Effects of Protocol Design on Lactate Minimum Power

Abstract

The aim of this investigation was to use a validated lactate minimum test protocol and evaluate whether blood lactate responses and the lactate minimum power are influenced by the starting power (study 1) and 1 min inter-stage rest intervals (study 2) during the incremental phase. Study 1: 8 subjects performed a lactate minimum test comprising a lactate elevation phase, recovery phase, and incremental phase comprising 5 continuous 4 min stages with starting power being 40% or 45% of the maximum power achieved during the lactate elevation phase, and with power increments of 5% maximum power.

Study 2: 8 subjects performed 2 identical lactate minimum tests except that during one of the tests the incremental phase included 1 min inter-stage rest intervals. The lactate minimum power was lower when the incremental phase commenced at 40% (175±29 W) compared to 45% (184±30 W) maximum power (p<0.01), and was increased when 1 min inter-stage rest intervals were included during the incremental phase (192±25 vs. 200±26 W, p<0.01). In conclusion, changes in lactate minimum power were small and thus unlikely to compromise test validity and therefore training status evaluation and exercise prescription.

Introduction

The maximal lactate steady state (MLSS) represents the highest exercise intensity at which blood lactate concentration ([lac\textsuperscript{−}]\textsubscript{b}) remains stable over time and is widely acknowledged as an important determinant of endurance exercise performance [11–13]. Unfortunately, direct MLSS determination is laborious, requiring multiple, constant-intensity exercise tests (typically lasting 30 min) over a range of exercise intensities. Numerous protocols have therefore been designed to predict MLSS using a single exercise test [11, 25]. The lactate minimum test was originally proposed by Tegtbur et al. [26] as a method to predict MLSS running velocity, and the protocol has since been adapted for MLSS prediction in cycling [11, 13, 21, 22, 24] and swimming [15]. The lactate minimum represents the workload corresponding to the nadir of a U-shaped [lac\textsuperscript{−}]\textsubscript{b} profile during an incremental exercise test (hereafter termed incremental phase) initiated with hyperlactataemia due to a preceding high-intensity exercise bout (hereafter termed lactate elevation phase) and short recovery (hereafter termed recovery phase). Collectively, the literature suggests good agreement between lactate minimum and MLSS workloads [1, 13, 20, 23, 26], and we recently described a novel cycling lactate minimum test that provides a good estimate of MLSS and a concurrent measure of maximal oxygen uptake [11]. Interestingly, however, temporal changes in [lac\textsuperscript{−}]\textsubscript{b} during the incremental phase failed to reflect the changes observed during constant power exercise at equivalent intensities, and [lac\textsuperscript{−}]\textsubscript{b} responses and the lactate minimum power were modified when the lactate elevation phase was performed using remote muscle groups (arm-cranking) [11]. These observations suggest that lactate kinetics during the incremental phase are modified by the preceding lactate elevation phase, and that altering certain aspects of the lactate minimum protocol may worsen the agreement between lactate minimum and MLSS powers [11].

Many variants of the lactate minimum protocol have been published. For example, inter-stage rest intervals during the incremental phase of either 30 s [16, 26], 45 s [15, 19], or more commonly 1 min [5, 10, 13, 17, 20, 23] have been used.
Whether the inclusion of rest intervals during the incremental phase affects the lactate minimum workload remains unknown. The intensity at which the incremental phase commences has also varied and Carter et al. [4] concluded that starting intensity affected lactate minimum running speed. However, during the incremental phase [lac\(^-\)\(_{1}\)] reached, or approached, that seen at rest, which prevents a genuine lactate minimum nadir being observed and therefore renders that study inconclusive [13]. Therefore, using our recently described, validated lactate minimum test protocol [11], the aims of this investigation were to examine whether [lac\(^-\)\(_{1}\)] responses and the lactate minimum power are affected by using different incremental phase starting powers (study 1) and the introduction of 1 min inter-stage rest intervals during the incremental phase (study 2).

Method

Participants, equipment and measurements

Studies 1 and 2 were approved by the Nottingham Trent University ethics committee, and were conducted in accordance with the principles of the Declaration of Helsinki and the International Journal of Sports Medicine [8]. 16 non-smoking recreationally active (performing 30–90 min of aerobic exercise, 3–4 times per week) male subjects provided written informed consent to participate in the study. Subjects refrained from strenuous exercise during the 24 h preceding an exercise test. On test days subjects abstained from alcohol and caffeine and reported to the laboratory at least 2 h post-prandial. Successive tests were separated by at least 48 h, but no more than 1 week, and were performed at a similar time of day. Tests within each study were performed in a counterbalanced order.

Exercise was performed on an electromagnetically-braked cycle ergometer (Excalibur Sport, Lode, Groningen, The Netherlands). The same self-selected cycling cadence (83 ± 8 revs · min\(^{-1}\)) was used during all tests. Arterialised venous blood samples were taken from a dorsal hand vein via an indwelling cannula and analysed for [lac\(^-\)\(_{1}\)] (P-GM7 MicroStat, Analox Instruments, London, UK). During the lactate elevation phase, subjects wore a facemask (model 7940, Hans Rudolph, Kansas City, Missouri) and respiratory variables were measured breath-by-breath (Pulmolab EX670, Ferraris Respiratory Europe, Hertford, UK).

Study 1 – effect of different starting powers during the incremental phase

Subjects (n=8; age 23.6 ± 4.5 years; height 179.4 ± 4.4 cm; body mass 79.5 ± 8.1 kg) performed 2 lactate minimum tests as described in Johnson et al. [11]. Each test comprised 3 consecutive phases: (I) lactate elevation phase comprising maximal incremental exercise; (II) 8 min recovery phase at 60 W; and (III) incremental phase comprising five 4 min exercise stages, starting at either 40% or 45% of the maximal power (Wmax) achieved during the lactate elevation phase, and with power increments of 5% Wmax (hereafter these tests are termed LM\(_{40-60}\) and LM\(_{45-65}\), respectively). During the lactate elevation phase power was increased every 1.5 s by a constant amount (8–10 W, depending upon the subjects training history) chosen to elicit exercise intolerance (cadence <60 revs · min\(^{-1}\) in ~10 min. The final power and the highest oxygen uptake recorded over any 30 s period defined Wmax and maximal oxygen uptake, respectively. During their second lactate minimum test, subjects were encouraged to exercise for the same duration during the lactate elevation phase as that achieved in the initial lactate minimum test, at which point the lactate elevation phase was terminated. Exercise duration during the lactate elevation phase was matched in an attempt to replicate the magnitude of lactate elevation, and thus the starting [lac\(^-\)\(_{1}\)] at the onset of the incremental phase. We did not conduct a lactate minimum test in which the incremental phase began at 50% Wmax because pilot work showed that 70% Wmax (which would represent the final stage of the incremental phase) represented an intolerable workload. Blood samples were taken every minute following the first minute of the recovery phase.

Study 2 – effect of inter-stage rest interval during the incremental phase

Subjects (n=8; age 24.5 ± 5.8 years; height 177.3 ± 6.0 cm; body mass 77.5 ± 7.0 kg) performed 2 lactate minimum tests as described for LM\(_{45-65}\). The 2 tests were performed with (LM\(_{INT}\)) and without (LM\(_{CONT}\)) 1 min inter-stage rest intervals during the incremental phase. During their second lactate minimum test, subjects were again encouraged to exercise for the same duration during the lactate elevation phase as that achieved in the initial lactate minimum test. Blood samples were taken every minute during the incremental phase.

Data analyses

Data analyses were performed using SPSS. The Shapiro-Wilk test was used to confirm that the data were normally distributed. The area under the [lac\(^-\)\(_{1}\)] curve in the recovery phase of study 1 was calculated using the trapezoidal rule. The lactate minimum power was determined from the zero gradient tangent to a cubic spline function fitting the [lac\(^-\)\(_{1}\)] (measured at the end of each stage) vs. power data. Rates of change of [lac\(^-\)\(_{1}\)] (Δ[lac\(^-\)\(_{1}\)]/Δt, where t = time) during each incremental phase stage were taken as the gradient of a linear regression of [lac\(^-\)\(_{1}\)] against time. Data were analysed using repeated measures ANOVA and paired t-tests where appropriate. Pearson product-moment correlation coefficients (r) were determined to assess the relationship between variables. Results are reported as mean ± SD. Statistical significance was set at p < 0.05.

Results

The [lac\(^-\)\(_{1}\)] profile during the incremental phase was well described by the cubic spline function (study 1: R\(^2\) = 0.96 ± 0.04 and 0.97 ± 0.04 in LM\(_{40-60}\) and LM\(_{45-65}\), respectively; study 2: R\(^2\) = 0.99 ± 0.02 and 0.92 ± 0.07 in LM\(_{CONT}\) and LM\(_{INT}\), respectively), with R\(^2\) values in LM\(_{CONT}\) being higher than those in LM\(_{INT}\) (p < 0.05). Lactate minimum power was correlated (r = 0.09) with Wmax in both LM\(_{40-60}\) and LM\(_{45-65}\), but r = 0.97 and 0.99 in LM\(_{CONT}\) and LM\(_{INT}\), respectively.

Study 1

The Wmax and maximal oxygen uptake during the first lactate minimum test were 344 ± 56 W and 3.67 ± 0.45 L · min\(^{-1}\), respectively. For each subject exercise duration during the lactate elevation phase was identical in both lactate minimum tests. All subjects demonstrated a declining [lac\(^-\)\(_{1}\)] during the recovery phase (see Fig. 1) and the calculated area under the [lac\(^-\)\(_{1}\)] curve in LM\(_{40-60}\) (46 ± 6 mmol) was not different from that determined in LM\(_{45-65}\) (47 ± 7 mmol). The [lac\(^-\)\(_{1}\)] at the end of the recovery phase was 6.8 ± 1.1 mmol · L\(^{-1}\) (range: 5.2–8.2 mmol · L\(^{-1}\)) in...
The lactate minimum power was lower in LM 40–60 (175 ± 29 W, 50.9 ± 0.9% Wmax) compared to LM 45–65 (184 ± 30 W, 53.5 ± 1.2% Wmax) (p < 0.01) (● ▶ Fig. 1). The lactate minimum [lac–B] was lower in LM 40–60 (4.3 ± 1.4 mmol·L⁻¹, range: 3.0–6.9 mmol·L⁻¹) compared to LM 45–65 (5.1 ± 1.5 mmol·L⁻¹, range: 3.8–8.4 mmol·L⁻¹) (p < 0.01). The time taken to reach the lactate minimum from the start of the incremental phase was longer in LM 40–60 (12.74 ± 0.77 min) compared to LM 45–65 (10.65 ± 0.91 min) (p < 0.01). Δ[lac–B]/Δt was higher in LM 40–60 compared to LM 45–65 at 50% and 55% Wmax (p < 0.01) (● ▶ Fig. 2).

Study 2

The Wmax and maximal oxygen uptake during the first lactate minimum test were 363 ± 49 W and 3.64 ± 0.49 L·min⁻¹, respectively. Exercise duration during the lactate elevation phase was identical in both lactate minimum tests. The [lac–B] at the end of the recovery phase was 6.7 ± 1.6 mmol·L⁻¹ (range: 4.6–9.8 mmol·L⁻¹) in LMCONT and 6.4 ± 1.1 mmol·L⁻¹ (range: 4.9–8.3 mmol·L⁻¹) in LMINT. The lactate minimum power was higher in LMINT (200 ± 26 W, 55.3 ± 1.1% Wmax) compared to LMCONT (192 ± 25 W, 52.9 ± 1.9% Wmax) (p < 0.01) (● ▶ Fig. 3). The lactate minimum [lac–B] was lower in LMINT (4.0 ± 1.1 mmol·L⁻¹, range:...
2.5–6.0 mmol·L⁻¹) compared to LM_CONT (4.9 ± 0.7 mmol·L⁻¹, range: 2.9–8.6 mmol·L⁻¹) (p < 0.05). Following the first stage of the incremental phase, [lac⁻]ᵢ was determined over 3–5 min of each stage. Values were mean ± SD. The main finding of this investigation was that despite [lac⁻]ᵢ responses being modified by starting the incremental phase at a lower power (study 1) and including 1 min inter-stage rest intervals during the incremental phase (study 2), the effects on lactate minimum power are small.

The 9W reduction in lactate minimum power when the incremental phase was initiated at a lower intensity is consistent with the findings of Carter et al. [4], although in this study [lac⁻]ᵢ reached, or approached, resting values during the incremental phase, which according to MacIntosh et al. [13] renders their study inconclusive. Conversely, in study 1 [lac⁻]ᵢ remained ≥3.0 mmol·L⁻¹ in all subjects throughout the incremental phase, thus indicating a genuine influence of starting intensity on the lactate minimum power. That our protocol [11] standardises starting intensity for the incremental phase is therefore advantageous in comparison to protocols in which knowledge of training status [1, 12, 26] or separate testing [5, 13, 15, 20, 22] was required. However, the difference in lactate minimum power between LM_40–60 and LM_45–65 is well within the 95% limits of agreement (22W) for the comparison of lactate minimum (from LM_45–65) and MLSS powers [11] and is also less than the resolution with which MLSS is typically determined (10–20W) [25]. Thus using LM_40–60 is unlikely to compromise test validity, and therefore training status evaluation or exercise prescription. On the contrary, given that LM_45–65 could be considered physically challenging to complete, the lower intensities performed during the incremental phase of LM_40–60 may provide a more tolerable test for certain individuals (e.g. untrained, elderly and/or clinical patients) [14, 18].

The greater decline in [lac⁻]ᵢ when the incremental phase commenced at a lower power was probably due to greater lactate clearance by oxidative muscle [2, 4] since low intensity exercise accelerates lactate clearance [2, 16]. Although the [lac⁻]ᵢ was lower throughout the incremental phase of LM_40–60 compared to LM_45–65, Δ[lac⁻]ᵢ/Δt was higher in LM_40–60 at equivalent intensities (50% and 55% Wmax). This supports the notion that Δ[lac⁻]ᵢ/Δt during each incremental phase stage does not solely reflect the metabolic demand of that exercise intensity, i.e., Δ[lac⁻]ᵢ/Δt during constant power exercise [11]. Rather, Δ[lac⁻]ᵢ/Δt is likely to be influenced by the underlying blood lactate recovery kinetics following the lactate elevation phase and preceding exercise stage [4, 11]. Therefore, at a given relative power, the greater exercise history during LM_40–60 compared to LM_45–65 may have promoted greater lactate release from passive lactate “reservoirs” (i.e., resting skeletal muscle) and active muscle [6], thus causing a slight leftward shift of the lactate minimum curve.

Discussion

The main finding of this investigation was that despite [lac⁻]ᵢ responses being modified by starting the incremental phase at a lower power (study 1) and including 1 min inter-stage rest intervals during the incremental phase (study 2), the effects on lactate minimum power are small.

The 9W reduction in lactate minimum power when the incremental phase was initiated at a lower intensity is consistent with the findings of Carter et al. [4], although in this study [lac⁻]ᵢ reached, or approached, resting values during the incremental phase, which according to MacIntosh et al. [13] renders their study inconclusive. Conversely, in study 1 [lac⁻]ᵢ remained ≥3.0 mmol·L⁻¹ in all subjects throughout the incremental phase, thus indicating a genuine influence of starting intensity on the lactate minimum power. That our protocol [11] standardises starting intensity for the incremental phase is therefore advantageous in comparison to protocols in which knowledge of training status [1, 12, 26] or separate testing [5, 13, 15, 20, 22] was required. However, the difference in lactate minimum power between LM_40–60 and LM_45–65 is well within the 95% limits of agreement (22W) for the comparison of lactate minimum (from LM_45–65) and MLSS powers [11] and is also less than the resolution with which MLSS is typically determined (10–20W) [25]. Thus using LM_40–60 is unlikely to compromise test validity, and therefore training status evaluation or exercise prescription. On the contrary, given that LM_45–65 could be considered physically challenging to complete, the lower intensities performed during the incremental phase of LM_40–60 may provide a more tolerable test for certain individuals (e.g. untrained, elderly and/or clinical patients) [14, 18].

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The purpose of the recovery phase is to allow $[\text{lac}^{-1}_B]$ to decrease prior to the incremental phase. A $[\text{lac}^{-1}_B]$ of $-8.0 \text{ mmol} \cdot \text{L}^{-1}$ at the onset of the incremental phase has been recommended [4], although a rationale was not provided. Although some subjects in our studies did not meet this criterion, a range of starting $[\text{lac}^{-1}_B]$ values evidently permit a valid test [11, 13]. It is, however, essential that the starting $[\text{lac}^{-1}_B]$ is sufficiently high to prevent $[\text{lac}^{-1}_B]$ approaching/reaching resting values during the incremental phase, which prevents the determination of a lactate minimum nadir.

Many studies have included inter-stage rest intervals during the incremental phase of either 30 s [16,26], 45 s [15,19], or more commonly 1 min [5,10,13,17,20,23]. The 8 W increase in lactate minimum power with 1 min inter-stage rest intervals concurs with a previous study in which MLSS cycling power increased from 278 W to 300 and 310 W when rest intervals of 0.5 and 1.5 min, respectively, were included every 5 min during each 30 min constant power MLSS determination test [3]. During incremental running exercise the speed at the 4.0 mmol · L$^{-1}$ $[\text{lac}^{-1}_B]$ threshold was unchanged with 0.5 min inter-stage rest intervals [7], but increased by $-0.70 \text{ km} \cdot \text{h}^{-1}$ with 1.5 min intervals [9]. Although increases in lactate transitions with test interruptions could result in an overestimate of endurance training status and an inappropriate prescription of training exercise intensity, the difference in lactate minimum power between $\text{LM}_{\text{CONT}}$ and $\text{LM}_{\text{INT}}$ is probably too small to have any practical significance (see above). Indeed, for reasons highlighted previously, the $\text{LM}_{\text{INT}}$ protocol would probably be more tolerable for certain individuals.

The lower $[\text{lac}^{-1}_B]$ during $\text{LM}_{\text{INT}}$ compared to $\text{LM}_{\text{CONT}}$ probably resulted from a reduced glycolytic rate during the rest interval combined with a sustained metabolic rate promoting whole-body lactate clearance [3,6]. 4 min may have thus provided insufficient time for $[\text{lac}^{-1}_B]$ to increase to the same value as that observed during $\text{LM}_{\text{CONT}}$, therefore causing a rightward shift of the lactate minimum curve. An increasing oscillatory response in $[\text{lac}^{-1}_B]$ was observed in the latter incremental stage of $\text{LM}_{\text{INT}}$, and $\Delta[\text{lac}^{-1}_B]/\Delta t$ (determined over 3–5 min of each stage) was higher compared to $\text{LM}_{\text{CONT}}$ (differences in $\Delta[\text{lac}^{-1}_B]/\Delta t$ also approached statistical significance at $55 \% (p=0.06)$ and $60 \% (p=0.09)$ $\text{Wmax}$). A rapid decline in metabolic rate during each 1 min rest interval would have predisposed a delayed kinetic response (i.e., metabolic inertia) at the onset of the subsequent exercise stage [18,27] and therefore, compared to $\text{LM}_{\text{CONT}}$, a larger contribution from non-oxidative energy pathways and thus greater lactate efflux from muscle.

Previous studies of lactate minimum test validity may have been influenced by the inclusion of rest intervals during the incremental phase and/or each 30 min constant load MLSS determination test. For example, MacIntosh et al. [13] included 1 min rest intervals during the incremental phase but not during MLSS determination tests. Although close agreement was observed between lactate minimum and MLSS powers, it is likely that lactate minimum power would have been lower had a continuous incremental phase been used (present study), whereas MLSS power would have been higher had 1 min rest intervals also accompanied each 5 min blood sample [3]. Furthermore, because changes in systemic lactate kinetics due to rest intervals are likely to differ between incremental exercise commencing with hyperlactataemia and 30 min square-wave constant load exercise [11], even identical rest interval durations may cause dissimilar shifts in lactate minimum and MLSS workloads [3,7], which may partly explain some of the individual discrepancies observed between lactate minimum and MLSS workloads [4,12,13,23,26].

A limitation of the present investigation (studies 1 and 2) was that particular measurements, including pulmonary gas exchange, heart rate, and rating of perceived exertion, were not taken at the end of the lactate elevation phase in both lactate minimum tests (within each study), nor were they taken during the incremental phase. For coaches and daily clinical practice such measures are valuable as they provide additional indicators of training status/physiological exertion and can inform exercise training prescription.

In summary, although the $[\text{lac}^{-1}_B]$ response to the incremental phase of our validated lactate minimum test protocol [11] is modified by a lower starting intensity and 1 min inter-stage rest intervals, the impact on lactate minimum power is minimal and unlikely to compromise test validity and therefore training status evaluation and exercise prescription. Our experience is that individuals of lower endurance training status with little familiarity of cycle exercise tolerate the validated protocol relatively poorly. Investigators may therefore wish to implement such protocol amendments for certain individuals/populations in order to make the test more tolerable. For daily practice and to inform future exercise prescription, concurrent measures of heart rate and rating of perceived exertion should also be taken. Furthermore, since $\text{LM}_{40-60}$ and $\text{LM}_{60-80}$ protocols decreased and increased, respectively, lactate minimum power by a similar amount (9 and 8 W, respectively), combining the amendments may yield a lactate minimum power more similar to that produced by the previously validated protocol [11] and further improve participants’ tolerance of the test. Future investigations should explore this possibility and evaluate the impact on MLSS prediction.

References