



Vitamin B₁₂ (Cobalamin) and Micronutrient Fortification in Food Crops Using Nanoparticle Technology

Soojin Oh¹, Gareth Cave² and Chungui Lu^{1*}

¹ School of Animal, Rural and Environmental Sciences, Nottingham Trent University, Nottingham, United Kingdom, ² School of Science and Technology, Nottingham Trent University, Nottingham, United Kingdom

OPEN ACCESS

Edited by:

Jung Eek Son,
Seoul National University,
South Korea

Reviewed by:

Dimitrios Savvas,
Agricultural University of Athens,
Greece
Hyo Gil Choi,
Kongju National University,
South Korea

*Correspondence:

Chungui Lu
Chungui.Lu@ntu.ac.uk

Specialty section:

This article was submitted to
Technical Advances in Plant Science,
a section of the journal
Frontiers in Plant Science

Received: 17 February 2021

Accepted: 22 June 2021

Published: 23 August 2021

Citation:

Oh S, Cave G and Lu C (2021)
Vitamin B₁₂ (Cobalamin)
and Micronutrient Fortification in Food
Crops Using Nanoparticle
Technology.
Front. Plant Sci. 12:668819.
doi: 10.3389/fpls.2021.668819

It is necessary to develop a resilient food supply that will withstand unexpected future shocks and deliver the required amounts of nutrients to consumers. By increasing the sustainability of food and agriculture, the food system will be able to handle challenges such as climate change, declining agricultural resources, growing population/urbanization, pandemics, and recessions/shortages. Micronutrient deficiency, otherwise called hidden hunger, is one of the major malnutrition consequences worldwide, particularly in middle- or low- income countries. Unlike essential mineral or nutrient compounds, micronutrients could be less of a priority due to their small levels of requirement. However, insufficient micronutrients caused critical adverse health symptoms and are excessively vital for young children's development. Therefore, there have been numerous attempts to enhance minerals and nutrients in food crops, including biofortification, food fortification, and supplementation. Based on several interventions involving micronutrients, modern technology, such as nanotechnology, can be applied to enhance sustainability and to reduce the food system's environmental impact. Previous studies have addressed various strategies or interventions to mitigate major micronutrient deficiency including iron, iodine, zinc, and vitamin A. Comparably small amounts of studies have addressed vitamin B₁₂ deficiency and its fortification in food crops. Vitamin B₁₂ deficiency causes serious adverse health effects, including in the nervous or blood systems, and occurs along with other micronutrient deficiencies, such as folate, iron, and zinc, worldwide, particularly in middle- and low-income countries. Mitigation for B₁₂ deficiency has mainly focused on developing pharmacological and medical treatments such as vitamin B₁₂ serum or supplements. Further studies are required to undertake a sustainable approach to fortify vitamin B₁₂ in plant-based food sources for public health worldwide. This review paper highlights nanoparticle application as a promising technology for enhancing vitamin B₁₂ without conventional genetic modification requirements. The nanoparticle can efficiently deliver the mineral/nutrient using coating techniques to targeted sites into the plant. This is mainly because nanoparticles have better solubility and permeability due to their nano size with high surface exposure. Vitamin B₁₂-coated nanoparticles would

be absorbed, translocated, and accumulated by the plant and eventually enhance the bioavailability in food crops. Furthermore, by reducing adverse environmental effects, such as leaching issues that mainly occur with conventional fertilizer usage, it would be possible to develop more sustainable food fortification.

Keywords: biofortification, cobalamin (Cbl), food fortification, nanoparticle, vitamin B₁₂, vitamin B₁₂ deficiency

INTRODUCTION

A healthy diet contains adequate amounts of macronutrients, including proteins, fats, and carbohydrates, and essential micronutrients, such as vitamins and minerals. However, the current global food system does not meet the universal requirement of adequate nutrition in food production, which directly influences human health. According to the Food and Agriculture Organization, over 690 million people worldwide are suffering hunger and nearly 750 million people have been exposed to food insecurity in 2019 (FAO et al., 2020). Globally, current food consumption shifts towards animal-based food and highly processed-products, while the consumption of fresh unprocessed-food products such as fruits and vegetables has increased inadequately (Bodirsky et al., 2020). The trends of food consumption have different patterns depending on socioeconomic status and geographical, cultural, and demographical traits. According to Agriculture and Rural Development in European Commission, the consumptions of high-value food, including meats and dairy products, has significantly increased in emerging economies like China. Meanwhile, the trends show a shift from red meat to plant-based food consumption including fruits and vegetables in developed economies countries such as Europe and North America (Agriculture and Rural Development, 2019). In middle- and high-income countries, plant-based diets (e.g., veganism or vegetarianism) have been rising along with increased concerns regarding climate change, environment issues, animal welfare, and health (Jones, 2020). According to Google Trends analysis, the interests regarding vegan/vegetarian diets have been simultaneously increasing worldwide, particularly in upper middle-income countries, along with the rising interest in

veganism (19.54%) and vegetarianism (15.09%). The actual rates of plant-based diets has increased from 1.4 to 2% for vegan and 5% for vegetarian diets in the United States (Kamiński et al., 2020). These trends of plant-based diets also contribute to an incremental increase of plant-based sales in the U.S., up to 31.3% from 2017 to 2019 (Forgrieve, 2019). Thus, there are rising concerns regarding plant-based diets with low bioavailability of essential mineral/micronutrients such as iron, zinc, vitamin D, and fatty acids (Rizzo et al., 2016).

The nutrition transition shows a significant relation to malnutrition-related health effects (Bodirsky et al., 2020). Malnutrition is one of the major forms of food insecurity that refers to deficiencies and excesses or imbalances of nutrients or energy in human beings. Malnutrition symptoms include undernutrition, micronutrient deficiency, and obesity (WHO, 2020). Micronutrient deficiency is also known as hidden hunger due to the fact that its deficiency problem is generally asymptomatic but influences human health critically. In addition, micronutrient deficiency is highly related to food insecurity, which is a widespread issue in low-income and lower-middle-income countries such as Sub-Saharan Africa and South-Central and South-East Asia (Muthayya et al., 2013). The rationale underlying this is that these countries heavily rely on starchy staple foods such as cereals, tubers, and roots, and the overall availability of animal-based food is lower than that in high-income countries (FAO et al., 2020). The population groups at high risk for micronutrient deficiency include children under five and pregnant women, who have a considerable need for micronutrient supply for healthy development and growth. Child growth stunting is one of the major adverse symptoms due to malnutrition and it occurred in 21.3% or 144 million children in 2019 worldwide. Over 90% of stunted children lived in African or Asian countries accounting for 40–54% of all stunted children worldwide (FAO et al., 2020).

Among micronutrient deficiency, four major micronutrient deficiencies have been highlighted, namely iron, iodine, zinc, and vitamin A, which affect 2 billion people in the world (Ruel-Bergeron et al., 2015). Various strategies for these key micronutrient interventions have been developed and conducted worldwide through biofortification, food fortification, and supplementation (Khush et al., 2012). However, fortification of vitamin B₁₂ has been less highlighted although its deficiency impacts on human health critically. Vitamin B₁₂ deficiency is particularly common among vegetarians because animal-based food is the natural source for the B₁₂. It can cause neurological, hematological, and psychiatric symptoms and affect the formation of red blood cells and the normal functioning of the nervous system (Nakos, 2016). There has been extensive medicinal and pharmacological research

Abbreviations: AAS, atomic absorption spectrophotometer; Ado-Cbl, adenosyl cobalamin; BSA NPs, bovine serum albumin nanoparticles; Cbl, cobalamin; CN-Cbl, cyano cobalamin; DLS, dynamic light scattering; EDS, energy-dispersive X-ray spectrometry; ELS, electrophoretic light scattering; ENMs, engineered nanomaterials; FAO, Food and Agriculture Organisation; FE-SEM, Field Emission Scanning Electron Microscope; FTIR, Fourier transform infrared; H-Cbl, hydroxo cobalamin; HCl, hydrochloric acid; HCY, homocysteine pathway; His, histidine; HPLC-ICP-MS, high performance liquid chromatography – inductively coupled plasma mass spectrometry; ICP-OES, inductively coupled plasma optical emission spectrometry; ICP-MS, inductively coupled plasma mass spectrometry; IF, intrinsic factor; LA-ICP-MS, laser ablation inductively coupled plasma mass spectrometry; MALDI-MS, matrix-assisted laser desorption/ionization mass spectrometry; Me-Cbl, methyl cobalamin; NIH, National Institutes of Health; NPK, nitrogen-phosphorus-potassium; NTA, nanoparticle tracking analysis; NTDs, neural tube defects; RID, radioisotope dilution; RT-PCR, reverse transcription polymerase chain reaction; SAM, S-adenosylmethionine; SEM, scanning electron microscopy; SEM-EDX, scanning electron microscopy – energy dispersive X-ray analysis; SFY, stirred functional yogurt; TEM, transmission electron microscopy; TC, transcobalamin; UV-DRS, UV differential reflectance spectroscopy; WHO, World Health Organisation; XANES, X-ray absorption near edge structure; XRD, X-ray diffraction; XRF, X-ray fluorescence.

focusing on vitamin B₁₂ supplementation. However, little is known about the specific uptake mechanism of vitamin B₁₂ enrichment in living plants. Therefore, this review paper aims to review strategies for micronutrient fortification of food crops, particularly vitamin B₁₂, using nanotechnology. The objectives are to: (i) provide an overview of micronutrient interventions, (ii) determine a sustainable approach or technology for micronutrient fortification such as nanoparticle applications, and (iii) highlight the hydroponic system as a sustainable micronutrient fortification method for food crops.

CURRENT STRATEGIES OR INTERVENTIONS FOR MICRONUTRIENT DEFICIENCY

Biofortification for Micronutrient

Biofortification, as a complex process of developing new varieties of staple food crops, focuses on enhancing bioavailability of micronutrients through agronomic practices, breeding, or biotechnology techniques (Bouis and Saltzman, 2017; FAO, 2017). Biofortification includes three main approaches: agronomic practices, conventional breeding, and genetic modification technology (Thompson and Amoroso, 2010). In comparison to fertilizer development, biofortification can enhance up to the sufficient level of micronutrient/mineral in food crops without adverse environmental effects and it provides a cost-effective strategy for the long-term application with feasibility for underserved rural populations (Kaur et al., 2020). Biofortified crops have been widely adopted in low- and middle-income countries where staple crops, including cereals, tuberous roots, and legumes crops, are mainly consumed. For instance, vitamin A is fortified in various crops such as sweet potato, maize, cassava, banana, and plantain along with the enhancement of various agronomic traits including increased harvest yield and stress resistance. Iron has been fortified in legumes crops and pearl millet along with increased yield and disease resistance. In addition, zinc fortification has also been undertaken with wheat, rice, and maize (HarvestPlus and FAO, 2019).

Biofortification presents two main advantages: being cost-effective in the long-term and having the ability to spread to rural populations which are underserved (Bouis and Saltzman, 2017). Through various stages of biofortification, such as exploration development and delivery stages, the crop improvement activities are conducted considering applicability and cost-effectiveness worldwide, especially for low- and middle-income countries (Bouis and Saltzman, 2017; FAO, 2017). The main challenges for biofortification development consider three critical elements which are supply, policy, and demand. Agronomic traits should meet the micronutrient requirements and also biofortified crops should be supplied to both rural and urban consumers through supportive policies (Bouis and Saltzman, 2017). Supportive policy is necessary for further intervention program with sustainability. Consumer and growers' participation can also limit the biofortified food application which presents as an acceptance

of biofortified crop produce. Also, conventional plant breeding or genetic engineering technology requires long-term development and adequate expenses. Utilizing the agronomic method has also highlighted the biofortification method which mainly focuses on fertilizers with micronutrients using nanotechnology. Davies (2018) stated that micronutrient-based nanoparticle application would be a great alternative for biofortification without further breeding or genetic variation by enhancing micronutrient content and crop quality. The study found that micronutrient contents including iron, zinc, and calcium are enhanced in potato, tomato, and chili pepper grown in hydroponic systems utilizing metal oxide nanoparticle which synthesized with amino acid, which enhances the stability of nanoparticle (Davies, 2018).

Food Fortification for Micronutrient

Food fortification refers to an approach to enhance essential micronutrient content by adding vitamins or minerals into food crops so that it contributes to fortifying the contents of nutrient/mineral and providing health benefits to the public (FAO, 2017). Food fortification technology positively regulates micronutrient deficiency and prevents malnutrition problems, especially in staple crops. Compared to supplementations, which generally require a large dose of micronutrients in the form of capsules, tablets, and syrups (Smith et al., 2006; Elemike et al., 2019), food fortification requires relatively small amounts of micronutrients. In comparison to supplementation which can easily cause overdoses (Elemike et al., 2019), food fortification provides adequate amounts of micronutrients to consumers from a mass scale to specific targeted scale (Smith et al., 2006). There are several opportunities and challenges in food fortification. First, fortified food would supply a better-quality diet within micronutrients which are necessary for women and children by reducing the requirements of supplementation. Second, fortified food for staple crops could contain micronutrients in a natural level. In addition, the application of fortified food could improve the nutritional status in a cost-effective way in a large population or targeted population worldwide.

Although food fortification has numerous impacts on the public diet, there are several challenges to be considered. First of all, although fortified food applies to the public or targeted group, a specific fortified food product might be consumed only by some part of the targeted group. It is required that adequate programs or advertisements regarding fortified food are given for the targeted group (Smith et al., 2006). Second, due to accessibility, there is a higher chance to access fortified food in urban areas compared to rural areas (Elemike et al., 2019). Therefore, research should focus not only on the fortification process but also on the distribution of fortified products. Third, fortified food generally targets young children and women due to their extra requirement of micronutrients. However, the amount of uptake of fortified food is relatively small in infants or young children. Therefore, they are less likely to consume recommended intakes compared to their requirement of micronutrients. In addition, further knowledge is required about the effect of interaction among nutrients when micronutrients are added. To achieve sustainable food fortification and its implementation, it is necessary to consider transdisciplinary aspects based on understanding the

effects of agriculture, environment, socioeconomic, and politics (Smith et al., 2006). Also, further supportive national scale involvement is vital for sustainable food fortification with increased bioavailability in the long term.

VITAMIN B₁₂ (COBALAMIN)

Background

Physiochemical Traits of Vitamin B₁₂ and Its Role in Human Health

Vitamin B₁₂, known as cobalamin (Cbl), is a water-soluble vitamin and it is synthesized by bacteria such as heterotrophic bacteria and most of the aerobic photosynthetic cyanobacteria (Tandon et al., 2017). Vitamin B₁₂ mainly involves two enzymatic reactions, DNA synthesis and assimilation of inorganic carbon, which consequently influence on gene regulation, formation of blood cells, and metabolism of the nervous system (National Institute of Health, 2020). According to the National Institute of Health, the recommended daily amount of vitamin B₁₂ is 0.4 mcg to 0.5–0.6 mcg for infants, 1.2–2.4 mcg for children to adolescence, and 2.4 mcg for adults (2.6 and 2.8 mcg for pregnancy and lactation respectively) (Pawlak et al., 2013; Nakos, 2016).

Vitamin B₁₂ is originated by bacteria mainly found in the gastrointestinal tract of animals and it should be synthesized via the fermentation process of lactic acid (Jedut et al., 2021). There are two different vitamin B₁₂-biosynthetic routes for prokaryotes in nature based on their oxygen-dependency: aerobic and anaerobic pathways. For instance, *Paracoccus denitrificans* takes an aerobic (oxygen-dependent) pathway, whilst *Bacillus megaterium*, *P. shermanii*, and *Salmonella typhimurium* take anaerobic (oxygen-independent) pathways (Martens et al., 2002). Enteric bacteria, as the main source of vitamin B₁₂, is responsible for most vitamin B₁₂ biosynthesis processes found in animals or humans. However, eukaryotes is merely linked to the metabolic process of vitamin B₁₂ so that nematodes or animals acquire dietary vitamin B₁₂ from vitamin B₁₂-producing bacteria (Lawrence et al., 2018). Vitamin B₁₂ is a nutritional requirement for animals, whereas plants neither require nor synthesize it. Animals contain a vitamin B₁₂-dependent enzyme called methionine synthase (METH), such as methyl malonyl-CoA mutase or methionine synthase, whilst plants contain vitamin B₁₂-independent methionine synthase (METE) enzyme (Tandon et al., 2017). Only traceable amounts of vitamin B₁₂ are found in plants due to a nitrogen-fixing actinobacteria, *Frankia annii*, which associates symbiotically with actinorhizal plants and contributes vitamin B₁₂ (M. Nakos et al., 2017). Thus it is mainly found in animal-derived sources, including meat (e.g., ruminant or omnivorous animals), dairy, fish, and shellfish (Watanabe and Bito, 2018).

Vitamin B₁₂ (Cobalamin) consists of central corrin rings containing four pyrrole rings around a central cobalt atom, a lower ligand (α -ligand) donated by 5,6-dimethylbenzimidazole (DMBI), and an upper ligand (β -ligand) synthesized from an adenosyl or methyl group (Nakos, 2016; Rizzo et al., 2016). Dependent on the different ligands on the upper surface of

cobalt atoms, vitamin B₁₂ is classified as four different chemical forms: (i) hydroxocobalamin (H-Cbl), (ii) Methylcobalamin (Me-Cbl), (iii) Cyanocobalamin (CN-Cbl), and (iv) adenosylcobalamin (Ado-Cbl). As coenzymes in the cell, Me-Cbl and Ado-Cbl are the active forms of vitamin B₁₂. CN-Cbl and H-Cbl are provitamin forms requiring Me-Cbl or Ado-Cbl to be utilized by the cells (Rizzo et al., 2016). By participating in the metabolic homocysteine pathway (HCY), Me-Cbl acts a cofactor of methionine synthases. HCY pathway impacts on DNA synthesis and is involved in regenerating methyl donor S-adenosylmethionine (SAM). Methionine is required not only for SAM formation but also for metabolisms of DNA, RNA, proteins, and lipids (NIH, 2021). Ado-Cbl, as a cofactor of methylmalonyl-CoA mutase, is involved in metabolism of amino acid and fatty acid. Among the different chemical forms of vitamin B₁₂, cyanocobalamin (CN-Cbl), which is chemically manufactured, is the most stable form and is obtained by reacting natural cobalamin with cyanide. Therefore, CN-Cbl is the most widely used in food fortification, in nutrient formulas, and for pharmaceutical purposes (Nakos, 2016; Rizzo et al., 2016).

Vitamin B₁₂ Absorption, Digestion, and Circulation in Humans

Vitamin B₁₂ is predominately contained in animal-based food. Following food-cobalamin intake, vitamin B₁₂ attaches to dietary animal protein produced from the salivary gland. In the stomach, dietary protein is released by the acidic environment of the stomach via proteolysis. Vitamin B₁₂ binds to haptocorrin (R-protein) which is secreted by the salivary glands protecting vitamin B₁₂ from acid degradation (Green et al., 2017). Pepsin and hydrochloric acid (HCl) are released from gastric secretion. In stomach, Intrinsic factor (IF) is also secreted, and it binds less strongly when gastric R-protein presents. In duodenum, haptocorrin degradation occurs and the pH is changed in favor of vitamin B₁₂ binding to IF. Pancreatic enzymes degrade dietary vitamin B₁₂ and haptocorrin complex to release free vitamin B₁₂ and free vitamin B₁₂ binds to IF and vitamin B₁₂-IF complex transport to ileum (Andrès et al., 2004). In the ileum, vitamin B₁₂-IF complex binds/enters to the cubam receptor which contains cubilin. Cubam receptor mediates endocytosis of vitamin B₁₂-IF complex (Stabler, 2013; Green et al., 2017; Surendran et al., 2018). After the IF and vitamin B₁₂ are detached, vitamin B₁₂ binds to transport proteins transcobalamin I, II, and III. Transcobalamin II (TCII) involves transportation of vitamin B₁₂ to all cells in the human body. Vitamin B₁₂ is consequently transported via the portal system. TCII-Cbl complex is absorbed by endocytosis and free vitamin B₁₂ is enzymatically converted into its two coenzymatic forms: methyl-cobalamin (Me-Cbl) and adenosylcobalamin (Ado-Cbl). Most vitamin B₁₂ is stored in the liver and some vitamin B₁₂ is secreted in bile which undergoes enterohepatic circulation (Andrès et al., 2004; Green, 2017; Green et al., 2017).

Previous studies pointed out several endogenous and exogenous factors that impact on the absorption of vitamin B₁₂ in gastrointestinal metabolism and enterohepatic circulation (Andrès et al., 2004). Firstly, achlorhydria in stomach may be associated with cobalamin malabsorption, which is a lack of

hydrochloric acid in the gastric secretion in the stomach. Vitamin B₁₂ malabsorption can be induced by insufficient IF along with chronic gastritis which may lead to megaloblastic anemia and neurological disorders (Andrès et al., 2004). Secondly, insufficient exocrine pancreatic can also influence vitamin B₁₂ malabsorption owing to low pH in the small intestine and impaired degradation of haptocorrin due to pancreatic enzymes (Green et al., 2017). Thirdly, bacterial overgrowth, such as *Pseudomonas* spp. and *Klebsiella* spp., can influence vitamin B₁₂ absorption in small intestine. Bacterial overgrowth may occur by gastrectomy, ileocolic intestinal resection, and secretion of impaired gastric acid (Andrès et al., 2004; Green et al., 2017). Lastly, genetic disorders associated with vitamin B₁₂ deficiency is involved in plasma transportation and conversion to coenzyme forms (Andrès et al., 2004; **Figure 1**).

Vitamin B₁₂ Deficiency

Vitamin B₁₂ deficiency is characterized by low levels of circulating serum vitamin B₁₂ and holo-transcobalamin (holoTC) along with elevated levels of total homocysteine in plasma and methylmalonic acid in serum or urine (Brito et al., 2018). Homocysteine, as an amino acid, is generated by the demethylation of methionine and is accumulated in blood when folate, vitamin B₁₂, and vitamin B₆ are insufficient (Tucker et al., 2004). Vitamin B₁₂ deficiency is prevalent not only in lower-middle-income countries, but also in upper-middle-income countries. Geographically, the deficiency generally occurs in middle- or low-income countries due to the staple crop and plant-based diets and insufficient consumption of animal-based food (Titcomb and Tanumihardjo, 2019). Deficiency of vitamin B₁₂/iron causes pernicious anemia, mainly threatening pregnant women in South Asia and Sub-Sahara countries reaching 60 and 42% of children under five globally (Ritchie and Roser, 2017; **Figures 2, 3**). Vitamin B₁₂ deficiency is also prevalent in elderly, affecting up to 20% of people over 60 and approximately 6% of adults (younger than 60 years) in the United States and United Kingdom (NIH, 2021).

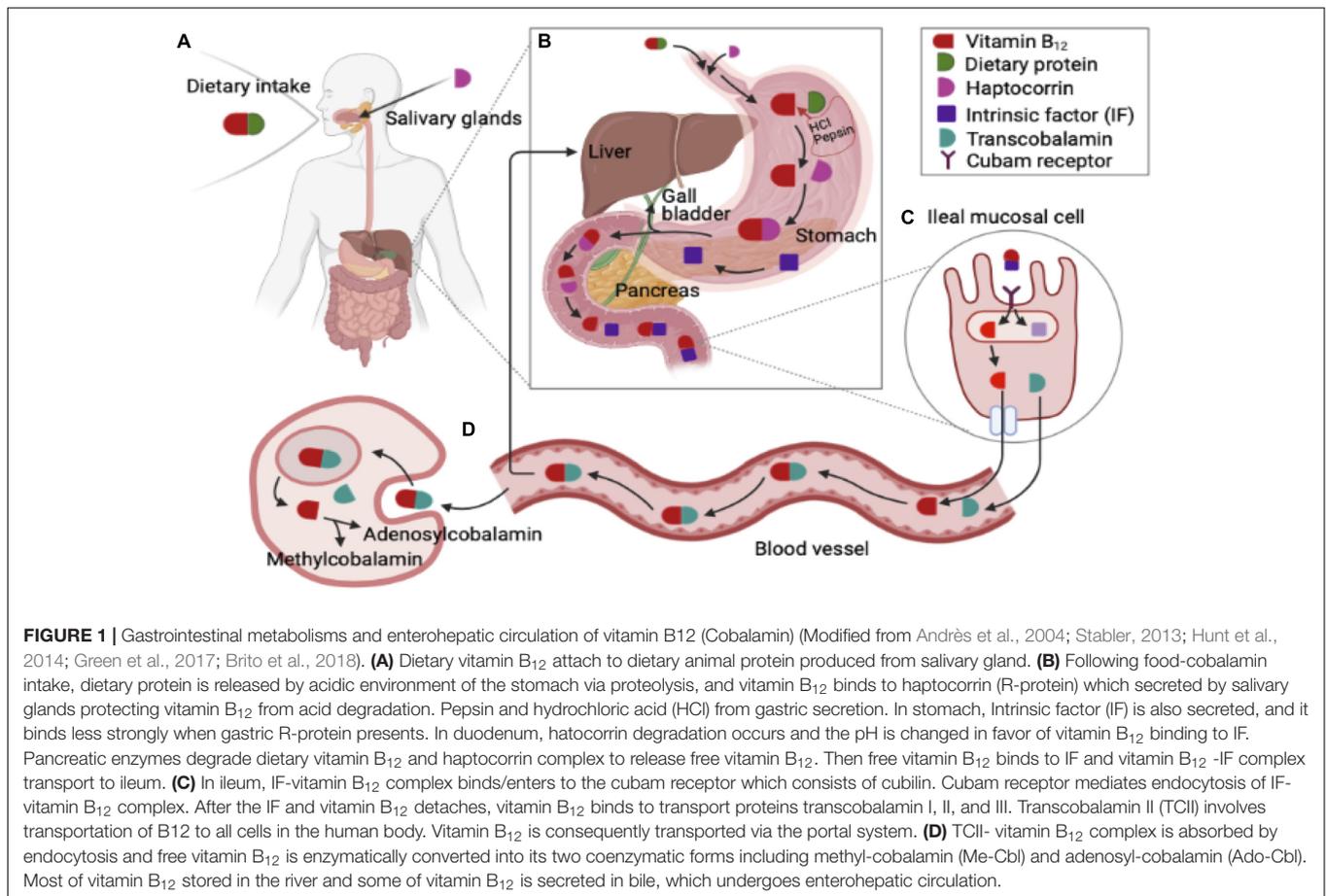
Populations at high risk for vitamin B₁₂ deficiency include the elderly, children, pregnant women of reproductive age, patients (e.g., autoimmune diseases including pernicious anemia and atrophic gastritis), and those with malabsorption of food-cobalamin including those on strict vegetarian or vegan diets (Stabler, 2020). Within the elderly population, vitamin B₁₂ malabsorption is mainly due to age-related gastric atrophy, which causes a reduction in acid and intrinsic factor (IF) (Rusher and Pawlak, 2013; **Figure 1**). Also, the populations with atrophic gastritis with low levels of stomach acid secretion generally have vitamin B₁₂ malabsorption with certain gastric dysfunctions from food-bound vitamin B₁₂ sources (Watanabe, 2007). Vitamin B₁₂ deficiency is generally measured by serum methylmalonic acid (MMA) or the level of total plasma homocysteine which is normally lower than 200 or 250 pg/mL (NIH, 2021). Maternal vitamin B₁₂ deficiency during the period and/or lactation increases the possibility of birth defects or growth retardation in infants (Brito et al., 2018). In addition, low levels of vitamin B₁₂ in breastfeeding and insufficient maternal intake of animal-based food causes brain development defects and overall developmental regressions, particularly 4–6-month-old infants (Stabler, 2013).

Pathophysiology of vitamin B₁₂ deficiency includes subclinical symptoms because vitamin B₁₂ is stored at 1–5 mg in the human body which makes it difficult to diagnose deficiency (NIH, 2021). Vitamin B₁₂ deficiency is often accompanied with other micronutrient deficiencies such as folate (as known as B₉), iron, zinc, and protein deficiencies (Stabler, 2020). In particular, various studies have highlighted the correlation between vitamin B₁₂ and folate (Allen et al., 2010; Oakley and Tulchinsky, 2010). Folic acid fortification neither prevents nor treats vitamin B₁₂ deficiency, but it also does not impact adversely on vitamin B₁₂ deficiency (Oakley and Tulchinsky, 2010). Severe vitamin B₁₂ deficiency induces megaloblastic anemia (abnormal nucleated red blood cells) and abnormal neurologic diseases (NIH, 2021). This deficiency also causes hematological and psychiatric adverse symptoms by affecting the formation of red blood cells and the normal functions of the nervous system (Nakos, 2016; Rizzo et al., 2016; Stabler, 2020). Previous research has highlighted several endogenous and exogenous effects on the absorption of vitamin B₁₂: (i) food-cobalamin malabsorption such as inadequate vitamin B₁₂ levels due to inappropriate dietary intake or low bioavailability; (ii) malabsorption due to chronic disorder or autoimmune diseases including genetic disorders, pernicious anemia, atrophic gastric, malabsorption due to the overgrowth of *Helicobacter pylori*, or chronic alcoholism; and (iii) competition for vitamin B₁₂ as a result of nitrous oxide exposure (Rusher and Pawlak, 2013; Stabler, 2013; Brito et al., 2018). Currently, patients suffering autoimmune diseases with vitamin B₁₂ deficiency are recommended to be injected with 1000 µg of vitamin B₁₂ several times weekly. Also, high-dose oral treatment is also recommended for patients providing effective treatments. Vitamin B₁₂ replacement therapy via intramuscular administration was also conducted for treating vitamin B₁₂ deficiency (Rusher and Pawlak, 2013; Stabler, 2013). However, it has been reported that deficiency symptoms are alleviated but not fully improved. Extensive medical and pharmacological research has focused on vitamin B₁₂ supplementation. A more comprehensive study would be conducted on vitamin B₁₂ enrichment in living plants.

Vitamin B₁₂ Sources: Supplementations and Natural Food Sources

It is common to treat vitamin B₁₂ deficiency with high-dose injection, oral treatments, and nasal gel spray with cyanocobalamin. Although the supplementations have a high level of vitamin B₁₂ doses, it is generally believed to be safe since only limited amounts of vitamin B₁₂ are stored in the human body (1.4–5.1 µg) (Doets et al., 2013; NIH, 2021). For instance, vitamin B₁₂ is absorbed 50% from 1 µg oral dose and 20% from 5 µg dose and the absorption is decreased by saturation from vitamin B₁₂-IF complex in the ileum from 1 to 2% of an oral dose (Brito et al., 2018). Daily losses of vitamin B₁₂ presents 3.8–20.7 µg in healthy populations (over 25 years old), which is 1.4–8.6 times higher than the required amount of vitamin B₁₂ preventing deficiency (Doets et al., 2013).

Vitamin B₁₂ is generally concentrated in animal-based sources such as meat, dairy products, eggs, fish, and shellfish (Bito et al.,



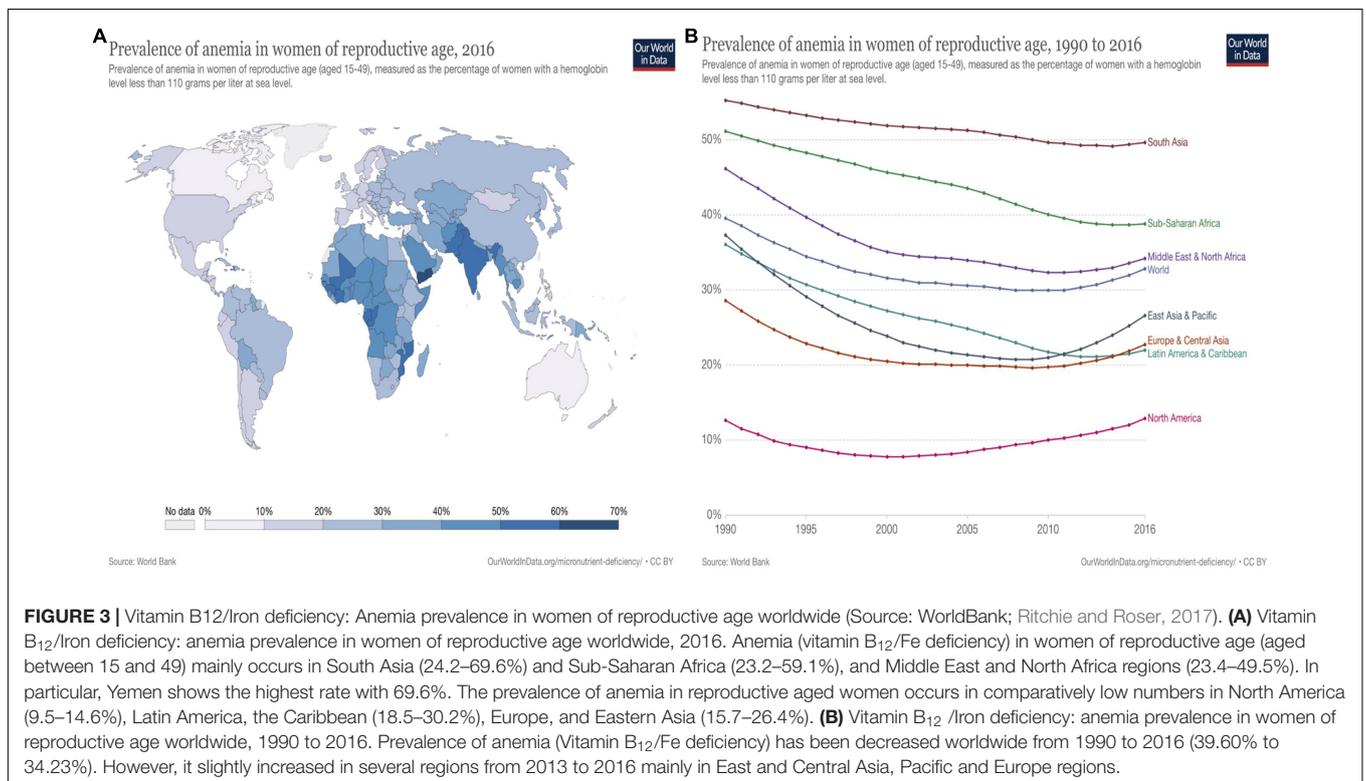
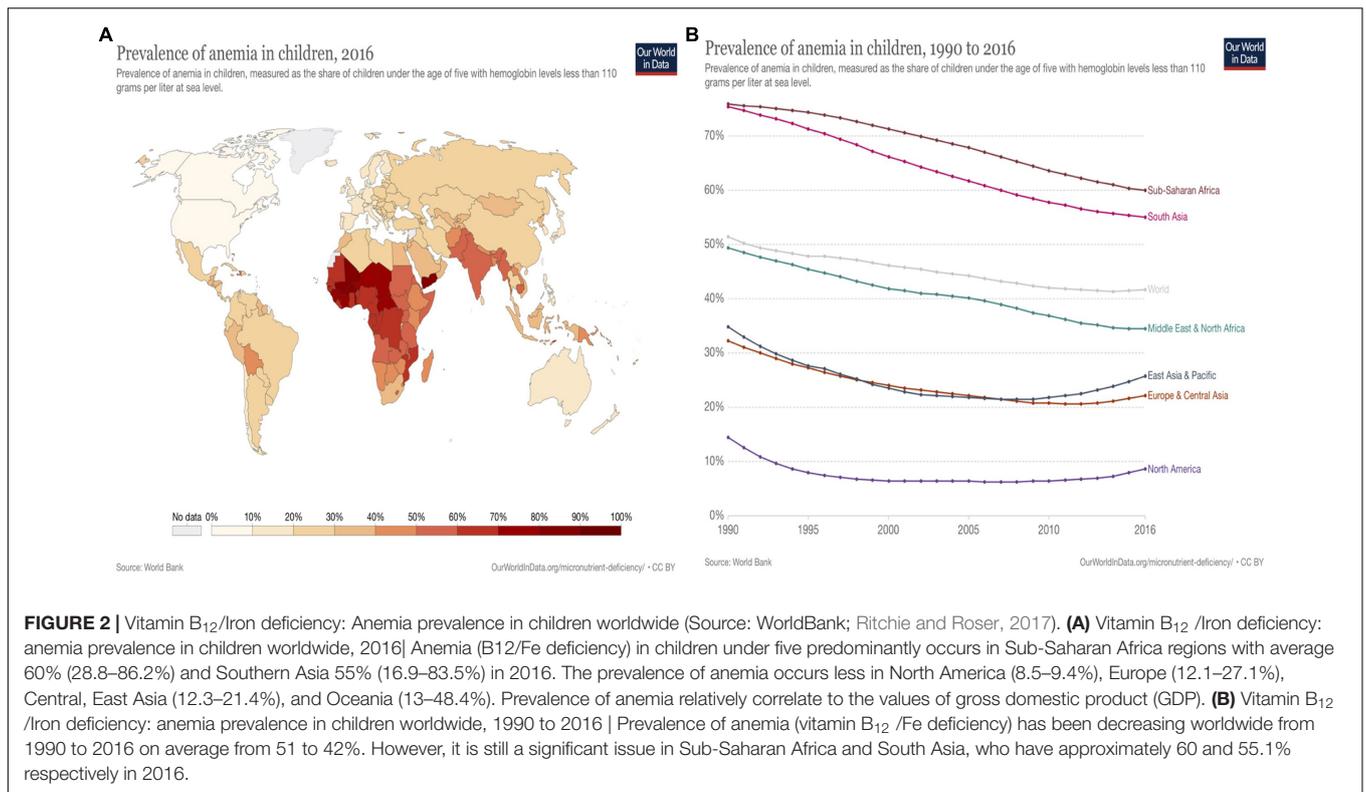
2013; Rizzo et al., 2016). According to Watanabe and Bito (2018), the meat and livers of ruminant animals contain higher amounts of vitamin B₁₂ (cooked beef liver contains 83 µg/100 g) than that found in omnivorous animals (Cooked chicken meat contains 0.4–0.6 µg/100 g). Traceable amounts of vitamin B₁₂ are found in dairy products, such as milk (0.3–0.4 µg/100 g), egg (0.9–1.4 µg/100 g), shellfish (104 µg/100 g), and fish including salmon, trout, and tuna (3.0–8.9 µg/100 g) (Watanabe and Bito, 2018). Compared to animal-based food, plant-based food contains very low or negligible amounts of vitamin B₁₂ (Nakos, 2016). For instance, only negligible amounts of vitamin B₁₂ was found (<0.1 µg/100 g) in vegetables, including broccoli, asparagus, and mung bean sprouts (Watanabe and Bito, 2018).

Several studies have focused on plants, fungi, and algae with traceable amounts of vitamin B₁₂ within these symbioses including edible mushrooms, edible algae, fermented soybeans (0.1–1.5 µg/100 g) and vegetables, and processed food such as cereal, bread, and beverages (Bito et al., 2016; Watanabe and Bito, 2018). The contents of vitamin B₁₂ in edible plants is generally very low with different degrees of stability and bioavailability which contribute complex analysis of vitamin B₁₂ (Nakos et al., 2017). Previous studies have emphasized that the biological activity of vitamin B₁₂ is uncertain in most cases due to the limited availability of natural sources of vitamin B₁₂ (Nakos et al., 2017). Therefore, further studies are required to focus on the

fortification of vitamin B₁₂ in food crops and evaluate their applicability in the dietary system.

Status of Vitamin B₁₂ Supplementation/Fortification

Among micronutrient deficiencies, the amelioration of vitamin B₁₂ deficiency is significantly less emphasized. Extensive research regarding vitamin B₁₂ fortification has focused on supplementation for medical or pharmacological purposes. Supplements of vitamin B₁₂ from multivitamin/minerals are generally 500–1,000 mcg of vitamin B₁₂ with 2 and 1.3% of absorption respectively (NIH, 2021). Due to the IF-mediated gastrointestinal absorption and circulation system, vitamin B₁₂ bioavailability decreased ranging from 1.5 to 2.0 µg per meal under physiologic conditions or food processing (Watanabe, 2007). In healthy populations, vitamin B₁₂ bioavailability presents 42% from fish or meat (beef, lamb, and chicken), ranging from 56 to 89%, and although traceable amounts of vitamin B₁₂ contents are found in the yolk, eggs have poor bioavailability compared to other vitamin B₁₂-contained food sources. Vitamin B₁₂ bioavailability can be significantly degraded by dysfunctions such as atrophic gastritis with low levels of stomach acid secretion. According to Dietary Reference Intake, it is assumed that a healthy population with a normal



gastrointestinal function can absorb 40–50% of vitamin B₁₂ from animal-based food sources (Watanabe, 2007). It is difficult to evaluate vitamin B₁₂ bioavailability by quantifying active vitamin

B₁₂ or inactive corrinoids in certain foods, such as vitamin B₁₂ fortified plant-based food. Therefore, for vitamin B₁₂ fortification strategies, it is necessary to enhance the contents of vitamin B₁₂

in food sources along with the high bioavailability which is actual amounts of available uptake (Watanabe, 2007).

Little is known about vitamin B₁₂ biofortification based on crops. However, a relatively limited number of studies have been conducted to investigate vitamin B₁₂ fortification and understand their mechanisms of uptake/distribution in living plants. According to previous studies, several plants, fungi, and algae (e.g., Japanese radish sprouts, mushrooms, dry seaweed, and garden cress) absorb and translocate vitamin B₁₂ if they are grown in nutrient-sufficient conditions with organic fertilizer or vitamin B₁₂-enriched growing media (Sato et al., 2004; Bito et al., 2013; Lawrence et al., 2018). One of the effective strategies to enhance the contents of vitamin B₁₂ is flour fortification for a national scale with uniform dose. There are various processed food products that have been fortified with vitamin B₁₂ such as cereal grain products (Tucker et al., 2004; Melo et al., 2020), dried soup powder, and powdered milk drink (Sanchez et al., 2013). Vitamin B₁₂ can also be supplemented with fresh-cut fruits and vegetables for alleviating its deficiency in the population groups who are at high risk. A combination of vitamin B₁₂ and chitosan is applied in fresh-cut salad mixes (melon, pineapple, and carrot) which are for 'ready-to-blend' beverages. According to Artés-Hernández et al. (2017), the study shows the fortified beverage contains up to 8.6 µg/1 kg of vitamin B₁₂ along with an enhanced shelf-life.

Unlike widespread folate fortified wheat flour, there is inadequate case for vitamin B₁₂ fortification with food crops in worldwide. Folic acid fortification has been successfully conducted, but it also increased the concerns about the possibility of neurological issues and deterioration of cognitive ability occurring with high folic acid supply and low level of vitamin B₁₂ (Garrod et al., 2019). The study conducted by Garrod et al. (2019) aimed to quantify the bioavailability of fortified vitamin B₁₂ in bread in five healthy elderly people aged over 60. The study shows that vitamin-B₁₂-fortified flour retains its bioavailability approximately 50% after the processes of fermentation and baking containing 2 µg/100 g. The study shows the healthy elderly can absorb sufficient vitamin B₁₂ from the fortified bread addressing that the vitamin B₁₂ fortified wheat (flour) can be a promising animal-based substitute with high-purity crystalline ¹⁴C-vitamin B₁₂ (Garrod et al., 2019). Further study would be required to investigate the absorption of fortified flour for different targeted subjects who have high risk of vitamin B₁₂ deficiency such as children or pregnant women or larger populations of the elderly. According to Tucker et al. (2004), vitamin B complex fortified breakfast cereal contribute enhancement of their contents including vitamin B₆, B₉ (folate), and B₁₂ (cobalamin). Fortified cereal significantly improves vitamin B₁₂ concentrations and reduces the prevalence of high levels of plasma homocysteine from 13 to 3%. This study highlights that fortified cereal would be great vitamin B₁₂ sources for general populations who frequently have poor vitamin status (Tucker et al., 2004). Tucker et al. (2004) pointed out that free vitamin B₁₂-fortified breakfast cereals presents better absorption compared to vitamin B₁₂-bound to proteins in food which may be due to the IF-mediated gastrointestinal metabolisms effects on vitamin B₁₂ absorption. Vitamin B₁₂ fortification is also targeted to adults aged over 50 years by

using the microencapsulation approach (Melo et al., 2020). The study found that the application of the encapsulated vitamin B₁₂ powder in yogurt (50 µg of vitamin B₁₂ was added into 175 g of yogurt) successfully fortified vitamin B₁₂. The study compared Me-Cbl and CN-Cbl and the outcomes of the study show that CN-Cbl presents better stability throughout shelf-life. This study undertakes encapsulation technique for vitamin B₁₂ fortification using spray-drying method to coat vitamin B₁₂ with a maize starch-derived polymeric materials (Melo et al., 2020).

There are several in situ fortification methods for vitamin B₁₂ in fermented plant-based food. In situ fortification with bacteria (*Propionibacterium freudenreichii* DSM 20271 and *Levilactobacillus brevis*) successfully enhances vitamin B₁₂ contents in fermented cereal, pseudo-cereal (such as rice bran and buckwheat bran), and legume plants (Xie et al., 2021). Fermented legume materials contain 300–400 ng/g (dry weight) of vitamin B₁₂ measured by ultra-high performance liquid chromatography and the results suggest that fermented crop materials can be a great potential alternative for plant-based food-cobalamin. This study also found that optimal pH condition for *P. freudenreichii* can increase the vitamin B₁₂ contents in fermented grain materials. Further research would be needed to examine the contents of other micronutrients and minerals in the fermented crop materials using in situ fortification which is necessary to be considered for vitamin B₁₂ amelioration along with other micronutrient deficiencies (Xie et al., 2021). Vitamin B₁₂ also successfully fortified up to 0.97 µg/100 g in tempeh by *in situ* approach using *Propionibacterium freudenreichii* (Log 7 CFU/g) and *Rhizopus oryzae* spores (Log 4 CFU/g) (Wolkers – Rooijackers et al., 2018). This research determines the effect of in situ strategy on tempeh quality by analyzing the consumer acceptance traits including microbial composition, firmness, and volatile organic compounds to measure aroma quality. The result of the study shows that the fortification using both food-grade bacterium does not have any negative impact on the quality traits. However, it is also necessary for this study to analyze other micronutrient/mineral components that are highly related to vitamin B₁₂ deficiency and fortification.

Vitamin B₁₂ supplement programs and fortification strategies target reduction of vitamin B₁₂ deficiency and further factors should be considered for successful vitamin B₁₂ fortification: (i) actual concentration and bioavailability of vitamin B₁₂ in fortified or supplemented sources; (ii) demographic, geographic, and socioeconomic traits of targeted groups, and (iii) effect of frequent- or over-dose of vitamin B₁₂ (Carmel, 2008).

NANOPARTICLE TECHNOLOGY: A SUSTAINABLE APPROACH FOR MICRONUTRIENT/MINERAL ENHANCEMENT

Background on Nanoparticle Technology

Engineered nanomaterials (ENMs), called nanoparticles, feature at least one dimension ranging from 1 to 100 nm and consist of organic, inorganic, or hybrid materials (Shang et al., 2019).

Owing to their small particle size, nanoparticles have a large surface area and exhibit high solubility and mobility that is widely exploited for smart delivery for pharmaceutical, medical, and agricultural purposes (Shang et al., 2019). The size of ENMs corresponds to biological barriers such as a plant cell wall or membrane after root or foliar applications and to enable new smart delivery of nutrients or pesticides (Nair et al., 2010; Lowry et al., 2019). The characteristics of nanoparticles, such as structure or surface chemistry traits, should be selected properly for different functions or nanotechnological strategies (Lowry et al., 2019). Through interactions between nanoparticles and plants, including nutrient interactions, it is possible to enhance the nutritional quality of food crops by accumulating specific macro- and micronutrients.

The agronomical biofortification of food crops with micronutrients/minerals is a promising technique with a fast and easy way to mitigate inadequate essential nutrients/minerals in plants (Elemike et al., 2019). Nano-agrochemicals can be classified by their types or delivered nutrients, such as macronutrient and micronutrient fertilizers and macronutrient carriers (Singh Sekhon, 2014). Macronutrients, in particular NPK fertilizers, can be applied with nanoparticles in nanocapsule form, or particles can be coated with nutrients/minerals (such as in urea-coated zeolite chips) with slow release application to enhance the uptake and efficiency of fertilizer (Singh Sekhon, 2014). Nano-NPK fertilizer promotes crop harvest yield and growth quality, including the size or weight of edible vegetative parts and physiochemical compounds in several crops, such as wheat (Abdel-Aziz et al., 2016; Al-juthery and Al-Shami, 2019), potato (Rop et al., 2019), maize, kale, and capsicum crops (Rop et al., 2019). Among techniques involving nano-NPK fertilizers, such as the slow-release method or coating with nutrient ions, nano-NPK fertilizers shorten the life cycles of crops compared to conventional fertilizer application, which is highly relevant to increasing crop yield (Liu and Lal, 2015). Micronutrient-loaded nanoparticles may provide more favorable uptake or distribution of micronutrients in plants by providing slow release of the nutrient by plants or soils and reducing environmental pollution (e.g., leaching) or agroecology degradation (Elemike et al., 2019). Compared to conventional fertilizers, nanofertilizer can triple the nutrient effectiveness, reduce the requirement or usage of fertilizers applications, develop crops with stress resistance, and cause less adverse environmental impacts (e.g., leaching) (Elemike et al., 2019).

Micronutrient fertilizers are being developed with various micronutrient ions, such as Fe (Rameshraddy et al., 2017); Mn, Zn (Liu and Lal, 2015); and Cu (Trujillo-Reyes et al., 2014). Depending on the application method, nanoparticles synthesized with micronutrients differentially translocate and accumulate in leaves, shoots, and grains along with various effects on growth performance. For instance, hydroponic cultivation enables more efficient and effective root application of nanofertilizer (Jeyasubramanian et al., 2016). Nanofertilizers are also widely applied for foliar applications using spraying methods. Furthermore, nanofertilizer treatments can be conducted together, such as the combination of seed priming and foliar application of zinc oxide nanoparticles (ZnO NPs), which showed

enhancement of seedling growth and increased biomass contents of chlorophyll and yield in rice (Rameshraddy et al., 2017).

Fortification Strategies of Vitamin B₁₂ and Its Deficiency Relevant Micronutrient

Vitamin B₁₂ Fortification Using Nanoparticle Technology

Vitamin B₁₂ deficiency may be accompanied by high folate status presenting a negative association with adverse health consequences such as cognitive impairment and delaying nerve conductivity (Brito et al., 2018). According to Prueksaritanond et al. (2013), intramuscular inject of vitamin B₁₂ along with the supplemented iron and folate significantly improve vitamin B₁₂ deficiency. Thus, further strategies for vitamin B₁₂ fortification should ameliorate folate, iron, and zinc deficiencies. Currently, nanocarriers have been applied for the pharmaceutical application of vitamin B₁₂ for targeted smart delivery. Nanocarriers consist of organic/inorganic nanoparticles and are utilized for smart delivery as commonly used in the pharmaceutical industry. Nanoparticle applications offer innovative solutions to improve the sensitivity of measurement by enhancing the electromagnetic signal in metal nanoparticle due to their nanosize with surface plasmon resonance effect (Fidaleo et al., 2021). For vitamin B₁₂ delivery, nanoparticles should be absorbed in the small intestinal tracts. There are several strategies for improving bioavailability using vitamin B₁₂ nanocarriers with thiolate polyacrylic acid particles and nanoengineered polymeric capsules. However, there are increasing concerns regarding potential toxicity of nanoparticles. Further studies should address not only the mechanisms of uptake and transport and metabolisms *in vivo*, but also the safety applications dealing with the potential toxicity of nanoparticles.

Unlike for medical/pharmacological purposes, comparatively limited studies have been conducted for food/agriculture strategies. Seed priming techniques (Sato et al., 2004; Keshavarz and Moghadam, 2017) and a hydroponic system with vitamin B₁₂-enriched solution (Bito et al., 2013) have been utilized for vitamin B₁₂ fortification in food crops (Table 1). Plants treated with high concentrations of vitamin B₁₂ exhibit more favorable growth performance and an increased content of vitamin B₁₂ in the crops (Keshavarz and Moghadam, 2017). The study found that vitamin B₁₂ increases the resistance capacity against abiotic stress and reduces oxidative stress by providing an effective antioxidant and regulating osmotic balance. Common bean seeds soaked with 11 and 22 μm of vitamin B₁₂ concentrations show enhanced chlorophyll contents, catalase, and peroxidase activity in the leaves in the condition of salt stress compared with control treatment. Also, the application of seed priming with 22 μm of vitamin B₁₂ under saline conditions increase the level of proteins in the bean plants, whilst there was no effect of protein improvement under non-saline conditions. The outcome of this study highlights that the application of vitamin B₁₂ enhances not only the salinity tolerance, but also effective photosynthetic biosynthesis by alleviating the adverse effect on photosynthesis pigments in salinity stress (Keshavarz and Moghadam, 2017).

Seed priming with vitamin B₁₂-enriched solution also increases the vitamin B₁₂ contents in kaiware daikon sprout up to 1.5 μm/g in any concentration of vitamin B₁₂ solutions ranging from 0 to 200 μg/ml (Sato et al., 2004). This study also addresses the possibility of reduction of vitamin B₁₂ by cooking processes, such as boiling, and also highlights that prolonged boiling (3–10 min) will decrease the vitamin B₁₂ contents in kaiware daikon. Based on this previous finding, further studies can be conducted to identify the effect of vitamin B₁₂ enriched coated with micronutrient/mineral in food crops and how much vitamin B₁₂ contents will be retained after cooking processes.

As demonstrated above, only a few previous studies focused on vitamin B₁₂ fortification in food crops (Sato et al., 2004; Bito et al., 2013; Keshavarz and Moghadam, 2017; **Table 1**). Various studies only focused on the quantification and determination of vitamin B₁₂ in food crops such as *Hippophae rhamnoides* berries (Nakos et al., 2017), edible algae (Kumudha, 2015), mushrooms (Watanabe et al., 2012, 2014; Bito et al., 2014, 2016), and fermented plant-based products (Watanabe et al., 2013). Titcomb and Tanumihardjo (2019) highlighted that high intake of vitamin B₁₂ did not show adverse effects on human bodies. However, there are limited studies on the effect of fortified vitamin B₁₂ in the food crops and its specific health effects on human health. Further research would be required to identify sustainable fortification methods for vitamin B₁₂ and its stability and effects on human health when it is digested or accumulated in the long term. Furthermore, it still remains challenging to quantify and determine exact vitamin B₁₂ contents in food crops because vitamin B₁₂ mainly exists as bound form in food crops with different degrees of stability (Nakos et al., 2017). Nakos et al. (2017) pointed out that immunoaffinity chromatography and HPLC analysis can provide quantitative chromatographic isolation of vitamin B₁₂ in food crops. Further studies should distinguish between active or inactive analogs of vitamin B₁₂ forms in food crops (Nakos et al., 2017).

Folate Fortification Using Nanoparticle Technology

In order to enhance the contents of vitamin B₁₂ in food crops, the interrelated deficiencies should also be alleviated. Vitamin B₉, as known as folate, is involved in the synthetic mechanisms and methylations of nucleotides by intervening in cell multiplications and tissue growth. Vitamin B₁₂ and folate presents an intimate connection via their cooperation in one-carbon metabolism and the hematological complications that are indistinguishable consequences/symptoms of deficiencies caused by either vitamin B₁₂ or folate (Green, 2017). In vitamin B₁₂ deficiency status, normal folate cycling disrupts the regeneration of methylenetetrahydrofolate, and it is required to sustain the synthesis of thymidine for replication of DNA. Since vitamin B₁₂ is required for its conversion to tetrahydrofolate within the reaction of methionine synthase, folate becomes trapped as methyl-folate which ultimately causes functional folate deficiency (Green, 2017). Vitamin B₁₂ deficiency reduces the activity of methionine synthase and subsequently reduces folate cycle intermediates, causing thymidine synthesis (Titcomb and Tanumihardjo, 2019).

TABLE 1 | Vitamin B₁₂ fortification in food crop.

Plant	Method	Analysis tools	Media	Germ.	Seedling phenotype	Seedling growth	Yield	Nutrient	Toxicity	References
Cereal, pseudo-cereal and legume plants	In situ fortification with <i>Protonibacterium freudenreichii</i> and <i>Levilactobacillus brevis</i>	ICP-MS and Determination of pH, total titratable acids and contents of acids	Fermentation inoculation	N/A	N/A	N/A	N/A	↑	N/A	Xie et al., 2021
Common bean	Seed priming (0, 11, and 22 μM) and salinity stress application	Enzyme activity assay, chlorophyll and carotenoid assay, malondialdehyde assay and proline assay	In vitro	N/A	↑	↑	N/A	↑ (phenolic and protein)	N/A	Keshavarz and Moghadam, 2017
Kaiware daikon (Japanese radish sprout)	Soaking vitamin B ₁₂ solutions (0–200 μg/ml) and Heat treatment	Chemiluminescence assay method, supernatant fluid assay (E. coli 215 and L. delbrueckii lactis ATCC 7839)	In vitro	N/A	N/A	N/A	N/A	↑	N/A	Sato et al., 2004
Lettuce	CN-Cbl-loaded solution	Spectrophotometer, HPLC, Bioautography, Immunoaffinity column, LC/ESH-MS/MS	Hydroponic	N/A	N/A	N/A	N/A	↑(CN-Cbl)	N/A	Bito et al., 2013

Folate deficiency is prevalent worldwide and over 1.6 billion people are struggling with these deficiencies. Folate deficiency occurs along with iron deficiency and induces megaloblastic anemia in severe deficiency status due to reduced oxygen-carrying capacity (Titcomb and Tanumihardjo, 2019). However, fortification of folic acid, as a synthetic form of folate, has successfully mitigated folate deficiency. Unlike limited bioavailability of vitamin B₁₂, folic acid has over 85% bioavailability and folic acid has approximately 70% higher bioavailability than food folate (Dary, 2008). There are various folate fortified cereals and grains products aiming to reduce the incidence of neural tube defections (NTDs) (Crider et al., 2011). NTDs, as birth defects, occur when the neural tube is exposed to underlying neural tissue owing to the failure of closure during the early embryonic development. Mandatory folic acid fortification programs have been carried out in 53 countries and wheat flour is most widely fortified with folate (Crider et al., 2011). Food fortification with folic acid provides sufficient amounts of folic acid to meet individual and global requirements. However, fortified cereal grain products do not adequately reach all women of reproductive age. Furthermore, there are emerging concerns about the excessive intake of folic acid from the fortified food adversary impacts on pernicious anemia, known as vitamin B₁₂ deficiency (Crider et al., 2011). For instance, vitamin B₁₂ deficient people have a higher possibility of developing neurologic disorders from increased folic acid intake (Carmel, 2011).

A recent study conducted by Darwish et al. (2021) shows that folic acid and iron can be fortified via bovine serum albumin-nanoparticles in stirred functional yogurt (SFY) (BSA-NPs). BSA-NPs are coated with amino acids (lysine) allowing the positive/negative charge of molecules to absorb electrostatically without any other compounds' intervention. BSA-NPs show stable applications and BSA-NPs loaded with folic acid/iron restore most of the monitored plasma iron parameters in SFY products. This fortified SFY retained iron and protein without adverse effects or architectural changes in the liver or kidney. Furthermore, it contributes to enhancing water-holding capacity, microstructure, and overall acceptability of sensors (Darwish et al., 2021). This study successfully introduces the nano-encapsulation technique for enhancing iron and folic acid addressing their physiochemical interaction between dairy food products.

Iron Fortification Using Nanoparticle Technology

As a co-factor in photosynthetic, iron is an essential nutrient for photosynthetic organisms involving various metabolic mechanisms such as electron transport chain (Davies, 2018). Iron deficiency is concomitant with vitamin B₁₂ deficiency masking the macrocytosis, typically seen in vitamin B₁₂ deficiency. Due to vitamin B₁₂/folate deficiencies, ineffective formation of the red cell is a block in iron utilization, causing increased serum iron levels. If the hemolytic anemia condition persisted, iron may be depleted and eventually cause iron deficiency anemia (Green, 2017). Iron and iron oxide nanoparticle (Fe- and Fe₂O₃ NPs) application has been widely conducted because Fe/Fe₂O₃ NPs enhance the development of shoots/roots, plant growth, and

yields in potato, tomato plant (Shankamma et al., 2016), chili pepper (Davies, 2018), and bean seedlings (Duran et al., 2018; see **Table 2**). Iron bioavailability depends on the ferrous sulfate standards indicating high bioavailability in highly water-soluble compounds (Clarke, 1995).

Davies (2018) highlights that the application of nanoparticles with innovative synthesis methods successfully fortify iron, zinc, and calcium in potato, tomato, and chili pepper without requiring conventional breeding. Potato tubers were propagated with iron/iron oxide nanoparticle (FeNP/Fe₃O₄) coated with histidine (His) with an average of 4.732 nm (n⁻²⁰). FeNP + His solution was applied via the foliar application and hydroponic nutrient solution with 8, 12, and 16 mg/L concentration. The application of FeNP+His 16mg/L significantly increased the Fe contents in both potato skin and tuber owing to the nano size of the Fe+His penetrating and accumulating in the tuber. FeNP+His is also treated in tomato and chili pepper with different concentrations. All concentrations (6, 12, and 24mg/L) of FeNP+His significantly increased Fe contents in tomato with the greatest increase obtained by 6 mg/L. Furthermore, tomato treated with FeNP+His (particularly with 12 mg/L dose) showed increased weight in the ripened fruits and produced 146.38% more than control. FeNP + His with 6 mg/L contributes significantly to increasing the fresh weight of tomato from 287.21 to 501.08 g. Fe-fortified tomato displayed no phytotoxicity effects on excessive amounts of Fe treatment. In chili pepper trials, FeNP+His 6mg/L treatment increases plant height (70 nm) and fresh weight. Among various varieties of chili peppers, C. Chinese varieties gained a significant increase in Fe content with 6mg/L treatment. This study highlights that the hydroponic propagation contributes to fortifying micronutrient levels with advantageous conditions by providing adequate soil and compost substrates.

Zinc Fortification Using Nanoparticle Technology

Over 20% of the worldwide population could have risks of zinc deficiency based on the zinc intake and bioavailability estimations from food balance data obtained by the FAO (Smith et al., 2006). Zinc deficiencies widely occur in different geographical regions including South Asia (particularly in India and Bangladesh), Africa, and the Western Pacific. Zinc deficiency occurs along with iron deficiency, which is inhibited by phytates presence (Smith et al., 2006). Zinc and zinc oxide have been fortified in various food crops such as maize (Itrotwar et al., 2020), potato (Davies, 2018), soybean (Hoe et al., 2018), rice (Rameshraddy et al., 2017; Kasivelu et al., 2020), and barely (Seaman, 2017; **Table 2**).

Several studies present successful ZnO fortification in food crops using nanoprimering and root/foliar application enhancing germination, plant growth, and harvest yield. Zn nanoparticle application also enhances the contents of zinc in potato and rice. Davies (2018) addressed the level of zinc in potato enhanced by zinc nanoparticle coated with histidine (ZnNP + His). The application of ZnNP + His presents a positive effect of tuber fortification due to significantly increased amounts of ZnO in potato tubers with 8 mg/L concentration in both hydroponic propagation system and compost media. However, ZnO is rapidly aggregated in an aqueous nutrient solution in hydroponic systems. The aggregation of nanoparticles is one

TABLE 2 | Micronutrient fortification relevant to vitamin B₁₂ deficiency in food crops using nanoparticles: Iron (Fe) and Zinc (Zn).

Plant	NPs	NPs mass/size	Analysis tools	Media	Germ.	Seedling phenotype	Root Vigor	Yield	Nutrient	Toxicity	References
Potato and tomato	Fe ₃ O ₄ NPs	8–32 mg/L, 3.62–20.18 nm	NPs synthesis (coating), SDR (spinning disc reactor), TEM, SEM, FTIR, ICP-OES and XRD	Hydroponic	N/A	↑ (Tuber)	↑	↑ (Tuber)	↑ (Fe in skin and tuber)	X (in Tomato)	Davies, 2018
Bean	Fe ₃ O ₄ NPs and Fe ₃ O ₄ -PEG NPs	1–1000 mg/11 nm and 12 nm	EDXRF, XRD, XRF, TEM, DLS, XANES and SEM-EDX	Seed priming	—	↑ (1 to 100 mg/L)	↑	↑	↑ (Seed coat and radicle)	O (in high concentration)	Duran et al., 2018
Tomato	Magnetic Fe ₂ O ₃ NPs	50–800 mg/L	UV-vis spectrophotometer, TEM and XRD	Greenhouse and hydroponic	↑	↑ (50–200 mg/L)	↑	N/A	↑ (Root hair, root tip)	O (in high concentration)	Shankramma et al., 2016
Maize	ZnO	10–200 mg/L, ~37 nm	TEM, HR-SEM, measurement of seed germination and seedling parameters	Seed priming, <i>in vitro</i>	↑	↑	↑	N/A	N/A	X	Itrotwar et al., 2020
Maize and Rice	γ-Fe ₂ O ₃ and ZnO	100–2000 ppm	SEM, UV-DRS, XRD, FT-IR, germination and vigor analysis	<i>In vitro</i>	↑	↑	↑	↑	N/A	N/A	Kasivelu et al., 2020
Potato	Zn	13.18–13.73 nm	NPs synthesis (coating), SDR (spinning disc reactor), TEM, SEM, FTIR, ICP-OES and XRD	Hydroponic	N/A	↑ (Tuber)	↓ (No. tuber)	↑	↑ (Zn in tubers)	↑ chance (in hydroponic)	Davies, 2018
Rice	Zn, ZnO and ZnSO ₄ NPs	500–1500 ppm, 30 nm	Seed priming (1000 ppm) and foliar application, ICP-OES and RT-PCR analysis	<i>In vitro</i> (drought stress)	↑ (Zn, ZnO seed priming)	↑ (ZnO seed priming), ↓ (In high ZnO, ZnSO ₄)	↑ (Zn seed priming)	↑ (ZnO > Zn)	↑ (ZnO > ZnSO ₄)	O (1500 ppm of ZnO and ZnSO ₄)	Rameshraddy et al., 2017
Soybean	Fe, ZnO, Cu and Co	5–500 mg/L, 40–40 nm	Morphological and cytological analysis,	<i>In vitro</i>	↑	↑	↑	N/A	N/A	↑ % of Zn	Hoe et al., 2018
Barely fodder	Teprosyn Zn/P	0.5–3.6 ml	LA-ICP-MS, MALDI-MS, HPLC-ICP-MS	Seed priming, hydroponic	↑	↑	↓	N/A	↑	N/A	(Seaman, 2017)

of the major issues because the size of the aggregated particle become larger which consequently reduces bioavailability of the nanoparticle. Also, increased ZnNP + His application can also cause phototoxic effects on food crops such as severe stunted leaves. Therefore, it is necessary to avoid ZnO aggregation and excessive applications for food fortification (Davies, 2018). Rameshraddy et al. (2017) also presents that ZnO nanoparticle application improves plant physiological growth (e.g., growth, yield, and quality), and enhances drought stress tolerance along with the increased contents of ZnO in rice. The study also highlights that high concentration of ZnO and ZnSO₄ (at 1500 ppm in both) significantly reduce rice seedling vigor which might cause toxicity (Rameshraddy et al., 2017).

Micronutrient Fortification for Plants Grown in Hydroponic and Aeroponic System

Hydroponic and aeroponic systems support effective fortification of micronutrients with high productivity and efficiency by providing optimized year-round production (Rouphael and Kyriacou, 2018). Hydroponic cultivation refers to the growing method based on the recirculated inorganic/organic nutrient solution instead of soil cultivation. In hydroponic systems, the plant roots are immersed partially or completely in a nutrient solution. On the other hand, aeroponic system exposes the plant roots to aerosol droplets containing micro-/macro-nutrient (Eldridge et al., 2020). In aeroponic systems, droplet size is one of the major parameters for determining the absorption effectiveness directly influencing plant growth and it can be classified into spray (over 100 μm), fog (1–100 μm), and mist (1–35 μm) (Niam and Sucahyo, 2020). Both hydroponic and aeroponic cultivations provide favorable environments for plant growth. Nanoparticles can be applied into nutrient solutions for hydroponic cultivation. Better absorption and translocation of nanoparticles was observed in plants grown in hydroponic systems compared to soil cultivation due to more conductive aggregation and dissolution of nanoparticles in roots zone in hydroponic system (Kranjc and Drobne, 2019). Some minerals/micronutrients present low mobility or are even unavailable to plants in the soil depending on the physicochemical characteristics including pH, composition, and electrical conductivity (Freire et al., 2020). Therefore, hydroponic cultivation systems can be an efficient strategy for vitamin B₁₂ fortification by applying vitamin B₁₂-enriched nutrient solution precisely with nanoparticles coating techniques.

Bito et al. (2013) shows that vitamin B₁₂-loaded nutrients significantly improve the level of vitamin B₁₂ in lettuce grown in hydroponic systems. Hydroponic growing systems have better management of water and nutrient supply without pathogen or bacteria risks or leaching issues. This study dissolves CN-Cbl into hydroponic nutrient solution at 5 μmol/L for lettuce growing. The results indicated the majority of CN-Cbl accumulated in leaves (86%) which may be a promising source of free CN-Cbl in food crops. Approximately 164.6 μg/g fresh weight of lettuce would provide the recommended daily allowance for vitamin B₁₂ (2.4 μg/g). This study addressed the expected costs for CN-Cbl-nutrition solution for lettuce fortification which was calculated to be approximately U.S \$0.06. Therefore, compared

to conventional supplementary programs, it would be a cost-effective fortification strategy with an excellent source of free CN-Cbl for the populations who have plant-based diets or the elderly (Bito et al., 2013). One of the main challenges was to maintain the stability of CN-Cbl in hydroponic nutrient solution for future application by controlling light conditions (Bito et al., 2013). Additionally, the different concentrations of vitamin B₁₂ variously impact crop growth performance and rate of vitamin B₁₂ accumulation, so it is critical to identify the optimal concentration of vitamin B₁₂ solution for fortification of living plants. Further study is required to undertake the actual bioavailability of vitamin B₁₂ in fortified food crops.

Previous studies present successful approaches for sustainable fortification on food crops grown in hydroponic systems by enhancing mineral/micronutrient and non-essential micronutrients such as folate, iodine, and selenium. Watanabe et al. (2017) present that the contents of folate significantly increased approximately 1.8-fold in spinach with the applications of folate and phenylalanine in hydroponic cultivation. As non-essential micronutrients, selenium and iodine have been notably investigated by several previous studies. Puccinelli et al. (2021) showed that iodine was fortified in both basil and lettuces grown in closed-loop hydroponic cultivation with 10 μM potassium iodide-loaded nutrient solution. The outcome of the study presents that the growth rate of lettuces in aeroponic systems is much higher than in the hydroponic system, due to higher levels of dissolved oxygen in the nutrient solution in aeroponic systems. Therefore, aeroponics can provide an efficient growing system for nutrient fortification in plants with greater oxygen availability in the root zone, enhancing water and nutrients use efficiency.

Ultrasonic atomization aeroponic, as a novel hybrid system, enables more precise and effective control by producing very fine micro-size droplets (1–5 μm) of nutrient solution generated by ultrasonic atomization disks (Niam and Sucahyo, 2020). Therefore, nanoparticles can be applied into nutrient solution within ultrasonic atomization disk, and it would be possible to increase targeted micronutrients in food crops due to their better solubility and permeability. Further study would be useful to investigate the optimal droplet size, flow rate, and nutrient solution conditions such as temperature and nanoparticle application in aeroponic systems (Niam and Sucahyo, 2020). It is required to identify optimal nanoparticle application to fortify micronutrients in closed soilless cultivation such as in hydroponic and aeroponic systems, especially in ultrasonic atomization aeroponics. Also, efforts should be made for identifying adequate growing systems for food crops based on understanding of their genotypes and physiological characteristics.

Mechanisms of Uptake and Translocation of Nanoparticles in Plants Factors Impacting the Uptake and Translocation of Nanoparticles in Plants

It is necessary to understand how the plant absorbs, transports, and accumulates the nanoparticle in order to enhance nutrient/mineral contents in food crops in targeted locations (such as edible parts of the food crops). Nanoparticle application

enables the effective smart-delivery functions owing to their high solubility and mobility (Shang et al., 2019). Several studies have been conducted to investigate the mechanisms of uptake, transport, and accumulation of micronutrient/mineral nanoparticle in living plants by applying integrated analytic methods such as microscopy and mass spectrometry. However, due to the complexity and interaction between nanoparticles and plants, exact mechanisms should be investigated. Additionally, various factors significantly impact the absorption and uptake of nanoparticles in living plants. First, the size of nanoparticles is one of the main restrictions for penetration into cell wall pores in plants, which are 5–20 nm wide (Pérez-de-Luque, 2017). Second, the type and chemical composition of nanoparticles have a great influence on the absorption or uptake of nanoparticles (Rico et al., 2011; Pérez-de-Luque, 2017).

Coating materials for nanoparticles and the chemical composition of their surfaces can also alter their absorption or accumulation in living plants (Schwab et al., 2016; Pérez-de-Luque, 2017). Third, plant species impact the speed of absorption and distribution of nanoparticles due to different plant physiological traits, such as the thickness or architecture of the barriers, including the size of the cell wall pore, Casparian strip, and xylem thickness (Cifuentes et al., 2010). For instance, paramagnetic-coated nanoparticles were applied to four different crops, wheat, pea, sunflower, and tomato, with results indicating that the nanoparticles accumulate in different plant locations, such as vascular tissues or trichomes (Schwab et al., 2016). The effects of ENMs on plants can be highlighted as dependent on crop life stages, including (i) germination and early seedling growth, (ii) post-transplant and further growth of seedlings, and (iii) mature/harvest stages (Servin and White, 2016). Finally, the method of application plays an important role in nanoparticle pathways in plants. Nanoparticles in terrestrial plants generally accumulate and aggregate at the root zone, which is greatly influenced by microenvironmental conditions, including symbiosis with bacteria or fungi (Schwab et al., 2016). According to Schwab et al. (2016), the application of foliar ENMs in soil via roots is particularly challenging due to the interaction between microorganisms in soils and additional complex soil conditions. Delivering ENMs via foliar application or seed coating may improve the uptake or distribution of ENMs in living plants. Further study is required to investigate the optimum application method of ENMs based on understanding the transdisciplinary factors of growing plants.

Uptake and Translocation of Nanoparticles in Plants

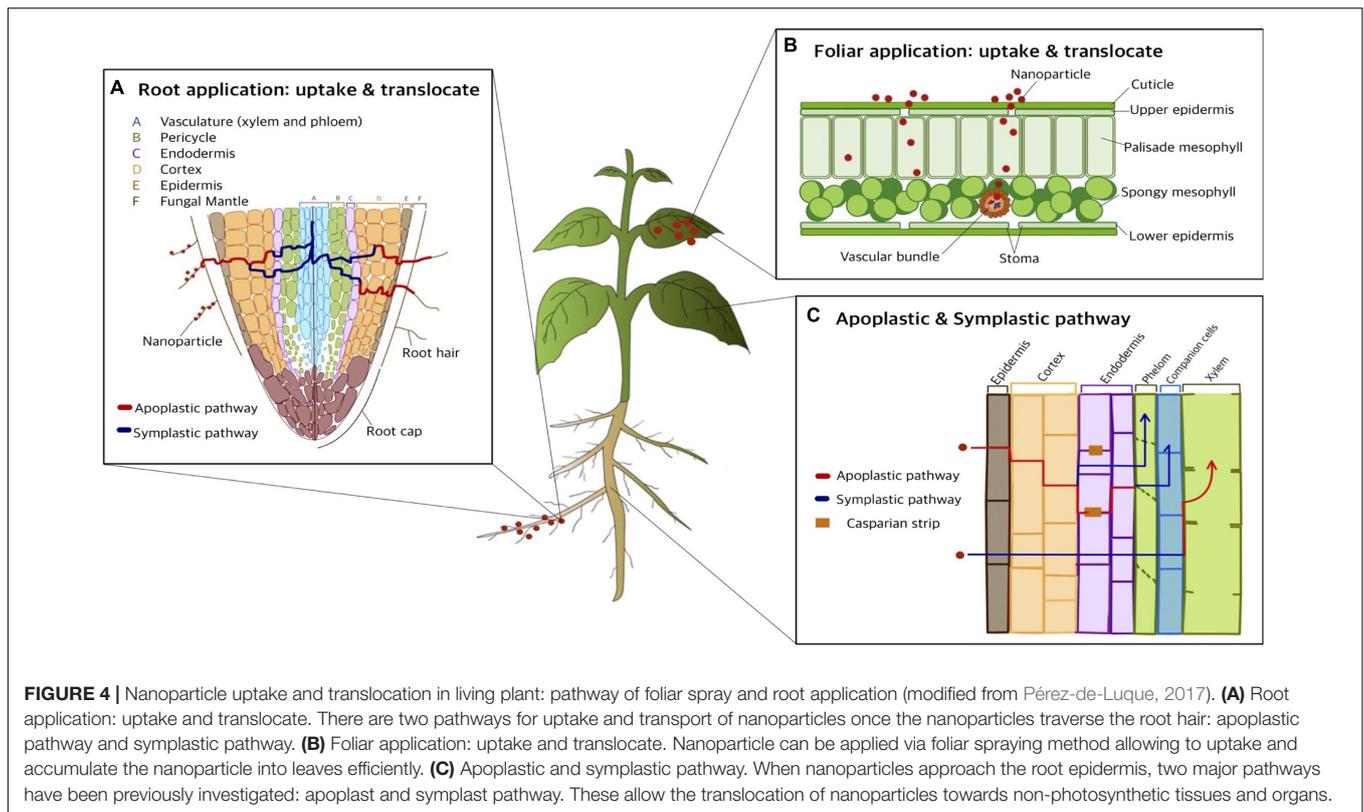
There are two primary paths for the uptake, translocation, and accumulation of nanoparticles: foliar and root (Figure 4). In foliar application, the cuticle is the major obstacle preventing nanoparticles from entering plant tissues due to the water resistance from waxy components. Therefore, the major route for nutrient uptake is via cuticular pathways, including lipophilic pathways. The lipophilic pathway involves diffusion via cuticular wax layers, whilst the hydrophilic pathway occurs via polar aqueous pores located in the cuticle or stomata in leaves: the “stomatal pathway.” Uptake via the hydrophilic pathway is influenced by stomatal openings, such as the morphological traits of stomatal apertures in various plant species. After

the stomatal pathway, nanoparticles are transported via the vascular pathway, consisting of the xylem and phloem systems. The direction of flow in the xylem system is from root to shoot, whilst the flow direction in the phloem system is from shoot to root. Thus, in foliar applications, nanoparticles are translocated only via the phloem system from leaves to roots. Accordingly, previous studies have found that nanoparticles can be transported via xylem and phloem systems. However, the mechanisms of nanoparticle translocation in xylem and phloem still require further investigation (Lv et al., 2019). Due to the size of the cuticular pore, which is approximately 2 nm in diameter, the stomatal pathway is the most likely route for nanoparticle translocation. Therefore, from foliar spraying application, nanoparticles can be found in leaf stomata and deeper plant tissues, such as phloem, with translocation (Lv et al., 2019).

Compared to foliar application, many more studies have focused on root application. In root application, nanoparticles passively cross the permeable cuticle region and the pores of the cell wall. There are several factors that influence the uptake of nanoparticles in plants via root application, such as nanoparticle size, fictionalization of surface (e.g., positive and negative charge), morphological traits, conditions of exposure, plant species and growth stage, root integrity, and rhizospheric processes (Lv et al., 2019). When applied in the root zone, there are several barriers that nanoparticles cross, including physiological barriers (e.g., root hair) or environmental conditions of the root (e.g., bacteria and mycorrhizae), mucilage/exudates, and cuticle of the root periderm. During interactions with barriers, nanoparticles penetrate the cell membrane by fluid-phase endocytosis, passive diffusion, or mechanical piercing and then accumulate in the mucilage, cuticle, and cell wall. Afterward, microorganisms, through processes such as mucilage exudation or biomineralization, assist nanoparticles in transport (Schwab et al., 2016).

Once nanoparticles penetrate the plant cell membrane through the pores of the roots, they start to diffuse by two pathways: (i) the space between the cell wall and plasma membrane and (ii) the intercellular space without penetrating the cell membrane (shown in Figure 4). Via penetration of roots, nanoparticles traverse from the root surface cuticle to intercellular structures, such as the epidermis, cortex, endodermis, and Casparian strip, and eventually penetrate the shoot via xylem (Lv et al., 2019). When nanoparticles approach the root epidermis, two major pathways have been previously investigated, namely the apoplast and symplast, connecting cell wall and intercellular spaces, and protoplasmic connections through ion channels (Pérez-de-Luque, 2017). The apoplastic pathway plays an important role in radial movement within plant tissues, and it allows nanoparticles to translocate upwards to the aerial part. In addition, once nanoparticles enter the central cylinder, they can move towards the aerial part via the transpiration stream through the xylem.

When the nanoparticles translocate through the apoplastic pathway, they face a barrier reaching the xylem through the root, which is called the Casparian strip and is mainly required in the symplastic pathway. The Casparian strip is a belt of cell wall components sealed by lipophilic hydrocarbons



located adjacent to the vascular system. Casparian strips hamper nanoparticle translocation from roots to shoots if the particle size is too large (Cai et al., 2017; Lv et al., 2019). The symplastic pathway, which involves the movement of water and nutrients, utilizes the sieve tube elements in the phloem, allowing the translocation of nanoparticles towards non-photosynthetic tissues and organs. In the symplastic route, nanoparticles penetrate the cytoplasm or adjacent cells through plasmodesmata, which enables intercellular communication by linking the cytoplasm between adjacent cells. There are several hypotheses regarding symplastic pathways, involving aquaporins, ion channel interconnection, endocytosis, and breaking of membrane tubulation. Endocytosis is the most feasible transmembrane pathway, including for nutrient uptake and microbial interactions (Lv et al., 2019). In root application, the nanoparticles are mainly translocated through the xylem but not the phloem and move from root to shoot and leaves (Pérez-de-Luque, 2017).

Nanoparticles are translocated and accumulate at different speeds and locations in plants based on their component materials (Pérez-de-Luque, 2017). Generally, nanoparticles accumulate in fruits, grains, flowers, or young leaves by traveling to the vascular system (Lu et al., 2008; Khot et al., 2012). Owing to advanced spectrometry or mass spectrometry techniques, it is possible to determine the overall uptake, translocation, and accumulation of nanoparticles in living plants. For instance, transmission electron microscopy (TEM), scanning electron microscopy (SEM), and X-ray spectroscopy analysis allow us

to determine the sizes, shapes, and locations of nanoparticles in plant cells (Khot et al., 2012; Davies, 2018). Inductively coupled plasma optical emission spectroscopy (ICP-OES) and ICP-mass spectrometry (ICP-MS) precisely analyze nanoparticle compounds (Pérez-de-Luque, 2017). However, further studies should be conducted to identify the interaction between nanoparticles that are absorbed by plants and animals or humans that consume plants treated with nanoparticles (Pérez-de-Luque, 2017). Additionally, the uptake, distribution, and accumulation mechanisms vary and are greatly influenced by several components. Mozafar and Oertli (1992) found the uptake of vitamin B₁₂ contents in soybean plants using the radioisotope dilution (RID) technique. By identifying vitamin B₁₂ and vitamin B₁₂-binding sites, the study showed that vitamin B₁₂ transport from vitamin B₁₂-enriched nutrient to soybean plants that was found in unifoliate, trifoliate leaves, and stems. Vitamin B₁₂ transports rapidly from root to shoot and more vitamin B₁₂ was accumulated in the leaves in the plants aerated with nitrogen.

Overall Opportunities and Challenges of Nanoparticle Applications for Food Crop

As a promising technology for sustainable agriculture, nanotechnology poses numerous opportunities. First, ENMs enhance the overall efficiency of water use and light and agrochemical use (Zhou et al., 2015). Increased water use efficiency is realized in nanofertilizer applications by increasing the retaining capacity, decreasing the loss amount, and

increasing the efficiency of utilization of water and nutrients (Giraldo et al., 2014). ENMs show the potential to increase light use efficiency in plants by augmenting chloroplast photosynthetic activities and enhancing chloroplast reactive oxygen species (ROS) (Giraldo et al., 2014). Agrochemical application shows significant enhancement of performance by decreasing adverse environmental effects, including leaching. Nanofertilizers constitute an alternative technique to the use of conventional fertilizers because they boost crop yield, quality, and tolerance to abiotic stress owing to their high efficiency (Kah et al., 2018). Compared to conventional fertilizer, nanofertilizer can increase crop production by approximately 20–30% and achieve similar levels of plant protection and nutrient enhancement with lower fertilizer usage (Kranjc and Drobne, 2019). Owing to their biochemical and physical characteristics, ENMs can be delivered precisely to the plant interior to enhance the uptake of nutrients and fertilizer (Lowry et al., 2019). Second, nanotechnology promotes soil sustainability by lowering additional fertilizer inputs via increased nutrient efficiency. Additionally, ENMs have the potential to increase fungi, which is beneficial for plant growth, or to encourage plant root vitality via nitrogen-fixing bacteria. Furthermore, nanotechnology enables the control of biotic stress such as pathogens or weed competitors and abiotic stress, including extreme temperature and water stress (Lowry et al., 2019).

There are several challenges for further deployment of nanotechnology in agri-food. One of the most significant challenges in nanotechnology is the potential risks and risk perceptions regarding nanoparticles (Lowry et al., 2019). Consumer perception and acceptance are negative towards ENMs in food crops. According to Giles et al. (2015), food product-integrated nanotechnology is less likely to be accepted by consumers than food packaging within nanotechnology. However, of most significant concern regarding ENM applications in agricultural food is that sectors have only focused on the input of nanoparticles as ingredients or additives to food products directly, whilst comparatively little concern has been aroused regarding nanoagrochemical applications, including nanofertilizers or nanopesticides. Furthermore, these observations indicate that the consumer accepts nanotechnology application in agricultural food production when perceived benefits outweigh the perceived risks. For the future application of nanotechnology in food crops, it is necessary to encourage marketing and commercialization for nanotechnology-processed food crops by increasing acceptance along with expert opinions regarding safety or acceptability (Giles et al., 2015). Further efforts should address and secure the availability of nanoagrochemicals within effective legislation and long-term risk assessment for the entire life cycle of nanoparticle application to food crops and further human/animal intake (Iqbal, 2019).

Additionally, although there have been several attempts, it is still necessary to investigate the interaction between ENMs and plants and to understand the mechanisms of uptake and translocation of nanoparticles in living plants (Nair et al., 2010; Pérez-de-Luque, 2017; Lowry et al., 2019). Precisely delivering ENMs remains a challenge considering precise target location, exact time of application, and appropriate dose of ENMs. Additionally, in nanoparticle applications, particle size plays an

important role, influencing the translocation of nanoparticles in living plants. Nanoparticle size can be increased beyond the nano range and attain different shapes due to continuous aggregation which also may result in Ostwald ripening. Thus, synthesis process of nanoparticles should be developed to control nanosize and composition of nanoparticles, particularly when scaling up for commercial usages (Davies, 2018; Akbar et al., 2020). Davies (2018) presents a successful synthesis approach for producing stable sizes of nanoparticles ranging 3.62–20.18 nm for calcium oxide, 3–7.6 nm for iron oxide, and 7.03–15.41 nm for zinc oxide. The study exploited spinning disk reactor (SDR) to synthesize nanoparticles, which provided relatively rapid, efficient, and cost-effective synthesis process. SDR method allows the production of large amounts of nanoparticles with uniform size, and electrostatic coating method with histidine (amino acid) also enhances the mobility and solubility of metal oxide nanoparticle by enhancing retention capabilities and decreasing leaching issue (Davies, 2018). Further guidance documents and validated methods for nanoparticle size measurement should be established (Parisi et al., 2014).

CONCLUSION

Overall, this review paper suggests that nanoparticle technology would be a great sustainable strategy to mitigate vitamin B₁₂ deficiency by providing efficient smart delivery of micronutrient/mineral to food crops. This review paper suggests several key findings:

- (1) A sustainable vitamin B₁₂ fortification approach should be balanced with other micronutrient/mineral deficiency focusing on not only increasing the contents but also the bioavailability of vitamin B₁₂.
- (2) Nanoparticle technologies, such as seed priming and nanoparticle applications, can accelerate sustainable food fortification with better quality and yield by targeted macro- or micronutrient enhancement without further plant breeding or genetic technology.
- (3) To develop food fortification along with nanoparticle applications, the participation of consumers and experts is significant. Further attempts should focus on enhancing the recognition of safety and applicability of nanotechnology for food crops along with investigations of the phytotoxicity and safety of nanoparticle application.
- (4) Integration of hydroponic/aeroponic systems and nanoparticle applications is expected to be a promising technology for not only better productivity but also micronutrient fortification (e.g., vitamin B₁₂), which can be applied to vertical farming or indoor farming in future.

AUTHOR CONTRIBUTIONS

SO contributed to conception and review of the study, wrote first draft of the manuscript. SO, GC, and CL contributed to manuscript revision, read and approved the submitted version. All the authors contributed to the article and approved the submitted version.

REFERENCES

- Abdel-Aziz, H., Hasaneen, M., and Omer, A. (2016). Nano chitosan-NPK fertilizer enhances the growth and productivity of wheat plants grown in sandy soil. *Span. J. Agric. Res.* 14:e0902. doi: 10.5424/sjar/2016141-8205
- Agriculture and Rural Development (2019). *Global Food Supply and Demand, Consumer Trends, Trade Challenges*. In *EU Agricultural Markets Briefs*. Rome: FAO, 16.
- Akbar, S., Tauseef, I., Subhan, F., Sultana, N., Khan, I., Ahmed, U., et al. (2020). An overview of the plant-mediated synthesis of zinc oxide nanoparticles and their antimicrobial potential. *Inorg. Nano Metal Chem.* 50, 257–271. doi: 10.1080/24701556.2019.1711121
- Al-juthery, H., and Al-Shami, Q. (2019). The effect of fertigation with nano NPK fertilizers on some parameters of growth and yield of potato (*Solanum tuberosum* L.). *J. Agric. Sci.* 9, 225–232. doi: 10.33794/qjas.Vol9.Iss2.93
- Allen, L. H., Rosenberg, I. H., Oakley, G. P., and Omenn, G. S. (2010). Considering the case for vitamin B12 fortification of flour. *Food Nutr. Bull.* 31(1 Suppl.), 36–46. doi: 10.1177/15648265100311s104
- Andrés, E., Loukili, N. H., Noel, E., Kaltenbach, G., Ben Abdelgheni, M., Perrin, A. E., et al. (2004). Vitamin B12 (cobalamin) deficiency in elderly patients. *CMAJ* 171, 251–259. doi: 10.1503/cmaj.1031155
- Artés-Hernández, F., Formica-Oliveira, A. C., Artés, F., and Martínez-Hernández, G. B. (2017). Improved quality of a vitamin B12-fortified 'ready to blend' fresh-cut mix salad with chitosan. *Food Sci. Technol. Int.* 23, 513–528. doi: 10.1177/1082013217705036
- Bito, T., Bito, M., Asai, Y., Takenaka, S., Yabuta, Y., Tago, K., et al. (2016). Characterization and quantitation of Vitamin B12 compounds in various *Chlorella* supplements. *J. Agric. Food Chem.* 64, 8516–8524. doi: 10.1021/acs.jafc.6b03550
- Bito, T., Ohishi, N., Hatanaka, Y., Takenaka, S., Nishihara, E., Yabuta, Y., et al. (2013). Production and characterization of cyanocobalamin-enriched lettuce (*Lactuca sativa* L.) grown using hydroponics. *J. Agric. Food Chem.* 61, 3852–3858. doi: 10.1021/jf305033s
- Bito, T., Teng, F., Ohishi, N., Takenaka, S., Miyamoto, E., Sakuno, E., et al. (2014). Characterization of vitamin B12 compounds in the fruiting bodies of shiitake mushroom (*Lentinula edodes*) and bed logs after fruiting of the mushroom. *Mycoscience* 55, 462–468. doi: 10.1016/j.myc.2014.01.008
- Bodirsky, B. L., Dietrich, J. P., Martinelli, E., Stenstad, A., Pradhan, P., Gabrysch, S., et al. (2020). The ongoing nutrition transition thwarts long-term targets for food security, public health and environmental protection. *Scientific Reports* 10:19778. doi: 10.1038/s41598-020-75213-3
- Bouis, H. E., and Saltzman, A. (2017). Improving nutrition through biofortification: a review of evidence from HarvestPlus, 2003 through 2016. *Glob. Food Sec.* 12, 49–58. doi: 10.1016/j.gfs.2017.01.009
- Brito, A., Habeych, E., Silva-Zolezzi, I., Galaffu, N., and Allen, L. H. (2018). Methods to assess Vitamin B12 bioavailability and technologies to enhance its absorption. *Nutrit. Rev.* 76, 778–792. doi: 10.1093/nutrit/nuy026
- Cai, F., Wu, X., Zhang, H., Shen, X., Zhang, M., Chen, W., et al. (2017). Impact of TiO₂ nanoparticles on lead uptake and bioaccumulation in rice (*Oryza sativa* L.). *NanoImpact* 5, 101–108. doi: 10.1016/j.impact.2017.01.006
- Carmel, R. (2008). Efficacy and safety of fortification and supplementation with vitamin B12: biochemical and physiological effects. *Food Nutr. Bull.* 29, 177–187. doi: 10.1177/15648265080292S121
- Carmel, R. (2011). Mandatory fortification of the food supply with cobalamin: an idea whose time has not yet come. *Bone* 23, 1–7. doi: 10.1007/s10545-010-9150-2.Mandatory
- Cifuentes, Z., Custardoy, L., de la Fuente, J. M. D., Marquina, C., Ibarra, R., Rubiales, D., et al. (2010). Absorption and translocation to the aerial part of magnetic carbon-coated nanoparticles through the root of different crop plants. *J. Nanobiotechnol.* 8:26. doi: 10.1186/1477-3155-8-26
- Clarke, R. (1995). *Annex 4—Micronutrient Fortification of Food: Technology and Quality Control*. In *Annex 4—Micronutrient Fortification of Food: Technology and Quality Control*. Rome: FAO.
- Crider, K. S., Bailey, L. B., and Berry, R. J. (2011). Folic acid food fortification—its history, effect, concerns, and future directions. *Nutrients* 3, 370–384. doi: 10.3390/nu3030370
- Darwish, A. M. G., Soliman, T. N., Elhendy, H. A., and El-Kholy, W. M. (2021). Nano-encapsulated iron and folic acid-fortified functional yogurt enhance anemia in albino rats. *Front. Nutr.* 8:654624. doi: 10.3389/fnut.2021.654624
- Dary, O. (2008). Establishing safe and potentially efficacious fortification contents for folic acid and vitamin B12. *Food Nutr. Bull.* 29 2(Suppl.), 214–224. doi: 10.1177/15648265080292s126
- Davies, K. (2018). *Biofortification of Potato (Solanum tuberosum) Using Metal Oxide Nanoparticles*. Available online at: <http://irep.ntu.ac.uk/> (accessed November 20, 2020).
- Doets, E. L., In't Veld, P. H., Szczecińska, A., Dhonukshe-Rutten, R. A. M., Cavelaars, A. E. J. M., Van 't Veer, P., et al. (2013). Systematic review on daily Vitamin B12 losses and bioavailability for deriving recommendations on Vitamin B12 intake with the factorial approach. *Ann. Nutr. Metab.* 62, 311–322. doi: 10.1159/000346968
- Duran, N. M., Medina-Llamas, M., Cassanji, J. G. B., De Lima, R. G., De Almeida, E., Macedo, W. R., et al. (2018). Bean seedling growth enhancement using magnetite nanoparticles. *J. Agric. Food Chem.* 66, 5746–5755. doi: 10.1021/acs.jafc.8b00557
- Eldridge, B. M., Manzoni, L. R., Graham, C. A., Rodgers, B., Farmer, J. R., and Dodd, A. N. (2020). Getting to the roots of aeroponic indoor farming. *New Phytol.* 228, 1183–1192. doi: 10.1111/nph.16780
- Elemike, E. E., Uzoh, I. M., Onwudiwe, D. C., and Babalola, O. O. (2019). The role of nanotechnology in the fortification of plant nutrients and improvement of crop production. *Appl. Sci.* 9:499. doi: 10.3390/app9030499
- FAO, IFAD, UNICEF, WFP and WHO (2020). *The State of Food Security and Nutrition in the World 2020*. In *The State of Food Security and Nutrition in the World 2020*. Rome: FAO, doi: 10.4060/ca9692en
- FAO (2017). *Nutrition-Sensitive Agriculture and Food Systems in Practice. Options for Intervention*. In *Nutrition-Sensitive Agriculture and food Systems in practice. Options for Intervention*. Rome: FAO.
- Fidaleo, M., Tacconi, S., Sbarigia, C., Passeri, D., Rossi, M., Tata, A. M., et al. (2021). Current nanocarrier strategies improve vitamin b12 pharmacokinetics, ameliorate patients' lives, and reduce costs. *Nanomaterials* 11:743. doi: 10.3390/nano11030743
- Forgrieve, J. (2019). *Plant-Based Food Sales Continue To Grow By Double Digits, Fueled By Shift In Grocery Store Placement*. *Forbes*. 1–6. Available online at: <https://www.forbes.com/sites/janetforgrieve/2019/07/16/plant-based-food-sales-pick-up-the-pace-as-product-placement-shifts/#119d3dd54f75> (accessed July 16, 2019)
- Freire, M., Pereira, M., Lange, N., and Batista, L. (2020). "Biofortification of crop plants: a practical solution to tackle elemental deficiency," in *Sustainable Solutions for Elemental Deficiency and Excess in Crop Plants*, eds M. Freire, M. Pereira, N. Lange, and L. Batista (Singapore: Springer), 135–182. doi: 10.1007/978-981-15-8636-1_7
- Garrod, M. G., Buchholz, B. A., Miller, J. W., Haack, K. W., Green, R., and Allen, L. H. (2019). Vitamin B12 added as a fortificant to flour retains high bioavailability when baked in bread. *Nucl. Instrum. Methods Phys. Res. B* 438, 136–140. doi: 10.1016/j.nimb.2018.05.042
- Giles, E. L., Kuznesof, S., Clark, B., Hubbard, C., and Frewer, L. J. (2015). Consumer acceptance of and willingness to pay for food nanotechnology: a systematic review. *J. Nanopart. Res.* 17:464. doi: 10.1007/s11051-015-3270-4
- Giraldo, J. P., Landry, M. P., Faltermeier, S. M., McNicholas, T. P., Iverson, N. M., Boghossian, A. A., et al. (2014). Plant nanobionics approach to augment photosynthesis and biochemical sensing. *Nat. Mater.* 13, 400–408. doi: 10.1038/nmat3890
- Green, R. (2017). Vitamin B12 deficiency from the perspective of a practicing hematologist. *Blood* 129, 2603–2612. doi: 10.1182/blood-2016-10-569186.In
- Green, R., Allen, L. H., Bjørke-Monsen, A. L., Brito, A., Guéant, J. L., Miller, J. W., et al. (2017). Vitamin B12 deficiency. *Nat. Rev. Dis. Prim.* 3:17040. doi: 10.1038/nrdp.2017.40
- HarvestPlus and FAO (2019). *Biofortification: a Food-Systems Solution to Help End Hidden Hunger*. Rome: FAO.
- Hoe, P. T., Mai, N. C., Lien, L. Q., Ban, N. K., Van Minh, C., Chau, N. H., et al. (2018). Germination responses of soybean seeds under Fe, ZnO, Cu and Co nanoparticle treatments. *Int. J. Agric. Biol.* 20, 1562–1568. doi: 10.17957/IJAB/15.0670
- Hunt, A., Harrington, D., and Robinson, S. (2014). Vitamin B12 deficiency. *BMJ (Online)* 349, 1–7.

- Iqbal, M. (2019). "Nano-fertilisers for sustainable crop production under changing climate_ A global perspective," in *Sustainable Crop Production*, Vol. 8, ed. M. Iqbal (London: InTechOpen), 1–13. doi: 10.5772/intechopen.89089
- Itroutwar, P. D., Kasivelu, G., Raguraman, V., Malaichamy, K., and Sevathapandian, S. K. (2020). Effects of biogenic zinc oxide nanoparticles on seed germination and seedling vigor of maize (*Zea mays*). *Biocatal. Agric. Biotechnol.* 29:101778. doi: 10.1016/j.bcab.2020.101778
- Jedut, P., Sz wajgier, D., and Glibowski, P. (2021). Some plant food products present on the polish market are a source of Vitamin B12. *Appl. Sci.* 11:3601. doi: 10.3390/app11083601
- Jeyasubramanian, K., Gopalakrishnan Thoppey, U. U., Hikku, G. S., Selvakumar, N., Subramania, A., and Krishnamoorthy, K. (2016). Enhancement in growth rate and productivity of spinach grown in hydroponics with iron oxide nanoparticles. *RSC Adv.* 6, 15451–15459. doi: 10.1039/c5ra23425e
- Jones, L. (2020). *Veganism: Why are Vegan Diets on the Rise?—BBC News*. In *BBC News*. London: BBC News, 1–2.
- Kah, M., Kookana, R. S., Gogos, A., and Bucheli, T. D. (2018). A critical evaluation of nanopesticides and nanofertilizers against their conventional analogues. *Nat. Nanotechnol.* 13, 677–684. doi: 10.1038/s41565-018-0131-1
- Kamiński, M., Skonieczna-Żydecka, K., Nowak, J. K., and Stachowska, E. (2020). Global and local diet popularity rankings, their secular trends, and seasonal variation in Google Trends data. *Nutrition* 79–80:11075. doi: 10.1016/j.nut.2020.110759
- Kasivelu, G., Selvaraj, T., Malaichamy, K., Kathickeyan, D., Shkolnik, D., and Chaturvedi, S. (2020). Nano-micronutrients [γ -Fe₂O₃(iron) and ZnO (zinc)]: green preparation, characterization, agro-morphological characteristics and crop productivity studies in two crops (rice and maize). *New J. Chem.* 44, 11373–11383. doi: 10.1039/d0nj02634d
- Kaur, T., Rana, K. L., Kour, D., Sheikh, I., Yadav, N., Kumar, V., et al. (2020). "Microbe-mediated biofortification for micronutrients: present status and future challenges," in *New and Future Developments in Microbial Biotechnology and Bioengineering*, eds A. Rastegari, N. Yadav, and A. Yadav (Amsterdam: Elsevier Inc), 1–17. doi: 10.1016/b978-0-12-820528-0.00002-8
- Keshavarz, H., and Moghadam, R. S. G. (2017). Seed priming with cobalamin (vitamin B12) provides significant protection against salinity stress in the common bean. *Rhizosphere* 3, 143–149. doi: 10.1016/j.rhisph.2017.04.010
- Khot, L. R., Sankaran, S., Maja, J. M., Ehsani, R., and Schuster, E. W. (2012). Applications of nanomaterials in agricultural production and crop protection: a review. *Crop Prot.* 35, 64–70. doi: 10.1016/j.cropro.2012.01.007
- Khush, G. S., Lee, S., Cho, J. Il, and Jeon, J. S. (2012). Biofortification of crops for reducing malnutrition. *Plant Biotechnol. Rep.* 6, 195–202. doi: 10.1007/s11816-012-0216-5
- Kranjc, E., and Drobne, D. (2019). Nanomaterials in plants: a review of hazard and applications in the agri-food sector. *Nanomaterials* 9:1094. doi: 10.3390/nano9081094
- Kumudha, A. (2015). Effect of different extraction methods on vitamin B12 from blue green algae, spirulina platensis. *Pharm. Anal. Acta* 6:337. doi: 10.4172/2153-2435.1000337
- Lawrence, A. D., Nemoto-Smith, E., Deery, E., Baker, J. A., Schroeder, S., Brown, D. G., et al. (2018). Construction of fluorescent analogs to follow the uptake and distribution of cobalamin (Vitamin B 12) in bacteria, worms, and plants. *Cell Chem. Biol.* 25, 941.e–951.e. doi: 10.1016/j.chembiol.2018.04.012 941-951.e6,
- Liu, R., and Lal, R. (2015). Potentials of engineered nanoparticles as fertilizers for increasing agronomic productions. *Sci. Total Environ.* 514, 131–139. doi: 10.1016/j.scitotenv.2015.01.104
- Lowry, G., Avellan, A., and Gilbertson, L. (2019). Opportunities and challenges for nanotechnology in the agri-tech revolution. *Nat. Nanotechnol.* 14, 517–522. doi: 10.1038/s41565-019-0461-7
- Lu, W., Dulal, S., Shuguang, W., Oleg, T., Anant Kumar, S., Hongtao, Y., et al. (2008). Effect of surface coating on the toxicity of silver nanomaterials on human skin keratinocytes. *Chem. Phys. Lett.* 23, 1–7. doi: 10.1038/jid.2014.371
- Lv, J., Christie, P., and Zhang, S. (2019). Uptake, translocation, and transformation of metal-based nanoparticles in plants: recent advances and methodological challenges. *Environ. Sci. Nano* 6, 41–59. doi: 10.1039/C8EN00645H
- Martens, J. H., Barg, H., Warren, M., and Jahn, D. (2002). Microbial production of vitamin B12. *Appl. Microbiol. Biotechnol.* 58, 275–285. doi: 10.1007/s00253-001-0902-7
- Melo, L., Ng, C., Tsang, R., Singh, A. P., Kitts, D., and Lamers, Y. (2020). Development of novel Vitamin B12 fortified yogurts using isolated and microencapsulated Vitamin B12. *Proc. Nutr. Soc.* 79:2021. doi: 10.1017/s0029665120002128
- Mozafar, A., and Oertli, J. J. (1992). Uptake of a microbially-produced vitamin (B12) by soybean roots. *Plant Soil* 139, 23–30. doi: 10.1007/BF00012838
- Muthayya, S., Rah, J. H., Sugimoto, J. D., Roos, F. F., Kraemer, K., and Black, R. E. (2013). The global hidden hunger indices and maps: an advocacy tool for action. *PLoS One* 8:e67860. doi: 10.1371/journal.pone.0067860
- Nair, R., Varghese, S. H., Nair, B. G., Maekawa, T., Yoshida, Y., and Kumar, D. S. (2010). Nanoparticulate material delivery to plants. *Plant Sci.* 179, 154–163. doi: 10.1016/j.plantsci.2010.04.012
- Nakos, M., Pepelanova, I., Beutel, S., Krings, U., Berger, R. G., and Scheper, T. (2017). Isolation and analysis of vitamin B12 from plant samples. *Food Chem.* 216, 301–308. doi: 10.1016/j.foodchem.2016.08.037
- Nakos, M. (2016). *Quantitative Determination of Vitamin B 12 in Plants [Gottfried Wilhelm Leibniz Universität Hannover]*. Available online at: <https://www.repo.uni-hannover.de/bitstream/handle/123456789/8806/1024528006.pdf?sequence=1&disAllowed=y> (accessed January 5, 2021).
- National Institute of Health (2020). *Vitamin B12—Consumer*. Bethesda MD: National Institutes of Health.
- Niam, A., and Suchayo, L. (2020). Ultrasonic atomizer application for low cost aeroponic chambers (LCAC): A review. *IOP Conf. Ser. Earth Environ. Sci.* 542:012034. doi: 10.1088/1755-1315/542/1/012034
- NIH (2021). *Vitamin B12_ Fact Sheet for Health professionals*. Office of Dietary Supplements (ODS); National Institutes of Health (NIH). Bethesda MD: National Institutes of Health, 1–13.
- Oakley, G. P., and Tulchinsky, T. H. (2010). Folic acid and vitamin B12 fortification of flour: a global basic food security requirement. *Public Health Rev.* 32, 284–295. doi: 10.1007/BF03391603
- Parisi, C., Vigani, M., and Rodriguez-Cerezo, E. (2014). *Proceedings of a Workshop on "Nanotechnology for the Agricultural sector?: From Research to the Field*. Brussels: European Union, 1–40. doi: 10.2791/80497
- Pawlak, R., James, P. S., Raj, S., Cullum-Dugan, D., and Lucus, D. (2013). Understanding Vitamin B12. *Am. J. Lifestyle Med.* 7, 60–65. doi: 10.1177/1559827612450688
- Pérez-de-Luque, A. (2017). Interaction of nanomaterials with plants: what do we need for real applications in agriculture? *Front. Environ. Sci.* 5:12. doi: 10.3389/fenvs.2017.00012
- Pruksaritanond, S., Barbaryan, A., Mirrahimov, A. E., Liana, P., Ali, A. M., and Gilman, A. D. (2013). A puzzle of hemolytic anemia, iron and vitamin B12 deficiencies in a 52-year-old male. *Case Rep. Hematol.* 2013:708489. doi: 10.1155/2013/708489
- Puccinelli, M., Landi, M., Maggini, R., Pardossi, A., and Incrocci, L. (2021). Iodine biofortification of sweet basil and lettuce grown in two hydroponic systems. *Sci. Hortic.* 276:109783. doi: 10.1016/j.scienta.2020.109783
- Rameshreddy, Pavithra, G. J., Rajashekar Reddy, B. H., Salimath, M., Geetha, K. N., and Shankar, A. G. (2017). Zinc oxide nano particles increases Zn uptake, translocation in rice with positive effect on growth, yield and moisture stress tolerance. *Indian J. Plant Physiol.* 22, 287–294. doi: 10.1007/s40502-017-0303-2
- Rico, C. M., Majumdar, S., Duarte-Gardea, M., Peralta-Videa, J. R., and Gardea-Torresdey, J. L. (2011). Interaction of nanoparticles with edible plants and their possible implications in the food chain. *J. Agric. Food Chem.* 59, 3485–3498. doi: 10.1021/jf104517j
- Ritchie, H., and Roser, M. (2017). *Micronutrient Deficiency*. In *Published online at OurWorldInData.org (Issue August 2017)*. 1. Available online at: <https://ourworldindata.org/micronutrient-deficiency#citation> (accessed December 18, 2020).
- Rizzo, G., Laganà, A. S., Rapisarda, A. M. C., La Ferrera, G. M. G., Buscema, M., Rossetti, P., et al. (2016). Vitamin B12 among vegetarians: status, assessment and supplementation. *Nutrients* 8:767. doi: 10.3390/nu8120767
- Rop, K., Karuku, G. N., Mbui, D., Njomo, N., and Michira, I. (2019). Evaluating the effects of formulated nano-NPK slow release fertilizer composite on the performance and yield of maize, kale and capsicum. *Ann. Agric. Sci.* 64, 9–19. doi: 10.1016/j.aos.2019.05.010
- Rouphael, Y., and Kyriacou, M. C. (2018). Enhancing quality of fresh vegetables through salinity eustress and biofortification applications facilitated

- by soilless cultivation. *Front. Plant Sci.* 9:1254. doi: 10.3389/fpls.2018.01254
- Ruel-Bergeron, J. C., Stevens, G. A., Sugimoto, J. D., Roos, F. F., Ezzati, M., Black, R. E., et al. (2015). Global update and trends of hidden hunger, 1995–2011: the hidden hunger Index. *PLoS One* 10:e0143497. doi: 10.1371/journal.pone.0143497
- Rusher, D. R., and Pawlak, R. (2013). A review of 89 published case studies of vitamin B12 deficiency. *J. Hum. Nutr. Food Sci.* 1:1008.
- Sanchez, H., Albala, C., Lera, L., Dangour, A. D., and Uauy, R. (2013). Effectiveness of the national program of complementary feeding for older adults in Chile on vitamin B12 status in older adults; secondary outcome analysis from the CENEX Study (ISRCTN48153354). *Nutr. J.* 12:124. doi: 10.1186/1475-2891-12-124
- Sato, K., Kudo, Y., and Muramatsu, K. (2004). Incorporation of a high level of vitamin B12 into a vegetable, kaiware daikon (Japanese radish sprout), by the absorption from its seeds. *Biochim. Biophys. Acta Gen. Subj.* 1672, 135–137. doi: 10.1016/j.bbagen.2004.03.011
- Schwab, F., Zhai, G., Kern, M., Turner, A., Schnoor, J. L., and Wiesner, M. R. (2016). Barriers, pathways and processes for uptake, translocation and accumulation of nanomaterials in plants—critical review. *Nanotoxicology* 10, 257–278. doi: 10.3109/17435390.2015.1048326
- Seaman, C. (2017). *Investigation of Nutrient Solutions for the Hydroponic Growth of Plants Doctoral*. Sheffield: Sheffield Hallam University.
- Servin, A. D., and White, J. C. (2016). Nanotechnology in agriculture: next steps for understanding engineered nanoparticle exposure and risk. *NanoImpact* 1, 9–12. doi: 10.1016/j.impact.2015.12.002
- Shang, Y., Hasan, M. K., Ahammed, G. J., Li, M., Yin, H., and Zhou, J. (2019). Applications of nanotechnology in plant growth and crop protection: a review. *Molecules* 24:2558. doi: 10.3390/molecules24142558
- Shankamma, K., Yallappa, S., Shivanna, M. B., and Manjanna, J. (2016). Fe₂O₃ magnetic nanoparticles to enhance *S. lycopersicum* (tomato) plant growth and their biomining. *Appl. Nanosci. (Switzerland)* 6, 983–990. doi: 10.1007/s13204-015-0510-y
- Singh Sekhon, B. (2014). Nanotechnology in agri-food production: an overview. *Nanotechnol. Sci. Appl.* 7, 31–53. doi: 10.2147/NSA.S39406
- Smith, M. E., Coffin, A. B., Miller, D. L., and Popper, A. N. (2006). Guidelines on food fortification with micronutrients. *J. Exp. Biol.* 209, 4193–4202. doi: 10.1242/jeb.02490
- Stabler, S. (2020). “Present knowledge in nutrition,” in *Present Knowledge in Nutrition*, 11th Edn, Vol. Volume 1, eds B. P. Marriott, D. F. Birt, V. A. Stallings, and A. A. Yates (Washington, DC: International Life Sciences Institute (ILSI)), 11. doi: 10.1016/c2018-0-02422-6
- Stabler, S. P. (2013). Clinical practice. Vitamin B12 deficiency. *N. Engl. J. Med.* 368, 149–160. doi: 10.1056/NEJMcp1113996
- Surendran, S., Adaikalakoteswari, A., Saravanan, P., Shatwan, I. A., Lovegrove, J. A., and Vimalaswaran, K. S. (2018). An update on vitamin B12-related gene polymorphisms and B12 status. *Genes Nutr.* 13, 1–35. doi: 10.1186/s12263-018-0591-9
- Tandon, P., Jin, Q., and Huang, L. (2017). A promising approach to enhance microalgae productivity by exogenous supply of vitamins. *Microb. Cell Factories* 16:219. doi: 10.1186/s12934-017-0834-2
- Thompson, B., and Amoroso, L. (2010). “Combating micronutrient deficiencies: food-based approaches,” in *Combating Micronutrient Deficiencies: Food-based Approaches*, eds B. Thompson and L. Amoroso (Rome: Food and Agriculture Organisation of the United Nations; CAB International), doi: 10.1079/9781845937140.0000
- Titcomb, T. J., and Tanumihardjo, S. A. (2019). Global concerns with B vitamin statuses: biofortification, fortification, hidden hunger, interactions, and toxicity. *Compr. Rev. Food Sci. Food Safe.* 18, 1968–1984. doi: 10.1111/1541-4337.12491
- Trujillo-Reyes, J., Majumdar, S., Botez, C. E., Peralta-Videa, J. R., and Gardea-Torresdey, J. L. (2014). Exposure studies of core-shell Fe/Fe₃O₄ and Cu/CuO NPs to lettuce (*Lactuca sativa*) plants: are they a potential physiological and nutritional hazard? *J. Hazard. Mater.* 267, 255–263. doi: 10.1016/j.jhazmat.2013.11.067
- Tucker, K. L., Olson, B., Bakun, P., Dallal, G. E., Selhub, J., and Rosenberg, I. H. (2004). Breakfast cereal fortified with folic acid, vitamin B-6, and vitamin B-12 increases vitamin concentrations and reduces homocysteine concentrations: a randomized trial. *Am. J. Clin. Nutr.* 79, 805–811. doi: 10.1093/ajcn/79.5.805
- Watanabe, F. (2007). Vitamin B12 sources and bioavailability. *Exp. Biol. Med.* 232, 1266–1274. doi: 10.3181/0703-MR-67
- Watanabe, F., and Bito, T. (2018). Vitamin B12 sources and microbial interaction. *Exp. Biol. Med.* 243, 148–158. doi: 10.1177/1535370217746612
- Watanabe, F., Schwarz, J., Takenaka, S., Miyamoto, E., Ohishi, N., Nelle, E., et al. (2012). Characterization of vitamin B12 compounds in the wild edible mushrooms black trumpet (*Craterellus cornucopioides*) and golden chanterelle (*Cantharellus cibarius*). *J. Nutr. Sci. Vitaminol.* 58, 438–441. doi: 10.3177/jnsv.58.438
- Watanabe, F., Yabuta, Y., Bito, T., and Teng, F. (2014). Vitamin B12-containing plant food sources for vegetarians. *Nutrients* 6, 1861–1873. doi: 10.3390/nu6051861
- Watanabe, F., Yabuta, Y., Tanioka, Y., and Bito, T. (2013). Biologically active vitamin B12 compounds in foods for preventing deficiency among vegetarians and elderly subjects. *J. Agric. Food Chem.* 61, 6769–6775. doi: 10.1021/jf401545z
- Watanabe, S., Ohtani, Y., Tatsukami, Y., Aoki, W., Amemiya, T., Sukekiyo, Y., et al. (2017). Folate biofortification in hydroponically cultivated spinach by the addition of phenylalanine. *J. Agric. Food Chem.* 65, 4605–4610. doi: 10.1021/acs.jafc.7b01375
- WHO (2020). *Fact Sheets: Malnutrition. Fact Sheets: Malnutrition*. Geneva: WHO.
- Wolkers – Rooijackers, J. C. M., Endika, M. F., and Smid, E. J. (2018). Enhancing vitamin B12 in lupin tempeh by in situ fortification. *LWT* 96, 513–518. doi: 10.1016/j.lwt.2018.05.062
- Xie, C., Coda, R., Chamlagain, B., Edelmann, M., Varmanen, P., Piironen, V., et al. (2021). Fermentation of cereal, pseudo-cereal and legume materials with *Propionibacterium freudenreichii* and *Levilactobacillus brevis* for vitamin B12 fortification. *LWT* 137:110431. doi: 10.1016/j.lwt.2020.110431
- Zhou, L., Cai, D., He, L., Zhong, N., Yu, M., Zhang, X., et al. (2015). Fabrication of a high-performance fertilizer to control the loss of water and nutrient using micro/nano networks. *ACS Sustain. Chem. Eng.* 3, 645–653. doi: 10.1021/acsuschemeng.5b00072

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Oh, Cave and Lu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.