

Autism as a Disorder of Neural Information Processing: Directions for Research and Targets for Therapy ¹

Copyright ©2004 by the authors listed below. All rights reserved.

**AN ABRIDGED VERSION OF THIS PAPER APPEARS IN
MOLECULAR PSYCHIATRY, 2004, doi:10.1038/sj.mp.4001499**

Matthew K. Belmonte
Autism Research Centre
Departments of Psychiatry and
Experimental Psychology
University of Cambridge, UK

George M. Anderson
Child Study Center
Yale University
New Haven, Connecticut, USA

William T. Greenough
Beckman Institute
University of Illinois
Urbana-Champaign, Illinois, USA

Eric Courchesne
Department of Neurosciences
University of California San Diego
La Jolla, California, USA

Susan B. Powell
Department of Psychiatry
University of California San Diego
San Diego, California, USA

Elaine K. Perry
Centre Development in Clinical Brain Ageing
University of Newcastle
Newcastle, UK

Timothy M. DeLorey
Molecular Research Institute
Mountain View, California, USA

Edwin H. Cook, Jr.
Departments of Psychiatry,
Pediatrics, and Human Genetics
University of Chicago
Chicago, Illinois, USA

John L.R. Rubenstein
Department of Psychiatry
University of California San Francisco
San Francisco, California, USA

Andrea Beckel-Mitchener
Beckman Institute
University of Illinois
Urbana-Champaign, Illinois, USA

Lisa M. Boulanger
Department of Neurobiology
Harvard Medical School
Boston, Massachusetts, USA

Pat R. Levitt
Kennedy Center for Research on
Human Development
Vanderbilt University
Nashville, Tennessee, USA

Yong-hui Jiang
Department of Molecular and Human Genetics
Baylor College of Medicine
Houston, Texas, USA

Elaine Tierney
Kennedy Krieger Institute
Baltimore, Maryland, USA

¹This paper is the report of the meeting ‘Pinpointing Autism: Neurochemical Targets and Research Directions in Developmental Neurobiology’ convened by Cure Autism Now in Santa Monica, California, April 2002. In addition to those workshop participants who have directly contributed to this review, we wish to acknowledge the participation of Michael Merzenich, Eric Hollander, Steven Watkins, Maja Bucan, and Mark Geyer, whose insights have helped shape this discussion.

Abstract

The broad variation in phenotypes and severities within autism spectrum disorders suggests the involvement of multiple predisposing factors, interacting in complex ways with normal developmental courses and gradients. Identification of these factors, and the common developmental path into which they feed, is hampered by the large degrees of convergence from causal factors to altered brain development, and divergence from abnormal brain development into altered cognition and behaviour. Genetic, neurochemical, neuroimaging and behavioural findings on autism, as well as studies of normal development and of genetic syndromes that share symptoms with autism, offer hypotheses as to the nature of causal factors and their possible effects on the structure and dynamics of neural systems. Such alterations in neural properties may in turn perturb activity-dependent development, giving rise to a complex behavioural syndrome many steps removed from the root causes. Animal models based on genetic, neurochemical, neurophysiological, and behavioural manipulations offer the possibility of exploring these developmental processes in detail, as do human studies addressing endophenotypes beyond the diagnosis itself.

The past several years of research on autism and brain development have produced a confluence of findings that point the way towards understanding this complex behavioural syndrome in terms of its developmental roots. Imaging and behavioural studies on one front and genetic and biochemical studies on the other are converging on a view of autism as the result of pervasive, early developmental abnormalities affecting neural information processing. Such dysfunctions may arise through the interactions of many determinants, each of which contributes differently to the final phenotypic profile. Examination of specific phenotypic components within the broad syndrome of autism promises to identify these underlying factors.

1 Autism and Abnormal Neural Dynamics

Key to the understanding of autism is its recognition as a developmental disorder – developmental not simply in terms of taxonomy but in terms of its detailed aetiology. The approach in most studies has been to attempt to dissect autism as if it were a lesion, a missing locus or capacity within an otherwise normal, fully developed brain in which all other factors have somehow been held constant. This approach is inappropriate to the study of developmental disorders because it assumes that the disorder is a function of a localised module, rather than an emergent property of developmental interactions between many brain regions and functions [129]. Cognitively, autism has been construed as a disorder involving fundamental deficits in central coherence [91], executive function [166], and theory of mind [21] or empathising [18] – these descriptions are neither mutually exclusive nor mutually independent, and a complete explanation will encompass all of them. Anatomically, abnormalities associated

with autism have been localised in cerebellum [74, 161, 73, 105], brain stem [105, 194], frontal lobes [42, 67, 215, 11], parietal lobes [69], hippocampus [12, 203] and amygdala [12]. While significant abnormalities are present in all these cognitive capacities and anatomical regions, it remains to be seen how these characteristics are related to each other and to autism's fundamental causes. Past research has focused on the leaves of the developmental tree – the collection of surface features that are diagnostic or most apparent in autism. In order to understand autism as a developmental syndrome it is necessary to explore development at its trunk and roots. It is in this way that the two fronts of autism will be joined, unifying abnormal structure and function with the underlying genetics and neurobiology, and providing specific targets for therapeutic intervention.

Looking beyond surface features is a challenge for autism research: primary dysfunctions can be masked by the evolution of compensatory processing strategies which normalise behaviour [200], and also by the induction of activity-dependent secondary dysfunctions [66, 2] which disrupt behaviour in new ways. There is thus a high degree of fan-out (divergence) from core dysfunctions in the developing brain to cognitive and behavioural symptoms in the developed brain. The sheer number of genetic susceptibility factors and loci that have been linked to autism implies that autism's core dysfunction admits a high degree of fan-in (convergence) from primary biasing factors to core dysfunctions. The 60% concordance for strictly diagnosed autism in identical twins [88] establishes a role for environmental interactions in this fan-in just as surely as it does for genetic predispositions.

The very breadth of autism's anatomical and functional abnormalities admits the possibility that its core dysfunctions may involve some pervasive alteration of neural processing. One route to such an alteration might be via abnormally low signal-to-noise in developing neural assemblies, a condition that may be produced by abnormal neural connectivity. The high incidence of epilepsy in autism [16] is consistent with this hypothesis, and there has been no shortage of relevant neuropathological findings. The numbers of Purkinje cells, and to a lesser extent granule cells, in cerebellar cortex are abnormally low [262, 192, 24, 25] – presumably leading to disinhibition of the cerebellar deep nuclei and consequent overexcitation of thalamus and cerebral cortex. Neurons in hippocampus, amygdala, and other limbic regions are abnormally densely packed [24, 25, 187], and a Golgi analysis of the hippocampus in two autistic brains [187] has revealed an abnormally low degree of dendritic branching. A recent photographic examination of neurons in several cortical regions suggests a reduction in the size of cortical minicolumns in the autistic brain, and an increase in cell dispersion within minicolumns – characteristics that could increase the total number of minicolumns and thus the degree of connectivity between minicolumns [44, 45]. It is important to note that a decrease in signal-to-noise can arise from abnormalities of connectivity in either direction: whereas an overconnected network passes so much noise that it swamps the signal, an underconnected network passes so little signal that it becomes lost in the noise. In either case, large segments of the network are constrained to either an all-on or an all-off state, and the network's information capacity is thereby reduced [54].

2 Neurophysiologic and Neuroanatomic Effects

2.1 Neurophysiological Consequences

Such a failure to delimit activation within an abnormally connected network may be observable as hyper-arousal in response to sensory input, and decreased ability to select among competing sensory inputs [29]. Cardiovascular, neuroendocrine and neurochemical indices of arousal in novel and stressful situations are consistent with this prediction [229, 124], as are physiological and behavioural observations of the extent and intensity of perceptual processing. Physiologically, functional imaging has demonstrated heightened activity in autism in brain regions associated with stimulus-driven, sensory processing, and decreased activity in regions that normally subserve higher-order processing: these results include abnormally high activity in ventral occipital visual areas during a visual task demanding separation of local features from global context (the Embedded Figures Test), even while prefrontal and parietal activations are abnormally low [190]; heightened activity during face processing in peristriate cortex [76], inferior temporal gyrus [205], and other areas outside the fusiform ‘face area’ [174] while fusiform activity is abnormally low; heightened activity in superior temporal gyrus during inference of mental state from pictures of eyes, while prefrontal and medial temporal activations are abnormally low [22]; and decreased connectivity between extrastriate visual areas and prefrontal and temporal areas associated with inference of mental state, while prefrontal and temporal activations are again abnormally low [48].

Supporting the idea of impaired selection of perceptual inputs, evoked potential and quantitative EEG studies reveal a pattern of abnormally distributed response in autism during tasks that demand selective attention. In adults with autism, the P1 evoked potential is either abnormally augmented in response to stimuli at the attended location, or abnormally generalised to stimuli distant from the attended location [233]. During shifts of attention between hemifields, the normal, spatiotopically selective augmentation of the visual steady-state evoked potential is absent, and instead both hemispheres activate indiscriminately during shifts of attention into either hemifield [27, 29]. In children with autism, the visual N2 to novel stimuli is augmented during task performance even when these stimuli are not relevant to the task [133]. When a response is required to an auditory stimulus, the P3 in these same children with autism is abnormally generalised to occipital sites overlying visual processing areas [134]. In general, both in children and in adults, perceptual filtering in autism seems to occur in an all-or-none manner, with little specificity in selecting for the location of the stimulus, for the behavioural relevance of the stimulus, or even for the sensory modality in which the stimulus appears.

Recent fMRI evidence suggests that the autistic brain may compensate for this deficit in selecting sensory inputs by suppressing any irrelevant sensory information at a later, less efficient stage of processing [29]. In other words, in the absence of a normally functional mechanism to bias sensory processing towards attended stimuli, all stimuli receive much the same degree of sensory evaluation, and the irrelevant stimuli must then be actively discarded

in a manner that creates a processing bottleneck. Perhaps because such a compensatory mechanism cannot be as quickly reset as the normal mechanism of selective attention, performance is impaired at tasks that demand rapid reconfiguration of perceptual resources [36]. An autistic deficit in rapid shifting of attention has been observed in cases of shifts between sensory modalities [71], between spatial locations [246, 247, 234, 236, 235, 104, 27], and between object features [72, 189]. Even when the demand for rapid shifting is eliminated and people with autism produce normal behavioural output, physiological measures remain abnormal, indicating the operation of compensatory processing: frontal negativities associated with sustained attention are reduced or absent in the autistic brain [68, 53], the frontal late positive component to peripheral visual stimuli is delayed [237], and the visual P3b is highly variable [68] with a somewhat low average amplitude [162, 53, 244, 237].

2.2 Developmental Connections

These abnormalities of perceptual processing point to a developmental explanation for higher-order abnormalities, an explanation that depends crucially on the idea of a developmental chain of abnormal function: when a developing brain is confronted with an abnormal constraint on information processing, it will evolve an abnormal organisation in order to accommodate that constraint [129]. It is via such a chain of dysfunction that a core deficit such as degraded neural signal-to-noise could fan out into a welter of autistic behavioural abnormalities. From the earliest months of infancy, the flood of input generated by over-aroused, under-selective primary processing would overload nascent higher-order cognitive processes [29] – systems whose development may be being independently sabotaged by the same neuropathology that affects primary regions. Such an effect may explain the extension of sensory and motor activations in adults with autism into territories that normally subserve later-maturing cognitive functions [160]. Depending on its timing, such an error of neurocognitive development might underlie loss of cognitive skills or failure to develop skills.

Faced with this bottleneck in higher-order cognition, the developing and plastic brain would likely evolve a cognitive style that avoids reliance on high-level integrative processing and instead emphasises low-level features – a pattern typical of autistic attention and perception which Frith has termed weak central coherence [91]. Central coherence, in Frith’s definition, is the drive that causes fragmentary features to lose their identity as separate fragments once they are assembled into a single object; it is the effect of context on meaning. Weak central coherence is of especial interest as part of an explanatory framework since it predicts not only autism’s cognitive impairments in integrative tasks but also its cognitive advantages in tasks that demand attention to detail. People with autism perform better than normal on the Embedded Figures Test [206], a task that demands recognition of individual features within spatial arrangements. Unlike normal subjects, they are not aided by pre-segmentation of patterns in the Wechsler Block Design subtest, and when they make errors, these are more likely to be ones of global configuration rather than local detail [207]. In many high-functioning children with autism, lexical-to-phonological decoding

based on words' small-scale features outpaces comprehension, producing a condition known as hyperlexia [261]. Perhaps because they process faces as collections of individual features rather than as centrally coherent gestalts, children with autism are impaired at recognising unfamiliar faces [33, 136] and show a reduced or absent face-inversion effect [111, 224]. Again presumably because of intense processing of individual features, the normal levels-of-processing effect on memory is inverted in autism, with rote memory superior to normal, and memory for semantic associations impaired [228]. This piecemeal approach to perception is mirrored by a failure to generalise special skills to broader domains – *e.g.* the calendar calculator who has no interest in other forms of arithmetic calculation.

Autism's weakened central coherence may thus be a secondary property, emerging in the interaction of normal cognitive development with abnormal neural information processing – an interaction that encourages an unusual cognitive dependence on low-level processing of individual details. Indeed, the presence of low-level perceptual abnormalities in autism [181] has contributed to a recognition that many higher-level cognitive abnormalities may best be characterised as the effects of a surfeit of local processing rather than any specific impairment in global processing [180], and to an appreciation of the role of compensatory developmental changes surrounding altered perceptual functioning [159]. The possibility that some of autism's cognitive symptoms may develop as compensatory or accommodative changes raises the possibility of targeted behavioural and pharmacological interventions – one could, for example, imagine an early intervention aimed at preventing the developing brain from relying so exclusively on its intact processing of local features, in somewhat the same manner as an eye patch prevents a child at risk for amblyopia from relying on the intact eye.

Weak central coherence may in turn form the cognitive underpinning of a wide array of developmental behavioural disturbances, by impairing the use of contextual information in complex perceptual and executive tasks [93]. The executive deficit defined by this inability to use context may impact performance on theory-of-mind tasks [201], which depends on an ability to apply the context contained in one's inner model of another person's mental state. Such an executive deficit may also impede the development of joint attention and shared affect [137, 195], processes in which response to stimuli depends crucially on social context. The resulting poverty of early social cognitive experience may perturb or prevent the further activity-dependent development [66, 2] of specialised modules or capacities for such tasks as face processing, language, and complex social relations involving theory-of-mind and empathising. This failure to use context and to apply a theory of mind may result in a style of unsupervised learning founded on statistical associations rather than learning that is directed by the intentionality of others [92], and to a preference for ritualised, scripted, and repeatable interactions. Autism's surface symptoms might thus be understood as the developmental reaction of a normal human mind to abnormal neural hardware.

2.3 Anatomical Correlates

Partly because of the difficulty in recruiting people with autism who can tolerate task demands within the scanner environment, functional MRI has begun to be applied to autism only recently. Structural MRI, on the other hand, is an established technique. In adolescents and adults with autism, MRI morphometry has revealed volume deficits in cerebellum [74, 161, 73, 105], brain stem [105], and posterior corpus callosum [84]. As is the case with behavioural measures of autism, anatomical measures reveal subgroups: a minority of patients show increased rather than decreased cerebellar volume [70], and nearly half have a volume deficit in the parietal lobe [69] which is associated with a narrowing of the spatial focus of attention [233]. This anatomical heterogeneity likely reflects different convergent routes to, divergent routes from, or correlates of a common dysfunction. Studies of adult subjects say comparatively little about what this common dysfunction may be, since observations in adults take place long after the developmental pathology has run its course: although the underlying dysfunction may still exist in these adults, it can be expected to have been hidden by a forest of secondary dysfunctions and adaptive or compensatory changes.

Longitudinal studies using MRI volumetric and head circumference data point to the conclusion that autism involves transient postnatal macrencephaly [64]. Although non-standardised measurements of head circumference must be viewed with some caution, the data currently available on newborns later diagnosed with autism or PDD-NOS suggest that head size at the time of birth is normal or perhaps even slightly smaller than normal [65]. By 2 to 4 years of age, 90% of these children have MRI-based brain volumes larger than the normal average [42, 67, 215, 11], and group measures of head circumference have increased correspondingly. This abnormal brain growth is due primarily to excessive enlargement of cerebellar and cerebral white matter and cerebral grey matter [67, 108]. Even in older children, enlargement of superficial white matter tracts containing cortico-cortical fibres may persist, while internal capsule and corpus callosum are proportionately reduced [109]. Deviation from normal is so common and extreme that cerebellar and cerebral white matter volumes together with cerebellar vermis size accurately distinguish 95% of autistic toddlers and young children from normal children, and also accurately predict whether a young child with autism will have a low or high functioning developmental outcome [67]. These two-to-four-year-old autism patients show an anterior-to-posterior gradient of overgrowth, with frontal lobes being the largest [43]. This early abnormality creates a lasting legacy of increased head circumference throughout life, but, in most cases, not a lasting macrencephaly. By adulthood, then, macrencephaly is not typically present in autism, and brain volumes tend to be near normal. In sum, an abnormally rapid increase in brain volume in early life in autism is followed by abnormally slowed or prematurely arrested growth in cerebellar and cerebral regions. This early brain overgrowth takes place during a period which in normal development involves changes in synaptic density [117]. The increase in the volume of cortical grey matter matter may reflect a failure of synaptic pruning or a surfeit of synaptogenesis, and may thus be the earliest gross anatomical manifestation of a pervasive abnormality in the development of neural assemblies. Of equal potential importance, the increased volume

of local white matter projections suggests abnormality in the normal postnatal process of rapid myelination.

In characterising this underlying abnormality, it is important to note that a general deficit in development may interact with normal developmental gradients to produce an anatomically and functionally specific pattern of deficits. Myelination, for example, is not uniformly controlled across the cerebral hemispheres, but instead develops in a posterior to anterior direction. The persistence of myelination into the third decade of life in frontotemporal but not occipitoparietal fibre tracts [264, 28] indicates that oligodendrocyte regulation differs regionally. Likewise, changes in the development of neuronal populations can arise in a regionally selective fashion: in a targeted mutation of the hepatocyte growth factor signalling system, for example, deficits of GABAergic neurons appear only in frontal and parietal cortices, and only in the parvalbumin-containing subpopulation of cells [184]. In terms of cellular and molecular relationships just as in the case of cognitive ones, it may be fruitful to view autism as a product of the interaction of normal development with abnormal constraints. Observations of abnormal patterns of early brain growth suggest that at least a portion of this interaction occurs postnatally. This postnatal time frame for pathogenesis offers hope of early biological interventions, if the genetic and neurochemical determinants of abnormal growth can be identified.

3 Neurobiological Bases

3.1 Genes

It has become increasingly clear that genetic factors are very significant determinants of autism pathophysiology. The risk that another sibling will be born with autism to parents who have already had an autistic child is approximately 4.5% [130]. Given recent estimates of population prevalence for autism (which have increased, partly as a result of more complete ascertainment) [50], this statistic represents a sibling recurrence risk over twenty times the risk for the general population. Moving from siblings in general to identical twins in particular, the increased risk of concordance for autism rises from twentyfold to over two hundredfold. Such a large differential suggests not only the involvement of multiple genes, but also a multiplicative effect among these genes. For example, if there were five genes involved, a twentyfold increase could be accumulated from risk multipliers as small as 1.8 for each gene. Although some estimates of the number of interacting genetic loci in autism are as low as two to ten [172], others range from over fifteen to over a hundred [191, 186].

This multiplicative mode of inheritance carries implications for the nature of the genetic factors involved, for the experimental discriminability of each individual factor, and for the potential of interventions targeting these factors. Multiplicative effects arise from interactions between individual genes, and the susceptibility factors for autism are therefore likely to influence gene expression or to encode subtle functional alterations in proteins that participate

in regulatory networks. Multiplicative inheritance also means that the small effect of each gene by itself will be difficult to identify and to confirm; each genetic variant is likely to act as “the straw that breaks the camel’s back,” innocuous by itself but pathological in concert. Although these small effect sizes will be difficult to discern, the advantage in terms of treatment is that intervening to restore regulation to a single gene or to a small set of genes may diminish the multiplicative effect enough to yield large preventative or therapeutic effects. This situation in which the disorder may be approached via any of several therapeutic targets stands in contrast to monogenic disorders where there may not be an obvious therapeutic strategy before decades of basic work (*e.g.* Huntington’s disease).

A further consequence of the multiplicative model is that the prevalence of individual risk alleles in the general population will be high. Most if not all of these autism ‘risk’ alleles are likely to have some adaptive value, in cases where they do not lead to autism. In some cases this adaptive value would be obvious (*e.g.* enhancement of focused attention or other elements of neuropsychological function). In other cases the benefits to humanity of the existence of these alleles may be more difficult to determine, but are likely to be present.

Although review of the current status of identifying genetic risk factors for autism is beyond the scope of this manuscript, several thorough reviews are available [59, 88, 143, 141, 60]. Two relatively small regions, on 15q11-q13 centred around the GABA_Aβ₃ receptor subunit gene (*GABRB3*) and near the serotonin transporter (*SLC6A4*) on 17q11.2, are locations where more than two, but not all, studies have shown family-based association with autism. In addition to 17q, several other regions of the genome including 2q and 7q appear to be involved in the larger syndrome based on convergence from several studies. Fine mapping of these regions should be a priority in addition to generating larger samples in which two or more siblings have autism in a family (*e.g.* the Autism Genetic Resource Exchange (AGRE) [95]). In Turner syndrome, an interesting finding of increased risk for autism after deletion of the paternal, but not maternal X chromosome, suggests that a protein expressed only when the paternal X chromosome is present may be protective from autism. Such a protein might be expressed only in the developing female brain, perhaps accounting in part for the approximate 4:1 ratio of males with autism to females with autism [212]. Another possible explanation is that some of autism’s several ‘risk’ alleles are on the X chromosome.

At least four X chromosome loci have already been implicated in autism, including neuroligin 3 (*NLGN3*), neuroligin 4 (*NLGN4*), *FMR1* (see below), and *MECP2*. Although rare, a possibly functional missense mutation has been identified in *NLGN3* in an affected sibling pair, and a *de novo* nonsense mutation of *NLGN4* has been found in affected siblings and their unaffected mother [126]. Discrepancies between the affected status of the mother and offspring are somewhat difficult to interpret since, as the authors note, strong sequence conservation exists between *NLGN4* and *NLGN4Y*. Nevertheless, the association of neuroligins with autism is of great interest in light of the idea that autism’s pathophysiology may begin with abnormal neural connectivity.

The first truncating mutations in an autism-related disorder were reported when mutations in *MECP2* were identified in Rett syndrome [5]. The function of *MECP2* is to turn off

several genes whose promoters have been methylated. Therefore, in addition to *FMR1* and *UBE3A* discussed below, mutations in *MECP2* contribute to neurobiological dysfunction due to dysregulation of networks of temporally and spatially regulated genes. The recent finding of 2 *de novo* mutations of *MECP2* in a screen of 69 females with autism [41] shows that MeCP2, and perhaps its regulatory targets, are likely to be a contributing factor in autistic pathology.

In summary of the current findings, there is as yet no confirmed genetic finding in autism that has been explained in terms of its specific relation to brain development. In order to understand what genetic factors may influence the developmental processes that lead to autism, it is useful to focus on syndromes whose symptoms overlap with those of autism and which may be more tractable in terms of our current knowledge of biological cause and effect.

3.2 Proteins that Regulate Gene Expression

One such symptomatically overlapping disorder is Fragile X syndrome (FXS). FXS is the most common inherited form of mental retardation and second only to Down syndrome overall. Its neurobehavioural phenotype is characterised by some combination of mild to severe cognitive impairment, attention deficit, anxiety, seizure susceptibility, communicative disorders, and stereotypic behaviours. In addition, recent reports have suggested that 25-40% of individuals with FXS also meet diagnostic criteria for autism [15, 196, 14]. Studies of children with autism have shown a concurrence of FXS with autism of approximately 2%, with one study finding the Fragile X mutation to be the most frequent chromosomal anomaly associated with diagnosed autistic cases [13, 256]. In many instances the behavioural profiles observed in FXS seem indistinguishable from those of idiopathic autism – although some studies have noted distinct differences especially in the realm of social and communicative skills [55].

FXS is caused by the silencing of a single gene (*FMR1*) [175] that codes for the Fragile X Mental Retardation Protein (FMRP), an RNA binding protein [9]. FMRP binds to numerous mRNA cargoes, including its own mRNA, and has been hypothesised to regulate the expression of a specific set of genes in neurons. When FMRP is absent, the expression of these genes is altered and the FXS phenotype is observed. A critical feature of FXS, therefore, is that what is in one sense a single-gene disorder is more proximally the result of a disruption of complex patterns of expression of many genes. Using a novel technique, Miyashiro *et al.* have recently demonstrated that multiple classes of mRNA molecules are bound to FMRP in neurons [158]. In light of the high incidence of autistic behaviours in FXS, the data set of approximately 80 genes whose mRNA's bind to FMRP was screened to reveal 15 that map closely to suspected autism susceptibility loci. It is interesting and reasonable to speculate that polymorphic variations in these genes, and perhaps others to be determined, may be related to the variable autistic phenotype observed in many patients with FXS.

An examination of FMRP's expression and association with polyribosomal machinery in the brain leads to three important concepts. First, some of the protein synthesis that involves FMRP occurs at synapses in response to Group I metabotropic receptor activation [257]. (Since not all FMRP-expressing cells receive synaptic input, though, it is likely that not all protein synthesis involving FMRP is synaptic.) Second, FMRP itself is synthesised in response to activation of metabotropic glutamate receptor subtypes 1 and/or 5. Third, this process requires FMRP, since synaptic activity dependent protein synthesis is greatly reduced in the FraX knockout mouse [99]. An interesting *in vivo* correlate of this observation is that the proportion of dendritic spines that contain polyribosomal aggregates is reduced in the cortices of knockout mice. Therefore, one physiologically based hypothesis is that FMRP regulates the local expression of a subset of genes by controlling their translation in response to neuronal activity.

Behavioural exposure to a complex environment increases FMRP expression in rat visual cortex [121], as do physical activity in the case of somatomotor cortex [121] and whisker stimulation in the case of barrel cortex [227]. Knockout of the *FMR1* gene in mice increases dendritic spine density in visual cortex [56]. Similarly, in barrel cortex, a natural process of developmental dendritic regression fails to occur in the knockout mouse [94]. Taken together, these findings provide strong evidence for a critical role of FMRP in protein syntheses involved in activity-dependent brain development, and specifically in the patterning of neural circuitry. Anatomically, in FXS there is an overabundance of long, thin dendritic spines and a relative reduction in the number of short, thicker spines in cortical regions examined (*e.g.* [110, 56, 122]). Although the FXS patients from which these microanatomical results arise were not specifically diagnosed with autism, the symptomatic and possible aetiological overlap between FXS and autism makes these observations relevant to the understanding of autistic neuropathology. The alteration in spine morphology suggests an abnormally large proportion of immature synapses. The fact that there are more long, thin spines, and more spines overall, may reflect a failure of the synapse elimination that would normally occur during early development. A study of the hippocampal formation in idiopathic autism [187] described reduced dendritic field size in pyramidal neurons in the Ammon's horn region, a result that contrasts with the FXS findings (in other brain regions) and suggests either a failure to fully reach adult form (the authors' interpretation) or an increase in the amount of pruning in the autistic cases, the opposite of what is reported in barrel cortex [94]. Either sort of change, an abnormal abundance of dendritic connections or an abnormal lack of them, could lead to difficulty in information processing due to reduced signal-to-noise. The difficulty in differentiating these two scenarios suggests that other methods are needed to converge on the capacity of structurally-altered brain regions for information processing.

The question remains as to why 25-40% of FXS cases have symptomatic similarity to autism and why other disorders such as Rett syndrome, Prader-Willi syndrome, and Angelman syndrome may have similar symptoms as well. This neurobehavioural similarity suggests something common to the aetiology or something common to the outcome in each disorder. One example of an aetiological commonality is that of interacting genetic factors, such as the FMRP cargo genes that map to loci associated with autism. The broad range

of severities and features in the FXS phenotype may reflect interaction with polymorphisms in such genes.

Common outcomes are perhaps more challenging to understand, but the possibility exists of relatively stable or “trapping” end points that may be reached via fan-in from multiple pathways and, once entered, are very difficult to overcome. For example, one might imagine that the autistic symptoms exhibited in FXS and in idiopathic autism serve a purpose with respect to their neurobehavioural contexts by reducing, say, high levels of anxiety. Thus any attempt to overcome these symptoms serves to increase anxiety and hence elicits opposition and persistence of symptoms. It is also possible that symptoms such as chronically repeated stereotypic movements offset some tension at the neural level rather than the cognitive or behavioural level. In either case, behaviour would become reinforced into a pathological state. Neural network modellers have used the term “stable attractor” to delineate states of their networks with similar properties. Whether this conceptualisation is useful remains to be determined, but it seems clear that understanding what underlies the similarities and differences between FXS and autism may offer significant insight into the complexity of both disorders.

FMRP, of course, is not the only regulatory protein with a possible association with autism. The abnormalities in 15q11-q13 are of especial interest because of the association of a known gene in 15q11-q13 with two neurogenetic disorders, Prader-Willi syndrome (PWS) and Angelman syndrome (AS), and overlapping clinical features between AS and autism. The AS gene, *UBE3A*, is maternally expressed only in brain tissue and codes for a ubiquitin protein ligase, an enzyme that can regulate the levels of many intracellular proteins by labelling them for ubiquitin-mediated degradation. *UBE3A* abnormalities thus are capable of producing a wide range of fan-out effects.

The majority of patients with PWS or AS have a deletion of 15q11-q13, on the paternal chromosome in PWS and on the maternal chromosome in AS [127]. In contrast, a smaller fraction of PWS and AS patients have uniparental disomy for chromosome 15, with two paternal chromosomes causing AS and two maternal chromosomes causing PWS. In each case the balance between maternally and paternally derived genes is altered; however, in the case of deletion the aetiology is clearly genetic, whereas in the case of uniparental disomy it is epigenetic: in these disomic individuals there is no abnormality of the genomic sequence, yet the phenotype is indistinguishable from that caused by genetic deletion. Epigenetic disease of this sort may be more common than previously considered [26]. Similar phenomena may be operating in autism, with both genetic and epigenetic alterations contributing to autism susceptibility.

Despite the association of autism with maternal duplication of 15q11-q13, interstitial duplications in the homologous region on the paternal chromosome are associated with a relatively normal phenotype. These data strongly suggest that there is an imprinted, maternally expressed gene or genes within 15q11-q13, epigenetically-induced over-expression of which can contribute to autism. The best studied form of epigenetic modification is DNA methylation, and abnormalities of DNA methylation and/or its effects are implicated in

several genetic diseases whose clinical features overlap with those of autism, including FXS, PWS, AS, and Rett syndrome. The dynamic changes in DNA methylation that occur during early embryonic development and during germ cell development provide ample opportunity for germ-line or somatic errors in epigenetic processes, ‘epimutations’ [188]. If an abnormality of DNA methylation were proven in autism, an immediate therapeutic intervention could be explored using drugs or dietary manipulations known to alter DNA methylation, such as folate supplementation.

Success in modelling Angelman syndrome in *UBE3A* knockout mice suggests that murine 15q11-q13 manipulations may also hold promise for the development of an animal model of autism. Mice with maternal deficiency of *UBE3A* recapitulate the human AS phenotype with motor dysfunction, seizures, deficiency in contextual learning, and impairment in hippocampal long-term potentiation [128]. The role of 15q11-q13 in autism can be dissected by making mice with various locus-specific duplications [267] of the homologous PWS/AS domain. This approach may produce a valid mouse model for autism even before a specific autism-related gene within 15q11-q13 is conclusively identified.

3.3 γ -Aminobutyric Acid

The 15q11-q13 region also codes for several subunits of the GABA_A receptor, an ionotropic receptor whose hyperpolarising effect is mediated by chloride. *GABRB3*, *GABRA5* and *GABRG3* (encoding the GABA_A receptor’s β_3 , α_5 , and γ_3 subunits, respectively) all are clustered within 15q11-q13, and there is evidence for association with autism of genes in this chromosomal region [38]. The increased incidence of epilepsy in patients with autism and 15q11-q13 duplications is consistent with the involvement of GABA. Hippocampal GABA_A receptor binding in autism is abnormally low [31], as are platelet GABA levels [198]. GABAergic interneurons have a special role in establishing the computational architecture of cortical columns; double bouquet interneurons, for example, send axon bundles from layers 2 and 3 to synapse on pyramidal and spiny stellate cells in columnar arrangements [80, 170]. In addition, GABAergic neurons may be particularly vulnerable to developmental errors, since they arise from a different portion of the neural tube than the excitatory cells with which they integrate: GABAergic cells migrate into neocortex from the primordium of the basal ganglia [8].

GABAergic transmission in the mature brain plays primarily an inhibitory function. In the forebrain GABAergic neurons have a number of central roles. For instance, GABAergic projection neurons of the basal ganglia are an essential link in the extrapyramidal circuit. GABAergic projection neurons of the reticular nucleus regulate thalamic function. GABAergic local circuit neurons have essential roles in information processing in the cerebral cortex, hippocampus and olfactory bulb. Thus, reduced function of GABAergic neurons would affect most aspects of forebrain function. Primary sensory information would reach the cerebral cortex, but the intrinsic and basal ganglia circuits that process this information would be abnormal. Furthermore, several regions of the forebrain would be hyper-excitable.

These characteristics resemble our present understanding of features of the autistic mind, and reflect the fact that many people with autism have epilepsy and high levels of anxiety.

Recent genetic studies in mice suggest molecular mechanisms that could underlie this pathological physiology [199]. Mice lacking the *GAD65* gene, encoding an enzyme that makes GABA, have defective cortical plasticity that can be corrected with a GABA agonist [107]. Mutations in the *Dlx1* and *Dlx2* genes disrupt development of most telencephalic GABAergic neurons [8, 153]. It is worth noting that the human locus that has the highest LOD score for autism susceptibility (D2S2188 on chromosome 2q) maps very close to both the *GAD65* and the *Dlx1* and *Dlx2* genes [118]. Furthermore, the autism susceptibility locus (D7S477) on chromosome 7q maps within about six megabases of the *Dlx5* and *Dlx6* genes. These genes are also implicated in regulating forebrain GABAergic development [218, 219, 148] [Cobos and Rubenstein, unpublished]. Also in this region is the gene for reelin, a protein expressed in cortical GABAergic neurons [3].

One of the GABA_A receptor subunit genes in particular, *GABRB3*, is highly expressed during brain development in rats [142], and its deletion in mice disrupts GABA-related pharmacology and electrophysiology and produces behaviours reminiscent of autism [82]. In mice homozygous for a disruption of *GABRB3*, GABA_A receptor binding in neonatal and adult brain is reduced by half, with most of this reduction occurring in cerebral cortex and thalamus [125]. GABA function is impaired in these knockout mice in dorsal root ganglia, hippocampus, and the thalamic reticular nucleus, with an 80% reduction in GABA-activated chloride current in homozygotes and a 25% reduction in heterozygotes [125]. As a result, *GABRB3* knockout mice have epileptiform EEG containing high-amplitude slow and sharp waves, and develop overt seizures as they mature [125, 82]. Behaviourally, *GABRB3* knockouts are hypersensitive to thermal and tactile stimuli [239]. They are hyperactive, exhibiting stereotyped circling [125, 82]. They lack motor coordination, often falling and failing to learn a rotating-rod task [82]. Their cycles of rest and activity are lengthened [82], and EEG spectral characteristics during slow-wave sleep are abnormal [263]. There are also cognitive deficits, as measured by abnormalities of passive-avoidance conditioning and contextual fear conditioning [82]. Mothers homozygous for the *GABRB3* knockout fail to engage in normal nurturing behaviour, even towards their wild-type pups [125]. These similarities to autism suggest either a direct involvement of *GABRB3* in autism, or some GABA pathology that shares physiological or developmental effects with the *GABRB3* knockout.

Collectively these observations suggest that reduced GABA signalling is a candidate mechanism that may contribute to the causation of autism. Perhaps the co-inheritance of a combination of hypomorphic alleles that all affect GABAergic tone is a basis for some forms of autism. Efforts should therefore focus on studying known genes in known autism susceptibility loci that affect development and/or function of the GABAergic system. In particular, the exons and known regulatory elements of the *Dlx1,2,5,6* and *GAD1* genes should be sequenced in autism patients and their family members. This analysis will identify SNPs and potential function-altering mutations. Furthermore, GABAergic agonists should

be considered as potential therapeutic agents for autism.

3.4 Acetylcholine

GABAergic dysfunction can be produced not only by direct alterations in GABA systems but also by changes in neuromodulation of GABAergic neurons. Several neuromodulators may be involved in such changes, potentially with synergistic effects. Abnormal developmental changes in the population of septal neurons in autism suggest a cholinergic abnormality: septal neurons are excessive in younger and reduced in older individuals with autism [25]. Acetylcholine affects performance during sustained attention [157], possibly by shifting the dynamics of cortical networks so that afferent influence predominates over intracortical inputs [135]. In mice, knockout of the α_4 nicotinic subunit produces reduced antinociception and increased anxiety [154, 171]; abnormal pain sensitivity and anxiety both are features of the autistic syndrome. In addition, nicotinic receptors are essential for normal early synaptic development [75, 231] and thus nicotinic dysfunction may produce abnormal neural microarchitecture.

The $\alpha_4\beta_2$ nicotinic acetylcholine receptor is decreased in autopsied brains of adults with autism [169, 145]. This deficit is most pronounced in the cerebral neocortex, where the $\alpha_4\beta_2$ receptor subtype has an important regulatory effect on GABAergic neurons [4]. The decrease is selective; the other principal nicotinic receptor subtype, α_7 , is not involved. Although normal levels of choline acetyltransferase suggest that cholinergic input to the cerebral cortex and the cerebellum is intact, the level of BDNF in the basal forebrain has been observed to be three times higher than normal [145]. In addition to its role in GABAergic development outlined above, BDNF is involved in enhancing cholinergic transmission [112] and in promoting the survival of developing cholinergic neurons of the basal forebrain [248]. This BDNF abnormality at the major source of forebrain cholinergic projection may therefore be significant for cholinergic function.

As these neurochemical results come from adult brains, it remains to be seen whether nicotinic abnormality is present at early stages of autistic development. Cholinergic dysfunction may be an indirect contributor to autistic development via its influence on GABAergic neurons, a correlate of prior GABAergic dysfunction, or a direct contributor via its influence on synaptic development. *In vivo* studies at early ages, using PET and MR spectroscopy, will give further information on the role of acetylcholine in autism. Pharmacologic therapies targeting the cholinergic system may be of value; there has been one report of success using the acetylcholinesterase inhibitor donepezil to treat autistic symptoms of irritability and hyperactivity [103].

3.5 Serotonin

There has been a growing interest in the role of serotonin (5-hydroxytryptamine, 5-HT) in autism since the initial report of elevated levels of platelet 5-HT in individuals with autism

[204]. An increasing awareness of 5-HT's critical involvement in guiding neurodevelopment and in modulating sensory input and arousal has provided a convincing theoretical basis for 5-HT's role in the aetiology and pathophysiology of pervasive developmental disorders. Serotonin's especially rich innervation of limbic areas critical for emotional expression and social behaviour and the extended ontogeny and apparent plasticity of the central serotonergic system have provided additional support for its possible involvement.

As might be expected from its phylogenetically ancient role in neurodevelopment and neural transmission, and its extensive CNS projections, 5-HT has been shown to play a key role in a variety of behaviours and processes. Much of the early expression of 5-HT appears related to its role as a growth factor and regulator of neuronal development. Thus, in addition to functioning as a modulator of neural transmission, 5-HT appears to have critical effects on neurogenesis, morphogenesis, and synaptogenesis in the developing brain [260].

In adult animals, 5-HT plays important roles mediating diverse autism-relevant behaviours, including sleep, mood, arousal, aggression, impulsivity, and affiliation [151]. Reduced serotonergic function has been associated with worsened sleep, depressed mood, altered arousal, increased aggression, greater impulsivity, and reduced social behaviour. Genetic data have connected 5-HT-related genes to disorders defined by symptoms in these areas of behaviour (*e.g.* mood, social phobia, obsessive-compulsive, and anxiety disorders), and it is clear that the serotonergic system is intimately interconnected with GABAergic and glutamatergic neurons throughout the brain.

Empirical studies of 5-HT in autism include pharmacological treatment and challenge studies, biochemical/neurochemical studies of 5-HT and related species, and genetic studies of 5-HT-related genes. Drugs targeting the 5-HT transporter, including the 5-HT reuptake inhibitors fluoxetine, fluvoxamine, and clomipramine, are now widely used in autism [183]. The reuptake inhibitors appear to affect most aspects of autistic behaviour. Risperidone, another frequently used medication, also acts predominantly through a serotonergic target, the 5-HT_{2A} receptor.

Platelet hyperserotonemia has been especially well studied and is generally considered the most robust and well-replicated biological finding in autism [7, 61, 6]. Most studies have reported group mean elevations of 25-50% in platelet serotonin in persons with autism. The mechanism of the alteration and its possible relationship to brain abnormalities remain unknown. The platelet does not appear to be exposed to greater amounts of 5-HT; attention has therefore focused on handling of 5-HT within the platelet. To date, no clear alteration in the platelet has been identified although there is some suggestion that uptake may be increased in some subjects with increased platelet levels.

Genes encoding a number of the components involved in 5-HT neural transmission have been examined as possible contributors to the potentially relevant behaviours and disorders mentioned. Research in autism has focused on the influence of 5-HT transporter gene (*SLC6A4*) variants on risk to autism. Although taken together the studies do not convincingly support a role for *SLC6A4* variants in determining overall risk to autism,

investigators are now examining allelic influences on the severity of specific aspects or domains of autistic behaviour [230]. There have been several reports of variants in the transporter promoter having modifying effects in neuropsychiatry, and a recent report of strong influence on vulnerability to stress [46]. Reports of effects of 5-HT-related alleles on therapeutic response to serotonergic antidepressants and atypical neuroleptics (in mood disorders and psychosis, respectively) also tend to link 5-HT and autism [243].

At present, the areas of neuroimaging and postmortem brain research seem to offer the greatest potential for elucidating the role of serotonin in autism. The recent availability of postmortem brain tissue has opened a wide window of opportunity. Reciprocal interchange between imaging, neuropsychological, and postmortem research should be especially useful and illuminating. Work on the mechanism of the platelet hyperserotonemia may provide critically important information regarding possible central 5-HT dysfunction; the advantages of having identified a specific biochemical alteration in a delineated cell type might be best exploited by applying gene array or expression technology to this question.

3.6 Lipids

Another genetic condition that shares autistic symptoms is Smith-Lemli-Opitz syndrome (SLOS), an autosomal recessive, multiple malformation/mental retardation syndrome [213] with an estimated incidence among individuals of European ancestry of one in 40,000 to one in 60,000 births [150, 202, 39, 132], and a probable average carrier frequency of 1% [132]. Principal abnormalities include a characteristic facial appearance, microcephaly, hypotonia, postnatal growth retardation, 2-3 toe syndactyly, and hypogenitalism. In 1993, SLOS was shown to be caused by a defect of cholesterol biosynthesis at the level of the 7-dehydrocholesterol reductase [120, 226]. This defect impairs the conversion of 7-dehydrocholesterol (7-DHC) to cholesterol, causing an increased level of 7-DHC in blood and tissues, and, in most patients, decreased blood and tissue cholesterol levels. A major consequence of these biochemical abnormalities is the alteration of normal embryonic and foetal somatic development, causing postnatal abnormalities of growth, learning, language, and behaviour.

Cholesterol is necessary for the proper functioning of cells, and a defect in cholesterol synthesis impairs multiple systems in the body. Decreased cellular cholesterol levels may impair the development of cellular components and cellular function. Hedgehog proteins, which are partially responsible for embryonic patterning, require cholesterol for proper functioning. Individuals with SLOS have been found to have various CNS structural abnormalities including holoprosencephaly. Studies *in vitro* have shown that the activity of receptors for serotonin and other ligands is impaired in cholesterol-deficient environments. Low cholesterol levels impair the function of G-proteins, and thus the operation of metabotropic receptors. Low cholesterol production in individuals with SLOS may thus illuminate the decreased serotonergic functioning associated with autism.

Clinical data show that SLOS is associated with autism, and suggest that cholesterol supplementation ameliorates autistic behavioural symptoms. Of 17 subjects with SLOS for

whom the Autism Diagnostic Interview (ADI-R) [149] algorithm questions were administered, 9 (53%) met the ADI-R criteria for autism [225]. Of the 9 subjects who began cholesterol supplementation before the age of 5.0 years, 2 (22%) met the ADI-R algorithm criteria for autism at age 4.0 to 5.0 years. Of the remaining 8 subjects who began supplementation after the age of 5.0 years or had not yet started supplementation, 7 (88%) met the ADI-R algorithm criteria for autism.

Despite these behavioural similarities, the extent to which SLOS may share neurobiological mechanisms with non-SLOS autism remains unclear. Individuals with SLOS manifest some anatomical abnormalities [132] reminiscent of those often seen in autism: hypoplasia of the corpus callosum, and of the cerebellum and particularly the vermis. However, the developing macrocephaly in autism contrasts with a microcephaly in SLOS present from birth and persisting into later life. The comparison between SLOS and autism at the level of microanatomy remains an open question which could be usefully explored by quantitative neuropathological studies. Even in the absence of proven shared mechanisms of neuropathology, SLOS's comorbidity with autism does suggest several lipid-associated pathways that may be disrupted in non-SLOS autism and which may be examined for further clues. Abnormalities of the lipid membrane may affect membrane-associated proteins involved in neural or developmental signalling [37]. Individuals may also have dysfunctions of sterol or steroid metabolism, beginning *in utero* or later in life.

3.7 Immunological Signalling

In addition to genes related to specific neurotransmitter systems and to factors that directly regulate gene expression, recent studies have suggested that genes within the major histocompatibility complex (MHC, or HLA in humans), on chromosome 6p21.3, may encode predisposing factors in autism [253, 78, 252] (but see [197]). Genes in this region encode products that are required for self-nonsel self recognition by the adaptive immune system. In particular, an unusual number of people with autism may share all or part of the extended MHC haplotype B44-SC30-DR4 [253, 78, 252, 232]. Evidence also exists for a link between autism and a null allele of the C4B gene, in the class III MHC region, [254], and correspondingly low levels of C4b protein [249] which is essential in activation of the classical complement pathway. The previously mentioned possible involvement of the *UBE3A* gene also ties into MHC functioning, since the peptide fragments presented by MHC molecules can be produced by the proteolytic activity of ubiquitin ligase. Alterations in ubiquitin-related proteins or in other aspects of the peptide-generating machinery can therefore be expected to have profound effects on the intensity and specificity of MHC-dependent immune signalling. These and other genetic correlates between autism and immune-related genes could be causal, or alternatively may result from linkage disequilibrium with nearby autism-associated genes that are as yet unknown.

In support of an immune signalling aetiology, there are many reports of elevated incidence of immune disorders in the autistic population and in their first-degree relatives (reviewed in [242, 138]). A subset of autism patients displays abnormal cell-mediated immunity and

abnormal T cell populations and functions [217, 251, 255, 265, 83], reduced NK cell activity [250], and lower Th1 and higher Th2-like cytokines [100]. In addition, autism patients often exhibit abnormal humoral immune and autoantibody responses [259, 211, 210, 58]. Genetic and symptomatic links between elevated serotonin levels and autism (see above) could also contribute to immune dysfunction in autism; besides being a neurotransmitter, serotonin is an immunomodulator [266]. Interestingly, immune dysfunction and strong genetic linkage to the MHC are seen in dyslexia, another neurodevelopmental disorder that is similarly far more prevalent in boys (*e.g.* [113, 40]). It appears that abnormal immune responses may be predisposing but not sufficient to cause autism, since non-autistic first-degree relatives often share these abnormalities [57]. Rather, an environmental insult such as an infection may interact with these genetically-based predisposing factors to cause autism.

The timing of any such environmental insult remains unclear, though the predictive relationship between autistic symptoms later in development and increased brain volume in the first 2 to 12 months [65] seems to suggest a perinatal or prenatal time frame. Some maternal viral infections are known to increase the risk for schizophrenia and autism (reviewed in [52]), and in mice, experimental maternal influenza infection produces profound anatomical, motor, and other behavioural defects reminiscent of autism spectrum disorders, including hyperanxiety in novel situations and early postnatal macrocephaly [86, 208]. These infectious links may be due to viral infection of the developing brain, or infection of the pregnant mother. Infection in turn could lead to changes in expression of cytokines [168] or neuronal class I MHC [116], or to production of autoantibodies [77]. While it is unknown if autism is correlated with a change in neuronal MHC expression, maternal neuronal autoantibodies have been associated with autism [77], and neuronal autoantibodies have also been detected in autistic individuals themselves [210, 58].

Together, reports of genetic linkage to the MHC and immune comorbidity point to the potential for an autoimmune aetiology for some cases of autism. It is worth noting that viral or bacterial infections are suspected in the induction of many autoimmune diseases, perhaps due to molecular mimicry or to sequelae of persistent subclinical infection [138]. Alternatively, it may be that the paired neurological and immunological symptoms of some autism patients reflect the impact of abnormal neuronal function on the immune system, with which it is known to have extensive crosstalk (reviewed in [1]). A third possibility is that immune and neurological symptoms are parallel manifestations of a single form of genetic or metabolic disorder.

This last possibility is interesting in light of the recent discovery that class I MHC, in addition to its well-characterised role in immune function, is expressed in neurons and is critical for normal brain development and function [62, 116]. This novel finding suggests that defects in MHC function could give rise, in parallel, to both immunological and neurodevelopmental dysfunctions (reviewed in [34]). Although originally not thought to be expressed by normal, uninjured neurons, class I MHC was identified in an unbiased screen for genes involved in activity-dependent refinement of developing visual projections [62]. Here, as in many other regions of the developing brain, initial projections are large and imprecise, and patterned

neuronal activity arising in the periphery is necessary for the removal of inappropriate connections and stabilisation of appropriate ones. This process is critical to achieving mature patterns of connectivity, and in mammals occurs in early postnatal life. Strikingly, class I MHC mRNA is regulated by the spontaneous activity that drives refinement of the developing visual system [62], and is expressed by neurons precisely at the times and places of widespread activity-dependent structural and functional plasticity, both during development and in the adult [62, 116]. This positive association between MHC signalling and synaptic development is reinforced by the negative effects of MHC knockouts. Mice deficient for class I MHC signalling retain the immature pattern of visual connections, presumably due to a failure of activity-dependent refinement [116]. In addition, hippocampal synapses in these mice are not weakened in response to patterned activity, and instead are inappropriately strengthened. Both of these defects are also found in mice lacking CD3 ζ , a component of many known receptors for class I MHC [116]. These changes in plasticity are not a nonspecific effect of immune compromise, since neural plasticity is normal in even more severely immunocompromised (RAG1^{-/-}) mutant mice [116]. Thus class I MHC is required, either developmentally or acutely, for activity-dependent synaptic weakening and removal of inappropriate connections, changes that are crucial for normal brain maturation and function [116, 34]. The idea that shared molecular mechanisms may underlie both neuronal plasticity and cellular immunity is further supported by the fact that much of the MHC signal transduction machinery found in the immune system is also expressed in neurons (reviewed in [34]).

In the context of the hypothesis of abnormal connectivity in autistic brain development, the implications of such a failure of synaptic weakening and synaptic specificity are straightforward. The combination of early brain overgrowth, immunological abnormalities, and genetic linkage to the MHC is consistent with the possibility that a defect in MHC signalling is causal for a subset of people with autism. Defects in MHC signalling lead to abnormal activity-dependent refinement and plasticity in the developing visual system, the adult hippocampus, and perhaps elsewhere in the brain. Abnormal information processing later in life could stem from the excessive connectivity laid down during development as well as persistent abnormal potentiation of synaptic strength. The patterns of expression of various MHC genes are diverse, varying with neuron type, brain region, and age [116]. This chemical diversity provides a route whereby a general abnormality in MHC signalling may interact with normal development to produce specific patterns of abnormality.

To date, the connections between autism and immune genes and dysfunction have been interpreted as support for an autoimmune or infectious aetiology. It may be that class I MHC expression by populations of neurons at a given time in development confers selective vulnerability to autoimmune attack, particularly if T cells are entering the brain in large numbers, as they do during some infections. It would therefore be of interest to determine if neurons affected early in autism, such as cerebellar Purkinje cells and granule cells, express high levels of MHC around the time of onset. In addition, it would be of great value to know if MHC is expressed in abnormal patterns or levels in the developing autistic brain, although such experiments await the further refinement and validation of animal models of autism.

The studies linking immune genes and immunological functions to autism are fragmentary and a topic of heated debate. Methodological differences, small sample sizes, and a lack of appropriate control groups, as well as likely heterogeneity in aetiology and pathology, all hamper interpretation of conflicting studies. Causal heterogeneity in particular will likely frustrate attempts to determine genetic, functional, and anatomical correlates until other, perhaps biochemical, diagnostic criteria are developed to classify causal subtypes of autism. To that end, the hypothesis that MHC signalling is involved in the aetiology of a subset of cases of autism suggests candidate markers and neural substrates that may identify one such population. The current data appear to link altered immune function to abnormal neuron development through shared molecular mechanisms, and offer testable hypotheses regarding the possible relationship to autism. Understanding the role of immune molecules in brain maturation and function may ultimately lead to unexpected new strategies for the screening, treatment, and prevention of autism and other neurodevelopmental disorders.

4 Animal Models

Although several primate models of autism have been developed [106, 152], research on neurodevelopmental abnormalities and treatments for autism has had to cope with a dearth of non-primate animal models [63, 193]. Targeting relatively simple behaviours in rodent models has proven useful in the development of pharmacological targets in other neuropsychiatric diseases such as schizophrenia and affective disorders [97, 147]. Establishing the face, predictive, and construct validity of an animal model of autism may prove extremely useful in advancing understanding of autism’s neuropathology and in screening potential therapies. Approaches to animal models of neuropsychiatric disease involve modelling the aetiology or pathophysiology of the disease (*e.g.* producing mice with cerebellar pathology) or modelling particular behavioural attributes (*e.g.* studying animals with abnormal social interactions or deficient sensorimotor gating).

If people with autism are hyper-responsive to sensory stimuli, then existing methods of examining sensory responsiveness may be useful in defining endophenotypes and developing animal models of autism. One approach to measuring sensory processing arises in the context of sensorimotor gating. Prepulse inhibition (PPI) of the startle response is a cross-species measure of the normal decrement in startle when a barely detectable prestimulus immediately precedes (30-500ms) a startling stimulus [123, 98, 79]. PPI appears to reflect the activation of a ubiquitous centrally mediated behavioural ‘gating’ process, and has been widely applied in studies of information processing and cognition in animals and humans. In particular, the inhibitory processes activated by the weak ‘prepulse’ and the resulting decrement in startle amplitude have been used as an operational measure of sensorimotor gating. Studying PPI has enabled a focus on well-defined measures of information processing having substantial homology among different species and, in the case of schizophrenia, significant predictive validity for treatments [221]. As reviewed elsewhere, schizophrenia-like deficits in PPI have been mimicked in animals using both pharmacological [96] and anatomical manipulations

[222].

Deficits in PPI have been observed in schizophrenia [35], obsessive-compulsive disorder [220], Huntington’s disease [223], nocturnal enuresis and Attention Deficit Disorder [164], and Tourette syndrome [47]. Although an earlier study of autism showed no abnormality in PPI [165], a more recent examination, parametrised in terms of prestimulus amplitude and prestimulus-to-stimulus interval, revealed a selective impairment in the inhibition of startle response at high prestimulus amplitude and longer interstimulus intervals [155]. While normal subjects showed the typical pattern of increased prepulse inhibition at higher prestimulus intensities (12 dB versus 4 dB) and longer interstimulus intervals (120ms versus 30ms), people with Asperger syndrome did not. This failure to make use of extra processing time to rapidly modify response bias is reminiscent of results on slowed shifting of attention in autism (*e.g.* [236]), and the absence of an effect of prepulse amplitude suggests abnormal sensory responsiveness – a possibility supported by skin conductance findings of abnormally high tonic arousal [216] and abnormally high phasic response to stimuli [241] in autism. Similar findings of high arousal [30] and high responsiveness [156] in FXS support the idea that these abnormal sensory phenomena may be markers of abnormal neural development. More comprehensive studies assessing startle plasticity in adults and children with autism are warranted, and are currently underway (William Perry, personal communication).

Sensorimotor gating abnormalities similar to those in human neuropsychiatric conditions have been produced in rats by a variety of neurodevelopmental manipulations. Models include alterations of the intrauterine environment (*e.g.* viral insult, neurotoxin exposure, prenatal maternal stress), birth complications [240], postnatal maternal and/or social deprivation [85, 258], and neonatal ventral hippocampal lesions [146]. The developmental model most extensively studied for its effects on PPI has been post-weaning social isolation, a manipulation that induces profound abnormalities in behaviour, drug responses, and neurochemistry [185]. Neurodevelopmental animal models like those developed for schizophrenia are now being brought to bear on autism. Some of these autism models are based on prenatal exposure to teratogens such as Borna disease virus [182] or valproic acid [119]. Others derive from findings of abnormal cerebellar anatomy in people with autism, and include both mutant animals with specific cerebellar malformations [49, 139] and animals with surgical lesions of the midline cerebellum [32]. Several such cerebellar models have begun to find application in the study of sensorimotor gating and other behaviours relevant to autism: deficits in PPI have been observed in heterozygous reeler mice [238] and in mice homozygous for the cerebellar deficient folia (*cdf*) mutation [167].

A further model is based on the hypothesis of low signal-to-noise as a basis of autistic brain development: if this is the case, developmental errors similar to those in autism ought to be able to be induced by manipulating signal-to-noise directly, independently of any neurochemical antecedents. A straightforward way of implementing such a direct manipulation is to supply the developing brain with consistently noisy sensory inputs. In rats reared in continuous 70dB acoustic noise, neurons in primary auditory cortex retain into adulthood the immature pattern of broad, high-frequency tuning curves and imprecise

tonotopy typical of the earliest stage of auditory development [51]. This physiological finding of broad tuning in the auditory system seems very much akin to the psychophysical finding of abnormal bandwidth of auditory filters in people with high-functioning autism [180]. In addition to retarding the emergence of normal activity-dependent patterns of neural tuning, developmental exposure to noise prolongs the critical period during which any input, regardless of behavioural context, is significant for activity-dependent plasticity [51]. A delay of this transition from unconditional plasticity to context-dependent plasticity may underlie the ease with which people with autism learn arbitrary associations, and the difficulty with which they incorporate behavioural context into such learning. Furthermore, a delayed and abnormal closing of this critical period may be the trigger for the onset of autistic regression during the second or third year of life.

Inevitably animal models will play an important role in future research on the causes and cures for autism. Creative translational brain-behaviour designs will be needed, wherein tests demonstrating specific brain-behaviour relationships in children with autism are formatted for animal model studies. For instance, a recent human study showed that when allowed to explore freely an open area containing novel objects, children with autism spent less time exploring the novel information than normal children, and that the more abnormal the exploration behaviour, the smaller the cerebellar vermis [173]. Such a study design can be easily formatted for testing animal models of candidate genetic or non-genetic causes of autism, and in fact several candidate models already exist. The GS guinea pig, a mutant with malformation of cerebellar vermal lobules VI and VII, shows decreased exploratory behaviour [49], as do Purkinje cell degeneration mutant mice [140]. Specific loss of Purkinje cells in cerebellar vermal lobules VI and VII in L1CAM knockout mice is associated with decreased exploration and stereotyped circling in a novel environment [131, 90]. Rats with cerebellar lesions show deficits in long-term habituation of the acoustic startle response [144] and increased spontaneous motor activity and perseverative behaviour [32]. As the definition of human behavioural endophenotypes in autism proceeds, measurable, homologous behaviours can be quantified in animal models. Prepulse inhibition and other forms of startle modulation (*e.g.* fear potentiated startle) may be useful in constructing such animal behavioural homologues.

5 Research Imperatives

5.1 Phenotypes within and beyond the Diagnosis

To those who treat patients with autism spectrum disorders, or who work with them as researchers or caregivers, it can often seem as if there are as many kinds of autism as there are people with autism. Since the diagnosis of autism rests on broad and entirely behavioural criteria, studies whose inclusion criteria follow the diagnostic criteria are liable to recruit subject groups that are heterogeneous in terms of ultimate causes. Reducing this heterogeneity demands a more detailed taxonomy, extending not only within the diagnosis but also outside it. The accepted wisdom has been to restrict studies to only the most

severe cases – for example those patients who satisfy both ADI-R and ADOS criteria and whose clinical diagnoses exclude Asperger syndrome and PDD-NOS. Although this strategy certainly reduces heterogeneity, it puts blinders on efforts to identify behavioural endophenotypes. In contrast, broadening studies beyond the strict diagnosis of autism holds a great deal of promise for identifying which components of the autistic syndrome are genetically transmitted, and how these components interact. The early results of such work reveal familial patterns of repetitive behaviours, impaired nonverbal communication, and impaired or delayed development of phrase speech, but not social interaction or verbal communication [209].

Autism may well occur when several root factors combine to produce a core dysfunction – a dysfunction which we have suggested may involve neural signal-to-noise – to a degree that brings on a grand change in developmental course and produces autism’s many symptoms. Information on the abnormal events surrounding this critical developmental event becomes much more valuable when it can be contrasted with information on what happens when the event is avoided. Such a contrast can be obtained by studying siblings and other family members of patients with autism, people who presumably share some of the genetic susceptibility factors but in whom those factors have not become magnified into the full syndrome of autism. A wealth of behavioural data suggests that such factors are operative in first-degree relatives and do produce subclinical abnormalities. Siblings of autistic children show a cognitive profile reminiscent of that of autism itself, with superior spatial and verbal span, poor set-shifting, poor planning, and poor verbal fluency [115]. Parents of autistic children perform in the superior range on the Embedded Figures Test [20], visual search tasks [163], and other tasks that require processing of parts and details [102], but are below normal performance at inferring mental state from facial expression [20], a task that demands configural and integrative processing. In addition, parents share difficulties in pragmatic language tasks [87], impairments in some tasks that tap executive function [114], and autistic personality characteristics such as rigidity, aloofness, and anxiety [179, 178]. Many first-degree relatives show a disparity between performance IQ and verbal IQ, with the performance score being lower [89, 177, 87] due to impairment on Picture Arrangement and Picture Completion, both of which demand attention to global, contextual information. These subtle characteristics of the broader autism phenotype are visible in the cognitive skills that relatives tend to develop: occupations in engineering, which demands meticulous attention to detail, are over-represented in the fathers and grandfathers of people with autism [23] and, conversely, the incidence of autism is increased in the families of engineers, mathematicians, and physicists [19]. Siblings are at greater risk than the general population for a wide array of cognitive and affective disorders [176, 10], and there is some epidaemiological evidence relating autism to familial history of affective disorder [81].

If the cognitive profile in siblings is, as it seems, reflective of that in autism, then the same may be true at the levels of neuroanatomy, neurophysiology, and neurochemistry. Studies of first-degree relatives at these levels of analysis can expose primary factors closer to the roots of autistic brain development, unobscured by so many of the secondary developmental sequelae that may overshadow or mask such factors in autism probands. In particular, there

is a wealth of electrophysiological and morphometric findings in autism whose value would be greatly augmented if the same experiments and measures were applied in autism relatives, and the results compared. Do relatives manifest to any degree the anatomically over-generalised activation found in autism probands during attentional tasks? Is focused spatial attention associated with abnormally modulated mid-latency visual evoked potentials as it is in autism probands? Are there any changes in the amplitude or latency of frontal negativities, or in the late positive component? Do relatives show any abnormality in cerebellar volume, or in cross-sectional area of the corpus callosum? What about head circumference and early overgrowth of the frontal lobes? Many of these comparative questions can be answered quite easily, and should be addressed. Where differences exist, they may be subtle, and large groups of subjects may be required in order to achieve sufficient statistical power. As autism susceptibility genes begin to be identified, these non-autistic relatives will be of the utmost value in defining the behavioural and physiological phenotypes with which single susceptibility factors are associated.

The potential for large-scale secondary dysfunctions to mask more primary abnormalities has implications for studies within autism probands, too. A great deal of research attention has been devoted to autism's most apparent and most debilitating symptoms, those that make up the diagnostic triad of impaired social interaction, impaired communication, and restricted and repetitive interests and behaviours. Even physiological studies have addressed complex social capacities such as theory-of-mind [22] and emotion perception [76]. Observations at all levels of behavioural and physiological complexity are useful in reverse-engineering autistic development. However, in concentrating exclusively on the most diagnostic features of autism, such work may be overlooking cognitive and perceptual features at low levels of processing that are closer to autism's core dysfunction. There is a great need for further exploration of the neurophysiology associated with low-level perceptual abnormalities. It is quite remarkable and difficult to fathom that we currently have more functional imaging data about how the autistic brain processes a face or a theory of mind than we do about the way it processes, say, location, colour, orientation, or spatial frequency; at what level of processing do the perceptual and cognitive abnormalities begin? It is also important to recognise that absence of behavioural performance or functional activation does not necessarily imply incapacity of the corresponding brain subsystems. Rather, an apparent lack may be due to failure to engage an intact capacity. The proper stimuli or experimental paradigm can bring out such hidden abilities.

The finding of early brain overgrowth illustrates the importance of looking for primary abnormalities at the youngest possible ages, before all of the secondary changes have occurred. Although autism currently cannot be decisively diagnosed until the age of 5 years, its sibling recurrence risk of 4.5% opens the possibility of conducting the study in advance of the diagnosis. After the collection of data from all at-risk siblings, observations of siblings who do not meet full criteria at age 5 can be excluded from the autism sample. These observations of siblings who turn out not to have autism are not wasted, since, as noted previously, the sibling group is an informative contrast both to the autism group and to unrelated control subjects.

In addition to these more traditional, cognitively based elements of behaviour, attention must be paid to even lower-level aspects of the phenotype. Parent reports are rife with anecdotal observations of low-level dysfunctions. Motor abnormalities, in particular, may be one of autism's earliest signs, appearing within the first year of life [17]. Abnormal gait is common in autism [245, 101], and motor clumsiness seems particularly prominent in Asperger syndrome [214]. Other oft-reported abnormalities (*e.g.* gastrointestinal problems, blink rate, pupillary dilation), and the unproven interventions that have been designed around some of them, may yet contain a kernel of truth and should be subjected to controlled study.

5.2 A Synthetic Approach

Most experimental observations of autism yield correlations rather than definite causal relationships, and these correlations often exist in a vacuum: it is possible to implicate a chromosomal locus without knowing what genes it contains, a gene without knowing the function of its protein product, a protein without knowing its physiologic effect, and so forth. Connecting the many stages from genetics and biochemistry to brain structure and function and behaviour demands studies that bridge these levels of analysis.

Postmortem studies and *in vivo* MRS studies aimed at elucidating the neuronal and molecular bases of early brain growth abnormalities in cerebellar and cerebral white matter and cerebral grey matter are needed. Studies can and should be designed to demonstrate explicitly relationships between such postmortem microscopic findings and *in vivo* MRI-based macroscopic effects. Also, detailed knowledge of brain growth defects may help guide the effort to create more developmentally realistic animal models of the disorder. Techniques such as MRI morphometry and event-related potentials make it possible to associate specific sites of brain abnormality with specific types of neurobehavioural deficits. Event-related potentials and other brain-behaviour correlation studies have elucidated relationships between anatomical sites of abnormality and deficits in visual selective attention, shifting attention, motor activation, motor learning, visuospatial exploration, and orienting attention. An important hypothesis for future research is that the age of onset, rate, and duration of aberrant brain growth are related to the severity and age of onset of autistic behaviours.

Like the individuals whom it seeks to understand, the field of autism research often falls victim to a sort of weak central coherence. Its challenge is to unify a complex set of local observations and details into coherent explanations of the autistic syndrome. Isolation of specific endophenotypes and examination of contributing genes will aid in uncovering the neurobiological roots of autism's core dysfunctions, while targeting low-level processes and examining early stages of development will highlight the steps through which those core dysfunctions unfold into autism's complex behavioural syndrome.

References

- [1] Ader R, Felton DL, Cohen N (eds). *Psychoneuroimmunology* 2/e. Academic Press: San Diego, 1991.
- [2] Akshoomoff N, Pierce K, Courchesne E. The neurobiological basis of autism from a developmental perspective. *Dev Psychopathol* 2002; **14**:613-634.
- [3] Alcantara S, Ruiz M, D’Arcangelo G, Ezan F, de Lecea L, Curran T, Sotelo C, Soriano E. Regional and cellular patterns of reelin mRNA expression in the forebrain of the developing and adult mouse. *J Neurosci* 1998; **18**:7779-7799.
- [4] Alkondon M, Albuquerque EX. Nicotinic acetylcholine receptor α_7 and $\alpha_4\beta_2$ subtypes differentially control GABAergic input to CA1 neurons in rat hippocampus. *J Neurophysiol* 2001; **86**:3043-3055.
- [5] Amir RE, Van den Veyver IB, Wan M, Tran CQ, Francke U, Zoghbi HY. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. *Nat Genet* 1999; **23**:185-188.
- [6] Anderson GM, Gutknecht L, Cohen DJ, Brailly-Tabard S, Cohen JH, Ferrari P, Roubertoux PL, Tordjman S. Serotonin transporter promoter variants in autism: functional effects and relationship to platelet hyperserotonemia. *Mol Psychiatry* 2002; **7**:831-836.
- [7] Anderson GM, Horne WC, Chatterjee D, Cohen DJ. The hyperserotonemia of autism. *Ann NY Acad Sci* 1990; **600**:331-340.
- [8] Anderson S, Qiu M, Bulfone A, Eisenstat D, Meneses JJ, Pedersen RA, Rubenstein, JLR. Mutations of the homeobox genes *Dlx-1* and *Dlx-2* disrupt the striatal subventricular zone and differentiation of late-born striatal cells. *Neuron* 1997; **19**:27-37.
- [9] Ashley CT Jr, Wilkinson KD, Reines D, Warren ST. FMR1 protein: conserved RNP family domains and selective RNA binding. *Science* 1993; **262**:563-566.
- [10] August GJ, Stewart MA, Tsai L. The incidence of cognitive disabilities in the siblings of autistic children. *Br J Psychiatry* 1981; **138**:416-422.
- [11] Aylward EH, Minshew NJ, Field K, Sparks BF, Singh N. Effects of age on brain volume and head circumference in autism. *Neurology* 2002; **59**:175-183.
- [12] Aylward EH, Minshew NJ, Goldstein G, Honeycutt NA, Augustine AM, Yates KO, Barta PE, Pearlson GD. MRI volumes of amygdala and hippocampus in non-mentally retarded autistic adolescents and adults. *Neurology* 1999; **53**:2145-2150.
- [13] Bailey A, Bolton P, Butler L, Le Couteur A, Murphy M, Scott S, Webb T, Rutter M. Prevalence of the fragile X anomaly amongst autistic twins and singletons. *J Child Psychol Psychiatry* 1993; **34**:673-688.
- [14] Bailey DB Jr, Hatton DD, Skinner M, Mesibov G. Autistic behavior, FMR1 protein, and developmental trajectories in young males with fragile X syndrome. *J Autism Dev Disord* 2001; **31**:165-174.
- [15] Bailey DB Jr, Mesibov GB, Hatton DD, Clark RD, Roberts JE, Mayhew L. Autistic behavior in young boys with fragile X syndrome. *J Autism Dev Disord* 1998; **28**:499-508.
- [16] Ballaban-Gil K, Tuchman R. Epilepsy and epileptiform EEG: association with autism and language disorders. *Ment Retard Dev Disabil Res Rev* 2000; **6**:300-308.
- [17] Baranek GT. Autism during infancy: a retrospective video analysis of sensory-motor and social behaviors at 9-12 months of age. *J Autism Dev Disord* 1999; **29**:213-224.
- [18] Baron-Cohen S. The extreme male brain theory of autism. *Trends Cogn Sci* 2002; **6**:248-254.
- [19] Baron-Cohen S, Bolton P, Wheelwright S, Scchall V, Short L, Mead G, Smith A. Autism occurs more often in families of physicists, engineers, and mathematicians. *Autism* 1998; **2**:296-301.
- [20] Baron-Cohen S, Hammer J. Parents of children with Asperger syndrome: what is the cognitive phenotype? *J Cogn Neurosci* 1997; **9**:548-554.
- [21] Baron-Cohen S, Leslie AM, Frith U. Does the autistic child have a “theory of mind”? *Cognition* 1985; **21**:37-46.
- [22] Baron-Cohen S, Ring HA, Wheelwright S, Bullmore ET, Brammer MJ, Simmons A, Williams SCR. Social intelligence in the normal and autistic brain: an fMRI study. *Eur J Neurosci* 1999; **11**:1891-1898.
- [23] Baron-Cohen S, Wheelwright S, Stott C, Bolton P, Goodyer I. Is there a link between engineering and autism? *Autism* 1997; **1**:153-163.
- [24] Bauman ML, Kemper TL. Histoanatomic observations of the brain in early infantile autism. *Neurology* 1985; **35**:866-874.
- [25] Bauman ML, Kemper TL. Neuroanatomic observations of the brain in autism. In: Bauman ML, Kemper TL. (eds) *The Neurobiology of Autism*. Johns Hopkins University Press: Baltimore, 1994,

- pp 119-145.
- [26] Beaudet AL. Is medical genetics neglecting epigenetics? *Genet Med* 2002; **4**:399-402.
 - [27] Belmonte MK. Abnormal attention in autism shown by steady-state visual evoked potentials. *Autism* 2000; **4**:269-285.
 - [28] Belmonte MK, Egaas B, Townsend J, Courchesne E. NMR intensity of corpus callosum differs with age but not with diagnosis of autism. *Neuroreport* 1995; **6**:1253-1256.
 - [29] Belmonte MK, Yurgelun-Todd DA. Functional Anatomy of Impaired Selective Attention and Compensatory Processing in Autism. *Cogn Brain Res* 2003; **17**:651-664.
 - [30] Belser RC, Sudhalter V. Arousal difficulties in males with fragile X syndrome: a preliminary report. *Dev Brain Dysfunct* 1995; **8**:270-279.
 - [31] Blatt GJ, Fitzgerald CM, Guptill JT, Booker AB, Kemper TL, Bauman ML. Density and distribution of hippocampal neurotransmitter receptors in autism: an autoradiographic study. *J Autism Dev Disord* 2001; **31**:537-543.
 - [32] Bobee S, Mariette E, Tremblay-Leveau H, Caston J. Effects of early midline cerebellar lesion on cognitive and emotional functions in the rat. *Behav Brain Res* 2000; **112**:107-117.
 - [33] Boucher J, Lewis V. Unfamiliar face recognition in relatively able autistic children. *J Child Psychol Psychiatry* 1992; **33**:843-859.
 - [34] Boulanger LM, Huh GS, Shatz CJ. Neuronal plasticity and cellular immunity: shared molecular mechanisms. *Curr Opin Neurobiol* 2001; **11**:568-578.
 - [35] Braff DL, Geyer MA, Swerdlow NR. Human studies of prepulse inhibition of startle: normal subjects, patient groups, and pharmacological studies. *Psychopharm* 2001; **156**:234-258.
 - [36] Burack JA, Enns JT, Stauder JEA, Motttron L, Randolph B. Attention and autism: behavioral and electrophysiological evidence. In: Cohen DJ, Volkmar FR. (eds) *Handbook of Autism and Pervasive Developmental Disorders 2/e*. Wiley: New York, 1997, pp 226-247.
 - [37] Burger K, Gimpl G, Fahrenholz F. Regulation of receptor function by cholesterol. *Cell Mol Life Sci* 2000; **57**:1577-1592.
 - [38] Buxbaum JD, Silverman JM, Smith CJ, Greenberg DA, Kilifarski M, Reichert J, Cook EH Jr, Fang Y, Song CY, Vitale R. Association between a GABRB3 polymorphism and autism. *Mol Psychiatry* 2002; **7**:311-316.
 - [39] Bzdúch V, Behulova D, Skodova. Incidence of Smith-Lemli-Opitz syndrome in Slovakia. *J Am Med Genet* 2000; **90**:260.
 - [40] Cardon LR, Smith SD, Fulker DW, Kimberling WJ, Pennington BF, DeFries JC. Quantitative trait locus for reading disability on chromosome 6. *Science* 1994; **266**:276-279.
 - [41] Carney RM, Wolpert CM, Ravan SA, Shahbazian M, Ashley-Koch A, Cuccaro ML, Vance JM, Pericak-Vance MA. Identification of MeCP2 mutations in a series of females with autistic disorder. *Pediatr Neurol* 2003; **28**:205-211.
 - [42] Carper RA, Courchesne E. Inverse correlation between frontal lobe and cerebellum sizes in children with autism. *Brain* 2000; **123**:836-844.
 - [43] Carper RA, Moses P, Tigue ZD, Courchesne E. Cerebral lobes in autism: early hyperplasia and abnormal age effects. *NeuroImage* 2002; **16**:1038-1051.
 - [44] Casanova MF, Buxhoeveden DP, Switala AE, Roy E. Minicolumnar pathology in autism. *Neurology* 2002; **58**:428-432.
 - [45] Casanova MF, Buxhoeveden DP, Switala AE, Roy E. Asperger's syndrome and cortical neuropathology. *J Child Neurol* 2002; **17**:142-145.
 - [46] Caspi A, Sugden K, Moffitt TE, Taylor A, Craig IW, Harrington H, McClay J, Mill J, Martin J, Braithwaite A, Poulton R. Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science* 2003; **301**:386-389.
 - [47] Castellanos FX, Fine EJ, Kaysen DL, Kozuch PL, Hamburger SD, Rapoport JL, Hallett M. Sensorimotor gating in boys with Tourette's syndrome and ADHD: preliminary results. *Biol Psychiatry* 1996; **39**:33-41.
 - [48] Castelli F, Frith C, Happé F, Frith U. Autism, Asperger syndrome and brain mechanisms for the attribution of mental states to animated shapes. *Brain* 2002; **125**:1839-1849.
 - [49] Caston J, Yon E, Mellier D, Godfrey HP, Delhaye-Bouchard N, Mariani J. An animal model of autism: behavioural studies in the GS guinea pig. *Eur J Neurosci* 1998; **10**:2677-2684.
 - [50] Chakrabarti S, Fombonne E. Pervasive developmental disorders in preschool children. *JAMA* 2001; **285**:3093-3099.
 - [51] Chang EF, Merzenich MM. Environmental noise retards auditory cortical development. *Science* 2003;

- 300:498-502.**
- [52] Ciaranello AL, Ciaranello RD. The neurobiology of infantile autism. *Annu Rev Neurosci* 1995; **18**:101-128.
 - [53] Ciesielski KT, Courchesne E, Elmasian R. Effects of focused selective attention tasks on event-related potentials in autistic and normal individuals. *EEG Clin Neurophysiol* 1990; **75**:207-220.
 - [54] Cohen I. An artificial neural network analogue of learning in autism. *Biol Psychiatry* 1994; **36**:5-20.
 - [55] Cohen, IL, Vietze PM, Sudhalter V, Jenkins EC, Brown WT. Effects of age and communication level on eye contact in fragile X males and non-fragile X autistic males. *Am J Med Genet* 1991; **38**:498-502.
 - [56] Comery TA, Harris JB, Willems PJ, Oostra BA, Irwin SA, Weiler IJ, Greenough WT. Abnormal dendritic spines in fragile X knockout mice: maturation and pruning deficits. *Proc Natl Acad Sci USA* 1997; **94**:5401-5404.
 - [57] Comi AM, Zimmerman AW, Frye VH, Law PA, Peeden JN. Familial clustering of autoimmune disorders and evaluation of medical risk factors in autism. *J Child Neurol* 1999; **14**:388-394.
 - [58] Connolly AM, Chez MG, Pestronk A, Arnold ST, Mehta S, Deuel RK. Serum autoantibodies to brain in Landau-Kleffner variant, autism, and other neurological disorders. *J Pediatrics* 1999; **134**:607-613.
 - [59] Cook EH Jr. Genetics of autism. *Child Adolesc Psychiatr Clin N Am* 2001; **10**:333-350.
 - [60] Cook EH Jr. Genetics of Autism. In: Cooper D. (ed) *Nature Encyclopedia of the Human Genome*. Nature Publishing Group: London, 2003.
 - [61] Cook EH Jr, Leventhal BL. The serotonin system in autism. *Curr Opin Pediatr* 1996; **8**:348-354.
 - [62] Corriveau RA, Huh GS, Shatz CJ. Regulation of class I MHC gene expression in the developing and mature CNS by neural activity. *Neuron* 1998; **21**:505-520.
 - [63] Courchesne E. Brainstem, cerebellar and limbic neuroanatomical abnormalities in autism. *Curr Opin Neurobiol* 1997; **7**:269-278.
 - [64] Courchesne E. Abnormal early brain development in autism. *Mol Psychiatry* 2002; **7**:S21-23.
 - [65] Courchesne E, Carper R, Akshoomoff N. Evidence of brain overgrowth in the first year of life in autism. *JAMA* 2003; **290**:337-344.
 - [66] Courchesne E, Chisum H, Townsend J. Neural activity-dependent brain changes in development: implications for psychopathology. *Dev Psychopathol* 1994; **6**:697-722.
 - [67] Courchesne E, Karns C, Davis HR, Ziccardi R, Carper RA, Tigue ZD, Chisum HJ, Moses P, Pierce K, Lord C, Lincoln AJ, Pizzo S, Schreibman L, Haas RH, Akshoomoff NA, Courchesne RY. Unusual brain growth patterns in early life in patients with autistic disorder: an MRI study. *Neurology* 2001; **57**:245-254.
 - [68] Courchesne E, Lincoln AJ, Yeung-Courchesne R, Elmasian R, Grillon C. Pathophysiologic findings in nonretarded autism and receptive developmental language disorder. *J Autism Dev Disord* 1989; **19**:1-17.
 - [69] Courchesne E, Press G, Yeung-Courchesne R. Parietal lobe abnormalities detected with MR in patients with infantile autism. *Am J Roentgenol* 1993; **160**:387-393.
 - [70] Courchesne E, Saitoh O, Yeung-Courchesne R, Press GA, Lincoln AJ, Haas RH, Schreibman L. Abnormality of cerebellar vermal lobules VI and VII in patients with infantile autism: identification of hypoplastic and hyperplastic subgroups with MR imaging. *Am J Roentgenol* 1994; **162**:123-130.
 - [71] Courchesne E, Townsend J, Akshoomoff N, Saitoh O, Yeung-Courchesne R, Lincoln AJ, Haas R, Schreibman L, Lau L. Impairment in shifting attention in autistic and cerebellar patients. *Behav Neurosci* 1994; **108**:848-865.
 - [72] Courchesne E, Townsend J, Akshoomoff N, Yeung-Courchesne R, Lincoln AJ, Press G, Murakami J, James H, Saitoh O, Egaas B, Haas R, Schreibman L. A new finding: impairment in shifting attention in autistic and cerebellar patients. In: Broman SH, Grafman J. (eds) *Atypical Cognitive Deficits in Developmental Disorders: Implications for Brain Function*. Lawrence Erlbaum: Hillsdale, New Jersey, 1994, pp 101-137.
 - [73] Courchesne E, Townsend J, Saitoh O. The brain in infantile autism: posterior fossa structures are abnormal. *Neurology* 1994; **44**:214-223.
 - [74] Courchesne E, Yeung-Courchesne R, Press G, Hesselink JR, Jernigan TL. Hypoplasia of cerebellar vermal lobules VI and VII in autism. *N Engl J Med* 1988; **318**:1349-1354.
 - [75] Court JA, Martin-Ruiz C, Graham A, Perry E. Nicotinic receptors in human brain: topography and pathology. *J Chem Neuroanat* 2000; **20**:281-298.
 - [76] Critchley HD, Daly EM, Bullmore ET, Williams SCR, Van Amelsvoort T, Robertson DM, Rowe A, Phillips M, McAlonan G, Howlin P, Murphy DGM. The functional neuroanatomy of social behaviour: changes in cerebral blood flow when people with autistic disorder process facial expressions. *Brain*

- 2000; **123**:2203-2212.
- [77] Dalton P, Deacon R, Blamire A, Pike M, McKinlay I, Stein J, Styles P, Vincent A. Maternal neuronal antibodies associated with autism and a language disorder. *Ann Neurol* 2003; **53**:533-537.
- [78] Daniels WW, Warren RP, Odell JD, Maciulis A, Burger RA, Warren WL, Torres AR. Increased frequency of the extended or ancestral haplotype B44-SC30-DR4 in autism. *Neuropsychobiology* 1995; **32**:120-123.
- [79] Davis M. The mammalian startle response. In: Eaton RC (ed) Neural mechanisms of startle behavior. Plenum Press: New York, 1984, pp 287-342.
- [80] DeFelipe J, Hendry SH, Hashikawa T, Molinari M, Jones EG. A microcolumnar structure of monkey cerebral cortex revealed by immunocytochemical studies of double bouquet cell axons. *Neurosci* 1990; **23**:622-631.
- [81] DeLong R, Nohria C. Psychiatric family history and neurological disease in autistic spectrum disorders. *Dev Med Child Neurol* 1994; **36**:441-448.
- [82] DeLorey TM, Handforth A, Anagnostaras SG, Homanics GE, Minassian BA, Asatourian A, Fanselow MS, Delgado-Escueta A, Ellison GD, Olsen RW. Mice lacking the β_3 subunit of the GABA_A receptor have the epilepsy phenotype and many of the behavioral characteristics of Angelman syndrome. *J Neurosci* 1998; **18**:8505-8514.
- [83] Denney DR, Frei BW, Gaffney GR. Lymphocyte subsets and interleukin-2 receptors in autistic children. *J Autism Dev Disord* 1996; **26**:87-97.
- [84] Egaas B, Courchesne E, Saitoh O. Reduced size of corpus callosum in autism. *Arch Neurol* 1995; **52**:794-801.
- [85] Ellenbroek BA, van den Kroonenberg PT, Cools AR. The effects of an early stressful life event on sensorimotor gating in adult rats. *Schiz Res* 1998; **30**:251-260.
- [86] Fatemi SH, Earle J, Kanodia R, Kist D, Emamian ES, Patterson PH, Shi L, Sidwell R. Prenatal viral infection leads to pyramidal cell atrophy and macrocephaly in adulthood: implications for genesis of autism and schizophrenia. *Cell Mol Neurobiol* 2002; **22**:25-33.
- [87] Folstein SE, Gilman SE, Landa R, Hein J, Santangelo SL, Piven J, Lainhart J, Wzorek M. Predictors of cognitive test patterns in autism families. *J Child Psychol Psychiatry* 1999; **40**:1117-1128.
- [88] Folstein SE, Rosen-Sheidley B. Genetics of autism: complex aetiology for a heterogeneous disorder. *Nat Rev Genet* 2001; **2**:943-955.
- [89] Fombonne E, Bolton P, Prior J, Jordan H, Rutter M. A family study of autism: cognitive patterns and levels in parents and siblings. *J Child Psychol Psychiatry* 1997; **38**:667-683.
- [90] Fransen E, D'Hooge R, Van Camp G, Verhoye M, Sijbers J, Reyniers E, Soriano P, Kamiguchi H, Willemsen R, Koekkoek SK, De Zeeuw CI, De Deyn PP, Van der Linden A, Lemmon V, Kooy RF, Willems PJ. L1 knockout mice show dilated ventricles, vermis hypoplasia and impaired exploration patterns. *Hum Mol Genet* 1998; **7**:999-1009.
- [91] Frith U, *Autism: Explaining the Enigma*. Blackwell: Oxford, 1989.
- [92] Frith U, Mind blindness and the brain in autism. *Neuron* 2001; **32**:969-979.
- [93] Frith U, Happé F, Autism: beyond "theory of mind". *Cognition* 1994; **50**:115-132.
- [94] Galvez R, Gopal AR, Greenough WT. Somatosensory cortical barrel dendritic abnormalities in a mouse model of the fragile X mental retardation syndrome. *Brain Res* 2003; **971**:83-89.
- [95] Geschwind DH, Sowiński J, Lord C, Iversen P, Shestack J, Jones P, Ducat L, Spence SJ. The autism genetic resource exchange: a resource for the study of autism and related neuropsychiatric conditions. *Am J Hum Genet* 2001; **69**:463-466.
- [96] Geyer MA, Krebs-Thomson K, Braff DL, Swerdlow NR. Pharmacological studies of prepulse inhibition models of sensorimotor gating deficits in schizophrenia: a decade in review. *Psychopharm* 2001; **156**:117-154.
- [97] Geyer MA, Markou A. Animal models of psychiatric disorders. In: Bloom FE, Kupfer D (eds) Psychopharmacology: The Fourth Generation of Progress, Raven Press: New York, 1995, pp 787-798.
- [98] Graham F. The more or less startling effects of weak prestimulation. *Psychophysiol* 1975; **12**:238-248.
- [99] Greenough WT, Klintsova AY, Irwin SA, Galvez R, Bates KE, Weiler IJ. Synaptic regulation of protein synthesis and the fragile X protein. *Proc Natl Acad Sci USA* 2001; **98**:7101-7106.
- [100] Gupta S, Aggarwal S, Roshanravan B, Lee T. Th1- and Th2-like cytokines in CD4+ and CD8+ T cells in autism. *J Neuroimmunol* 1998; **85**:106-109.
- [101] Hallett M, Lebedowska MK, Thomas SL, Stanhope SJ, Denckla MB, Rumsey J. Locomotion of autistic adults. *Arch Neurol* 1993; **50**:1304-1308.

- [102] Happé F, Briskman J, Frith U. Exploring the cognitive phenotype of autism: weak “central coherence” in parents and siblings of children with autism: I. Experimental tests. *J Child Psychol Psychiatry* 2001; **42**:299-307.
- [103] Hardan AY, Handen BL. A retrospective open trial of adjunctive donepezil in children and adolescents with autistic disorder. *J Child Adolesc Psychopharmacol* 2002; **12**:237-241.
- [104] Harris NS, Courchesne E, Townsend J, Carper RA, Lord C. Neuroanatomic contributions to slowed orienting of attention in children with autism. *Cogn Brain Res* 1999; **8**:61-71.
- [105] Hashimoto T, Tayama M, Murakawa K, Yoshimoto T, Miyazaki M, Harada M, Kuroda Y. Development of the brainstem and cerebellum in autistic patients. *J Autism Devel Disord* 1995; **25**:1-18.
- [106] Hemby SE, Sanchez MM, Winslow JT. Functional genomics approaches to a primate model of autistic symptomology. *J Autism Dev Disord* 2001; **31**:551-555.
- [107] Hensch TK, Fagiolini M, Mataga N, Stryker MP, Baekkeskov S, Kash, SF. Local GABA circuit control of experience-dependent plasticity in the developing visual cortex. *Science* 1998; **282**:1504-1508.
- [108] Herbert MR, Ziegler DA, Deutsch CK, O’Brien LM, Lange N, Bakardjiev A, Hodgson J, Adrien KT, Steele S, Makris N, Kennedy DN, Harris GJ, Caviness VS Jr. Dissociations of cerebral cortex, subcortical and cerebral white matter volumes in autistic boys. *Brain* 2003; **126**:1182-1192.
- [109] Herbert MR, Ziegler DA, Makris N, Sanders HA, Normandin JJ, Deutsch C, Kennedy DN, Caviness VS. White matter increases in autism are largely in superficial radiate regions. International Meeting for Autism Research, Orlando, Florida, 2002.
- [110] Hinton VJ, Brown WT, Wisniewski K, Rudelli RD. Analysis of neocortex in three males with the fragile X syndrome. *Am J Med Genet* 1991; **41**:289-294.
- [111] Hobson RP, Ouston J, Lee A. What’s in a face? The case of autism. *Br J Psychol* 1988; **79**:441-453.
- [112] Hohmann CF, Berger-Sweeney J. Cholinergic regulation of cortical development and plasticity. New twists to an old story. *Perspect Dev Neurobiol* 1998; **5**:401-425.
- [113] Hugdahl K, Synnevag B, Satz P. Immune and autoimmune diseases in dyslexic children. *Neuropsychologia* 1990; **28**:673-679.
- [114] Hughes C, Leboyer M, Bouvard M. Executive function in parents of children with autism. *Psychol Med* 1997; **27**:209-220.
- [115] Hughes C, Plumet MH, Leboyer M. Towards a cognitive phenotype for autism: increased prevalence of executive dysfunction and superior spatial span amongst siblings of children with autism. *J Child Psychol Psychiatry* 1999; **40**:705-718.
- [116] Huh GS, Boulanger LM, Du H, Riquelme PA, Brotz TM, Shatz CJ. Functional requirement for class I MHC in CNS development and plasticity. *Science* 2000; **290**:2155-2159.
- [117] Huttenlocher PR, Dabholkar AS. Regional differences in synaptogenesis in human cerebral cortex. *J Comp Neurol* 1997; **387**:167-178.
- [118] IMGSAC. A genomewide screen for autism: strong evidence for linkage to chromosomes 2q, 7q, and 16p. *Am J Hum Genet* 2001; **69**:570-581.
- [119] Ingram JL, Peckham SM, Tisdale B, Rodier PM. Prenatal exposure of rats to valproic acid reproduces the cerebellar anomalies associated with autism. *Neurotox and Teratology* 2000; **22**:319-324.
- [120] Irons M, Elias ER, Salen G, Tint GS, Batta AK. Defective cholesterol biosynthesis in Smith-Lemli-Opitz syndrome. *Lancet* 1993; **341**:1414.
- [121] Irwin SA, Swain RA, Christmon CA, Chakravarti A, Weiler IJ, Greenough WT. Evidence for altered Fragile-X mental retardation protein expression in response to behavioral stimulation. *Neurobiol Learn Mem* 2000; **73**:87-93.
- [122] Irwin SA, Patel B, Idupulapati M, Harris JB, Crisostomo RA, Larsen BP, Kooy F, Willems PJ, Cras P, Kozlowski PB, Swain RA, Weiler IJ, Greenough WT. Abnormal dendritic spine characteristics in the temporal and visual cortices of patients with fragile-X syndrome: a quantitative examination. *Am J Med Genet* 2001; **98**:161-167.
- [123] Ison JR, McAdam DW, Hammond GR. Latency and amplitude changes in the acoustic startle reflex produced by variations in auditory prestimulation. *Physiol Behav* 1973; **10**:1035-1039.
- [124] Hirstein W, Iversen P, Ramachandran VS. Autonomic responses of autistic children to people and objects. *Proc R Soc Lond B* 2001; **268**:1883-1888.
- [125] Homanics GE, DeLorey TM, Firestone LL, Quinlan JJ, Handforth A, Harrison NL, Krasowski MD, Rick CEM, Korpi ER, Mäkelä R, Brilliant MH, Hagiwara N, Ferguson C, Snyder K, Olsen RW. Mice devoid of γ -aminobutyrate type A receptor β_3 subunit have epilepsy, cleft palate, and hypersensitive behavior. *Proc Natl Acad Sci USA* 1997; **94**:4143-4148.

- [126] Jamain S, Quach H, Betancur C, Råstam M, Colineaux C, Gillberg IC, Soderstrom H, Giros B, Leboyer M, Gillberg C, Bourgeron T, Paris Autism Research International Sibpair Study. Mutations of the X-linked genes encoding neuroligins NLGN3 and NLGN4 are associated with autism. *Nat Genet* 2003; **34**:27-29.
- [127] Jiang YH, Tsai TF, Bressler J, Beaudet AL. Imprinting in Angelman and Prader-Willi syndromes. *Curr Opin Genet Dev* 1998; **8**:334-342.
- [128] Jiang YH, Armstrong D, Albrecht U, Atkins CM, Noebels JL, Eichele G, Sweatt JD, Beaudet AL. Mutation of the Angelman ubiquitin ligase in mice causes increased cytoplasmic p53 and deficits of contextual learning and long-term potentiation. *Neuron* 1998; **21**:799-811.
- [129] Johnson MH, Halit H, Grice SJ, Karmiloff-Smith A. Neuroimaging of typical and atypical development: a perspective from multiple levels of analysis. *Dev Psychopathol* 2002; **14**:521-536.
- [130] Jorde L, Hasstedt S, Ritvo E, Mason-Brothers A, Freeman B, Pingree C, McMahon W, Peterson B, Jenson W, Moll A. Complex segregation analysis of autism. *Am J Hum Genet* 1991; **49**:932-938.
- [131] Kamiguchi H, Hlavín ML, Lemmon V. Role of L1 in neural development: what the knockouts tell us. *Mol Cell Neurosci* 1998; **12**:48-55.
- [132] Kelley RI, Hennekam RCH. Smith-Lemli-Opitz Syndrome. In: Scriver CR, Beaudet AL, Sly WS, Valle D. (eds) *The Metabolic and Molecular Basis of Inherited Disease 8/e*. McGraw Hill: New York, 2000.
- [133] Kemner C, Verbaten MN, Cuperus JM, Camfferman G, van Engeland H. Visual and somatosensory event-related brain potentials in autistic children and three different control groups. *EEG Clin Neurophysiol* 1994; **92**:225-237.
- [134] Kemner C, Verbaten MN, Cuperus JM, Camfferman G, van Engeland H. Auditory event-related brain potentials in autistic children and three different control groups. *Biol Psychiatry* 1995; **38**:150-165.
- [135] Kimura F. Cholinergic modulation of cortical function: a hypothetical role in shifting the dynamics in cortical network. *Neurosci Res* 2000; **38**:19-26.
- [136] Klin A, Sparrow SS, de Bildt A, Cicchetti DV, Cohen DJ, Volkmar FR. A normed study of face recognition in autism and related disorders. *J Autism Dev Disord* 1999; **29**:499-508.
- [137] Klin A, Volkmar FR, Sparrow SS. Autistic social dysfunction: some limitations of the theory of mind hypothesis. *J Child Psychol Psychiatry* 1992; **33**:861-876.
- [138] Krause I, He X-S, Gershwin ME, Shoenfeld Y. Immune factors in autism: a critical review. *J Autism Devel Disord* 2002; **32**:337-345.
- [139] Kuwamura M, Morikawa T, Yamate J, Kato K, Kotani T, Sakuma S. Glial pathology in development of cerebellar dysplasia in the hereditary cerebellar vermis defect (cvd) rat. *Acta Neuropathol* 2000; **99**:305-309.
- [140] Lalonde R, Manseau M, Boetz MI. Exploration and habituation in Purkinje cell degeneration mutant mice. *Brain Res* 1989; **479**:201-203.
- [141] Lamb JA, Parr JR, Bailey AJ, Monaco AP. Autism: in search of susceptibility genes. *Neuromolecular Med* 2002; **2**:11-28.
- [142] Laurie DJ, Wisden W, Seeburg PH. The distribution of thirteen GABA_A receptor subunit mRNAs in the brain. III. Embryonic and postnatal development. *J Neurosci* 1992; **12**:4151-4172.
- [143] Lauritsen M, Ewald H. The genetics of autism. *Acta Psychiatr Scand* 2001; **103**:411-427.
- [144] Leaton RN, Supple WF Jr. Medial cerebellum and long-term habituation of acoustic startle in rats. *Behav Neurosci* 1991; **105**:804-816.
- [145] Lee M, Martin-Ruiz C, Graham A, Court J, Jaros E, Perry R, Iversen P, Bauman M, Perry E. Nicotinic receptor abnormalities in the cerebellar cortex in autism. *Brain* 2002; **125**:1483-1495.
- [146] Lipska BK, Swerdlow NR, Geyer MA, Jaskiw GE, Braff DL, Weinberger DR. Neonatal excitotoxic hippocampal damage in rats causes post-pubertal changes in prepulse inhibition of startle and its disruption by apomorphine. *Psychopharmacol* 1995; **122**:35-43.
- [147] Lipska BK, Weinberger DR. To model a psychiatric disorder in animals: schizophrenia as a reality test. *Neuropsychopharm* 2000; **23**:223-239.
- [148] Long JE, Garel S, Depew M, Tobet S, Rubenstein JLR. DLX5 regulates development of peripheral and central components of the olfactory system. *J Neurosci* 2003; **23**:568-578.
- [149] Lord C, Rutter M, Le Couteur A. Autism Diagnostic Interview-Revised: a revised version of a diagnostic interview for caregivers of individuals with possible pervasive developmental disorders. *J Autism Dev Disord* 1994; **24**:659-685.
- [150] Lowry RB, Yong SL. Borderline normal intelligence in the Smith-Lemli-Opitz (RSH) syndrome. *Am J Med Genet* 1980; **5**:137-143.

- [151] Lucki I. The spectrum of behaviors influenced by serotonin. *Biol Psychiatry* 1998; **44**:151-162.
- [152] Machado CJ, Bachevalier J. Non-human primate models of childhood psychopathology: the promise and the limitations. *J Child Psychol Psychiatry* 2003; **44**:64-87.
- [153] Marin O, Rubenstein JLR. A long, remarkable journey: cellular and molecular mechanisms of tangential migration in the telencephalon. *Nat Neurosci Rev* 2001; **2**:780-790.
- [154] Marubio LM, del Mar Arroyo-Jimenez M, Cordero-Erausquin M, Lena C, Le Novere N, de Kerchove d'Exaerde A, Huchet M, Damaï MI, Changeux JP. Reduced antinociception in mice lacking neuronal nicotinic receptor subunits. *Nature* 1999; **398**:805-810.
- [155] McAlonan GM, Daly E, Kumari V, Critchley HD, van Amelsvoort T, Suckling J, Simmons A, Sigmundsson T, Greenwood K, Russell A, Schmitz N, Happe F, Howlin P, Murphy DG. Brain anatomy and sensorimotor gating in Asperger's syndrome. *Brain* 2002; **125**:1594-1606.
- [156] Miller LJ, McIntosh DN, McGrath J, Shyu V, Lampe M, Taylor AK, Tassone F, Neitzel K, Stackhouse T, Hagerman RJ. Electrodermal responses to sensory stimuli in individuals with fragile X syndrome: a preliminary report. *Am J Med Genet* 1999; **83**:268-279.
- [157] Mirza NR, Stolerman IP. The role of nicotinic and muscarinic acetylcholine receptors in attention. *Psychopharmacology* 2000; **148**:243-250.
- [158] Miyashiro KY, Beckel-Mitchener A, Purk TP, Becker KG, Barret T, Liu L, Carbonetto S, Weiler IJ, Greenough WT, Eberwine J. RNA cargoes associating with FMRP reveal deficits in cellular functioning in FMR1 null mice. *Neuron* 2003; **37**:417-431.
- [159] Motttron L, Burack JA. Enhanced perceptual functioning in the development of autism. In: Burack JA, Charman T, Yirmiya N, Zelazo PR. (eds) *The Development of Autism: Perspectives from Theory and Research*. Lawrence Erlbaum Associates: Mahwah, New Jersey, 2001.
- [160] Müller R-A, Kleinhans N, Kemmotsu N, Pierce K, Courchesne E. Abnormal variability and distribution of functional maps in autism: an fMRI study of visuomotor learning. *Am J Psychiatry* 2003; **160**:1847-1862.
- [161] Murakami J, Courchesne E, Press G, Yeung-Courchesne R, Hesselink J. Reduced cerebellar hemisphere size and its relationship to vermal hypoplasia in autism. *Arch Neurol* 1989; **46**:689-694.
- [162] Novick B, Kurtzberg D, Vaughn HG Jr. An electrophysiologic indication of defective information storage in childhood autism. *Psychiatry Res* 1979; **1**:101-108.
- [163] O'Riordan MA, Plaisted KC, Driver J, Baron-Cohen S. Superior visual search in autism. *J Exp Psychol Hum Percept Perform* 2001; **27**:719-730.
- [164] Ornitz EM, Hanna GL, de Traversay J. Prestimulation-induced startle modulation in attention-deficit hyperactivity disorder and nocturnal enuresis. *Psychophysiol* 1992; **29**:437-451.
- [165] Ornitz EM, Lane SJ, Sugiyama T, de Traversay J. Startle modulation studies in autism. *J Autism Dev Disord* 1993; **23**:619-637.
- [166] Ozonoff S, Pennington B, Rogers SJ. Executive function deficits in high-functioning autistic individuals: relationship to theory of mind. *J Child Psychol Psychiatry* 1991; **32**:1081-1105.
- [167] Park C, Falls W, Finger JH, Longo-Guess CM, Ackerman SL. Deletion in *Catna2*, encoding α N-catenin, causes cerebellar and hippocampal lamination defects and impaired startle modulation. *Nat Genet* 2002; **31**:279-284.
- [168] Patterson PH. Maternal infection: window on neuroimmune interactions in fetal brain development and mental illness. *Curr Opin Neurobiol* 2002; **12**:115-118.
- [169] Perry E, Martin-Ruiz C, Lee M, Griffiths M, Johnson M, Piggott M, Haroutunian V, Buxbaum JD, Nasland J, Davis K, Gotti C, Clementi F, Tzartos S, Cohen O, Soreq H, Jaros E, Perry R, Ballard C, McKeith I, Court J. Nicotinic receptor subtypes in human brain ageing, Alzheimer and Lewy body diseases. *Eur J Pharmacol* 2000; **393**:215-222.
- [170] Peters A, Sethares C. The organization of double bouquet cells in monkey striate cortex. *J Neurocytol* 1997; **26**:779-797.
- [171] Picciotto MR, Brunez DH, Caldarone BJ. Effect of nicotine and nicotinic receptors on anxiety and depression. *Neuroreport* 2002; **13**:1097-1106.
- [172] Pickles A, Bolton P, Macdonald H, Bailey A, Le Couteur A, Sim CH, Rutter M. Latent-class analysis of recurrence risks for complex phenotypes with selection and measurement error: A twin and family history study of autism. *Am J Hum Genet* 1995; **57**:717-726.
- [173] Pierce K, Courchesne E. Evidence for a cerebellar role in reduced exploration and stereotyped behavior in autism. *Biol Psychiatry* 2001; **49**:655-664.
- [174] Pierce K, Müller R-A, Ambrose J, Allen G, Courchesne E. Face processing occurs outside the fusiform 'face area' in autism: evidence from functional MRI. *Brain* 2001; **124**:2059-2073.

- [175] Pieretti M, Zhang FP, Fu YH, Warren ST, Oostra BA, Caskey CT, Nelson DL. Absence of expression of the FMR-1 gene in fragile X syndrome. *Cell* 1991; **23**:817-822.
- [176] Piven J, Gayle J, Chase GA, Fink B, Landa R, Wzorek MM, Folstein SE. A family history study of neuropsychiatric disorders in the adult siblings of autistic individuals. *J Am Acad Child Adolesc Psychiatry* 1990; **29**:177-183.
- [177] Piven J, Palmer P. Cognitive deficits in parents from multiple-incidence autism families. *J Child Psychol Psychiatry* 1994; **38**:1011-1021.
- [178] Piven J, Palmer P, Landa R, Santangelo S, Jacobi D, Childress D. Personality and language characteristics in parents from multiple-incidence autism families. *Am J Med Genet* 1997; **74**:398-411.
- [179] Piven J, Wzorek M, Landa R, Lainhart J, Bolton P, Chase GA, Folstein S. Personality characteristics of the parents of autistic individuals. *Psychol Med* 1994; **24**:783-795.
- [180] Plaisted KC, Saksida L, Alcántara J, Weisblatt E. Towards an understanding of the mechanisms of weak central coherence effects: experiments in visual configural learning and auditory perception. *Phil Trans R Soc Lond B* 2003; **358**:375-386.
- [181] Plaisted KC, Swettenham J, Rees L. Children with autism show local precedence in a divided attention task and global precedence in a selective attention task. *J Child Psychol Psychiatry* 1999; **40**:733-742.
- [182] Pletnikov MV, Rubin SA, Vasudevan K, Moran TH, Carbone KM. Developmental brain injury associated with abnormal play behavior in neonatally Borna disease virus-infected Lewis rats: a model of autism. *Behav Brain Res* 1999; **100**:43-50.
- [183] Posey DJ, McDougle CJ. The pharmacotherapy of target symptoms associated with autistic disorder and other pervasive developmental disorders. *Harvard Review of Psychiatry* 2000; **8**:45-63.
- [184] Powell EM, Campbell DB, Stanwood GD, Davis C, Noebels JL, Levitt P. Genetic Disruption of Cortical Interneuron Development Causes Region- and GABA Cell Type-Specific Deficits, Epilepsy, and Behavioral Dysfunction *J Neurosci* 2003; **23**:622-631.
- [185] Powell SB, Geyer, MA. Developmental markers of psychiatric disorders as identified by sensorimotor gating. *Neurotoxicity Res* 2002; **4**:489-502.
- [186] Pritchard JK. Are rare variants responsible for susceptibility to complex diseases? *Am J Hum Genet* 2001; **69**:124-137.
- [187] Raymond G, Bauman M, Kemper T. Hippocampus in autism: a Golgi analysis. *Acta Neuropathol* 1996; **91**:117-119.
- [188] Reik W, Dean W, Walter J. Epigenetic reprogramming in mammalian development. *Science* 2001; **293**:1089-1093.
- [189] Rinehart NJ, Bradshaw JL, Moss SA, Brereton AV, Tonge BJ. A deficit in shifting attention present in high-functioning autism but not Asperger's disorder. *Autism* 2001; **5**:67-80.
- [190] Ring HA, Baron-Cohen S, Wheelwright S, Williams SCR, Brammer MJ, Andrew C, Bullmore ET. Cerebral correlates of preserved cognitive skills in autism. *Brain* 1999; **122**:1305-1315.
- [191] Risch N, Spiker D, Lotspeich L, Nouri N, Hinds D, Hallmayer J, Kalaydjieva L, McCague P, Dimiceli S, Pitts T, Nguyen L, Yang J, Harper C, Thorpe D, Vermeer S, Young H, Hebert J, Lin A, Ferguson J, Chiotti C, Wiese-Slater S, Rogers T, Salmon B, Nicholas P, Petersen PB, Pingree C, McMahon W, Wong DL, Cavalli-Sforza LL, Kraemer HC, Myers RM. A genomic screen of autism: evidence for a multilocus etiology. *Am J Hum Genet* 1999; **65**:493-507.
- [192] Ritvo ER, Freeman BJ, Scheibel AB, Duong T, Robinson H, Guthrie D, Ritvo A. Lower Purkinje cell counts in the cerebella of four autistic subjects: Initial findings of the UCLA-NSAC autopsy research report. *Am J Psychiatry* 1986; **143**:862-866.
- [193] Rodier PM, Hyman SL. Early environmental factors in autism. *Ment Retard Dev Disability Res Reviews* 1998; **4**:121-128.
- [194] Rodier PM, Ingram JL, Tisdale B, Nelson S, Romano J. Embryological origin for autism: developmental anomalies of the cranial nerve motor nuclei. *J Comp Neurol* 1996; **370**:247-261.
- [195] Rogers SJ, Pennington BF. A theoretical approach to the deficits in infantile autism. *Dev Psychopathol* 1991; **3**:137-162.
- [196] Rogers SJ, Wehner DE, Hagerman R. The behavioral phenotype in fragile X: symptoms of autism in very young children with fragile X syndrome, idiopathic autism, and other developmental disorders. *J Dev Behav Pediatr* 2001; **22**:409-417.
- [197] Rogers T, Kalaydjieva L, Hallmayer J, Peterson PB, Nicholas P, Pingree C, McMahon WM, Spiker D, Lotspeich L, Kraemer H, McCague P, Dimiceli S, Nouri N, Peachy T, Yang J, Hinds D, Risch N, Meyers RM. Exclusion of linkage to the HLA region in ninety multiplex sibships with autism. *J Autism Dev Disord* 1999; **29**:195-201.

- [198] Rolf LH, Haarmann FY, Grotemeyer KH, Kehrer H. Serotonin and amino acid content in platelets of autistic children. *Acta Psychiatr Scand* 1993; **87**:312-316.
- [199] Rubenstein JLR, Merzenich MM. Model of autism: increased ratio of excitation/inhibition in key neural systems. *Genes Brain Behav* 2003; **2**:255-267.
- [200] Rubia K. The dynamic approach to neurodevelopmental psychiatric disorders: use of fMRI combined with neuropsychology to elucidate the dynamics of psychiatric disorders, exemplified in ADHD and schizophrenia. *Behav Brain Res* 2002; **130**:47-56.
- [201] Russell J, Saltmarsh R, Hill E. What do executive factors contribute to the failure on false belief tasks by children with autism? *J Child Psychol Psychiatry* 1999; **40**:859-868.
- [202] Ryan AK, Bartlett K, Clayton P, Eaton S, Mills L, Donnai D, Winter RM, Burn J, Smith-Lemli-Opitz syndrome: a variable clinical and biochemical phenotype. *J Med Genet* 1998; **35**:558-565.
- [203] Saitoh O, Karns CM, Courchesne E. Development of the hippocampal formation from 2 to 42 years: MRI evidence of smaller area dentata in autism. *Brain* 2001; **124**:1317-1324.
- [204] Schain RJ, Freedman DX. Studies on 5-hydroxyindole metabolism in autistic and other mentally retarded children. *J Pediatr* 1961; **58**:315-320.
- [205] Schultz RT, Gauthier I, Klin A, Fulbright RK, Anderson AW, Volkmar F, Skudlarski P, Lacadie C, Cohen DJ, Gore JC. Abnormal ventral temporal cortical activity during face discrimination among individuals with autism and Asperger syndrome. *Arch Gen Psychiatry* 2000; **57**:331-340.
- [206] Shah A, Frith U. An islet of ability in autistic children: a research note. *J Child Psychol Psychiatry* 1983; **24**:613-620.
- [207] Shah A, Frith U. Why do autistic individuals show superior performance on the block design task? *J Child Psychol Psychiatry* 1993; **34**:1351-1364.
- [208] Shi L, Fatemi SH, Sidwell RW, Patterson PH. Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring. *J Neurosci* 2003; **23**:297-302.
- [209] Silverman JM, Smith CJ, Schmeidler J, Hollander E, Lawlor BA, Fitzgerald M, Buxbaum JD, Delaney K, Galvin P. Symptom domains in autism and related conditions: evidence for familiarity. *Am J Med Genet* 2002; **114**:64-73.
- [210] Singh VK, Warren R, Averett R, Ghaziuddin M. Circulating autoantibodies to neuronal and glial filament proteins in autism. *Pediatr Neurol* 1997; **17**:88-90.
- [211] Singh VK, Warren RP, Odell JD, Warren WL, Cole P. Antibodies to myelin basic protein in children with autistic behavior. *Brain Behav Immun* 1993; **7**:97-103.
- [212] Skuse D, James R, Bishop D, Coppin B, Dalton P, Aamodt-Leeper G, Bacarese-Hamilton M, Creswell C, McGurk R, Jacobs P. Evidence from Turner's syndrome of an imprinted X-linked locus affecting cognitive function. *Nature* 1997; **387**:705-708.
- [213] Smith DW, Lemli L, Opitz JM. A newly recognized syndrome of multiple congenital anomalies. *J Pediatr* 1964; **64**:210-217.
- [214] Smith IM. Motor functioning in Asperger's syndrome. In: Klin A, Volkmar FR, Sparrow SS. (eds) *Asperger's Syndrome*. The Guildford Press: New York, 2000.
- [215] Sparks BF, Friedman SD, Shaw DW, Aylward EH, Echelard D, Artru AA, Maravilla KR, Giedd JN, Munson J, Dawson G, Dager SR. Brain structural abnormalities in young children with autism spectrum disorder. *Neurology* 2002; **59**:184-192.
- [216] Stevens S, Gruzelier J. Electrodermal activity to auditory stimuli in autistic, retarded, and normal children. *J Autism Dev Disord* 1984; **14**:245-260.
- [217] Stubbs EG, Crawford ML. Depressed lymphocyte responsiveness in autistic children. *J Autism Child Schizophr* 1977; **7**:49-55.
- [218] Stühmer T, Anderson SA, Ekker M, Rubenstein JLR. Ectopic expression of the *Dlx* genes induces glutamic acid decarboxylase and *Dlx* expression. *Development* 2002; **129**:245-252.
- [219] Stühmer T, Puelles L, Ekker M, Rubenstein JLR. Expression from a *Dlx* gene enhancer marks adult mouse cortical GABAergic neurons. *Cereb Cortex* 2002; **12**:75-85.
- [220] Swerdlow NR, Benbow CH, Zisook S, Geyer MA, Braff DL. A preliminary assessment of sensorimotor gating in patients with obsessive compulsive disorder. *Biol Psychiatry* 1993; **33**:298-301.
- [221] Swerdlow NR, Braff DL, Taaid N, Geyer MA. Assessing the validity of an animal model of deficient sensorimotor gating in schizophrenic patients. *Arch Gen Psychiatry* 1994; **51**:139-154.
- [222] Swerdlow NR, Geyer MA, Braff DL. Neural circuit regulation of prepulse inhibition of startle in the rat: current knowledge and future challenges. *Psychopharm* 2001; **156**:194-215.
- [223] Swerdlow NR, Paulsen J, Braff DL, Butters N, Geyer MA, Swenson MR. Impaired prepulse inhibition of acoustic and tactile startle response in patients with Huntington's disease. *J Neurol Neurosurg*

- Psychiatry* 1995; **58**:192-200.
- [224] Tantam D, Monaghan L, Nicholson H, Stirling J. Autistic children's ability to interpret faces: a research note. *J Child Psychol Psychiatry* 1989; **30**:623-630.
- [225] Tierney E, Nwokoro NA, Porter FD, Freund LS, Ghuman JK, Kelly RI. The behavior phenotype in the RSH/ Smith-Lemli-Opitz syndrome. *Am J Med Genet*, 2001; **98**:191-200.
- [226] Tint GS, Irons M, Elias ER, Batta AK, Frieden R, Chen TS, Salen G. Defective cholesterol biosynthesis associated with the Smith-Lemli-Opitz syndrome. *N Engl J Med* 1994; **330**:107-113.
- [227] Todd PK, Mack KJ. Sensory stimulation increases cortical expression of the fragile X mental retardation protein in vivo. *Mol Brain Res* 2000; **80**:17-25.
- [228] Toichi M, Kamio Y. Long-term memory and levels-of-processing in autism. *Neuropsychologia* 2002; **40**:964-969.
- [229] Tordjman S, Anderson GM, McBride PA, Hertzog ME, Snow ME, Hall LM, Thompson SM, Ferrari P, Cohen DJ. Plasma β -endorphin, adrenocorticotropin hormone, and cortisol in autism. *J Child Psychol Psychiatry* 1997; **38**:705-715.
- [230] Tordjman S, Gutnecht L, Carlier M, Spitz E, Antoine C, Slama F, Cohen DJ, Ferrari P, Roubertoux PL, Anderson GM. Role of the serotonin transporter in the behavioral expression of autism. *Mol Psychiatry* 2001; **6**:434-439.
- [231] Torrao AS, Britto LR. Neurotransmitter regulation of neural development: acetylcholine and nicotinic receptors. *An Acad Bras Cienc* 2002; **74**:453-461.
- [232] Torres AR, Maciulis A, Stubbs EG, Cutler A, Odell D. The transmission disequilibrium test suggests that HLA-DR4 and DR13 are linked to autism spectrum disorder. *Hum Immunol* 2002; **63**:311-316.
- [233] Townsend J, Courchesne E. Parietal damage and narrow "spotlight" spatial attention. *J Cogn Neurosci* 1994; **6**:220-232.
- [234] Townsend J, Courchesne E, Egaas B. Slowed orienting of covert visual-spatial attention in autism: specific deficits associated with cerebellar and parietal abnormality. *Dev Psychopathol* 1996; **8**:563-584.
- [235] Townsend J, Courchesne E, Covington J, Westerfield M, Harris NS, Lyden P, Lowry TP, Press GA. Spatial attention deficits in patients with acquired or developmental cerebellar abnormality. *J Neurosci* 1999; **19**:5632-5643.
- [236] Townsend J, Singer-Harris N, Courchesne E. Visual attention abnormalities in autism: delayed orienting to location. *J Int Neuropsychol Soc* 1996; **2**:541-550.
- [237] Townsend J, Westerfield M, Leaver E, Makeig S, Jung T, Pierce K, Courchesne E. Event-related brain response abnormalities in autism: evidence for impaired cerebello-frontal spatial attention networks. *Cogn Brain Res* 2001; **11**:127-145.
- [238] Tueting P, Costa E, Dwivedi Y, Guidotti A, Impagnatiello F, Manev R, Pesokd C. The phenotypic characteristics of heterozygous reeler mouse. *Neuroreport* 1999; **10**:1329-1334.
- [239] Ugarte SD, Homanics GE, Firestone LL, Hammond DL. Sensory thresholds and the antinociceptive effects of GABA receptor agonists in mice lacking the β_3 subunit of the GABA_A receptor. *Neuroscience* 2000; **95**:795-806.
- [240] Vaillancourt C, Boksa P. Birth insult alters dopamine-mediated behavior in a precocial species, the guinea pig. Implications for schizophrenia. *Neuropsychopharm* 2000; **23**:654-666.
- [241] van Engeland H. The electrodermal orienting response to auditory stimuli in autistic children, normal children, mentally retarded children, and child psychiatric patients. *J Autism Dev Disord* 1984; **14**:261-279.
- [242] Van Gent T, Heijnen CJ, Treffers PD. Autism and the immune system. *J Child Psychol Psychiatry* 1997; **38**:337-349.
- [243] Veenstra-VanderWeele J, Anderson GM, Cook EH Jr. Pharmacogenetics and the serotonin system: initial studies and future directions. *Eur J Pharmacol* 2000; **410**:165-181.
- [244] Verbaten MN, Roelofs JW, van Engeland H, Kenemans JK, Slangen JL. Abnormal visual event-related potentials of autistic children. *J Autism Dev Disord* 1991; **21**:449-470.
- [245] Vilensky JA, Damasio AR, Maurer RG. Gait disturbances in patients with autistic behavior: a preliminary study. *Arch Neurol* 1981; **38**:646-649.
- [246] Wainwright-Sharp JA, Bryson SE. Visual orienting deficits in high-functioning people with autism. *J Autism Dev Disord* 1993; **23**:1-13.
- [247] Wainwright-Sharp JA, Bryson SE. Visual-spatial orienting in autism. *J Autism Dev Disord* 1996; **26**:423-438.
- [248] Ward NL, Hagg T. BDNF is needed for postnatal maturation of basal forebrain and neostriatum cholinergic neurons in vivo. *Exp Neurol* 2000; **162**:297-310.

- [249] Warren RP, Burger RA, Odell D, Torres AR, Warren WL. Decreased plasma concentrations of the C4B complement protein in autism. *Arch Pediatr Adolesc Med* 1994; **148**:180-183.
- [250] Warren RP, Foster A, Margaretten NC. Reduced natural killer cell activity in autism. *J Amer Acad Child Adolesc Psychiatry* 1987; **26**:333-335.
- [251] Warren RP, Foster A, Margaretten NC, Pace NC. Immune abnormalities in patients with autism. *J Autism Dev Disord* 1986; **16**:189-197.
- [252] Warren RP, Odell JD, Warren WL, Burger RA, Maciulis A, Daniels WW, Torres AR. Strong association of the third hypervariable region of HLA-DR β_1 with autism. *J Neuroimmunol* 1996; **67**:97-102.
- [253] Warren RP, Singh VK, Cole P, Odell JD, Pingree CB, Warren WL, DeWitt CW, McCullough M. Possible association of the extended MHC haplotype B44-SC30-DR4 with autism. *Immunogenetics* 1992; **36**:203-207.
- [254] Warren RP, Singh VK, Cole P, Odell JP, Pingree CB, Warren WL, White E. Increased frequency of the null allele at the complement C4b locus in autism. *Clin Exp Immunol* 1991; **83**:438-440.
- [255] Warren RP, Yonk LJ, Burger RA, Cole P, Odell JD, Warren WL, White E, Singh VK. Deficiency of suppressor-inducer (CD4+CD45RA+) T cells in autism. *Immunol Invest* 1990; **19**:245-251.
- [256] Wassink TH, Piven J, Patil SR. Chromosomal abnormalities in a clinic sample of individuals with autistic disorder. *Psychiatr Genet* 2001; **11**:57-63.
- [257] Weiler IJ, Irwin SA, Klintsova AY, Spencer, CM Brazelton AD, Miyashiro K, Comery TA, Patel B, Eberwine J, Greenough WT. Fragile X mental retardation protein is translated near synapses in response to neurotransmitter activation. *Proc Natl Acad Sci USA* 1997; **94**:5395-5400.
- [258] Weiss IC, Feldon J. Environmental animal models for sensorimotor gating deficiencies in schizophrenia: a review. *Psychopharm* 2001; **156**:305-326.
- [259] Weizman A, Weizman R, Szekely GA, Wijisenbeek H, Livni E. Abnormal immune response to brain tissue antigen in the syndrome of autism. *Am J Psychiatry* 1982; **139**:1462-1465.
- [260] Whitaker-Azmitia PM. Serotonin and brain development: role in human developmental diseases. *Brain Res Bull* 2001; **56**:479-485.
- [261] Whitehouse D, Harris JC. Hyperlexia in infantile autism. *J Autism Dev Disord* 1984; **14**:281-289.
- [262] Williams RS, Hauser SL, Purpura DP, DeLong GR, Swisher CN. Autism and mental retardation: Neuropathologic studies performed in four retarded persons with autistic behavior. *Arch Neurol* 1980; **37**:749-753.
- [263] Wisor JP, DeLorey TM, Homanics GE, Edgar DM. Sleep states and sleep electroencephalographic spectral power in mice lacking the β_3 subunit of the GABA_A receptor. *Brain Res* 2002; **955**:221-228.
- [264] Yakovlev PI, Lecours A-R. The myelogenetic cycles of regional maturation of the brain. In: Minkowski A. (ed) *Regional Development of the Brain in Early Life*. Blackwell: Oxford, 1967, pp 3-65.
- [265] Yonk LJ, Warren RP, Burger RA, Cole P, Odell J, Warren WL, White E, Singh VK. CD4+ helper T cell depression in autism. *Immunol Lett* 1990; **25**:341-345.
- [266] Young MR, Kut JL, Coogan MP, Wright MA, Young ME, Matthews J. Stimulation of splenic T-lymphocyte function by endogenous serotonin and by low-dose exogenous serotonin. *Immunology* 1993; **80**:395-400.
- [267] Yu Y, Bradley A. Engineering chromosomal rearrangements in mice. *Nat Rev Genet* 2001; **2**:780-790.