

ORIGINAL RESEARCH

A case controlled study of the oncological safety of fat grafting

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Abstract

Introduction

Fat grafting to the breast fulfils an increased clinical demand for a biocompatible filler in contour refinement, volume adjustment and tissue rejuvenation both in cosmetic and reconstructive procedures, and has been used for total breast reconstruction¹⁻⁵.

Whilst many clinical studies have reported on the efficacy of fat grafting for breast cancer patients in terms of its various indications^{2,6-9}, technical advancements^{7,9-13,14}, volume stability and graft survival^{5,8,9,12, 14,15-19}, radiologic safety^{2,3,12,20-22,29,30}, and complication rates^{9,12,13,23,24,25,29,30}, few studies focus on oncologic safety. Fewer still have adequate data and follow-up period to reach meaningful conclusions^{9,19,20,26-30}. Others report outcomes for cosmetic breast patients only^{5,24,31,32}. Where oncologic safety is assessed, many reviews focus on radiological sequelae interfering with mammographic surveillance^{1,12,13,15,21}.

Meta-analyses evaluating the oncologic safety of FG have begun to emerge in an effort to resolve this debate with a greater sample size^{20,33-35}. However, there are limitations to the previous studies, particularly the consideration for confounding variables such as tumour histology, resection margins, receptor status and adjuvant treatments, which can only be corrected for by comparison with an appropriate control group³⁵⁻³⁷.

Petit et al³⁹ reported the first case-matched retrospective series, (level 2b evidence). In this study we aimed to study a large series of breast cancer patients treated with FG in one institution using a matched cohort approach, and then systematically review relevant published cohort studies to which our results could be compared.

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Method

Between January 2007 and August 2013, 396 patients were treated with fat grafting (FG) to the breast at the Nottingham Breast Institute (NBI) for a variety of indications, including; breast asymmetry, contour deformity, correction of radiotherapy induced fibrosis and volume enhancement. Excluded from this study were; benign conditions (68), women whose primary oncologic surgery was performed elsewhere with missing cancer data (51), disease recurrence prior to fat grafting (35), and failure to identify a suitable case control match

(31). Data for 211 patients (DCIS = 27, Invasive carcinoma = 184) treated for breast cancer between 1977 and 2013 was included.

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For each of the 211 FG patients included in this study, two control subjects were matched from a prospective database of women who were treated for primary breast cancer at NBI and did not undergo FG intervention. Each control was matched for five variables; date of primary cancer operation (within 2 years), age (within 5 years), type of surgery, tumour histology, oestrogen receptor (ER) status and disease free interval by time of FG (Table 1). Further cancer variables were compared between the two populations to ensure homogeneity (Table 2 & 3). Similar to the Petit et al case-controlled series³⁴, the selected control patient had a disease-free period (*extrapolated* time A'-B') at least as long as the interval between oncologic surgery and FG procedure (time A-B) of the corresponding study patient (Figure 1). If the matched control had a recurrence prior to the end of this estimated time interval, then that patient was excluded, and another appropriate control was selected. The primary

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endpoint of the study was tumour recurrence, where the type and date were noted. The secondary endpoint was breast cancer-related death.

Data for women with recurrent disease prior to FG intervention (35) were collected for descriptive purposes and not included in the case-matched series, but were separately analysed. Patients treated for breast cancer at NBI were followed up annually with clinical examination and mammography for 2-5 years, depending on primary pathology.

Figure 1.

Fat grafting was performed according to the Coleman technique^{7,16} without stem cell enhancement. Tumescence included 150mg laevobupivacaine in 1L 0.9% Normal Saline with 1: 1 000 000 adrenaline injected with a blunt cannula. Donor site selection was dependent on surgeon and patient preference, but was most

commonly the abdomen and upper thigh area. Fat was injected in thin strips. Some patients had more than one FG procedure with a mean of 1.28 per patient (range 1-4).

A systematic literature review included all studies with adequate descriptions of oncologic events and follow up, reporting on patients treated for breast cancer with subsequent fat grafting. Patients who had a recurrent event prior to FG intervention were excluded.

Statistical method

The difference in prognostic variables between study and control groups was assessed using the chi-squared test. The main outcomes were recurrent oncological events in terms of local, regional and distant recurrences and death. Log Rank Kaplan-Meyer curves were used to calculate disease free survival (DFS).

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Any event occurring simultaneously with a local relapse, such as a regional recurrence or synchronous metastases, was counted as a single event. In case of no events, the endpoint of the study was censored at the last follow-up. Statistical significance was considered at a probability of $p < 0.05$. The impact of FG on risk of a recurrence was evaluated using the multivariate Cox proportion hazard regression model and expressed as hazard ratio (HR) with 95% confidence intervals (CIs).

Tables 1-4

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Results

Patient characteristics and follow-up are presented in Tables 1-4. The majority of patients in both the FG and control groups were treated by mastectomy; of these, almost 50% were skin sparing or nipple sparing mastectomies. Although the breast conserving surgery (BCS)

rate in the FG group was 16.6% (35 patients), this included only one patient with DCIS (0.7%) (Table 3).

Tables 2 & 3 show the distribution of non-matched variables in the FG and control groups for invasive cancer and DCIS. The populations are considered well matched, with Her-2 status ($p=0.013$) and Herceptin treatment ($p=0.001$) being the only different variables. This was most likely due to a large quantity of missing data in both groups as a result of non-routine Her-2 testing prior to 2005.

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Outcome analysis

The cumulative incidence of local recurrence in the FG and control groups was 0.95% and 1.90% respectively ($p=0.744$) (Tables 5&6).

The locoregional recurrence (LRR) was equal between the FG and control groups (4/211, 1.9% and 8/422, 1.9%), at 0.7% per year (Table 6).

The characteristics of 4 women experiencing LRR events after the FG intervention are shown in Table 7.

Tables 5-7, Figures 2-3

Locoregional recurrence or distant metastases *prior* to fat grafting, (n=35)

The mean age was of patients in this cohort was 42 years (range 27-64) and mean follow-up after oncological surgery and fat grafting was 170 months and 30 months respectively, with mean time to baseline 140 months. There were 33 invasive cancers, and two cases of DCIS. Eleven patients (31%) initially had a mastectomy, but all patients eventually underwent mastectomy following a recurrence, so all patient had a mastectomy prior to fat grafting. Table 8 demonstrates the recurrent events prior to FG in this subgroup. Despite 40% of these patients having a previous ipsilateral local recurrence, no patient suffered further local event following FG

procedure. Three patients developed further non-local recurrent events (8.6%). One patient developed a palpable supraclavicular node 20 months after the FG intervention, one patient developed a distant metastasis 6 months after FG and one patient developed a new contralateral breast cancer.

Table 8

Comparison with Case-Controlled Series in the Literature

Table 9 compares this study series and the Petit³⁴ series. There were a significantly greater proportion of patients undergoing BCS in the Petit series compared to the current series (39% v 17%).

Table 9

Systematic Review of Fat Grafting Case Series after Breast Cancer

Tables 10a & 10b

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Discussion

The results of this study show no significant association between fat grafting and disease recurrence in women previously treated for breast cancer. This was the case both for LRR (0.7% per year for both FG and control groups) and distant metastases (1.2% per year vs 0.9% per year for FG and control groups respectively). These rates are lower than those reported in the only other case-controlled series³⁸, where LRR was 1.9% per year in FG, 1.7% per year in controls and distant metastases 1.9% per year in both groups. This difference may be explained by the larger number of breast conserving surgery cases in their series.

Fat grafting after previous breast conserving surgery for breast cancer may be the best model for understanding the interaction between the fat graft and breast parenchyma in addressing the safety of fat grafting³⁷. In our series, most of the indications for fat grafting

were in post-mastectomy patients, although half of these patients received a skin-sparing or nipple-sparing mastectomy. For breast conserving cases, the LRR was higher in the FG group compared to controls although not significantly so (2.1% vs 1.1% per year, $p = 0.533$). In the Petit multicenter series³⁰, comparable cumulative incidence curves were observed when BCS and mastectomy patients were analyzed separately. In their case-controlled series, Petit et al³⁸ reported a LRR of 2.2% per year in patients who had FG after BCS, which was not significantly different from controls.

For the mastectomy group in our series, there was no significant difference in LRR amongst FG and control groups; 0.4% per year vs 0.6% per year ($p=0.848$). Riggio²⁸ reported a LRR of 0.88% per year, Rigotti et al²⁶ a rate of 0.75% per year and Petit et al³⁸ a LRR of 1.4% per year for FG after mastectomy (Table 11b).

Unlike the Petit series^{30,38,39}, we observed no recurrences amongst patients treated with FG following surgery for DCIS. This could be

due to a number of factors. There was a long disease-free interval between primary oncologic surgery and FG in these patients (54 months), which is the critical window period for recurrences to occur. This may therefore be a select low risk group, although, a relatively short follow-up after FG (mean 32 months) was observed. Breast conserving surgery for DCIS was low for both the FG and control arms (0.5% & 1.7% respectively), although the rate of skin-sparing mastectomy (SSM) was 57.6% for FG and 49.0% for the control arms. In the Petit et al DCIS series³⁹, 6 recurrences occurred in FG patients and 3 in controls (5 year cumulative incidence 18% & 3% respectively). All recurrences in both groups had either BCS or some form of skin sparing mastectomy. Furthermore, the mean time interval between oncologic surgery and FG in their series was relatively short, at 25 months, and mean time to recurrence from FG was 12 months. The rate of positive or close margins in the FG group was 42% and 22% in the controls (p 0.38) and there was no specific information provided regarding re-excisions or adjuvant radiotherapy for these patients. Margin control and early FG intervention could be

factors in the high LRR observed in the FG group in this series involving DCIS patients³⁹. In our series, all margins were clear.

In the Petit (DCIS) series³⁹, the likelihood of a recurrence was greatest if fat grafting occurred within 24 months of the primary oncological event. It is possible that early fat grafting may be associated with a greater risk of recurrence if performed in women with more risk factors for it. In this respect, our series may represent a low risk group. Ihrai et al²⁵ suggest a 36 month interval between primary surgery and fat grafting, and Riggio et al²⁸ 55 months. The mean time to fat grafting in our series and Petit et al³⁸ was similar (Table 10). The mean time to recurrence in the Petit DCIS series³⁹ was 12 months (range 5-24 months) and in the current series 50 months for invasive cancer (range 41-58 months).

There are limitations in interpreting individual studies reported in the literature. They are heterogeneous, retrospective, non-matched and many include cosmetic breast patients without exposure to breast

cancer. With respect to oncologic safety, it is important to focus on breast cancer patients only^{35,36}. Case-matching supports the validity of the results^{30,38}. Confounding variables such as resection margins, cancer histology, and receptor status can also directly influence the outcome. The FG and control groups in the current study are well matched (Tables 1-3).

However, case series may yield helpful information, if confounding variables are controlled for. Table 11a&b give a summary of studies presenting data for patients previously treated for invasive breast cancer or DCIS with FG. Our review of 1573 patients reported in the literature shows no evidence of an increased risk of breast cancer recurrence following fat grafting in terms of local (0.95% per year) or distant metastases (1.01% per year). Reassuringly, most series report both local and metastatic event rates between 1-2% per year. Petit et al³⁹ found a LRR of 3.2% per year in their case-controlled DCIS study which increased their overall LRR.

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Fat grafting is not a new technique, and has been exposed to criticism and controversy throughout its evolution^{5, 42-47}. In 1987, a position paper released by the American Society of Plastic and Reconstructive Surgeons (ASPRS) Ad-Hoc Committee on New Procedures was 'unanimous in deploring the use of fat injection in breast augmentation' as its radiological sequelae would compromise the detection of breast cancer on surveillance mammography⁴⁸. This issue has largely been resolved, as microcalcifications related to fat necrosis may be diagnosed after any type of breast cancer surgery and can be confidently distinguished from suspicious calcifications^{21,22}. Despite the 'veil of silence' the ASPRS paper imposed, many surgeons were continuing to report on the use of FG for breast augmentation and reconstruction^{5-9,12,14-16,18,21,23}. Coleman attempted to resolve these fears, by advocating meticulous planning, 'atraumatic liposuction', centrifugation and graft placement to provide 'pure, intact parcels of fat' to encourage integration and long-term graft survival^{1,6,7,10,11,15,16}. He also became interested in the mechanisms of fat graft survival and stability, and postulated whether

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adipose-derived stems cells in the lipoaspirate may be involved, given their regenerative effect in experimental research and in 'replenishing natural tissues'^{7,16}. Consequently, in a cautious public statement, both the American Society of Plastic Surgeons and American Society of Aesthetic Plastic Surgeons (ASPS and ASAPS) issued statements in 2007 that 'strongly support the ongoing research efforts that will establish the safety and efficacy of the procedure'^{13,49,50}. [Similarly, the French Society of Plastic, Reconstructive and Aesthetic Surgeons (SOFCPRE)^{51,52} and the UK NICE group⁵⁴ have issued caution with regard to FG after breast cancer. These societies acknowledged the lack of high level evidence in the literature demonstrating a link between fat grafting and breast cancer relapse. Based on recent reviews, the French have updated their recommendations⁵², and a phase III multicentre randomized, controlled trial is currently taking place in France with the goal of investigating this issue⁵⁴.

Despite some concerns, it seems that FG is increasingly popular in breast surgery^{4,55}. In a questionnaire to UK surgeons, 69% (48/70) of plastic surgeons and 11% (17/158) of breast surgeons are utilizing the technique (Ref). Most attitudes were positive with over 60% surgeons agreeing that the benefits of FG outweighed the risks. Similarly, the American College of Plastic Surgeons has reported 62% of their members regularly use FG for reconstructive breast surgery⁴.

Fat is a metabolically active tissue consisting of a heterogeneous cell population secreting cytokines, hormones and growth factors⁵⁶⁻⁵⁹. A fat graft specimen contains mature adipocytes and preadipocytes, also known as adipose derived stem cells (ASC) ^{56,58,59}. ASCs have considerable angiogenic and antiapoptotic features and constitute 10% of the cell population, however graft survival is largely dependent on them given their huge proliferative contribution^{14,40,57-59}. Adipocytes and ASCs release cytokines ('adipokines') to communicate with resident tissue for stimulating angiogenesis,

reducing apoptosis, and modulating the immune response during tissue repair^{57,58,60,61}. The initial apprehension regarding FG and cancer resurgence came from obesity studies observing altered adipokine signaling in resident adipocytes that could facilitate cancer initiation and progression^{62,62}. These 'adipokines' have been extensively studied, and include; leptin, adiponectin, resistin, metalloproteinase 11, hepatocyte growth factor (HGF), adipocyte-derived collagen VI, Interleukins, TGR-a, TGF-b, and VEGF^{57,58,60-64}. Concern was raised that placement of these physiochemical factors at the site of a previous tumour microenvironment, may fuel the 'tumour-stromal interaction' through autocrine, paracrine and exocrine/endocrine that can result in tumour recurrence^{14,56-61,64-66} and metastasis^{58,67,68}. The direct influence of ASCs on residual or dormant tumour cells has been investigated with variable results. Co-cultures of human epithelial and adenocarcinoma tumour cell lines with MSCs in murine *in vitro*^{58,67,69} and *in vivo*^{66,68,69} models resulted in increased tumour cell viability, enhanced proliferation and reduced apoptosis of tumour cells. However, other groups found ASCs

capable of inhibiting proliferation, down-regulating cell signalling and abrogating tumour progression⁷⁰⁻⁷². Zimmerman et al found that ASCs did not activate dormant cancer cells, but promoted residual active cancer cells into tumour growth and expansion. The group suggested that reconstructive therapy utilizing ASC-augmented whole fat should be postponed until there is no evidence of active disease and clear margins⁶⁹.

Local and systemic recurrence of breast cancer is observed at similar rates in our case controlled series, and may occur irrespective of the fat grafting intervention. Local recurrence and metastasis may be a manifestation of residual cancer stem cell activation and dissemination for reasons largely unknown, although the underlying molecular microenvironment may play a role^{56,60,73}. Long term dormancy of cancer cells is a known phenomenon, but it is particularly evident in breast cancer patients in whom, even after 8 years of disease-free survival, a significant rate of late recurrence has been observed^{74,75}. This suggests that many cancer types can

persist as 'minimal residual disease' and putative cancer stem cells can remain dormant for years, but are reactivated by still unknown mechanisms, often leading to rapid disease progression after a latent period^{76,77}.

The reason for the discordance between in vitro studies and clinical observations could be due to a number of reasons. The initial question of adipocytes secreting adipokines that could fuel cancer cell progression came from hypertrophied resident adipocytes in obese patients. This is different to the clinical setting regarding fat grafting, where normal adipocytes are transplanted into the breast in the mastectomy or prepectoral plane, and not into the parenchyma. In vitro and in vivo studies involve highly controlled microenvironments in constructed scaffolds or immunosuppressed animal models with many variables accounted for or eliminated. Furthermore, these studies have variable results with some adipokines stimulating cancer cell growth, and other suppressing it. The human subject is more complex, and there may be additional

unrecognized pathways that abrogate adipokine signaling after fat grafting into a previous cancer environment in the clinical setting.

As with any study, there are limitations with the current paper. This type of study would be difficult to design and run prospectively. The retrospective nature of this study is one drawback, but it facilitates adequate case matching. Case matching prospectively would be difficult due to the 5 variables requiring matching from entry into the study. If either of the two control patients had a recurrence prior to time B' (fat grafting intervention of the study patient), then they would need to be excluded, requiring that study patient also to be excluded, or two retrospective matches made at this point, rendering the study non-prospective in nature. The study is non-randomized, although randomization would be difficult due to the lack of a comparable alternative to fat grafting, and patients tend to know the benefits of the procedure and will ask for it, particularly if it enhances their reconstructive option. The ideal candidate to study the oncological safety of fat grafting is in the breast conserving patient. In our study,

there were low numbers of patients treated with BCS – around 16% in each arm (35 patients in FG group), however amongst our mastectomy group, around half of patients received either a skin-sparing or nipple-sparing mastectomy, with the small potential to leave breast tissue remaining (quote paper). There is controversy surrounding FG in patients with previous DCIS, particularly those treated with BCS, created by the Petit et al DCIS paper³⁹. Our study cannot reliably contribute evidence to this question, as we had low numbers of DCIS patients treated with BCS.

However, despite these warnings from basic science, **and our study's limitations**, the current clinical study shows no evidence of increased oncological risk associated with fat grafting in women previously treated for breast cancer. This evidence should be interpreted with other similarly case controlled studies in establishing safe indications for fat grafting in this setting.

References

1. Coleman, SR, Saboeiro AP. Fat grafting to the breast revisited: safety and efficacy. *Plast Reconstr Surg.* 2007;119(3): 775-785
2. Serra-Renom JM, Muñoz-Olmo JL, Serra-Mestre JM. Fat grafting in postmastectomy breast reconstruction with expanders and prostheses in patients who have received radiotherapy: formation of new subcutaneous tissue. *Plast Reconstr Surg.* 2010;125(1):12-18
3. Rigotti G, Marchi A, Galie M, Baroni G, Benati D, Krampera M., ... & Sbarbati A. Clinical treatment of radiotherapy tissue damage by lipoaspirate transplant: a healing process mediated by adipose-derived adult stem cells. *Plast Reconstr Surg.* 2007;119(5):1409-1422
4. Kling RE, Mehrara BJ, Pusic AL, Young VL, Hume KM, Crotty CA, & Rubin JP. Trends in autologous fat grafting to the breast: a national

survey of the American Society of Plastic Surgeons. *Plast Reconstr Surg.* 2013;132(1):35-46

5. Bircoll M. Cosmetic breast augmentation utilizing autologous fat and liposuction techniques. *Plast Reconstr Surg.* 1987;79(2):267-271

6. Coleman SR. Structural fat grafting. *Aesthetic Surg J* 1998; 18:386-8

7. Coleman SR. Structural fat grafts: the ideal filler?. *Clin Plast Surg.* 2001; 28(1): 111-119

8. Missana MC, Laurent I, Barreau L, Balleyguier C. Autologous fat transfer in reconstructive breast surgery: indications, technique and results. *Eur J Surg Oncol (EJSO).* 2007;33(6):685-690

9. Delay E, Garson S, Tousson G, Sinna R. Fat injection to the breast: technique, results, and indications based on 880 procedures over 10 years. *Aesth Surg J.* 2009;29(5):360-376.

10. Coleman SR. Facial recontouring with lipostructure. *Clin Plast Surg.* 1997; 24(2): 347-367

11. Coleman SR, Carraway JH. Hand rejuvenation with structural fat grafting. *Plast Reconstr Surg.* 2002; 110(7): 1731-1744

12. Chan CW, McCulley SJ, Macmillan RD. Autologous fat transfer—a review of the literature with a focus on breast cancer surgery. *J Plast Reconstr Aesth Surg.* 2008;61(12):1438-1448

13. Gutowski KA, Force AFGT. Current applications and safety of autologous fat grafts: a report of the ASPS fat graft task force. *Plast Reconstr Surg.* 2009;124(1): 272-280

14. Rigotti G, Marchi A, Sbarbati A. Adipose-derived mesenchymal stem cells: past, present, and future. *Aesth Plast Surg.* 2009;33(3):271-273.

15. Coleman SR. Long-term survival of fat transplants: controlled demonstrations. *Aesth Plast Surg.* 1995;19(5): 421-425

16. Coleman SR. Structural fat grafting: More than a permanent filler. *Plast Reconstr Surg.* 2006;118.3S:108S-120S

17. Illouz YG. Body contouring by lipolysis: a 5-year experience with over 3000 cases. *Plast Reconstr Surg.* 1983;72:591-7

18. Illouz YG, Sterodimas A. Autologous fat transplantation to the breast: a personal technique with 25 years of experience. *Aesth Plast Surg.* 2005;33(5): 706-715

19. Beck M, Amar O, Bodin F, Lutz JC, Lehmann S, Bruant-Rodier C.

Evaluation of breast lipofilling after sequelae of conservative treatment for cancer. *Eur J Plast Surg.* 2012;35(3):221-228

20. Claro F, Figueiredo JCA, Zampar AG, Pinto-Neto AM. (2012).

Applicability and safety of autologous fat for reconstruction of the breast. *Br J Surg.* 2012;99(6):768-780

21. Pulagam SR, Poulton T, Mamounas EP. Long-Term Clinical and

Radiologic Results with Autologous Fat Transplantation for Breast Augmentation: Case Reports and Review of the Literature. *Breast J.* 2006;12(1):63-65

22. Veber M, Tourasse C, Toussoun G, Moutran M, Mojallal A, Delay

E. Radiographic findings after breast augmentation by autologous fat transfer. *Plast Reconstr Surg.* 2001;127(3):1289-1299

23. Spear SL, Wilson HB, Lockwood MD. Fat injection to correct contour deformities in the reconstructed breast. *Plast Reconstr Surg.* 2005;116(5):1300-1305

24. Largo RD, Tchang LA, Mele V, Scherberich A, Harder Y, Wettstein R, Schaefer DJ. Efficacy, safety and complications of autologous fat grafting to healthy breast tissue: A systematic review. *J Plast Reconstr Aesth Surg.* 2013;67(4):437-448

25. Ihrai T, Georgiou C, Machiavello JC, Chignon-Sicard B, Figl A, Raoust I, Bourgeon Y, Fouche Y, Flipo B. Autologous fat grafting and breast cancer recurrences: retrospective analysis of a series of 100 procedures in 64 patients. *J Plast Surg Hand Surg.* 2013;47:273-275

26. Rigotti G, Marchi A, Stringhini P, Baroni G, Galiè M, Molino A M, Mercanti A, Micciolo R, Sbarbati A. Determining the oncological risk of autologous lipoaspirate grafting for post-mastectomy breast reconstruction. *Aesth Plastic Surg.* 2010;34(4):475-480

27. Rietjens MF, De Lorenzi F, Rossetto F, Brenelli A, Manconi S, Martella M, Intra et al. Safety of fat grafting in secondary breast reconstruction after cancer. *J Plas Reconstr Aesth Surg*. 2011;64(4):477-483

28. Riggio E, Bordoni D, Nava MB. Oncologic Surveillance of Breast Cancer Patients After Lipofilling. *Aesth Plastic Surg*. 2013;37(4):728-735

29. Losken A, Pinell XA, Sikoro K, Yezhelyev MV, Anderson E, Carlson GW. Autologous fat grafting in secondary breast reconstruction. *Ann Plastic Surg*. 2011;66(5):518-522

30. Petit JY, Lohsiriwat V, Clough KB, Sarfati I, Ihrai T, Rietjens M, ... Delay E. The oncologic outcome and immediate surgical complications of lipofilling in breast cancer patients: a multicenter

study—Milan-Paris-Lyon experience of 646 lipofilling procedures.

Plast Reconstr Surg. 2011;128(2):341-346

31. Yoshimura K, Sato K, Aoi N, Kurita M, Hirohi T, Harii K. Cell-assisted lipotransfer for cosmetic breast augmentation: supportive use of adipose-derived stem/stromal cells. Aesth Plastic Surg.

2008;32(1):48-55

32. Khouri RK, Eisenmann-Klein M, Cardoso E, Clooney B, Cooley BC, Kacher D, Gombos E, et al. Brava and autologous fat transfer is a safe and effective breast augmentation alternative: results of a 6-year, 81 patient, prospective multicentre study. Plast Reconstr Surg

2012;129:1173-87

33. Rosing JH, Wong G, Wong MS, Sahar D, Stevenson TR, Pu LL. Autologous fat grafting for primary breast augmentation: a systematic review. Aesth Plastic Surg.

2011;35(5):882-890

34. Saint-Cyr M, Rojas K, Colohan S, Brown S. The role of fat grafting in reconstructive and cosmetic breast surgery: a review of the literature. *J Reconstr Microsurg*. 2012; 28(2):99-110

35. Krastev TK, Jonasse Y, Kon M. Oncological safety of autologous lipoaspirate grafting in breast cancer patients: a systematic review. *Ann Surg Oncol*. 2013; 2:111-119

36. Petit, JY, Clough KB, Sarfati I, Lohsiriwat V, de Lorenzi F, Rietjens M. Lipofilling in breast cancer patients: from surgical technique to oncologic point of view. *Plast Reconstr Surg*. 2010;126(5):262e-263e

37. De Lorenzi F, Lohsiriwat V, Petit JY. *In Response To*: Rigotti G, Marchi A, Stringhini P, et al. Determining the Oncological Risk of Autologous Lipoaspirate Grafting for Post-Mastectomy Breast Reconstruction. *Aesth Plast Surg* 2010; 34: 475. *Aesth Plast Surg* 2011;35(1):132-133

38. Petit JY, Botteri E, Lohsiriwat V, Rietjens M, De Lorenzi F, Garusi C, Rossetto F, et al. Locoregional recurrence risk after lipofilling in breast cancer patients. *Ann Oncol.* 2012; 23(3): 582-588

39. Petit JY, Rietjens M, Botteri E, Rotmensz N, Bertolini F, Curigliano G, Rey P, et al. Evaluation of fat grafting safety in patients with intra epithelial neoplasia: a matched-cohort study. *Ann Oncol.* 2013;24(6):1479-1484

40. Pérez-Cano R, Vranckx JJ, Lasso JM, Calabrese C, Merck B, Milstein AM., ... & Weiler-Mithoff EM. Prospective trial of adipose-derived regenerative cell (ADRC)-enriched fat grafting for partial mastectomy defects: the RESTORE-2 trial. *Eur J Surgical Oncol (EJSO).* 2012; 38(5):382-389

41. Brenelli F, Rietjens M, De Lorenzi F, Pinto-Neto A, Rossetto F, Martella S, ... & Barbalho D. Oncological Safety of Autologous Fat

Grafting after Breast Conservative Treatment: A Prospective
Evaluation. Breast J. 2014;20(2):159-165

42. Neuber F. Fettransplantation. Chir Kongr Verhandl Dsch Gesellch
Chir. 1893;22:66

43. Czerny A. Plastischer Ersatz der Brustdruse durch ein Lipoma.
Chir Kongr Verhandl. 1895;216:2

44. Hartrampf Jr CR, Bennett GK. Autologous fat from liposuction for
breast augmentation. Plast Reconstr Surg. 1987;80:646-7

45. Peer LA. Loss of weight and volume in human fat grafts. Plast
Reconstr Surg. 1950;5:217

46. Letter: Ousterhout D. Plast Reconstr Surg. 1987;80:868

47. Perrot P, Rousseau J, Bouffaut AL, Rédini F, Cassagnau E, Deschaseaux F, Heymann MF et al. Safety concern between autologous fat graft, mesenchymal stem cell and osteosarcoma recurrence. PLoS One. 2010; 5(6):e10999

48. ASPRS Ad-Hoc Committee on New Procedures. Report on autologous fat transplantation. Plast Surg Nurs. 1987;7:140-1

49. American Society of Plastic Surgeons. Plastic surgery societies issue caution on fat grafting for breast augmentation 2007
<http://www.plasticsurgery.org/media/Patient-Safety-Press-Kit-Index.cfm> - site now superseded by 2011 statement

50. American Society of Plastic Surgeons and American Society of Aesthetic Plastic Surgeons (ASPS & ASAPS Position Statements, May 2011). http://www.plasticsurgery.org/Documents/medical-professionals/health-policy/guiding-principles/Joint_Stem_cells.pdf

51. SFdCPre Esthetique. FAQ – recommandation concernant les injections de graisse dans le sein 2008

52. SOFCPRE. Recommendations concernant les injections de graisse dans le sein February 2012

53. NICE Guidelines; Interventional procedure overview of breast reconstruction using lipomodelling after breast cancer treatment.

26/28 October 2010

<http://www.nice.org.uk/nicemedia/live/12575/53761/53761.pdf>

54. GRATSEC. Adipose tissue transfer for moderate breast cancer conservative treatment sequella (GRATSEC). NCT01035268.

Clinical Trials.gov.GRATSEC NCT01035268.

55. Skillman J, Hardwicke J, Whisker L, England D. Attitudes of UK breast and plastic surgeons to lipomodelling in breast surgery.

Breast. 2013;22(6):1200-1204

56. Lohsiriwat V, Curigliano G, Rietjens M, Goldhirsch A, Petit JY. Autologous fat transplantation in patients with breast cancer: “silencing” or “fueling” cancer recurrence?. *Breast*. 2011; 20(4):351-357

57. Vona-Davis L, Rose DP. Adipokines as endocrine, paracrine, and autocrine factors in breast cancer risk and progression. *Endocr Rel Cancer*. 2007;14(2):189-206

58. Manabe Y, Toda S, Miyazaki K, Sugihara H. Mature adipocytes, but not preadipocytes, promote the growth of breast carcinoma cells in collagen gel matrix culture through cancer–stromal cell interactions. *J Pathol*. 2003; 201(2):221-228

59. Pearl RA, Leedham SJ, Pacifico MD. The safety of autologous fat transfer in breast cancer: lessons from stem cell biology. *J Plast Reconstr Aesth Surg*. 2012;65(3) 283-288

60. Llanos AA, Dumitrescu RG, Marian C, Makambi KH, Spear SL, Kallakury BV, ... & Shields PG. Adipokines in plasma and breast tissues: associations with breast cancer risk factors. *Cancer Epidemiol Biomarkers & Prevention*. 2012;21(10):1745-1755

61. Motrescu ER, Rio MC. Cancer cells, adipocytes and matrix metalloproteinase 11: a vicious tumor progression cycle. *Biolog Chem*. 2008;389(8):1037-1041

62. Bertolini F, et al. Adipose tissue cells, lipotransfer and cancer: a challenge for scientists, oncologists and surgeons. *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*. 2012;1826.1: 209-214

63. Schäffler A, Schölmerich J, Buechler C. Mechanisms of disease: adipokines and breast cancer—endocrine and paracrine mechanisms that connect adiposity and breast cancer. *Nat Clin Pract Endocrinol & Met*. 2007; 3(4):345-354

64. Chandler EM, Seo BR, Califano JP, Andresen Eguiluz RC, Lee JC, Yoon CJ, Tims DT et al. Implanted adipose progenitor cells as physicochemical regulators of breast cancer. *Proc Natl Acad Sciences*. 2012;109(25): 9786-9791

65. Krumboeck A, Giovanoli P, Plock JA. Fat grafting and stem cell enhanced fat grafting to the breast under oncological aspects—Recommendations for patient selection. *Breast*. 2013;22(5):579-584

66. Zhang Y, Daquinag A, Traktuev DO, Amaya-Manzanares F, Simmons PJ, March KL, ... & Kolonin MG. White adipose tissue cells are recruited by experimental tumors and promote cancer progression in mouse models. *Cancer Res*. 2009;69(12):5259-5266

67. Martin-Padura I, Gregato G, Marighetti P, Mancuso P, Calleri A, Corsini C, Pruneri G et al. The white adipose tissue used in

lipotransfer procedures is a rich reservoir of CD34+ progenitors able to promote cancer progression. *Cancer Res.* 2012;72(1):325-334

68. Yu J, Jun E, Bae Y, Jung J. Mesenchymal stem cells derived from human adipose tissues favour tumour cell growth in vivo. *Stem cells Dev.* 2008;17:463-73

69. Zimmerlin L, Donnenberg AD, Rubin JP, Basse P, Landreneau RJ, Donnenberg VS. Regenerative therapy and cancer: in vitro and in vivo studies of the interaction between adipose-derived stem cells and breast cancer cells from clinical isolates. *Tissue Eng Part A.* 2011;17:93-106

70. Otsu K, Das S, Houser SD, Quadri SK, Battacharya J. Concentration dependent inhibition of angiogenesis by mesenchymal stem cells. *Blood.* 2009;113:4197-205

71. Sun B, Roh KH, Park JR et al. Therapeutic potential of

mesenchymal stromal cells in a mouse breast cancer metastasis model. *Cytotherapy*. 2009;11: 289-98

72. Rahimi N, Tremblay E, McAdam L, Roberts A, Elliott B. Autocrine secretion of TGF- β 1 and TGF- β 2 by pre-adipocytes and adipocytes: a potent negative regulator of adipocyte differentiation and proliferation of mammary carcinoma cells. *In Vitro Cell & Dev Biol-Animal*. 1998; 34(5):412-420

73. Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc. Natl. Acad. Sci. USA*. 2003;100:3983–3988

74. Mansi JL; Gogas H, Bliss JM, Gazet JC, Berger U, Coombes RC. Outcome of primary breast-cancer patients with micrometastases: A long-term follow-up study. *Lancet* .1999;354:197–202

75. Braun S, Vogl FD, Naume B, Janni W, Osborne MP, Coombes RC, Schlink G, Diel IJ, Gerber B, Gebauer G, et al. A pooled analysis of bone marrow micrometastasis in breast cancer. *N. Engl. J. Med.* 2005;353:793–802

76. Trumpp A, Wiestler OD. Mechanisms of disease: cancer stem cells—targeting the evil twin. *Nat Clin Prac Oncol.* 2008;5(6):337-347

77. Banys M, Krawczyk N, Fehm T. The Role and Clinical Relevance of Disseminated Tumor Cells in Breast Cancer. *Cancers.* 2014; 6(1): 143-152

