

Effects of opioids on immune function in patients with cancer pain: from bench to bedside

Abstract

In patients with cancer, opioids are principally used for the management of acute surgical and chronic cancer-related pain. Opioids have many non-analgesic effects, including direct and indirect effects on cancer cells and on anti-tumour immunity. Direct effects on immune cells are elicited via opioid and non-opioid Toll-like receptors, and indirect effects *via* the sympathetic nervous system and hypothalamic-pituitary-adrenal axis. Opioids can also decrease/alter immune cell infiltration into the tumour microenvironment. Animal models have shown that this is not a class effect: morphine and fentanyl suppress natural killer (NK) cell cytotoxicity, thereby increasing tumour burden; buprenorphine does not affect NK cell cytotoxicity, whereas tramadol increases NK cell cytotoxicity, thereby reducing metastasis. In healthy individuals, morphine suppresses and fentanyl enhances NK cell cytotoxicity. Although clinical outcomes were not determined, fentanyl has been shown to decrease NK cell cytotoxicity whereas tramadol increases cytotoxicity in patients undergoing surgery. Meta-analyses of opioid-sparing surgical studies have reported an association between improved recurrence-free and/or overall survival with regional/neuraxial anaesthesia compared with systemic opioids. In people receiving opioids for non-surgical cancer-related pain, morphine has variable effects on immunity; clinical outcomes were not assessed. Although there is a potential association between strong systemic opioids and shorter survival in people with cancer and a prognosis of months to years, studies have not been designed to primarily assess survival therefore causality cannot be apportioned. Pain is also immunosuppressive and adequate analgesia is important. The use of opioids for cancer-related pain continues to be recommended, until definitive data is available on the effects of opioids on clinical outcomes in specific patient groups becomes available.

Opioids and the immune system

Opioids are a diverse range of drugs that act on opioids receptors. In patients with cancer, opioids are principally used in the long-term management of cancer-related pain as well as in the shorter term for cancer surgery pain. They can produce a range of undesired effects

(Boland *et al.*, 2013), including effects on the immune system (Grace *et al.*, 2015; Ramaswamy *et al.*, 2016; Wigmore *et al.*, 2016).

In patients with, or developing, cancer, the immune system has a crucial role in controlling and potentially eradicating cancer cells. Many immune cells are involved in anti-tumour immunity including natural killer (NK) cells, T cells, mast cells, dendritic cells and macrophages, as well as cytokines and chemokines (Table 1) (Boland *et al.*, 2014b; Liang *et al.*, 2016; Maghazachi, 2010; Mittal *et al.*, 2014; Nguyen *et al.*, 2014). It has been suggested that NK cell activity is a critical endpoint in the immunotoxicological evaluation of pharmaceuticals (European Medicines Evaluation Agency. Note for Guidance on Repeated Dose Toxicity. CPMP/SWP/1042/99. EMEA: London, 2000) and a consensus statement for opioids for chronic pain in elderly people proposes that the effects of opioids on immune function should be considered (Pergolizzi *et al.*, 2008).

Table 1: Role and function of the main immune cells.

Cell	Role	Function	Arm
Dendritic cell	Antigen presentation	Presentation of antigenic peptides in the context of MHC class I and II molecules and the delivery of essential co-stimulatory molecules	Innate
Natural killer cell	Anti-tumour Anti-viral	Release of cytotoxic molecules (granzymes, perforin)	Innate
Neutrophil	Anti-bacterial / fungal	Phagocytosis and oxidative burst	Innate
Monocyte-Macrophage lineage	Anti-bacterial / fungal	Phagocytosis and oxidative burst	Innate
CD4⁺ T cell	Immune coordination / regulation	Regulating the activity of other immune cells	Adaptive
Regulatory T cell (CD4⁺, CD25⁺, FOXP3⁺, CD127^{low}, plus other markers)	Immune regulation/ suppression	Immune system modulation, maintaining tolerance to self-antigens, preventing autoimmune disease, potential barrier to the development of protective anti-tumour immunity	Adaptive
CD8⁺ T cell	Cytotoxicity	Eradication of virally-infected cells and cancer cells. Induction of apoptosis by i) release of cytotoxins (perforin, granzymes) ii) direct cell-cell contact	Adaptive
B cell	Antibody production	Antibody production	Adaptive

Adapted with permission from (Boland *et al.*, 2014b)

Given that patients with cancer are also at risk of infection, it is thus important that the anti-infection arm of the immune system to be maintained at an adequate level. There is evidence that opioids could increase the risk of infection in patients with cancer (Dublin *et al.*, 2011; Salimi *et al.*, 2013; Shao *et al.*, 2016; Suzuki *et al.*, 2013), as a retrospective study has shown that patients treated with morphine developed more infections than those treated with oxycodone (Suzuki *et al.*, 2013). Furthermore, the risk of infection risk has been shown to increase by 2% per 10 mg rise in the oral morphine equivalent daily dose; with no difference between opioids (Shao *et al.*, 2016).

Opioids can affect cancer via many mechanisms, including indirect effects on cancer growth via angiogenesis and host immunity (Afsharimani *et al.*, 2011; Gach *et al.*, 2011; Koodie *et al.*, 2014; Yamamizu *et al.*, 2015). Opioids might directly influence the growth of cancer cells as cancer tissue overexpresses *mu* opioid receptors (MOR) (Lennon *et al.*, 2012; Mathew *et al.*, 2011; Nguyen *et al.*, 2014; Singleton *et al.*, 2014; Zylla *et al.*, 2013). Overexpression of MOR on cancer cells in murine models has been shown to increase the growth of lung cancer (Lennon *et al.*, 2014), and decreasing MOR activity (using MOR small hairpin RNA mice or the opioid receptor antagonist methylnaltrexone) has been shown to reduce the growth of lung cancer (Mathew *et al.*, 2011). Furthermore, MOR overexpression has been associated with metastasis in patients with oesophageal and lung cancer (Singleton *et al.*, 2014; Zhang *et al.*, 2015). Morphine has been shown to have direct effects on cancer cells influencing their proliferation and survival via effects on tumour cell DNA cleavage, mitogen-activated protein kinase, Src, Gab-1, PI3 kinase, Akt and STAT3 signalling pathways (Lennon *et al.*, 2014; Lennon *et al.*, 2012; Mathew *et al.*, 2011).

The remainder of this review will concentrate on the cancer-related immune effects of opioids.

Mechanisms of immune effects of opioids

Numerous mechanisms underlying the influence of opioids on immune cells have been described (Figure 1). *In vivo*, opioids can modulate immune function by direct effects on immune cells, and via indirect effects which involve the central nervous system and its release of immune mediators (Al-Hashimi *et al.*, 2013; Borner *et al.*, 2008; Campana *et al.*, 2010).

For opioids to have direct effects on immune cells, immune cells must express opioid receptors, or opioids must be able to have effects via non-opioid receptors present on immune cells (e.g. Toll-like receptor-4). although *in vitro* morphine can directly interact with MORs on immune cells (Borner *et al.*, 2008), the presence of functional opioid receptors on immune cells, as determined using radioligand antibody binding studies, Western blot and polymerase chain reaction analysis continues to be disputed (Al-Hashimi *et al.*, 2016; Al-Hashimi *et al.*, 2013; Borner *et al.*, 2009; Campana *et al.*, 2010; Glattard *et al.*, 2010; Kraus, 2009; Langsdorf *et al.*, 2011; Williams *et al.*, 2007). One possibility for the discrepancy is that MOR expression depends upon immune cell activation - expression of MOR in T cells is non-constitutive (Borner *et al.*, 2008; Kraus *et al.*, 2001). In activated human T cells, MOR mRNA is increased to levels about 1% of those in neurons and can produce functional MORs in T cells (Borner *et al.*, 2007). Many extracellular signals, including the cytokines IL-1, IL-4, IL-6, TNF and IFN- γ , control MOR gene transcription and cell surface opioid receptor expression in immune cells (Kraus, 2009; Langsdorf *et al.*, 2011; Mohan *et al.*, 2010). Toll-like receptor-4 is involved in innate immune system activation. *In vitro* and *in silico* techniques have shown that many commonly used opioids activate this receptor and can have direct effects on innate immunity (Franchi *et al.*, 2012; Grace *et al.*, 2015; Hutchinson *et al.*, 2010).

There are two principal pathways via which the central immunosuppressive effects of morphine can be mediated; this is mostly based on animal data and might differ between species (Al-Hashimi *et al.*, 2013; Fecho *et al.*, 1996; Wang *et al.*, 2002). Acute morphine administration acts on the peri-aqueductal grey and sympathetic nervous system (which innervates lymphoid organs) to release immunosuppressive biological amines that can suppress NK cell cytotoxicity (Fecho *et al.*, 1993; Fecho *et al.*, 1996; Gomez-Flores *et al.*, 2000; Hernandez *et al.*, 1993; Irwin *et al.*, 1988; Lysle *et al.*, 1996; Weber *et al.*, 1989). An additional arm to the sympathetic nervous system pathway is the action of acute morphine/diamorphine via D1 receptors in the nucleus accumbens shell. This increases the release of neuropeptide Y from the sympathetic nervous system, which acts on peripheral Y1 receptors to inhibit splenic NK cell cytotoxicity (Saurer *et al.*, 2006a; Saurer *et al.*, 2009; Saurer *et al.*, 2006b). Prolonged use of opioids increases activity in the hypothalamic-pituitary-adrenal axis, thereby increasing the release of immunosuppressive glucocorticoids, and decreasing T cell function and NK cell cytotoxicity (Borman *et al.*, 2009; Hernandez *et al.*, 1993; Mellon *et al.*, 1998; Zhang *et al.*, 2011).

As well as having direct and indirect effects on immune cell activation, MOR activation can inhibit NK cell migration. Fewer NK cells are in the tumours of wild-type compared to MOR knockout mice (Boehncke *et al.*, 2011), morphine has been shown to reduce leukocyte migration in a murine lung tumour model (Koodie *et al.*, 2014), and NK cell infiltration into breast cancer tissue is decreased in women who have received more perioperative systemic opioids (Desmond *et al.*, 2015). For effective anti-tumour immunity, immune cells need to be in the tumour microenvironment, and a lack of immune cell infiltration could therefore be detrimental to anti-tumour immunity.

Not all immune cells are beneficial to the anti-tumour response; regulatory T cells can suppress the immune response (Takeuchi *et al.*, 2016). Meta-analyses have reported increased regulatory T cell infiltration into breast cancers is associated with a poorer survival (Shou *et al.*, 2016; Wang *et al.*, 2016). Regulatory T cells are increased in blood from patients with gastric cancer by *in vitro* morphine (Hou *et al.*, 2016), *in vitro* and *in vivo* by sufentanil and fentanyl in women undergoing breast cancer surgery (Gong *et al.*, 2014); these could inhibit anti-tumour immunity (Takeuchi *et al.*, 2016).

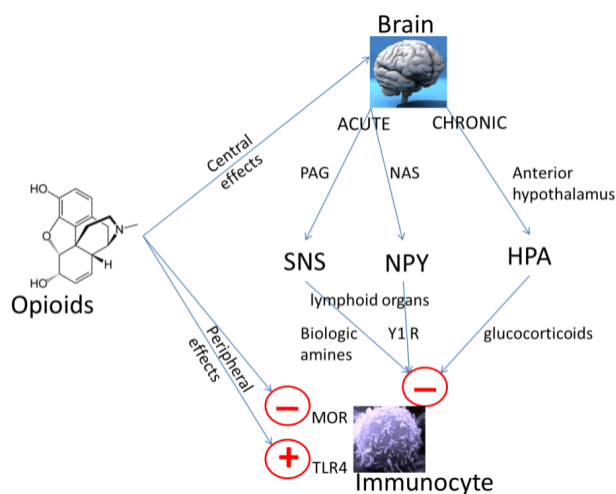


Figure 1: Peripheral and central mechanisms of opioid-induced immune suppression.

Opioids can have direct effects on immune cells which express appropriate receptors such as mu opioid receptors (MORs) and Toll-like receptor-4 (TLR-4). They can also have immunosuppressive effects via central mechanisms. Acute opioid administration enhances activity in the periaqueductal gray (PAG) matter which activates the sympathetic nervous system (SNS). The SNS innervates lymphoid organs, such as the spleen, and this activation induces the release of biological amines which suppress splenic lymphocyte proliferation and NK cell cytotoxicity (Fecho *et al.*, 1996; Irwin *et al.*, 1988). Second, prolonged use of opioids increases hypothalamic pituitary adrenal (HPA) axis activity and glucocorticoid production, which decrease NK cell cytotoxicity (Fecho *et al.*, 1996; Mellon *et al.*, 1998). Morphine can also act via D1 dopamine receptors in the nucleus accumbens

shell, increasing the release of neuropeptide Y (NPY) and reducing splenic NK cell cytotoxicity in rodent models (Saurer *et al.*, 2006b). Reproduced with permission from (Boland *et al.*, 2014b).

Animal studies: NK cells and tumour growth

NK cells are the principal immune cell involved in tumour immunosurveillance. Suppression of NK cell cytotoxicity has been demonstrated to correlate with increased tumour growth and metastasis in animal models (Franchi *et al.*, 2007; Gaspani *et al.*, 2002; Shavit *et al.*, 2004). The published literature generally reports that morphine (Franchi *et al.*, 2007; Weber *et al.*, 2006; Yeager *et al.*, 1995) and fentanyl (Franchi *et al.*, 2007; Martucci *et al.*, 2004; Shavit *et al.*, 2004) decrease NK cell cytotoxicity, that buprenorphine does not affect NK cell cytotoxicity (Franchi *et al.*, 2007; Martucci *et al.*, 2004) and that tramadol enhances it (Gaspani *et al.*, 2002; Sacerdote *et al.*, 2000) (Table 2). There may be different effects on NK cell cytotoxicity depending on doses, as in pigs low-dose morphine stimulated, and high-dose induced an initial increase followed by suppression (Borman *et al.*, 2009).

There are many studies describing how opioids can affect immunity, and these have mostly using acute administration of morphine. However, the duration of opioid administration (as well as the opioid itself) can lead to different effects on immune function (Liang *et al.*, 2016). The differential effects of opioids on different aspects of immune and how tolerance develops differently is illustrated in a series of murine experiments by Martucci *et al* (Martucci *et al.*, 2004). In these studies, single dose fentanyl only decreased lymphocyte proliferation, whereas chronic fentanyl reduced lymphocyte proliferation and also splenic NK cell cytotoxicity, IL-2 and IFN- γ production at 24 hours. NK cell cytotoxicity recovered by day 3, whereas lymphocyte proliferation, IL-2 and IFN- γ production recovered by day 7. After the 7 day infusion, the administration of twice the dose of fentanyl for 1 or 3 days to these mice did not affect any immune parameter, thereby suggesting that once immunological tolerance had developed, increasing the dose did not overcome it (Martucci *et al.*, 2004). Buprenorphine did not affect any of the measured parameters at any time point (Martucci *et al.*, 2004).

The effect of the timing of morphine administration on cancer progression has been studied in a murine model of breast adenocarcinoma. If given before the cancer had developed, morphine had no effect on cancer growth. However, once the cancer had developed, morphine, via the MOR, increased cancer progression and decreased survival (Nguyen *et al.*, 2014).

Several surgical studies have assessed how different opioids, and the timing of opioid administration, affect cancer progression. In a series of rodent experiments, (Franchi *et al.*, 2007) showed that morphine and fentanyl decreased NK cell cytotoxicity and increased MADB106 lung metastases, whereas buprenorphine did not. Surgery decreased NK cell cytotoxicity and increased MADB106 lung metastases. In the surgical model, perioperative buprenorphine reversed the suppressive effects of surgery on NK cell cytotoxicity and MADB106 lung metastases, whereas morphine and fentanyl did not (Franchi *et al.*, 2007). Decreased NK cell cytotoxicity correlated with high corticosterone levels, thereby suggesting the involvement of the hypothalamic-pituitary-adrenal axis (Franchi *et al.*, 2007). Fentanyl has been shown to suppress NK cell cytotoxicity in rats, and this suppression correlated to increased tumour load when fentanyl was administered close to the time of MADB106 tumour cell inoculation, but not when fentanyl was administered 6 hours before tumour inoculation (Shavit *et al.*, 2004). Thus, the acute administration of morphine and fentanyl around the time of tumour resection in animals could be detrimental to immune function and cancer outcomes.

In a series of rodent experiments, Gaspani *et al.* demonstrated that tramadol increased and morphine decreased NK cell cytotoxicity in non-operated rats (Gaspani *et al.*, 2002). Surgery reduced NK cell cytotoxicity, and this correlated with increased numbers of MADB106 lung metastases. Morphine did not influence surgery-induced NK cell cytotoxicity suppression, whereas tramadol prevented this and reduced lung metastasis (Gaspani *et al.*, 2002). The (+) enantiomer of tramadol inhibits serotonin uptake and is immunostimulatory – this effect is inhibited by the serotonergic antagonist metergoline (Sacerdote *et al.*, 1999). It has also been shown that an increased serotonergic tone stimulates NK cell cytotoxicity (Clancy *et al.*, 1991; Mossner *et al.*, 1998). Thus, the effect of tramadol on NK cell cytotoxicity following surgery might be the summation of stimulation of NK activity, which is also present in non-operated rats, due to serotonin re-uptake inhibition, and the reduction of surgical pain which is itself immunosuppressive (Gaspani *et al.*, 2002; Page, 2003; Page *et al.*, 2001). For potentially immunosuppressive opioids, there is a balance between the immunosuppressive effect of the opioid and the reduction of putative immunosuppression of pain (Figure) (Page, 2005; Rittner *et al.*, 2008).

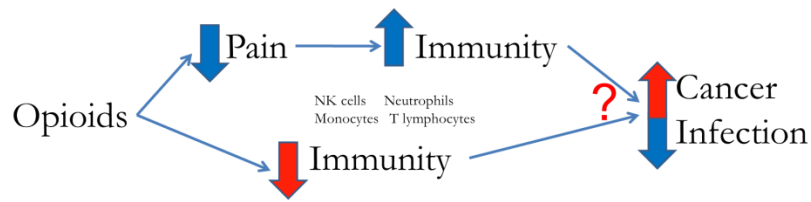


Figure 2: Opioids, pain, immunity and cancer.

Pain is immunosuppressive and may worsen rodent cancer outcome (Page, 2003; Page *et al.*, 2001). Opioids, by reducing pain, might have a beneficial effect on immune function and cancer (Gaspani *et al.*, 2002; Page, 2005). However, opioids which suppress immune function, may decrease anti-tumour immunity and promote the development of cancer; the balance of these effects is critical (Shavit *et al.*, 2004; Wang *et al.*, 2008).

A recent study using breast cancer mouse models has shown that perioperative morphine did not affect tumour growth, peripheral blood or tumour-infiltrating immune cells (although NK cells were not assessed), even when surgery was performed (Doornebal *et al.*, 2015). The difference between this and the other presented studies, could be due to orthotopic rather than intravenous inoculation of cancer cells, female mice being used (different species and genders could respond differently) and the 14-day dosing schedule of morphine which was initiated once mammary tumours had developed, or 1 day after mastectomy (Doornebal *et al.*, 2015).

Healthy volunteer studies:

Several studies assessing the effect of opioids in healthy volunteers have been performed (Table 2). These have the advantage of being able to evaluate the effect of opioids on both direct and indirect opioid-immune pathways in humans, as opposed to *in vitro* studies which can only assess direct pathways, albeit in a very controlled way (Boland *et al.*, 2014a).

Healthy volunteer study results cannot be extrapolated to people with cancer, as inflammation and immune cell activation are different.

In a volunteer study, morphine decreased NK cell cytotoxicity, the recovery of which was prolonged beyond 24 hr after the morphine was stopped with higher doses (Yeager *et al.*, 1995). In two volunteer studies, acute administration of intravenous fentanyl increased NK cell cytotoxicity, and this effect was shown to be due to an increase in the proportion of NK cells in the peripheral blood, rather than an increase in the cytotoxicity of individual NK cells (Yeager *et al.*, 2002) (Jacobs *et al.*, 1999). Although these studies suggest that morphine suppresses NK cell cytotoxicity in healthy subjects, with a more prolonged suppression at a

higher dose, and that fentanyl increases NK cell cytotoxicity, we cannot extrapolate healthy volunteer studies to the surgical/clinical setting.

Human cancer surgical studies

There is pre-clinical evidence that the immune response at the time of surgical removal of cancer is critical, that some opioids can be detrimental to the immune response and that this can promote cancer progression (Byrne *et al.*, 2016; Gaspani *et al.*, 2002; Shavit *et al.*, 2004). As *in vitro* conditions of most studies do not reproduce the biology of cancer *in vivo*, and that there are differences between opioids (Borner *et al.*, 2013), immune response and clinical outcomes to different opioids need to be studied in humans undergoing surgical resection of cancer. During surgery, opioids are just one of many factors that might affect cancer progression/metastasis (Byrne *et al.*, 2016). Furthermore, opioids have numerous mechanisms of affecting cancer growth in animal models, with the potential influence on anti-tumour immunity being just one of these (Afsharimani *et al.*, 2011; Gach *et al.*, 2011; Jaura *et al.*, 2014).

In patients undergoing surgery, surgical stress variably reduced NK cell cytotoxicity and different opioids (and the dose used) have different effects on NK cell cytotoxicity. In 30 patients undergoing surgery for uterine carcinoma to whom were administered fentanyl, anaesthetic medications and either morphine or tramadol, NK cell cytotoxicity was not affected by the surgery or by morphine (trend for inhibition only). However, tramadol increased NK cell cytotoxicity at 2 hours post-treatment despite similar analgesia (Sacerdote *et al.*, 2000). In a randomised controlled trial of 40 patients undergoing surgery for malignant or benign conditions, post-operative NK cell cytotoxicity was reduced, with high-dose fentanyl delaying the post-operative recovery of NK cell cytotoxicity (Beilin *et al.*, 1996). The long-term impact or overall outcome of different opioids and doses were not determined.

The influence of morphine with or without flurbiprofen (the flurbiprofen group consumed less morphine) on immune status has been evaluated in 60 patients undergoing surgery for gastric cancer. In the morphine treated patients, T and NK cell numbers decreased at 2 hours after incision, and NK cell numbers had not returned to baseline at 5 days after surgery. In the flurbiprofen group, NK cells at 2 hours and T and NK cells at 1 day were higher, despite similar levels of analgesia (Shen *et al.*, 2014). In a randomised controlled trial of 25 patients undergoing neck surgery, fentanyl suppressed NK cell cytotoxicity more than flurbiprofen on

day 1, but not day 2 postoperatively (Narahara *et al.*, 2013). The difference could either be a beneficial effect of the flurbiprofen or a negative effect of the opioid (Boland *et al.*, 2016; Hooijmans *et al.*, 2015).

Although NK cell cytotoxicity correlates with tumour growth and metastasis in animal models, cancer outcomes have not been assessed in human cancer surgery models (Table 2). Furthermore, the magnitude of change in NK cell cytotoxicity which is needed to produce a clinically relevant effect is unknown. It is therefore difficult to definitively attribute a decrease in NK cell cytotoxicity with poorer outcomes in patients with cancer.

Table 2: Summary of the effects of opioids on NK cell cytotoxicity and cancer outcomes in the different models

Opioid	Animal		Healthy volunteer	Human cancer surgery	
	NK cell cytotoxicity	Cancer	NK cell cytotoxicity	NK cell cytotoxicity	Cancer
Morphine	Decrease/no effect on suppressive effects of surgery	Increase	Decrease	No effect/delayed post-operative recovery	Not assessed
Fentanyl	Decrease/no effect on suppressive effects of surgery	Increase	Increase	Delayed post-operative recovery	Not assessed
Buprenorphine	No effect/reversed the suppressive effects of surgery	No effect	Not assessed	No effect/reversed the suppressive effects of surgery	Not assessed
Tramadol	Increase/reversed the suppressive effects of surgery	Decrease	Not assessed	Increase/reversed the suppressive effects of surgery	Not assessed

Systemic opioid sparing surgical studies

Many studies have assessed the effects of regional analgesia vs. systemic opioids on clinical outcomes. In general, these compare clinical outcomes between pain control with regional/neuraxial analgesia and systemic opioids in patients having cancer surgery.

Meta-analyses for the effect of neuraxial blockade on cancer surgery have generated mixed findings from the included heterogeneous and mostly retrospective studies. One concluded that there was no advantage for overall or progression-free survival (Cakmakkaya *et al.*, 2014), another suggested there might be a benefit for neuraxial blockade for prostate cancer surgery, but not for colonic cancer (Pei *et al.*, 2014). The most recent and largest meta-analysis (21 included studies) reported an association between improved recurrence-free and overall survival with neuraxial anaesthesia (compared to general anaesthetic alone), especially in patients having colorectal cancer surgery (Weng *et al.*, 2016).

There are many reviews assessing the various perioperative factors which might influence cancer-related outcomes. They suggest that although pre-clinical studies indicate a benefit of regional anaesthesia and stress response reduction in cancer formation, there is no clear association between regional/neuraxial anaesthesia for cancer surgery and tumour recurrence and cancer-related survival benefit from the heterogeneous and mostly retrospective studies. However, there might be an association with improved overall survival (Cata *et al.*, 2014; Sun *et al.*, 2015; Vogelaar *et al.*, 2016). This needs confirmation and causality explored in prospective studies.

A randomised controlled trial of 503 patients undergoing abdominal surgery for cancer resection compared the effect of general anaesthesia with either epidural analgesia or postoperative systemic opioids (median 3-day morphine dose: 0mg epidural group; 107mg opioid group). This showed no difference in cancer recurrence and mortality at 2-3 years between the two groups (Myles *et al.*, 2011). In a prospective cohort study of 34,188 cancer survivors who had resections for early stage breast cancer, opioid prescriptions were not associated with breast cancer recurrence (Cronin-Fenton *et al.*, 2015). This echoes previous studies indicating that opioids do not increase *de novo* cancer risk.

As surgical stress response and increased glucocorticoids can cause immune suppression, some of the benefits from regional anaesthesia may be because of better analgesia (reducing pain-associated immune suppression), along with attenuation of the surgical stress response, decreases in levels of endogenous opioids and the lower doses of systemic opioids (Al-Hashimi *et al.*, 2013; Byrne *et al.*, 2016; Juneja, 2014).

Data on the influence of opioid-sparing techniques for the surgical resection of cancer are mixed, and are primarily derived from retrospective studies. Although the most recent meta-analysis reported an association between improved recurrence-free and overall survival with

neuraxial anaesthesia (Weng *et al.*, 2016), more randomised controlled trials are needed in different cancers to elucidate causation, some of which are underway (Buggy *et al.*, 2015; Byrne *et al.*, 2016; Connolly *et al.*, 2016). In the meantime, there is consensus that there is currently insufficient evidence to change perioperative practice until the results of definitive randomised controlled trials are available (Buggy *et al.*, 2015; Byrne *et al.*, 2016; Connolly *et al.*, 2016).

Effects of opioids on anti-tumour immunity and survival in patients with cancer

Many people receive long-term opioids for cancer-related pain who are not undergoing surgery, and this is a different scenario in terms of the immune effects of opioids and clinical consequences.

Effects of opioids on immunological parameters that are relevant to anti-tumour immune potential in patients with cancer have been reviewed previously (Boland *et al.*, 2014b). Five human studies which assessed the immune effects of morphine in patients with cancer showed variable effects on immunologic end points. Given that none of these studies measured the clinical effects, it is not possible to know the clinical significance of these (Boland *et al.*, 2014b).

Opioids, via immune (and non-immune) effects can influence cancer progression in animal studies and other patient groups. The effect of opioids on prognosis in patients with cancer not undergoing surgery has been systematically reviewed (Boland *et al.*, 2015). In the 13 studies of people in the last days/weeks of life, there were mixed effects of opioids on survival. These studies were short term with poor methodology, sometimes only including patients with a very limited life expectancy such as from hospice admission, or just measured opioids taken in the last day(s) of life (Boland *et al.*, 2015). The best quality end of life study, a secondary data analysis, showed that an intravenous morphine-equivalent dose (IVME) above 20 mg/day was associated with a shorter survival compared with ≤ 17 mg/day (the lowest dose group); mean survival: 27 days for patients on ≤ 17 mg/day IVME vs 12 days for patients on 20-25 mg/day IVME (Portenoy *et al.*, 2006). Studies in patients with a longer prognosis (months to years) tended to be larger and of better quality. Six out of seven studies described an association between strong systemic opioid use or increasing dose and shorter survival. However, these studies did not have survival as a primary, appropriately powered,

endpoint. Furthermore they were limited in that they included variable populations, starting points for opioid use, durations of opioid administration and from when survival was measured. The main confounding factor is that the control groups were not directly matched (i.e. not patients who had refractory severe symptoms, but did not choose opioids), and that greater analgesic requirements and shorter survival is likely to be mediated by painful progressive cancer. As a consequence, these studies cannot show causality, only associations (Boland *et al.*, 2015).

Pharmacogenetic factors are also important. In 2039 women with breast cancer, the A118G MOR polymorphism, which confers a reduced receptor response to opioids, was associated with increased cancer-related survival in invasive breast cancer (Bortsov *et al.*, 2012).

Furthermore, Chinese people with the A118G MOR polymorphism had a lower incidence of oesophageal cancer (Wang *et al.*, 2013).

Future studies

In view of variable findings between study models, these cannot be extrapolated from *in vitro*, animal or healthy volunteer to clinical settings, or between the different clinical settings or between opioids (Juneja, 2014; Ramaswamy *et al.*, 2016). Therefore, appropriate studies in each of the clinical settings where opioids are used are needed. These need to focus on long term clinical studies, like those in progress in the surgical setting, and these need to be done in people needing long term opioids for non-surgical cancer-related pain. *In vitro*, animal and healthy volunteer studies should be used principally to explore mechanisms, as the environments can be carefully controlled. Furthermore, subset analysis of NK cells is needed (as CD56^{bright} and CD56^{dull} NK cells have different function) (Tabellini *et al.*, 2014), and pharmacogenomics, such as the A118G MOR polymorphism, should be taken into account.

People taking opioids for cancer-related pain could be on opioids for a long time, thus tolerance could develop. Given that they might also be on opioids intermittently, it is important to know if regular or intermittent opioids are more immune protective and how this influences clinical outcomes. As pain is immunosuppressive, the effects of pain in patients and how opioids influence immunity and clinical outcomes in these populations need further study. Appropriately designed and powered studies assessing clinical outcomes of opioid use

in people with cancer are required to inform the optimal use of opioids in these patient groups.

Conclusion

Evidence from preclinical, healthy volunteer, clinical and surgical models suggests that different opioids variably influence protective anti-tumour immunity. There are discrepancies in the results of these studies which might be partly explained by differing methodologies, species used, opioid used and the dose and duration of administration. However, in general the literature reports that morphine and fentanyl decrease NK cell cytotoxicity *in vivo* (although short-term *in vivo* exposure to fentanyl in healthy humans increases NK cell numbers and cytotoxicity), buprenorphine does not affect NK cell cytotoxicity and tramadol enhances it. This change in NK cell cytotoxicity correlates with tumour growth and metastasis in rodent models. Opioid dose and duration of administration can influence outcome.

Although clinical evidence is sparse, data suggest that perioperative opioid sparing may lead to better long-term outcomes. However, high-quality perioperative and chronic cancer-related pain studies are needed. Given that current data from patients with cancer are inconclusive, definitive recommendations about how adequate analgesia is best achieved cannot be made and opioids for cancer-related pain continues to be recommended.

References

- Afsharimani B, Cabot P, Parat MO (2011). Morphine and tumor growth and metastasis. *Cancer Metastasis Rev* **30**(2): 225-238.
- Al-Hashimi M, McDonald J, Thompson JP, Lambert DG (2016). Evidence for nociceptin/orphanin FQ (NOP) but not micro (MOP), delta (DOP) or kappa (KOP) opioid receptor mRNA in whole human blood. *Br J Anaesth* **116**(3): 423-429.
- Al-Hashimi M, Scott SW, Thompson JP, Lambert DG (2013). Opioids and immune modulation: more questions than answers. *Br J Anaesth* **111**(1): 80-88.
- Beilin B, Shavit Y, Hart J, Mordashov B, Cohn S, Notti I, *et al.* (1996). Effects of anesthesia based on large versus small doses of fentanyl on natural killer cell cytotoxicity in the perioperative period. *Anesth Analg* **82**(3): 492-497.
- Boehncke S, Hardt K, Schadendorf D, Henschler R, Boehncke WH, Duthey B (2011). Endogenous mu-opioid peptides modulate immune response towards malignant melanoma. *Experimental dermatology* **20**(1): 24-28.
- Boland J, Boland E, Brooks D (2013). Importance of the correct diagnosis of opioid-induced respiratory depression in adult cancer patients and titration of naloxone. *Clinical medicine* **13**(2): 149-151.
- Boland JW, Bennett MI (2016). Opioids do not influence metastasis in experimental animal cancer models. *Pain* **157**(5): 1173.
- Boland JW, Foulds GA, Ahmedzai SH, Pockley AG (2014a). A preliminary evaluation of the effects of opioids on innate and adaptive human in vitro immune function. *BMJ Support Palliat Care* **4**(4): 357-367.
- Boland JW, McWilliams K, Ahmedzai SH, Pockley AG (2014b). Effects of opioids on immunologic parameters that are relevant to anti-tumour immune potential in patients with cancer: a systematic literature review. *Br J Cancer* **111**(5): 866-873.
- Boland JW, Ziegler L, Boland EG, McDermid K, Bennett MI (2015). Is regular systemic opioid analgesia associated with shorter survival in adult patients with cancer? A systematic literature review. *Pain*.
- Borman A, Ciepiewski Z, Wrona D, Stojek W, Glac W, Leszkowicz E, *et al.* (2009). Small doses of morphine can enhance NK cell cytotoxicity in pigs. *Int.Immunopharmacol.* **9**(3): 277-283.

Borner C, Kraus J, Bedini A, Schraven B, Hollt V (2008). T-cell receptor/CD28-mediated activation of human T lymphocytes induces expression of functional mu-opioid receptors. *Mol Pharmacol* **74**(2): 496-504.

Borner C, Lanciotti S, Koch T, Hollt V, Kraus J (2013). mu opioid receptor agonist-selective regulation of interleukin-4 in T lymphocytes. *J Neuroimmunol* **263**(1-2): 35-42.

Borner C, Stumm R, Hollt V, Kraus J (2007). Comparative analysis of mu-opioid receptor expression in immune and neuronal cells. *J. Neuroimmunol.* **188**(1-2): 56-63.

Borner C, Warnick B, Smida M, Hartig R, Lindquist JA, Schraven B, *et al.* (2009). Mechanisms of opioid-mediated inhibition of human T cell receptor signaling. *J Immunol.* **183**(2): 882-889.

Bortsov AV, Millikan RC, Belfer I, Boortz-Marx RL, Arora H, McLean SA (2012). mu-Opioid receptor gene A118G polymorphism predicts survival in patients with breast cancer. *Anesthesiology* **116**(4): 896-902.

Buggy DJ, Borgeat A, Cata J, Doherty DG, Doornebal CW, Forget P, *et al.* (2015). Consensus statement from the BJA Workshop on Cancer and Anaesthesia. *Br J Anaesth* **114**(1): 2-3.

Byrne K, Levins KJ, Buggy DJ (2016). Can anesthetic-analgesic technique during primary cancer surgery affect recurrence or metastasis? *Can J Anaesth* **63**(2): 184-192.

Cakmakkaya OS, Kolodzie K, Apfel CC, Pace NL (2014). Anaesthetic techniques for risk of malignant tumour recurrence. *Cochrane Database Syst Rev*(11): CD008877.

Campana G, Sarti D, Spampinato S, Raffaelli W (2010). Long-term intrathecal morphine and bupivacaine upregulate MOR gene expression in lymphocytes. *International immunopharmacology.*

Cata JP, Hernandez M, Lewis VO, Kurz A (2014). Can regional anesthesia and analgesia prolong cancer survival after orthopaedic oncologic surgery? *Clinical orthopaedics and related research* **472**(5): 1434-1441.

Clancy J, Jr., Petrovic LM, Gordon BH, Handa RJ, Campbell DB, Lorens SA (1991). Effects of subchronic d-fenfluramine on splenic immune functions in young and old male and female Fischer 344 rats. *Int J Immunopharmacol* **13**(8): 1203-1212.

Connolly C, Buggy DJ (2016). Opioids and tumour metastasis: does the choice of the anesthetic-analgesic technique influence outcome after cancer surgery? *Current opinion in anaesthesiology* **29**(4): 468-474.

Cronin-Fenton DP, Heide-Jorgensen U, Ahern TP, Lash TL, Christiansen PM, Ejlertsen B, *et al.* (2015). Opioids and breast cancer recurrence: A Danish population-based cohort study. *Cancer* **121**(19): 3507-3514.

Desmond F, McCormack J, Mulligan N, Stokes M, Buggy DJ (2015). Effect of anaesthetic technique on immune cell infiltration in breast cancer: a follow-up pilot analysis of a prospective, randomised, investigator-masked study. *Anticancer research* **35**(3): 1311-1319.

Doornebal CW, Vrijland K, Hau CS, Coffelt SB, Ciampricotti M, Jonkers J, *et al.* (2015). Morphine does not facilitate breast cancer progression in two preclinical mouse models for human invasive lobular and HER2(+) breast cancer. *Pain* **156**(8): 1424-1432.

Dublin S, Walker RL, Jackson ML, Nelson JC, Weiss NS, Von Korff M, *et al.* (2011). Use of opioids or benzodiazepines and risk of pneumonia in older adults: a population-based case-control study. *J Am Geriatr Soc* **59**(10): 1899-1907.

Fecho K, Dykstra LA, Lysle DT (1993). Evidence for beta adrenergic receptor involvement in the immunomodulatory effects of morphine. *J Pharmacol Exp Ther* **265**(3): 1079-1087.

Fecho K, Maslonek KA, Dykstra LA, Lysle DT (1996). Evidence for sympathetic and adrenal involvement in the immunomodulatory effects of acute morphine treatment in rats. *J Pharmacol Exp Ther* **277**(2): 633-645.

Franchi S, Moretti S, Castelli M, Lattuada D, Scavullo C, Panerai AE, *et al.* (2012). Mu opioid receptor activation modulates Toll like receptor 4 in murine macrophages. *Brain Behav Immun* **26**(3): 480-488.

Franchi S, Panerai AE, Sacerdote P (2007). Buprenorphine ameliorates the effect of surgery on hypothalamus-pituitary-adrenal axis, natural killer cell activity and metastatic colonization in rats in comparison with morphine or fentanyl treatment. *Brain Behav. Immun.* **21**(6): 767-774.

Gach K, Wyrebska A, Fichna J, Janecka A (2011). The role of morphine in regulation of cancer cell growth. *Naunyn Schmiedebergs Arch Pharmacol* **384**(3): 221-230.

Gaspani L, Bianchi M, Limiroli E, Panerai AE, Sacerdote P (2002). The analgesic drug tramadol prevents the effect of surgery on natural killer cell activity and metastatic colonization in rats. *J Neuroimmunol* **129**(1-2): 18-24.

Glattard E, Welters ID, Lavaux T, Muller AH, Laux A, Zhang D, *et al.* (2010). Endogenous morphine levels are increased in sepsis: a partial implication of neutrophils. *PloS one* **5**(1): e8791.

Gomez-Flores R, Weber RJ (2000). Differential effects of buprenorphine and morphine on immune and neuroendocrine functions following acute administration in the rat mesencephalon periaqueductal gray. *Immunopharmacology* **48**(2): 145-156.

Gong L, Qin Q, Zhou L, Ouyang W, Li Y, Wu Y, *et al.* (2014). Effects of fentanyl anesthesia and sufentanil anesthesia on regulatory T cells frequencies. *International journal of clinical and experimental pathology* **7**(11): 7708-7716.

Grace PM, Maier SF, Watkins LR (2015). Opioid-induced central immune signaling: implications for opioid analgesia. *Headache* **55**(4): 475-489.

Hernandez MC, Flores LR, Bayer BM (1993). Immunosuppression by morphine is mediated by central pathways. *J.Pharmacol.Exp.Ther.* **267**(3): 1336-1341.

Hooijmans CR, Geessink FJ, Ritskes-Hoitinga M, Scheffer GJ (2015). A systematic review and meta-analysis of the ability of analgesic drugs to reduce metastasis in experimental cancer models. *Pain* **156**(10): 1835-1844.

Hou M, Zhou NB, Li H, Wang BS, Wang XQ, Wang XW, *et al.* (2016). Morphine and ketamine inhibit immune function of gastric cancer patients by increasing percentage of CD4(+)CD25(+)Foxp3(+) regulatory T cells in vitro. *The Journal of surgical research* **203**(2): 306-312.

Hutchinson MR, Zhang Y, Shridhar M, Evans JH, Buchanan MM, Zhao TX, *et al.* (2010). Evidence that opioids may have toll-like receptor 4 and MD-2 effects. *Brain Behav Immun* **24**(1): 83-95.

Irwin M, Hauger RL, Brown M, Britton KT (1988). CRF activates autonomic nervous system and reduces natural killer cytotoxicity. *Am J Physiol* **255**(5 Pt 2): R744-747.

Jacobs R, Karst M, Scheinichen D, Bevilacqua C, Schneider U, Heine J, *et al.* (1999). Effects of fentanyl on cellular immune functions in man. *Int J Immunopharmacol* **21**(7): 445-454.

Jaura AI, Flood G, Gallagher HC, Buggy DJ (2014). Differential effects of serum from patients administered distinct anaesthetic techniques on apoptosis in breast cancer cells in vitro: a pilot study. *Br J Anaesth* **113** Suppl 1: i63-67.

Juneja R (2014). Opioids and cancer recurrence. *Curr Opin Support Palliat Care* **8**(2): 91-101.

Koodie L, Yuan H, Pumper JA, Yu H, Charboneau R, Ramkrishnan S, *et al.* (2014). Morphine inhibits migration of tumor-infiltrating leukocytes and suppresses angiogenesis associated with tumor growth in mice. *Am J Pathol* **184**(4): 1073-1084.

Kraus J (2009). Regulation of mu-opioid receptors by cytokines. *Front Biosci.(Schol.Ed)* **1**: 164-170.

Kraus J, Borner C, Giannini E, Hickfang K, Braun H, Mayer P, *et al.* (2001). Regulation of mu-opioid receptor gene transcription by interleukin-4 and influence of an allelic variation within a STAT6 transcription factor binding site. *J Biol Chem.* **276**(47): 43901-43908.

Langsdorf EF, Mao X, Chang SL (2011). A role for reactive oxygen species in endotoxin-induced elevation of MOR expression in the nervous and immune systems. *J Neuroimmunol* **236**(1-2): 57-64.

Lennon FE, Mirzapoiazova T, Mambetsariev B, Poroyko VA, Salgia R, Moss J, *et al.* (2014). The Mu opioid receptor promotes opioid and growth factor-induced proliferation, migration and Epithelial Mesenchymal Transition (EMT) in human lung cancer. *PLoS one* **9**(3): e91577.

Lennon FE, Mirzapoiazova T, Mambetsariev B, Salgia R, Moss J, Singleton PA (2012). Overexpression of the mu-opioid receptor in human non-small cell lung cancer promotes Akt and mTOR activation, tumor growth, and metastasis. *Anesthesiology* **116**(4): 857-867.

Liang X, Liu R, Chen C, Ji F, Li T (2016). Opioid System Modulates the Immune Function: A Review. *Translational perioperative and pain medicine* **1**(1): 5-13.

Lysle DT, Hoffman KE, Dykstra LA (1996). Evidence for the involvement of the caudal region of the periaqueductal gray in a subset of morphine-induced alterations of immune status. *J Pharmacol Exp Ther* **277**(3): 1533-1540.

Maghazachi AA (2010). Role of chemokines in the biology of natural killer cells. *Current topics in microbiology and immunology* **341**: 37-58.

Martucci C, Panerai AE, Sacerdote P (2004). Chronic fentanyl or buprenorphine infusion in the mouse: similar analgesic profile but different effects on immune responses. *Pain* **110**(1-2): 385-392.

Mathew B, Lennon FE, Siegler J, Mirzapoiazova T, Mambetsariev N, Sammani S, *et al.* (2011). The novel role of the mu opioid receptor in lung cancer progression: a laboratory investigation. *Anesth Analg* **112**(3): 558-567.

Mellon RD, Bayer BM (1998). Role of central opioid receptor subtypes in morphine-induced alterations in peripheral lymphocyte activity. *Brain Res.* **789**(1): 56-67.

Mittal D, Gubin MM, Schreiber RD, Smyth MJ (2014). New insights into cancer immunoeediting and its three component phases--elimination, equilibrium and escape. *Curr Opin Immunol* **27**: 16-25.

Mohan S, Davis RL, DeSilva U, Stevens CW (2010). Dual regulation of mu opioid receptors in SK-N-SH neuroblastoma cells by morphine and interleukin-1beta: evidence for opioid-immune crosstalk. *Journal of neuroimmunology* **227**(1-2): 26-34.

Mossner R, Lesch KP (1998). Role of serotonin in the immune system and in neuroimmune interactions. *Brain Behav Immun* **12**(4): 249-271.

Myles PS, Peyton P, Silbert B, Hunt J, Rigg JR, Sessler DI (2011). Perioperative epidural analgesia for major abdominal surgery for cancer and recurrence-free survival: randomised trial. *Bmj* **342**: d1491.

Narahara H, Kadoi Y, Hinohara H, Kunimoto F, Saito S (2013). Comparative effects of flurbiprofen and fentanyl on natural killer cell cytotoxicity, lymphocyte subsets and cytokine concentrations in post-surgical intensive care unit patients: prospective, randomized study. *J Anesth* **27**(5): 676-683.

Nguyen J, Luk K, Vang D, Soto W, Vincent L, Robiner S, *et al.* (2014). Morphine stimulates cancer progression and mast cell activation and impairs survival in transgenic mice with breast cancer. *Br J Anaesth* **113** Suppl 1: i4-13.

Page GG (2003). The immune-suppressive effects of pain. *Adv Exp Med Biol* **521**: 117-125.

Page GG (2005). Immunologic effects of opioids in the presence or absence of pain. *J.Pain Symptom.Manage.* **29**(5 Suppl): S25-S31.

Page GG, Blakely WP, Ben-Eliyahu S (2001). Evidence that postoperative pain is a mediator of the tumor-promoting effects of surgery in rats. *Pain* **90**(1-2): 191-199.

Pei L, Tan G, Wang L, Guo W, Xiao B, Gao X, *et al.* (2014). Comparison of combined general-epidural anesthesia with general anesthesia effects on survival and cancer recurrence: a meta-analysis of retrospective and prospective studies. *PloS one* **9**(12): e114667.

Pergolizzi J, Boger RH, Budd K, Dahan A, Erdine S, Hans G, *et al.* (2008). Opioids and the management of chronic severe pain in the elderly: consensus statement of an International Expert Panel with focus on the six clinically most often used World Health Organization step III opioids (buprenorphine, fentanyl, hydromorphone, methadone, morphine, oxycodone). *Pain Pract* **8**(4): 287-313.

Portenoy RK, Sibirceva U, Smout R, Horn S, Connor S, Blum RH, *et al.* (2006). Opioid use and survival at the end of life: a survey of a hospice population. *J Pain Symptom Manage* **32**(6): 532-540.

Ramaswamy S, Langford R (2016). Antinociceptive and immunosuppressive effect of opioids in an acute postoperative setting: an evidence-based review. *BJA Education*.

Rittner HL, Brack A, Stein C (2008). Pain and the immune system. *Br.J.Anaesth.* **101**(1): 40-44.

Sacerdote P, Bianchi M, Gaspani L, Manfredi B, Maucione A, Terno G, *et al.* (2000). The effects of tramadol and morphine on immune responses and pain after surgery in cancer patients. *Anesth Analg* **90**(6): 1411-1414.

Sacerdote P, Bianchi M, Gaspani L, Panerai AE (1999). Effects of tramadol and its enantiomers on Concanavalin-A induced-proliferation and NK activity of mouse splenocytes: involvement of serotonin. *Int J Immunopharmacol* **21**(11): 727-734.

Salimi V, Hennis MP, Mokhtari-Azad T, Shokri F, Janssen R, Hodemaekers HM, *et al.* (2013). Opioid receptors control viral replication in the airways. *Crit Care Med* **41**(1): 205-214.

Saurer TB, Carrigan KA, Ijames SG, Lysle DT (2006a). Suppression of natural killer cell activity by morphine is mediated by the nucleus accumbens shell. *J.Neuroimmunol.* **173**(1-2): 3-11.

Saurer TB, Ijames SG, Lysle DT (2009). Evidence for the nucleus accumbens as a neural substrate of heroin-induced immune alterations. *J Pharmacol Exp.Ther.* **329**(3): 1040-1047.

Saurer TB, Ijames SG, Lysle DT (2006b). Neuropeptide Y Y1 receptors mediate morphine-induced reductions of natural killer cell activity. *J.Neuroimmunol.* **177**(1-2): 18-26.

Shao YJ, Liu WS, Guan BQ, Hao JL, Ji K, Cheng XJ, *et al.* (2016). Contribution of Opiate Analgesics to the Development of Infections in Advanced Cancer Patients. *Clin J Pain.*

Shavit Y, Ben-Eliyahu S, Zeidel A, Beilin B (2004). Effects of fentanyl on natural killer cell activity and on resistance to tumor metastasis in rats. Dose and timing study. *Neuroimmunomodulation* **11**(4): 255-260.

Shen JC, Sun HL, Zhang MQ, Liu XY, Wang Z, Yang JJ (2014). Flurbiprofen improves dysfunction of T-lymphocyte subsets and natural killer cells in cancer patients receiving post-operative morphine analgesia. *Int J Clin Pharmacol Ther* **52**(8): 669-675.

Shou J, Zhang Z, Lai Y, Chen Z, Huang J (2016). Worse outcome in breast cancer with higher tumor-infiltrating FOXP3+ Tregs : a systematic review and meta-analysis. *BMC cancer* **16**: 687.

Singleton PA, Mirzapioazova T, Hasina R, Salgia R, Moss J (2014). Increased mu-opioid receptor expression in metastatic lung cancer. *Br J Anaesth* **113** Suppl 1: i103-108.

Sun X, Yang C, Li K, Ding S (2015). The impact of anesthetic techniques on survival for patients with colorectal cancer: evidence based on six studies. *Hepato-gastroenterology* **62**(138): 299-302.

Suzuki M, Sakurada T, Gotoh K, Watanabe S, Satoh N (2013). Correlation between the administration of morphine or oxycodone and the development of infections in patients with cancer pain. *Am J Hosp Palliat Care* **30**(7): 712-716.

Tabellini G, Borsani E, Benassi M, Patrizi O, Ricotta D, Caimi L, *et al.* (2014). Effects of opioid therapy on human natural killer cells. *International immunopharmacology* **18**(1): 169-174.

Takeuchi Y, Nishikawa H (2016). Roles of regulatory T cells in cancer immunity. *Int Immunol* **28**(8): 401-409.

Vogelaar FJ, Lips DJ, van Dorsten FRC, Lemmens VE, Bosscha K (2016). Impact of anaesthetic technique on survival in colon cancer: a review of the literature. *Gastroenterology Report* **4**(1): 30-34.

Wang J, Barke RA, Ma J, Charboneau R, Roy S (2008). Opiate abuse, innate immunity, and bacterial infectious diseases. *Arch.Immunol.Ther.Exp.(Warsz.)* **56**(5): 299-309.

Wang J, Charboneau R, Balasubramanian S, Barke RA, Loh HH, Roy S (2002). The immunosuppressive effects of chronic morphine treatment are partially dependent on corticosterone and mediated by the mu-opioid receptor. *J Leukoc Biol* **71**(5): 782-790.

Wang S, Li Y, Liu XD, Zhao CX, Yang KQ (2013). Polymorphism of A118G in mu-opioid receptor gene is associated with risk of esophageal squamous cell carcinoma in a Chinese population. *International journal of clinical oncology* **18**(4): 666-669.

Wang Y, Sun J, Zheng R, Shao Q, Gao W, Song B, *et al.* (2016). Regulatory T cells are an important prognostic factor in breast cancer: a systematic review and meta-analysis. *Neoplasma* **63**(5): 789-798.

Weber RJ, Gomez-Flores R, Sora I, Uhl GR (2006). Loss of Morphine-induced Suppression of NK Cell activity and T-cell Functions in mu Opioid Receptor Knockout Mice. *Am J Immunol* **2**(2): 35-39.

Weber RJ, Pert A (1989). The periaqueductal gray matter mediates opiate-induced immunosuppression. *Science* **245**(4914): 188-190.

Weng M, Chen W, Hou W, Li L, Ding M, Miao C (2016). The effect of neuraxial anesthesia on cancer recurrence and survival after cancer surgery: an updated meta-analysis. *Oncotarget* **7**(12): 15262-15273.

Wigmore T, Farquhar-Smith P (2016). Opioids and cancer: friend or foe? *Curr Opin Support Palliat Care* **10**(2): 109-118.

Williams JP, Thompson JP, McDonald J, Barnes TA, Cote T, Rowbotham DJ, *et al.* (2007). Human peripheral blood mononuclear cells express nociceptin/orphanin FQ, but not mu, delta, or kappa opioid receptors. *Anesth.Analg.* **105**(4): 998-1005, table.

Yamamizu K, Hamada Y, Narita M (2015). kappa Opioid receptor ligands regulate angiogenesis in development and in tumours. *Br J Pharmacol* **172**(2): 268-276.

Yeager MP, Colacchio TA, Yu CT, Hildebrandt L, Howell AL, Weiss J, *et al.* (1995). Morphine inhibits spontaneous and cytokine-enhanced natural killer cell cytotoxicity in volunteers. *Anesthesiology* **83**(3): 500-508.

Yeager MP, Procopio MA, DeLeo JA, Arruda JL, Hildebrandt L, Howell AL (2002). Intravenous fentanyl increases natural killer cell cytotoxicity and circulating CD16(+) lymphocytes in humans. *Anesth Analg* **94**(1): 94-99.

Zhang EY, Xiong J, Parker BL, Chen AY, Fields PE, Ma X, *et al.* (2011). Depletion and recovery of lymphoid subsets following morphine administration. *Br J Pharmacol* **164**(7): 1829-1844.

Zhang YF, Xu QX, Liao LD, Xu XE, Wu JY, Wu ZY, *et al.* (2015). Association of mu-opioid receptor expression with lymph node metastasis in esophageal squamous cell carcinoma. *Diseases of the esophagus : official journal of the International Society for Diseases of the Esophagus / I.S.D.E* **28**(2): 196-203.

Zylla D, Gourley BL, Vang D, Jackson S, Boatman S, Lindgren B, *et al.* (2013). Opioid requirement, opioid receptor expression, and clinical outcomes in patients with advanced prostate cancer. *Cancer* **119**(23): 4103-4110.