters may suffer from the dual conditions of harmful algal blooms and release of As, driven by algal-induced hypoxia/anoxia. Here, we investigate the use of interfacial oxygen (O₂) nanobubble technology to combat the hypoxia and control As exposure in simulated mesocosm experiments. It was observed that remediation of algal-induced hypoxia at the sediment-water interfaces (SWI) by application of O₂ nanobubbles reduced the level of dissolved As from 23.2 µg L⁻¹ to <10 µg L⁻¹ and stimulated the conversion of As(III) to the less toxic As(V) (65–75%) and methylated As (10–15%) species. More than half of the oxidation and all the methylation of As(III) resulted from the manipulation by O₂ nanobubbles of microbes responsible for As(III) oxidation and methylation. Hydroxyl radicals were generated during the oxidation of reductive substances at the SWI in darkness, and should be dominant contributors to As(III) abiotic oxidation. X-ray absorption near-edge structure (XANES) spectroscopic

27 analysis demonstrated that surface sediments changed from being sources to acting as
28 sinks of As, due to the formation of Fe-(hydr)oxide. Overall, this study suggests that
29 interfacial O₂ nanobubble technology could be a potential method for remediation of
30 sediment As pollution through the manipulation of O₂-related microbial and geochemical
31 reactions.

32 **Keywords:** Eutrophic waters; Arsenic oxidation; Nanobubble; Hypoxia remediation;
33 Arsenic metabolism functional genes; Hydroxyl radicals

34

35 **1. Introduction**

36 Levels of Arsenic (As) are increasing in the environment and posing threats in
37 surface water ecosystems (e.g. rivers and lakes) through both natural and anthropogenic
38 activities (Weber et al., 2010; Zhang et al., 2018a). Drinking water **directly** derived from
39 such contaminated sources poses a human health risk for more than 150 million people
40 globally (Kay, 2011). Moreover, **As-bioaccumulation and magnification of As** through
41 aquatic foods web-**magnification**, and uptake **it** by crops through irrigation with As-
42 contaminated water also significantly contribute to public health issues (**Huang,**
43 **2016Zavala and Duxbury, 2008**). ~~Thus, development of As mitigation strategies to~~
44 ~~achieve and maintain low As concentrations in surface waters is crucial towards~~
45 ~~environmental sustainability, food security and in safeguarding global human health.~~

46 In **aquatic environments surface waters**—where the **bottom water—benthic**
47 **environment** is well-oxygenated, As is commonly bound with Fe, Mn, and organic carbon
48 within the sediment (Couture et al., 2010). ~~However,~~ Harmful algal blooms (HABs) are

49 ~~also~~ increasing in aquatic ecosystems globally, owing to increases in episodic
50 eutrophication and as a consequence of global warming (Huisman et al., 2018; Pan et al.,
51 2018). The subsequent oxygen (O₂) depletion by mineralization of senescent algal blooms
52 induces hypoxia/anoxia at the ~~sediment-water interfaces (SWI), potentially causing the~~
53 ~~mobilization of various nutrients and pollutants from sediment to water column~~ (Lei et
54 al., 2019; Tang et al., 2019; ~~Wang et al., 2018~~). Previous field investigations have
55 demonstrated that eutrophication ~~induced hypoxia/anoxia~~ might favour the ~~reductive~~
56 ~~dissolution of As(V)/As(III)-bearing iron oxides in the surface sediments,~~
57 ~~biogeochemical cycling of As~~ and thus induce ~~its~~ endogenous release of As to the water
58 ~~column from lake sediment~~ (Hasegawa et al., 2009; Yan et al., 2016). ~~During algal induced~~
59 ~~periods of hypoxia/anoxia, As levels in eutrophic waters can exceed the drinking water~~
60 ~~limits (10 µg L⁻¹), set by the World Health Organization (WHO), by several orders of~~
61 ~~magnitude (Hartland et al., 2015; Martin and Pedersen, 2004)~~. Moreover, reduction of
62 As(V) to As(III) (a more toxic As species) could be stimulated by the activity of
63 indigenous As(V)-~~reducing~~ ~~tive~~ microbes under hypoxic/anoxic conditions (Tang et al.,
64 2019). Therefore, combating the hypoxic/anoxic conditions at the SWI could be an
65 efficient strategy by which to control As levels and toxicity in eutrophic surface waters.

66 Traditional oxygenation methods, such as deep water aeration and the artificial
67 mixing of surface and bottom waters (Bierlein et al., 2017), have been reported to be
68 effective to some extent. However, these approaches have been hindered when used in
69 large and/or deep waters, due to their capital costs and energy consumption (Bormans et
70 al., 2016; Funkey et al., 2014). The capital cost of installation of pumps in a deep water
71 body is estimated to cost approximately €0.3 million/km² (Stigebrandt and Gustafsson,

72 2007; Zhang et al., 2021). Additional operational costs of nearly €0.2-0.25
73 million/km²/year may be incurred, due to the continuous operation required during the
74 summer season, or even the whole year, in order to maintain the O₂ supply and resultant
75 nutrients/pollutants control at the SWI (Bormans et al., 2015; Johnston et al., 2011).
76 Moreover, the induced massive hydraulic disturbance caused by traditional aeration
77 methods through pumps may trigger the re-suspension of anoxic sediments. Such re-
78 suspended sediments could release the long-term buried pollutants/nutrients back to the
79 water column and increase the O₂ consumption (Bormans et al., 2015). Furthermore, the
80 change of the status could affect the stable habitats for the benthic biota. ~~even affect~~
81 ~~benthic biota and lead to increased O₂ consumption and secondary release of~~
82 ~~pollution/nutrients, resulting from the re-suspension of anoxic sediments~~ (Zhang et al.,
83 2021).

84 In order to overcome these drawbacks, ~~an~~ the interfacial O₂ nanobubble technology
85 has been developed and attempted to address this issue in a cost-effective and
86 environmental-friendly way (Zhang et al., 2018b). ~~to~~ This approach utilises natural
87 minerals loaded with O₂, which could deliver nanoscale O₂ bubbles (bubble size < 1 µm)
88 after settling into the sediment surface by natural gravity, ~~a cost effective and~~
89 ~~environmental friendly way (Zhang et al., 2018b)~~. This earlier study also demonstrated
90 synergy between the diffusion of O₂ nanobubbles and retention of O₂ in the surficial
91 sediment layer, which successfully reversed the hypoxic conditions and maintained high
92 levels of dissolved O₂ (6 mg L⁻¹) at the SWI over a period of 4 months (Zhang et al.,
93 2018b). The successful performance could be attributed mainly to the novel physical and
94 chemical features of nanobubbles including long lifespan, high gas transfer efficiency,

95 and the capability to generate free radicals, as compared to larger bubbles (Atkinson et
96 al., 2019; Lyu et al., 2019). ~~Following this concept, further studies have been conducted~~
97 ~~by different researchers, however, these have been mainly focused on the treatment of~~
98 ~~hypoxia/anoxia conditions (Yu et al., 2019), nutrient turnover control (including nitrogen~~
99 ~~and phosphorus) (Zhang et al., 2020; Zhang et al., 2021), and modulation of~~
100 ~~mineralization of organics (Shi et al., 2018).~~ However, The effects of interfacial
101 nanobubble technology on the control, and biogeochemical cycling, of toxic metal
102 pollutants, such as As, during sediment remediation have not, hitherto, been well
103 understood.

104 Once the O₂ nanobubbles were delivered to the hypoxic/anoxic SWI through
105 minerals acting as carriers, we expected that the resultant hypoxic/anoxic-oxic transition
106 could trigger As(III) oxidation through both biotic and abiotic processes. On the one hand,
107 the supply of substantially dissolved O₂ ~~supply~~ may have stimulated the growth of the
108 indigenous As(III) oxidizing ~~ase~~ microbes and induced biotic oxidation of As(III) to less
109 toxic As(V) (Maguffin et al., 2015), which is prone to be immobilized by iron oxides
110 minerals (Luong et al., 2018). On the other hand, the abiotic process of As(III) oxidation
111 may have been caused by the generated hydroxyl radicals ($\bullet\text{OH}$), as a highly reactive and
112 unselective oxidant, ~~generated~~ during the nanobubble treatment. Previous studies have
113 proved that $\bullet\text{OH}$ could be generated along with the natural bursting of O₂ nanobubbles
114 (Atkinson et al., 2019). Moreover, the oxygenation of anoxic sediment/water containing
115 abundant ferrous minerals and reduced organic matter may also produce abundant $\bullet\text{OH}$
116 through Fenton or Fenton-like processes in darkness (Liao et al., 2019a; Liao et al., 2019b;
117 Tong et al., 2016). ~~Compared to the biotic oxidation of As(III), the abiotic oxidation~~

118 ~~process of As(III) is usually a rapid reaction that might occur within hours (Luong et al.,~~
119 ~~2018; Page et al., 2013; Tong et al., 2016).~~ The accompanied variation in **concentration**
120 **and speciation of** Fe and organic matter may influence their bioavailability to microbial
121 activity (Tang et al., 2019; Zhu et al., 2017), and thereby indirectly affect the biotic
122 oxidation of As(III). Thus far, the hypothesised biotic and abiotic mechanisms and their
123 quantitative contributions on As speciation in eutrophic waters have not been investigated.

124 The goal of this study was to evaluate the effectiveness, and to reveal the
125 mechanism for the mitigation of As pollution by interfacial O₂ nanobubble technology in
126 a simulated eutrophic sediment-water system. The overlying water quality and As
127 speciation were recorded in order to evaluate the performance of hypoxic/anoxic
128 reversion and As immobilization at the SWI. As and Fe K-edge **X-ray absorption near-**
129 **edge structure (XANES)** spectral analyses were used to investigate sequestration
130 mechanisms of As in the sediment, and genes responsible for As metabolism (Tang et al.,
131 2019) (i.e., As(III) oxidation (*aioA*), As(V) respiratory reduction (*arrA*), As(V) reduction
132 (*arsC*), and As(III) methylation (*arsM*)) extracted from the SWI were quantified to aid in
133 the understanding of the biotic processes of As transformation. Additionally, •OH
134 generation and its effect on As oxidation were studied in order to evaluate abiotic effects
135 on As speciation.

136 **2. Materials and methods**

137 ***2.1 Sample collection and construction of mesocosm columns***

138 Lake water and sediments were collected from Lake Dianchi (Fig. S1), which is the
139 sixth largest freshwater lake in China and, historically, has suffered from severe As

140 contamination and the annual occurrence of cyanobacterial blooms (Zhou et al. 2016).
141 Concentrations of As in the sediment and water column have been recorded to reach 154
142 mg kg⁻¹ and 7.23 µg L⁻¹, respectively (Tang et al., 2019). After transferring to the
143 laboratory, the original lake water collected from Dianchi was filtrated through a 0.2 µm
144 membrane (Sartorius AG, Germany), in order to minimize the background concentration
145 of algal biomass prior to any treatment. Then the filtrated lake water (600 mL) and
146 sediments (220 mL) were then placed in 24 plexiglass cylinders (Ø64 mm, length 300
147 mm) in order to simulate water-sediment systems (Fig. S1). All of the mesocosm columns
148 were stabilized under dark conditions for three weeks, then algal bloom scum
149 (*Microcystis aeruginosa*, 250 mg dry weight L⁻¹ water), collected from Lake Dianchi, was
150 added to all columns and flocculated by starch-modified soil (Jin et al., 2019) and then
151 allowed to settle under gravity to the sediment surface, in order to simulate algal-induced
152 hypoxia/anoxia at the SWI.

153 ***2.2 Interfacial nanobubble materials preparation and operation***

154 Both O₂ and N₂ nanobubble-modified zeolites were prepared by a previously-
155 described method (Zhang et al., 2018b), the details of which are also included in the
156 Supporting Information. After preparation of the algae biomass and of the zeolite
157 materials, six groups of columns (four replicates of each) were prepared as (Table S1): i)
158 the control group; ii) the **pristine zeolite (Air-Ze) treatment** group; iii) the **N₂ nanobubble-**
159 **modified zeolite (N₂-Ze) treatment** group; iv) the **O₂ nanobubble-modified zeolite (O₂-**
160 **Ze) treatment** group; v) the **sterilized control (S-control)** group; and vi) the **sterilized O₂**
161 **nanobubble-modified zeolite (S-O₂-Ze) treatment** group. Among these mesocosms, the

162 Air-Ze and N₂-Ze groups were designed to identify As transformations resulting solely
163 from the physical capping treatment or from any synergistic effects accompanying
164 nanobubble treatment, during the experimental period of 18 days at 25 °C under dark
165 conditions. The S-control and S-O₂-Ze groups were sterilized prior to commencement of
166 the experiment groups, to avoid any microbial contribution to the As transformation
167 process to the greatest extent, as well as to aid in the identification of any abiotic effects.
168 In these latter two groups, all materials, including sediments, water, zeolites and algae
169 biomass were sterilized as described in our previous study (Tang et al., 2019). The
170 experiment conditions for the different groups are summarized in Table S1 in the
171 Supporting Information.

172 ***2.3 Sampling and measurement***

173 In order to monitor water quality and As mobilization during the interfacial O₂
174 nanobubble treatment, DO (dissolved oxygen) and Eh (redox potential) at the SWI were
175 measured at days 0, 2, 4, 7, 12 and 18. At the same time, the overlying water at 4 cm
176 above the SWI was sampled from the columns in groups i-iv and filtered through a 0.45
177 µm membrane (Whatman, UK), to measure the total dissolved As and the As speciation,
178 including As(III), As(V), and methylated As (i.e. monomethylarsonate (MMA(V)) and
179 dimethylarsinate (DMA(V))). ~~Filtration by 0.45 µm membrane is a common method for~~
180 ~~the isolation of dissolved chemical species from overlying water and associated~~
181 ~~suspended solids (Johnston et al., 2011; Tang et al., 2019; Yan et al., 2016; Zhang et al.,~~
182 ~~2021).~~

183 Due to the clearly superior performance of O₂-Ze for As mitigation, the control (i)

184 and O₂-Ze (iv) groups were employed for further mechanistic investigations. Sediment
185 porewater samples were collected by Rhizons MOM samplers (Rhizosphere Research
186 Products B.V., Wageningen, The Netherlands) from 1 cm above the SWI to 4 cm below
187 with 1-cm intervals, at days 0, 2, 4, 7, 12, and 18. As this lab-scale study was conducted
188 in isolated, well-controlled columns, the effects of dissolved forms of Fe and organic
189 matter as suspended particles and/or in the colloidal size range, during water sample
190 analysis, can be neglected (Wu et al., 2001). Sediment porewater samples and the
191 aforementioned overlying water samples were filtered through a 0.45 µm membrane prior
192 to determination of dissolved organic carbon (DOC), dissolved Fe(II) and total dissolved
193 Fe. At the conclusion of the experiment, the mixed sediments from depths of 0-2 cm
194 (surface), 2-4 cm (middle) and below 4 cm (deep) were sampled in an anaerobic glove
195 chamber. These sediment samples were used for determining total organic carbon (TOC),
196 total Fe(II), total Fe (TFe), As(III) and total As (TAs) concentrations. From these data, the
197 concentrations of Fe(III) and As(V) can be calculated by difference between total Fe(II)
198 and TFe, and between As(III) and TAs, respectively. Additionally, surficial sediments
199 from these two groups were analysed by Synchrotron Radiation microscopic X-ray
200 Fluorescence (SR-µXRF) in order to obtain the distributions of As and Fe, and by XANES
201 in order to quantify the major speciation of As and Fe bound in sediments under various
202 redox conditions. The abundances of genes associated with As metabolism in the
203 overlying water column, surficial and mid-depth sediments were also analysed for these
204 two groups, in order to ascertain the biotic mechanisms of As speciation. Moreover, to
205 further identify the different contributions of biotic and abiotic processes to As oxidation,
206 overlying water and surface sediment samples from both sterilised groups (v and vi) were

207 also taken under the same aforementioned regime and measured for total As and As(V).
208 These levels were then compared with those from the corresponding unsterilized groups.
209 All details about methods of measurement may be found in the Supporting Information.

210 ***2.4 Hydroxyl radical determination***

211 As support for the estimation of the effect of •OH on abiotic As oxidation, two batch
212 experiments were further conducted in order to discriminate whether the •OH could be
213 generated directly from bursting nanobubbles or by simple hypoxic sediment/water
214 oxygenation. Because lake water usually contains other chemical species, such as reduced
215 Fe and organic matter, which create the potential to generate •OH when exposed to O₂
216 (Liao et al., 2019a; Liao et al., 2019b; Tong et al., 2016), ultrapure water was used to
217 identify the •OH forming capacity of bursting nanobubbles directly sourced from O₂-Ze
218 materials. Specifically, samples of the prepared O₂-Ze materials (10 g) were placed into
219 degassed ultrapure water (50 mL) in a sealed amber flask. A sample of the water was then
220 immediately removed and again at 5 minutes (mins), 0.5 hour, 24 hours and 7 days, for
221 measurement by electron spin resonance (ESR), of •OH production (Tong et al., 2016),
222 details are shown in the Supporting Information. Moreover, given that O₂-Ze can support
223 abundant dissolved O₂ for oxygenation at the SWI, oxygenation of sterilized sediment
224 slurry was additionally conducted in order to qualitatively verify the production of •OH
225 and its potential for As oxidation. The experimental setup was similar to that employed
226 by Tong et al. (2016), with some modification. Briefly, sterilised sediment (30 g) and
227 sterilised lake water (150 mL) were mixed and stored in an anaerobic chamber filled with
228 nitrogen (99.999%), before stirring magnetically in an open amber flask for oxygenation

229 with dissolved O₂ from the air under dark conditions. Three treatment groups with three
230 replicates for each were performed: (I) A blank group, carried out without oxygenation;
231 (II) An O₂ group, conducted under continuous magnetic stirring of the slurry for
232 oxygenation; (III) An O₂-ethanol group, to which was added ethanol (100 mmol L⁻¹) for
233 •OH inhibition (Tong et al., 2016) accompanying the oxygenation. Slurry samples from
234 groups I and II were taken at 5 mins and 30 mins and filtered through a 0.45 µm membrane
235 prior to •OH determination by ESR (Tong et al., 2016), details are shown in the
236 Supporting Information. In addition, at 0, 3, 7, 17, 24, and 48 hours after the oxygenation,
237 slurry waters (filtered through a 0.45 µm membrane) from all groups were taken for the
238 determination of As(V) and total As, as previously detailed.

239 **2.5 Calculation**

240 The total dissolved As concentrations during the 18-day experimental period were
241 used to calculate total dissolved As fluxes across the SWI. The average flux of total
242 dissolved As was evaluated according to the mass balance equation (Tang et al., 2019),
243 and the accumulated total dissolved As flux over the 18-day experimental period was
244 calculated using MATLAB 9.4 software (MathWorks, Inc, USA) by the trapezoidal
245 method (Shi et al., 2018), based on the average flux data.

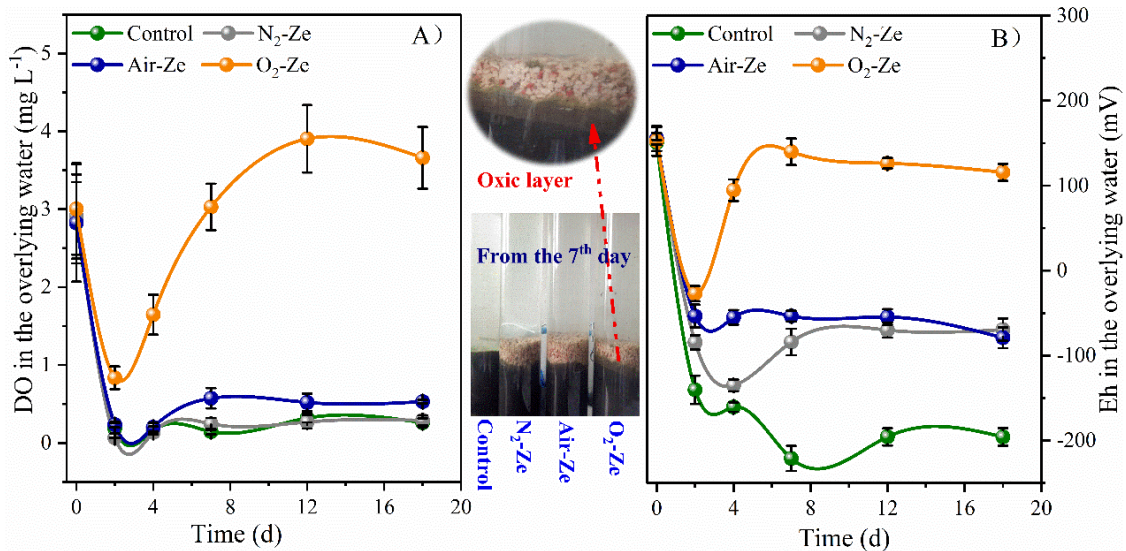
246 **3. Results and discussion**

247 **3.1 Hypoxia and As mitigation**

248 At the early stage, some labile, simple and low molecular weight carbon compounds,
249 such as alcohols, ketones, sugars or amino acids, in the settled algal biomass and surface

250 sediments are easily been biodegraded (Catalán et al., 2017; Guillemette et al., 2013),
251 leading to rapid O₂ consumption and a consequent swift decrease of the DO to 0.1 mg L⁻¹
252 and Eh to -140 mV at day 2 (Fig. 1A and 1B). This hypoxic condition was maintained
253 during the entire experimental period in the control system, which indicated the potential
254 high risk to the aquatic ecosystem if this situation had occurred in natural waters (Pan et
255 al., 2019). Sediment capping treatments, with pristine and modified zeolites, have
256 previously been used for control of internal phosphorus loading through the blocking of
257 buried anaerobic substances (e.g. reduced iron, sulfide and organic matter) (Liao et al.,
258 2017). However, the physical barrier thus formed at the SWI exhibited limited effect on
259 the remediation of hypoxia, which was indicated by the low levels of DO (<0.5 mg L⁻¹)
260 and Eh (<-50 mV) observed in the Air-Ze and N₂-Ze groups throughout the duration of
261 the experiment (Fig. 1). When O₂-Ze was applied and allowed to naturally settle to the
262 sediment surface, it was observed to partly mix with the soft algal bloom scum and
263 sediments at the SWI (O₂-Ze group). The effects of sediment capping and the sustainable
264 O₂ nanobubble release from the O₂-Ze matrix gradually compensated for the O₂
265 consumption at the SWI, resulting in the increase of DO to 3.2 mg L⁻¹ and Eh to 150 mV
266 beginning from day 7. This clear reversal of hypoxia, consistent with the previous studies
267 (Shi et al., 2018; Yu et al., 2019; Zhang et al., 2021), ~~could be~~ was directly visualized by
268 formation of a light brown-coloured oxic layer (~1 cm thickness) at the SWI ~~after seven~~
269 ~~days~~ (Fig. 1 and S2). ~~The observation of such oxic layer was well aligned with the~~
270 ~~previous findings (Zhang et al., 2018b), up to 4-cm depth of sediments turned to light~~
271 ~~brown color after the treatment by O₂-Ze for 127 days.~~ This oxic layer was formed by the
272 downward penetration of O₂, from O₂ nanobubbles, into surficial sediments, ~~which-It~~

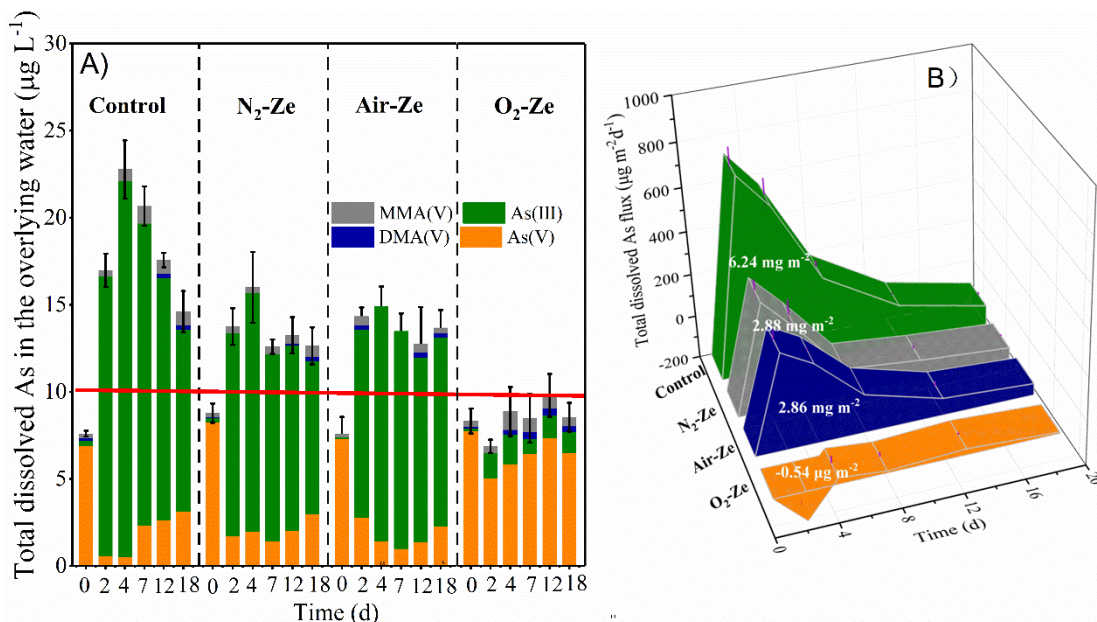
273 could, furthermore, provide an element of physical isolation, preventing the upwards
 274 diffusion of reduced substances from the anoxic layer into the water column and
 275 subsequent consumption of O₂. Therefore, the generated oxic layer was contributed
 276 contributing to an oxic condition persistent for weeks or even months (Zhang et al.,
 277 2018b).



278
 279 **Fig. 1.** Variation of DO (A) and Eh (B) in the overlying water during the experimental period

280 The simulated algal-induced hypoxia triggered an acute release of As from the
 281 sediment with total dissolved As levels reaching 23.2 $\mu\text{g L}^{-1}$ in the overlying water by day
 282 4 (Fig. 2A), and exceeding twice the WHO's recommended As safety limit (10 $\mu\text{g L}^{-1}$)
 283 (Yan et al., 2016). The total dissolved As levels in the overlying water from the Air-Ze
 284 and N₂-Ze groups were lower than those in the control group, but still remained higher
 285 than 10 $\mu\text{g L}^{-1}$ during the experiment (Fig. 2A), indicating that the physical capping by
 286 zeolites alone was not enough to control the endogenous release of As. Accompanying
 287 the hypoxic-oxic transition in the O₂-Ze group, the total dissolved As in the overlying
 288 water remained at low levels, within the range of 6.97-9.72 $\mu\text{g L}^{-1}$. Moreover, the

289 cumulative total dissolved As fluxes from sediment to water switched from positive (net
 290 As release) in the control (6.24 mg m⁻²), N₂-Ze (2.88 mg m⁻²) and Air-Ze (2.86 mg m⁻²)
 291 groups to negative (net As retention) at -0.54 μg m⁻² in the O₂-Ze group over the 18-day
 292 incubation period (Fig. 2B), demonstrating the successful change of sediment from As
 293 source to sink due to the synergistic effects of the formation of a physical barrier and the
 294 effects of O₂ nanobubbles originating from O₂-Ze. It should be noted that the oxidizing
 295 conditions at the SWI induced by O₂ nanobubbles also favoured As oxidation and
 296 methylation. The predominant As species changed from the more toxic form of As(III) in
 297 the control, N₂-Ze and Ze groups to lower toxicity forms of As(V) (65–75%) and
 298 methylated As (MMA(V) and DMA(V)) (10–15%) in the O₂-Ze group (Fig. 2A) thus
 299 further reducing the As exposure risk in the water column. Therefore, further investigation
 300 into possible mechanisms was focused on the comparison between control and O₂-Ze
 301 groups.



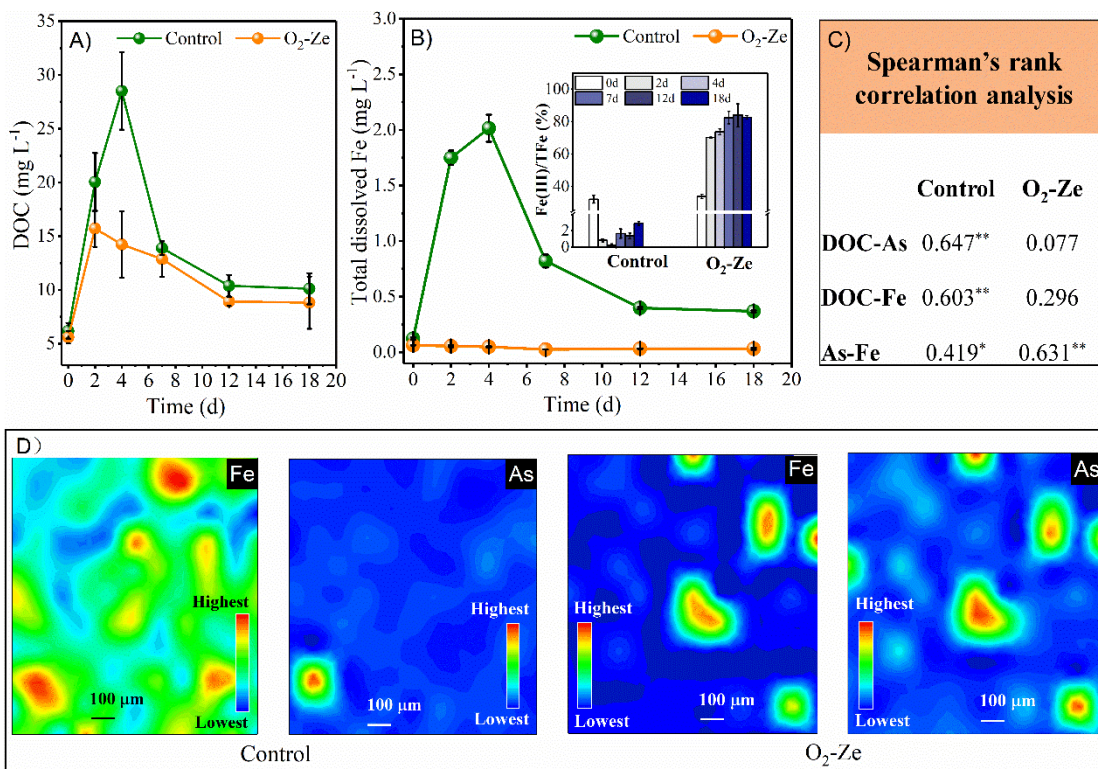
302
 303 **Fig. 2.** As dynamics during the experimental period. (A) Total dissolved As in the overlying water.
 304 The red line indicates the WHO's recommended As safety limits of 10 μg L⁻¹ in drinking water.

305 (B) Total dissolved As flux across the sediment-water interface. The data appears in a white font
306 in each bar, indicating the cumulative total dissolved As flux during the 18-day incubation period.

307 ***3.2 As mitigation controlled by geochemistry***

308 It has long been recognised that the geochemical cycle of As is strongly linked to
309 complex interactions, with co-mobilisation of some other elements, such as iron and
310 carbon, also present in the sediment (Zhu et al., 2017). In the control group,
311 accompanying the endogenous release of total dissolved As (mainly As(III)) (Fig. 2A),
312 the algal-induced hypoxia also triggered the release of large amounts of DOC (Fig. 3A)
313 and total dissolved Fe (mainly Fe(II)) (Fig. 3B) from sediments to the overlying water.
314 The dynamics of DOC and total dissolved Fe in the overlying water (Fig. 3A and 3B) and
315 sediment porewaters (Fig. S2S3) followed the same trend and ~~were~~ ~~was~~ significantly
316 correlated with concentration changes in total dissolved As in these waters ($P < 0.05$; Fig.
317 3C). This result indicated the occurrence of microbial reductive dissolution of As-bearing
318 Fe oxides with subsequent As release (Zhou et al., 2016), as triggered by the intensified
319 hypoxic conditions and the presence of an abundance of organic matter acting as an
320 electron donor at the SWI (Postma et al., 2012; Tang et al., 2019). However, during
321 treatment with O₂-Ze, a distinct decrease of DOC in the overlying water (Fig. 3A) ~~and~~
322 ~~organic carbon in surface sediments (Table S3)~~ was observed during the experimental
323 period, when compared to the control. This may attribute to the oxidation reactions
324 between DOC and dissolved O₂ or the possibly generated reactive oxidizing species
325 (ROs), such as •OH (Liao et al., 2019a; Page et al., 2013) at the SWI. The decrease of
326 reactive organic carbon (e.g. glucose) at the SWI could further have inhibited the activity
327 and growth of iron-reducing bacteria (Stuckey et al., 2016). Thus, oxidation of Fe(II)

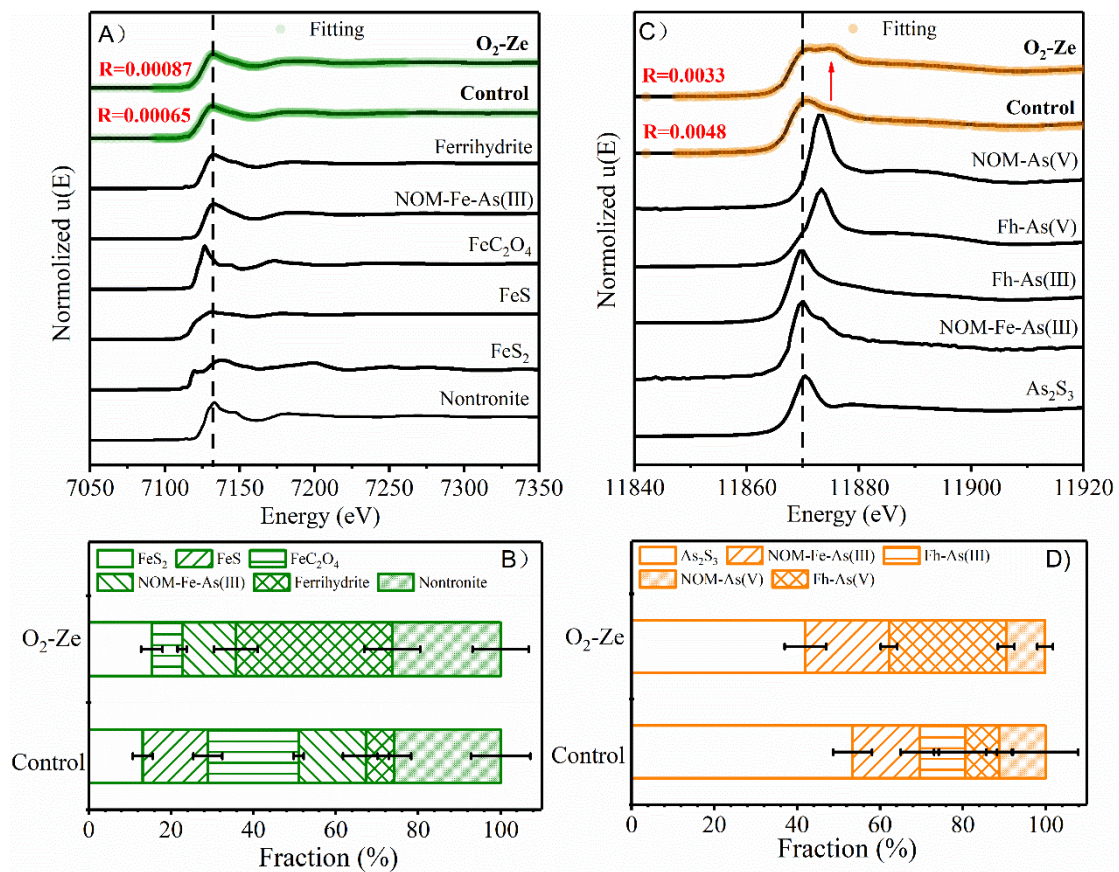
328 occurred both in the overlying water (Fig. 3B) and in surficial sediments (Fig. S4B)
 329 compared to the control, and the regenerated Fe(III)-(hydr)oxides may have been
 330 responsible for As sequestration at the oxic SWI (Tong et al., 2016; Stuckey et al., 2016),
 331 leading to the combination of reduction in total dissolved As levels in overlying water
 332 and the inhibition of As release from surface sediments. This speculation was not only
 333 supported by the highly significant correlation between total dissolved Fe and As
 334 concentrations in water profiles ($r = 0.631$, $P < 0.01$; Fig. 3C), but also by a strong
 335 correlation between the spatial distribution of As and that of Fe in surface sediments in
 336 O_2 -Ze group, observed through SR- μ XRF analysis (Fig. 3D).



337
 338 **Fig. 3.** Dynamic Geochemistry of As. (A) DOC (Dissolved Organic Carbon) in the overlying
 339 water; (B) Total dissolved Fe in the overlying water. The inset indicates the proportion of
 340 dissolved Fe (III) in the total dissolved Fe. (C) Spearman's rank correlation between total
 341 dissolved As, total dissolved Fe and DOC concentrations during the experimental period. Data
 342 for each correlation (n=24) consisted of concentrations taken from the overlying water and pore

343 water profiles. Values of $P < 0.05$ and < 0.01 indicate significant (*) and highly significant (**)
344 correlations, respectively. (D) As and Fe distributions in surficial sediments, observed by SR-
345 μ XRF analysis.

346 In order to further quantify the variation of interaction between As and Fe in surficial
347 sediments after O₂-Ze treatment, As and Fe K-edge XANES spectral analyses were
348 conducted. Panels A and C of Fig. 4 illustrate the linear combination fitting (LCF) of
349 Fe and As K-edge XANES spectra of surface sediments sampled after completion of the
350 simulation experiment, and Fig. 4B and 4D summarize the LCF results, respectively. The
351 results indicated that the oxic layer, generated on the surficial sediments after O₂-Ze
352 treatment, induced the rapid formation of ferrihydrite, the proportion of which increased
353 from 6.9% to 38.1% (Fig. 4B). Ferrihydrite is one of the amorphous Fe(III)-
354 oxyhydroxides that is ubiquitous in natural waters, especially at the redox boundaries of
355 the SWI (Hoffmann et al., 2014). It is known to be an effective As absorbent, due to
356 characteristics of nanometer-sized particles and high surface area (Tong et al., 2016;
357 Stuckey et al., 2016). In this investigation, accompanied by the distinct oxidation of As(III)
358 to As(V) in oxic surface sediments induced by O₂ nanobubbles (Fig. 4C and S34B), a
359 significant increase (from 8.1% to 28.3%) in the generated As(V) adsorbed to ferrihydrite,
360 compared with the control (Fig. 4D), was also observed. These results strongly confirmed
361 that Fe-controlled sequestration was the primary inhibition mechanism of endogenous As
362 release from As-bearing sediments under O₂-Ze treatment.



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Fig. 4. As and Fe speciation in surface sediments. (A) and (C), Fe and As K-edge XANES spectra of the raw samples and reference compounds used for linear combination fitting (LCF) analyses, respectively. Experimental data are shown as solid black lines, and symbols represent LCF of the XANES spectra. R values show the mean square misfit between the data and the fit. (B) and (D), speciation of Fe and As in surface sediments, respectively. Speciation was obtained by LCF of K-edge XANES data and the component sums were normalized to 100%. Fh-As(III)/As(V) indicates ferrihydrite adsorbed As(III) or As(V).

371

3.3 Biotic- and abiotic-mediated As speciation

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3.3.1 Biotic methylation of As

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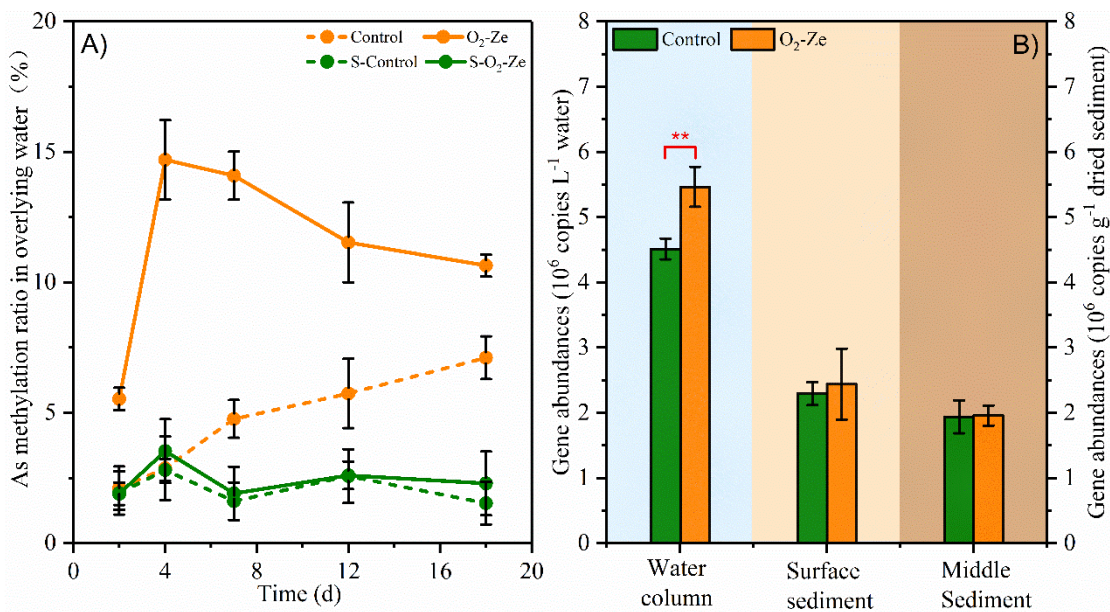
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In unsterilized systems, the As methylation ratios along the whole experimental period achieved to 2.1-7.1% and 5.5%-14.7%. A high methylation ratio of As in the overlying water in unsterilized systems was observed after of control and O₂-Ze treatment when compare to the control group, respectively (Fig. 5A). However, similar As

377 methylation ratios (1.9-3.5%) were found in the overlying water in the two sterilized
378 systems, i.e. S-O₂-Ze and S-control groups. In particular, the As methylation ratios were
379 significantly higher in the unsterilized systems compare with the corresponding sterilized
380 systems from day 7 ~~capping with O₂-Ze in the S-O₂-Ze group did not promote As~~
381 ~~methylation in the overlying water compared to the S-control group~~ (Fig. 5A). These
382 results supported the premise that methylation of As(III) to MMA(V) and DMA(V) **after**
383 **capping with O₂-Ze** was a microbially-mediated process (Maguffin et al., 2015). Known
384 as an effective method of As detoxification, As methylation has been demonstrated to be
385 catalysed by a family of As(III) S-adenosylmethionine methyltransferase enzymes
386 encoded by the *arsM* gene (Zhu et al., 2017). **There are more than 30,000 entries for *arsM***
387 **sequences in the NCBI database in members of all kingdoms, including both aerobic and**
388 **anaerobic microbes (Zhang et al., 2013; Zhu et al., 2014).** In our investigation,
389 abundances of the *arsM* gene in the water column increased significantly from 4.51×10⁶
390 to 5.46×10⁶ copies L⁻¹ after the O₂-Ze treatment (Fig. 5B), **demonstrating. It may attribute**
391 **to the** enhanced growth of As(III) methylated **aerobic microbes, under the oxic condition**
392 **at SWI induced by capping with O₂-Ze. Those methylated microbes can further which**
393 ~~may have been~~ responsible for the methylation of As(III) to DMA(V) and MMA(V) **in**
394 **the overlying water (Fig. 2A) (Zhu et al., 2014).**

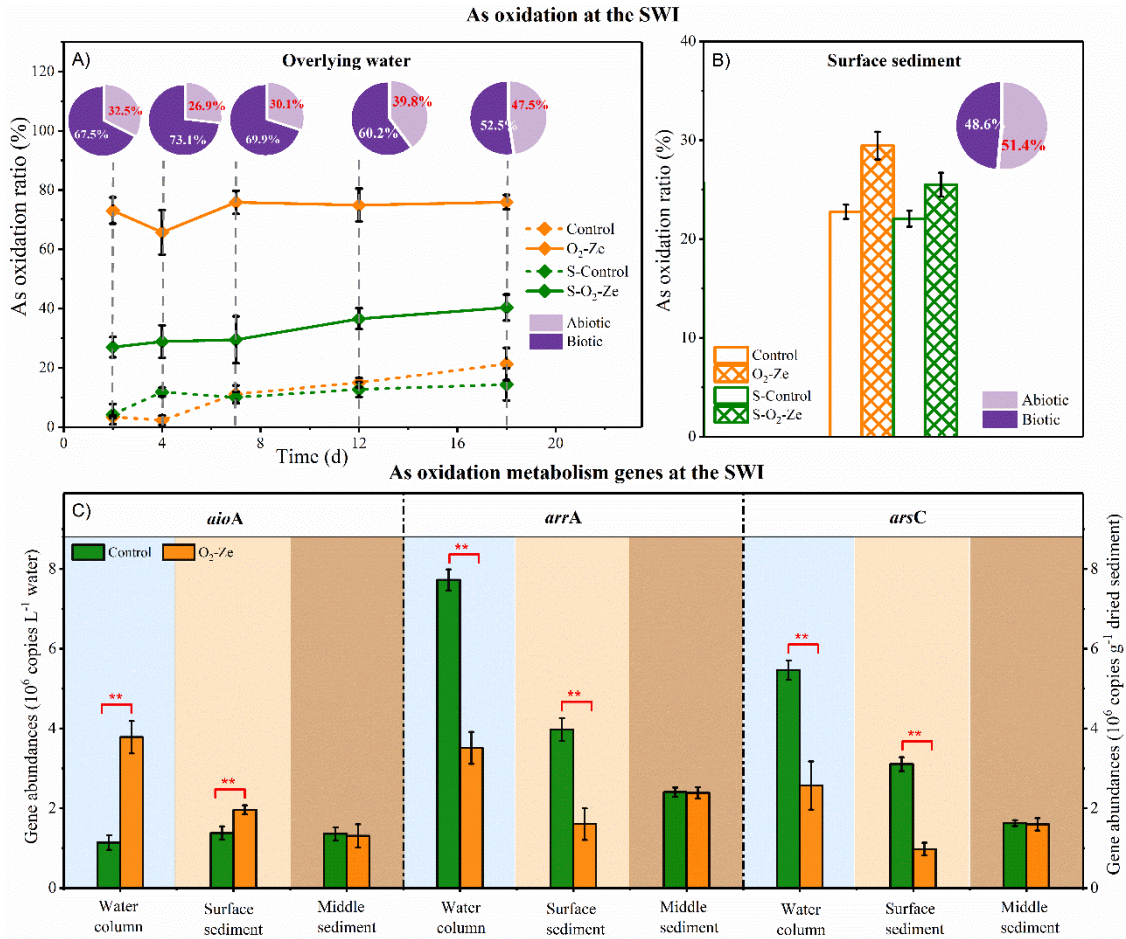


395
 396 **Fig. 5.** Methylation of As at the SWI. (A) As methylation ratio in the overlying water in the
 397 sterilized and unsterilized systems. Methylation ratio of As was calculated by the concentration
 398 ratio of methylated As (i.e. DMA and MMA) and total dissolved As in the overlying water. (B)
 399 Gene abundances of *arsM* at the SWI. The symbol “**” represents $p < 0.01$ (independent t-test).

400 3.3.2 Biotic oxidation of As

401 Significantly higher As oxidation ratios in both overlying water and surface
 402 sediments were observed after hypoxia remediation by O₂ nanobubbles compared to the
 403 control in unsterilized systems (Fig. 6A and 6B). More importantly, the As oxidation ratio
 404 in the sterilised system under treatment by O₂-Ze also showed much higher levels,
 405 compared with the sterilised control group (Fig. 6A and 6B). These results supported the
 406 hypothesis that As oxidation induced by O₂ nanobubbles technology can be categorised
 407 as both a biotic process, mediated by functional microbial activity, as well as direct abiotic
 408 chemical oxidation at the oxic SWI. Generally, As oxidation in the sterilised systems can
 409 be considered an abiotic process, but one which accounted for 26.9-51.4% of the total As
 410 oxidation in the unsterilized system, based on the mass balance calculation (Fig. 6A and
 411 6B). The remaining contribution of 52.5-73.1% could then be attributed to a biotic

412 processes. The biotic oxidation ~~through microbial metabolism~~ of As depends on the
413 ~~microbes capable of,~~ involves As(III) oxidation and As(V) reduction., ~~usually mediated by~~
414 ~~enzymes encoded by the *aioA* (oxidation), and the *arrA* and *arsC* (reduction) genes,~~
415 ~~respectively~~ As(III) oxidation is usually catalysed by As(III) oxidases encoded by *aioA*
416 genes, and As(V) reduction involves respiratory pathway mediated by *arrA* genes and
417 detoxification pathway mediated by *arsC* genes (Tang et al., 2019; Zhang et al., 2015). In
418 comparison to the control groups, capping with O₂-Ze facilitated a highly significant
419 increase in *aioA* gene abundances from 1.14×10⁶ to 3.78×10⁶ copies L⁻¹ in the water
420 column, and from 1.38 ×10⁶ to 1.96×10⁶ copies g⁻¹ in surface sediments (Fig. 6C). This
421 result indicated that the oxic SWI, induced by the introduction of O₂ nanobubbles,
422 favoured the growth of indigenous As(III) oxidizing ~~ative~~ microbes, contributing to the
423 biotic oxidation pathway of As from As(III) to As(V) (Fig. 6A and 6B) (Zhu et al., 2014).
424 Simultaneously, the corresponding abundances of *arrA* and *arsC* genes at the SWI
425 decreased significantly, implying that the oxic SWI also inhibited the microbial As(V)
426 respiratory and detoxification reduction pathways (Zhang et al., 2015).



427
 428 **Fig. 6.** Oxidation of As at the SWI. (A) As oxidation ratio in the overlying water for the sterilized
 429 and unsterilized systems. It was calculated by the concentration ratio of dissolved As(V) and total
 430 dissolved As in the overlying water. (B) As oxidation ratio in surface sediments for the sterilized
 431 and unsterilized systems. It was calculated by the concentration ratio of As(V) and TAs in the
 432 surface sediments. (C) Gene abundances of *aioA*, *arrA* and *arsC* at the SWI. The symbol“***”
 433 represents $p < 0.01$ (independent t-test). The inset pie charts in (A) and (B) indicate the calculated
 434 contributions of abiotic and biotic As oxidation.

435 3.3.3 Abiotic oxidation of As

436 When O₂ nanobubbles were released from O₂-Ze at the hypoxic SWI, they may have
 437 influenced the abiotic oxidation of As(III) by directly forming •OH during natural bubble
 438 rupture (Atkinson et al., 2019), in addition to providing sufficient dissolved O₂ for
 439 sediment/water oxygenation at the SWI (Fig. 1A). Therefore, we initially attempted to

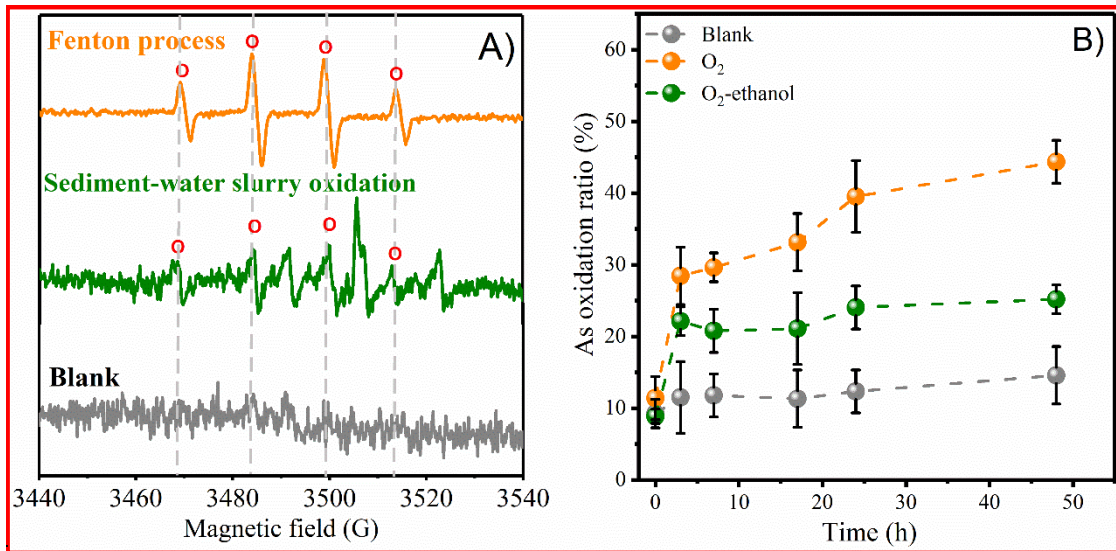
440 measure the •OH forming capacity of pure water with O₂-Ze, but no distinct ESR •OH
441 spectrum was detected (Figure S5). Given that the detection limit of ESR technique was
442 usually 10⁻⁸ mol L⁻¹ for paramagnetic species such as ROSs (Wang et al., 2011), this result
443 indicated no, or extremely low, generation of •OH by natural bubble rupture from O₂-Ze,
444 possibly due to the lack of •OH potentiating factors (e.g. UV irradiation or chemicals
445 (such as H₂O₂)) in the waters (Liu et al., 2016). Therefore, it was considered that the
446 dissolved O₂ supply was likely to be the dominant driver of abiotic oxidation of As(III) at
447 the SWI.

448 Generally, abiotic oxidation of As(III) by dissolved O₂ is considered to be quite a
449 slow reaction, with half-lives ranging from several months to a year (Smedley and
450 Kinniburgh, 2002). However, this reaction can be catalysed in natural systems by the
451 presence of redox-sensitive elements, such as Fe, Mn and C (Gorny et al., 2015). Recent
452 studies have discovered the formation of •OH through Fenton or Fenton-like processes
453 under dark conditions, once either Fe(II) or reduced organic matter in the anoxic
454 water/sediments was exposed to, and oxidized by, dissolved O₂ (Tong et al., 2016). These
455 reduced substances were abundant at the algal-induced hypoxic overlying water (Fig. 3A
456 and 3B) and surficial sediments (Fig. S34 and Table S3), but subsequently underwent
457 rapid oxidation after encountering dissolved O₂ originating from O₂ nanobubbles in the
458 O₂-Ze group, suggesting the possibility of generation of •OH at the SWI. To further verify
459 production of •OH during the oxidation of hypoxic water/sediment at the SWI and to
460 qualitatively clarify its potential for As abiotic oxidation, an oxygenation experiment was
461 conducted using a sterilized sediment-water slurry. The results indicated that when the
462 hypoxic slurry was exposed to dissolved O₂, an ESR spectrum, containing four peaks with

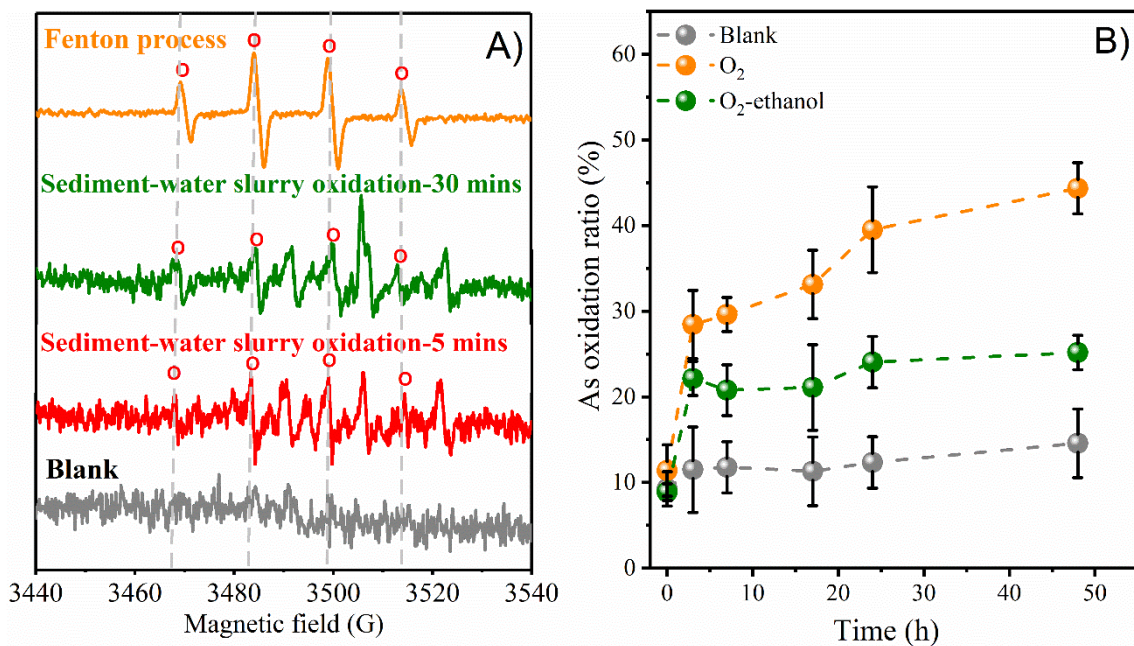
463 intensity ratios of 1:2:2:1, was observed, which could be compared to the ESR $\bullet\text{OH}$
464 spectrum produced by the traditional Fenton process (Fig. 7A), implying the generation
465 of $\bullet\text{OH}$.

466 Moreover, in ~~comparison with the blank control~~ group, ~~the cumulative As oxidation~~
467 ~~ratio was approximately 14% within 48 h (Fig. 7B). After exposure to dissolved O_2 ,~~
468 ~~significantly higher achieved~~ cumulative As oxidation ratios of nearly ~~4430%~~ ~~was~~
469 ~~achieved~~ in the water. However, ~~the oxidation corresponding~~ ratios sharply decreased to
470 ~~2410%~~ when ethanol, a $\bullet\text{OH}$ scavenger (Tong et al., 2016), was added during the
471 oxygenation process (Fig. 7B). These results confirmed that $\bullet\text{OH}$, generated from the
472 oxidation of abundant reduced substances present in the sediment/water, rather than as a
473 direct result of nanobubble rupture, would be the predominant contributor to abiotic
474 oxidation of As in the O_2 nanobubble treatment. In addition to $\bullet\text{OH}$, Fe(IV) may also play
475 a role in the residual abiotic oxidation of As(III). Several studies have reported that Fe(IV)
476 can be generated during interactions between Fe(II) and O_2 at neutral pH, leading to the
477 co-oxidation of As(III) in interfaces at natural redox environments (Ding et al., 2018; Hug
478 and Leupin, 2003). Thus, further research should focus on the in-site monitoring of $\bullet\text{OH}$
479 and Fe(IV), in order to assessing their contributions to As speciation.

480



481



482 **Fig. 7.** Abiotic-mediated As speciation at the SWI. (A) ESR evidence of •OH production from
483 oxygenation of **sterilized** sediment-water slurry. (B) As oxidation ratio in water during **sterilized**
484 sediment-water slurry oxygenation (sediment/water=30 g/150 mL).

485 3.4 As fate under the treatment of O₂-Ze

486 In As-polluted eutrophic waters, where HABs problems are increasing (Hasegawa
487 et al., 2009; Tang et al., 2019), anaerobic degradation of algae-induced organic matter at
488 the SWI stimulates a contemporaneous release of As(III), Fe(II) and reduced organic
489 matter into the water column (Fig. 2 and 3), leading to the risk of As pollution. Application

490 of O₂ -Ze at the hypoxic SWI can induce rapid remediation of hypoxia. The resultant
491 oxidizing conditions favored the growth of indigenous As(III) methylated (carrying the
492 *arsM* gene) and oxidative (through the *aioA* gene) microbes at the SWI. The former
493 microbes catalyzed the biotransformation of As(III) to MMA(V) and DMA(V), and the
494 latter catalyzed the biotic oxidation of As(III) to As(V). The As(III) oxidation process was
495 also promoted by the reaction of As(III) with •OH, during which •OH can be produced
496 by the oxidation of ferrous compounds and/or reduced organic matter in darkness. The
497 resultant As(V), MMA(V) and DMA(V) have much lower toxicity than As(III) (Tang et
498 al., 2019), which potentially reduce the risk of As exposure. Moreover, the rejuvenated
499 ferrihydrite, at the oxidizing SWI, is highly efficient not only in stripping dissolved As(V)
500 from the overlying water, but also in hampering the flux of As from sediments to the water
501 column, thus further mitigating As pollution.

502 ***3.5 Environmental implications***

503 Globally, HABs have been the cause of serious environmental disasters in
504 hypereutrophic waters over recent years (Huisman et al., 2018). The resultant
505 hypoxic/anoxic events have not only induced extensive mortality of benthos and fish, but
506 have also caused elevated exposure of pollutants, including As (Tang et al., 2019). This
507 study demonstrates that the proposed interfacial O₂ nanobubble technology has the
508 capability to reverse the hypoxic conditions, by manipulating O₂-related microbial and
509 geochemical processes to minimize the algal-induced As pollution risk in eutrophic
510 waters. The hypoxia reversal effectiveness of this technology has proven to last for
511 months in the earlier study (Zhang et al., 2018b), which may guarantee the long-term As

512 mitigation effect. Although a cost of €0.5-1.5 million per km² was roughly calculated by
513 using the interfacial O₂ nanobubble technology for sediment remediation in a previous
514 study (Zhang et al., 2021), this estimation was also based on a lab-scale experiment.
515 Therefore, optimised methodology for the material preparation and longer-term
516 experiments in large-scale systems should be further investigated, in order to move
517 towards field application of the approach. This will enable a full cost-benefit analysis of
518 the technology to be carried out. ~~However, in order to support the future real~~
519 ~~implementation, other potential factors, such as dosage of O₂-Ze and ecological impact~~
520 ~~of this technology, should be further in-situ studied.~~

521 Apart from the significant influence of O₂ nanobubbles on functional microbial
522 activity of As, we also highlight the production of •OH in darkness, at the hypoxic SWI
523 enriched with reductive substances and its dominant role in abiotic oxidation of As(III)
524 during hypoxic-oxic transitions induced by dissolved O₂ from O₂ nanobubbles. This may
525 provide insights for abiotic oxidative transformations of other redox-sensitive elements
526 and chemical species (e.g. Hg, Cr, U, S, Fe, and antibiotics) in systems facing naturally-
527 or artificially-hypoxic/anoxic-oxic transitions, for example, wetting and drying
528 phenomena due to irrigation-evaporation cycles in wetlands (Gambrell, 1994) or tidal
529 processes in estuaries and mudflats (Gonneea et al., 2014).

530 **4. Conclusions**

531 This study applied the use of interfacial O₂ nanobubble technology to combat the
532 algal-induced hypoxia that achieves good performance for As mitigation in eutrophic
533 waters. The hypoxic-oxic transition at SWI under O₂-Ze treatment manipulated microbes

534 responsible for As oxidation and methylation, leading to a conversion of As(III) to As(V)
535 and methylated As species with less toxicity. It can also stimulate the dark formation of
536 •OH, which further dominantly contributed to abiotic oxidation of As(III). The generated
537 As(V) was sequestered by Fe-(hydr)oxide under the oxic condition, making surface
538 sediments changed from being sources to acting as sinks of As. Thus it suggested that the
539 proposed technology could provide an eco-friendly and promising method for restoration
540 of algal-induced As pollution in eutrophic waters.

541 **Declaration of Competing Interest**

542 The authors declare that they have no known competing financial interests or
543 personal relationships that could have appeared to influence the work reported in this
544 paper

545 **Author contributions**

546 G.P. and Y.T. conceived the experiments, which were carried out by Y.T., who also
547 carried out data interpretation and drafted the manuscript. M.Y.Z. provided support for
548 the synchrotron radiation measurements, genetic testing, and for manuscript preparation.
549 T.L., J.Z. and M.C. contributed to the paper preparation, modification and language
550 polishing.

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