

Recognition of stream drying based on benthic macroinvertebrates: a new tool in Central Europe

Michal Straka^{a,b,*}, Marek Polášek^{a,b}, Vít Syrovátka^b, Rachel Stubbington^c, Světlana Zahrádková^b, Denisa Němejcová^a, Lenka Šikulová^b, Pavla Řezníčková^b, Libuše Opatřilová^{d,e}, Thibault Datry^f & Petr Pařil^b

^aT. G. Masaryk Water Research Institute, p. r. i., Mojmírovo náměstí 16, 612 00, Brno, Czech Republic

^bDepartment of Botany and Zoology, Faculty of Science, Masaryk University, Kotlářská 2, 611 37, Brno, Czech Republic

^cSchool of Science and Technology, Nottingham Trent University, Nottingham, NG11 8NS, UK

^dT. G. Masaryk Water Research Institute, p. r. i., Podbabská 2582/30, 160 00, Praha 6, Czech Republic

^ePovodí Vltavy, State Enterprise, Holečkova 3178/8, 150 00, Praha 5, Czech Republic

^fIRSTEA, UR RIVERLY, Centre de Lyon-Villeurbanne, France

*corresponding author: michal.straka@centrum.cz

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Abstract

Many streams in the extensive Central European region have an intermittent flow regime. Conventional hydrological methods used to identify zero-flow conditions, and in particular drying events, have limited use when assessing large areas dominated by low-order streams. We developed a novel multimetric index to recognise antecedent stream drying based on the analysis of benthic macroinvertebrate communities. The data used to develop the index were collected in pristine streams with different flow permanence regimes between 2012 and 2014, using standard sampling methods for ecological status assessment. The data include 64 perennial, 19 near-perennial and 27 intermittent benthic macroinvertebrate samples. Metrics considered for the index included variables based on (i) the occurrence of indicator taxa, (ii) the proportion of biological and ecological traits, and (iii) structural community metrics. Linear discriminant analysis identified the metric combinations that best discriminated among the three flow permanence categories. Different metrics were used in the final multimetric index calculation for the autumn and spring season that followed stream drying. In both seasons, the index included the proportion of indicator taxa and the proportion of taxa with high body flexibility. In addition, the autumn index included the proportion of taxa with a preference for organic substrates, whereas in spring the index included total abundance. Independent data from regulatory monitoring activity were used to validate the accuracy of the index. The correct classification of independent samples was 92% and 96% for samples from perennial and non-perennial sites, respectively. The index can be calculated using data collected by routine monitoring programmes used to assess ecological status and provides information about stream intermittence where conventional hydrological monitoring is limited. As intermittent streams increase in extent in global regions including Central Europe, the tool may be of particular interest to those who use invertebrates to monitor or manage these ecosystems.

1. Introduction

Intermittent rivers and ephemeral streams (i.e. streams with periodic flow cessation and/or drying; IRES) are mostly associated with arid and semi-arid regions. However, they are also common and widespread in temperate and continental regions with cooler, wetter climates (Datry, et al. 2014a; Stubbington et al., 2017). IRES are typically managed using methods developed for perennial waterways, or as if they were part of the terrestrial ecosystems (Acuña et al., 2014; Stubbington et al., 2018), and the need for more effective policies and management strategies has been highlighted around the world (Datry et al., 2017; Marshall et al., 2018). In the European Union, a major objective for water management is the assessment of ecological status in water bodies, to meet the legislative requirements of the EU Water Framework Directive (European Commission, 2000). The characterisation of 'biological quality elements' such as benthic macroinvertebrates is one important approach used to evaluate ecological status.

Aquatic macroinvertebrates are routinely used as indicator organisms for determining ecological status and diagnosing specific environmental pressures (e.g. Rosenberg and Resh, 1993; Birk et al., 2012). However, even short-term (i.e. days to weeks) stream drying can substantially alter benthic macroinvertebrate community composition (Datry et al., 2014b;

Hille et al., 2014; Lancaster and Ledger, 2015). The interpretation of ecological status based on metrics developed in perennial streams can thus be misleading (Munné and Prat, 2009; Menció and Mas-Pla, 2010; Wilding et al., 2018). Information about flow permanence is crucial to accurately interpret data used to evaluate status and thus inform effective IRES management. In addition, knowledge about stream intermittence is needed, to anticipate the effects of climate change, to assess human pressures such as water abstraction and to manage protected species and habitats (Wilby et al., 2010).

Using flow gauging stations to monitor the flow permanence of small streams is expensive and technically impractical, because small streams (Strahler orders 1-4) comprise a substantial part of the stream network length (e.g. 92% in Czech Republic; Zahrádková et al., 2015). Moreover, even where hydrological data are collected, they may not distinguish between lentic and dry zero-flow conditions, and alternative methods are therefore needed to recognize IRES and characterize their water regimes (Gallart et al., 2016; Beaufort et al., 2018), for example using stream biota. Loss of surface water acts as an ecological filter (Poff 1997) and benthic macroinvertebrates have taxon-specific quantitative responses to drying (Datry et al., 2014b; Leigh and Datry, 2017). Dry phases can therefore be detected through both structural (i.e. taxonomic) and functional (i.e. trait-based) changes in community composition (Bogan et al., 2013; Schriever et al., 2015; Leigh et al., 2016; Chadd et al., 2017). Presence/absence of indicator taxa of benthic invertebrates has also been used to recognize stream flow duration (NC Division of Water Quality, 2010; Nadeau, 2015), and both taxonomic structure and species trait information have been used to assess flow connectivity in Mediterranean regions (Cid et al., 2016). Such tools may enable the data routinely collected during biomonitoring programmes to provide information about flow intermittence, even if hydrological data are absent. However, no method to detect drying events has been developed for the extensive continental-climate region of Central Europe.

Different metrics can be sensitive to different aspects of stream drying, and a combination of structural, functional and indicator-taxa metrics may therefore provide robust and sensitive insight into the responses of an assemblage to drying (Li et al., 2010). Here, our aim was to develop and test a new multimetric index to identify antecedent stream drying events based on the structural and functional composition of benthic macroinvertebrate communities. We evaluated three groups of metrics as indicators of stream drying: (i) the occurrence of indicator taxa; (ii) the proportion of biological and ecological traits; and (iii) structural community metrics. We identified the metrics and metric combinations that most effectively indicated antecedent stream drying.

2. Materials and Methods

2.1. Study area

The study was conducted in 23 small (order 2-4) streams in the Czech Republic (Fig. 1). All were classified as reference streams, i.e. exposed to minimal anthropogenic impacts. Catchment land use was dominated by woodland, with smaller proportions of agricultural land, and with at least 10-m riparian buffer strips of deciduous bushes and trees. The study area has a warm-summer humid continental climate (Köppen-Geiger classification *Dfb*; Peel et al., 2007) and spans the Hercynian (17 streams) and West-Carpathian (6 streams) biogeographical subprovinces (Culek, 2013), which approximately correspond with

ecoregions 9 (Central Highlands) and 10 (Carpathians; Illies, 1967). To facilitate comparison of macroinvertebrate communities from different sites, streams were paired, with each pair comprising one non-perennial and one perennial stream of comparable altitude, size, geology and hydromorphology. In one case, a single perennial stream was paired with two non-perennial streams. A sampling site representative of typical conditions was selected in each stream. The altitude of the sampling sites ranged from 250 m to 560 m a.s.l. and the maximum distance between paired sites was 13 km.

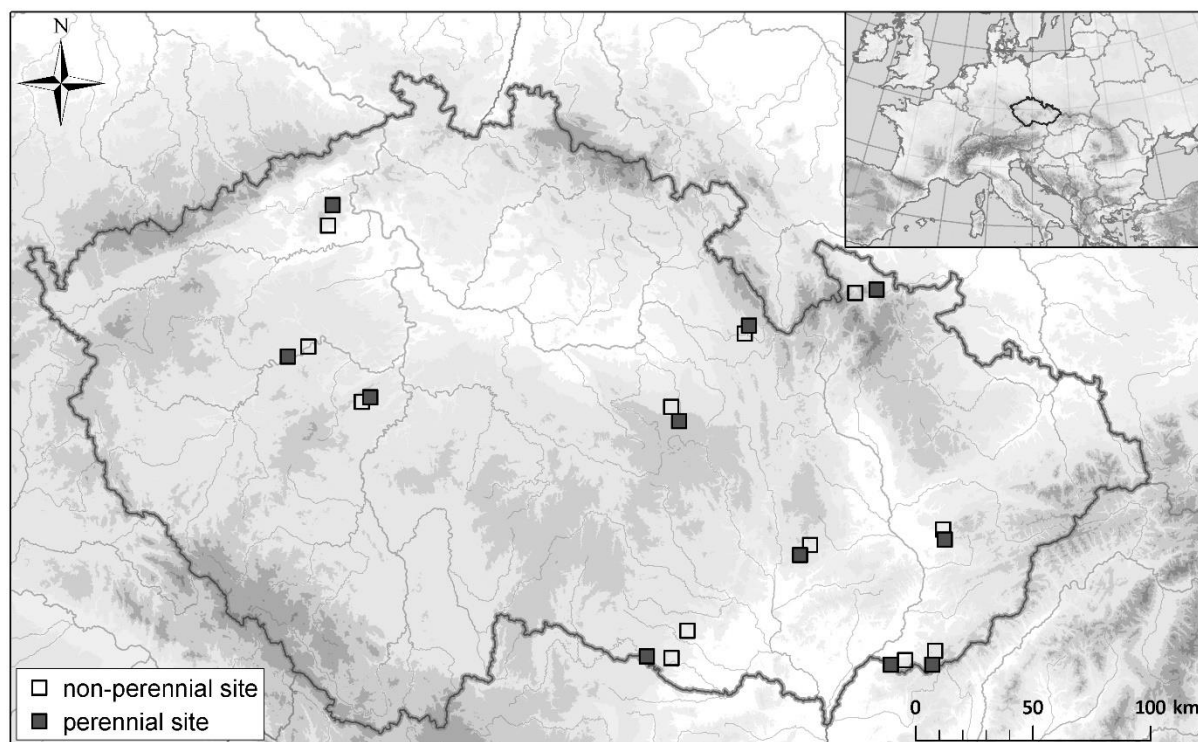


Fig. 1. Location of the study sites in the Czech Republic. Open squares represent non-perennial (i.e. near-perennial and intermittent) sites and filled squares represent perennial sites.

2.2. Sampling strategy

Benthic macroinvertebrate assemblages were sampled in spring (March-April) in 2013 and 2014 and in autumn (September-October) in 2012, 2013 and 2014. Proportional multi-habitat three-minute kick samples were collected with a hand net (25 × 25 cm aperture, 0.5-mm mesh size) according to the standard method used for ecological status assessments in the Czech Republic (Kokeš et al., 2006; Kokeš and Němejcová, 2006). Samples were preserved in 4% formaldehyde and processed in the laboratory. All macroinvertebrates were identified to the lowest possible taxonomic level: 292 out of 421 to species level, 117 to genus level, and the remaining 12 to a higher taxonomic level.

Sites were classified as non-perennial or perennial a priori, based on expert knowledge. In addition, observed instream conditions were recorded from summer 2012 to autumn 2014, including determination of the dry period duration using a water-level logger (Solinst Levellogger Edge) and photo-trap (Acorn 5310MG) installed at each site. Each water-level

logger was placed in the lowest part of the streambed, to account for the persistence of surface water in isolated pools. Photo-traps were installed in trees in the adjacent riparian zone (at 3 m height) and were facing a water-level gauge board. Logger and photo-trap data were collected every 15 minutes and every 4 hours, respectively.

We divided individual samples into three intermittence categories: *perennial*, *intermittent* and an intermediate *near-perennial* category. The allocation of samples into the three categories was based primarily on the dry period duration: i) perennial sites were defined as those which experienced more than one year of continuous flow prior to sample collection; ii) near-perennial sites lost surface water for hundreds of meters for <7 d in the summer prior to sampling, with disconnected pools occasionally present; and iii) intermittent sites dried for >1 km for 7-86 d before sampling. The cut-off between near-perennial and intermittent sites was arbitrary, reflecting the gradual nature of stream drying. In total, we collected 27 perennial, 7 near-perennial and 12 intermittent samples in spring and 34 perennial, 12 near-perennial and 15 intermittent samples in autumn. Spring samples from intermittent and near-perennial streams were taken 175-224 d after flow resumption and autumn samples were taken 2-63 d after flow resumption.

2.3. Identification of perennial flow indicators and flow intermittence tolerant taxa

To identify taxa associated with either perennial or intermittent streams, we performed Indicator Species Analysis (IndVal; Dufrêne and Legendre, 1997) using samples from spring 2013 and autumn 2012 and 2013. IndVal was performed using the `multipatt` function in the R package `indicspecies` (Cáceres and Legendre, 2009), which identifies taxa with a strong association with each group based on their abundance and frequency. The calculated indicator value (IV) of a given taxon for its preferred group ranges from zero (no affinity) to one (strongest affinity). Best-matching patterns were tested for statistical significance of the associations. Taxa significantly associated with either perennial or intermittent and near-perennial samples were considered as reliable indicators and were assigned a reliability value of 2 (see “Proportion of indicator taxa” section). Taxa with non-significant IV but found exclusively in either perennial or intermittent and near-perennial streams in at least 10% of samples were considered as indicators with lower reliability and were assigned a reliability value of 1.

To support the IndVal analyses, we conducted a literature review of primary research papers comparing macroinvertebrate assemblages at perennial and non-perennial sites during flowing phases. From these studies, a taxon was identified as indicative of perennial flow if it was absent from intermittent stream assemblages. A taxon was considered as intermittence tolerant if it was found in intermittent stream assemblages. Literature sources describing the autecology of macroinvertebrate taxa were also reviewed to explore taxon-specific responses to intermittence. If a taxon’s relationship with stream intermittence was described once in data-based published sources then the species was assigned a reliability value of 1, and if it was published two or more times, a value of 2 was assigned. The review focused on taxa living in the Czech Republic area; consulted literature is listed in Appendix A. If information in different published sources were contradictory or unclear, then a taxonomic expert decided the reliability of the taxa based on the available evidence.

2.4. Metrics used for index development

Three groups of potential metrics that indicate flow intermittence were calculated, to inform subsequent development of the multimetric index:

i) Proportion of indicator taxa

The proportion of indicator taxa was calculated as the sum of the reliability values of perennial flow indicators present in a sample divided by the sum of the reliability values of all (perennial + intermittence-tolerant) indicators present. The proportion of indicators was rescaled to zero and the resultant values are within the range -1 to +1.

$$\text{Proportion of indicator taxa} = \left(\frac{\sum \text{values of perennial flow indicators}}{\sum \text{values of all indicators}} \times 2 \right) - 1$$

ii) Proportion of macroinvertebrate traits in community

The macroinvertebrate community was characterized using traits extracted from available databases (IS ARROW, 2014; Schmidt-Kloiber and Hering, 2015), supplemented by information from the literature review and from experts in different taxonomic groups. Information about fine-sediment-sensitive invertebrates was taken from Extence et al. (2013). To describe the ability to penetrate substrata, taxonomic experts assigned each taxon to one of three body flexibility modalities: (i) none, body flexibility is limited by shell or exoskeleton, e.g. molluscs, adult beetles; (ii) low, body is flexible but cannot turn around, e.g. mayflies, stoneflies; (iii) high, body can turn 360 degrees, e.g. Oligochaetes, Chironomidae (Omesová et al. 2008). Preferences for organic substrate were calculated as preferences for pelal (organic mud) and particulate organic matter. For each trait coded in a single-category assignment system or presence/absence assignment system (Schmidt-Kloiber and Hering, 2015), the community-level trait value was calculated as the number of individuals within the trait modality divided by the total macroinvertebrate abundance of a sample. The community-level trait value of traits coded in ten-point assignment system (Schmidt-Kloiber and Hering, 2015) was calculated as the sum of the trait values multiplied by the abundance of each taxon and divided by the total abundance (Schmera et al., 2014).

iii) Community structure metrics

The number of taxa, $\ln(x+1)$ -transformed total abundance, and abundances and proportion in the community of the most abundant higher taxonomic units was calculated for each sample. This was done also for three drying-sensitive taxonomic groups (Bonada et al., 2007; Datry et al., 2014b): the family Heptageniidae, ET taxa (Ephemeroptera and Trichoptera) and EPT taxa (Ephemeroptera, Plecoptera and Trichoptera).

2.5. Index development

We performed exploratory data analysis to inform later identification of metrics that discriminate among intermittence classes. We used dot charts to assess metric distribution, and pair plots to assess their collinearity. We used nonmetric multidimensional scaling (NMDS) of a Bray-Curtis distance matrix calculated using $\ln(x+1)$ -transformed abundance data to visualize variability in community composition in relation to year, season, subprovince and intermittence. We used the adonis function from the vegan package (Oksanen et al., 2017) to run permutational multivariate ANOVA (PERMANOVA; Anderson,

2001) to identify differences in community composition between seasons (*spring, autumn*), subprovinces (*Hercynian, West-Carpathian*) and intermittence categories (*intermittent, near-perennial, perennial*). Because only autumn samples were available for 2012, the difference between years was not tested. One-way ANOVA was used to examine differences in total abundance and taxa richness between perennial, near-perennial, and intermittent samples.

To reduce the number of potential metrics selected for inclusion in the final multimetric index (hereafter, the *Biodrought index*) a pre-selection process was conducted. To identify candidate metrics, values of each potential metric were plotted against intermittence categories. Those that effectively discriminated between intermittent and perennial samples were retained as candidate metrics (Table 1), with Mann-Whitney U tests used to identify differences in metric values between perennial and intermittent samples.

To identify the metric combination that best discriminated among perennial, near-perennial and intermittent sites, we identified linear combinations of candidate metrics using a sequence of linear discriminant analyses (LDA). Because macroinvertebrate community composition differs among seasons (e.g. Straka et al., 2012) we expected different metrics to reflect stream intermittence in autumn and spring. Therefore, all possible combinations of 2-4 metrics were tested for their discriminatory power using the leave-one-out cross-validation procedure (lda function, MASS package; Venables and Ripley, 2002) in each season. The combination of metrics with the best discrimination power was selected and its equation noted. The final index is this equation centred to zero and can be used to calculate a 'discrimination score' for an independent sample. To relate the discrimination score to the intermittence categories, we calculated discrimination scores for all samples and fitted probability density functions of a normal distribution to each category, from which the probability of belonging to each category could be derived. Normality of distribution of LDA discriminant scores for each sample category was tested by Shapiro-Wilk tests. The effects of season and sample category on LDA discriminant score were tested using ANOVA. All statistical analyses and graphs were performed using R open-source software version 3.4.4 (R Core Team, 2018).

2.6. Validation by independent samples

The ability of the index to identify antecedent stream drying was tested on an external dataset, provided by the former Czech Agricultural Water Management Authority, and collected using the same method as the original dataset (Kokeš and Němejcová, 2006). The dataset comprised 117 (59 autumn + 58 spring) samples from 59 perennial sites and 26 (16 autumn + 10 spring) samples from 16 non-perennial sites. We could not distinguish between intermittent and near-perennial sites, and all samples for at which stream drying was observed were therefore classified as non-perennial. Samples were taken from 2nd to 4th order streams distributed across the Czech Republic (Fig. 2), at an altitude of 210 to 460 m a.s.l. The streams were not polluted or morphologically modified and monthly observations of hydrological instream conditions in the summer prior to sampling were available. The value of the Biodrought index was calculated for each sample and the probability of its classification as intermittent, near-perennial or perennial was calculated using the probability density function of the normal distribution. The sample was assigned to the category with the highest probability.

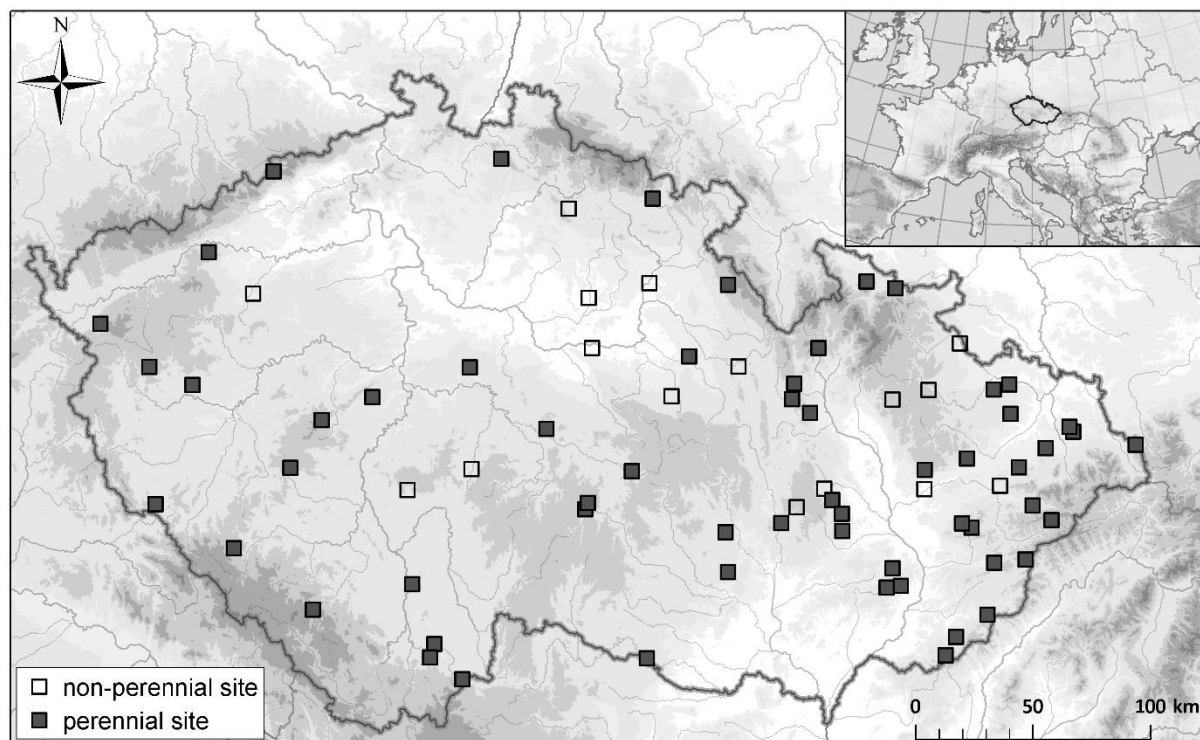


Fig. 2. Location of the sites used for validation within the Czech Republic. Open squares represent non-perennial sites and filled squares represent perennial sites.

3. Results

3.1 Community structure across years, seasons, and regions

In total, 532 552 benthic macroinvertebrate individuals from 421 taxa were collected (Appendix B). Individual sample abundance values ranged from 307 to 57 581 (mean \pm SE, 4997 ± 591) individuals per sample and taxa richness values from 28 to 108 (60.7 ± 1.5) taxa per sample. Macroinvertebrate abundance decreased with increasing flow intermittence (Fig. 3), with significant differences among intermittence categories (one-way ANOVA, $F_{2,104} = 3.71$, $p = 0.028$). Taxa richness was also lowest in intermittent samples ($F_{2,104} = 15.11$, $p < 0.001$). The five most abundant taxa were *Gammarus fossarum* (30% of all individuals), *Micropsectra atrofasciata*-Gr. (9.5%), *Nemoura* sp. (3.5%), *Baetis rhodani* s.l. (3.1%) and *Habroleptoides confusa* (2.9%). Twenty-six taxa were identified as significant indicators of intermittent and near-perennial samples and 33 were identified as significant indicators of perennial samples (Appendix A). The five taxa with the highest IV were *Eiseniella tetraedra*, *Brachyptera risi*, *Parametriocnemus stylatus*, *Paraphaenocladus* sp. and *Marionina* sp. for non-perennial sites and *Dugesia gonocephala*, *Baetis muticus*, *Baetis rhodani* s.l., *Leuctra* sp. and *Hydropsyche* sp. for perennial sites.

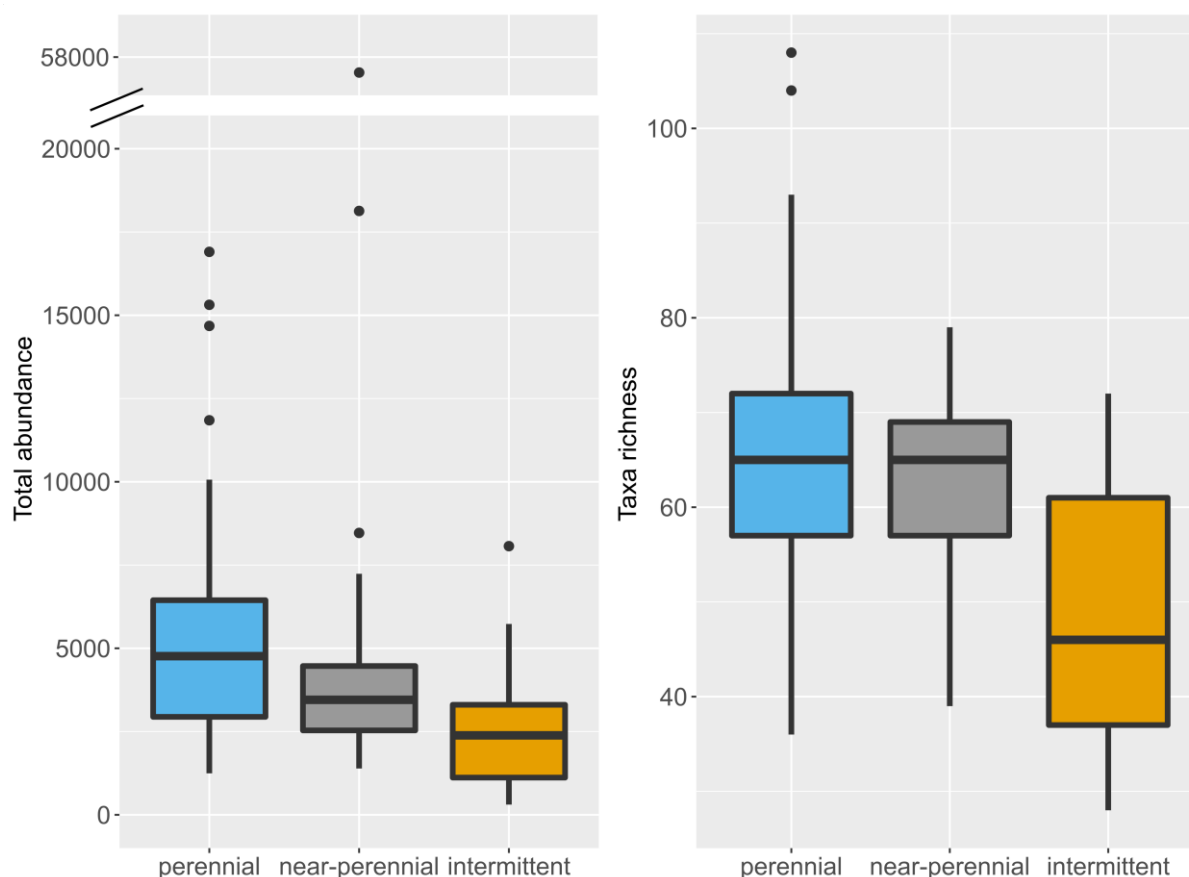


Fig. 3. (colour figure) Total invertebrate abundance (number of individuals) and taxa richness (number of taxa) in samples classified into three flow intermittence categories: i) perennial; ii) near-perennial; iii) intermittent. The box area indicates the first and third quartiles, the central line indicates the median, whiskers represent 95% confidence intervals and circles indicates outliers.

Non-metric multidimensional scaling analyses produced a two-dimensional solution (stress = 0.1998; Fig. 4). Community composition differed among perennial, near-perennial and intermittent samples (PERMANOVA; $F_{2,104} = 7.4$, $p < 0.001$). Despite considerable overlap, we detected differences in community composition both between seasons (PERMANOVA; $F_{1,105} = 6.0$, $p < 0.001$) and subprovinces ($F_{1,105} = 5.6$, $p < 0.001$).

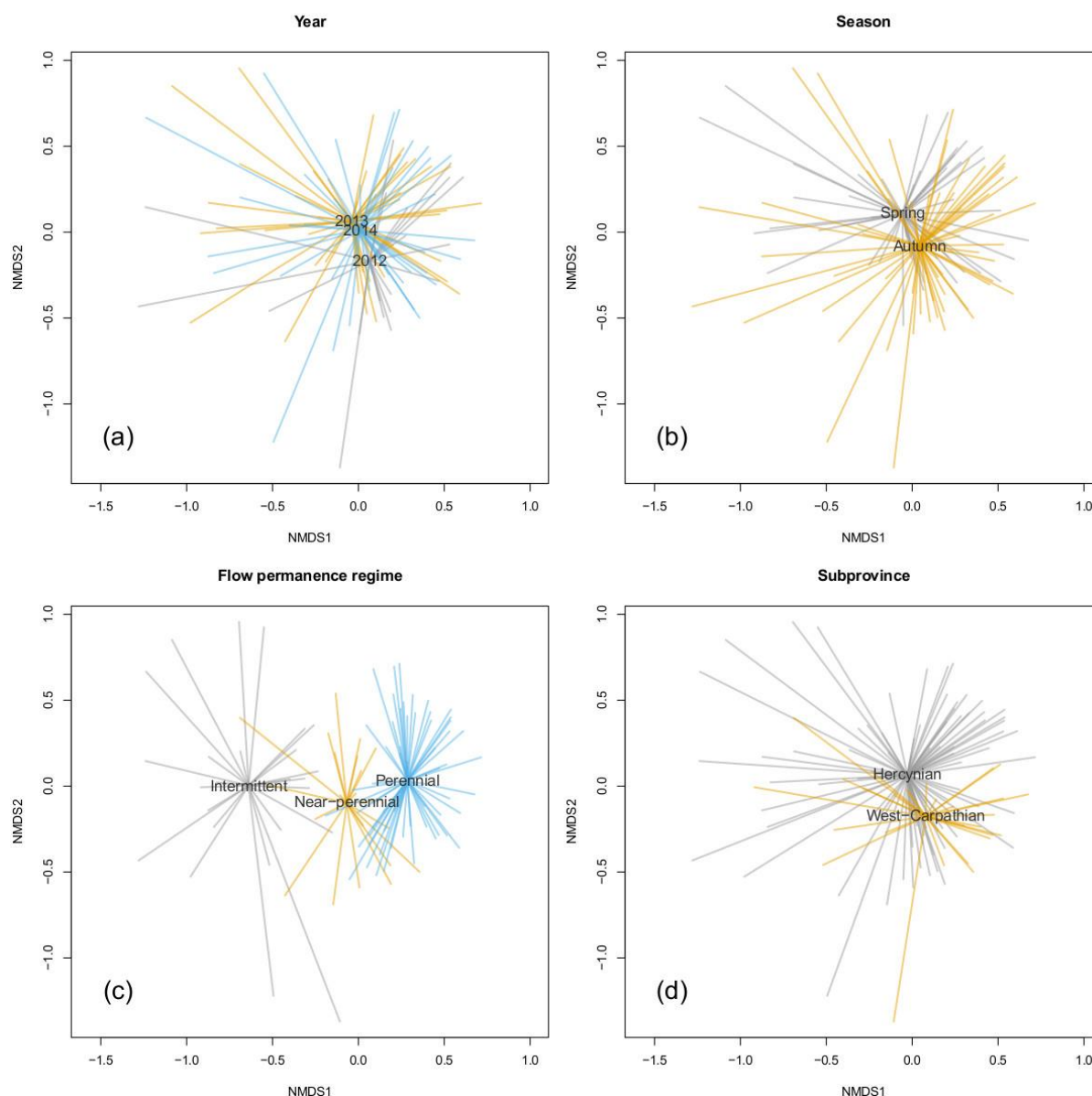


Fig. 4. (colour figure) Non-metric multidimensional scaling ordinations of all samples, showing differences in community composition between: (a) years (2012, 2013, 2014); (b) seasons (autumn, spring); (c) intermittence categories (intermittent, near-perennial, perennial); and (d) biogeographical subprovinces (Hercynian, West-Carpathian). Each point (sample) is connected to the group centroid.

3.2. Discrimination of samples from each intermittence category

Because PERMANOVA identified differences in community composition among perennial, near-perennial and intermittent samples, we investigated the metrics that most effectively discriminated between these intermittence categories. Because many potential metrics differed between intermittent and perennial samples, we plotted all calculated potential metrics and selected a subset of candidate metrics based on their ability to discriminate between categories (Table 1).

Table 1: Candidate metrics tested as discriminators of macroinvertebrate communities from sites in different intermittence categories. Differences between perennial/intermittent samples were tested with Mann-Whitney U-tests. ***<0.001, **<0.01, *<0.05

Metrics selected for index development	Autumn	Spring	Observed state at intermittent sites
% of lithal preference	***	***	lower
% of organic substrate preference	**	**	higher
% of rheophilic preference	***	n.s.	lower
% taxa with respiration with gills	***	***	lower
% taxa with high body flexibility	***	**	higher
% of rhithral preference	***	*	lower
% of epirhithral preference	***	*	lower
Weighted mean of sediment-sensitive invertebrates	***	n.s.	higher
Number of taxa	**	**	lower
Number of ET taxa	***	***	lower
Number of EPT taxa	***	***	lower
Total abundance	***	*	lower
Abundance of Heptageniidae	***	***	lower
Abundance of EPT	***	n.s.	lower
Proportion of Heptageniidae	***	***	lower
Proportion of Oligochaeta	***	n.s.	higher
Proportion of indicator taxa	***	***	lower

From the set of candidate metrics (Table 1), successive LDA revealed important metrics for sample separation. Three metrics were selected for discrimination in autumn: the proportion of indicator taxa, the proportion of individuals with high body flexibility, and the proportion of taxa with a preference for organic substrates. The formula for calculating the autumn index is:

$$\text{Biodrought index} = (4.23 * \text{Proportion_of_indicator_taxa}) + (-1.69 * \text{Proportion_of_high_body_flexibility}) + (-2.94 * \text{Proportion_of_organic sediment preference}) - 0.26$$

Three metrics were also selected as discriminators of spring samples: the proportion of indicator taxa, the proportion of individuals with high body flexibility, and the total abundance. The formula for calculating the spring season index is:

$$\text{Biodrought index} = (4.09 * \text{Proportion_of_indicator_taxa}) + (-1.76 * \text{Proportion_of_high_body_flexibility}) + (0.47 * \text{Total_abundance}) - 4.01$$

The Biodrought index was calculated for each sample (Fig. 5b) and the probability function for perennial, near-perennial and intermittent samples was approximated based on the distribution of index values within the intermittence categories. LDA discriminant scores (i.e. the Biodrought index) for each category were normally distributed (Shapiro-Wilk tests, $p >$

0.05). Mean discriminant scores varied between samples in the three intermittence categories ($F_{2, 101} = 215.9$; $p < 0.001$) but not between spring and autumn samples ($F_{1, 101} = 0.009$; $p = 0.926$), and there was no interaction between intermittence category and season ($F_{2, 101} = 0.328$, $p = 0.721$). Because there was no difference in the distribution of sample discriminant scores between spring and autumn, it was possible to calculate probability functions independently of season (Fig. 5a). Values for the probability function were derived from the dataset for intermittent (mean = -3.36, SD = 1.398), near-perennial (mean = -0.231, SD = 0.926) and perennial (mean = 1.559, SD = 0.777) sites.

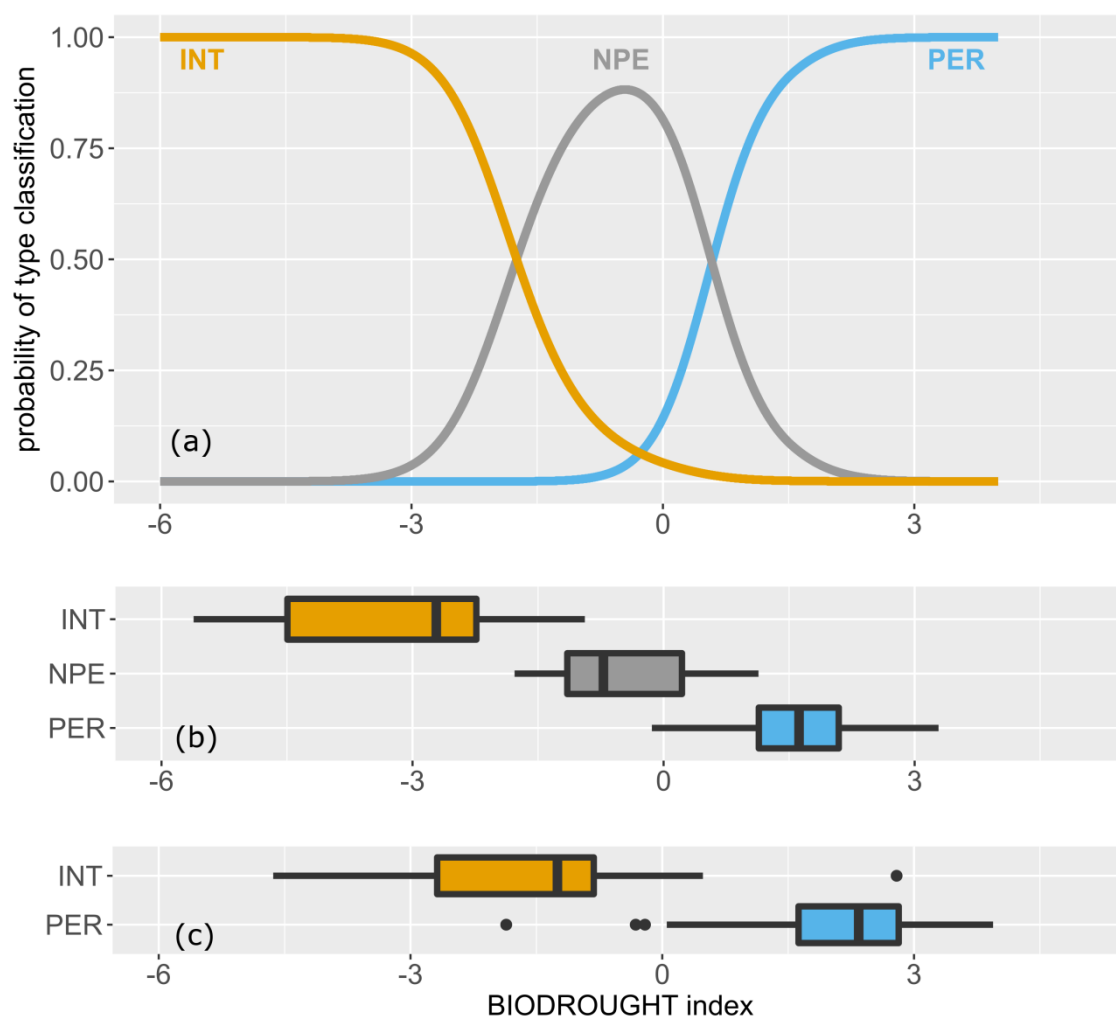


Fig. 5. (colour figure) Biodrought index values for each flow intermittence category: (a) Probability of classification to each category according to the index; index values for each category, for (b) samples used for index development and (c) independent samples used for index validation. INT – intermittent, NPE – near-perennial, PER – perennial.

3.3. Validation by independent samples

To evaluate the performance of the method, we calculated the Biodrought index values for a dataset not used for index development (Fig. 5c). Of 117 perennial samples, 108 (92%) had a probability >50% of being classified correctly by the index, with eight and one samples classified as near-perennial and intermittent, respectively. The index classified 1 of 26

samples from non-perennial streams incorrectly, with 15 and 10 correctly classified as near-perennial and intermittent, respectively. Because it was not possible to distinguish between intermittent and near-perennial categories in this independent dataset, the correct classification of non-perennial samples was 96%.

4. Discussion

Our results agree with previous research demonstrating that benthic macroinvertebrate community composition reflects stream flow intermittence (Arscott et al., 2010; Datry, 2012; Soria et al., 2017) and that the effects of relatively short (7-86 day) drying events can be detected even after seven months after flow resumption (Ledger and Hildrew, 2001; Chester and Robson, 2011). Previous studies from other regions have distinguished perennial and non-perennial streams using the proportional representation of different macroinvertebrate taxonomic groups (Mazzacano and Black, 2009; Nadeau, 2015; Cid et al., 2016; Cañedo-Argüelles et al., 2016) and using traits (Serra et al., 2017; Kelso and Entrekkin, 2018). Our results confirm that macroinvertebrate communities in Central Europe can also be explored to identify stream intermittence, and specifically drying events. Our Biodrought index provides a new tool to identify antecedent drying events, which may facilitate interpretation of ecological data including the results of ecological status assessments, especially when hydrological data are missing. The method was developed and validated using Czech Republic data, but it has considerable potential for wider uptake due to the extensive distribution of the continental climate zone across Central Europe

4.1. Metrics used for index calculations

Biotic responses to environmental stress (e.g. water loss) can be species-specific (Lake, 2003) and we examined available information about responses to flow continuity mainly at the species level, and summarized the most sensitive/tolerant indicator taxa (Appendix A). The proportion of these indicator taxa was one of the most effective metrics to distinguish among intermittence categories. Flow intermittence tolerant taxa usually possess behavioural, morphological, physiological and/or life-history adaptations to survive dry phase (Lytle and Poff, 2004). For example, one species strongly associated with intermittent streams in this study was *Brachyptera risi*. This stonefly has the eggs with summer diapause, allowing the species to remain in the sediments in a viable egg stage during dry phases (Khoo, 1964). However, the selective loss of taxa sensitive to flow intermittence (and specifically drying) rather than selection for desiccation-resistant specialists is the primary driver of differences in community composition between perennial and non-perennial sites (Datry, 2012; Vidal-Abarca et al., 2013; Cid et al., 2016).

Although high body flexibility is not typically reported as a crucial trait in relation to stream drying (e.g. Bonada et al., 2007; Díaz et al., 2008; Walters, 2011) our results indicated its relevance. Organisms with flexible body shapes can respond to drying by moving into the subsurface interstices, which may maintain high humidity and can thus act as refuge for aquatic invertebrates during dry phases (Stubbington, 2012; Strachan et al., 2015). Moreover, a highly flexible body enables organisms including oligochaetes and leeches to form globular, desiccation-tolerant cysts (Montalto and Marchese, 2005; Shikov, 2011). Intermittent streams also supported a higher proportion of invertebrates with a preference

for organic substrates, which may reflect their dry-phase survival in moisture-retaining patches of organic matter (Stubbington et al., 2009). Invertebrates that use such organic-rich habitats during flowing conditions can therefore persist in this substrate, which acts as a refuge that limits desiccation during dry phases (Boulton, 1989).

The Biodrought index calculated for the spring season also included total abundance. The relationship between total abundance and flow permanence in streams has previously been indicated by several studies (Rüegg and Robinson, 2004; Fenoglio et al., 2007; Datry, 2012). The lower abundance in our study was caused in particular by high mortality of *Gammarus* species, known to be sensitive to stream drying (Meyer and Meyer, 2000; Smith et al., 2003; Pařil et al., 2019).

4.2. Utility of the developed tool

Our index addresses the recognized need (Datry et al., 2011; Cid et al., 2016) for methods to identify antecedent drying events using biological metrics, and thus without using gauging stations or other prohibitively costly infrastructure. Even where gauging station data are available, biotic approaches such as our index complement the continuous, long-term hydrological information by distinguishing dry from lentic zero-flow states (Gallart et al., 2012). Sampling and processing of benthic invertebrate samples is methodologically well-managed and is routinely practised within the scope of monitoring programmes (Smith et al., 1999; Birk et al., 2012; Hill et al., 2017). The method used for taking samples (Kokeř et al., 2006) is comparable to other commonly used European methods such as AQEM/STAR method (Lorenz and Clarke, 2006), and the Biodrought index therefore has high potential for testing and use in other countries within the extensive continental climate zone that spans Central and Eastern Europe. The study dataset spans two biogeographical subprovinces (Hercynian and West-Carpathian; Culek, 2013) and the discriminatory power of the index was sufficient to correctly classify evaluated samples from the independent dataset from whole Czech Republic with high probability. Although working at a finer geographical resolution (e.g. within one subprovince) may achieve higher levels of correct assignment, we also recommend testing of our index in other Central European countries to explore the geographical limits of its reliability. A saprobic system based on common taxa is used in many Central and Eastern European countries (Rolauuffs et al., 2004) and the Biodrought index may therefore be useable in Central European region without extensive taxonomic adjustment.

Our index was developed in pristine streams where only a single, natural stressor (drying) was known to influence macroinvertebrate community composition. Elsewhere, factors that may affect index performance include interactions between the hydrological stressor of drying (both natural and anthropogenic) with other human pressures, interactions which can have antagonistic or additive effects on biological communities (Folt et al., 1999; Matthaei et al., 2010; Ormerod et al., 2010). In particular, the typical sequence of hydrological conditions from low flows, to flow cessation and gradual drying can reduce oxygen concentrations, and intermittence and organic pollution can therefore have comparable effects by eliminating sensitive invertebrates (Pardo and Garcia, 2016). Moreover, a reduction in oxygen availability and low flow can have synergistic effects on benthic invertebrates (Calapez et al., 2017). Disentangling the effects of stream drying and anthropogenic stressors (including

organic pollution) is therefore challenging, but consideration of the Biodrought index alongside physico-chemical quality elements may improve the accuracy of ecological status assessments that might otherwise incorrectly classify IRES as polluted. Wider testing is needed to explore how the index performs in streams exposed to human stressors.

The spatial distribution of intermittent reaches in relation to perennial reaches and other refuges may have a considerable influence on the rate and extent of community recovery after drying events (Sedell et al., 1990; Bogan et al., 2017; but see Datry et al., 2014b). The spatial proximity of extensive refuges, in particular upstream perennial reaches, may enable rapid recolonization and community reestablishment within weeks to obscure the occurrence of an antecedent dry phase (Fowler, 2004; Fritz and Dodds, 2004; Pařil et al., 2019). In our dataset, the intermittent sites were at least 0.5 km downstream of a perennial reach and the samples were taken at most 7 months after flow resumed. However, recovery at highly isolated sites (e.g. >10 km from perennial refuges) may take multiple years with repeated drying events often preventing aquatic communities from reaching a stable state (Bogan and Lytle, 2011; Bogan et al., 2017). Our index and comparable tools thus have higher potential for strong performance in IRES with relatively short dry phases, compared to those which may remain dry for years.

4.3. Conclusions and recommendations for index use

We developed the Biodrought index: a novel index to characterize the effect of stream drying on aquatic invertebrate communities. The index enable calculation of the probability of the antecedent stream drying, based on benthic invertebrates sampled using standard methods. We demonstrated the index as robust at national scale. The robustness of our index is evidenced by its registration by the Ministry of Environment of the Czech Republic, and we recommend its testing and adoption (with required adaptation) in other European countries within continental-climate region. In particular, the Biodrought index is intended for use by those with responsibility for monitoring and management of river ecosystems, including water management boards, environmental agencies and private water companies. We recommend adoption of the Biodrought index by such managers, to improve their interpretation of the macroinvertebrate assemblage data collected during ecological status assessments. The index can help avoid misinterpreting a deviation of community composition from reference status caused by a preceding dry phase (Reyjol et al., 2014; Cid et al., 2016; Stubbington et al., 2018). Environmental agencies could employ this tool for characterization of dry phase effects on macroinvertebrate communities in protected areas or for evaluation of the effectiveness of projects intended to restore naturally perennial flow at sites impacted by water resource pressures such as over-abstraction. The index can also be useful when identifying the impacts of water resource use in naturally perennial streams. However, since the method was developed in naturally intermittent streams, its use in non-natural IRES must be carefully assessed and the performance of Biodrought index in such conditions remains to be tested.

Our approach was essentially qualitative, identifying if a drying event had or had not occurred in the the one-year period preceding sample collection. Further research is needed to explore variability within 'intermittent', 'near-perennial' and other flow intermittence categories, in particular to identify species-specific responses to dry phases of differing

457 durations. Identification of such thresholds is crucial to predict biotic responses to increasing
 458 intermittence in the context of ongoing climate change, which is interacting with increasing
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469 6. References

- 470 Acuña, V., Datry, T., Marshall, J., Barceló, D., Dahm, C.N., Ginebreda, A., McGregor, G.,
 471 Sabater, S., Tockner, K., Palmer, M.A., 2014. Why should we care about temporary
 472 waterways? *Science* 343, 1080-1081. <https://doi.org/10.1126/science.1246666>.
- 473 Anderson, M. J., 2001. A new method for non-parametric multivariate analysis of variance.
 474 *Austral Ecology* 26, 32-46. <https://doi.org/10.1111/j.1442-9993.2001.01070.pp.x>
- 475 Arscott, D.B., Larned, S., Scarsbrook, M.R., Lambert, P., 2010. Aquatic invertebrate
 476 community structure along an intermittence gradient: Selwyn River, New Zealand. *J. N.*
 477 *Am. Benthol. Soc.* 29, 530-545. <https://doi.org/10.1899/08-124.1>.
- 478 Beaufort, A., Lamouroux, N., Pella, H., Datry, T., Sauquet, E., 2018. Extrapolating regional
 479 probability of drying of headwater streams using discrete observations and gauging
 480 networks. *Hydrol. Earth Syst. Sci.* 22: 3033-3051. [https://www.hydrol-earth-syst-](https://www.hydrol-earth-syst-sci.net/22/3033/2018/)
 481 [sci.net/22/3033/2018/](https://www.hydrol-earth-syst-sci.net/22/3033/2018/).
- 482 Birk, S., Bonne, W., Borja, A., Brucet, S., Courrat, A., Poikane, S., Solimini, A., Van De Bund,
 483 W., Zampoukas, N., Hering, D., 2012. Three hundred ways to assess Europe's surface
 484 waters: an almost complete overview of biological methods to implement the Water
 485 Framework Directive. *Ecological Indicators* 18, 31-41.
 486 <https://doi.org/10.1016/j.ecolind.2011.10.009>.
- 487 Bogan, M.T., Lytle, D. A., 2011. Severe drought drives novel community trajectories in desert
 488 stream pools. *Freshwater Biology* 56, 2070-2081. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2427.2011.02638.x)
 489 [2427.2011.02638.x](https://doi.org/10.1111/j.1365-2427.2011.02638.x)
- 490 Bogan, M.T., Boersma, K.S., Lytle, D.A., 2013. Flow intermittency alters longitudinal patterns
 491 of invertebrate diversity and assemblage composition in an arid-land stream network.
 492 *Freshwater Biology* 58: 1016-1028. <https://doi.org/10.1111/fwb.12105>.
- 493 Bogan, M.T., Chester, E.T., Datry, T., Murphy, A.L., Robson, B.J., Ruhi, A., Stubbington, R.,
 494 Whitney, J.E., 2017. Resistance, resilience, and community recovery in intermittent rivers
 495 and ephemeral streams, in: Datry, T., Bonada, N., Boulton, A. (Eds.), *Intermittent rivers*
 496 *and ephemeral streams. Ecology and management. Elsevier, Netherlands*, pp. 349-376.

- 497 Bonada, N., Rieradevall, M., Prat, N., 2007. Macroinvertebrate community structure and
 498 biological traits related to flow permanence in a Mediterranean river network.
 499 *Hydrobiologia* 589, 91-106. <https://doi.org/10.1007/s10750-007-0723-5>.
- 500 Boulton, A.J., 1989. Over-summer refuges of aquatic macroinvertebrates in two intermittent
 501 streams in central Victoria. *Transactions of the Royal Society of South Australia* 113: 22 –
 502 34.
- 503 Calapez, A.R., Branco, P., Santos, J.M., Ferreira, T., Hein, T., Brito, A.G., Feio, M.J., 2017.
 504 Macroinvertebrate short-term responses to flow variation and oxygen depletion: A
 505 mesocosm approach. *Science of the Total Environment* 599-600, 1202-1212.
 506 <https://doi.org/10.1016/j.scitotenv.2017.05.056>
- 507 Cañedo-Argüelles, M., Bogan, M.T., Lytle, D.A., Prat, N., 2016. Are Chironomidae (Diptera)
 508 good indicators of water scarcity? Dryland streams as a case study. *Ecological Indicators*
 509 71, 155-162. <https://doi.org/10.1016/j.ecolind.2016.07.002>
- 510 Chadd, R.P., England, J.A., Constable, D., Dunbar, M.J., Extence, Ch.A., Leeming, D.J., Murray-
 511 Bligh, J.A., Wood, P.J., 2017. An index to track the ecological effects of drought
 512 development and recovery on riverine invertebrate communities. *Ecological Indicators*
 513 82, 344-356. <http://dx.doi.org/10.1016/j.ecolind.2017.06.058>
- 514 Chester, E.T., Robson, B.J., 2011. Drought refuges, spatial scale and recolonisation by
 515 invertebrates in non-perennial streams. *Freshwater Biology* 56, 2094-2104.
 516 <https://doi.org/10.1111/j.1365-2427.2011.02644.x>
- 517 Cid, N., Verkaik, I., García-Roger, E.M., Rieradevall, M., Bonada, N., Sánchez-Montoya, M.M.,
 518 Gómez, R., Suárez, M.L., Vidal-Abarca, M.R., Demartini, D., Buffagni, A., Erba, S.,
 519 Karaouzas, I., Skoulidakis, N., Prat, N., 2016. A biological tool to assess flow connectivity in
 520 reference temporary streams from the Mediterranean Basin. *Science of the Total*
 521 *Environment* 540, 178-190. <https://doi.org/10.1016/j.scitotenv.2015.06.086>
- 522 Culek, M., 2013. Biogeographical provinces, subprovinces and bioregions of the Czech
 523 Republic. *Journal of Landscape Ecology* 6, 5-16. [https://doi.org/10.2478/v10285-012-](https://doi.org/10.2478/v10285-012-0065-5)
 524 [0065-5](https://doi.org/10.2478/v10285-012-0065-5)
- 525 Datry, T., Arscott, D.B., Sabater, S., 2011. Recent perspectives on temporary river ecology.
 526 *Aquatic Sciences* 73, 453-457. <https://doi.org/10.1007/s00027-011-0236-1>
- 527 Datry, T., 2012. Benthic and hyporheic invertebrate assemblages along a flow intermittence
 528 gradient: effects of duration of dry events. *Freshwater Biology* 57: 563-574.
 529 <https://doi.org/10.1111/j.1365-2427.2011.02725.x>
- 530 Datry, T., Larned, S.T., Tockner, K., 2014a. Intermittent rivers: A challenge for freshwater
 531 ecology. *BioScience* 64, 229-235. <https://doi.org/10.1093/biosci/bit027>
- 532 Datry, T., Larned, S.T., Fritz, K. M., Bogan, M.T., Wood, P.J., Meyer, E.I., Santos, A.N., 2014b.
 533 Broad-scale patterns of invertebrate richness and community composition in temporary
 534 rivers: effects of flow intermittence. *Ecography* 37, 69-104.
 535 <https://doi.org/10.1111/j.1600-0587.2013.00287.x>
- 536 Datry, T., Boulton, A.J., Bonada, N., Fritz, K., Leigh, C., Sauquet, E., Tockner, K., Hugueny, B.,
 537 Dahm, C.N., 2017. Flow intermittence and ecosystem services in rivers of the
 538 Anthropocene. *Journal of Applied Ecology* 55, 353-364. [https://doi.org/10.1111/1365-](https://doi.org/10.1111/1365-2664.12941)
 539 [2664.12941](https://doi.org/10.1111/1365-2664.12941)
- 540 De Cáceres, M., Legendre, P., 2009. Associations between species and groups of sites:
 541 indices and statistical inference. *Ecology* 90, 3566–3574, [http://dx.doi.org/10.1890/08-](http://dx.doi.org/10.1890/08-1823.1)
 542 [1823.1](http://dx.doi.org/10.1890/08-1823.1).

- 543 Díaz, A.M., Alonso, M.L.S., Gutiérrez, M.R.V.-A., 2008. Biological traits of stream
544 macroinvertebrates from a semi-arid catchment: patterns along complex environmental
545 gradients. *Freshwater Biology* 53, 1-21. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2427.2007.01854.x)
546 [2427.2007.01854.x](https://doi.org/10.1111/j.1365-2427.2007.01854.x)
- 547 Dufrêne, M., Legendre, P., 1997. Species assemblages and indicator species: The need for a
548 flexible asymmetrical approach. *Ecological Monographs* 67, 345-366. DOI:
549 10.2307/2963459
- 550 European Commission, 2000. Directive 2000/60/EC of the European Parliament of the
551 Council of 23rd October 2000 establishing a framework for community action in the field
552 of water policy. *Official J. Eur. Commun.* 327, 1-72.
- 553 Extence, C.A., Chadd, R.P., England, J., Dunbar, M.J., Wood, P.J., Taylor, E.D., 2013. The
554 assessment of fine sediment accumulation in rivers using macro-invertebrate community
555 response. *River Res. Applic.* 29, 17-55. <https://doi.org/10.1002/rra.1569>
- 556 Fenoglio, S., Bo, T., Cucco, M., Malacarne, G., 2007. Response of benthic invertebrate
557 assemblages to varying drought conditions in the Po river (NW Italy). *Italian Journal of*
558 *Zoology* 74, 191-201. <https://doi.org/10.1080/11250000701286696>
- 559 Folt, C.L., Chen, C.Y., Moore, M.V., Burnaford, J., 1999. Synergism and antagonism among
560 multiple stressors. *Limnology and oceanography* 44, 864-877.
561 https://doi.org/10.4319/lo.1999.44.3_part_2.0864
- 562 Fowler, R.T., 2004. The recovery of benthic invertebrate communities following dewatering
563 in two braided rivers. *Hydrobiologia* 523: 17-28.
564 <https://doi.org/10.1023/B:HYDR.0000033077.13139.7f>
- 565 Fritz, K.M., Dodds, W.K., 2004. Resistance and resilience of macroinvertebrate assemblages
566 to drying and flood in a tallgrass prairie stream system. *Hydrobiologia* 527, 99-112.
567 <https://doi.org/10.1023/B:HYDR.0000043188.53497.9b>
- 568 Gallart, F., Prat, N., García-Roger, E.M., Latron, J., Rieradevall, M., Llorens, P., Barberá, G.G.,
569 Brito, D., De Girolamo, A.M., Lo Porto, A., Buffagni, A., Erba, S., Neves, R., Nikolaidis, N.P.,
570 Perrin, J.L., Querner, E.P., Quiñonero, J.M., Tournoud, M.G., Tzoraki, O., Skoulidakis, N.,
571 Gómez, R., Sánchez-Montoya, M.M., Froebrich, J., 2012. A novel approach to analysing
572 the regimes of temporary streams in relation to their controls on the composition and
573 structure of aquatic biota. *Hydrol. Earth Syst. Sci.* 16, 3165-3182.
574 <https://doi.org/10.5194/hess-16-3165-2012>
- 575 Gallart, F., Llorens, P., Latron, J., Cid, N., Rieradevall, M., Prat, N., 2016. Validating alternative
576 methodologies to estimate the regime of temporary rivers when flow data are
577 unavailable. *Science of the Total Environment* 565, 1001-1010.
578 <https://doi.org/10.1016/j.scitotenv.2016.05.116>
- 579 Hill, R.A., Fox, E.W., Leibowitz, S.G., Olsen, A.R., Thornbrugh, D.J., Weber, M.H., 2017.
580 Predictive mapping of the biotic condition of conterminous U.S. rivers and streams.
581 *Ecological Applications* 27, 2397-2415. <https://doi.org/10.1002/eap.1617>
- 582 Hille, S., Kristensen, E.A., Graeber, D., Riis, T., Jørgensen, N.K., Baattrup-Pedersen, A., 2014.
583 Fast reaction of macroinvertebrate communities to stagnation and drought in streams
584 with contrasting nutrient availability. *Freshwater Science* 33, 847-859.
585 <https://doi.org/10.1086/677554>
- 586 Illies, J. (ed.), 1967. *Limnofauna Europaea*. Eine Zusammenstellung aller die europäischen
587 Binnengewässer bewohnenden mehrzelligen Tierarten m. Angaben über d. Verbreitung u.
588 Ökologie. 1st Edition, Gustav Fischer Verlag, Stuttgart, 474pp.

- 589 IS ARROW (Assessment and Reference Reports of Water Monitoring), 2014, Praha, Český
590 hydrometeorologický ústav. <http://hydro.chmi.cz/isarrow/> (accessed 20.06.2014).
- 591 Kelso, J.E., Entekin, S.A., 2018. Intermittent and perennial macroinvertebrate communities
592 had similar richness but differed in species trait composition depending on flow duration.
593 *Hydrobiologia* 807, 189-206. doi: 10.1007/s10750-017-3393-y
- 594 Khoo, S.G., 1964. Studies on the biology of stoneflies. Unpublished Ph.D. thesis, University of
595 Liverpool.
- 596 Kokeš, J., Němejcová, D., 2006. Metodika odběru a zpracování vzorků makrozoobentosu
597 tekoucích vod metodou PERLA [Methodology for macrozoobenthos sampling in running
598 waters by PERLA method]. Metodika VÚV TGM Praha.
- 599 Kokeš, J., Zahradková, S., Němejcová, D., Hodovský, J., Jarkovský, J., Soldán, T., 2006. The
600 PERLA system in the Czech Republic: A multivariate approach to assess ecological status
601 of running waters. *Hydrobiologia* 566, 343–354. [https://doi.org/10.1007/978-1-4020-](https://doi.org/10.1007/978-1-4020-5493-8_24)
602 [5493-8_24](https://doi.org/10.1007/978-1-4020-5493-8_24)
- 603 Lake, P.S., 2003. Ecological effects of perturbation by drought in flowing waters. *Freshwater*
604 *Biology* 48, 1161-1172. <https://doi.org/10.1046/j.1365-2427.2003.01086.x>
- 605 Lancaster, J., Ledger, M.E., 2015. Population-level responses of stream macroinvertebrates
606 to drying can be density-independent or density-dependent. *Freshwater Biology* 60,
607 2559-2570. <https://doi.org/10.1111/fwb.12643>
- 608 Ledger, M.E., Hildrew, A.G., 2001. Recolonization by the benthos of an acid stream following
609 a drought. *Arch. Hydrobiol.* 152, 1-17. DOI: 10.1127/archiv-hydrobiol/152/2001/1
- 610 Leigh, C., Datry, T., 2017. Drying as a primary hydrological determinant of biodiversity in river
611 systems: a broad-scale analysis. *Ecography* 40, 487-499.
612 <https://doi.org/10.1111/ecog.02230>
- 613 Leigh, C., Bonada, N., Boulton, A.J., Hugueny, B., Larned, S.T., Vander Vorste, R., Datry, T.,
614 2016. Invertebrate assemblage responses and the dual roles of resistance and resilience
615 to drying in intermittent rivers. *Aquatic Sciences*, 78, 291-301.
616 <https://doi.org/10.1007/s00027-015-0427-2>
- 617 Li, L., Zheng, B., Liu, L., 2010. Biomonitoring and bioindicators used for river ecosystems:
618 Definitions, approaches and trends. *Procedia Environmental Sciences* 2, 1510-1524.
619 <https://doi.org/10.1016/j.proenv.2010.10.164>
- 620 Lorenz, A., Clarke, R.T., 2006. Sample coherence – a field study approach to assess similarity
621 of macroinvertebrate samples. *Hydrobiologia* 566, 461-476. [https://doi.org/10.1007/978-](https://doi.org/10.1007/978-1-4020-5493-8_32)
622 [1-4020-5493-8_32](https://doi.org/10.1007/978-1-4020-5493-8_32)
- 623 Lytle, D.A., Poff, N.L., 2004. Adaptation to natural flow regimes. *TRENDS in Ecology and*
624 *Evolution* 19, 94-100. <https://doi.org/10.1016/j.tree.2003.10.002>.
- 625 Marshall, J.C., Acuña, V., Allen, D.C., Bonada, N., Boulton, A.J., Carlson, S.M., Dahm, C.N.,
626 Datry, T., Leigh, C., Negus, P., Richardson, J.S., 2018. Protecting U.S. temporary
627 waterways. *Science*, 361, 856-857. DOI: 10.1126/science.aav0839
- 628 Matthaei, C.D., Piggott, J.J., Townsend, C.R., 2010. Multiple stressors in agricultural streams:
629 interactions among sediment addition, nutrient enrichment and water
630 abstraction. *Journal of Applied Ecology*, 47, 639-649. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2664.2010.01809.x)
631 [2664.2010.01809.x](https://doi.org/10.1111/j.1365-2664.2010.01809.x)
- 632 Mazzacano, C., Black, S.H., 2009. Using Aquatic Macroinvertebrates as Indicators of Stream
633 Flow Duration. The Xerces Society for Invertebrate Conservation, Portland, USA.

- Menció, A., Mas-Pla, J., 2010. Influence of groundwater exploitation on the ecological status of streams in a Mediterranean system (Selva Basin, NE Spain). *Ecological Indicators* 10, 915-926. <https://doi.org/10.1016/j.ecolind.2010.02.001>
- Meyer, A., Meyer, E. I., 2000. Discharge regime and the effect of drying on macroinvertebrate communities in a temporary karst stream in East Westphalia (Germany). *Aquatic Sciences* 62, 216-231. <https://doi.org/10.1007/PL00001333>
- Montalto, L., Marchese, M., 2005. Cyst formation in Tubificidae (Naidinae) and Opistocystidae (Annelida, Oligochatea) as an adaptive strategy for drought tolerance in fluvial wetlands of the Paraná River, Argentina. *Wetlands* 25, 488-494. <https://doi.org/10.1672/23>.
- Munné, A., Prat, N., 2009. Use of macroinvertebrate-based multimetric indices for water quality evaluation in Spanish Mediterranean rivers: an intercalibration approach with IBMWP index. *Hydrobiologia* 628, 203-225. <https://doi.org/10.1007/s10750-009-9757-1>
- Nadeau, T.-L., 2015. Streamflow duration assessment method for the Pacific northwest. US Environmental Protection Agency, region 10, Seattle, WA.
- NC Division of Water Quality, 2010. Methodology for Identification of Intermittent and Perennial Streams and their Origins, Version 4.11. North Carolina Department of Environment and Natural Resources, Division of Water Quality. Raleigh, NC.
- Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P.R., O'Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., Wagner, H., 2017. *vegan: Community Ecology Package*. R package version 2.4-3. <https://CRAN.R-project.org/package=vegan>
- Omesová, M., Horsák, M., Helešic, J., 2008. Nested patterns in hyporheic meta-communities: the role of body morphology and penetrability of sediment. *Naturwissenschaften* 95, 917-926. <https://doi.org/10.1007/s00114-008-0399-3>
- Ormerod, S.J., Dobson, M., Hildrew, A.G., Townsend, C., 2010. Multiple stressors in freshwater ecosystems. *Freshwater Biology* 55, 1-4. <https://doi.org/10.1111/j.1365-2427.2009.02395.x>
- Pardo, I., García, I., 2016. Water abstraction in small lowland streams: Unforeseen hypoxia and anoxia effects. *Science of the Total Environment* 568, 226-235. <https://doi.org/10.1016/j.scitotenv.2016.05.218>
- Pařil, P., Leigh, C., Polášek, M., Sarremejane, R., Řezníčková, P., Dostálová, A., Stubbington, R., 2019. Short-term streambed drying events alter amphipod population structure in a central European stream. *Fundamental and Applied Limnology*. <https://doi.org/10.1127/fal/2019/1164>
- Peel, M.C., Finlayson, B.L., McMahon, T.A., 2007. Updated world map of the Köppen–Geiger climate classification. *Hydrol. Earth Syst. Sci.* 11, 1633–1644. <https://doi.org/10.5194/hess-11-1633-2007>
- Poff, N.L., 1997. Landscape filters and species traits: towards mechanistic understanding and prediction in stream ecology. *J. N. Am. Benthol. Soc.* 16, 391-409. <https://doi.org/10.2307/1468026>
- Reyjol, Y., Argillier, Ch., Bonne, W., Borja, A., Buijse, A.D., Cardoso, A.C., Daufresne, M., Kernan, M., Ferreira, M.T., Poikane, S., Prat, N., Solheim, A.-L., Stroffek, S., Usseglio-Polatera, P., Villeneuve, B., van de Bund, W., 2014. Assessing the ecological status on the context of the European Water Framework Directive: Where do we go now? *Science of Total Environment* 497-498, 332-344. <https://doi.org/10.1016/j.scitotenv.2014.07.119>

- 680 Rolauffs, P., Stubauer, I., Zahrádková, S., Brabec, K., Moog, O., 2004. Integration of the
681 saprobic system into the European Union Water Framework Directive. *Hydrobiologia* 516,
682 285-298.
- 683 R Core Team, 2018. R: A language and environment for statistical computing. Retrieved from
684 <http://www.R-project.org/>.
- 685 Rosenberg, D.M., Resh, V.H. (Eds.), 1993. *Freshwater biomonitoring and benthic*
686 *macroinvertebrates*. Chapman and Hall, New York.
- 687 Rüegg, J., Robinson, Ch.T., 2004. Comparison of macroinvertebrate assemblages of
688 permanent and temporary streams in an Alpine flood plain, Switzerland. *Arch. Hydrobiol.*
689 161, 489-510. DOI: 10.1127/0003-9136/2004/0161-0489
- 690 Schmera, D., Podani, J., Erös, T., Heino, J., 2014. Combining taxon-by-trait and taxon-by-site
691 matrices for analysing trait patterns of macroinvertebrate communities: a rejoinder to
692 Monaghan & Soares (2014). *Freshwater Biology* 59, 1551-1557.
693 <https://doi.org/10.1111/fwb.12369>
- 694 Schmidt-Kloiber, A., Hering, D., 2015. www.freshwaterecology.info – An online tool that
695 unifies, standardises and codifies more than 20, 000 European freshwater organisms and
696 their ecological preferences. *Ecological Indicators* 53: 271-282.
697 <https://doi.org/10.1016/j.ecolind.2015.02.007>
- 698 Schriever, T.A., Bogan, M.T., Boersma, K.S., Cañedo-Argüelles, M., Jaeger, K.L., Olden, J.D.,
699 Lytle, D.A., 2015. Hydrology shapes taxonomic and functional structure of desert stream
700 invertebrate communities. *Freshwater Science* 34, 399-409. <https://doi.org/10.1086/680518>
- 701 Sedell, J.R., Reeves, G.H., Hauer, F.R., Stanford, J.A., Hawkins, Ch.P., 1990. Role of refugia in
702 recovery from disturbances: Modern fragmented and disconnected river systems.
703 *Environmental Management* 14, 711-724. <https://doi.org/10.1007/BF02394720>
- 704 Serra, S.R.Q., Graça, M.A.S., Dolédec, S., Feio, M.J., 2017. Discriminating permanent from
705 temporary rivers with traits of chironomid genera. *Ann. Limnol. – Int. J. Lim.* 53: 161-174.
<https://doi.org/10.1051/limn/2017004>
- 706 Shikov, E.V., 2011. *Haemopsis sanguisuga* (Linnaeus, 1758) (Hirudinea) – The first observation
707 of a leech predation on terrestrial gastropods. *Folia Malacologica* 19, 103-106.
708 <http://dx.doi.org/10.2478/v10125-011-0016-5>
- 709 Smith, M.J., Kay, W.R., Edward, D.H.D., Papas, P.J., Richardson, K.StJ., Simpson, J.C., Pinder,
710 A.M., Cale, D.J., Horwitz, P.H.J., Davis, J.A., Yung, F.H., Norris, R.H., Halse, S.A., 1999.
711 AusRivAS: using macroinvertebrates to asses ecological condition of rivers in Western
712 Australia. *Freshwater Biology* 41, 269-282. <https://doi.org/10.1046/j.1365-2427.1999.00430.x>
- 714 Smith, H., Wood, P.J., Gunn, J., 2003. The influence of habitat structure and flow
715 permanence on invertebrate communities in karst spring systems. *Hydrobiologia* 510, 53-
716 66. <https://doi.org/10.1023/B:HYDR.00000008501.55798.20>
- 717 Soria, M., Leigh, C., Datry, T., Bini, L.M., Bonada, N., 2017. Biodiversity in perennial and
718 intermittent rivers: a meta-analyses. *Oikos* 126, 1078-1089.
719 <https://onlinelibrary.wiley.com/doi/full/10.1111/oik.04118>
- 720 Strachan, S.R., Chester, E.T., Robson, B.J., 2015. Freshwater invertebrate life history
721 strategies for surviving desiccation. *Springer Science Reviews* 3, 57-75.
722 <https://doi.org/10.1007/s40362-015-0031-9>
- 723 Straka, M., Syrovátka, V., Helešic, J., 2012. Temporal and spatial macroinvertebrate variance
724 compared: crucial role of CPOM in a headwater stream. *Hydrobiologia* 686, 119-134.
725 <https://doi.org/10.1007/s10750-012-1003-6>

- Stubbington, R., Wood, P.J., Boulton, A.J., 2009. Low flow controls on benthic and hyporheic macroinvertebrate assemblages during supra-seasonal drought. *Hydrological Processes* 23, 2252-2263. <https://doi.org/10.1002/hyp.7290>
- Stubbington, R., 2012. The hyporheic zone as an invertebrate refuge: a review of variability in space, time, taxa and behaviour. *Freshwater Research*, 63, 293-311. <http://dx.doi.org/10.1071/MF11196>
- Stubbington, R., England, J., Wood, P.J., Sefton, C.E.M., 2017. Temporary streams in temperate zones: recognizing, monitoring and restoring transitional aquatic-terrestrial ecosystems. *Wiley Interdisciplinary Reviews, Water* 2017, 4, Article e1223 <https://doi.org/10.1002/wat2.1223>
- Stubbington, R., Chadd, R., Cid, N., Csabai, Z., Miliša, M., Morais, M., Munné, A., Pařil, P., Peřić, V., Tziortzis, I. Verdonschot, R.C.M., Datry, T., 2018. Biomonitoring of intermittent rivers and ephemeral streams in Europe: Current practice and priorities to enhance ecological status assessments. *Science of the Total Environment* 618, 1096-1113. DOI: 10.1016/j.scitotenv.2017.09.137
- Usseglio-Polatera, P., Tachet, H., 1994. Theoretical habitat templates, species traits, and species richness: Plecoptera and Ephemeroptera in the Upper Rhône River and its floodplain. *Freshwater Biology* 31, 417-437. <https://doi.org/10.1111/j.1365-2427.1994.tb01749.x>
- Venables, W.N., Ripley, B.D., 2002. *Modern Applied Statistics with S*. Fourth Edition. Springer, New York. ISBN 0-387-95457-0.
- Vidal-Abarca, M.R., Sánchez-Montoya, M.M., Guerrero, C., Gómez, R., Arce, M.I., García-García, V., Suárez, M.L., 2013. Effects of intermittent stream flow on macroinvertebrate community composition and biological traits in a naturally saline Mediterranean stream. *J. Arid Environ.* 99, 28–40. <https://doi.org/10.1016/j.jaridenv.2013.09.008>
- Walters, A.W., 2011. Resistance of aquatic insects to a low-flow disturbance: exploring a trait-based approach. *Freshwater Science* 30, 346-357. <https://doi.org/10.1899/10-041.1>
- Wilby, R.L., Orr, H., Watts, G., Battarbee, R.W., Berry, P.M., Chadd, R., Dugdale, S.J., Dunbar, M.J., Elliott, J.A., Extence, C., Hannah, D.M., Holmes, N., Johnson, A.C., Knights, B., Milner, N.J., Ormerod, S.J., Solomon, D., Timlett, R., Whitehead, P.J., Wood, P.J., 2010. Evidence needed to manage freshwater ecosystems in a changing climate: Turning adaptation principles into practice. *Science of the Total Environment* 408, 4150-4164. <https://doi.org/10.1016/j.scitotenv.2010.05.014>
- Wilding, N.A., White, J.C., Chadd, R.P., House, A., Wood, P.J., 2018. The influence of flow permanence and drying pattern on macroinvertebrate biomonitoring tools used in the assessment of riverine ecosystems. *Ecological Indicators* 85, 548-555. <https://doi.org/10.1016/j.ecolind.2017.10.059>
- Zahrádková, S., Hájek, O., Tremel, P., Pařil, P., Straka, M., Němejcová, D., Polášek, M., Ondráček P., 2015. Hodnocení rizika vysychání drobných vodních toků v České republice. *VTEI* 6, 4-16.

Appendix A

Table A.1: List of flow intermittence tolerant taxa and the reasons for their inclusion in the list. The reliability of indicators is based on stronger (value 2) or weaker (value 1) evidence. Indicator values (IV) and their significance are shown separately for spring and autumn seasons for taxa that were associated with intermittent and near-perennial samples in Indicator Species Analysis. The frequency of taxa at intermittent and near-perennial samples is shown. Published sources in which information was found is shown. * - $p < 0.05$, ** - $p < 0.01$, *** - $p < 0.001$, n.s. – not significant

Taxon	Reliability	IV Spring/Autumn	Significance of IV Spring/Autumn	Frequency Spring/Autumn	References
Platyhelminthes					
<i>Mesostoma lingua</i>	1				Young, 2001
Nematomorpha					
Gordiidae Gen. sp.	1	-/0.06	-/n.s.	0/0.06	Clifford, 1966
<i>Gordius</i> sp.	1				Clifford, 1966
Mollusca					
<i>Anisus leucostoma</i>	2				Piechocki, 1979; Jurkiewicz-Karnkowska, 2011; Costil et al., 2001
<i>Anisus septemgyratus</i>	2				Jurkiewicz-Karnkowska, 2008; Jurkiewicz-Karnkowska, 2011
<i>Anisus</i> sp.	1				Costil et al., 2001
<i>Anisus spirorbis</i>	1				Jurkiewicz-Karnkowska, 2008
<i>Aplexa hypnorum</i>	2				Jurkiewicz-Karnkowska, 2008; Costil et al., 2001
<i>Bathyomphalus contortus</i>	1				Jurkiewicz-Karnkowska, 2011
<i>Bithynia leachii</i>	2				Jurkiewicz-Karnkowska, 2008; Piechocki, 1979
<i>Bythinella austriaca</i> agg.	2	0.44/0.46	*/*	0.45/0.56	
<i>Galba truncatula</i>	2	0.24/0.18	n.s./n.s.	0.27/0.19	Meyer and Meyer, 2000; Acuña et al., 2005; Stubbington et al., 2009; Costil et al., 2001; Falkner et al., 2001
<i>Gyraulus riparius</i>	1				Jurkiewicz-Karnkowska, 2008
<i>Musculium lacustre</i>	1				Piechocki, 1993
<i>Pisidium casertanum</i>	1				Piechocki, 1993
<i>Pisidium obtusale</i>	1				Piechocki, 1993
<i>Pisidium personatum</i>	1				Piechocki, 1993
<i>Planorbis corneus</i>	1				Jurkiewicz-Karnkowska, 2011
<i>Planorbis planorbis</i>	2				Jurkiewicz-Karnkowska, 2011; Falkner et al., 2001
<i>Potamopyrgus antipodarum</i>	1	0.09/-	n.s./-	0.09/0.06	Costil et al., 2001
<i>Radix peregra</i>	2	-/0.06	-/n.s.	0/0.06	Costil et al., 2001; Extence, 1981
<i>Segmentina nitida</i>	2				Jurkiewicz-Karnkowska, 2011; Falkner et al., 2001
<i>Stagnicola fuscus</i>	1				Falkner et al., 2001
<i>Stagnicola occultus</i>	1				Falkner et al., 2001
<i>Stagnicola palustris</i>	1				Jurkiewicz-Karnkowska, 2011
<i>Valvata cristata</i>	2				Jurkiewicz-Karnkowska, 2008; Jurkiewicz-Karnkowska, 2011; Falkner et al., 2001.
<i>Valvata macrostoma</i>	2				Jurkiewicz-Karnkowska, 2008; Jurkiewicz-Karnkowska, 2011; Falkner et al., 2001
Oligochaeta					

<i>Eiseniella tetraedra</i>	2	0.54/0.64	n.s./*	0.82/0.75	Fenoglio et al., 2007; Hynes, 1958; Dumnicka and Kozsalka, 2005
<i>Enchytraeus</i> sp.	2	0.24/0.25	n.s./*	0.27/0.25	
<i>Epirodillus pygmaeus</i>	1	0.18/0.2	n.s./n.s.	0.18/0.25	
<i>Haplotaxis gordioides</i>	2	0.29/0.32	n.s./*	0.36/0.38	
<i>Marionina</i> sp.	2	0.61/0.12	*/n.s.	0.73/0.13	
<i>Nais elinguis</i>	2	0.18/0.3	n.s./**	0.18/0.31	
<i>Stylodrilus heringianus</i>	2	0.56/0.52	n.s./n.s.	0.64/0.63	Dumnicka and Koszalka, 2005; Nijboer et al., 2004
Hirudinea					
<i>Dina lineata</i>	1				Elliott and Mann, 1979
<i>Haemopsis sanguisuga</i>	2	0.18/0.19	n.s./*	0.18/0.19	Elliott and Mann, 1979
Crustacea					
<i>Asellus aquaticus</i>	2	0.07/0.04	n.s./n.s.	0.09/0.06	Iversen et al., 1978; Extence, 1981
Ephemeroptera					
<i>Cloeon dipterum</i> s. lat.	1	0.09/0.13	n.s./n.s.	0.09/0.13	
<i>Ecdyonurus picteti</i>	1	0.18/-	n.s./-	0.18/0	
<i>Habrophlebia fusca</i>	2	-/-	-/-	0.09/0.06	Bohle, 2000; Řezníčková et al., 2010
<i>Metreletus balcanicus</i>	2	0.09/-	n.s./-	0.09/0	Bauernfeind and Soldán, 2012; Soldán and Zahrádková, 2000; Bohle, 2000
<i>Paraleptophlebia werneri</i>	1				Soldán and Zahrádková, 2000
<i>Siphonurus aestivalis</i>	2	0.25/-	n.s./-	0.27/0	Bauernfeind and Soldán, 2012; Řezníčková et al., 2010
<i>Siphonurus armatus</i>	2	0.09/-	n.s./-	0.09/0	Bauernfeind and Soldán, 2012; Soldán and Zahrádková, 2000; Bohle, 2000
<i>Siphonurus lacustris</i>	2				Fenoglio et al., 2007; Hynes, 1958
Odonata					
<i>Pyrrhosoma nymphula</i>	1				Opatrilova et al., 2005
Plecoptera					
<i>Brachyptera risi</i>	2	0.76/0.06	*/n.s.	0.82/0.06	Bohle, 2000; Hynes, 1941; Wagner et al., 2011; Mackereth, 1957
<i>Capnia bifrons</i>	2	0.45/0.5	*/n.s.	0.45/0.56	Bohle, 2000
<i>Isoperla tripartita</i>	2	0.55/0.19	**/n.s.	0.55/0.25	
<i>Nemoura cinerea</i>	2				Bohle, 2000; Malicky, 1989; Meyer and Meyer, 2000; Iversen et al., 1978; Hynes, 1958
Coleoptera					
<i>Agabus guttatus</i>	2	0.36/0.54	*/**	0.36/0.56	Bohle, 2000; Hynes, 1958
<i>Agabus</i> sp.	2				Bohle, 2000; Hynes, 1958; Meyer and Meyer, 2000
<i>Anacaena globulus</i>	2	-/0.25	-/*	0/0.25	Bohle, 2000; Hynes, 1958
<i>Cyphon</i> sp.	1	-/0.13	-/n.s.	0/0.13	
<i>Esolus angustatus</i>	2	0.15/0.25	n.s./*	0.27/0.25	
<i>Helophorus aquaticus</i>	1				Bohle, 2000
<i>Helophorus brevipalpis</i>	2				Bohle, 2000; Smith and Wood, 2002; Stubbington et al., 2009
<i>Helophorus flavipes</i>	1				Hynes, 1958
<i>Helophorus grandis</i>	1				Bohle, 2000
<i>Helophorus</i> sp.	2				Wood et al., 2005
<i>Hydraena melas</i>	1	-/-	-/-	0/0.06	Fenoglio et al., 2007
<i>Hydraena nigrita</i>	1	0.3/0.19	n.s./n.s.	0.55/0.38	Bohle, 2000
<i>Hydroporus planus</i>	1	-/0.06	-/n.s.	0/0.06	Opatrilova et al., 2005
<i>Hydroporus pubescens</i>	1				Opatrilova et al., 2005
<i>Limnebius truncatellus</i>	2	-/-	-/-	0/0.06	Bohle, 2000; Jackson, 1973

<i>Platambus maculatus</i>	2	0.18/0.53	n.s./*	0.18/0.63	Bohle, 2000; Hynes, 1958; Meyer and Meyer, 2000
Trichoptera					
<i>Agrypnia varia</i>	1				Moretti et al., 1976
<i>Beraea pullata</i>	1				Bohle, 2000
<i>Beraeidae</i> Gen. sp.	1				Fenoglio et al., 2007
<i>Beraeodes minutus</i>	2	0.09/0.19	n.s./*	0.09/0.19	Bohle, 2000
<i>Glyphotaelius pellucidus</i>	2	0.25/0.19	n.s./n.s.	0.27/0.19	Bohle, 2000; Acuña et al., 2005; Svensson, 1974
<i>Grammotaulius nigropunctatus</i>	2				Novák and Sehnal, 1963; Hiley, 1978
<i>Chaetopteryx villosa</i>	1	0.18/0.15	n.s./n.s.	0.18/0.19	
<i>Ironoquia dubia</i>	2	0.09/-	n.s./-	0.09/0	Bohle, 2000; Sommerhäuser et al., 1995
<i>Leptocerus tineiformis</i>	1				Moretti et al., 1976
<i>Limnephilus affinis</i>	1				Hiley, 1978
<i>Limnephilus auricula</i>	2				Novák and Sehnal, 1963; Hiley, 1978; Bohle, 2000; Smith et al., 2003; Pastuchová, 2008; Sommerhäuser et al., 1995; Svensson, 1974
<i>Limnephilus bipunctatus</i>	2				Novák and Sehnal, 1963; Moretti et al., 1976; Bohle, 2000; Meyer and Meyer, 2000
<i>Limnephilus centralis</i>	2				Hiley, 1978; Bohle, 2000; Smith et al., 2003; Wood et al., 2005; Svensson, 1974
<i>Limnephilus coenosus</i>	1				Hiley, 1978
<i>Limnephilus flavicornis</i>	1				Novák and Sehnal, 1963
<i>Limnephilus griseus</i>	2				Novák and Sehnal, 1963; Hiley, 1978; Svensson, 1974; Gislason, 1993
<i>Limnephilus incisus</i>	1				Hiley, 1978
<i>Limnephilus lunatus</i>	2				Bohle, 2000; Meyer and Meyer, 2000; Smith et al., 2003
<i>Limnephilus sparsus</i>	2				Novák and Sehnal, 1963; Hiley, 1978; Moretti et al., 1976; Bohle, 2000; Svensson, 1974
<i>Limnephilus stigma</i>	2				Novák and Sehnal, 1963; Hiley, 1978; Svensson, 1974
<i>Limnephilus vittatus</i>	2				Novák and Sehnal, 1963; Hiley, 1978; Moretti et al., 1976; Svensson, 1974
<i>Micropterna lateralis</i>	2				Bohle, 2000; Smith et al., 2003; Wood et al., 2005; Pastuchová, 2008; Sommerhäuser et al., 1995
<i>Micropterna nycterobia</i>	2	0.49/-	*/-	0.55/0	Pastuchová, 2008; Meyer and Meyer, 2000; Moretti et al., 1976
<i>Micropterna sequax</i>	2				Bohle, 2000; Meyer and Meyer, 2000; Smith et al., 2003; Wood et al., 2005
<i>Micropterna testacea</i>	1				Meyer and Meyer, 2000
<i>Oligostomis reticulata</i>	1				Sommerhäuser et al., 1995
<i>Rhadicoleptus alpestris</i>	1				Hiley, 1978
<i>Stenophylax permistus</i>	2				Bohle, 2000; Meyer and Meyer, 2000; Smith et al., 2003; Wood et al., 2005
<i>Trichostegia minor</i>	2				Novák and Sehnal, 1963; Svensson, 1974
Diptera					
<i>Diplocladius cultriger</i>	2	0.34/0.55	n.s./**	0.36/0.56	
<i>Eukiefferiella claripennis</i>	1	-/0.13	-/n.s.	0/0.13	
<i>Hydrobaenus</i> sp.	1	0.27/-	n.s./-	0.27/0	
<i>Chironomus</i> sp.	2	-/0.31	-/**	0/0.31	
<i>Krenosmittia camptophleps</i>	1	0.27/-	n.s./-	0.27/0	
<i>Macropelopia</i> sp.	2	0.45/0.08	*/n.s.	0.45/0.13	
<i>Micropsectra atrofasciata</i> -Gr.	2	-/0.89	-/*	1/0.94	
<i>Natarsia</i> sp.	2	0.58/0.42	*/*	0.64/0.5	
<i>Orthocladus rivicola</i> -Gr.	2	0.28/0.19	n.s./*	0.36/0.19	
<i>Parametriocnemus stylatus</i>	1	0.71/0.35	n.s./n.s.	0.91/0.63	Rüegg and Robinson, 2004
<i>Paraphaenocladus</i> sp.	2	0.64/0.25	*/*	0.82/0.25	

<i>Paratrichocladius rufiventris</i>	1	-/-	-/-	0.18/0	Moller Pillot, 2013
<i>Procladius</i> sp.	1	0.18/0.24	n.s./n.s.	0.18/0.25	
<i>Rheocricotopus atripes</i>	1	0.27/0.13	n.s./n.s.	0.27/0.13	
<i>Rheocricotopus effusus</i>	2	0.44/0.3	*/*	0.45/0.31	
<i>Rheocricotopus effusus</i>	2	0.44/0.3	*/*	0.45/0.31	
<i>Rhypholophus haemorrhoidalis</i>	1	0.41/0.13	n.s./n.s.	0.45/0.13	
<i>Simulium noelleri</i>	1	0.09/0.19	n.s./n.s.	0.09/0.19	
<i>Zavrelimyia</i> sp.	2	0.7/-	**/-	0.82/0	

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777 Table A.2: List of perennial flow indicators and the reasons for their inclusion in the list. The reliability
 778 of indicators is based on stronger (value 2) or weaker (value 1) evidence. Indicator values (IV) and
 779 their significance are shown separately for spring and autumn seasons for taxa that were associated
 780 with perennial samples in Indicator Species Analysis. The frequency of taxa at perennial samples is
 781 shown. Published sources in which information was found are shown. * - $p < 0.05$, ** - $p < 0.01$, *** -
 782 $p < 0.001$, n.s. – not significant

Taxon	Reliability	IV Spring/Autumn	Significance of IV spring/autumn	Frequency spring/autumn	References
Platyhelminthes					
<i>Crenobia alpina</i>	2				Fenoglio et al., 2007; Rügge and Robinson, 2004; Hynes, 1958
<i>Dugesia gonocephala</i>	2	0.82/0.73	**/**	0.83/0.74	Meyer and Meyer, 2000
<i>Dugesia</i> sp.	1				Fenoglio et al., 2007
<i>Phagocata vitta</i>	1				Hynes, 1958
<i>Polycelis nigra</i>	1				Sarriquet et al., 2007
<i>Polycelis</i> sp.	1				Fenoglio et al., 2007
<i>Polycelis tenuis</i>	1	-/-	-/-	0/0.04	Sarriquet et al., 2007
Mollusca					
<i>Ancylus fluviatilis</i>	2	0.46/0.25	*/n.s.	0.5/0.26	Meyer and Meyer, 2000; Wood et al., 2005; Stubbington et al., 2009; Fenoglio et al., 2007
<i>Lithoglyphus naticoides</i>	2				Jurkiewicz-Karnkowska, 2008; Falkner et al., 2001
<i>Lymnaea stagnalis</i>	1				Falkner et al., 2001
<i>Myxas glutinosa</i>	1				Falkner et al., 2001
Oligochaeta					
<i>Aulodrilus japonicus</i>	1	0.17/0.12	n.s./n.s.	0.17/0.15	
<i>Nais alpina</i>	2	0.33/0.47	n.s./*	0.33/0.48	
<i>Ophidonais serpentina</i>	1	0.25/0.11	n.s./n.s.	0.25/0.11	
<i>Pristinella rosea</i> Gr.	1	0.17/0.35	n.s./n.s.	0.17/0.37	
<i>Propappus volki</i>	2	0.25/0.3	n.s./*	0.25/0.3	
<i>Rhyacodrilus coccineus</i>	1	0.25/0.05	n.s./n.s.	0.25/0.15	
Hirudinea					
<i>Erpobdella octoculata</i>	2	0.17/0.14	n.s./n.s.	0.17/0.15	Meyer a Meyer, 2001; Sarriquet et al., 2007
<i>Glossiphonia complanata</i>	1				Sarriquet et al., 2007
<i>Piscicola geometra</i>	2				Wood et al., 2005; Elliott and Mann, 1979
Acari					
Hydracarina	2	0.56/0.53	n.s./*	0.58/0.59	
Crustacea					
<i>Gammarus fossarum</i>	1	-/0.46	-/n.s.	0.75/0.78	Meyer and Meyer, 2000
Ephemeroptera					
<i>Baetis alpinus</i>	1				Rügge and Robinson, 2004
<i>Baetis muticus</i>	2	0.87/0.8	**/**	0.92/0.81	Řezníčková et al., 2010
<i>Baetis niger</i>	1	0.17/0.17	n.s./n.s.	0.17/0.19	
<i>Baetis rhodani</i> s.l.	2	0.96/0.87	**/**	1/0.93	Iversen et al., 1978; Pastuchová, 2008; Morrison, 1990
<i>Baetis</i> sp.	2				Bohle, 2000; Wood et al., 2005; Stubbington et al., 2009; Pastuchová, 2008
<i>Baetis vernus</i>	1	-/0.11	-/n.s.	0/0.22	Pastuchová, 2008
<i>Centroptilum luteolum</i>	1	0.5/-	n.s./-	0.75/0.67	Fenoglio et al., 2007

<i>Ecdyonurus austriacus</i>	1	-/0.11	-/n.s.	0.08/0.11	
<i>Ecdyonurus starmachi</i>	1				Řezníčková et al., 2010
<i>Ecdyonurus subalpinus</i>	1	0.16/0.26	n.s./n.s.	0.17/0.3	Pastuchová, 2008
<i>Ecdyonurus torrentis</i>	2	0.33/0.48	n.s./*	0.33/0.48	
<i>Ecdyonurus venosus</i>	1	-/0.04	-/n.s.	0/0.04	Hynes, 1958
<i>Epeorus assimilis</i>	1	0.33/0.22	n.s./n.s.	0.33/0.22	
<i>Ephemera danica</i>	2	0.65/0.63	*/**	0.75/0.78	Meyer and Meyer, 2000; Pastuchová, 2008; Řezníčková et al., 2010
<i>Ephemerella mucronata</i>	1	0.17/0.07	n.s./n.s.	0.17/0.07	
<i>Habroleptoides confusa</i>	2	0.55/0.7	n.s./*	0.67/0.81	Řezníčková et al., 2010; Pastuchová, 2008
<i>Habrophlebia lauta</i>	2	0.69/0.63	*/*	0.92/0.81	Řezníčková et al., 2010; Meyer and Meyer, 2000
<i>Heptagenia sulphurea</i>	1				Wood et al., 2005
<i>Leptophlebia marginata</i>	1				Opatrilova et al., 2005
<i>Paraleptophlebia submarginata</i>	2	0.48/0.59	*/n.s.	0.5/0.67	
<i>Rhithrogena carpatoalpina</i>	2	0.21/0.19	n.s./n.s.	0.25/0.19	Pastuchová, 2008; Řezníčková et al., 2010
<i>Rhithrogena iridina</i>	2				
<i>Rhithrogena iridina/picteti</i>	2	0.76/0.63	*/**	0.83/0.63	
<i>Rhithrogena picteti</i>	2				Meyer and Meyer, 2000; Pastuchová, 2008
<i>Rhithrogena semicolorata</i>	2	0.17/0.07	n.s./n.s.	0.17/0.07	Pastuchová, 2008; Řezníčková et al., 2010; Hynes, 1958
<i>Serratella ignita</i>	2				Meyer and Meyer, 2000; Stubbington et al., 2009; Pastuchová, 2008; Hynes, 1958
<i>Torleya major</i>	2	0.25/0.33	n.s./*	0.25/0.33	Meyer and Meyer, 2000
Plecoptera					
<i>Amphinemura</i> sp.	1	0.41/0.22	n.s./n.s.	0.42/0.22	
<i>Dinocras cephalotes</i>	2	0.08/0.07	n.s./n.s.	0.08/0.07	Fenoglio et al., 2007; Hynes, 1941
<i>Dinocras</i> sp.	2				Fenoglio et al., 2007; Hynes, 1941
<i>Diura bicaudata</i>	2	-/0.04	-/n.s.	0/0.04	Stubbington et al., 2009; Hynes, 1958
<i>Chloroperla</i> sp.	2				Fenoglio et al., 2007; Hynes, 1941
<i>Chloroperla tripunctata</i>	2				Fenoglio et al., 2007; Hynes, 1941
<i>Isoperla oxylepis</i>	1	0.25/0.17	n.s./n.s.	0.25/0.22	
<i>Isoperla rivulorum</i>	1	0.06/0.11	n.s./n.s.	0.08/0.11	
<i>Leuctra digitata</i>	1				Pastuchová, 2008
<i>Leuctra fusca</i>	1				Wood et al., 2005; Hynes, 1941
<i>Leuctra handlirschi</i>	1				Hynes, 1941
<i>Leuctra inermis</i>	2				Hynes, 1958
<i>Leuctra pseudosignifera</i>	1	0.08/0.11	n.s./n.s.	0.08/0.11	
<i>Leuctra</i> sp.	2	0.5/0.87	n.s./**	0.5/0.89	Stubbington et al., 2009
<i>Perla</i> sp.	2				Fenoglio et al., 2007; Hynes, 1941; Mackereth, 1957
<i>Perlodes microcephalus</i>	1	0.17/0.11	n.s./n.s.	0.17/0.11	
<i>Protonemura</i> sp.	2	0.38/0.44	n.s./**	0.42/0.44	Pastuchová, 2008
<i>Siphonoperla</i> sp.	1	0.17/0.25	n.s./n.s.	0.17/0.26	
Heteroptera					
<i>Velia saulii</i>	1	-/0.11	-/n.s.	0/0.11	
Coleoptera					
<i>Elmis aenea</i>	2	0.49/0.44	*/*	0.5/0.44	Wood et al., 2005; Stubbington et al., 2009
<i>Elodes marginata</i>	2	0.33/0.3	n.s./*	0.33/0.3	
<i>Esolus parallelepipedus</i>	1	0.08/0.15	n.s./n.s.	0.08/0.15	
<i>Hydraena saga</i>	1	-/0.11	-/n.s.	0/0.11	
<i>Hydraena schuleri</i>	1	0.17/0.09	n.s./n.s.	0.17/0.11	
<i>Hydroporus nigrita</i>	1				Opatrilova et al., 2005
<i>Limnius perrisi</i>	2	0.5/0.33	*/*	0.5/0.33	
<i>Limnius volckmari</i>	1	0.27/0.46	n.s./n.s.	0.33/0.74	Wood et al., 2005
<i>Orectochilus villosus</i>	2	0.17/0.11	n.s./n.s.	0.17/0.11	Meyer and Meyer, 2000
<i>Oreodytes sanmarkii</i>	1	-/0.07	-/n.s.	0/0.07	Fenoglio et al., 2007
<i>Riolus subviolaceus</i>	1	0.07/0.09	n.s./n.s.	0.08/0.11	Wood et al., 2005

Trichoptera

<i>Agapetus fuscipes</i>	2	0.25/-	n.s./-	0.25/0	Sommerhäuser et al., 1995; Wood et al., 2005
<i>Anabolia nervosa</i>	1				Sommerhäuser et al., 1995
<i>Athripsodes aterrimus</i>	2				Hiley, 1978; Sommerhäuser et al., 1995
<i>Athripsodes bilineatus</i>	1	0.18/0.21	n.s./n.s.	0.25/0.3	Sommerhäuser et al., 1995
<i>Athripsodes cinereus</i>	1				Sommerhäuser et al., 1995
<i>Cyrnus flavidus</i>	1				Bohle, 2000
<i>Drusus annulatus</i>	1	0.16/-	n.s./-	0.25/0.04	Stubbington et al., 2009
<i>Ecclisopteryx madida</i>	1	0.17/-	n.s./-	0.17/0	
<i>Goera pilosa</i>	2				Iversen et al., 1978; Sommerhäuser et al., 1995
<i>Hagenella clathrata</i>	1				Opatrilova et al., 2005
<i>Halesus digitatus</i>	2				Pastuchová, 2008; Sommerhäuser et al., 1995
<i>Halesus radiatus</i>	2				Hiley, 1978; Pastuchová, 2008; Sommerhäuser et al., 1995
<i>Hydropsyche angustipennis</i>	2				Iversen et al., 1978; Sommerhäuser et al., 1995
<i>Hydropsyche bulbifera</i>	1				Pastuchová, 2008
<i>Hydropsyche contubernalis</i>	1				Sommerhäuser et al., 1995
<i>Hydropsyche fulvipes</i>	1				Pastuchová, 2008
<i>Hydropsyche instabilis</i>	1				Pastuchová, 2008
<i>Hydropsyche pellucidula</i>	2				Meyer and Meyer, 2000; Sommerhäuser et al., 1995; Pastuchová, 2008
<i>Hydropsyche saxonica</i>	2				Sommerhäuser et al., 1995; Pastuchová, 2008
<i>Hydropsyche siltalai</i>	1				Sommerhäuser et al., 1995
<i>Hydropsyche</i> sp.	2	0.87/0.89	**/**	0.92/0.93	
Hydropsychidae Gen. sp.	1				Bohle, 2000
Hydroptilidae Gen. sp.	2				Fenoglio et al., 2007; Wallace et al., 2003
<i>Chaetopteryx fusca/villosa</i>	1				Pastuchová, 2008
<i>Lasiocephala basalis</i>	2	0.08/0.11	n.s./n.s.	0.08/0.11	Sommerhäuser et al., 1995
<i>Lepidostoma hirtum</i>	1	0.17/0.07	n.s./n.s.	0.17/0.07	
<i>Lype phaeopa</i>	1				Sommerhäuser et al., 1995
<i>Lype reducta</i>	1				Sommerhäuser et al., 1995
<i>Lype</i> sp.	1	0.46/0.31	n.s./n.s.	0.5/0.41	Sommerhäuser et al., 1995
<i>Micrasema longulum</i>	1	-/0.11	-/n.s.	0/0.11	
<i>Molanna angustata</i>	1				Sommerhäuser et al., 1995
<i>Mystacides azurea</i>	2	0.08/0.09	n.s./n.s.	0.08/0.11	Sommerhäuser et al., 1995
<i>Neureclipsis bimaculata</i>	1				Sommerhäuser et al., 1995
<i>Notidobia ciliaris</i>	1				Sommerhäuser et al., 1995
<i>Odontocerum albicorne</i>	2	0.55/0.64	*/**	0.67/0.67	Pastuchová, 2008; Meyer and Meyer, 2000; Fenoglio et al., 2007
<i>Oecismus monedula</i>	1	0.21/0.19	n.s./n.s.	0.25/0.19	
Philopotamidae Gen. sp.	1				Bohle, 2000
<i>Philopotamus montanus</i>	2	0.33/0.22	n.s./n.s.	0.33/0.22	Pastuchová, 2008
<i>Polycentropus flavomaculatus</i>	2	0.57/0.5	*/*	0.58/0.56	Meyer and Meyer, 2000; Wood et al., 2005
<i>Polycentropus irroratus</i>	2	0.17/0.11	n.s./n.s.	0.17/0.11	Sommerhäuser et al., 1995
<i>Potamophylax cingulatus</i>	2				Hiley, 1978; Sommerhäuser et al., 1995
<i>Potamophylax luctuosus</i>	1				Sommerhäuser et al., 1995
<i>Potamophylax nigricornis</i>	2				Pastuchová, 2008; Meyer and Meyer, 2000
<i>Potamophylax rotundipennis</i>	2				Sommerhäuser et al., 1995; Pastuchová, 2008
<i>Potamophylax</i> sp.	2	0.44/0.4	n.s./*	0.75/0.41	
<i>Psychomyia pusilla</i>	1				Meyer and Meyer, 2000
Psychomyiidae Gen. sp.	1				Fenoglio et al., 2007
<i>Rhyacophila dorsalis</i>	2				Wood et al., 2005; Hynes, 1958
<i>Rhyacophila fasciata</i>	2				Pastuchová, 2008; Sommerhäuser et al., 1995
<i>Rhyacophila nubila</i>	2	0.25/-	n.s./-	0.25/0	Meyer and Meyer, 2000
<i>Rhyacophila oblitterata</i>	1				Hynes, 1958

<i>Rhyacophila polonica</i>	1	0.14/-	n.s./-	0.17/0	Pastuchová, 2008
<i>Rhyacophila</i> sp.	1				Bohle, 2000
<i>Rhyacophila tristis</i>	1	0.25/-	n.s./-	0.25/0	
<i>Sericostoma personatum</i>	2				Pastuchová, 2008; Sommerhäuser et al., 1995
<i>Sericostoma</i> sp.	2	0.52/0.64	n.s./*	0.83/0.81	Meyer and Meyer, 2000
<i>Silo pallipes</i>	1	0.44/-	n.s./-	0.58/0	Pastuchová, 2008
<i>Silo piceus</i>	1				Pastuchová, 2008
<i>Synagapetus iridipennis</i>	1	-/-	-/-	0.08/0	Pastuchová, 2008
<i>Tinodes assimilis</i>	1				Sommerhäuser et al., 1995
<i>Tinodes pallidulus</i>	1				Sommerhäuser et al., 1995
<i>Tinodes rostocki</i>	1				Pastuchová, 2008
<i>Tinodes waeneri</i>	1				Sommerhäuser et al., 1995
<i>Wormaldia occipitalis</i>	1				Pastuchová, 2008

Diptera

<i>Ablabesmyia</i> sp.	1	0.17/-	n.s./-	0.17/0.04	
<i>Atherix ibis</i>	2	-/0.11	-/n.s.	0/0.11	Meyer and Meyer, 2000
<i>Berdeniella</i> sp.	2	0.42/0.15	*/n.s.	0.42/0.15	
<i>Brillia longifurca</i>	1	0.22/0.15	n.s./n.s.	0.33/0.15	
<i>Ceratopogonidae</i> Gen. sp.	1	0.33/-	n.s./-	0.33/11	
<i>Dicranota</i> sp.	2	0.82/0.47	**/n.s.	0.92/0.67	Meyer and Meyer, 2000
<i>Eloeophila</i> sp.	1	-/0.41	-/n.s.	0.67/0.7	Meyer and Meyer, 2000
<i>Eukiefferiella devonica/ilkeyensis</i>	1	0.17/0.04	n.s./n.s.	0.17/0.04	
<i>Hemerodromia</i> sp.	2	0.5/0.24	**/n.s.	0.5/0.26	Opatrilova et al., 2005
<i>Ibisia marginata</i>	2	0.57/0.44	**/*	0.58/0.44	
<i>Microtendipes rydalensis-Gr.</i>	1	0.25/0.19	n.s./n.s.	0.25/0.19	
<i>Paratrissocladius excerptus</i>	1	0.27/0.19	n.s./n.s.	0.42/0.19	
<i>Peripsychoda</i> sp.	1	0.17/0.04	n.s./n.s.	0.17/0.04	
<i>Pilaria</i> sp.	1	-/0.15	-/n.s.	0/0.15	
<i>Polypedilum albicorne</i>	1	0.13/0.11	n.s./n.s.	0.25/0.11	
<i>Polypedilum convictum-Gr.</i>	2	-/0.49	-/*	0.25/0.56	
<i>Polypedilum pedestre-Gr.</i>	1	-/0.15	-/n.s.	0/0.15	
<i>Potthastia longimana</i>	2	0.25/0.37	n.s./*	0.25/0.37	
<i>Prosimulium hirtipes</i>	1	0.33/-	n.s./-	0.33/0	
<i>Rheotanytarsus</i> sp.	2	0.17/0.22	n.s./n.s.	0.17/0.22	Meyer and Meyer, 2000
<i>Scleroprocta</i> sp.	1	0.17/0.1	n.s./n.s.	0.17/0.11	
<i>Simulium angustitarse</i>	1	0.17/-	n.s./-	0.17/0.07	
<i>Simulium variegatum</i>	1	0.25/0.11	n.s./n.s.	0.25/0.11	
<i>Stempellinella brevis</i>	1	0.3/0.15	n.s./n.s.	0.67/0.15	
<i>Stratiomyidae</i> Gen. sp.	1				Fenoglio et al., 2007
<i>Thienemanniella</i> sp.	2	0.58/0.24	*/n.s.	0.58/0.3	
<i>Thienemannimyia</i> sp.	2	0.42/0.12	*/n.s.	0.42/0.15	
<i>Tvetenia bavarica/calvescens</i>	2	0.44/0.41	n.s./*	0.5/0.41	Moller Pillot, 2013

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References for Appendix A

- Acuña, V., Muñoz, I., Giorgi, A., Omella, M., Sabater, F., Sabater, S. 2005. Drought and postdrought recovery cycles in an intermittent Mediterranean stream: structural and functional aspects. *J. N. Am. Benthol. Soc.* 54, 919-933.
- Bauernfeind, E., Soldán, T., 2012. *The Mayflies of Europe (Ephemeroptera)*, Apollo Books, Ollerup, Denmark, 781 pp.
- Bohle, H.W., 2000. Anpassungsstrategien ausgewählter Organismen an temporäre Wasserführung - Insekten periodischer Fließgewässer Mitteleuropas. *Gewässer ohne Wasser? Ökologie, Management und Schutz temporärer Gewässer*, NUA Seminarbericht Band 5, 53-71.
- Clifford, H.F., 1966. The Ecology of Invertebrates in an Intermittent Stream. *Invest. Indiana lakes and streams*, vol. VII, No. 2.
- Costil, K., Dussart, G.B.J., Daguzan, J., 2001. Biodiversity of aquatic gastropods in the Mont St-Michel basin (France) in relation to salinity and drying of habitats. *Biodiversity and Conservation* 10, 1–18.
- Dumnicka, E., Kozsalka, J., 2005. The effect of drought on Oligochaeta communities in small woodland streams. *Biologia, Bratislava* 60, 143-150.
- Elliott, J.M., Mann, K.H., 1979. A key to the British freshwater leeches : with notes on their life cycles and ecology. *Freshwater Biological Association, Cumbria UK*, 72 p.
- Extence, C.A., 1981. The effect of drought on benthic invertebrate communities in a lowland river. *Hydrobiologia* 83, 217-224.
- Falkner, G., Obrdlík, P., Castella, E., Speight, M.C.D., 2001. *Shelled Gastropoda of Western Europe*, Verlag der Friedrich-Held-Gesellschaft, München. pp 267.
- Fenoglio, S., Bo, T., Malacarne, G., 2007. Response of benthic invertebrate assemblages to varying drought conditions in the Po river (NW Italy). *Italian Journal of Zoology* 74, 191-201.
- Hiley, P.D., 1978. Some aspects of the life histories of Limnephilidae (Trichoptera) related to the distribution of their larvae. *Proc. of the 2nd Int. Symp. on Trichoptera, 1977*, Junk, Hague.
- Hynes, H.B.N., 1941. The taxonomy and ecology of the nymphs of British Plecoptera with notes on the adults and eggs. *Trans. R. ent. Soc. Lond.* 91, 459-557.
- Hynes, H.B.N., 1958. The effect of drought on the fauna of a small mountain stream in Wales. *Verh. internat. Ver. Limnol.* 13: 826-833.
- Iversen, T.M., Wiberg-Larsen, P., Hansen, S.B., Hansen, F.S., 1978. The effect of partial and total drought on the macroinvertebrate communities of three small Danish streams. *Hydrobiologia* 60: 235-242.
- Jackson, D.J., 1973. The influence of flight capacity on the distribution of aquatic Coleoptera in Fife and Kinross-shire. *Entomologist's Gazette* 24, 247-293.
- Jurkiewicz-Karnkowska, E., 2008. Aquatic mollusc communities in riparian sites of different size, hydrological connectivity and succession stage. *Polish Journal of Ecology*, 56, 99-118.
- Jurkiewicz-Karnkowska, E., 2011. Diversity of aquatic malacofauna of temporary water bodies within the lower Bug river floodplain. *Folia malacologica*, 19, 9-18.
- Mackereth, J.C., 1957. Notes on the Plecoptera from stony stream. *J. Anim. Ecol.* 26: 341-350.
- Malicky, H., 1989. Ein ungewöhnlicher Biotop von *Nemoura cinerea* Retz. (Plecoptera). *Z. Arbgem. öst. Entomol.* 40:119.

- 831 Meyer, A., Meyer, E.I., 2000. Discharge regime and the effect of drying on macroinvertebrate
832 communities in a temporary karst stream in East Westphalia (Germany). *Aquatic Sciences*
833 62: 216-231.
- 834 Moller Pillot, H.K.M., 2013. Chironomidae larvae of the Netherlands and adjacent lowlands.
835 Biology and ecology of the aquatic Orthoclaadiinae. Zeist. The Netherlands. KNNV
836 Publishing., 314 pp.
- 837 Moretti, G.P., Cianficconi, F., Pirisinu, Q., 1976. The Trichoptera population of a temporary
838 ecosystem of the Umbrian Apennines (Perugia, Italy): Proc. Of the First Int. Symp. On
839 Trichoptera, 1947, Junk, The Hague.
- 840 Morrison, B.R. S., 1990. Recolonisation of four small streams in central Scotland following
841 drought conditions in 1984. *Hydrobiologia* 208: 261-267.
- 842 Nijboer, R.C., Wetzel, M.J., Verdonschot P.F.M., 2004. Diversity and distribution of
843 Tubificidae, Naididae, and Lumbriculidae (Annelida: Oligochaeta) in the Netherlands: an
844 evaluation of twenty years of monitoring data. *Hydrobiologia* 520: 127–141.
- 845 Novák, K., Sehnal, F., 1963. The development cycle of some species of the genus *Limnephilus*
846 (Trichoptera). *Časopis Československé společnosti entomologické* 60, 68-80.
- 847 Opatrilova, L., Verdonschot, P., Ofenböck, T., Murphy, J., Erba, S., Sandin, L., Lorenz, A.,
848 Postolache, C., Gevrey, M., 2005. Report – review of existing information on key taxa and
849 functional groups relevant to the eight study catchments, Deliverable No. 13, Euro-
850 limpacs project. 205 pp.
- 851 Pastuchová, Z., 2008. Ephemeroptera, Plecoptera and trichoptera communities of streams in
852 Cerová vrchovina highland, Slovakia. *Lauterbornia* 62: 121-127.
- 853 Piechocki, A., 1979. Mieczeniaki (Mollusca), Ślimaki (Gastropoda). Warszawa – Poznań:
854 Państwowe Wydaw. naukowe (Fauna słodkowodna Polski), 186 pp.
- 855 Řezníčková, P., Soldán, T., Pařil, P., Zahrádková, S., 2010. Comparison of mayfly
856 (Ephemeroptera) taxocenes of permanent and intermittent Central European small
857 streams via species traits. *Biologia* 65: 720-729.
- 858 Rüttger, J., Robinson, Ch.T., 2004. Comparison of macroinvertebrate assemblages of
859 permanent and temporary streams in an Alpine flood plain, Switzerland. *Arch. Hydrobiol.*
860 161, 489-510.
- 861 Sarriquet, P.E., Bordenave, P., Marmonier, P., 2007. Effects of bottom sediment restoration
862 on interstitial habitat characteristics and benthic macroinvertebrate assemblages in a
863 headwater stream. *River Research Application* 23: 815–828.
- 864 Smith, H., Wood, P.J., 2002. Flow permanence and macroinvertebrate community variability
865 in limestone spring systems. *Hydrobiologia* 487: 45-58
- 866 Smith, H., Wood, P.J., Gunn, J., 2003. The influence of habitat structure and flow
867 permanence on invertebrate communities in karst spring systems. *Hydrobiologia* 510: 53-
868 66.
- 869 Soldán, T., Zahrádková, S., 2000. Ephemeroptera of the Czech Republic: Atlas of Distribution.
870 In: Helešic J. and Zahrádková S. (Eds): *Fauna Aquatica Europae Centralis I*. Masaryk
871 University, Brno, 401 pp.
- 872 Sommerhäuser, M., Robet, B., Schuhmacher, H., 1995. Flight Periods and Life Strategies of
873 Caddisflies in Temporary and Permanent Woodland Brooks in the Lower Rhine Area. Proc.
874 8th Int. Symp. Trich., Minneapolis/St.Paul, 1995.
- 875 Stubbington, R., Greenwood, A.M., Wood, P.J., Armitage, P.D., Gunn, J., Robertson, A.L.,
876 2009. The response of perennial and temporary headwater stream invertebrate
877 communities to hydrological extremes. *Hydrobiologia* 630: 299-312.

- 878 Svensson, B.W., 1974. Population movements of adult Trichoptera at a South Swedish
879 stream. *Oikos*, 25: 157-175.
- 880 Wagner, R., Marxsen, J., Zwick, P., Cox, E. J. (eds.), 2011. Central European Stream
881 Ecosystems: The long term study of the Breitenbach. Wiley-Blackwell, 672 pp.
- 882 Wood, P.J., Gunn, J., Smith, H., Abas-Kutty, A., 2005. Flow permanence and
883 macroinvertebrate community diversity within groundwater dominated headwater
884 streams and springs. *Hydrobiologia* 545: 55-64.
- 885 Young, J.O., 2001. Keys to the freshwater Microturbellarians of Britain and Ireland with notes
886 on their ecology. Freshwater Biological Association Scientific Publication 59, 142 pp.,
887 Ambleside, Cumbria, kart
888

889 **Appendix B**

890 Table B.1: List of benthic macroinvertebrates collected during the study and their frequency of
 891 occurrence and mean density in three flow intermittence categories.

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
Coelenterata						
<i>Hydra</i> sp.	0.02	0.00	0.00	0.13	0.00	0.00
Turbellaria						
<i>Crenobia alpina</i>	0.02	0.05	0.04	0.33	0.11	0.07
<i>Dendrocoelum lacteum</i>	0.02	0.05	0.00	0.02	0.42	0.00
<i>Dugesia gonocephala</i>	0.80	0.26	0.07	42.82	1.89	0.15
<i>Dugesia polychroa</i>	0.10	0.00	0.00	2.66	0.00	0.00
<i>Polycelis tenuis</i>	0.02	0.05	0.00	0.03	0.42	0.00
Mollusca						
<i>Ancylus fluviatilis</i>	0.31	0.32	0.07	12.79	6.11	0.19
<i>Bythinella austriaca</i> agg.	0.16	0.42	0.56	2.51	2.58	29.22
<i>Galba truncatula</i>	0.11	0.16	0.33	0.36	3.16	5.81
<i>Gyraulus albus</i>	0.02	0.05	0.00	0.13	0.11	0.00
<i>Gyrinus substriatus</i>	0.03	0.00	0.00	0.11	0.00	0.00
<i>Musculium lacustre</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Pisidium</i> sp.	0.77	1.00	0.89	113.05	254.16	76.67
<i>Potamopyrgus antipodarum</i>	0.05	0.16	0.04	0.89	0.84	6.56
<i>Radix ovata</i>	0.08	0.05	0.00	6.90	0.26	0.00
<i>Radix peregra</i>	0.00	0.00	0.11	0.00	0.00	5.67
Nematoida						
Nematoda	0.31	0.26	0.26	1.89	0.95	3.04
Nematomorpha	0.02	0.00	0.04	0.02	0.00	0.11
Oligochaeta						
<i>Achaeta</i> sp.	0.00	0.00	0.07	0.00	0.00	0.22
<i>Aulodrilus japonicus</i>	0.15	0.16	0.04	2.57	10.53	0.30
<i>Aulodrilus limnobius</i>	0.00	0.05	0.00	0.00	0.11	0.00
<i>Aulodrilus plurisetia</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Bryodrilus</i> sp.	0.00	0.00	0.04	0.00	0.00	1.04
<i>Cernosvitoviella</i> sp.	0.07	0.11	0.04	0.43	1.26	0.59
<i>Cognettia glandulosa</i>	0.13	0.16	0.33	1.15	1.74	6.52
<i>Cognettia sphagnetorum</i>	0.23	0.00	0.19	2.82	0.00	7.70
<i>Eiseniella tetraedra</i>	0.57	0.79	0.89	5.34	6.74	41.30
<i>Enchytraeus</i> sp.	0.05	0.11	0.22	0.39	0.21	22.59
<i>Epirodilus pygmaeus</i>	0.08	0.32	0.15	0.36	4.21	3.33
<i>Haplotaxis gordioides</i>	0.20	0.16	0.41	0.46	0.42	1.85
<i>Henlea/Fridericia</i> sp.	0.52	0.47	0.89	6.48	4.05	56.63
<i>Limnodrilus claparedeanus</i>	0.05	0.00	0.00	0.48	0.00	0.00
<i>Limnodrilus hoffmeisteri</i>	0.38	0.37	0.22	7.69	9.16	3.89
<i>Limnodrilus profundicola</i>	0.00	0.05	0.00	0.00	0.05	0.00

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Limnodrilus udekemianus</i>	0.03	0.05	0.00	0.16	0.16	0.00
Lumbricidae Gen. sp.	0.26	0.21	0.52	0.57	0.32	1.89
<i>Lumbriculus variegatus</i>	0.33	0.37	0.11	4.10	2.26	0.93
<i>Marionina</i> sp.	0.13	0.26	0.33	1.48	1.58	6.44
<i>Mesenchytraeus armatus</i>	0.10	0.00	0.33	0.72	0.00	2.48
<i>Nais alpina</i>	0.38	0.16	0.04	17.67	10.26	0.07
<i>Nais barbata</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Nais bretscheri</i>	0.10	0.05	0.00	0.92	0.95	0.00
<i>Nais communis</i>	0.36	0.32	0.30	6.02	20.37	5.63
<i>Nais elinguis</i>	0.18	0.37	0.33	11.13	22.58	122.59
<i>Nais christinae</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Nais pardalis</i>	0.02	0.05	0.00	0.13	1.00	0.00
<i>Nais stolci</i>	0.05	0.00	0.00	2.56	0.00	0.00
<i>Nais variabilis</i>	0.02	0.16	0.07	0.03	14.53	2.74
<i>Ophidonais serpentina</i>	0.16	0.05	0.04	3.77	0.32	1.19
<i>Pristina aequisetata</i>	0.03	0.11	0.00	0.13	1.05	0.00
<i>Pristinella rosea</i> Gr.	0.25	0.16	0.22	3.74	1.00	0.81
<i>Propappus volki</i>	0.28	0.00	0.00	33.89	0.00	0.00
<i>Psammoryctides barbatus</i>	0.26	0.32	0.11	7.39	16.84	3.00
<i>Rhyacodrilus coccineus</i>	0.15	0.16	0.00	2.00	4.26	0.00
<i>Rhyacodrilus falciformis</i>	0.23	0.26	0.26	1.08	0.58	1.59
<i>Rhyacodrilus subterraneus</i>	0.05	0.00	0.00	0.79	0.00	0.00
<i>Rhypholophus haemorrhoidalis</i>	0.03	0.05	0.37	0.10	0.42	1.19
<i>Slavina appendiculata</i>	0.05	0.05	0.00	3.84	0.42	0.00
<i>Spirosperma ferrox</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Stylodrilus brachystylus</i>	0.28	0.26	0.07	47.69	82.42	38.67
<i>Stylodrilus heringianus</i>	0.64	0.53	0.67	48.97	37.63	289.52
<i>Trichodrilus</i> sp.	0.23	0.05	0.15	6.74	0.63	0.59
<i>Trichodrilus strandi</i>	0.11	0.11	0.04	12.98	3.26	0.89
<i>Tubifex tubifex</i>	0.54	0.63	0.22	72.00	7.00	2.89

Hirudinea

<i>Erpobdella octoculata</i>	0.13	0.11	0.07	0.87	0.16	0.19
<i>Erpobdella vilnensis</i>	0.20	0.21	0.07	0.54	0.47	0.19
<i>Glossiphonia complanata</i>	0.03	0.11	0.04	0.10	0.16	0.07
<i>Glossiphonia nebulosa</i>	0.02	0.05	0.00	0.07	0.11	0.00
<i>Glossosoma conformis</i>	0.05	0.00	0.00	0.34	0.00	0.00
<i>Haemopsis sanguisuga</i>	0.02	0.11	0.11	0.03	0.21	0.48
<i>Helobdella stagnalis</i>	0.02	0.00	0.00	0.02	0.00	0.00

Acari

Hydracarina	0.43	0.32	0.15	9.43	1.21	0.48
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Crustacea

<i>Asellus aquaticus</i>	0.05	0.16	0.07	0.21	8.32	0.26
Copepoda	0.07	0.00	0.11	0.56	0.00	1.07

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Gammarus fossarum</i>	0.79	0.89	0.63	1811.13	2533.32	43.26
<i>Gammarus roeselii</i>	0.10	0.16	0.11	3.39	3.37	0.96
<i>Niphargus aquilex</i>	0.02	0.00	0.19	0.02	0.00	4.89
Ephemeroptera						
<i>Baetis alpinus</i>	0.02	0.00	0.00	0.15	0.00	0.00
<i>Baetis buceratus</i>	0.03	0.00	0.00	0.18	0.00	0.00
<i>Baetis fuscatus</i>	0.02	0.11	0.00	0.03	1.37	0.00
<i>Baetis liebenauae</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Baetis muticus</i>	0.84	0.47	0.11	83.92	9.68	1.70
<i>Baetis niger</i>	0.16	0.16	0.00	3.95	0.37	0.00
<i>Baetis rhodani</i>	0.97	0.89	0.44	229.98	103.63	11.93
<i>Baetis vernus</i>	0.11	0.26	0.07	3.07	9.47	0.48
<i>Caenis luctuosa</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Caenis macrura</i>	0.02	0.05	0.04	0.13	0.05	0.15
<i>Caenis pseudorivulorum</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Centroptilum luteolum</i>	0.64	0.53	0.44	26.44	58.63	43.44
<i>Centroptilum pennulatum</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Cloeon dipterum</i> s. lat.	0.00	0.05	0.11	0.00	0.68	11.19
<i>Ecdyonurus aurantiacus</i>	0.02	0.05	0.00	0.28	0.21	0.00
<i>Ecdyonurus austriacus</i>	0.07	0.00	0.07	0.66	0.00	1.07
<i>Ecdyonurus dispar</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Ecdyonurus helveticus</i> -Gr.	0.07	0.05	0.04	0.80	0.21	0.04
<i>Ecdyonurus macani</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Ecdyonurus picteti</i>	0.05	0.05	0.04	0.75	0.37	0.04
<i>Ecdyonurus starmachi</i>	0.00	0.00	0.04	0.00	0.00	0.11
<i>Ecdyonurus subalpinus</i>	0.36	0.26	0.22	50.46	13.63	1.78
<i>Ecdyonurus submontanus</i>	0.02	0.00	0.00	0.20	0.00	0.00
<i>Ecdyonurus torrentis</i>	0.43	0.32	0.00	24.41	2.95	0.00
<i>Ecdyonurus venosus</i>	0.07	0.00	0.00	0.57	0.00	0.00
<i>Electrogena lateralis</i>	0.05	0.11	0.00	0.62	0.21	0.00
<i>Electrogena quadrilineata</i>	0.03	0.00	0.07	6.64	0.00	1.30
<i>Electrogena ujhelyii</i>	0.57	0.58	0.26	87.93	118.37	2.81
<i>Epeorus assimilis</i>	0.28	0.00	0.00	7.72	0.00	0.00
<i>Ephemerella danica</i>	0.75	0.63	0.04	40.80	26.68	0.44
<i>Ephemerella mucronata</i>	0.10	0.00	0.00	5.02	0.00	0.00
<i>Ephemerella notata</i>	0.02	0.00	0.04	0.33	0.00	0.07
<i>Habroleptoides confusa</i>	0.79	0.74	0.44	211.72	120.89	16.96
<i>Habrophlebia fusca</i>	0.11	0.32	0.15	1.95	5.84	5.15
<i>Habrophlebia lauta</i>	0.87	0.79	0.30	78.84	72.63	16.48
<i>Leptophlebia marginata</i>	0.02	0.00	0.00	0.02	0.00	0.00
<i>Metreletus balcanicus</i>	0.00	0.00	0.07	0.00	0.00	0.63
<i>Paraleptophlebia submarginata</i>	0.56	0.53	0.22	18.00	7.11	10.19
<i>Rhithrogena carpatoalpina</i>	0.28	0.32	0.04	29.02	1.68	0.04

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Rhithrogena iridina/picteti</i>	0.70	0.32	0.30	121.93	1.16	1.89
<i>Rhithrogena semicolorata</i>	0.08	0.00	0.04	5.61	0.00	0.07
<i>Siphonurus aestivalis</i>	0.05	0.05	0.19	0.25	0.68	18.52
<i>Siphonurus armatus</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Siphonurus lacustris</i>	0.00	0.05	0.00	0.00	0.21	0.00
<i>Torleya major</i>	0.26	0.00	0.00	11.51	0.00	0.00

Odonata

<i>Aeshna cyanea</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Calopteryx virgo</i>	0.15	0.05	0.07	0.66	0.11	0.26
<i>Cordulegaster bidentata</i>	0.00	0.05	0.00	0.00	0.11	0.00
<i>Cordulegaster boltonii</i>	0.13	0.11	0.00	0.66	0.11	0.00

Plecoptera

<i>Amphinemura</i> sp.	0.30	0.16	0.07	21.28	0.53	1.44
<i>Brachyptera risi</i>	0.28	0.21	0.41	8.03	26.63	83.52
<i>Brachyptera seticornis</i>	0.03	0.00	0.00	1.00	0.00	0.00
<i>Capnia bifrons</i>	0.26	0.37	0.41	11.39	44.58	94.26
<i>Dinocras cephalotes</i>	0.08	0.00	0.00	2.84	0.00	0.00
<i>Diura bicaudata</i>	0.05	0.00	0.00	0.08	0.00	0.00
<i>Isoperla grammatica</i>	0.11	0.16	0.19	6.64	7.00	27.19
<i>Isoperla oxylepis</i>	0.23	0.21	0.00	11.70	20.42	0.00
<i>Isoperla rivulorum</i>	0.10	0.05	0.00	1.77	0.47	0.00
<i>Isoperla tripartita</i>	0.11	0.37	0.41	4.61	16.63	35.85
<i>Leuctra braueri</i>	0.05	0.00	0.04	0.18	0.00	0.07
<i>Leuctra nigra</i>	0.15	0.11	0.04	1.11	0.68	0.22
<i>Leuctra pseudosignifera</i>	0.10	0.00	0.00	0.46	0.00	0.00
<i>Leuctra</i> sp.	0.72	0.79	0.33	163.98	7.74	2.56
<i>Nemoura</i> sp.	0.84	1.00	0.74	179.26	188.84	152.00
<i>Nemurella picteti</i>	0.02	0.00	0.00	0.15	0.00	0.00
<i>Perla burmeisteriana</i>	0.07	0.00	0.00	0.25	0.00	0.00
<i>Perla marginata</i>	0.03	0.00	0.00	0.13	0.00	0.00
<i>Perlodes microcephalus</i>	0.10	0.00	0.00	0.13	0.00	0.00
<i>Protonemura</i> sp.	0.48	0.11	0.11	47.62	0.63	1.93
<i>Siphonoperla</i> sp.	0.20	0.16	0.00	7.59	1.79	0.00
<i>Taeniopteryx</i> sp.	0.02	0.00	0.00	0.03	0.00	0.00

Heteroptera

<i>Aquarius paludum</i>	0.02	0.00	0.00	0.02	0.00	0.00
<i>Gerris lacustris</i>	0.05	0.05	0.00	0.18	0.05	0.00
<i>Gerris odontogaster</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Micronecta</i> sp.	0.07	0.05	0.04	2.44	1.58	0.11
<i>Velia caprai</i>	0.03	0.05	0.04	0.07	0.05	0.04
<i>Velia saulii</i>	0.05	0.05	0.07	0.07	0.26	0.30

Neuroptera

<i>Osmylus fulvicephalus</i>	0.02	0.00	0.00	0.02	0.00	0.00
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	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
Megaloptera						
<i>Sialis fuliginosa</i>	0.49	0.63	0.26	4.30	10.32	2.44
Trichoptera						
<i>Agapetus fuscipes</i>	0.08	0.00	0.00	4.79	0.00	0.00
<i>Allogamus auricollis</i>	0.00	0.05	0.00	0.00	0.05	0.00
<i>Allogamus uncatus</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Athripsodes bilineatus</i>	0.26	0.21	0.19	3.98	1.47	0.33
<i>Beraeodes minutus</i>	0.00	0.16	0.15	0.00	0.32	0.22
<i>Brachycentrus montanus</i>	0.08	0.00	0.00	1.02	0.00	0.00
<i>Crunoecia irrorata</i>	0.00	0.00	0.07	0.00	0.00	0.19
<i>Cyrnus trimaculatus</i>	0.07	0.05	0.00	0.21	0.11	0.00
<i>Drusus annulatus</i>	0.08	0.26	0.11	0.61	1.58	0.22
<i>Ecclisopteryx dalecarlica</i>	0.07	0.00	0.07	0.79	0.00	0.07
<i>Ecclisopteryx madida</i>	0.05	0.05	0.00	1.34	0.11	0.00
<i>Glyphotaelius pellucidus</i>	0.05	0.05	0.30	0.16	0.05	2.30
<i>Halesus</i> sp.	0.31	0.32	0.22	12.25	5.26	1.78
<i>Hydatophylax infumatus</i>	0.02	0.00	0.00	0.02	0.00	0.00
<i>Hydropsyche</i> sp.	0.93	0.63	0.19	93.03	11.63	0.96
<i>Chaetopteryx fusca</i>	0.02	0.05	0.00	0.02	3.63	0.00
<i>Chaetopteryx major</i>	0.16	0.05	0.11	3.20	5.53	0.96
<i>Chaetopteryx villosa</i>	0.07	0.11	0.30	0.46	0.21	0.48
<i>Ironoquia dubia</i>	0.00	0.00	0.07	0.00	0.00	0.11
<i>Lasiocephala basalis</i>	0.11	0.00	0.00	0.48	0.00	0.00
<i>Lepidostoma hirtum</i>	0.10	0.00	0.00	7.30	0.00	0.00
<i>Limnephilus lunatus</i>	0.00	0.05	0.00	0.00	0.11	0.00
<i>Limnephilus rhombicus</i>	0.02	0.05	0.00	0.13	0.05	0.00
<i>Limnephilus</i> sp.	0.02	0.11	0.04	0.07	0.16	0.04
<i>Lithax obscurus</i>	0.03	0.00	0.00	0.05	0.00	0.00
<i>Lype</i> sp.	0.43	0.32	0.04	2.75	1.47	0.07
<i>Micrasema longulum</i>	0.05	0.00	0.00	0.30	0.00	0.00
<i>Micrasema minimum</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Micropterna nycterobia</i>	0.02	0.26	0.26	0.07	1.32	1.41
<i>Micropterna</i> sp.	0.02	0.05	0.26	0.03	0.42	12.37
<i>Mystacides azurea</i>	0.10	0.05	0.00	1.05	0.21	0.00
<i>Odontocerum albicorne</i>	0.62	0.32	0.07	8.33	1.16	0.30
<i>Oecismus monedula</i>	0.21	0.00	0.04	1.16	0.00	0.15
<i>Oligostomis reticulata</i>	0.02	0.00	0.00	0.02	0.00	0.00
<i>Philopotamus ludificatus</i>	0.08	0.00	0.00	2.03	0.00	0.00
<i>Philopotamus montanus</i>	0.26	0.00	0.00	20.95	0.00	0.00
<i>Plectrocnemia brevis</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Plectrocnemia conspersa</i>	0.18	0.32	0.30	7.97	12.53	8.11
<i>Polycentropus flavomaculatus</i>	0.49	0.37	0.04	14.56	4.84	0.07
<i>Polycentropus irroratus</i>	0.11	0.05	0.00	1.13	0.42	0.00

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Potamophylax</i> sp.	0.59	0.37	0.15	8.95	2.53	0.89
<i>Psychomyia pusilla</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Ptilocolepus granulatus</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Rhyacophila</i> cf. <i>fasciata</i>	0.21	0.11	0.04	1.77	0.84	0.07
<i>Rhyacophila nubila</i>	0.11	0.05	0.00	1.66	0.79	0.00
<i>Rhyacophila polonica</i>	0.08	0.00	0.07	0.69	0.00	0.22
<i>Rhyacophila tristis</i>	0.11	0.05	0.00	2.33	0.05	0.00
<i>Sericostoma</i> sp.	0.82	0.68	0.19	21.85	15.63	0.41
<i>Silo nigricornis</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Silo pallipes</i>	0.21	0.11	0.00	1.93	1.05	0.00
<i>Stenophylax permistus</i>	0.00	0.11	0.11	0.00	0.26	0.19
<i>Synagapetus iridipennis</i>	0.03	0.00	0.04	0.26	0.00	0.15
<i>Synagapetus moselyi</i>	0.07	0.11	0.22	0.23	1.89	18.67
<i>Tinodes</i> sp.	0.46	0.37	0.04	7.34	2.79	0.15

Coleoptera

<i>Agabus guttatus</i>	0.03	0.32	0.67	0.13	1.21	12.22
<i>Anacaena globulus</i>	0.02	0.11	0.11	0.03	0.16	0.19
<i>Contacyphon</i> sp.	0.00	0.00	0.11	0.00	0.00	1.11
<i>Deronectes latus</i>	0.05	0.00	0.00	0.10	0.00	0.00
<i>Elmis aenea</i>	0.46	0.37	0.15	25.64	1.16	1.19
<i>Elmis latreillei</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Elmis maugetii</i>	0.93	0.84	0.67	86.97	138.89	14.37
<i>Odeles</i> sp.	0.25	0.00	0.04	1.34	0.00	0.15
<i>Elodes</i> sp.	0.43	0.84	0.26	5.74	19.16	0.48
<i>Esolus angustatus</i>	0.11	0.21	0.22	2.07	1.84	7.78
<i>Esolus parallelepipedus</i>	0.16	0.00	0.15	1.72	0.00	0.30
<i>Haliphus lineatocollis</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Haliphus</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Hydraena belgica</i>	0.05	0.00	0.00	1.20	0.00	0.00
<i>Hydraena dentipes</i>	0.11	0.00	0.04	4.23	0.00	0.07
<i>Hydraena excisa</i>	0.23	0.32	0.41	3.20	13.63	5.04
<i>Hydraena gracilis</i>	0.89	0.79	0.52	71.30	58.37	13.33
<i>Hydraena melas</i>	0.05	0.00	0.07	0.46	0.00	0.15
<i>Hydraena nigrita</i>	0.23	0.42	0.41	2.36	5.00	2.00
<i>Hydraena paganettii</i>	0.00	0.05	0.00	0.00	0.21	0.00
<i>Hydraena pygmaea</i>	0.05	0.00	0.00	0.38	0.00	0.00
<i>Hydraena riparia</i>	0.41	0.32	0.26	9.13	5.37	1.93
<i>Hydraena rufipes</i>	0.02	0.00	0.04	0.07	0.00	0.07
<i>Hydraena saga</i>	0.13	0.00	0.00	0.66	0.00	0.00
<i>Hydraena schuleri</i>	0.11	0.05	0.00	0.74	0.21	0.00
<i>Hydraena subimpressa</i>	0.03	0.00	0.00	0.15	0.00	0.00
<i>Hydraena truncata</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Hydrocyphon deflexicollis</i>	0.02	0.00	0.00	0.03	0.00	0.00

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Hydroporus palustris</i>	0.00	0.05	0.00	0.00	0.05	0.00
<i>Hydroporus planus</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Ilybius</i> sp.	0.02	0.05	0.00	0.07	0.32	0.00
<i>Laccobius bipunctatus</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Laccobius obscuratus</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Limnebius truncatellus</i>	0.13	0.00	0.04	0.49	0.00	0.04
<i>Limnius perrisi</i>	0.39	0.11	0.07	54.25	0.37	0.96
<i>Limnius volckmari</i>	0.82	0.79	0.33	34.20	49.63	4.48
<i>Orectochilus villosus</i>	0.15	0.05	0.00	2.57	0.32	0.00
<i>Oreodytes sanmarkii</i>	0.03	0.00	0.00	0.05	0.00	0.00
<i>Oulimnius tuberculatus</i>	0.28	0.21	0.04	3.41	1.58	0.11
<i>Platambus maculatus</i>	0.33	0.68	0.44	2.80	26.74	7.07
<i>Riolus cupreus</i>	0.02	0.05	0.00	0.07	1.58	0.00
<i>Riolus subviolaceus</i>	0.08	0.16	0.00	3.72	4.53	0.00

Diptera (excl. Chironomidae)

<i>Acanthocnema glaucescens</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Antocha</i> sp.	0.10	0.05	0.00	0.62	0.11	0.00
<i>Atherix ibis</i>	0.05	0.00	0.00	0.34	0.00	0.00
<i>Berdeniella</i> sp.	0.15	0.00	0.00	2.21	0.00	0.00
<i>Beris vallata</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Bezzia</i> sp.	0.51	0.47	0.33	6.18	10.84	3.33
Ceratopogonidae	0.21	0.16	0.19	1.54	0.84	1.37
<i>Clinocera/Wiedemannia</i> sp.	0.11	0.05	0.00	1.87	0.53	0.00
<i>Clytocerus</i> sp.	0.00	0.00	0.07	0.00	0.00	0.37
<i>Dicranomyia</i> sp.	0.00	0.00	0.04	0.00	0.00	0.30
<i>Dicranota</i> sp.	0.70	0.63	0.30	20.48	11.21	0.81
<i>Dixa</i> sp.	0.28	0.21	0.07	1.48	1.32	0.15
Dolichopodidae	0.10	0.11	0.04	0.49	0.21	0.07
<i>Eloeophila</i> sp.	0.69	0.63	0.19	8.95	14.26	0.59
Ephydridae	0.03	0.00	0.00	0.10	0.00	0.00
<i>Erioptera</i> sp.	0.02	0.00	0.00	0.07	0.00	0.00
<i>Fannia</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Gonomyia</i> sp.	0.03	0.00	0.00	0.10	0.00	0.00
<i>Hemerodromia</i> sp.	0.28	0.11	0.04	11.26	0.37	0.15
<i>Hexatoma</i> sp.	0.36	0.16	0.37	2.49	3.05	3.30
<i>Chelifera</i> sp.	0.44	0.42	0.37	11.87	7.95	15.63
<i>Chrysopilus auratus</i>	0.10	0.05	0.26	0.48	0.11	0.74
<i>Chrysops</i> sp.	0.38	0.47	0.33	3.05	9.47	1.89
<i>Ibisia marginata</i>	0.46	0.16	0.07	47.87	0.42	0.26
<i>Jungiella</i> sp.	0.02	0.00	0.00	0.46	0.00	0.00
<i>Limnophila</i> sp.	0.03	0.00	0.07	0.20	0.00	0.11
<i>Limonia</i> sp.	0.03	0.00	0.11	0.16	0.00	0.30
<i>Molophilus</i> sp.	0.11	0.26	0.26	0.54	1.16	22.56

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
Muscidae	0.08	0.05	0.11	0.46	0.11	0.56
<i>Nemotelus</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Neolimnomyia</i> sp.	0.05	0.11	0.00	0.13	0.21	0.00
<i>Orimarga</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Oxycera</i> sp.	0.10	0.11	0.22	0.31	1.42	6.89
<i>Pedicia</i> sp.	0.31	0.53	0.19	1.90	3.95	1.00
<i>Pericoma</i> sp.	0.11	0.37	0.19	2.18	6.26	0.56
<i>Peripsychoda</i> sp.	0.07	0.05	0.00	0.59	0.84	0.00
<i>Pilaria</i> sp.	0.11	0.11	0.11	0.46	0.26	0.41
<i>Pneumia</i> sp.	0.07	0.11	0.00	0.52	0.16	0.00
<i>Prosimulium hirtipes</i>	0.10	0.00	0.00	19.67	0.00	0.00
<i>Prosimulium latimucro</i>	0.00	0.00	0.04	0.00	0.00	1.96
<i>Prosimulium rufipes</i>	0.00	0.00	0.04	0.00	0.00	1.04
<i>Prosimulium tomosvaryi</i>	0.33	0.42	0.37	130.51	65.21	86.96
<i>Psychoda</i> sp.	0.02	0.00	0.04	0.03	0.00	0.04
<i>Ptychoptera</i> sp.	0.41	0.32	0.04	13.93	17.58	0.07
Sciomyzidae	0.00	0.00	0.07	0.00	0.00	0.15
<i>Scleroprocta</i> sp.	0.20	0.16	0.04	0.85	0.32	0.04
<i>Simulium angustipes</i>	0.15	0.26	0.19	7.20	6.11	4.63
<i>Simulium angustitarse</i>	0.08	0.05	0.11	0.28	0.84	0.30
<i>Simulium argyreatum</i>	0.08	0.05	0.07	5.84	0.11	0.11
<i>Simulium brevidens</i>	0.02	0.00	0.00	1.05	0.00	0.00
<i>Simulium carpathicum</i>	0.00	0.00	0.04	0.00	0.00	0.04
<i>Simulium costatum</i>	0.10	0.05	0.04	2.48	0.05	0.15
<i>Simulium cryophilum</i>	0.16	0.11	0.07	3.89	1.16	12.78
<i>Simulium equinum</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Simulium noelleri</i>	0.00	0.11	0.11	0.00	1.16	1.33
<i>Simulium ornatum</i>	0.28	0.42	0.19	3.10	131.68	0.63
<i>Simulium reptans</i>	0.05	0.00	0.00	1.44	0.00	0.00
<i>Simulium trifasciatum</i>	0.00	0.00	0.04	0.00	0.00	0.33
<i>Simulium variegatum</i>	0.11	0.00	0.04	7.67	0.00	0.11
<i>Simulium vernum</i>	0.31	0.58	0.37	7.54	10.11	5.26
<i>Tabanus miki</i>	0.00	0.00	0.04	0.00	0.00	0.07
<i>Tabanus</i> sp.	0.03	0.00	0.04	0.05	0.00	0.07
<i>Tipula lateralis</i>	0.00	0.05	0.00	0.00	0.21	0.00
<i>Tipula maxima</i>	0.28	0.68	0.44	1.66	9.37	1.41
<i>Tipula saginata</i>	0.20	0.16	0.07	0.82	1.89	0.15
<i>Tipula unca</i>	0.02	0.00	0.07	0.07	0.00	0.15
<i>Tipula vittata</i>	0.00	0.00	0.04	0.00	0.00	0.19
Chironomidae						
<i>Ablabesmyia</i> sp.	0.08	0.05	0.07	1.41	0.47	0.37
<i>Apsectrotanypus trifascipennis</i>	0.39	0.53	0.15	3.28	51.00	0.78
<i>Brillia bifida</i>	0.62	0.89	0.78	25.57	69.26	29.11

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Brillia longifurca</i>	0.21	0.11	0.07	0.87	0.32	0.15
<i>Bryophaenocladus</i> sp.	0.02	0.05	0.00	0.13	0.11	0.00
<i>Cladotanytarsus vanderwulpi</i> -Gr.	0.03	0.00	0.00	0.26	0.00	0.00
<i>Conchapelopia</i> sp.	0.74	0.74	0.48	24.93	36.63	10.96
<i>Corynoneura coronata</i> -Gr.	0.02	0.00	0.04	0.23	0.00	0.07
<i>Corynoneura lobata</i>	0.31	0.37	0.37	2.95	5.21	4.41
<i>Cricotopus albiforceps</i>	0.00	0.05	0.00	0.00	0.11	0.00
<i>Cricotopus bicinctus</i>	0.03	0.00	0.04	0.30	0.00	0.15
<i>Cricotopus patens/flavocinctus</i>	0.02	0.00	0.00	0.26	0.00	0.00
<i>Cricotopus</i> sp.	0.10	0.05	0.00	1.66	0.11	0.00
<i>Cricotopus sylvestris</i> -Gr.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Cricotopus tremulus</i> -Gr.	0.07	0.11	0.00	1.21	0.47	0.00
<i>Cricotopus trifascia</i>	0.02	0.00	0.00	0.79	0.00	0.00
<i>Cryptochironomus</i> sp.	0.03	0.00	0.00	0.10	0.00	0.00
<i>Demicryptochironomus</i> sp.	0.03	0.00	0.00	0.20	0.00	0.00
<i>Diamesa cinerella</i> -Gr.	0.20	0.26	0.30	30.69	21.68	51.41
<i>Dicrotendipes</i> sp.	0.00	0.00	0.04	0.00	0.00	0.59
<i>Diplocladius cultriger</i>	0.16	0.53	0.52	0.97	19.95	24.37
<i>Epoicocladius ephemerae</i>	0.51	0.32	0.00	5.10	9.32	0.00
<i>Eukiefferiella breviculcar</i>	0.34	0.32	0.30	7.11	5.84	5.52
<i>Eukiefferiella claripennis</i>	0.00	0.21	0.04	0.00	2.42	0.37
<i>Eukiefferiella</i> cf. <i>clypeata</i>	0.03	0.00	0.00	0.08	0.00	0.00
<i>Eukiefferiella devonica/ilkeleyensis</i>	0.10	0.16	0.00	0.98	1.37	0.00
<i>Eukiefferiella gracei</i> -Gr.	0.07	0.05	0.04	0.39	0.95	0.15
<i>Eukiefferiella minor/fittkau</i>	0.00	0.00	0.04	0.00	0.00	0.30
<i>Eukiefferiella rectangularis</i> -Gr.	0.03	0.05	0.00	0.23	0.11	0.00
<i>Eukiefferiella similis</i>	0.02	0.00	0.00	0.20	0.00	0.00
<i>Georthocladus luteicornis</i>	0.02	0.00	0.00	0.03	0.00	0.00
<i>Gymnometriocnemus</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Heleniella ornaticollis</i>	0.33	0.32	0.11	7.54	2.47	2.30
<i>Heterotrissocladius</i> cf. <i>scutellatus</i>	0.00	0.00	0.04	0.00	0.00	0.15
<i>Heterotrissocladius marcidus</i>	0.18	0.16	0.19	3.13	2.42	4.93
<i>Hydrobaenus</i> sp.	0.00	0.00	0.11	0.00	0.00	1.85
<i>Chaetocladius dentiforceps</i> -Gr.	0.02	0.00	0.04	0.07	0.00	0.11
<i>Chaetocladius piger</i> -Gr.	0.05	0.11	0.19	0.67	1.16	1.59
<i>Chaetocladius vitellinus</i> -Gr.	0.03	0.00	0.00	0.16	0.00	0.00
<i>Chironomus</i> sp.	0.05	0.11	0.15	0.21	0.68	47.89
<i>Krenopsectra</i> sp.	0.11	0.05	0.19	6.49	1.58	28.78
<i>Krenosmittia camptophleps</i>	0.00	0.00	0.15	0.00	0.00	4.44
<i>Limnophyes</i> sp.	0.11	0.26	0.30	0.62	0.84	5.00
<i>Macropelopia</i> sp.	0.05	0.21	0.26	0.43	0.89	15.30
<i>Metriocnemus fuscipes</i> -Gr.	0.00	0.05	0.00	0.00	0.42	0.00
<i>Micropsectra apposita</i> -Gr.	0.13	0.21	0.44	3.21	14.74	13.93

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Micropsectra atrofasciata</i> -Gr.	0.80	0.89	0.89	161.33	1816.00	223.70
<i>Microtendipes pedellus</i> -Gr.	0.59	0.63	0.30	19.36	54.00	19.48
<i>Microtendipes rydalensis</i> -Gr.	0.13	0.05	0.00	11.80	0.11	0.00
<i>Monodiamesa bathyphila</i>	0.05	0.00	0.00	0.20	0.00	0.00
<i>Nanocladius parvulus/rectinervis</i>	0.05	0.11	0.07	0.16	0.95	0.15
<i>Natarsia</i> sp.	0.25	0.42	0.59	0.90	3.74	4.44
<i>Nilotanytus dubius</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Orthocladius lignicola</i>	0.10	0.05	0.19	0.36	0.42	0.63
<i>Orthocladius obumbratus/oblidens</i>	0.34	0.37	0.33	69.77	21.16	25.07
<i>Orthocladius rivicola</i> -Gr.	0.11	0.26	0.26	6.31	2.42	39.56
<i>Orthocladius rivulorum</i>	0.07	0.11	0.04	4.57	0.63	0.22
<i>Orthocladius rubicundus</i>	0.11	0.26	0.11	3.31	5.05	0.37
<i>Paracladopelma</i> sp.	0.02	0.00	0.00	0.02	0.00	0.00
<i>Paracricotopus niger</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Parakiefferiella</i> sp.	0.02	0.11	0.00	0.03	0.47	0.00
<i>Paramerina</i> sp.	0.03	0.05	0.00	0.10	0.42	0.00
<i>Parametriocnemus stylatus</i>	0.80	0.95	0.70	41.02	60.05	193.67
<i>Paraphaenocladius</i> sp.	0.05	0.21	0.48	2.00	3.47	14.44
<i>Parasmittia</i> sp.	0.00	0.00	0.04	0.00	0.00	0.04
<i>Paratendipes albimanus</i> -Gr.	0.03	0.00	0.04	0.08	0.00	0.15
<i>Paratrachocladius rufiventris</i>	0.13	0.21	0.07	5.70	0.95	0.44
<i>Paratrissocladius excerptus</i>	0.25	0.16	0.22	3.43	3.58	1.63
<i>Parorthocladius</i> sp.	0.00	0.00	0.07	0.00	0.00	3.74
<i>Phaenopsectra</i> sp.	0.07	0.05	0.11	0.15	0.16	1.70
<i>Polypedilum albicorne</i>	0.18	0.11	0.26	1.25	0.53	3.26
<i>Polypedilum convictum</i> -Gr.	0.48	0.42	0.33	27.10	10.95	5.48
<i>Polypedilum cultellatum</i>	0.02	0.00	0.00	0.07	0.00	0.00
<i>Polypedilum laetum</i> -Gr.	0.00	0.05	0.00	0.00	0.68	0.00
<i>Polypedilum pedestre</i> -Gr.	0.10	0.05	0.07	0.38	0.11	0.52
<i>Polypedilum scalaenum</i> -Gr.	0.28	0.21	0.11	1.67	2.11	0.44
<i>Potthastia longimana</i>	0.26	0.00	0.00	2.46	0.00	0.00
<i>Procladius</i> sp.	0.08	0.26	0.22	0.28	6.16	1.52
<i>Prodiamesa olivacea</i>	0.33	0.42	0.41	5.34	4.11	17.52
<i>Psectrocladius</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Pseudorthocladius</i> sp.	0.05	0.11	0.07	0.15	0.63	0.74
<i>Pseudosmittia</i> sp.	0.00	0.00	0.04	0.00	0.00	0.22
<i>Rheocricotopus atripes</i>	0.02	0.05	0.19	0.07	0.21	1.41
<i>Rheocricotopus effusus</i>	0.07	0.26	0.63	0.23	5.00	9.48
<i>Rheocricotopus fuscipes</i>	0.62	0.79	0.63	19.79	35.89	13.19
<i>Rheocricotopus chalybeatus</i>	0.00	0.05	0.00	0.00	0.11	0.00
<i>Rheotanytarsus</i> sp.	0.20	0.11	0.04	6.97	0.16	0.04
<i>Smittia</i> sp.	0.00	0.00	0.04	0.00	0.00	0.07
<i>Stempellinella brevis</i>	0.48	0.26	0.37	24.36	41.47	2.22

	Frequency			Density		
	Perennial	Near-perennial	Intermittent	Perennial	Near-perennial	Intermittent
<i>Stictochironomus</i> sp.	0.15	0.11	0.00	0.84	2.37	0.00
<i>Synorthocladius semivirens</i>	0.20	0.11	0.11	3.38	0.53	0.30
<i>Tanytarsus brundini/curticornis</i>	0.11	0.11	0.04	4.23	0.32	0.07
<i>Tanytarsus</i> sp.	0.48	0.74	0.22	27.61	50.58	3.07
<i>Thienemannia</i> sp.	0.02	0.11	0.00	0.03	0.84	0.00
<i>Thienemanniella</i> sp.	0.34	0.21	0.19	7.20	8.53	0.70
<i>Thienemannimyia</i> sp.	0.21	0.11	0.04	5.02	0.63	0.15
<i>Trissopelopia</i> sp.	0.13	0.26	0.19	2.25	34.42	4.63
<i>Tvetenia bavarica/calvescens</i>	0.39	0.32	0.07	40.90	7.79	2.89
<i>Tvetenia discoloripes</i> -Gr.	0.49	0.42	0.22	17.28	19.26	2.00
<i>Zavrelimyia</i> sp.	0.43	0.47	0.59	7.07	22.89	19.15

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