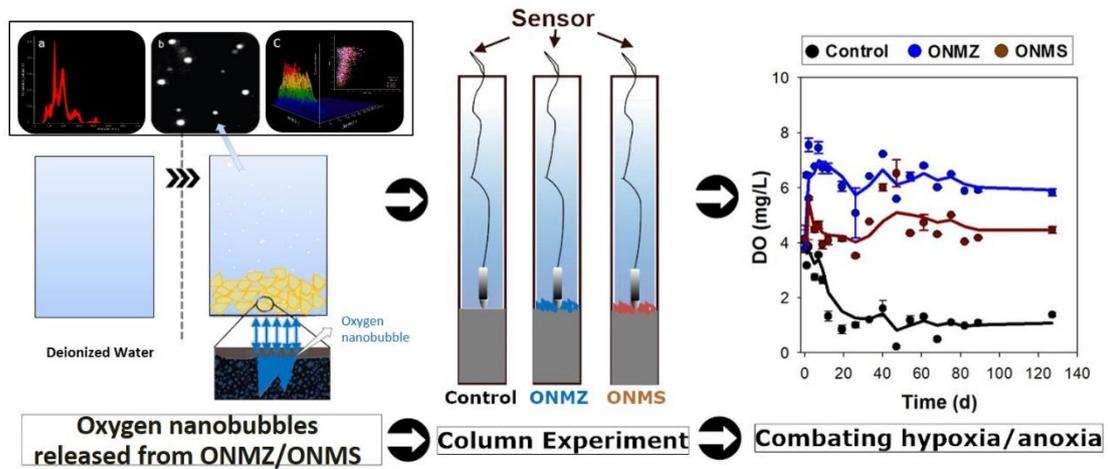


## Graphic Abstract



## Highlights

- Oxygen nanobubble is a potentially promising technique to mitigate hypoxia/anoxia
- Oxygen nanobubble modified zeolite can effectively deliver oxygen to bottom water
- The oxygen-locking surface sediment layer is crucial in reducing sediment anoxia
- Oxygen-locking sediment layer can switch the anoxia sediment from P source to sink

1 **Combating hypoxia/anoxia at sediment-water interfaces: a preliminary**  
2 **study of oxygen nanobubble modified clay materials**

3

4 Honggang Zhang<sup>1</sup>, Tao Lyu<sup>2</sup>, Lei Bi<sup>1</sup>, Grant Tempero<sup>3</sup>, David P. Hamilton<sup>3</sup>, Gang Pan<sup>\*1,2</sup>

5 <sup>1</sup>*Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences, Beijing 100085, China*

6 <sup>2</sup>*School of Animal, Rural and Environmental Sciences, Nottingham Trent University, Brackenhurst Campus,*

7 *NG250QF, UK*

8 <sup>3</sup>*School of Science, University of Waikato, Private Bag 3105, Hamilton 3240, New Zealand*

9 \*Corresponding author: [gpan@rcees.ac.cn](mailto:gpan@rcees.ac.cn) (GP)

10 **Abstract**

11 Combating hypoxia/anoxia is an increasingly common need for restoring natural waters suffering  
12 from eutrophication. The oxygen nanobubble modified natural particles were investigated for  
13 mitigating hypoxia/anoxia at sediment-water interfaces (SWI) in a simulated column experiment. By  
14 adding oxygen nanobubble modified zeolites (ONMZ) and local soils (ONMS), the oxygen  
15 nanobubble concentrations ( $10^5$ - $10^7$  particles/mL) were several orders of magnitude higher in the  
16 water than the original water solution ( $10^4$  particles/mL) within 24 hours. In the column experiment,  
17 an oxygen-locking surface sediment layer were formed after capping with ONMZ and ONMS  
18 particles. The synergy of diffusion of oxygen nanobubbles and retention of oxygen in this layer  
19 contributes to both the increase of DO and reversal of hypoxic conditions. The overlaying water  
20 maintained significantly higher dissolved oxygen (DO) values (4-7.5 mg/L) over the experimental

21 period of 127 days in ONMZ and ONMS compared with the control systems (around 1 mg/L).  
22 Moreover, the oxidation-reduction potential (ORP) was reversed from -200 mV to 180-210 mV and  
23 maintained positive values for 89 days in ONMZ systems. In the control systems, ORP was  
24 consistently negative and decreased from -200 mV to -350 mV. The total phosphorus (TP) flux from  
25 sediment to water across SWI was negative in the ONMZ and ONMS treated systems, but positive  
26 in the control system, indicating the sediment could be switched from TP source to sink. The oxygen-  
27 locking capping layer was crucial in preventing oxygen consumption caused by the reduced  
28 substances released from the anoxic sediment. The study outlines a potentially promising technology  
29 for mitigating sediment anoxia and controlling nutrients release from sediments, which could  
30 contribute significantly to addressing eutrophication and ecological restoration.

31 **Keywords:** deep water, eutrophication control, harmful algae blooms, nutrient flux, oxygen deliver

## 32 **1. Introduction**

33 Hypoxia/anoxia is a global threat to aquatic ecosystems, often inducing "dead zones" at the  
34 sediment-water interface (SWI) (Diaz and Rosenberg, 2008; Feist et al., 2016; Stramma et al., 2008).  
35 In the dead zones, sediment release rates may be accelerated for many constituents, including  
36 phosphorus, nitrogen, iron, manganese, methyl-mercury and hydrogen sulfide (Beutel et al., 2008;  
37 De Vittor et al., 2016; Gantzer et al., 2009; Testa and Kemp, 2012; Zhu et al., 2013). Among the  
38 released substances, phosphorus and nitrogen can lead to eutrophication, which is often associated  
39 with harmful algal blooms (Funkey et al., 2014). Moreover, the hypoxic/anoxic condition can be  
40 exacerbated by the additional oxygen demand from the mineralization of dead algal biomass (Diaz

41 and Rosenberg, 2008; Testa and Kemp, 2012). Thus, mitigation of hypoxia/anoxia at the SWI is  
42 crucial for both water quality improvement and eutrophication control.

43 Current efforts de-signed to replenish benthic dissolved oxygen (DO) and remove the anoxic  
44 environment are often based on the directly injecting either air (aeration) (Henares et al., 2015), or  
45 oxygen gas (oxygenation) (Bierlein et al., 2017) sometimes using oxygen-supersaturated water (Forth  
46 et al., 2015) into the hypoxic region near the SWI. Although these techniques have been reported to  
47 be effective to some extent, they are still limited by high cost and efficiency at large scale (Bormans  
48 et al., 2016). Additionally, gas or water pumped into the SWI region may disturb the settled sediment  
49 and induce internal releases of nutrients and other contaminants to the water column, as well as  
50 potentially leading to additional oxygen consumption and increase hypoxia (Bierlein et al., 2017).  
51 The pump system also needs to be continuously operated to maintain the oxygen supply to the SWI,  
52 otherwise DO may be rapidly consumed, leading to rapid return of anoxia (Bryant et al., 2010). In  
53 the Baltic Sea, where hypoxic waters have expanded in area from 5,000 to > 60,000 km<sup>2</sup> in the last  
54 century (Carstensen et al., 2014), enhanced ventilation of deep waters through additional input of  
55 oxygenated surface water has been suggested (Conley et al., 2009). However, this method will require >  
56 30 years to take effect and may cause a drastic change in stratification and alteration of the  
57 biodiversity (Funkey et al., 2014). Ventilation by pumping oxygen-rich water downward to alleviate  
58 hypoxia in the Baltic Sea is estimated to require more than 100 pump stations (0.6 MW each) at a  
59 cost of around 20,000 million Euros (Stigebrandt and Gustafsson, 2007). Therefore, developing a  
60 more cost-effective and sustainable technique for hypoxia/anoxia mitigation in bottom waters and at  
61 the SWI is vitally important.

62 Oxygen nanobubbles have attracted increasing attention in recent years due to the  
63 characteristics of high gas solubility and long lifetime of oxygen in the liquid (Ebina et al., 2013;  
64 Peng et al., 2015). As opposed to oxygen gas (Cavalli et al., 2009), nano-scale oxygen bubbles could  
65 slowly diffuse oxygen into the surrounding water phase and last more than 70 days when diameter is  
66 <200 nm (Ebina et al., 2013). The oxygen nanobubble technique has already been widely used in  
67 medicine (Cai et al., 2015), physiology (Ebina et al., 2013) and water treatment (Agarwal et al., 2011).  
68 However, a cost-effective method to deliver the oxygen nanobubble into SWI for hypoxia/anoxia  
69 mitigation remains a bottleneck. It was recently reported that oxygen nanobubbles can be generated  
70 and persist at solid particle-water interfaces (i.e., surface nanobubbles) (Pan et al., 2016; Wang et al.,  
71 2016; Yang et al., 2013). The presence of oxygen nanobubbles has been proven and quantified at the  
72 rough and irregular surfaces of clay particles (Pan et al., 2016). It is a means to increase total oxygen  
73 content in a suspension by adding clay particles loading with oxygen nanobubbles (Pan and Yang,  
74 2012; Pan et al., 2011). Sedimentation of a carrier loaded with oxygen nanobubbles due to the gravity  
75 effect provides a mechanism to alter the hypoxia/anoxia near the SWI but has not been investigated  
76 systematically.

77 Many geo-engineering methods, such as adding phosphorus-adsorbing materials, have been  
78 demonstrated to significantly contribute to remediating eutrophication control and contributing to  
79 lake restoration (Huser et al., 2016; Noyma et al., 2016; Spears et al., 2014; Waajen et al., 2016).  
80 However, the sinking materials cover the sediment and their effect on redox potential at the SWI may  
81 be temporary (Pan et al., 2012). Additionally, most of the adsorbing materials, e.g., metal salts and  
82 Phoslock®, are synthesized artificially and may have potential side-effects on the environment.  
83 Natural sediments entering lakes through weathering and runoff, have high microporous surface area

84 (Pan et al., 2013). These natural particles have potentially as oxygen nanobubble carriers to deliver  
85 oxygen to the SWI. However, no previous study has applied such technology and there is little  
86 knowledge about the effects of oxygen nanobubbles on the oxygen conditions, redox potential and  
87 nutrient fluxes at the SWI.

88 The objective of this study is to investigate for the first time the efficacy and sustainability of a  
89 surface oxygen nanobubble technique for mitigating hypoxia/anoxia and its effect on nutrient fluxes  
90 across the SWI. Local soil and natural zeolite were selected as the oxygen nanobubble carriers in the  
91 experiment. After oxygen nanobubble modified zeolite (ONMZ) or oxygen nanobubble modified  
92 soils (ONMS) were applied in simulated eutrophic water-sediment systems in the laboratory columns,  
93 oxygen levels in the overlying water and redox potential at SWI were monitored. Nutrient  
94 concentrations, including total phosphorus (TP), total nitrogen (TN), ammonium ( $\text{NH}_4^+\text{-N}$ ), nitrate  
95 ( $\text{NO}_3^-\text{-N}$ ), and nitrite ( $\text{NO}_2^-\text{-N}$ ), were measured in the overlying water and nutrient fluxes across the  
96 SWI were calculated.

## 97 **2. Materials and methods**

### 98 **2.1 Preparation of oxygen nanobubble modified materials**

99 Natural zeolite and local soil were selected as the carrier materials to investigate the effect of  
100 surface oxygen nanobubble technology on hypoxia/anoxia mitigation at the SWI. Zeolite with particle  
101 size of 1-2 mm was purchased from Yongjia Natural Minerals Ltd., Hebei, China. Local soil from  
102 Lake Ngaroto, Waikato, New Zealand, was sieved through a mesh sieve to remove particles  $>380\ \mu\text{m}$ .  
103 Ngaroto is the largest peat lake in Waikato region, with a surface area of about 108 ha, maximum  
104 depth of 4 m and average depth of c. 2 m. Land in the catchment of this lake is mostly used for pastoral

105 grazing. The lake is hypertrophic and has major cyanobacteria harmful algal blooms throughout  
106 summer. The specific surface area and micropore size of the natural zeolite and local soil were  
107 determined by the Brunauer–Emmett–Teller (BET) method with a Micromeritics ASAP-2020  
108 apparatus (Micromeritics Inc., USA) (Zhang et al., 2014).

109 The zeolite and soil, were washed with deionized water and dried for 10 h at 90 °C. The  
110 preparation of oxygen nanobubble modified zeolite (ONMZ) and soil (ONMS) followed a modified  
111 method based on exposure to oxygen supersaturating ambient conditions (Pan et al., 2016). Briefly,  
112 the materials (zeolite or soil) were placed into a pressure-resistant and airtight container. A vacuum  
113 was created to hold pressure to -0.08 to -0.1 MPa for 2 h to remove gas from the micropores of zeolite  
114 and soil. Thereafter, pure O<sub>2</sub> (99.99%) was pumped into the container and held at a pressure of 0.12  
115 to 0.15 MPa for 4 h to load the O<sub>2</sub>. The oxygen nanobubble loading process, including the creation  
116 of the vacuum, was repeated three times to achieve supersaturation of O<sub>2</sub> in the particle micropores.

## 117 **2.2 Nanobubbles analysis**

118 Prior to the column experiment, the release potential of oxygen nanobubbles into water from  
119 the modified solid particles was tested via a flask experiment. Twenty grams of the oxygen  
120 nanobubble modified zeolite or soil was put into 250 mL flasks with 200 mL deionized water and  
121 sealed by gas-permeable sealing film (0.3 μm). Controls consisted of flasks of 250 mL filled with 200  
122 mL of deionized water. Each control and treatment flask experiment were conducted in triplicate. To  
123 confirm the sequential changes of nanobubble release, size (detection range; 10-1000 nm) and  
124 concentration of oxygen nanobubbles, measurements were conducted at 1 min, 6 h and 24 h in a  
125 nanoparticle-tracking analysis instrument (NanoSight NS500 & NTA 2.0 Analytical Software,

126 Malvern Instrument Ltd, Salisbury, UK) at room temperature ( $24 \pm 1^\circ\text{C}$ ).

### 127 **2.3 Column experiment**

128 The column experiment was conducted in an indoor laboratory in University of Waikato, New  
129 Zealand, over a total duration of 127 days. Six plexiglass cylinders with identical inner diameter of  
130 12 cm and height of 150 cm were used as incubation columns (Fig. S1 **Error! Reference source not**  
131 **found.**). Each column was filled in the bottom 20 cm with the lake sediment and with filtered (mesh  
132 size of 25  $\mu\text{m}$ ) lake water (also from Lake Ngaroto) to a height of 120 cm. Each experimental column  
133 was wrapped with black plastic to shield the system from ambient light. The columns included  
134 duplicates of a control, treatment by oxygen nanobubble modified natural zeolite (ONMZ) and  
135 oxygen nanobubble modified local soil (ONMS). The oxidation-reduction potential (ORP) meter  
136 (HANNA, HI2001) was placed lightly on the sediment surface in each column to monitor the change  
137 of ORP at the SWI throughout the experiment. After a 3-day stabilization period, the ONMZ and  
138 ONMS treatment systems were pretreated by flocculation using 3 mg/L chitosan modified soils (Li  
139 and Pan, 2013) following by application of approximately 100 g of ONMZ and ONMS, resulting a 2  
140 cm depth capping layer.

### 141 **2.4 Sampling and analysis**

142 During the experiment, overlying water samples (100 mL) from each column were carefully  
143 collected from 5 cm above the sediment using a syringe with a siphon. The collected water samples  
144 were evenly separated into three parts (c.33 mL) which were measured for turbidity and nutrient  
145 concentrations (TP, TN,  $\text{NH}_4^+\text{-N}$ ,  $\text{NO}_3^-\text{-N}$ , and  $\text{NO}_2^-\text{-N}$ ). After each sample collection, all columns  
146 were slowly replenished with the original filtered lake water to compensate for the sampling losses.

147 Turbidity was analyzed with portable turbidity meter (HANNA, HI98713). TP was determined using  
148 a potassium persulfate digestion-Mo-Sb-Vc colorimetric method, TN using an alkaline potassium  
149 persulphate digestion–ultraviolet spectrometer,  $\text{NH}_4^+$ -N with Nessler's colorimetric, and  $\text{NO}_3^-$ -N and  
150  $\text{NO}_2^-$ -N with ultraviolet colorimetric method with and without cadmium column reduction,  
151 respectively (APHA, 1998). The DO was measured using a Yellow Springs Instruments (YSI, Proplus)  
152 by carefully putting the meters into the overlying water and holding at 1-2 cm above the SWI. To  
153 avoid cross contamination, the meters were carefully cleaned with Milli-Q water and ethanol between  
154 measurements. The DO and turbidity were measured simultaneously with ORP from the in-site meters  
155 (HANNA, HI2001) at days 0, 1, 2, 5, 7, 9, 12 and then every around 7 days until day 89, although  
156 DO was measured until day 127. The concentrations of TP, TN,  $\text{NH}_4^+$ -N,  $\text{NO}_3^-$ -N, and  $\text{NO}_2^-$ -N were  
157 measured at 7-day frequency until 47 days and then monitored every 14 days. The nutrient fluxes  
158 were calculated from day 19 in order to minimize the influence caused by suspended substances  
159 sedimentation as indicated by relative low turbidity in control. All samples were tested in triplicate  
160 for each duplicate column, values were averaged and standard deviations for the samples from the  
161 same treatment system.

## 162 2.5 Calculation

163 The monitoring data on nutrient concentrations obtained after day 19 was used to calculate  
164 nutrient fluxes at the SWI. The average nutrient flux was calculated according to the following mass  
165 balance Equation (1):

$$166 \quad F_n = \frac{[V(c_n - c_m) + \sum_{j=1, i=1}^n V_j(c_n - c_i)]}{S \times t} \quad (1)$$

167 Where  $F_n$  is the flux on  $n^{\text{th}}$  sampling day ( $\text{mgm}^{-2}\text{d}^{-1}$ ),  $V$  is the volume of overlying water (L),  $c_n$  is the nutrient

168 concentration ( $\text{mgL}^{-1}$ ) on  $n^{\text{th}}$  sampling day ( $n > 19$ ),  $c_m$  is the nutrient concentration ( $\text{mgL}^{-1}$ ) on  $m^{\text{th}}$  sampling day  
169 when turbidity had stabilized in the three columns (i.e., the 19<sup>th</sup> day in this study),  $c_i$  is the nutrient  
170 concentration of the compensating water for maintaining the volume of water in the columns ( $\text{mgL}^{-1}$ ),  $V_j$  is volume of sampling water (L),  $S$  is the cross section area of each column ( $\text{m}^2$ ) and  $t$  is incubation time  
171  
172 (d).

## 173 **2.6 Statistical Analysis**

174 Sigmaplot software (version 12.5, Sigma, Inc.) and SPSS16.0 (Statistical Program for Social  
175 Sciences) were used for plotting and data analyses, respectively. A one-way ANOVA and post hoc  
176 Tukey's HSD test were used to compare water quality parameters (DO, ORP and turbidity) and  
177 nutrient concentrations between different treatment systems (control, ONMZ and ONMS) at each  
178 corresponding sampling point, with differences accepted at a significance level  $< 0.05$ . To fully  
179 understand the effect of surface oxygen nanobubble technology on the hypoxia/anoxia mitigation in  
180 the column experiment, principal component analysis (PCA) was used to provide an overview of the  
181 system performance using a visualization method to normalize all parameters. PCA was used to  
182 identify different performance patterns between control, ONMZ and ONMS treatment systems during  
183 from day 19 to day 89 of the experiment. PCA was conducted using all measured parameters,  
184 including DO, ORP, turbidity, TP, TN,  $\text{NH}_4^+\text{-N}$ ,  $\text{NO}_3^-\text{-N}$  and  $\text{NO}_2^-\text{-N}$ . The data was standardized (to  
185 a Z score with a mean = 0 and S.D. = 1) to ensure that each variable had the same influence in the  
186 analysis. Multiple correlation analysis was carried out to assess the relationships between all  
187 measured parameters in the column experiment. The data was checked for normality and  
188 homogeneity of variance prior to all statistical analysis. If variables were not normally distributed,  
189 they were log-transformed.

## 190 **3. Results**

### 191 **3.1 Release of oxygen nanobubbles from ONMZ and ONMS**

192 In order to verify the release of oxygen nanobubbles from the modified particles, the water  
193 solution in the flask experiment after ONMZ and ONMS addition was analyzed in a nanoparticle-  
194 tracking analysis instrument to detect the concentration and size of the nanobubbles. The  
195 concentrations of nanobubbles were both around  $10^7$  particles/mL immediately after the addition (1  
196 min) of ONMZ (Fig. 1) and ONMS (Fig. 2) into the water, and approximately 70% of the released  
197 nanobubbles were  $<200$  nm in diameter. The concentration of nanobubbles in the water with ONMZ  
198 addition was maximal at 24 h (still around  $10^7$  particles/mL), and the concentration in the water with  
199 ONMS addition decreased to  $10^5$  particles/mL. Nevertheless, the concentrations were clearly elevated  
200 compared with the original deionized water, which remained around  $10^4$  particles/mL (Fig. S2). The  
201 peaks in the concentration vs. size graph (Fig. 1) move toward the right direction of the x axis, which  
202 indicates that the size of nanobubbles gradually increased along with the culture time in both ONMZ  
203 and ONMS (Fig. 2) treated water. The 3D graph shows two distinct nanobubble populations, clearly  
204 confirmed by the higher light scattering intensity of the nanobubbles in ONMZ treated water  
205 compared to the ONMS treated water.

### 206 **3.2 DO and ORP dynamics at the sediment-water interfaces**

207 At the beginning of the column experiment, the DO and ORP of the overlying water in all  
208 systems were around 4 mg/L (Fig. 3a) and -200 mV (Fig. 3b), respectively. After the oxygen  
209 nanobubble modified particles application, DO increased in the first 5 days and reached around 7.5  
210 and 5.5 mg/L in ONMZ and ONMS systems, respectively. Concentrations gradually decreased to 6  
211 and 4.3 mg/L and generally remained around this level until the conclusion of the experiment on day

212 127 in the ONMZ and ONMS systems, respectively. However, the control system maintained a  
213 hypoxic condition with DO declining to 1 mg/L at day 20 of the experiment, and remaining around  
214 this level throughout the tested period of 127 days. Significant differences of DO concentrations in  
215 the overlaying water between each system were observed ( $P < 0.05$ ) and followed the order of ONMZ >  
216 ONMS > control (**Error! Reference source not found.3a**). ORP values showed a similar pattern as  
217 DO with significantly higher values in ONMZ, followed by ONMS and control systems along the  
218 experiment (**Error! Reference source not found.3b**). ORP values were increased from -200 mV to  
219 210 and 180 mV in the ONMZ and ONMS systems, respectively, in the first 5 days. During the  
220 experiment, ORP decreased until day 20 and remained reasonably stable at 150 mV and -160 mV in  
221 the ONMZ and ONMS columns, respectively, until day 89. In the control columns, the ORP showed  
222 a continuous decrease from -200 mV to -350 mV by day 89.

### 223 **3.3 Nutrient dynamics in bottom water**

224 After 19 days of the experimental set-up, the turbidity of the overlying waters had decreased  
225 from 40 NTU to around 5 NTU in the control columns and < 1 NTU in ONMZ and ONMS treated  
226 systems (**Error! Reference source not found.4a**). Turbidity remained around this level until day 89.  
227 The control systems had elevated TP until day 50 and significantly higher TP concentrations (0.08  
228 mg/L) in the overlying water compared with the ONMZ and ONMS treated systems (Fig. 4b). The  
229 TP concentrations in the ONMZ and ONMS treated columns maintained below 0.02 mg/L and did  
230 not show significant differences between the two treatments.

231 TN concentrations in the ONMS columns increased slightly and the values (around 2 mg/L)  
232 become significantly higher than those in ONMZ columns (around 1.5 mg/L) and in the control

233 columns (around 1.2 mg/L) at day 89 (**Error! Reference source not found.4c**).  $\text{NH}_4^+\text{-N}$   
234 concentrations showed a similar tendency of increase from day 19, with the highest concentration at  
235 day 40; concentrations were around 0.1, 0.3 and 0.65 mg/L in ONMZ, ONMS and control systems,  
236 respectively (**Error! Reference source not found.4d**). Concentrations of  $\text{NH}_4^+\text{-N}$  gradually  
237 decreased to <0.02, 0.18 and 0.13 mg/L by day 89 in ONMZ, ONMS and control systems, respectively.  
238 The concentrations of  $\text{NO}_3^-\text{-N}$  (**Error! Reference source not found.5e**) and  $\text{NO}_2^-\text{-N}$  (**Error!**  
239 **Reference source not found.4f**) were significantly higher in the ONMZ and ONMS systems than  
240 that in control after 19 days of incubation. However, the concentrations in ONMZ and ONMS systems  
241 showed a clear decreasing tendency after day 80 and became similar to levels in the control systems  
242 (0.01 mg/L of  $\text{NO}_3^-\text{-N}$  and 0.03 mg/L of  $\text{NO}_2^-\text{-N}$ ).

### 243 **3.4 Nutrients fluxes across the sediment-water interfaces**

244 Fig. 5 shows the nutrient (TP, TN,  $\text{NH}_4^+\text{-N}$ ,  $\text{NO}_3^-\text{-N}$ , and  $\text{NO}_2^-\text{-N}$ ) fluxes across from the  
245 sediment to the overlying water (effluxes) from day 26 calculated by Equation (1). The flux of TP  
246 were negative in ONMZ and ONMS systems (-0.1 to -2.3 mg/m<sup>2</sup>/d). The control system had positive  
247 values of TP flux, with a decreasing tendency through time and around 0.3 mg/m<sup>2</sup>/d at day 89 (**Error!**  
248 **Reference source not found.5a**).

249 Generally, flux of TN was negative (average of -10 mg/m<sup>2</sup>/d) in the control during the  
250 experiment (**Error! Reference source not found.5b**), while values in the ONMZ systems were  
251 generally positive (average of 5 mg/m<sup>2</sup>/d). In ONMS systems, TN flux reversed from negative to  
252 positive at day 47 and finally reached around 0 at day 89. The  $\text{NH}_4^+\text{-N}$  flux was positive (5-20  
253 mg/m<sup>2</sup>/d) in all systems (**Error! Reference source not found.5c**) at day 26, however, the values

254 declined with time, and reached around 3 mg/m<sup>2</sup>/d in the control system and -1 and -4 mg/m<sup>2</sup>/d in the  
255 ONMZ and ONMS systems at day 89. **Error! Reference source not found.5d** shows that the flux of  
256 NO<sub>3</sub><sup>-</sup>-N in the ONMZ and ONMS systems was consistently positive (4-15 mg/m<sup>2</sup>/d). However, the  
257 flux of NO<sub>3</sub><sup>-</sup>-N in the control systems was always negative (< -0.8mg/m<sup>2</sup>/d). The flux of NO<sub>2</sub><sup>-</sup>-N was  
258 around 0 in the control system, however, values reversed from positive to negative in the ONMZ and  
259 ONMS systems at day 89 (**Error! Reference source not found.5e**).

### 260 3.5 Environmental factors identified with statistical analysis

261 Water quality parameters (DO, ORP and turbidity) and nutrient concentrations (TP, TN, NH<sub>4</sub><sup>+</sup>-  
262 N, NO<sub>3</sub><sup>-</sup>-N and NO<sub>2</sub><sup>-</sup>-N) in the overlying water were analysed with principal component analysis  
263 (PCA) to examine differences between treatment systems (control, ONMZ and ONMS) and sampling  
264 times from day 19 to day 89 (**Error! Reference source not found.6**). The eigenvalues of all the  
265 components of the PCA are shown in Table S1. The first two components (PC 1 and PC 2) contribute  
266 high proportions (57.6% and 17.7%) of the variance in both experimental phases, thus, they were  
267 extracted as principal components for further analysis. Clear group differences between three systems  
268 were shown in the visualized figures (**Error! Reference source not found.6**), suggesting  
269 performances differed by treatment. The control systems tended to locate to the left side of the  
270 ordination, and the ONMS and ONMZ systems located in the upper right lower right side, respectively.  
271 TP and turbidity contribute substantially to the control system performance patterns due to higher  
272 levels of TP and turbidity in overlying water of the control systems. DO and ORP contribute more to  
273 the positive side of PC1 (the location of the ONMS and ONMZ systems), which means the oxygen  
274 nanobubble modified particle application reduced hypoxia/anoxia at the SWI. The DO, ORP and NO<sub>3</sub><sup>-</sup>  
275 -N loading factors were higher for the ONMZ than the ONMS system but this was reversed for TN,

276  $\text{NH}_4^+\text{-N}$  and  $\text{NO}_2^-\text{-N}$ .

277 Multiple correlation analysis was carried out as a blind test by homogenizing all the data from  
278 all columns to assess the relationships between all the parameters (Table 1). The DO and ORP showed  
279 a significant negative correlation with TP concentrations and significant positive correlation with  
280  $\text{NO}_3^-\text{-N}$  concentrations. TP concentrations were also negatively correlated with TN and positively  
281 correlated with turbidity.

## 282 **4. Discussion**

### 283 **4.1 Instant increase of DO by oxygen nanobubble delivery**

284 Bottom water oxygenation is an increasingly common lake management strategy for mitigating  
285 hypoxia/anoxia and associated deleterious effects on water quality in deep lakes and reservoirs  
286 (Bierlein et al., 2017). The current methods may require a number of large pumps with very high  
287 costs and energy consumption (Funkey et al., 2014; Stigebrandt and Gustafsson, 2007). In the present  
288 study, surface oxygen nanobubble technology was demonstrated to deliver oxygen into the water or  
289 to the SWI using a carrier of modified zeolite (ONMZ) or soils (ONMS) through gravity settling with  
290 minimum energy consumption. During the ONMZ and ONMS treatments, part of the oxygen loaded  
291 on the microporous surfaces of zeolite and soil quickly diffused into water through both visible  
292 microbubbles (Fig. S3) and non-visible oxygen nanobubbles (Fig. 1 and 2), and instantly increased  
293 the DO concentrations from 4 mg/L to 7.5 and 5.5 mg/L in ONMZ and ONMS treated systems,  
294 respectively, in the first 5 days (**Error! Reference source not found.**3a). Oxygen nanobubbles were  
295 directly detected in the water through the flask experiment, which verified that they could be  
296 generated/delivered and then released to the surrounding bulk water from the ONMZ and ONMS

297 (**Error! Reference source not found.** 1 and 2). The formed oxygen-locking sediment capping layer  
298 not only released oxygen into the water, but also retained oxygen which could significantly mitigate  
299 and even reverse the anoxic condition at the sediment-water interfaces (Fig. 3b). Currently,  
300 hypolimnetic oxygenation need to develop some novel oxygen diffuser equipment in order to prevent  
301 any sediment resuspension (Gafsi et al., 2016). The settling delivery method via clay or soil particle  
302 carriers can fundamentally avoid the typical problems of resuspension of sediment and rigorous  
303 turbulence to the SWI faced by conventional deep water aeration methods, which may be important  
304 for lake restoration in maintaining natural stratification conditions (Beutel and Horne, 1999; Gachter  
305 and Wehrli, 1998). The ONMZ was more efficient in supplying oxygen to the SWI than the ONMS,  
306 which may due to the locking ability of oxygen by zeolite particles as well as its larger  
307 specific surface area than the natural local soils (Table S2).

#### 308 **4.2 Sustained reversal of hypoxia/anoxia at the SWI**

309 The enhanced DO levels in bottom water resulting from oxygenation by conventional pumping  
310 methods may be rapidly negated after turning the pump off (Bierlein et al., 2017; Bryant et al., 2010;  
311 Gachter and Wehrli, 1998). Rapid depletion of DO occurs through oxidation of both organic detritus  
312 and reduced chemical substances from the sediment and thus restoration of oxygenation is a long-  
313 lasting project (Liboriussen et al., 2009). The maintenance of higher DO levels in the ONMZ and  
314 ONMS treated systems in the present study can be attributed to the long lifespan of the oxygen  
315 nanobubbles and oxygen retention within the capping layer. It was reported that when aerated bubbles  
316 are in nano size (<200 nm), they can have much longer life than the macro bubbles (Ebina et al.,  
317 2013). These oxygen nanobubbles can slowly diffuse oxygen into the water column and maintain the  
318 higher DO level (4-7.5 mg/L) in ONMZ and ONMS. The reversed ORP values from -200 mV to 180-

319 210 mV in ONMZ for 89 days indicated that oxidation status can be sustained in the capping layer  
320 for very long time beyond months. Part of oxygen nanobubbles can be generated and stable exist at  
321 the zeolite/soil particle-water interfaces in the experiment which can be inferred from the evidence  
322 that a stable cloud of O<sub>2</sub> nanobubbles could be found at the diatomite particle-water interface after  
323 oxygen loading (Pan et al., 2016). Oxygen retention in the capping layer and downward penetration  
324 of oxygen into the deeper sediment formed an oxygen-locking sediment layer and contribute to a  
325 persistent reversal of anoxic condition. The oxygen nanobubbles appeared to either be active or have  
326 provided sufficient oxidation to maintain elevated levels of DO and redox potential for 3-4 months in  
327 the present study. At the end of present study, it was visually evidenced that around 4 cm of sediment  
328 at the SWI showed a light yellow color in the ONMZ systems, which was in sharp contrast to an  
329 unchanged black anoxic layer in the control systems (Fig. 7).

330 Although the previous study shows that the chitosan modified soils could form a capping layer  
331 on the sediment to decrease the internal nutrients loading, the capper layer could not provide extra  
332 oxygen delivery into the water and sediment and thus only can remediate the hypoxia for a short  
333 period (Pan et al., 2012).The color difference of the sediment in the present study indicated that in  
334 addition to direct diffusion of oxygen into the water column, there was significant amount of oxygen  
335 loaded in ONMZ and ONMS that can penetrate into the sediment at a considerable depth. This likely  
336 facilitated oxidation of organic matter as well as reduced substances. What is most important is that  
337 such a layer provides physical isolation to prevent reduced substances from the anoxic layer from  
338 diffusing upwards into the water column. Most hypoxia is formed when DO in bottom water is  
339 consumed by the anaerobic substances from the sediment while there is not enough oxygen  
340 replenishing from the surface water. The reversal of ORP at SWI in the ONMZ and ONMS systems

341 (Fig. 3b) could be expected to lead to oxidation of reduced forms of iron, manganese, methyl-mercury  
342 and sulfide that would otherwise be released from the sediment (Beutel et al., 2008; De Vittor et al.,  
343 2016; Gantzer et al., 2009; Testa and Kemp, 2012; Zhu et al., 2013). The nanobubble technology  
344 demonstrated in this study appeared to provide a novel principle for the remediation of  
345 hypoxia/anoxia in the bottom water.

### 346 **4.3 Manipulating nutrients fluxes at SWI**

347 Reversal of hypoxia/anoxia may not only prevent the release of reduced species release and  
348 facilitate organic matter mineralization (Beutel et al., 2008; De Vittor et al., 2016; Gantzer et al., 2009;  
349 Testa and Kemp, 2012), but can also influence nutrient recycling across SWI. In the present study,  
350 evidence of the positive effect of alleviation of hypoxia remediation on nutrient recycling was found  
351 in the significant differences in nutrient concentrations and fluxes across SWI between the ONMZ  
352 and ONMS treated and control systems (**Error! Reference source not found.4** and **Error!**  
353 **Reference source not found.5**). Obvious P release from the sediment to the overlying water occurred  
354 in the control system, however, there was a net flux of TP from water to the sediments in the ONMZ  
355 and ONMS systems (**Error! Reference source not found.4b** and **Error! Reference source not**  
356 **found.5a**). Although, in some previous reports, capping with unmodified materials could also  
357 influence nutrient fluxes in short period (Faithfull et al., 2008; Pan et al., 2012), no evidence showed  
358 these materials could reversal the sediment hypoxia/anoxia which can contribute to P fixation in  
359 sediment. The main mechanism of the P release from sediments is generally related to changes in  
360 redox-sensitive iron and manganese oxide minerals and the associated P (Funes et al., 2016; Jordan  
361 et al., 2008), with inorganic phosphorus generally adsorbed by the metal oxide–hydroxide complexes  
362 under oxic conditions (Tang et al., 2013; Xu et al., 2013). Reversal of the ORP sign (i.e., positive

363 values) in ONMZ treated systems indicated that the SWI changed from anoxic to oxic. Jilbert et al.  
364 (2011) found preferential remineralization of P in relation to carbon and nitrogen during  
365 decomposition of organic substances induced by reducing conditions plays a key role leading to  
366 surplus bioavailable P in the Baltic Sea, and was likely why TP concentrations in the overlying water  
367 were significantly positively correlated with DO and ORP (Table 1). Reversal of ORP in ONMZ  
368 induced a conversion of sediment from P sources to sinks compared with that in the control.

369 Changes in redox potential affect N transformations from bacterial activities at the SWI,  
370 particularly nitrification, denitrification and anammox reactions (Brzozowska and Gawronska, 2009).  
371 The significantly higher oxygen condition induced by surface oxygen nanobubble materials (**Error!**  
372 **Reference source not found.**3) may facilitate nitrification, which converts  $\text{NH}_4^+\text{-N}$  to  $\text{NO}_3^-\text{-N}$   
373 (Rassamee et al., 2011). Thus, lower  $\text{NH}_4^+\text{-N}$  occurred in ONMZ. Moreover, nitrification can supply  
374 necessary  $\text{NO}_2^-\text{-N}$  as an electron acceptor to anammox bacteria which can reduce  $\text{NH}_4^+\text{-N}$  to nitrogen  
375 gas (Kim et al., 2016). Nitrification-anammox coupling reactions often occur between aerobic and  
376 anaerobic zones (Brzozowska and Gawronska, 2009), notably in the SWI region. The co-existence  
377 of nitrification and anammox in the treated systems may reduce the efflux of  $\text{NH}_4^+\text{-N}$  in the  
378 experimental systems compared with those in the control at the end of the experiment (**Error!**  
379 **Reference source not found.**5c). In control systems, anaerobic conditions not only hinder  
380 nitrification and supply of  $\text{NO}_2^-\text{-N}$  for anammox bacteria but may also stimulate sulfide accumulation,  
381 further inhibiting nitrification (Jensen et al., 2008). Thus, the lowest concentrations of  $\text{NO}_3^-\text{-N}$  in the  
382 control systems were likely to have been through denitrification (**Error! Reference source not**  
383 **found.**4e). Beyond denitrification, dissimilatory  $\text{NO}_3^-\text{-N}$  reduction to  $\text{NH}_4^+\text{-N}$  can also be facilitated  
384 under anoxic conditions (McCarthy et al., 2008). The sum of these effects contributed to the reduced

385 TN efflux in the treated systems (**Error! Reference source not found.5b**). It should be noted that  
386 local soils, collected from the lake and used in our experiment, may contain more organic matter than  
387 zeolite. The higher  $\text{NH}_4^+$ -N concentrations in the ONMS treated systems might be produced by  
388 mineralization of organic N from the soils (**Error! Reference source not found.4d**). Previous studies  
389 point out that organic matter in the anoxic sediment could be degraded together with N  
390 transformations from organic N to  $\text{NH}_4^+$ -N within two weeks (Han et al., 2015; Xu et al., 2013). This  
391 is consistent with our results that  $\text{NH}_4^+$ -N increased between 19 and 33 days and decreased in the  
392 subsequent days in all the three systems (**Error! Reference source not found.4d**). As a result of  
393 these processes above mentioned, using surface oxygen nanobubble technology can significantly  
394 regulate the biogeochemical processes that regulate species of phosphorus and nitrogen at the SWI.  
395 It would be an important topic to study the role of oxygen nanobubbles in relation to microbial  
396 communities under controlled laboratory and field conditions.

#### 397 **4.4 Implementation perspective for lake geo-engineering**

398 The present study has, for the first time, indicated that oxygen nanobubble modified clays can  
399 deliver amounts of oxygen into both water and sediment where there has previously been  
400 hypoxia/anoxia. By using the geo-engineering method developed based on oxygen nanobubbles, it is  
401 possible to deliver oxygen into bottom water/sediment through gravity settling, which can achieve  
402 both replenishing oxygen consumption in the "dead zone" with minimum energy consumption and  
403 minimizing the disturbance to the water stratification and surface sediment. The prolonged time  
404 effects of oxygen nanobubbles, denoted by improving oxygen levels and reversal of ORP, may further  
405 trigger a series changes in physico-chemical and microbial responses at SWI. Nevertheless, the in-  
406 site experiment of the surface nanobubble technology in lakes and sea waters need to be further

407 investigated.

## 408 **5. Conclusions**

409         The study verified a novel principle for combating hypoxia/anoxia at the sediment-water  
410 interfaces using oxygen nanobubble technology. The synergistic effects of diffusion and retention of  
411 oxygen in the nanobubble modified natural zeolite (ONMZ) and local soils (ONMS) contributed to  
412 mitigating sediment anoxia and controlling phosphorus release from bottom sediments. It was found  
413 that amount of oxygen loaded in clay particles could release via both macro- and nano-scale bubbles  
414 and quickly increase the DO levels in water column. Moreover, the oxygen nanobubbles in the  
415 modified particles could stably retain at SWI and penetrate oxygen downward to the sub-layer  
416 sediment, and then form an oxygen-locking sediment layer between the anoxic sediment and water  
417 column. This layer can sustainably reverse hypoxia/anoxia condition at SWI for several months.  
418 Finally, nutrient fluxes across the SWI could be regulated by capping with oxygen nanobubble  
419 modified materials in which the bottom sediments become adsorptive for phosphorus, rather than  
420 releasing it. In this study we have demonstrated the potential for a major breakthrough in remediation  
421 of aquatic systems via geo-engineering and delivery of oxygen and form oxygen-locking later into  
422 the deep sediment-water interface, which is crucial for eutrophication control and ecological  
423 restoration.

## 424 **Acknowledgments**

425         The research was supported by the Strategic Priority Research Program of CAS  
426 (XDA09030203), National Natural Science Foundation of China (41401551), Beijing Natural  
427 Science Foundation (8162040) and National Key Research and Development Program of China

428 (2017YFA0207204). It was further supported by the Bay of Plenty Regional Council and the New  
429 Zealand Ministry of Business, Innovation and Employment (UOWX1503; Enhancing the health and  
430 resilience of New Zealand lakes).

431

432 **References**

- 433 Agarwal A, Ng WJ, Liu Y, 2011. Principle and applications of microbubble and nanobubble  
434 technology for water treatment. *Chemosphere* 84 (9), 1175-1180.
- 435 APHA, 1998. Standard methods for the examination of water and wastewat. American Public Health  
436 Association, Washington DC 20th Edition.
- 437 Beutel MW, Horne AJ, 1999. A Review of the Effects of Hypolimnetic Oxygenation on Lake and  
438 Reservoir Water Quality. *Lake Reservoir Manag.* 15 (4), 285-297.
- 439 Beutel MW, Leonard TM, Dent SR, Moore BC, 2008. Effects of aerobic and anaerobic conditions on  
440 P, N, Fe, Mn, and Hg accumulation in waters overlaying profundal sediments of an oligo-  
441 mesotrophic lake. *Water Res.* 42 (8-9), 1953-1962.
- 442 Bierlein KA, Rezvani M, Socolofsky SA, Bryant LD, Wüest A, Little JC, 2017. Increased sediment  
443 oxygen flux in lakes and reservoirs: The impact of hypolimnetic oxygenation. *Water Resources*  
444 *Research.*
- 445 Bormans M, Marsalek B, Jancula D, 2016. Controlling internal phosphorus loading in lakes by  
446 physical methods to reduce cyanobacterial blooms: a review. *Aquat. Ecol.* 50 (3), 407-422.
- 447 Bryant LD, Lorrai C, McGinnis DF, Brand A, Wuest A, Little JC, 2010. Variable sediment oxygen  
448 uptake in response to dynamic forcing. *Limnol. Oceanogr.* 55 (2), 950-964.
- 449 Brzozowska R, Gawronska H, 2009. The influence of a long-term artificial aeration on the nitrogen  
450 compounds exchange between bottom sediments and water in Lake Dlugie. *Oceanological and*  
451 *Hydrobiological Studies* 38 (1), 113-119.
- 452 Cai WB, Yang HL, Zhang J, Yin JK, Yang YL, Yuan LJ, et al., 2015. The Optimized Fabrication of  
453 Nanobubbles as Ultrasound Contrast Agents for Tumor Imaging. *Sci Rep-Uk* 5, 13725.

454 Carstensen J, Andersen JH, Gustafsson BG, Conley DJ, 2014. Deoxygenation of the Baltic Sea during  
455 the last century. *Proc. Natl. Acad. Sci. U. S. A.* 111 (15), 5628-5633.

456 Cavalli R, Bisazza A, Rolfo A, Balbis S, Madonnaripa D, Caniggia I, et al., 2009. Ultrasound-  
457 mediated oxygen delivery from chitosan nanobubbles. *Int. J. Pharm.* 378 (1–2), 215-217.

458 Conley DJ, Bonsdorff E, Carstensen J, Destouni G, Gustafsson BG, Hansson LA, et al., 2009.  
459 Tackling Hypoxia in the Baltic Sea: Is Engineering a Solution? *Environ. Sci. Technol.* 43 (10),  
460 3407-3411.

461 De Vittor C, Relitti F, Kralj M, Covelli S, Emili A, 2016. Oxygen, carbon, and nutrient exchanges at  
462 the sediment–water interface in the Mar Piccolo of Taranto (Ionian Sea, southern Italy). *Environ.*  
463 *Sci. Pollut. Res.* 23 (13), 12566-12581.

464 Diaz RJ, Rosenberg R, 2008. Spreading dead zones and consequences for marine ecosystems. *Science*  
465 321 (5891), 926-929.

466 Ebina K, Shi K, Hirao M, Hashimoto J, Kawato Y, Kaneshiro S, et al., 2013. Oxygen and Air  
467 Nanobubble Water Solution Promote the Growth of Plants, Fishes, and Mice. *PLoS One* 8 (6).

468 Faithfull CL, Hamilton DP, Burger DF, Duggan IC. Waikato peat lakes sediment nutrient removal  
469 scoping exercise. CBER Contract Report. The University of Waikato, 2008.

470 Feist TJ, Pauer JJ, Melendez W, Lehrter JC, DePetro PA, Rygwelski KR, et al., 2016. Modeling the  
471 Relative Importance of Nutrient and Carbon Loads, Boundary Fluxes, and Sediment Fluxes on  
472 Gulf of Mexico Hypoxia. *Environ. Sci. Technol.* 50 (16), 8713-8721.

473 Forth M, Liljebldh B, Stigebrandt A, Hall POJ, Treusch AH, 2015. Effects of ecological engineered  
474 oxygenation on the bacterial community structure in an anoxic fjord in western Sweden. *ISME J.*  
475 9 (3), 656-669.

476 Funes A, de Vicente J, Cruz-Pizarro L, Alvarez-Manzaneda I, de Vicente I, 2016. Magnetic  
477 microparticles as a new tool for lake restoration: A microcosm experiment for evaluating the  
478 impact on phosphorus fluxes and sedimentary phosphorus pools. *Water Res.* 89, 366-374.

479 Funkey CP, Conley DJ, Reuss NS, Humborg C, Jilbert T, Slomp CP, 2014. Hypoxia Sustains  
480 Cyanobacteria Blooms in the Baltic Sea. *Environ. Sci. Technol.* 48 (5), 2598-2602.

481 Gachter R, Wehrli B, 1998. Ten years of artificial mixing and oxygenation: No effect on the internal  
482 phosphorus loading of two eutrophic lakes. *Environ. Sci. Technol.* 32 (23), 3659-3665.

483 Gafsi M, Kettab A, Djehiche A, Goteicha K, 2016. Study of the efficiency of hypolimnetic aeration  
484 process on the preservation of the thermal stratification. *Desalin Water Treat* 57 (13), 6017-6023.

485 Gantzer PA, Bryant LD, Little JC, 2009. Controlling soluble iron and manganese in a water-supply  
486 reservoir using hypolimnetic oxygenation. *Water Res.* 43 (5), 1285-1294.

487 Han C, Ding SM, Yao L, Shen QS, Zhu CG, Wang Y, et al., 2015. Dynamics of phosphorus-iron-  
488 sulfur at the sediment-water interface influenced by algae blooms decomposition. *J. Hazard.*  
489 *Mater.* 300, 329-337.

490 Henares MNP, Preto BD, Rosa FRT, Valenti WC, Camargo AFM, 2015. Effects of artificial substrate  
491 and night-time aeration on the water quality in *Macrobrachium amazonicum* (Heller 1862) pond  
492 culture. *Aquac. Res.* 46 (3), 618-625.

493 Huser BJ, Futter M, Lee JT, Perniel M, 2016. In-lake measures for phosphorus control: The most  
494 feasible and cost-effective solution for long-term management of water quality in urban lakes.  
495 *Water Res.* 97, 142-152.

496 Jensen MM, Kuypers MMM, Lavik G, Thamdrup B, 2008. Rates and regulation of anaerobic  
497 ammonium oxidation and denitrification in the Black Sea. *Limnol. Oceanogr.* 53 (1), 23-36.

498 Jilbert T, Slomp CP, Gustafsson BG, Boer W, 2011. Beyond the Fe-P-redox connection: preferential  
499 regeneration of phosphorus from organic matter as a key control on Baltic Sea nutrient cycles.  
500 *Biogeosciences* 8 (6), 1699-1720.

501 Jordan TE, Cornwell JC, Boynton WR, Anderson JT, 2008. Changes in phosphorus biogeochemistry  
502 along an estuarine salinity gradient: The iron conveyor belt. *Limnol. Oceanogr.* 53 (1), 172-184.

503 Kim H, Ogram A, Bae H-S, 2016. Nitrification, Anammox and Denitrification along a Nutrient  
504 Gradient in the Florida Everglades. *Wetlands*, 1-9.

505 Li L, Pan G, 2013. A universal method for flocculating harmful algal blooms in marine and fresh  
506 waters using modified sand. *Environ. Sci. Technol.* 47 (9), 4555-4562.

507 Liboriussen L, Sondergaard M, Jeppesen E, Thorsgaard I, Grunfeld S, Jakobsen TS, et al., 2009.  
508 Effects of hypolimnetic oxygenation on water quality: results from five Danish lakes.  
509 *Hydrobiologia* 625, 157-172.

510 McCarthy MJ, McNeal KS, Morse JW, Gardner WS, 2008. Bottom-water hypoxia effects on  
511 sediment-water interface nitrogen transformations in a seasonally hypoxic, shallow bay (Corpus  
512 christi bay, TX, USA). *Estuaries and Coasts* 31 (3), 521-531.

513 Noyma NP, de Magalhães L, Furtado LL, Mucci M, van Oosterhout F, Huszar VLM, et al., 2016.  
514 Controlling cyanobacterial blooms through effective flocculation and sedimentation with  
515 combined use of flocculants and phosphorus adsorbing natural soil and modified clay. *Water Res.*  
516 97, 26-38.

517 Pan G, Dai LC, Li L, He LC, Li H, Bi L, et al., 2012. Reducing the Recruitment of Sedimented Algae  
518 and Nutrient Release into the Overlying Water Using Modified Soil/Sand Flocculation-Capping  
519 in Eutrophic Lakes. *Environ. Sci. Technol.* 46 (9), 5077-5084.

- 520 Pan G, He G, Zhang M, Zhou Q, Tyliczszak T, Tai R, et al., 2016. Nanobubbles at Hydrophilic  
521 Particle-Water Interfaces. *Langmuir* 32 (43), 11133-11137.
- 522 Pan G, Krom MD, Zhang MY, Zhang XW, Wang LJ, Dai LC, et al., 2013. Impact of Suspended  
523 Inorganic Particles on Phosphorus Cycling in the Yellow River (China). *Environ. Sci. Technol.*  
524 47 (17), 9685-9692.
- 525 Pan G, Yang B, 2012. Effect of Surface Hydrophobicity on the Formation and Stability of Oxygen  
526 Nanobubbles. *Chemphyschem* 13 (8), 2205-2212.
- 527 Pan G, Yang B, Lei L, Lei, Liao L, Ding C, Wang D, 2011. A method for remediating of anoxic  
528 sediment in lakes by using nanobubbles. Patent NO. 200910080563.5.
- 529 Peng H, Birkett GR, Nguyen AV, 2015. Progress on the Surface Nanobubble Story: What is in the  
530 bubble? Why does it exist? *Adv. Colloid Interface Sci.* 222, 573-580.
- 531 Rassamee V, Sattayatewa C, Pagilla K, Chandran K, 2011. Effect of Oxidic and Anoxic Conditions on  
532 Nitrous Oxide Emissions from Nitrification and Denitrification Processes. *Biotechnol. Bioeng.*  
533 108 (9), 2036-2045.
- 534 Spears BM, Maberly SC, Pan G, Mackay E, Bruere A, Corker N, et al., 2014. Geo-Engineering in  
535 Lakes: A Crisis of Confidence? *Environ. Sci. Technol.* 48 (17), 9977-9979.
- 536 Stigebrandt A, Gustafsson BG, 2007. Improvement of Baltic proper water quality using large-scale  
537 ecological engineering. *Ambio* 36 (2-3), 280-286.
- 538 Stramma L, Johnson GC, Sprintall J, Mohrholz V, 2008. Expanding oxygen-minimum zones in the  
539 tropical oceans. *Science* 320 (5876), 655-658.
- 540 Tang WZ, Zhang H, Zhang WQ, Wang C, Shan BQ, 2013. Biological invasions induced phosphorus  
541 release from sediments in freshwater ecosystems. *Colloid Surface A* 436, 873-880.

- 542 Testa JM, Kemp WM, 2012. Hypoxia-induced shifts in nitrogen and phosphorus cycling in  
543 Chesapeake Bay. *Limnol. Oceanogr.* 57 (3), 835-850.
- 544 Waajen G, van Oosterhout F, Douglas G, Lürling M, 2016. Geo-engineering experiments in two  
545 urban ponds to control eutrophication. *Water Res.* 97, 69-82.
- 546 Wang L, Miao XJ, Pan G, 2016. Microwave-Induced Interfacial Nanobubbles. *Langmuir* 32 (43),  
547 11147-11154.
- 548 Xu D, Chen YF, Ding SM, Sun Q, Wang Y, Zhang CS, 2013. Diffusive Gradients in Thin Films  
549 Technique Equipped with a Mixed Binding Gel for Simultaneous Measurements of Dissolved  
550 Reactive Phosphorus and Dissolved Iron. *Environ. Sci. Technol.* 47 (18), 10477-10484.
- 551 Yang CW, Lu YH, Hwang IS, 2013. Condensation of Dissolved Gas Molecules at a  
552 Hydrophobic/Water Interface. *Chinese J Phys* 51 (1), 174-186.
- 553 Zhang ZS, Wang G, Li Y, Zhang X, Qiao NL, Wang JH, et al., 2014. A new type of ordered  
554 mesoporous carbon/polyaniline composites prepared by a two-step nanocasting method for high  
555 performance supercapacitor applications. *J Mater Chem A* 2 (39), 16715-16722.
- 556 Zhu G, Wang S, Wang W, Wang Y, Zhou L, Jiang B, et al., 2013. Hotspots of anaerobic ammonium  
557 oxidation at land-freshwater interfaces. *Nature Geosci* 6 (2), 103-107.

558

559

560