INTERNATIONAL SURVEY OF CRONOBACTER SAKAZAKII AND OTHER CRONOBACTER SPP. IN FOLLOW UP FORMULAS AND INFANT FOODS.

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1 Abstract

2 A coordinated survey for *Cronobacter* and related organisms in powdered infant formula, follow up formula and infant foods was undertaken by 8 laboratories in 7 3 4 countries in recognition of and in response to the data needs identified in an FAO/WHO call for data in order to develop global risk management guidance for 5 these products. The products (domestic and imported) were purchased from the 6 local market and were categorised according to their principle ingredients. A total of 7 8 290 products were analysed using a standardised procedure of pre-enrichment in 225 ml Buffered Peptone Water (BPW), followed by enrichment in 9 Enterobacteriaceae Enrichment (EE) broth, plating on the chromogenic Cronobacter 10 11 Druggan-Forsythe-Iversen (DFI) agar and presumptive identification with ID 32 E. Presumptive Cronobacter isolates were identified using 16S rRNA gene sequence 12 analysis. Aerobic plate counts (APC) of the products were also determined on 13 nutrient agar. Fourteen samples had APC >10⁵ cfu/g, 3 of which contained probiotic 14 cultures. C. sakazakii was isolated from 27 products; 3/91 (3%) follow up formulas 15 16 (as defined by Codex Alimentarius Commission), and 24/199 (12%) infant foods and drinks. Hence C. sakazakii was less prevalent in follow up formula than other foods 17 given to infants over the same age range. A range of other bacteria were also 18 isolated from follow up formulas, including Acinetobacter baumannii, Enterobacter 19 cloacae, Klebsiella pneumoniae, Citrobacter freundii, and Serratia ficaria. There was 20 significant variation in the reconstitution instructions for follow up formulas. These 21 22 included using water at temperatures which would enable bacterial growth. Additionally, the definition of follow up formula varied between countries. 23

1 1. Introduction

Cronobacter is a recently defined genus comprising of at least five species, and was 2 previously known as Enterobacter sakazakii (Iversen et al., 2008). They are motile 3 peritrichous Gram-negative rod-shaped non-spore forming bacteria, which belong to 4 5 the Enterobacteriaceae family. Cronobacter species have been implicated in neonatal intensive care unit outbreaks of meningitis, septicaemia and necrotizing 6 enterocolitis (van Acker et al., 2001; Himelright et al., 2002; Caubilla-Barron et al., 7 8 2007). Bowen and Braden (2006) considered 46 cases of Cronobacter infections in neonates. They reported that the symptoms of very low birth weight neonates (age of 9 onset ca. 1 month) tend to be bacteraemia, whereas those of birth weight ca. 2000g 10 suffered from meningitis and an onset age of a few days. Cronobacter species are 11 frequently isolated from the environment, plant material (wheat, rice, herbs and 12 spices) and various other food products (Iversen and Forsythe, 2003 & 2004; 13 Friedemann, 2007; Shaker et al., 2007; Osaili and Forsythe, 2009). The 14 microbiological safety of powdered infant formula (PIF) is a major concern to 15 regulatory agencies and formula producers as their intended use includes newborn 16 infants who have undeveloped immune systems and lack a competing intestinal flora 17 (Townsend and Forsythe 2008). Subsequently the control of the organism in these 18 products has been studied intensively, and various detection methods have been 19 developed (Cordlier, 2008; Fanning and Forsythe, 2008). It is known that starches 20 from wheat and rice can be a source of Cronobacter and are PIF ingredients 21 (FAO/WHO, 2004 & 2006). Muytjens et al. (1988) analysed 141 PIF samples from 22 35 countries and reported that 14% contained 'E. sakazakii'. It should be noted that 23 recent re-identification of the strains using 16S rDNA sequence analysis, has 24

revealed that while most were C. sakazakii, some were E. hormaechei (Townsend et 1 al., 2008). Iversen and Forsythe (2004) surveyed 486 PIF and other food products 2 for the presence of Cronobacter (then E. sakazakii). They isolated the bacterium 3 from 2/82 (2.4%) infant formulas, 5/49 (10.2%) infant weaning foods as well as 4 40/122 (37.8%) herbs and spices. More recently, Restaino et al. (2006) isolated 5 Cronobacter from 2/6 dried infant cereals, and Shaker et al. (2007) isolated the 6 7 organism from 2/8 infant formulas, and 2/15 infant wheat-based follow up formulas. Osaili et al. (2009) reported that C. sakazakii and C. muytjensii grew in infant wheat-8 based formulas whether they were reconstituted with water, UHT milk, pasteurized 9 grape or apple juices. The *Cronobacter* grew more (>5 log₁₀) in formulas 10 reconstituted with water or milk than those prepared with grape or apple juices (ca. 11 12 $2-3 \log_{10}$).

Follow up formula (also known as 'follow on formulas') are defined as 'a food 13 intended for use as a liquid part of the weaning diet for the infant from the 6th month 14 on and for young children' (CAC 1987, CCFH 2009). Similar to PIF, follow up 15 formulas are non-sterile products. The microbiological criteria for PIF used to be 16 based on the 1979 Codex Alimentarius Commission (CAC) guidelines which covered 17 infant formulas for intended use up to 12 months. These criteria have recently been 18 amended (CAC, 2008 a & b). However the issue of whether criteria for Cronobacter 19 20 spp. were necessary for the follow up formulas, which by definition only form part of an infant's diet, had not been decided. This issue was the focus of the FAO/WHO 21 (2008) risk assessment which also reviewed the known cases Cronobacter infections 22 which have been reported in infants >6 months in age. It is of particular interest, that 23 the well publicized Tennessee outbreak of C. sakazakii on a neonatal intensive unit 24

in which one neonate died was attributed to the accidental feeding of a non-infant
formula. This product was marketed for children and adults and was not intended for
use with neonates or infants (Himelright et al., 2002).

The objective of this study was to survey and compare a wide range of infant 4 5 follow up formulas and weaning foods for Cronobacter spp. using a specific chromogenic agar (Iversen et al., 2004) and general microbial load (aerobic plate 6 count) using laboratories in number of countries. The products were chosen 7 according the general description of 'follow up formulas' in each country with an 8 intended use by infants aged between 6-12 months. However, it was subsequently 9 found that many products did not meet the CAC definition of 'follow up formula' 10 especially in that they were not consumed as a liquid (CAC, 1987). In this paper, the 11 CAC definition of 'follow up formulas' has been applied and other products are 12 referred to as 'infant foods'. The latter may also be called 'weaning foods' in some 13 countries and are commonly cereal-based products. 14

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16 **2. Materials and Methods**

17 2.1. Participants and sample collection

Eight laboratories participated in seven countries; Brazil, Indonesia, Jordan (2 independent laboratories), Korea, Malaysia, Portugal, and UK. One hundred and thirty-six follow up formulas and 179 other infant foods were purchased from local markets. These for primarily products with intended use of infants aged 6-12 months, and were both imported and domestically produced. Three herbal tea samples were included in the survey as these are given to infants as drinks. The list of ingredients
and preparation instructions were recorded for each sample.

3 2.2. Microbiological analysis

The microbiological analysis was limited to the aerobic plate count (APC), and 4 5 Cronobacter detection using standardised methods for all participants. For APC 6 enumeration, 25 gram samples were aseptically taken from each product and mixed with 225 ml Buffered Peptone Water (BPW; Oxoid Thermo Fisher, UK). After 10 7 8 minutes, 1 ml aliquots were aseptically removed, decimally diluted in sterile saline, 9 and plated on Nutrient Agar (Oxoid Thermo Fisher) using the spread plate technique (0.1ml volume). The plates were incubated overnight at 37°C. Discrete colonies 10 were enumerated to determine the aerobic plate count. The remaining BPW-11 sample mixture was incubated at 37°C, overnight as a pre-enrichment step. After 12 incubation, a 10 ml aliquot was transferred to 90ml Enterobacteriaceae Enrichment 13 14 (EE) broth (Oxoid Thermo Fisher, UK), and after a further overnight incubation at 37°C, the broth was streaked on the Brilliance Enterobacter sakazakii chromogenic 15 DFI agar (Oxoid Thermo Fisher, UK). The plates were incubated at 37°C, for 18 16 hours. 17

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19 2.3. Identification of isolates

Presumptive *Cronobacter* colonies (blue-green colouration) were picked from DFI
 agar plates for phenotypic identification using ID 32 E (bioMerieux), and were
 speciated using 16S rDNA sequence analysis (Accugenix, Delaware, USA) and

phenotyping. Other non-*Cronobacter* colonies were also picked for phenotypic
 identification using ID 32 E.

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5 3. Results

In total, 318 products were sampled in seven countries by eight laboratories,
comprising 136 follow up formulas (according to CAC 2008a definition), and 182
other products. *C. sakazakii* was isolated from 1/136 (1%) follow up formulas, and
22/179 (12%) of infant foods, and none of the three herbal teas. Table 1
summarises the results of the survey for aerobic plates counts, and incidence of *Cronobacter* spp. for each country.

Cronobacter spp. were not isolated from any products in Brazil, Korea or 12 Malaysia (Table 1). Brazil analysed 31 follow up formulas. One sample contained 13 aerobic plate count 10^2 - 10^3 range, all the others were < 10^2 cfu/g. Other organisms 14 isolated included Acinetobacter baumannii, E. amnigenus, E. cloacae, and Pantoea 15 spp. All products had labels informing the user that the product should be 16 reconstituted with water that had been boiled, but not the specific temperature of 17 rehydration. Korea did not detect any Cronobacter in the 30 products sampled; 24 18 follow up formulas, and 6 infant foods. High aerobic plate counts (>10⁵ cfu/g) were 19 obtained with cereal-based infant foods. All products had labels advising the use of 20 water >70°C for reconstituting. Malaysia found high APC values for 7/12 follow up 21 formulas; four were 10^4 - 10^5 cfu/g and three were > 10^5 cfu/g. One sample with APC 22 >10⁵ cfu/g contained a probiotic culture. The packaging instructions for 7 follow up 23

formulas advised reconstituting with water that was 40-45°C, one stated 'lukewarm'
water and the remainder advised 50-55°C. The infant foods also had a wide range
of APC, with 4 having values >10⁵ cfu/g.

C. sakazakii was isolated from infant foods, but not follow up formulas in the 4 UK, Indonesia, and Portugal (Table 1). The UK isolated C. sakazakii from 6/64 of 5 6 infant foods, and these products had instructions advising the use of warm or cold 7 milk for reconstitution. Other organisms isolated included Aeromonas sobria, K. 8 pneumoniae, Pantoea spp., E. cloacae, Stenotrophomonas maltophila, A. baumannii, Ps. oryzihabitans, Citrobacter amalonaticus, and Esch. vulneris. The 9 APC ranged from $<10^2$ to $>10^5$ cfu/g for 55/64 and 2/64 infant foods, respectively. 10 11 Indonesia recovered C. sakazakii from 6/15 infant foods. A range of other organisms were also isolated; K. pneumoniae, K. terrigena, Pantoea spp., Esch. vulneris, S. 12 ficaria, S. plymuthycia, S. rubidaea, E. cloacae, and Cirobacter freundii. For 13 Portugal, the packaging instructions for two Cronobacter positive samples advised 14 the use of warm milk (50°C) for reconstitution. Leclercia adecarboxylata and E. 15 *helveticus* were also isolated from infant foods. The APC were primarily $<10^2$ cfu/g, 16 with 8/30 in the range 10²-10³ cfu/g. C. sakazakii was isolated from a transition 17 infant milk formula. This is a product for special medical purposes, and is subject to 18 the same microbiological criteria as PIF by CAC (2008b). The instructions advised 19 reconstituting with water that had been boiled for 5 minutes, and had cooled to 40°C. 20 The APC of this product was 23 cfu/g. 21

Jordan analysed 11 follow up formulas, 46 infant foods, and 3 herbal teas which are given to infants >4 months. It was the only country to isolate *Cronobacter* spp. from a follow up formula sample (Table 1). No other Enterobacteriaceae were

isolated from this sample, and the APC was $<10^2$ cfu/g. The instructions on the 1 packaging advised that reconstitution should use water which had been boiled for 5 2 minutes, and allowed to cool to an unspecified temperature. C. sakazakii was also 3 isolated from 7/46 infant foods, and the highest APC was $>10^5$ cfu/g. Other 4 5 Enterobacteriaceae isolated were E. hormaechei subsp. steigerwaltii and E. helveticus. The labelling on infant food products advised using boiled water, but gave 6 no specific temperature for reconstitution. No Cronobacter were isolated from the 7 herbal teas. These had APC $<10^2$ cfu/g. 8

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11 4. Discussion

This study was focused on follow up formula and other foods given to infants >6 12 months in response to the FAO/WHO (2008) call for appropriate microbiological 13 data. A major advantage of our study was the use of laboratories in seven countries 14 which sampled imported and domestically-produced formulas in local retail, and 15 16 therefore a wide range of products were analysed. However on compilation of the data, the study revealed that the term 'follow up formula' varied between countries. 17 For consistency, the description of follow up formulas from CAC (1987, 2009) was 18 followed, that is products with the intended age >6 months which are part of the 19 infant's diet and consumed as a liquid. Consequently some products which had 20 been were purchased as follow up formulas were re-categorised as 'infant foods' in 21 Table 1. 22

C. sakazakii was the only *Cronobacter* species isolated from any products. It
was isolated from 1/136 (0.7%) follow up formulas, and 22/182 (12%) other products.
This is the first large study of follow up formulas and shows the organism to be
present at a lower frequency than infant foods which are given to the same age
range. Previously it was shown that *Cronobacter* spp. were present in 5/49 (10%)
infant foods (Iversen and Forsythe, 2004).

Since the FAO/WHO (2006) and WHO (2007) recommended the use of water
>70°C for reconstitution of powdered infant formula, it was of interest to record the
guidance instructions on the follow up formula packaging. The only country which
clearly stated the use of water >70°C for reconstitution was Korea. The advice given
in other countries varied considerably including non-specified temperature
(lukewarm) and as low as 40°C which is within the growth range of *Cronobacter* spp.
and other Enterobacteriaceae.

In addition, other bacteria which the FAO/WHO (2004) listed as Category B
(causality plausible-but not yet demonstrated) were also isolated from follow up
formulas and infant foods; *E. cloacae*, *K. pneumoniae*, *C. freundii*, *Esch. vulneris*, *Pantoea* spp., *S. ficaria* and *A. baumannii*. These opportunistic pathogens would not
have been killed by the mild temperatures used on reconstitution, and could multiply
if the feed was left at ambient temperature.

The APC of the follow up formulas ranged from $<10^{2}$ for 102/136 (75%) samples to $>10^{5}$ cfu/g for 3/136 (2%). These can be compared with 103/179 (58 %) and 11/179 (6%) for infant foods. A similar APC range was reported by Iversen and Forsythe (2004) for infant foods; 35/49 (71%) $<10^{2}$ cfu/g, and 3/49 (6%) 10^{4} - 10^{5} cfu/g. The latter data is included in Table 1 for comparative purposes of samples

taken 4 years previously in the UK. In the current study, 7 products contained 1 probiotics, three of which gave very high APC (>10⁵ cfu/g). Whether the presence of 2 these organisms affects the detection of *Cronobacter* is unknown and is a matter 3 warranting further investigation. It is important to note that Joosten et al (2006) 4 reported the need to modify the Salmonella detection method due the presence of 5 probiotic cultures. The three herbal tea samples were included in the survey, as 6 7 these are also given to infants (>4 months) and Cronobacter have been isolated from similar products (Tamura et al., 1995; Friedemann 2007; Osaili and Forsythe, 2009). 8 These products are prepared with boiled water which is cooled to about 50°C. 9

Infants >6 months are exposed to a wider range of sources of micro-10 organisms (environmental and foodstuffs) than neonates, who are more susceptible 11 to infection due to the lack of a developed immune system and intestinal flora 12 (Townsend and Forsythe, 2008). Nevertheless infants are still prone to infections 13 and general good hygienic practices in food preparation are necessary to reduce this 14 risk. This survey has shown the presence of C. sakazakii in follow up formulas and 15 infants foods. In addition, it has shown that the packaging instructions are 16 inconsistent with regard to the temperature of reconstitution, which is a known step 17 in reducing bacterial load. Cronobacter spp. infections have been reported for all 18 age groups. Worldwide there have been ca. 120 reported cases in infants and 19 children <3 years in age, of which 8 were cases aged between 6-35 months 20 (FAO/WHO, 2008). In the UK between 1999-2007, 15/570 laboratory reported 21 infections were from infants (<12 months in age), and 16/570 were from children (1-4 22 years in age) (FAO/WHO, 2008). Previous surveys of PIF have shown the incidence 23 of Cronobacter spp. to be between 2-22 % (FAO/WHO 2006). In contrast, this 24

international study has shown that the incidence of *Cronobacter* spp. is lower (0.7%)
in follow up formula. Since follow up formula is only part of the infant's diet, this data
supports the recent CAC (2009) decision for microbiological criteria for follow up
formula to include *Salmonella* and not *Cronobacter* spp. Nevertheless good hygienic
practices are always required in the preparation of such foods.

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14 **References**

15	Bowen, A.B., Braden, C.R., 2006. Invasive Enterobacter sakazakii disease in
16	infants. Emerging Infectious Diseases. Date last accessed 08/05/09.
17	Caubilla-Barron, J., Hurrell, E., Townsend, S., Cheetham, P., Loc-Carrillo, C., Fayet,
18	O., Prere, MF., Forsythe, S.J., 2007. Genotypic and phenotypic analysis of
19	Enterobacter sakazakii strains from an outbreak resulting in fatalities in a

1	neonatal intensive care unit in France. Journal of Clinical Microbiology 45,
2	3979-3985.

- 3 Codex Alimentarius Commission (CAC), 1979. Recommended international code of
- 4 hygienic practice for foods for infants and children. CAC/ RCP-21. Alinorm
- 5 79/38. Codex Alimentarius Commission, Rome.
- 6 Codex Alimentarius Commission (CAC), 1987. Codex standard for follow-up formula.
- 7 Codex Stan 156-1987. Available at
- 8 http://www.codexalimentarius.net/download/standards/293/CXS_156e.pdf.
- 9 Last accessed 08/05/09.
- 10 Codex Alimentarius Commission (CAC), 2008a. Report of the thirty-first session of
- 11 the Codex Alimentarius Commission. Geneva, Switzerland, 30 June 4 July.
- 12 Alinorm 08/31/REP. Available at:
- ftp://ftp.fao.org/codex/Alinorm08/al31REP_adv.pdf. Date last accessed
 08/05/09.
- 15 Codex Alimentarius Commission (CAC), 2008b. Code of hygienic practice for
- powdered formulae for infants and young children. CAC/RCP 66-2008.
- 17 http://www.codexalimentarius.net/download/standards/11026/cxp_066^e.pdf.
- 18 Date last accessed 08/05/09.
- 19 Codex Alimentarius Commission (CAC), 2009. Microbiological criteria for powdered
- 20 follow-up formulae and formulae for special medical purposes for young
- 21 children (Annex II to the Code of hygienic practice for powdered formulae for
- infants and young children (CAC/RCP 66-2008) at Step 5/8 (ALINORM
- 23 09/32/13 paras 45-47 and Appendix III). CL 2009/1-FH. ALINORM 09/32/13.

1	http://www.codexalimentarius.net/download/report/714/a132_13e.pdf. Date
2	last accessed 08/05/09.
3	Cordier, J.L, 2008. Production of powdered infant formulae and microbiological
4	control measures. In: J. Farber, Forsythe, S.J. (Eds.), Enterobacter
5	sakazakii. ASM Press, Washington, DC, pp. 145-185.
6	Fanning, S., Forsythe, S.J., 2008. Isolation and identification of Enterobacter
7	sakazakii. In: J. Farber, Forsythe, S.J. (Eds.), Enterobacter sakazakii. ASM
8	Press, Washington, DC, pp. 27-59.
9	Food and Agriculture Organization/World Health Organization (FAO/WHO), 2004.
10	Enterobacter sakazakii and other microorganisms in powdered infant formula.
11	Meeting report, MRA series 6. World Health Organization, Geneva,
12	Switzerland. Available at
13	http://www.who.int/foodsafety/publications/micro/mra6/en/index.html. Date last
14	accessed 08/05/09.
15	Food and Agriculture Organization/World Health Organization (FAO/WHO), 2006.
16	Enterobacter sakazakii and Salmonella in powdered infant formula. Second
17	risk assessment workshop. Meeting report, MRA series 10. World Health
18	Organization, Geneva, Switzerland. Available at
19	http://www.who.int/foodsafety/publications/micro/mra10/en/index.html. Date

2	Food and Agriculture Organization/World Health Organization (FAO/WHO), 2008.
3	Enterobacter sakazakii (Cronobacter spp.) in powdered follow-up formulae.
4	MRA series. Available at
5	http://www.who.int/foodsafety/publications/micro/MRA_followup.pdf. Date
6	last accessed 08/05/09.
7	Friedemann, M., 2007. Enterobacter sakazakii in food and beverages (other than
8	infant formula and milk powder). International Journal of Food Microbiology.
9	116, 1–10.
10	Himelright, I., Harris, E., Lorch, V., Anderson M., 2002. Enterobacter sakazakii
11	infections associated with the use of powdered infant formula -Tennessee,
12	2001. The Journal of the American Medical Association 287, 2204–2205.
13	Iversen, C., Forsythe, S.J., 2003. Risk profile of Enterobacter sakazakii, an
14	emergent pathogen associated with infant milk formula. Trends in Food
15	Science and Technology. 11, 443-454.
16	Iversen, C., Forsythe, S.J., 2004. Isolation of Enterobacter sakazakii and other
17	Enterobacteriaceae from powdered infant formula milk and related products
18	Food Microbiology 21, 771–776.
19	Iversen, C., Druggan, P., Forsythe, S.J., 2004. A selective differential medium for

Enterobacter sakazakii. International Journal of Food Microbiology. 96, 133–
 139.

3	Iversen, C., Mullane, N., McCardell, B. D., Tall, B.D., Lehner, A., Fanning, S.,
4	Stephan, R. and Joosten, H., 2008. Cronobacter gen. nov., a new genus to
5	accommodate the biogroups of <i>Enterobacter sakazakii</i> , and proposal of
6	Cronobacter sakazakii gen. nov., comb. nov., Cronobacter malonaticus sp.
7	nov., Cronobacter turicensis sp. nov., Cronobacter muytjensii sp. nov.,
8	Cronobacter dublinensis sp. nov., Cronobacter genomospecies 1, and of
9	three subspecies, Cronobacter dublinensis subsp. dublinensis subsp. nov.,
10	Cronobacter dublinensis subsp. lausannensis subsp. nov. and Cronobacter
11	dublinensis subsp. lactaridi subsp. nov. International Journal of Systematic
12	and Evolutionary Microbiology 58, 1442–1447.
13	Joosten H, Bidlas E, Garofalo N., 2006. Salmonella detection in probiotic products.
14	International Journal of Food Microbiology110,104-7.
15	Muytjens, H. L., Roelofs-Willemse, H., Jaspar, G.H., 1988. Quality of powdered
16	substitutes for breast milk with regard to members of the family
17	Enterobacteriaceae. Journal of Clinical Microbiology 26, 743–746.
18	Osaili, T.M., Forsythe, S.J., 2009. Desiccation and persistence of Cronobacter
19	species in infant formula. International Journal of Food Microbiology (In
20	submission)
21	Osaili, T.M., Shaker, R.R., Ayyash, M. M., Al-Nabulsi, A. A., Forsythe, S. J., 2009.

1	Survival and growth of Cronobacter species (Enterobacter sakazakii) in
2	wheat-based infant follow on formulas. Letters in Applied Microbiology
3	48, 408-412.
4	Restaino, L., Frampton, E.W., Lionberg, W.C., Becker, R.J., 2006. A chromogenic
5	plating medium for the isolation and identification of Enterobacter sakazakii
6	from foods, food ingredients, and environmental sources. Journal of Food
7	Protection 69, 315–322.
8	Shaker, R., Osaili, T. Al-Omary, W., Jaradat, Z., Al-Zuby, M., 2007. Isolation of
9	Enterobacter sakazakii and other Enterobacter sp. from food and food
10	production environments. Food Control 18, 1241–1245
11	Tamura, A., Kato, M., Omori, M., Nanba, A., Miyagawa, K., Wang, C. R., Zhou, W.
12	H., 1995. Flavor components and microorganisms isolated from Suancha
13	(sour tea, Takeutsu-sancha in Japanese). Nippon Kasei Gakkaishi 46, 759–
14	764.
15	Townsend, S., Forsythe, S.J., 2008. The neonatal intestinal microbial flora,
16	immunity, and infections. In: J. Farber, Forsythe, S.J. (Eds.)
17	Enterobacter sakazakii. ASM Press. Washington, DC, pp. 61-100.
18	Townsend, S.M., Hurrell, E., Caubilla-Barron, J., Loc-Carrillo, C., Forsythe, S.J.,
19	2008. Characterization of an extended-spectrum beta-lactamase
20	Enterobacter hormaechei nosocomial outbreak, and other Enterobacter

1	hormaechei misidentified as Cronobacter (Enterobacter) sakazakii.
2	Microbiology 154, 3659-3667.
3	van Acker, J., de Smet, F., Muyldermans, G., Bougatef, A., Naessens, A.,
4	Lauwers, S., 2001. Outbreak of necrotizing enterocolitis associated with
5	Enterobacter sakazakii in powdered milk formula. Journal of Clinical
6	Microbiology 39, 293-297.
7	World Health Organisation (WHO), 2007. Safe preparation, storage and handling of
8	powdered infant formula guidelines. Available at
9	http://www.who.int/foodsafety/publications/micro/pif2007/en/index.html.
10	Date last accessed 08/05/09.

	······	Number of samples	Aerobic plate counts (cfu/g)					Cronobactor positivo
Product	Country		<10 ²	10 ² -10 ³	10 ³ -10 ⁴	10 ⁴ -10 ⁵	>10 ⁵	samples (%)
Follow up formula ^a	Brazil	31	30	1				
	Korea	24	21	3				
	Malaysia	12	5(1) [°]			4	3 (1)	
	UK	38	22	12	4(1)			
	Indonesia	0	ND^{d}					
	Portugal	20	18	2				
	Jordan 1	6	6					1
	Jordan 2	5	ND					
	Total	136	102	18	4	4	3	1 (0.7)
Infant foods	Brazil	0						
	Korea	6	2				4	
	Malaysia	18	11	1	1	1(1)	4(2)	
	UK	64	55	5	2		2	6
	Indonesia	15	ND					6
	Portugal	30	23	8				3
	Jordan 1	21	12(1)	7	1		1	4
	Jordan 2	25	ND					3
	Total	179	103	21	4	1	11	22(12)
Herbal tea	Jordan 1	3	3					
Previous infant food published data ^b	UK	49	35	8	3	3		5 (10)

Table 1. Microbiological analysis of follow up formulas and infant foods.

- a. Description matches that of CAC (1987 & 2009) definition of 'follow up formulas'.
- b. Iversen and Forsythe, 2004.
- c. Number in parenthesis indicates the number of samples which contained probiotic cultures.
- d. Not done.