

**Genome sequence of the
hyperinvasive *Campylobacter jejuni*
strains**

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ABSTRACT

Campylobacter jejuni is the world's major cause of gastroenteritis in humans. Although motility, toxin production, adhesion and invasion are some of the key factors associated with *C. jejuni* pathogenesis, their mechanism in the disease process remains unclear. The key aim of this project is to study the genetic basis of hyperinvasiveness in a group of six *C. jejuni* strains which have been reported as hyperinvasive into human intestinal cell lines.

Here, genotyping of the hyperinvasive *C. jejuni* was performed by comparative genomic hybridization (CGH) against four low invasive *C. jejuni* strains. A group of 67 genes were identified as being present or highly divergent/absent in the hyperinvasive versus low invasive *C. jejuni* strains. Of these, nine genes were present and six genes were highly divergent/absent in all hyperinvasive *C. jejuni*. The PCR screening of these 15 genes in nine additional low invasive *C. jejuni* strains showed a significant association with the hyperinvasive phenotype. The majority of identified genes encoded proteins with essential cellular and metabolic functions along with some genes with known virulence related roles. Thus, the hyperinvasive phenotype is characterised by different functional networks rather than a single gene or gene cluster. All strains showed an overall genetic variability and the capsule, lipooligosaccharide, flagellar biosynthesis and restriction modification regions were the most diverse. The hierarchical clustering based on comparative genomic hybridization (CGH) did not group together the hyperinvasive *C. jejuni* as a single group and these strains possessed different MLST profiles.

The hyperinvasive *C. jejuni* strains were shown to contain additional genetic content by pooled suppressive subtractive hybridization (PSSH). Eleven inserts were identified in total which were variably distributed in the hyperinvasive *C. jejuni* strains. Of these four sequences were specific to the hyperinvasive *C. jejuni* as these were absent from all thirteen low invasive *C. jejuni* strains tested. The majority of sequences matched with genes in *Campylobacter* and other bacteria and one sequence had no homology with anything in the databases today. Since, there is no insert identified as present in all the hyperinvasive *C. jejuni* strains it can be suggested that each strain might have evolved a different mechanism for hyperinvasiveness and that this phenotype is a multifactorial process.

C. jejuni 01/10 and 01/51 whole genome sequences identified no unique genetic content in either strain except for a prophage in *C. jejuni* 01/51. *C. jejuni* 01/10 was found to contain two prophages. *C. jejuni* 01/51 has a highly mosaic capsule locus with genes similar to *C. jejuni* subsp. *doylei* and *C. lari* capsular polysaccharide genes. Some genes with homology to the *C. jejuni* subsp. *doylei* capsule genes were also identified in *C. jejuni* 01/10 capsule region. This is evidence of genetic recombination with capsule genes from other pathogenic *Campylobacter* species which is not reported in the capsule region of other *Campylobacter* strains sequenced to date. This suggests that the highly diverse capsule in *C. jejuni* 01/10 and 01/51 is required for the hyperinvasive phenotype in these strains.

This study has provided detailed insight into the genomic structure of the hyperinvasive *C. jejuni* strains and has highlighted genetic factors involved in their hyperinvasive phenotype.

DECLARATIONS

Section 1: This is an original piece of research work carried out by the author in the School of Science and Technology at Nottingham Trent University. The adhesion and invasion profiling of hyperinvasive and low invasive *C. jejuni* strains used in this study was originally reported by Fearnley *et al* (2008) and later re-confirmed by my predecessor post-graduate researcher, Dr Afzal Javed at NTU (Javed., 2009). The phylogeny of hyperinvasive *C. jejuni* 01/51 and 01/10 strains (chapter 6) was performed by Dr Alan McNally at NTU. There is no material contained within this thesis that has been submitted for any other degree, or at any other institution.

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LIST OF ABBREVIATIONS

ATCC	American Type Culture Collection
AFLP	Amplified fragment length polymorphism
ATP	Adenosine triphosphate
BA	Butzler agar
BB	Bolton broth
BMDCs	Bone marrow derived murine dendritic cells
bp	Base pair
CAPs	Cationic antimicrobial peptides/proteins
CDS	Coding sequence
CDT	Cytolethal distending toxin
CEB	<i>Campylobacter</i> Enrichment Broth
cfu	Colony forming unit
CGH	Comparative genomic hybridization
Cia	<i>Campylobacter</i> invasion antigen
CJIE	<i>Campylobacter jejuni</i> integrated element
CMLP	<i>Campylobacter</i> Mu like prophage
CPS/CAP	Capsular polysaccharides
cy-dCTP	Cyanine 2'-deoxycytidine 5'-triphosphate
DCs	Dendritic cells
DNA	Deoxyribonucleic acid
dNTP	2'-deoxynucleoside 5'-triphosphate
EDTA	Ethylenediamine tetra-acetic acid
ERK	Extracellular signal regulated kinase
<i>feds</i>	Flagellar coexpressed determinants
FISH	Fluorescence <i>In situ</i> Hybridization
<i>fla</i> -SVR	<i>fla</i> -amplified short variable regions
FM	Flagellar modification

GBS	Guillain Barré syndrome
G + C	Guanine and cytosine
HMW	High molecular weight
HS	Heat stable
Ig	Immunoglobulin
IL	Interleukin
IPEC	Intestinal pig epithelial cells
kbp	Kilobase pair
LB	Luria-Bertani broth
LegAm	Legionaminic acid
LOS	Lipooligosaccharide
LPS	Lipopolysaccharides
MAP	Mitogen activated protein
(m)CCDA	(modified) charcoal cefoperazone deoxycholate
MCP	Methyl-accepting chemotaxis protein
MeOPN	O-methyl phosphoramidate
MFS	Miller Fisher syndrome
MHA	Muller Hinton agar
MHB	Muller Hinton broth
MLST	Multi-locus sequence typing
MOMP	Major outer membrane protein
NCTC	National Collection of Type Cultures
NF-kB	Nuclear factor-kappa-enhancer of activated B cells
OD	Optical density
ORF	Open reading frame
<i>p</i> value	Probability value
PAMPs	Pathogen associated molecular patterns
PB	Preston broth
PBS	Phosphate buffered saline solution

PCR	Polymerase chain reaction
PFGE	Pulsed-field gel electrophoresis
PseAc	Pseudaminic acid
PSSH	Pooled suppressive subtractive hybridization
RAPD	Random amplification polymorphic DNA
RFLP	Restriction fragment length polymorphism
RM	Restriction modification
ROS	Reactive oxygen species
rpm	Revolutions per minute
rRNA	Ribosomal ribonucleic acid
TAE	Tris/acetate/EDTA
TLRs	Toll-like receptors
sIg	Secretory immunoglobulin
σ_{28}	Sigma factor 28 or fliA (promoter)
σ_{54}	Sigma factor 54 or rpoN (promoter)
SSH	Suppressive subtractive hybridization
T _m	Melting temperature
v/v	Volume per volume
w/v	Weight per volume

Chapter One

INTRODUCTION

INTRODUCTION

1.1 *Campylobacter* species in general

Campylobacter species are small Gram negative (0.5-5.0 μm long and 0.2-0.8 μm wide), spiral shaped, non-spore forming fermentative bacteria. These bacteria exhibit darting motility due to the presence of unipolar or dipolar flagella. *Campylobacter* species grow under microaerobic conditions in an atmosphere containing 3-15% CO_2 and 3-5% O_2 . *Campylobacter* species have a growth temperature range of maximum $\sim 46^\circ\text{C}$ and minimum 30°C (Davis and DiRita., 2008). Levin *et al* (2007) proposed that *Campylobacter* species should be referred to as “thermotolerant” as they do not represent characteristics of a classic thermophile with a growth temperature of 55°C . *Campylobacter* species are known to lack the glycolytic enzyme 6-phosphofructokinase, hence are unable to metabolize carbohydrates. Recently Stahl *et al* (2011) showed that *C. jejuni* is able to utilize L-fucose released by intestinal mucin glycoproteins. *Campylobacter* species use amino acids, mainly serine (Velayudhan *et al.*, 2004), and some organic acids including pyruvate and lactate (Thomas *et al.*, 2011) as carbon sources for energy production.

1.2 History of *Campylobacter* species

Campylobacter species were described as “*Vibrio*” like organisms in 1913, causing disease in bovines and ovines (McFadyean and Stockman., 1913, Vandamme *et al.*, 2010). Later Smith and Taylor (1919), isolated spiral shaped bacteria from bovine abortions. These bacteria had similar morphology as described before by McFadyean and Stockman and were named “*Vibrio fetus*” (Smith and Taylor., 1919). These *Vibrio* like bacteria from human infections were studied in detail by King (1957). She noticed that although these bacteria were “*Vibrio* like” they showed growth at much higher temperatures hence they were later called “*Vibrio jejuni*”. *Vibrio* like organisms were also found in the faeces of pigs suffering from diarrhoea and were known as *Vibrio coli* (Doyle., 1944).

The first microscopic evidence of *Campylobacter* species came from Escherich (1886) who observed spiral bacteria, simply named “*Cholera infantum*”, in the samples taken from colon of infants who died of cholera like disease as well as from stools of infants

suffering from diarrhoea (Escherich., 1886). These bacteria were considered unculturable. It was not until 1963, when Sebald and Veron classified *Campylobacter* as a separate genus based mainly on the small genome size, anaerobic growth and fermentative metabolism. *Campylobacter* were first isolated from stool samples of patients with enteritis by a filtration method (Dekeyser *et al.*, 1972). Later on, *Campylobacter jejuni* and *Campylobacter coli* were also differentiated from each other (Skirrow., 1977). Now, *C. jejuni* and *C. coli* are considered as the major enteric pathogens of humans causing disease in the developing (Coker *et al.*, 2002) and developed world (Altekruse *et al.*, 1998).

1.3 Taxonomy of *Campylobacter* species

Since the first taxonomical classification of the *Campylobacter* genus by Sebald and Veron in 1963, the genus has undergone considerable changes (On., 2001, Debruyne *et al.*, 2005). The very early classification described four species in the *Campylobacter* genus including *C. jejuni*, *C. coli*, *C. fetus* and *C. sputorum*. The diversity in the *Campylobacter* genus was first defined by studying the 16S rRNA gene sequence in these bacteria. The 16S rRNA gene is highly conserved in bacteria and is of considerable length (~1500 bp) that makes it a useful tool for phylogenomic classification (Man *et al.*, 2010). The study of 16S rRNA gene sequences, fatty acid profiles and flagellar structure defined a genus called *Helicobacter* that included the formerly named *C. pylori* and *C. mustelae* (Goodwin *et al.*, 1989). The 16S rRNA sequence divergence also classified the oral anaerobes *Wolinella curva* and *W. recta* into the *Campylobacter* genus (Vandamme *et al.*, 1991). These techniques together with other immunotyping data and whole cell protein profile classified the aero-tolerant campylobacters *A. nitrofigilis* and *A. cryoaerophilus* into another genus called *Arcobacter* (Vandamme *et al.*, 1991 and 1992). Later on, a new bacterial family called *Campylobacteraceae* was defined that included genera *Arcobacter*, *Campylobacter*, *Sulfurospirillum* and *Bacteroides ureolyticus* (Vandamme., 2000). Recently, Vandamme *et al* (2010) studied 26 *Bacteroides ureolyticus* strains using 16S rRNA and *cpn60* gene sequences, amplified fragment length polymorphism and protein profiling. Vandamme *et al* (2010) reclassified *B. ureolyticus* as a species in the *Campylobacter* genus, *Campylobacter ureolyticus*. The bacterial families Camplobacteraceae and Helicobacteraceae are included in the rRNA super-family VI and form a part of the Epsilonobacteria or Proteobacteria (Cavalier-Smith., 2002).

The *Campylobacter* genus consists of 20 species and sub-species (Fernández *et al.*, 2008). In the same year, Debruyne *et al* (2008) reported 17 species and 6 sub-species in the *Campylobacter* genus with the number of species in this genus continuously increasing.

C. jejuni, *C. coli*, *C. lari*, *C. fetus*, *C. hyointestinalis*, *C. upsaliensis*, *C. gracilis*, *C. showae* are some well-known species included in the *Campylobacter* genus (Debruyne *et al.*, 2005). *C. jejuni* contains two sub-species, *C. jejuni* subsp. *jejuni* and *C. jejuni* subsp. *doylei*. *C. jejuni* subsp. *jejuni* is present as commensal bacteria in the intestinal tract of domestic and commercial animals and it is also the most common cause of gastroenteritis in humans (Miller *et al.*, 2007) whereas *C. jejuni* subsp. *doylei* has only been isolated from human cases of septicaemia and bacteraemia (Lastovica., 2006). Other phenotypic and biochemical characteristics that distinguish *C. jejuni* subsp. *doylei* from *C. jejuni* subsp. *jejuni* include the inability of *C. jejuni* subsp. *doylei* to reduce nitrate, variable growth rate at 42 °C, absence of γ -glutamyl transferase (GGT) and L-arginine arylamidase enzymatic activity and susceptibility to cephalothin (Miller *et al.*, 2007). The two sub-species show significant sequence divergence at the genome level (Parker *et al.*, 2007). The sequence variation at the *nap* gene locus has been used to develop a multiplex PCR assay for *Campylobacter* diagnosis at the sub-species level (Miller *et al.*, 2007). Using the readily available genome sequence data for *C. jejuni* strains, Taboada *et al* (2012) has reported a highly sensitive comparative genomic fingerprinting method for *C. jejuni* sub-speciation. This method relies on identifying variability at multiple alleles widely distributed in the accessory gene pool across the whole genome (Taboada *et al.*, 2012).

1.4 Isolation and further classification of *Campylobacter* species

The first step in detection of *Campylobacter* species is their isolation from samples. Further classification is carried out by phenotyping or genotyping. Some of these methods are discussed below briefly.

1.4.1 Isolation and phenotyping

All *Campylobacter* species except for *C. gracilis* show oxidase activity. This phenotype has been used for the development of selective media containing one or more oxygen scavengers (*i.e.* blood, ferrous ions) with selective antibiotics (Corry *et al.*, 1995). A

number of selective broths including Bolton Broth (BB), *Campylobacter* Enrichment Broth (CEB) and Preston Broth (PB) with oxygen limiting enzyme oxyrase have been successfully used for the isolation of *Campylobacter* species (Abeyta *et al.*, 1997, Baylis *et al.*, 2000). The selective agars *i.e.* Preston, charcoal cefoperazone deoxycholate (CCDA) and Butzler Agar (BA) have been used efficiently for the isolation of *Campylobacter* species. The standard method used at present for the detection of *Campylobacter* species is by plating and enumeration directly on mCCDA. The Bolton Broth (BB) and Muller Hinton Broth (MHB) are used for growth in liquid culture or as an enrichment step (ISO 2006a and 2006b).

Other phenotypes differentiating *Campylobacter* species include catalase activity, growth at 25 °C or 42 °C and resistance to antibiotics including naladixic acid, cephalothin and fluoroquinolones. *C. jejuni* can hydrolyse sodium hippurate, indoxyle acetate and reduces nitrate. *C. coli* are unable to hydrolyse hippurate and it is the lack of this activity that differentiates *C. jejuni* from *C. coli*. Still there are some hippurate negative *C. jejuni* isolates which are deficient of hippurate activity (Koenraad *et al.*, 1995). *C. fetus* is the only member of *Campylobacter* species that can survive in the presence of glycine and can produce hydrogen sulphide (On., 1996). A series of biochemical tests are commercially available as rapid identification kits (e.g. Campy API kits) that can differentiate all *Campylobacter* species (Hoosain and Lastovica., 2009, Reina *et al.*, 1995). Other rapid methods that can detect and confirm *Campylobacter* species include Fluorescence *In situ* Hybridization (FISH) (Lehtola *et al.*, 2006) and latex autoagglutination (Wilma *et al.*, 1992).

Another phenotypic method used to discriminate between *Campylobacter* species is serotyping. The Penner serotyping scheme is the most acceptable and widely used phenotypic scheme for typing of *Campylobacter* species. It differentiates strains based on the soluble heat stable (HS) antigen. In Penner serotyping, a passive haemagglutination reaction is observed by mixing the bacterial cell suspension with different antisera (Penner and Hennessy., 1980). The *C. jejuni* capsular polysaccharides (Karlyshev *et al.*, 2000, Wren *et al.*, 2001) have been shown as the serodeterminant of Penner serotyping. The Penner serotyping scheme contains 48 antisera for *C. jejuni* and 15 antisera for *C. coli* and recognizes 63 serotypes (Penner *et al.*, 1983). The main drawbacks of Penner serotyping are non-typeability of a large number of *Campylobacter* strains compared to modern day genotyping methods and cross reactivity with more than

one antiserum (Cornelius *et al.*, 2010). Another method used for *Campylobacter* species serotyping is called Lior serotyping. It is based on heat labile antigens. It can differentiate between 150 serotypes of *C. jejuni*, *C. coli* and *C. lari* (Lior *et al.*, 1982).

Penner serotyping is generally combined with another typing technique called phage typing for diagnosis of *Campylobacter* species. In phage typing, the phage pattern of an isolate is compared to the profile of other virulent bacteriophage. Two or more strains with identical bacteriophage patterns form a phage type (Frost *et al.*, 1999). Serotyping and phage typing have poor resolution. These techniques have to be combined with advanced molecular typing for epidemiological and surveillance studies (Hopkins *et al.*, 2004).

1.4.2 Genotyping

Molecular typing methods are used for bacterial identification and speciation in both pure and mixed cultures. These techniques are sensitive, rapid and highly discriminatory compared to the conventional phenotypic methods. A number of genotypic methods have been developed for typing of *Campylobacter* species. The advantages and drawbacks of these techniques for *Campylobacter* species typing have been recently reviewed (Ahmed *et al.*, 2012). The pulsed-field gel electrophoresis (PFGE), ribotyping and flagellin gene typing, restricted and amplified chromosomal fragment length polymorphism (RFLP and AFLP) are a few examples of the genotyping techniques (Nielsen *et al.*, 2000).

PFGE is based on digesting the chromosomal DNA with different restriction enzymes *e.g.* *Sma*I, *Kpn*I and *Sal*I. This digestion generates DNA fragments of variable length. The variation in length of resulting fragments is compared to generate a PFGE profile for the isolate (O’Leary *et al.*, 2011, Gilpin *et al.*, 2006, Peters., 2009). The electrophoresis profiles generated for a large number of strains can be analysed by computerized software (*e.g.* BioNumerics by Applied Maths, Ghent, Belgium). PFGE is a useful technique which successfully discriminates closely related strains but it requires technical expertise (Peters., 2009, Pittenger *et al.*, 2009).

Restricted Fragment Length Polymorphism (RFLP) is a technique that detects variation in a selected locus in the genome *e.g.* *fla*, *porA*. The flagellin and MOMP are stable but highly genetically diverse regions in *Campylobacter* species (Cody *et al.*, 2009). For example, in *fla*-typing, the *fla* gene is PCR amplified and digested with one or more

restriction enzymes (*fla*-RFLP) or the PCR product is sequenced using the gene specific primers *fla*-amplified short variable regions (*fla*-SVR) (On *et al.*, 2008). Amplified fragment length polymorphism (AFLP) like RFLP involves digesting the genomic DNA with two or more restriction enzymes which are ligated to specific adaptors. These labelled fragments are then PCR amplified using fluorescent labelled primers (On *et al.*, 2008). AFLP has been used as a highly discriminatory and reproducible molecular typing technique used in epidemiological studies (Johnsen *et al.*, 2007, Siemer *et al.*, 2005). Another technique used for typing of *Campylobacter* species is Random Amplification of Polymorphic DNA (RAPD). This method involves PCR amplification of genomic regions by using one 10-15 bp primer that will produce several fragments of variable lengths in different strains as a result of primer binding at different sites (On *et al.*, 2008). Ribotyping detects variations in the ribosomal DNA genes. The genomic DNA is digested with two enzymes followed by hybridization with probed rRNA specific primers. The characteristic ribosomal pattern of each isolate is then visualised (On *et al.*, 2008). A fully automated ribotyping method is also developed now that has further improved the speed and reproducibility of this method (Pavlic and Griffiths., 2009). However, ribotyping lacks discriminatory ability for *Campylobacter* species (Pavlic and Griffiths., 2009, Ge *et al.*, 2006).

Currently, Multi-locus Sequence Typing (MLST) is frequently used as a genotyping technique to study *Campylobacter* species. MLST is based on studying the DNA sequence variation in *Campylobacter* housekeeping (usually seven) genes. Each sequenced gene is given an arbitrary allele number. The allele number for all the genes are combined to assign a unique sequence type number to each strain. *C. jejuni* has been shown to be diverse and have a weakly clonal structure (Dingle *et al.*, 2002, Manning *et al.*, 2003).

With an increasing number of genome sequences of *Campylobacter* species becoming available, a number of high-throughput genome based typing techniques have been designed. These techniques are based on studying differences between *Campylobacter* strains at the whole genome level. DNA microarray technology is also being developed as a diagnostic tool for use in epidemiological and phylogenetic studies (Marotta *et al.*, 2012, Pittenger *et al.*, 2012, Taboada *et al.*, 2012).

1.5 Campylobacteriosis: a disease burden on public health

Campylobacter species has been identified as a major zoonotic pathogen causing gastroenteritis in humans worldwide. According to the European Centre of Disease Control (ECDC) and European Foods Safety Agency (EFSA) *Campylobacter* species caused gastroenteritis was most common in the European Union (EU) followed by salmonellosis and yersiniosis in a five years period (EFSA., 2007, 2010a). There were just around 200,000 human cases of campylobacteriosis recorded in 2009 and 2010 (EFSA., 2009 and 2010a). This number represented a 14.2% rise from 2006 which is a significant burden on the economy.

The Foodborne Diseases Active Surveillance Network (FoodNet) of the Centres for Disease Control and Prevention (CDC) estimated that *Campylobacter* species cause approximately 845,000 cases of illness in the USA each year. This number is still an underestimate as a lot of cases remain unreported and undiagnosed (Anonymous., 2010).

Similarly, the number of human cases of campylobacteriosis in the EU is not an actual representation of true disease burden implicated by *Campylobacter* species. It is estimated that in EU approximately 2 to 20 million people may get campylobacteriosis per year (EFSA., 2010b). *Campylobacter* species are the most common cause of foodborne outbreaks in the UK. There were approximately 321,000 human cases of *Campylobacter* food poisoning, over 15,000 hospitalizations and 76 deaths in England and Wales in 2008. The food poisoning caused by *Campylobacter* species costs £583 million which represents one third of the cost of foodborne illness in England and Wales in 2008 (<http://www.food.gov.uk/multimedia/pdfs/campylobacterstrategy.pdf>). In 2010, 62,684 human cases of enteric and non-enteric *Campylobacter* species caused infections were reported in England and Wales. This figure was six times higher than human food poisoning cases of *Salmonella* (9133 cases) reported in 2010 (<http://www.hpa.org.uk/Topics/InfectiousDiseases/>). Chicken and related products are the major reservoir in most cases (50-80% of cases) with broiler chickens accounting for 20-30% of these cases (EFSA., 2010b).

Based on the significance of disease burden of *Campylobacter* caused infections in the UK, the government has introduced an “Innovation Strategy for *Campylobacter*” from 2010 through to 2015. The program aims to research ways to control *Campylobacter* species at all levels in the food chain from farms to consumers,

(<http://www.food.gov.uk/multimedia/pdfs/campylobacterstrategy.pdf>). The European Food Standards Agency has also advised on conducting an active surveillance of campylobacteriosis in all European member states with an emphasis on detecting unreported cases of campylobacteriosis. This surveillance strategy also aims to identify the unknown reservoirs of *Campylobacter* caused infections by advanced molecular techniques.

Huge costs are incurred with *Campylobacter* associated infections mainly due to medical costs, long absences of employees from work, expenses incurred as a result of product recalls, legal charges etc. It is estimated that the total costs linked to campylobacteriosis is \$1billion per annum in USA (CAST., 1994). In a surveillance conducted in Netherlands recently, it has been reported that the costs associated with campylobacteriosis is 21 million euros per annum (Havelaar *et al.*, 2005).

1.6 *Campylobacter* species sources and transmission of infection

Campylobacter species form part of normal flora of wildlife and domestic birds. This may be because of the higher body temperature of avian hosts favours their survival (Skirrow, 1977) and chicken is responsible for harbouring *Campylobacter* species (Corry and Atabay, 2001). Most cases of campylobacteriosis are related to the consumption of raw or undercooked poultry products or by cross contamination of raw or undercooked foods. Chicken is a rich source of essential proteins and minerals, and requires shorter preparation time compared to cooking pork and beef. These factors influence their popularity and enhance the chances of *Campylobacter* species related outbreaks (Corry and Atabay, 2001). Among *Campylobacter* species, *C. coli* and *C. jejuni* are the most prevalent in outbreaks. *C. jejuni* is responsible for 12 times the human cases compared to *C. coli* in England and Wales (Friedman *et al.*, 2000). *C. coli* is dominant in free range and organic chickens whereas *C. jejuni* is mainly isolated from caged chicken breeds (El-Shibiny *et al.*, 2005).

Campylobacter species are present in the intestine of chicken and mainly located in the cecum and colon (Berrang *et al.*, 2001). During slaughter and processing of chicken, the intestine may rupture and bacteria spread all over on the skin and get trapped in skin pores and cracks. *Campylobacter* species can persist on the carcass under frozen conditions and at 4 °C. Under favourable conditions these bacteria can grow on the skin and can be spread by cross contamination (Chantarapanont *et al.*, 2003). Even under

controlled packaging conditions *Campylobacter* species can persist on chicken skin and can spread by improper handling and storage at the consumer end (Scherer *et al.*, 2006).

Other processed animal meats have been shown to harbour *Campylobacter* species and these include cattle, sheep, pigs, turkey and ducks (Humphrey *et al.*, 2007). The highest prevalence of *Campylobacter* species is on pig carcasses, compared to sheep and beef (Nesbakken *et al.*, 2003). This may be because the skin remains on pig carcass during most of the slaughter and processing of meat (Moore *et al.*, 2005). *Campylobacter* species are present in the digestive tract of healthy cattle (Atabay and Corry., 1998) whereas the lowest prevalence of *Campylobacter* species is in sheep (Zweifel and Stephan., 2004). Shellfish obtained from water systems contaminated with *Campylobacter* species is also a major source of dissemination of infection (Wilson and Moore., 1996). The transmission of *Campylobacter* species to the human hosts is also attributed to the consumption of unpasteurised milk and milk products. The cross contamination with animal faeces is the likely cause of transmission of *Campylobacter* species in milk especially during the milking process (Zilbauer *et al.*, 2008, Hänninen *et al.*, 2000, Shane., 2000). Consumption of untreated water and direct contact with domestic and farm animals are high risk factors for *Campylobacter* species transmission to humans (Gilpin *et al.*, 2008, Coker *et al.*, 2002, Shane., 2000). Environmental contamination with animal faeces is also a major factor contributing to the transmission of *Campylobacter* species infection to humans (Ridley *et al.*, 2008). Wills and Murray (1997) reported that *Campylobacter* species were present in large numbers in chicken intestines during summer months (May through to October) and a significant reduction in number was observed in winter months. A similar trend was noticed when chickens were sampled in summer months showing 87-97% of samples tested positive for *C. jejuni*. In December and in January only 7% and 33% of chickens sampled were positive for *C. jejuni* respectively. This may also correlate with the number of *Campylobacter* species associated outbreaks in humans and the outbreaks in humans are thought to be seasonal too (Wills and Murray., 1997).

1.7 Enteric *C. jejuni* infection and clinical symptoms

The most common symptoms of *C. jejuni* infections are typical of gastroenteritis. Variability in the clinical symptoms of *C. jejuni* have been reported ranging from watery diarrhoea with no inflammation to mucous containing bloody diarrhoea. The severe

diarrhoeal attack is characterised by abdominal cramps and fever within 48 hours of the initial attack (Zilbauer *et al.*, 2008). The illness is self-limiting and the duration of illness is less than 7 days. Recurrence of illness can be observed in patients with low immune responses (Wassenaar and Blaser., 1999). The incidence of *C. jejuni* infections is very high in immunocompromised people e.g. AIDS patients (Coker *et al.*, 2002). In developing countries, children in early childhood are found to be most susceptible to *C. jejuni* infections because of their undeveloped immune system with the incidence of infection decreases in late childhood (Wassenaar and Blaser., 1999). Asymptomatic *C. jejuni* infections are also more frequently reported in the developing countries where individuals are in close contact with animals and are exposed to bad sanitation conditions (Coker *et al.*, 2002). In contrast, in developed countries only farm and slaughter house workers and consumers of raw milk etc represented asymptomatic carriers of *C. jejuni* (Wassenaar and Blaser., 1999).

1.7.1 Complex disease

A prolonged and life-threatening consequence of *C. jejuni* infection is the development of a neurological disease which is characterized by the ascending paralysis of peripheral and cranial nerves. This disease is called Guillain Barré syndrome. The first case of *C. jejuni* caused GBS was reported in 1982 as a post infection complication of *C. jejuni* enteritis (Rhodes and Tattersfield., 1982). Later research showed that *C. jejuni* produces lipooligosaccharides (LOS) that mimic the gangliosides of the nervous system initiating a host inflammatory immune response against the pathogen that causes nerve damage (Hadden and Gregson., 2001, Moran and Prendergast., 2001, Aspinall *et al.*, 1994). In addition to the nervous system damage other body organs can also get affected and even death occurs in severe cases (Korinthenberg and Monting., 1996). The global GBS incidence rate is between 0.4-4.0 per 100,000 cases annually with more cases reported in infants and elderly patients (McGrogan *et al.*, 2009, Hadden and Gregson., 2001, Asbury and Cornblath., 1990). A recent review of GBS reported cases in the literature associated *Campylobacter* species with 31% of GBS incidents arising from gastrointestinal infections (Poropatich *et al.*, 2010). The development of GBS is dependent on strain type and fitness of host cells. In some cases, *C. jejuni* causes Miller Fisher Syndrome (MFS) which is a less severe subtype of GBS. The typical symptoms of MFS include weakness of gait (ataxia), dysfunction of reflexes and defects in eye

movement. Around 5% of GBS cases are of MFS (Hughes and Cornblath., 2005, Govoni and Granieri., 2001).

Recently, *C. jejuni* has been reported to cause endocarditis in patients with artificial heart valve transplants (Dinant *et al.*, 2011) and reactive arthritis (Hannu *et al.*, 2004).

1.8 Treatment

The disease caused by the *Campylobacter* species is generally self-limiting. In case of systemic infections, macrolides (*e.g.* erythromycin), fluoroquinolones (*e.g.* ciprofloxacin) and tetracyclines are used for treatment. The antimicrobial resistance is recognized as a major factor in persistence of *Campylobacter* species caused infections in public health (EFSA., 2012, Moore *et al.*, 2006, McDermott *et al.*, 2005). The antibiotic susceptibility testing methods and the interpretation of test results for *Campylobacter* species vary considerably in different countries which makes it difficult to establish a list of effective antibiotics for treating infections caused by this bacteria (EFSA., 2012).

The most recent EFSA (2012) report indicated that the human clinical *C. jejuni* isolates showed highest frequency of resistance against ciprofloxacin (51.6%) followed by resistance to nalidixic acid (49.8%). Among the human *C. coli* strains, the highest resistance was observed against nalidixic acid (69%) whereas the second highest resistance was reported against ciprofloxacin (66%) (EFSA., 2012).

1.9 Host immune response to *C. jejuni* infections

C. jejuni is a commensal organism in chicken gut but causes disease when inside the human intestinal tract. To establish infection in humans *C. jejuni* has to overcome several host defence mechanisms. These bacteria have to pass through the acidic stomach environment, and when inside the intestinal tract they have to withstand high bile salt concentrations (Dasti *et al.*, 2010). Indeed, the mucin rich mucosal lining acts as a physical barrier for the underlying epithelium against foreign intrusion (Hugdahl *et al.*, 1988). *C. jejuni* has developed a number of factors to survive these hostile host gut conditions and initiate infection. *C. jejuni* peptidoglycan cell envelope that maintains the helical corkscrew shape of the bacterial cell contributes to overcoming the mucosal cell lining (Friedrich *et al.*, 2012). A transcriptional study of *C. jejuni* strains has reported upregulation of flagella, LOS and capsule related genes when in contact with the

intestinal mucosa suggesting a role of these surface structures in penetrating mucus (Tu *et al.*, 2008). The majority of *C. jejuni* strains have developed resistance to the potent effects of bile salts in intestine (Van Deun *et al.*, 2007).

The human body has a specialised immune defence system to provide protection against microbial infection. The human immune system can be broadly divided into the innate immune response and adaptive immune response. The factors and mechanisms of these immune systems in response to the *C. jejuni* infection will be briefly discussed here;

1.9.1 Innate immune system

The innate immune response is the first line of defence that bacteria encounter after entering the human body. When bacteria invade the intestinal mucosa the underlying epithelial cells respond to the bacterial invasion by releasing cytokines, chemokines and antimicrobial peptides. These elements of the innate immune system initiate an inflammatory response by recruiting macrophages and dendritic cells (DCs). *C. jejuni* strains have shown to initiate a pro-inflammatory response in cultured human epithelial cells by the production of cytokines interleukin 8 (IL-8) (Hickey *et al.*, 1999, Mellits *et al.*, 2002, Zheng *et al.*, 2008).

C. jejuni infection leads to the activation of transcription factor NF- κ B. The release of NF- κ B is coordinated with IL-8 production (Zheng *et al.*, 2008). The NF- κ B/rel family of transcription factors forms part of the early immune response to microbial infection and controls transcription of genes encoding for cytokines and chemokines (Silverman and Maniatis., 2001). Using a gnotobiotic IL-10; NF- κ B mouse infection model Lippert *et al* (2009) showed that *C. jejuni* infection triggered NF- κ B transcriptional activity that resulted in rapid recruitment of cytokines in the infected mouse colon. This immune response resulted in rapid and severe inflammatory colitis in the infected animal. In addition, *C. jejuni* infection promoted NF- κ B production initiating cytokine gene expression in bone marrow derived DCs (Lippert *et al.*, 2009). The vertebrates recognise products of microbial infection by specific membrane receptors called TLRs (Toll-like receptors) activating an immune response (Dunne and O'Neill., 2005). There are 10 protein receptors in the TLR family that interact with “pathogen associated molecular patterns” (PAMPs) (Medzhitov., 2001, Schnare *et al.*, 2001). The TLRs work individually or in combination to recognize microbial patterns. The expression of TLRs is tightly regulated to prevent an inappropriate activation of pro-inflammatory response.

When required host cells block TLR activity through expression of TLR-inhibitory protein Tollip (Melmed *et al.*, 2003). The microbial triggers for TLRs include surface polysaccharides such as LOS, capsule, flagellin and DNA (de Zoete *et al.*, 2010). Recently, the sialylated flagellin of *C. jejuni* has been shown as a molecular trigger for activating TLR-4 (Kuijf *et al.*, 2010). Al-Sayeqh *et al* (2010) has reported that the transcription factor NF-kB can directly detect *C. jejuni* infection and does not require other surface proteins including TLRs.

Different types of proinflammatory (IL-1 α , IL-6, IL-8, TNF α) and anti-inflammatory (TGF- β 1, TGF- β 2, TGF- β 3, IL-4, IL-10) cytokines are recruited as an innate immune response to *C. jejuni* infection (Bahrami *et al.*, 2011, Hu *et al.*, 2006a). The trigger of *C. jejuni* invading epithelial cells results in a rapid proinflammatory response as the first line of defence. Once released these cytokines are transported to T-cells where they act as a stimulus for adaptive immunity bridging with innate immunity (Hu *et al.*, 2006b). *C. jejuni* surface structures, mainly LOS, interact with the TLR-2 receptors present on epithelial cells to activate IL-6 production. IL-8 is released when *C. jejuni* is recognised by extracellular signal regulated kinase (ERK) and p38 mitogen activated protein (MAP) kinase pathways (Borrmann *et al.*, 2007). More recently Zheng *et al* (2008) have reported that cytolethal distending toxin was important in IL-8 production. The interaction with host epithelial cells also contributed to IL-8 release (MacCallum *et al.*, 2006, Hickey *et al.*, 1999). MacCallum *et al* (2006) also showed that the type of infected cell line and *C. jejuni* strain played a critical role in adhesion/invasion mediated IL-8 response. A recent study showed a *C. jejuni* mutant in *pgp1* (peptidoglycan peptidase 1) was unable to maintain the helical cell shape, was deficient in chick colonization and promoted chemokine IL-8 production in epithelial cells. This suggests that *C. jejuni* helical shape morphology also contributes to the innate immune response against *C. jejuni* infection (Friedrich *et al.*, 2012). *C. jejuni* infection acts as a stimulus for the release of anti-inflammatory IL-4 and IL-10 cytokines. IL-4 and IL-10 have immunoregulatory effects as they prevent tissue damage by terminating the production of the proinflammatory host response. IL-4 has been reported to down regulate the proinflammatory effects by IL-1 α , IL-1 β and TNF α chemokines (Bogdan *et al.*, 1993). IL-10 has been shown to provide resistance against infection to the host and plays a critical role in clearing out infection by immunostimulatory activities (Asadullah *et al.*, 2003, Lindsay and Hodgson., 2001) . More recently, *C. jejuni* infection of IL-4 pre-

infused intestinal pig epithelial cells (IPEC-1) has been shown to change their physiology allowing increased invasion and damaging the paracellular junctions which suggests a link between invasion and IL-4 production (Parthasarathy and Mansfield., 2009). In another study, an IL-10 deficient mouse was orally inoculated with *C. jejuni* 11168 that showed enhanced colonization of colon and development of severe enteritis within 2-35 days of infection. This showed IL-10 to be important in *C. jejuni* gastroenteritis (Mansfield *et al.*, 2007).

The inflammatory response to microbial infection in humans is characterised by the production of neutrophils in the intestine. In cases of severe colitis, neutrophils damage the mucosal structure, migrate across epithelial cells and cause complete dysfunction of the intestine (Anderson *et al.*, 1986). In *C. jejuni* infected human epithelial cells increased amounts of neutrophils were recruited that move from the basolateral to apical side of the epithelium concentrating in intestinal crypts (Murphy *et al.*, 2011). The cationic antimicrobial peptides/proteins (CAPs) are produced by the host innate immune system in response to the microbial infection (Eckmann., 2004). Defensins and cathelicidins are two example of CAPs frequently recruited in an innate immune response to bacterial infection (Lehrer., 2004). Cathelicidins are secreted by neutrophils targeting at the infected tissue (Iimura *et al.*, 2005). Another cationic antimicrobial peptide, Bactericidal/Permeability Increasing Protein (BPI) is produced in response to neutrophil recruitment and is accumulated in neutrophil vacuoles. BPI mediates killing by interacting with the lipid A structure of LOS in Gram negative bacteria (Weiss., 2003, Eckmann., 2004). An *in vitro* study with *C. jejuni* strain 11168 reported enhanced β - defensin production. The bactericidal activity of β -defensins killed bacteria by damaging their cell wall suggesting an important role of defensins in innate immune response (Zilbauer *et al.*, 2005). Recently, a *C. jejuni* mutant in the *waaF* gene was deficient in β -defensin production showing that *C. jejuni* lipooligosaccharides contribute to mediating innate immune protection in human body (Keo *et al.*, 2011). The innate complement system in humans also has an inhibitory and antibacterial response to *Campylobacter* species infection (Fernández *et al.*, 1995). Capsule polysaccharides in *C. jejuni* have stimulatory effects on complement based immunity in the human host (Guerry *et al.*, 2012).

1.9.2 Adapted antibody immunity

Adapted or acquired immunity is an essential defence strategy developed by vertebrates against microbial infection. The primary response of adapted immunity to *Campylobacter* species infection is the secretion of sIgA (immunoglobulin A) in the human gastrointestinal tract. This antibody immune response is dependent on age and geographical location of subjects. The epidemiological evidence of acquired immunity mediated by *C. jejuni* infection in developing countries showed that campylobacteriosis cases decreased with an increasing age accompanied with milder symptoms to illness. This is because in the developing world infants are frequently exposed to food borne pathogens including *Campylobacter* species due to the lack of hygiene standards which results in early development of an anti-*Campylobacter* antibody mediated immune response in early childhood (2-5 years of age). The increasing level of IgA in adults is accompanied by milder and self-limiting *C. jejuni* disease profile with an overall decrease in the number of *C. jejuni* mediated illnesses (Tribble *et al.*, 2008). In comparison, in industrialized nations general health conditions and public awareness of food borne infections is better which results in more incidents of *Campylobacter* enteritis reported in the adult population (Tribble *et al.*, 2007). The epidemiological data obtained from individuals in contact with dairy farms and frequently consuming raw milk showed a lower incidence of *C. jejuni* mediated diarrhoea compared to people not exposed to raw milk on daily basis (Blaser, *et al.*, 1987). Both a symptomatic and an asymptomatic *Campylobacter* infection mounted an equally intense *Campylobacter*-specific IgA antibody response (Tribble *et al.*, 2007). In *Campylobacter* disease, the adaptive immunity is stimulated by bacterial surface structures including flagellin (Baqar *et al.*, 2008), LOS and capsule polysaccharides (Moran and Prendergast., 2001).

The *C. jejuni* colonization of the chicken gut also mediates an adaptive antibody response that varies with the age of the chicken. In 1-2 week old birds, the levels of maternal derived IgY (Immunoglobulin Y) antibodies are high and this amount levels out just after 3 weeks making chicken flocks more susceptible to colonization by *C. jejuni* (Cawthraw and Newell., 2010). *C. jejuni* surface polysaccharides mainly flagella and the surface expressed CadF adhesin are major stimulants for antibody production in chicken within first three weeks (Shoaf-Sweeney *et al.*, 2008, Cawthraw *et al.*, 2000).

1.10 Pathogenesis mechanisms of *C. jejuni* infection

C. jejuni is a versatile organism. It has developed specialized mechanisms to survive in the environment and to cause pathogenesis in the host. For mediating an enteric or blood infection *C. jejuni* has to pass through a number of physical and chemical barriers in the host gastrointestinal tract. *C. jejuni* has to penetrate the mucosal protective layer of the gut, adhere to or/and invade intestinal epithelial cells, survive and multiply inside the cells and in some severe cases migrate into the blood. A brief description of the key virulence associated factors studied in *C. jejuni* is stated below.

1.10.1 Flagella

Campylobacter species possess one or two polar flagella that make them motile. This motility is essential for colonization of the avian gut and invasion of the mucous layer of the intestine in humans during diarrhoeal infection (Jagannathan and Penn., 2005, Grant *et al.*, 1993, Nachamkin *et al.*, 1993, Morooka *et al.*, 1985). The *Campylobacter* flagellum is composed of two main homologous units, flagellin A (FlaA) and flagellin B (FlaB). The gene *flaA* is regulated by σ_{28} (sigma factor 28) and *flaB* is regulated by σ_{54} (sigma factor 54) (Jagannathan and Penn., 2005). *In vitro*, the role of FlaA in colonization, adherence and invasion of *C. jejuni* in human intestinal cells has been reported (Wassenaar *et al.*, 1991, Jain *et al.*, 2008). However, the *flaB* *C. jejuni* mutant formed flagella with a normal structure and had no influence on virulence (Wassenaar *et al.*, 1991, Guerry., 2007). *C. jejuni* produces eight different types of proteins when it comes in contact with the host surface (Konkel *et al.*, 2004).

Due to the lack of type III secretion system in *C. jejuni*, the secretion and transport of flagellated and non-flagellated proteins in the host cells is controlled by flagella. The secretion of the key proteins called *Campylobacter* invasion antigen (Cia) that allows the growth of *C. jejuni* in INT-407 cells in serum free medium or in the presence of bile salts was found to be under the control of flagellin signals (Christensen *et al.*, 2009, Ko and Park., 2000). CiaB is an important virulence protein secreted during initial *C. jejuni* infection and is translocated through the intestinal cells. However, the mechanism of this translocation is still unknown (Guerry., 2007, Poly and Guerry., 2008). Recently, a set of four genes called flagellar coexpressed determinants (*feds*) have been identified as a part of the flagellar regulatory system in *C. jejuni*. The transcription of *feds* and *ciaI* are coregulated that in turn is dependent on the expression of *flaA* (Barrero-Tobon and

Hendrixon., 2012). *C. jejuni* flagella are unique structures. Even though these are highly motile, they are not recognised by the Toll-like receptor sites (TLR-5) on human epithelial cells, thereby avoiding the innate host immunity and invade epithelial cells (Andersen-Nissen *et al.*, 2005). Glycosylation of flagella is an essential phenomenon adopted by *C. jejuni* which will be discussed in a later section (Guerry., 2007).

1.10.2 Chemotaxis

Motile bacteria contain a chemosensory system which they use to move towards favourable conditions and away from harmful environments (Lux and Shi., 2004). As a pathogen inside the living host, *C. jejuni* is attracted towards the glycoprotein mucin of the mucosal lining of epithelial cells, amino acids (e.g. aspartate, serine, cysteine) and other organic salts (citrate, fumarate, malate) (Hugdahl *et al.*, 1988, Baserisalehi and Bahador., 2011). Methyl accepting chemotaxis proteins (MCP) are recognized to sense these chemicals in *C. jejuni* (Vegge *et al.*, 2009). A mutagenesis study by Hendrixson and DiRita (2004) identified the role of *C. jejuni* methyl accepting chemotaxis receptors (Cj0019c and Cj0262c) in chicken gut colonization. A *C. jejuni* mutant in *cheY*, a chemotaxis regulatory gene that controls flagellar rotation, was deficient in chick colonization (Hendrixson and DiRita., 2004) but displayed a hyperinvasive phenotype (Golden and Acheson., 2002). On the contrary, *C. jejuni* strains with two copies of the *cheY* gene were unable to cause disease in ferret infection model but successfully colonized the mouse intestine (Yao *et al.*, 1997). In a recent study, the elevated expression of a chemotaxis related gene *cheW* reduced the subcellular translocation of *C. jejuni* (van Alphen *et al.*, 2008).

It can be proposed that chemotaxis facilitates *C. jejuni* migration to the mucin in intestine but has no role in invasion through the mucosal lining (Konkel *et al.*, 2001). Similarly, mutation in an autoinducer-2 (AI-2) production gene (*luxS*) abolished *in vitro* adherence and colonization of chick hepatoma (LMH) cells (Quinones *et al.*, 2009). In *C. jejuni*, LuxS is an important enzyme of the methyl recycling pathway and produces AI-2 as its metabolic byproduct (Plummer., 2012). Inactivation of chemoreceptor, CetB (Cj1189) in *C. jejuni* completely eliminated their ability to invade cultured human epithelial cell lines (Golden and Acheson., 2002). A transposon mutagenesis study has reported a transposon inserted in a chemoreceptor gene, Cj0952c that together with an adjacent chemotaxis related gene Cj0951c was important in host cell invasion, motility

and chemotaxis response toward formic acid (Tareen *et al.*, 2010). Recently, a tissue culture study using INT-407 cells showed that a *C. jejuni* 81-176 mutant in a chemoreceptor gene *cheR*, resulted in a hyperadherent and hyperinvasive phenotype whereas deletion of another chemoreceptor gene *cheB* was responsible for a non-adherent and non-invasive profile (Kanungpean *et al.*, 2011). The same study reported that a *cheRB* mutant was unable to colonize the chick cecum. Therefore, the research in *Campylobacter* chemotaxis system shows that it not only allows survival of bacteria under environmental stresses but it also has an important role in virulence particularly in host cell interaction.

1.10.3 Adhesion and Invasion

C. jejuni interaction with host cells has been studied as a critical process in the mediation of disease in human hosts (Hu and Kopecko., 2008). However, the mechanism that *C. jejuni* uses for adhesion and invasion is not understood fully. It was suggested that *C. jejuni* adheres to the epithelial cells before internalization. It was observed that when the bacteria were present in high cell density, the cell shape and flagellar orientation changes which allows bacteria to penetrate the mucus that protects the intestinal epithelium. This process was considered related to the adhesion of *C. jejuni* to the host cells (Ferrero and Lee., 1988). Everest *et al* (1992) showed that *C. jejuni* and *C. coli* strains were able to adhere to the epithelial cells without invading them afterwards. During binding to the host cells, *C. jejuni* preferentially associated with the intercellular junctions that would allow bacteria to transcytose the cell monolayers (Konkel *et al.*, 1992a, Oelschlaeger *et al.*, 1993) and this attachment to the host cell membrane was promoted by centrifugation (DeMelo *et al.*, 1989; Konkel *et al.*, 1992b). However, the colonization studies showed the adhesion of host cells is not an essential step as *C. jejuni* was successfully able to stay in the intestine after colonizing the mucus suggesting that *C. jejuni* might invade the epithelial cells directly (Lee *et al.*, 1986).

Early research proposed different mechanisms which *C. jejuni* uses to invade host cells but there is no consensus on these mechanisms. Like other invasive pathogens including *Listeria*, *Shigella* and *Salmonella*, *C. jejuni* may invade the host cells in a microfilament dependant manner (DeMelo *et al.*, 1989, Konkel and Joens, 1989; Konkel *et al.*, 1992b) while the other studies found that the invasion remained unaffected in the presence of microfilament inhibitors (Oelschlaeger *et al.*, 1993). This suggested that the entry into

host cells was via a microtubule dependant pathway (Oelschlaeger *et al.*, 1993). During the *C. jejuni* 81-176 invasion process into INT-407 cells, actin filament depolymerisation and polymerisation of microtubule bundles took place in response to the signal transduction from bacterial cells bound to the cell surface. This resulted in the formation of finger like protrusions by the cell surface which engulf bacteria. *C. jejuni* survives within these membrane-bound endosomes which are then transported along the microtubules to the perinuclear region of the cell (Hu *et al.*, 2008). A novel host signal transduction pathway has also been proposed for *C. jejuni* invasion into the differentiated enterocyte like cells (Wooldridge *et al.*, 1996). Wooldridge *et al* (1996) showed that *Campylobacter* species stimulate a receptor on host cell membrane that interacts with molecules associated with caveolae. This interaction results in a signal transduction event that causes ruffling across the membrane and subsequent actin filament dependent endocytosis of *C. jejuni* (Figure 1.1).

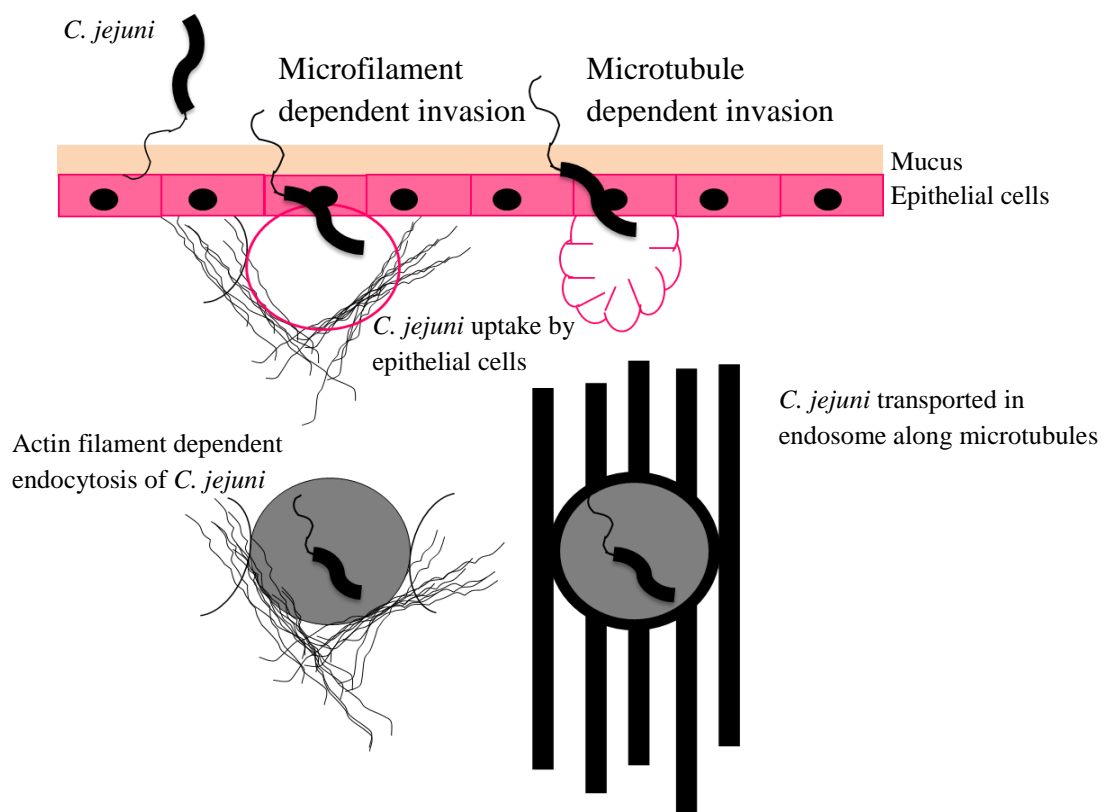


Figure 1.1: A diagrammatic representation of the proposed mechanisms of *C. jejuni* invasion into the epithelial cell lining.

The ability of *C. jejuni* to attach to and invade epithelial cells is generally strongly correlated with severity of clinical symptoms presented (Fauchere *et al.*, 1986). Several studies have used *in vitro* cultured cell lines of human and non-human origin to investigate *C. jejuni* interaction with host cells (Konkel *et al.*, 2001, Ketley., 1997, Prasad *et al.*, 1996, Konkel and Joens., 1989). In addition, *in vivo* animal infection models including primates have been used to study the role of invasion and adhesion in *C. jejuni* pathogenesis (Senior *et al.*, 2011, Yao *et al.*, 1997, Babakhani and Joens., 1993, Russell *et al.*, 1993). All these studies have reported adhesion and invasion as important factors contributing to *C. jejuni* pathogenesis. *C. jejuni*'s ability to adhere to and invade epithelial cells very strongly depends on strain type and the infection model used (for example type of intestinal cell line used) (Prasad *et al.*, 1996, Konkel and Joens., 1989, Newell *et al.*, 1985).

Research has also suggested that clinical human isolates of *C. jejuni* are generally more adherent and invasive than non-clinical isolates. Prasad *et al* (1996) demonstrated that human isolates are more pathogenic than chicken isolates for their invasion, adhesion and cytotoxin production phenotypes using the human cell lines HeLa and HEP-2 cells as *in vitro* models. Similar results were reported by Konkel and Joens (1989) for nine clinical and four nonclinical strains of *C. jejuni* tested for their adherence and invasion to cultured HEP-2 cells. Newell *et al* (1985) compared two main phenotypes (*i.e.* adherence and invasion) of *C. jejuni* environmental isolates and strains obtained from diarrhoea patients to HeLa cells using electron microscopy and immunofluorescence. The authors concluded that the clinical isolates were more invasive than the nonclinical strains. This difference in the invasion potential between the human and animal isolates was also reported by Biswas *et al.*, 2000. A detailed study classified *C. jejuni* strains isolated from clinical, poultry and environmental sources as hyper, high and low invasive using an *in vitro* invasion assay. It was reported that the hyperinvasive category had a greater proportion of clinical isolates compared to strains from other sources (Fearnley *et al.*, 2008). Based on the above studies it can be suggested that adhesion and invasion are important phenotypes contributing to *C. jejuni* pathogenesis. In contrast, another research group (van Deun *et al.*, 2007) did not find any difference in the invasion potential among the set of human and poultry isolates investigated. However, *C. jejuni* adhesion and invasion is a multifactorial process and other bacterial factors that contribute to this phenotype still need investigation.

1.10.3.1 Factors influencing adhesion and invasion in *C. jejuni*

Campylobacter species do not possess a Type III secretion system and other structures including fimbriae or pilli as in other Gram negative bacteria such as *Salmonella* and *E. coli*. Instead adherence and invasion of intestinal epithelial cells is mediated by several other bacterial structures. Some of these factors directly influence adhesion and invasion of *C. jejuni* to the host surface receptors while others play an indirect role in host cell interaction. The role of these host cell interaction factors in *C. jejuni* pathogenesis has only started to be understood (Hu and Kopecko., 2008, Dasti *et al.*, 2010).

A functional *C. jejuni* flagellar export apparatus is required for delivery of effector proteins into host epithelial cells. A flagellin protein, FlaC which is structurally homologous to FlaA and FlaB has been shown to support the flagellar export apparatus. FlaC has also been found to have a role in *in vitro* invasion of host epithelial cells (Song *et al.*, 2004). Two *Campylobacter* invasion antigen (Cia) effector proteins, CiaB with limited homology to SipB of *Salmonella* species and CiaC with no known similarity to any other proteins that have been characterised (Christensen *et al.*, 2009). An insertion mutation into gene, Cj1242 which was annotated as CiaC in *C. jejuni* clinical strain F38011 showed defects in motility, adherence and internalization of INT-407 cells (Christensen *et al.*, 2009). In contrast, mutation in *ciaB* had no effect on the invasion potential of *C. jejuni* 81-176 in T84 cells (Novik *et al.*, 2010) suggesting that the role of CiaB in invasion is strain and cell line dependant. Another Cia protein called CiaI (Cj1450) has been recently identified in *C. jejuni*. CiaI contains an amino terminal type III secretion sequence and is transported via the flagellar type III secretion system into host epithelial cells. A *C. jejuni* *ciaI* mutant was attenuated for survival in cultured INT-407 and HeLa epithelial cells which suggests CiaI might have a role in intracellular survival (Buelow *et al.*, 2011). Not all Cia proteins are known yet nor their exact mechanism in *C. jejuni* host interaction is fully understood however a *C. jejuni* strain lacking CiaB secretion ability inhibited secretion of other Cia proteins into the epithelial cells (Ko and Park., 2000).

In bacteria, the main role of σ_{28} genes is in the functioning of flagella (Carrillo *et al.*, 2004, Goon *et al.*, 2006, Poly *et al.*, 2007a). However, *C. jejuni* expresses some σ_{28} genes that are not important in motility but have been involved in invasion and damage to epithelial cells (Goon *et al.*, 2006, Poly *et al.*, 2007a). A σ_{28} dependent gene *fspA* is

expressed as two alleles *fspA1* and *fspA2* in some *C. jejuni* strains. FspA1 and FspA2 proteins are not directly involved in *C. jejuni* invasion however FspA2 has been implicated in apoptosis of INT-407 cells (Poly *et al.*, 2007a). Another flagella dependent gene with a role in invasion of intestinal epithelial cells is *cj0977* (Goon *et al.*, 2006). At first the Cj0977 protein was shown to be secreted in the cytoplasm. Its secretion was independent of the flagellar export apparatus and had no effect on motility (Goon *et al.*, 2006). A later study contradicted the initial research and showed a *cj0977* mutant to be deficient in motility in a liquid culture and was unable to invade epithelial cells (Novik *et al.*, 2010). Inactivation of *fliS* (σ_{28}) and *rpoN* (σ_{54}) resulted in reduced motility and defects in invasion of HeLa cells (Fernando *et al.*, 2007). A flagellar coexpressed determinant, FedA, together with CiaI has been reported recently to be involved in the invasion of human epithelial cells (Barrero-Tobon and Hendrixson., 2012). Chemotaxis and the glycosylation systems (O and N-linked) also have an important role in influencing the adhesion and invasion phenotypes in *C. jejuni* and these have been discussed in detail in sections 1.10.2 and 1.10.9 respectively.

Other surface polysaccharides including LOS and capsule have been implicated in adherence and invasion of epithelial cells and *in vitro* ferret model of infection (Karlyshev *et al.*, 2000, Bacon *et al.*, 2001, Kanpies *et al.*, 2004, Bachtiar *et al.*, 2007, Louwen *et al.*, 2008). An initial study investigated the role of *C. jejuni* LOS in host cell interaction and internalization. This study showed that the purified extract of LOS promoted adhesion to epithelial cells and this interaction was eliminated with the oxidation of LOS (McSweegan and Walker., 1986). Kanpies *et al* (2004) reported that a mutant in *C. jejuni* 81-176 LOS locus lost its ability to invade cultures human epithelial cell lines. Bacon *et al* (2001) demonstrated the role of *C. jejuni* 81-176 LOS in invasion and serum resistance using INT-407 cell lines in an *in vitro* model. Sialylated LOS exposed on the outer surface of *C. jejuni* has been shown to influence invasion (Louwen *et al.*, 2008, Habib *et al.*, 2009). Louwen *et al* (2008) showed that the mutation in *cst-II* gene encoding for sialyltransferase in GBS associated *C. jejuni* strains made them defective in invasion of intestinal epithelial cells. To further investigate the role of LOS in *C. jejuni* pathogenesis, Marsden *et al* (2009) showed that a *C. jejuni* mutant lacking a functional LOS locus was unable to invade Caco-2 cells. Mutations in one aminotransferase encoding gene *wlaRG* and two glycosyltransferase encoding genes *wlaTB* and *wlaTC* in LOS core in *C. jejuni* 81116 deprived the mutants in their ability to invade *in vitro* human epithelial cell lines. Moreover, mutants in these three genes

showed reduced immunogenicity and invasion into chick embryonic fibroblasts (Holden *et al.*, 2012). Recently, Javed *et al.* (2012) showed that a transposon inserted in a LOS gene, *cj1136*, which encodes for a putative galactosyltransferase in *C. jejuni* NCTC11168 significantly reduced the ability of the mutant *C. jejuni* 01/51 strain to invade into cultured INT-407 and Caco-2 cells. All the above studies strongly support the role of LOS in *C. jejuni* interaction to the human cells.

Campylobacter species contain adhesins or binding factors that are required to adhere and internalize the host epithelium. The CadF (*Campylobacter* adhesion to fibronectin) is an outer membrane protein present in all *C. jejuni* and *C. coli* strains and establishes bacterial interaction with the host extracellular fibronectin receptors (Konkel *et al.*, 1997). The *cadF* gene sequence in *C. jejuni* has a 39 bp insertion compared to the *cadF* in *C. coli*. This sequence difference in the *cadF* gene resulted in *C. jejuni* being more adherent and invasive to the INT-407 epithelial cells compared to *C. coli* strains. Furthermore, isogenic *cadF* mutant strains showed significantly impaired host cell interaction ability confirming its role in *C. jejuni* pathogenesis (Krause-Gruszczynska *et al.*, 2007). A recent study reported that *C. jejuni* fibronectin binding proteins, CadF and FlaA, mediate invasion of host epithelial cells via an epidermal growth factor (EGF) pathway. This process involves activation of *C. jejuni* cytoskeleton related proteins mainly, P13 kinase, c-Src and focal adhesion kinase (FAK) (Eucker and Konkel., 2012). Two surface exposed lipoprotein adhesins, CapA and JlpA, have been shown to mediate *C. jejuni in vitro* adhesion to the human epithelial cell lines and *in vivo* colonization of chicken gut (Ashgar *et al.*, 2007, Jin *et al.*, 2001, Pei and Blaser., 1993).

Campylobacter species contain different periplasmic proteins PEB 1-4 that serve as adhesins (Pei *et al.*, 1991). PEB 1 is a 28-kDa protein and is conserved in *C. jejuni* and *C. coli* strains (Pei and Blaser., 1993, Pei *et al.*, 1991). PEB 1 has immunogenic and immunoprotective properties (Du *et al.*, 2008) and is required for *C. jejuni* binding to HeLa cells (Pei *et al.*, 1998). It is functionally homologous to periplasmic binding proteins from other Gram negative bacteria with a role in glutamate/aspartate transport (Leon-Kempis Mdel *et al.*, 2006, Pei and Blaser., 1993). PEB 1 was detected in culture supernatants by cell fractionation and immunoblotting techniques confirming that PEB1 in *C. jejuni* is a periplasmic associated protein and not an inner or outer membrane protein (Leon-Kempis Mdel *et al.*, 2006). PEB 3 has antigenic properties similar to PEB 1 and shares homology with class 1 pili in *Neisseria meningitidis* and heat-labile

enterotoxin B subunit in *E. coli*, and is required for interaction with intestinal epithelial cells (Pei *et al.*, 1991). PEB 4 is another periplasmic chaperone in *C. jejuni* as antigenic in nature as PEB 1. It is involved in adhesion to INT-407 cells, biofilm formation and mice colonization (Asakura *et al.*, 2007) as well as invasion of cultured human epithelial cells (Kervela *et al.*, 1993). Recently, another periplasmic-binding protein encoding gene, Cj1289, has been identified in *C. jejuni*. This protein is structurally similar to PEB 4 in *C. jejuni* and may have a role in virulence (Kale *et al.*, 2011). HtrA is a periplasmic protein in *C. jejuni* with dual functionality: a chaperone and a protease. HtrA chaperone activity is required for interaction with host epithelial cells and binding to macrophages (Baek *et al.*, 2011a).

Other bacterial factors recently identified that influence adhesion and invasion phenotypes include a *C. jejuni* 11168 antigen encoding gene, *cj0034c*, with a role in invasion of INT-407 epithelial cell lines. Furthermore, mutants in *cj0034c*, *cj0404*, *cj0525c* were reduced in invasion of spleen and liver (Nielsen *et al.*, 2012). Oxidoreductase encoding genes *cj0004c* and *cj0005c* involved in sulphite metabolism in cytoplasm (Tareen *et al.*, 2011), polyphosphate kinase 2 encoding gene (*ppk2*) required for inorganic polyphosphate (poly P) synthesis as an alternate energy source in stress conditions (Gangaiah *et al.*, 2010) are important *C. jejuni* metabolic pathways that influence *C. jejuni* attachment and internalization in *in vitro* tissue culture assays. A formate metabolism related operon (*fdhTU*) expressed as two alleles *fdhT* and *fdhU* has been recently identified in *C. jejuni* to have a role in internalization of bacterial cells in an *in vitro* colony based assay and by using fluorescence microscopy (Pryjma *et al.*, 2012).

The bacterial factors reported as influencing the adhesion and invasion phenotypes of *C. jejuni* strains are summarized in table 1.1.

Table 1.1: Summary of the bacterial factors influencing adhesion and invasion of *C. jejuni*.

Bacterial factor	Function	Phenotype		Reference
		Adhesion	Invasion	
Flagella				
FlaA	Flagellin A		√	Wassenaar <i>et al.</i> , 1991, Jain <i>et al.</i> , 2008, Eucker and Konkel., 2012
FlaC	Flagellar export apparatus		√	Song <i>et al.</i> , 2004
<i>cj0977</i>	Flagellum		√	Goon <i>et al.</i> , 2006
Sigma factors				
$\sigma 28$ (<i>flhS</i>)	Transcription of flagellar genes		√	Carrillo <i>et al.</i> , 2004, Fernando <i>et al.</i> , 2007
$\sigma 54$ (<i>rpoN</i>)			√	Fernando <i>et al.</i> , 2007
Chemotaxis				
<i>luxS</i>	Autoinducer-2 (AI-2) producing gene (methyl recycling pathway)	√		Quinones <i>et al.</i> , 2009
CetB	Chemoreceptor		√	Golden and Acheson., 2002
<i>cj0952c</i>	Formic acid chemoreceptor		√	Tareen <i>et al.</i> , 2010
CheR, CheB	Chemoreceptor	√	√	Kanungpean <i>et al.</i> , 2011
LOS				
Cst-II	Sialyltransferase		√	Louwen <i>et al.</i> , 2008
WlaRG	Aminotransferase		√	Holden <i>et al.</i> , 2012
WlaTB, WlaTC	Glycosyltransferase		√	
<i>cj1136</i>	Putative galactosyltransferase		√	Javed <i>et al.</i> , 2012
Periplasmic adhesins				
CadF	<i>Campylobacter</i> adhesion to fibronectin	√	√	Krause-Gruszczynska <i>et al.</i> , 2007, Eucker and Konkel., 2012
CapA, JlpA	Lipoprotein adhesins		√	Asgar <i>et al.</i> , 2007
PEB1	Periplasmic adhesins	√		Pei <i>et al.</i> , 1998
PEB3		√		Pei <i>et al.</i> , 1991
PEB4	Periplasmic chaperone	√	√	Asakura <i>et al.</i> , 2007, Kervela <i>et al.</i> , 1993
HtrA	Periplasmic protein	√		Baek <i>et al.</i> , 2011a
Toxin				
CdtA, CdtC	Cytolethal distending toxin (CDT) subunits	√		Lee <i>et al.</i> , 2003
Capsule				
KpsE	Capsular polysaccharide ABC transporter	√	√	Bachtiar <i>et al.</i> , 2007
JJD26997_1801	Capsular polysaccharide biosynthesis		√	Javed <i>et al.</i> , 2010
O-linked glycosylation system				
PseAc	Pseudaminic acid	√	√	Ewing <i>et al.</i> , 2009
N-linked glycosylation system				
PglH	General glycosylation pathway protein	√	√	Karlyshev <i>et al.</i> , 2004
PglB	Oligosaccharide transferase	√	√	Szymanski <i>et al.</i> , 2002
PglE	Putative aminotransferase	√	√	
<i>cj1121c</i>	Aminotransferase	√		Vijayakumar <i>et al.</i> , 2006
Other				
FedA	Flagellar coexpressed determinant		√	Barrero-Tobon and Hendrixson., 2012
CiaC	<i>Campylobacter</i> invasion antigen (Cia)	√		Christensen <i>et al.</i> , 2009
CiaI			√	Barrero-Tobon and Hendrixson., 2012
<i>cj0034c</i> , <i>cj0404</i> , <i>cj0525c</i>	Antigen encoding genes		√	Nielsen <i>et al.</i> , 2012
<i>cj0004c</i>	Oxidoreductase	√	√	Tareen <i>et al.</i> , 2011
<i>cj0005c</i>	Sulphite metabolism	√	√	
<i>ppk2</i>	Polyphosphate kinase 2 in inorganic polyphosphate (poly P) synthesis	√	√	Gangaiah <i>et al.</i> , 2010
FdhTU	Formate metabolism		√	Pryjma <i>et al.</i> , 2012
<i>feoB</i>	Ferrous iron uptake		√	Raphael and Joens., 2003

1.10.4 Translocation

Campylobacter infections are initiated by interaction of bacteria with the intestinal mucus lining followed by invasion of epithelial cells. *Campylobacter* strains are also isolated from cases of bacteraemia which suggests the ability of bacteria to cross epithelium into the extra-intestinal locations. *C. jejuni* are able to translocate through the otherwise impermeable epithelial cell layer by using three reported systems. *Campylobacter* strains have been observed to translocate by paracellular pathway or paracytosis in which bacteria move through the epithelial cells without invading them via tight junctions between them (Everest *et al.*, 1992, Konkel *et al.*, 1992a, Grant *et al.*, 1993). In the second system, *C. jejuni* are observed to use specialized intestinal epithelial cells called M-cells to translocate through the epithelial cell layer into the underlying lymphoid tissue (Walker *et al.*, 1988). Finally, the third translocation mechanism called transcellular translocation or transcytosis involves bacterial invasion into the epithelial cells from where they travel through the cytoplasm appearing at the other side (Hu *et al.*, 2008, Brás and Ketley., 1999). Within 60 minutes of post-infection, *C. jejuni* 81-176 was internalised into endosomes. These bacteria containing endosomes travelled for 8-10 hours from the apical surface to the basolateral surface where they were released by exocytosis (*i.e.* endosome fusion to the basolateral surface) Hu *et al.*, 2008. The transcellular movement of *C. jejuni* 81-176 (Hu *et al.*, 2008a) and more recently of *C. fetus* 11686 (Baker and Graham., 2010) carried on without compromising the integrity of polarized Caco-2 cells monolayer. By contrast, a study that infected polarized human colonic HCA-7 cells with *C. jejuni* (Beltinger *et al.*, 2008) observed complete loss of colonic cells integrity and disruption of cellular tight junctions within 6 hours of infection. A modification of transcellular transport called subvasion has been reported in *C. jejuni* recently where bacteria travel for a few hours at the cell basal layer of infected cells before finally invading the epithelial cells (van Alphen *et al.*, 2008, Pogacar *et al.*, 2010).

1.10.5 Toxin production

Campylobacter species are known to produce several toxins but the cytolethal distending toxin (CDT) is the only one studied in detail (Wassenaar and Blaser., 1999). CDT is composed of three subunits Cdt A, B and C. The presence of these protein subunits is necessary for the function of CDT (Pickett *et al.*, 1996). CdtB acts as a nuclease (Lee *et*

al., 2003) and has shown structural similarity to the DNAase I of the mammalian eukaryotic cells (Lara-Tejero and Galan., 2000). Lee *et al* (2003) demonstrated that only CdtA and CdtC but not CdtB show close affinity and bind to the HeLa cell surface. This might suggest that CdtB might be transported to the nucleus to digest DNA and kill the cell. CDT causes HeLa and Caco-2 cells to be blocked in the G2 phase during the mitotic cell cycle thus resulting in eventual cell death (Whitehouse *et al.*, 1998). It has been recently reported that *C. jejuni* survives and multiplies in human monocytes and utilizes CDT to initiate an apoptotic state in cells that leads to the cell death (Hickey *et al.*, 2005). *C. jejuni* CDT induces the release of the cytokine Interleukin-8 (IL-8) from the intestinal epithelial cells by two mechanisms (Hickey *et al.*, 2000). In the first mechanism, the live *C. jejuni* cells are needed to adhere and invade the epithelial cells and then induce IL-8 production which is responsible for the proinflammatory response seen in diarrhoeal disease (Hickey *et al.*, 1999). The other mechanism for IL-8 production requires the production of CDT itself and requires the presence of all the three CDT subunits (*i.e.* CdtA, B and C) (Hickey *et al.*, 2000). This mechanism of CDT activity has been demonstrated by the Hickey group (Hickey *et al.*, 1999, Hickey *et al.*, 2000) *in vitro* studies using human epithelial cell lines. However, there is no *in vivo* confirmation of the pathogenicity of CDT inside the human host.

1.10.6 Iron homeostasis

Iron is an essential nutrient of bacteria required for the survival in diverse environments. It is also required for electron transport, oxidative stress responses and expression of pathogenic genes (Johnson *et al.*, 2005, McHugh *et al.*, 2003, Bou-Abdallah *et al.*, 2002, Escolar *et al.*, 1999, Mason and Cammack., 1992, Hantke., 1981).

Campylobacter species have evolved an efficient iron uptake, transport and storage system. *C. jejuni* acquires iron in different ways. The *C. jejuni* genome sequence contains only one ferrous iron uptake gene *feoB* (Raphael and Joens., 2003). *C. jejuni* 81-176 with an inactivated *feoB* gene showed reduced invasion of INT-407 human epithelial cells and porcine IPEC-1 small intestine cells. *C. jejuni* NCTC11168 *feoB* mutants were unable to colonize and persist in the rabbit ileum. Mutants in the *feoB* gene of *C. jejuni* NCTC11168, 81-176 and ACTC 43431 were also deficient in colonization of the chick cecum and showed reduced infection in piglet virulence models (Naikare *et al.*, 2006). An outer surface receptor, CfrB has been recently identified in

many *C. jejuni* strains. The CfrB receptor has ~34% similarity with CfrA. The synthesis of the CfrB receptor is essential for ferric iron uptake. A *C. jejuni* mutant in the *cfrB* gene was defective in colonization of the chick intestinal tract (Xu *et al.*, 2010).

When *C. jejuni* is present inside the human or avian host it has to obtain iron from complex molecules including heme, haemoglobin, transferrin and lactoferrin (Palyada *et al.*, 2004). The genes responsible for iron uptake from these compounds are present in two transcriptional sets and are located together upstream of the *ctuA* gene that encodes for an outer membrane receptor (Holmes *et al.*, 2005). The protein, CtuA has a role in promoting chick colonization (Palyada *et al.*, 2004). The loss of the *ctuA* gene does not abolish ferri-lactoferrin transport that suggests alternate processes used by *C. jejuni* for iron uptake inside the host (Miller *et al.*, 2008a).

C. jejuni produces many exogenous siderophores (Guerry *et al.*, 1997, Richardson and Park., 1995, Baig *et al.*, 1986). Among these siderophores, a ferri-enterochelin transport system in *C. jejuni* consists of a number of outer membrane associated proteins. A periplasmic receptor protein (CfrA), an outer membrane binding protein (CeueE), and an iron transport complex (CeueBCD) (Richardson and Park., 1995). This iron associated transport system is conserved in many *C. jejuni* and *C. coli* strains.

C. jejuni contains a ferritin like protein, Cft that has a role in excessive iron storage and survival under oxidative stress conditions (Palyada *et al.*, 2004). In addition to Cft, a single putative bacterioferritin protein (Dps) is present in *C. jejuni* (Wai *et al.*, 1995). The Dps protein has ~40 iron and oxygen binding sites and has a role in iron storage and protection against hydrogen peroxide damage. Dps expression is observed under both iron excess and depleted conditions (Ishikawa *et al.*, 2003). The piglets inoculated with wild type *C. jejuni* strains showed upregulation of *dps* gene with symptoms similar to human campylobacteriosis (Theoret *et al.*, 2011). Another gene, Cj0241c, encoding for a probable iron binding protein, has been identified in sequenced *C. jejuni* strains with putative role in iron storage which also reflects the importance to safeguard bacteria under potent amount of metals including iron (van Vliet *et al.*, 2002).

Iron homeostasis in bacteria is a well observed process which is regulated at the transcriptional level. The transcriptional factor Fur regulates the amount of iron available to bacteria in the form of ferrous ions. *C. jejuni* encodes for two homologous transcriptional regulators (Fur and PerR). PerR, regulates the expression of the enzymes

catalase (KatA) and alkyl hydroperoxidase (AhpC) that have a role in the oxidative stress response (van Vliet *et al.*, 1999). Mutation of the *perR* gene in *C. jejuni* resulted in defects of colonization in the chick intestinal tract (Palyada *et al.*, 2009). Iron metabolism and storage is an essential phenomenon in *C. jejuni* which involves several genes from multiple cellular processes particularly energy metabolism and oxidative stress response systems. The specific role of these genes in iron homeostasis and *C. jejuni* pathogenesis still need characterization.

1.10.7 Lipopolysaccharide

C. jejuni like many other Gram negative bacteria, has diverse surface polysaccharide materials of variable length and density that form lipooligosaccharide (LOS) and capsule (Wassenaar and Blaser., 1999, Moran., 1997). These surface structures have a key role in serological identification of *C. jejuni* and also contribute to its pathogenesis (Zilbauer *et al.*, 2008). LPS are high molecular weight units with an outer core composed of 10-15 repeating oligosaccharide residues. LOS, in contrast, has a low molecular weight and lacks repeating oligosaccharide units in the outer core. Both LOS and LPS are anchored to lipid A on the surface. The lipid A of *C. jejuni* is an endotoxin (Moran., 1997) which induces an immune response in human hosts (Bax *et al.*, 2011, Heikema *et al.*, 2010, Kuijf *et al.*, 2010). The electron microscopic characterization of *C. jejuni* lipopolysaccharides (LPS) by Karlyshev *et al* (2001) demonstrated, for the first time, that the high molecular weight lipopolysaccharides (HMWs) in *C. jejuni* are capsular polysaccharides (CPS). The outer core of LOS is a highly variable in different *C. jejuni* strains (Dorrell *et al.*, 2001, Parker *et al.*, 2008). The LOS outer core mimics the human gangliosides as it coated with the sialic acid residues. The structural similarity of LOS to human brain gangliosides develops into the neuropathies called GBS and MFS (Yu *et al.*, 2011, Guerry and Szymanski., 2008, Yuki *et al.*, 1995, Aspinall *et al.*, 1994). The role of bacterial LOS in causing neurological disorders in humans has recently been reviewed (Yuki., 2010). The sialylation of LOS in *C. jejuni* is responsible for serum resistance by the pathogen against host antigens (Keo *et al.*, 2011, Guerry *et al.*, 2000). Also, sialylated LOS triggers a variety of immune response systems in *in vitro* studies and in mice infection model which showed sialylation to be a complex but essential phenomenon in *C. jejuni* mediated neurological complications (Huizinga *et al.*, 2012). Recently, Naito *et al* (2010) showed the importance of the LOS outer core genes (*waaF* and *lgtF*) in stress response, intraepithelial survival and pathogenesis of *C. jejuni*. By

generating mutants in the LOS outer core genes these authors were able to show that a fully functional outer core was required to initiate immune response and biofilm formation (Naito *et al.*, 2010). The role of LOS in biofilm formation has also been studied earlier by Corcoran and Moran (2007). Marsden *et al* (2009) showed that deletion of LOS biosynthesis genes (*cj1132c* and *cj1152c*) in *C. jejuni* NCTC11168 resulted in loss of growth and natural transformation of mutants.

1.10.8 Capsule biosynthesis locus

Capsule is the high molecular weight polysaccharide surface structure in *C. jejuni* (Karlyshev *et al.*, 2000). The synthesis of capsule polysaccharides is independent of the low molecular weight polysaccharides, LOS and is attached to the bacterial cell surface via phospholipid linkage (Oldfield *et al.*, 2002). The capsular region in *C. jejuni* is composed of conserved *kps* genes which border the central variable region (Figure 1.2).

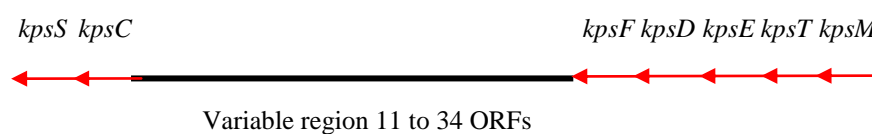


Figure 1.2: Schematic representation of the capsule region of *C. jejuni*.

The capsule region varies in size between 15 to 34 kbp as the central variable region consists of 11 to 34 ORFs. This variability in the capsule region is reflected in diverse capsular structures in different *C. jejuni* strains (Guerry *et al.*, 2012).

Capsule has antigenic properties and it is the major serodeterminant in the Penner serotyping scheme (Karlyshev *et al.*, 2000). Capsule is a complex and phase variable structure in different *C. jejuni* strains. The *C. jejuni* switches on/off the expression of its capsule which is due to the slip strand mismatching in one or more of the conserved *kps* genes (Guerry and Szymanski., 2008, Karlyshev *et al.*, 2005 a and b, Bacon *et al.*, 2001). *C. jejuni* capsules show variability in sugar molecules and possess other structural extensions that are responsible for different serotypes of *C. jejuni* strains. The structural variability of capsule is associated with the expression of heptoses of unusual configuration (*i.e.* *altro*, *ido*, *gulo*, *talo*) and O-methyl phosphoramidate (MeOPN) modifications (McNally *et al.*, 2006b and 2007). The capsular structure gets more

complicated by the introduction of a deoxy modification at C-6 position of a polysaccharide chain in addition to a normal heptose sugar (Guerry *et al.*, 2012). For example, the *C. jejuni* strains of serotype complex HS23/36 contain both the *glycero-D-altro*-heptose and its complementary 6-deoxy-*altro*-heptose (Aspinall *et al.*, 1993, Kanipes *et al.*, 2006). Similarly, MeOPN can be located on different sugar residues in different CPS structures (Guerry *et al.*, 2012).

Capsule has a role in serum resistance, survival to phagocytic activity and complement mediated immunity (Keo *et al.*, 2011, Guerry and Szymanski., 2008). *C. jejuni* mutants in the capsule polysaccharide transport protein encoding gene (*kpsM*) were more susceptible to the host complement immune system whereas LOS mutant showed sensitivity to the cationic antimicrobial drugs including polymixin B (Keo *et al.*, 2011). A previous study showed that the capsulated *C. jejuni* strains were resistant to killing to by β -defensins 1 and lysozyme activity (Zilbauer *et al.*, 2008). A further study showed that the CPS loci in *C. jejuni* provide resistance to the innate host immunity. Recently, Rose *et al* (2012) showed that only a slight variation in the capsule composition of *C. jejuni* NCTC11168, for example the lack of MeOPN modification, increased cytokine production by bone marrow derived murine dendritic cells (BMDCs). This confirms the role of *C. jejuni* capsule in survival against the host innate immunity.

The capsule has a role in virulence mechanisms including adhesion and invasion of *C. jejuni* (Bachtiar *et al.*, 2007, Bacon *et al.*, 2001). *C. jejuni* mutant in *kpsE* capsule gene in *C. jejuni* 81116 resulted in reduced adhesion and invasion of the mutant however, colonization of the chicken intestine remained unaffected (Bachtiar *et al.*, 2007). The capsular gene mutants in *C. jejuni* also resulted in impaired virulence profile in the ferret disease model (Bacon *et al.*, 2001). A recent study showed that surface related structures, primarily capsular polysaccharides, are subjected to genetic variation in the chicken intestine which is the main reservoir of *C. jejuni* strains (Wilson *et al.*, 2010). These changes may increase the virulence of some *C. jejuni* strains by improving their ability to invade the human intestine (Pope *et al.*, 2007). This provides an interesting link between *C. jejuni* surface polysaccharide, invasion and virulence mechanism.

1.10.9 Protein glycosylation systems in Campylobacter species

Glycosylation is a well-known phenomenon in eukaryotes. The study of surface structures for their role in bacterial virulence has also identified glycosylation as a prominent mechanism in prokaryotes. There are two glycosylation systems studied in *Campylobacter* species. The O-linked glycosylation system that glycosylates the flagella (Guerry *et al.*, 2006, McNally *et al.*, 2006a, Goon *et al.*, 2003, Thibault *et al.*, 2001, Doig *et al.*, 1996) and the N-linked glycosylation system that decorates other periplasmic proteins (Linton *et al.*, 2002, Young *et al.*, 2002, Szymanski *et al.*, 1999). The O-linked glycosylation system links the glycans to the hydroxyl oxygen of serine or threonine amino acids in the target flagellin protein. The N-linked glycosylation system modifies asparagine residues on many outer membrane proteins (Szymanski *et al.*, 1999, Szymanski *et al.*, 2003, Szymanski and Wren., 2005).

In *Campylobacter* species, O-glycan modification is an essential process for flagella synthesis and motility. This post translational modification of flagellins was first identified in *C. coli* and then this process was studied in *C. jejuni* and *C. coli* (Alm *et al.*, 1992, Logan *et al.*, 1989). There are two major O-glycans that modify flagellins in *Campylobacter* species, both having a similar structure and each composed of similar 9-carbon sugars. These saccharides are pseudaminic acid (PseAc) and legionaminic acid (LegAm) (McNally *et al.*, 2007, Schirm *et al.*, 2003, Thibault *et al.*, 2001). The major glycan coating *C. jejuni* flagellins is PseAc whereas LegAm modifies *C. coli* flagellins (Young *et al.*, 2007). In *C. jejuni* the PseAc glycans are synthesized by proteins encoded by several *pse* genes. It has been reported recently that the PseAc in *C. jejuni* have a variety of substitutions including an acetamidino, O-acetyl or N-acetylglutamine attachments (Schirm *et al.*, 2005, Thibault *et al.*, 2001). Genome sequence analysis of *C. jejuni* 81-176 revealed a group of genes encoding for the O-glycosylation system located adjacent to the flagellin synthesis genes and this region was highly variable (Guerry and Szymanski., 2008). The glycosylation at seven specific serine or threonine residues is important for the motility and autoagglutination phenotypes in *C. jejuni* (Ewing *et al.*, 2009). Since O-glycan coated flagellins are surface exposed they may play a role in promoting interaction between flagella of neighbouring bacteria and facilitate autoagglutination. The two processes; flagellin glycosylation and autoagglutination together may also allow survival of bacteria inside the host. *C. jejuni* 81-176 mutants in PseAc glycans resulted in defects in motility and autoagglutination.

In vitro adhesion and invasion of cultured epithelial cells and virulence in the ferret disease model was decreased (Ewing *et al.*, 2009, Guerry *et al.*, 2006). Another study showed that *C. jejuni* mutants in LegAm had defects in colonization of chicks (Howard *et al.*, 2009). The glycan coated flagella showed antigenic properties which suggests that the O-linked glycosylation system may have a role in survival in host cells by escaping the host immune responses (Thibault *et al.*, 2001, Guerry., 2007, Young *et al.*, 2007).

The N-linked glycosylation system in *Campylobacter* species is encoded by *pgl* genes. The *pgl* genes are conserved in *C. jejuni* and *C. coli*. The translational products of *pgl* system are multi-sugar molecules (usually 7 sugars) (Jervis *et al.*, 2012). A number of studies have suggested that the N-linked glycosylation system in *C. jejuni* is required to mediate host interactions. Mutants in *pgl* genes were unable to adhere to and invade human epithelial cells and were unsuccessful in colonizing the avian intestinal tract (Kelly *et al.*, 2006, Hendrixson and DiRita., 2004, Karlyshev *et al.*, 2004, Szymanski *et al.*, 2002). Kakuda and DiRita (2006) showed that glycosylation of the periplasmic protein Cj1496c was required for the adherence and invasion of INT-407 cells and colonization of the chicken intestinal tract. Karlyshev *et al* (2004) reported a *pglH* *C. jejuni* mutant with reduced adherence and invasion to Caco-2 cells. Similarly, mutational inactivation of the glycosylation system genes, oligosaccharide transferase (*pglB*) and putative aminotransferase (*pglE*) in *C. jejuni* 81-176 impaired its ability to adhere and invade INT-407 cells (Szymanski *et al.*, 2002). Another aminotransferase encoding gene, *cj1121c* which is a glycoprotein of N-linked glycosylation system was reported to be involved in motility and *in vitro* host cell interaction (Vijayakumar *et al.*, 2006).

Currently, over 70 N-linked glycosylation proteins have been identified in *C. jejuni* (Kowarik *et al.*, 2006, Young *et al.*, 2002). The N-linked glycosylation system modifies the VirB10 protein which is encoded on the pVir plasmid in some *C. jejuni* strains (Larsen *et al.*, 2004). VirB10 is homologous to the proteins of T4SS and is involved in DNA transformation (Bacon *et al.*, 2000). The N-linked glycans modify VirB10 at two asparagine residues providing stability to the protein. However, the glycosylation of only one asparagine residue of VirB10 is required for the expression of this protein (Larsen *et al.*, 2004). A recent study suggested that the Pgl system releases free oligosaccharides (fOS) in the periplasmic space. The amount of these fOS produced is

influenced by the salt concentration. This suggests that fOS may provide protection under osmotic stress conditions (Nothaft *et al.*, 2009).

The *C. jejuni* N-linked glycosylation system also influences the host immune system by recognizing specific lectins present on macrophages and some dendritic cells. A *C. jejuni* mutant of *pglA* promoted excessive release of IL-6 (interleukin-6) from human dendritic cells which suggests that the N-linked glycosylation system may function to enable the organism to evade the immune response during infection in human hosts (van Sorge *et al.*, 2009).

1.11 Project background and key aims of research

The ability to attach to and internalize the human epithelial cells is an essential phenomenon used by *C. jejuni* to cause an asymptomatic or diarrhoeal disease (Young *et al.*, 2007).

In a recent development to differentiate the adhesion and invasion virulence traits of clinical and non-clinical *C. jejuni* strains, Fearnley *et al* (2008) studied 74 strains of poultry origin and 39 clinical strains isolated from patients with enteritis and bacteraemia. This study categorised the *C. jejuni* strains into three classes; low invasive, highly invasive and hyperinvasive strains using INT407 cell lines, Caco-2 and HT29-Cl.16E mucous secreting cell lines. The invasion potential for these *C. jejuni* strains into INT-407 cells is presented in Figure 1.3.

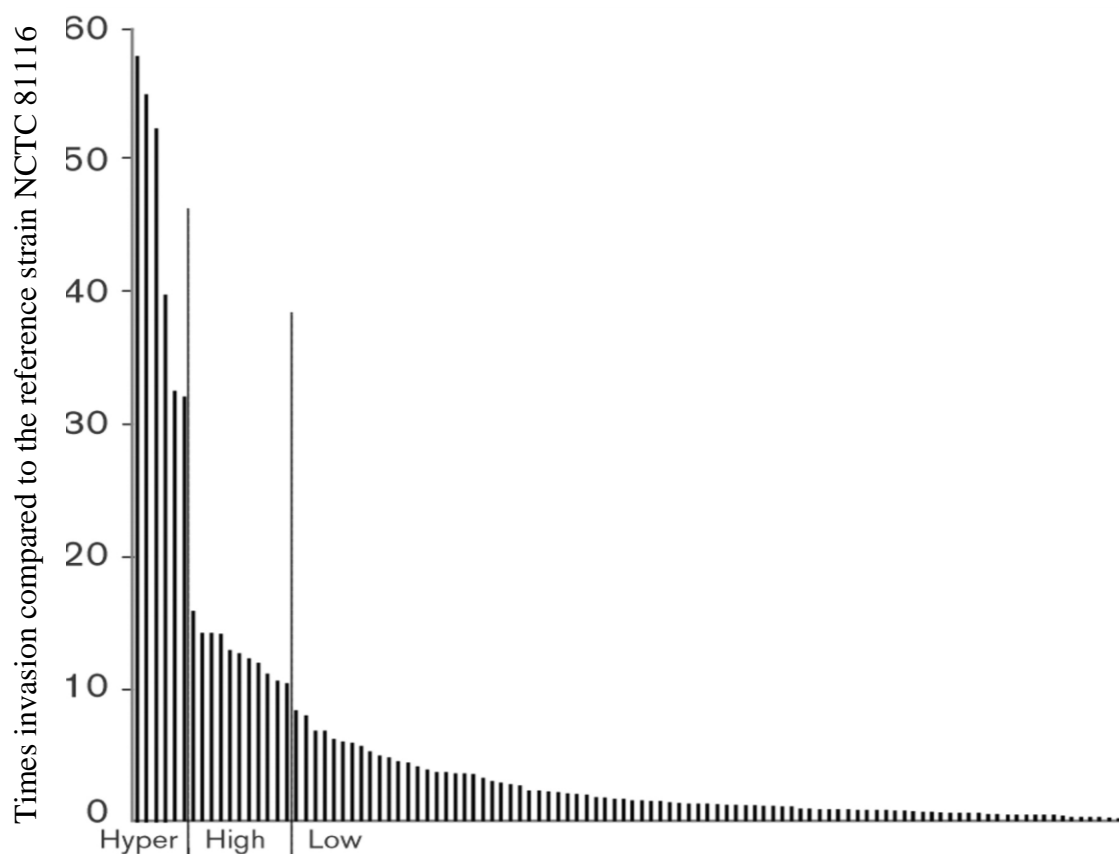


Figure 1.3: Distribution of 113 *C. jejuni* isolates tested for their ability to invade into INT-407 cells compared to the low invasive reference *C. jejuni* strain NCTC 81116. © 2012 Crown copyright.

The hyperinvasive strains were >25 times more invasive compared to the reference strain. The high invasive strains were at least 10 times as invasive and the low invasive *C. jejuni* strains were less than 10 times as invasive as the reference strain (Fearnley *et al.*, 2008).

Six clinical hyperinvasive *C. jejuni* isolates (01/10, 01/35, 01/04, 01/41, 01/51, EX114) were identified in this study which showed significantly higher levels of invasion (>25 -fold) into the human intestinal cell lines INT-407 and Caco-2 compared to the low invasive reference *C. jejuni* 81116 (Fearnley *et al.*, 2008).

A hyperinvasive *C. jejuni* strain 01/51 has been characterised in detail by transposon mutagenesis (Javed *et al.*, 2010). A transposon mutant library composed of 800 mutants was screened. An initial study identified 26 mutants with high motility and reduced invasiveness which were selected for further investigation. Six genes (*cipA*, *cj0690c*, *cj1136*, *cj1245c*, *cj1305c* and *cj1539c*) were identified with potential roles in

invasiveness and this was confirmed by targeted mutagenesis, complementation and phenotype studies. These genes with a role in invasiveness were also studied for colonization of the chick gut (Javed *et al*, 2010).

In this PhD project, all six hyperinvasive *C. jejuni* strains identified by Fearnley *et al* (2008) were studied. The initial hypothesis was that this group of strains possesses extra genetic material to make them hyperinvasive. Therefore, this study was designed to investigate the genomic basis of invasiveness in these hyperinvasive *C. jejuni* strains using advanced molecular techniques such as Comparative Genomic Hybridization (CGH), Suppressive Subtractive Hybridization (SSH) and genome sequencing.

The main aims of this project are;

- To determine differences between selected *C. jejuni* hyperinvasive strains compared with low invasive strains at the genomic level.
- To establish the function of genes of interest identified in the hyperinvasive *C. jejuni* strains.
- To relate any genomic variations identified by using the molecular techniques to the phenotypic characteristics of the strains.
- To understand more about how this major human pathogen has evolved based on the knowledge acquired in this study.

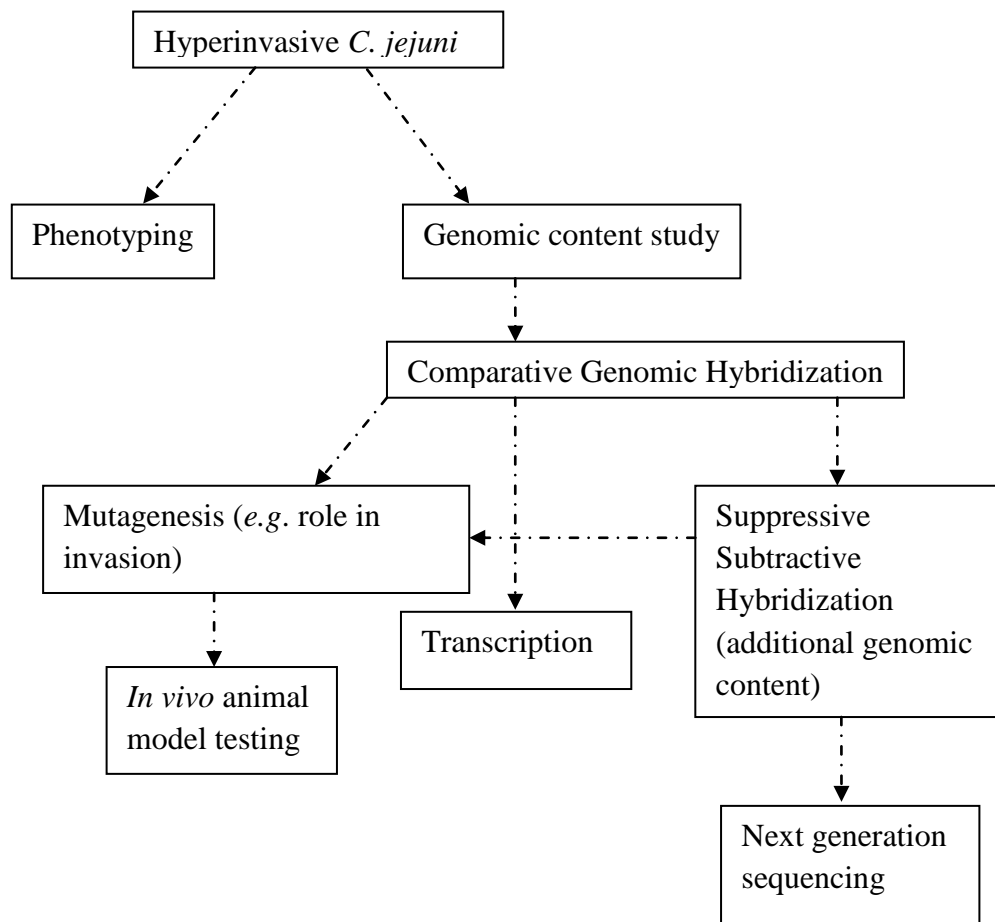


Figure 1.4: The proposed research plan for this project.

Chapter Two

MATERIALS AND METHODS

MATERIALS AND METHODS

2.1 Bacterial strains

This study employed 24 *Campylobacter jejuni* strains and *Campylobacter coli* RM2228. All *Campylobacter jejuni* isolates used in this study were obtained from the Veterinary Laboratory Agency, Weybridge, UK. *Campylobacter coli* RM2228 was provided by Prof Ian Connerton, University of Nottingham, UK (Table 2.1).

Table 2.1: The bacterial strains used in this study.

Bacterial strain	Source	Isolated from	Invasion profile*	MLST		Reference
				Clonal complex	ST complex	
01/10	Human clinical	Blood	Hyperinvasive	21	21	Fearnley <i>et al.</i> , 2008
01/35		Faecal	Hyperinvasive	21	22	
01/04		Blood	Hyperinvasive	677	677	
01/41		Faecal	Hyperinvasive	21	916	
01/51		Faecal	Hyperinvasive	21	21	
EX114		Environment (Puddle isolate from outside a boiler house in South East of England)	Hyperinvasive	682	914	
01/30			Low invasive	45	45	
01/32			Low invasive	22	22	
01/46			Low invasive	21	21	
01/39			Low invasive	21	53	
01/44			Low invasive	21	21	
01/05			Low invasive	508	508	
01/08			Low invasive	61	61	
01/11			Low invasive	45	137	
01/36			Low invasive	354	324	
C2/3			Low invasive	21	21	
01/46		Low invasive	21	21	Fearnley <i>et al.</i> , 2008 MLST-this study	
01/44		Low invasive	21	21	Fearnley <i>et al.</i> , 2008 MLST-this study	
C12/11	Poultry		Low invasive	658	908	Fearnley <i>et al.</i> , 2008
C27/14			Low invasive	257	257	
C69/2			Low invasive	443	393	
C110/4			Low invasive	433	433	
81116	Human		Low invasive	283	267	Isolated from patient suffering with diarrhoea from a water-borne outbreak in 1981 (Palmer <i>et al.</i> , 1983)
RM1221	Poultry	Chicken carcass	Not known	354	354	This is a fully genome sequenced <i>C. jejuni</i> strain (Fouts <i>et al.</i> , 2005)
<i>C. coli</i> RM2228	Poultry	Chicken	Not known	828	1063	This is the first genome sequenced <i>C. coli</i> (Fouts <i>et al.</i> , 2005)
<i>E. coli</i> TOP10F**	Not applicable	Invitrogen, UK	Not known	Not applicable		FL {lacIq Tn10 (TetR)} mcrA <i>f</i> _ϕ (mrr hsdRMS-mcrBC) <i>f</i> ³⁸⁰ lacZ <i>f</i> _ϕ M15 <i>f</i> _ϕ lac <i>f</i> '74 recA1 araD139 <i>f</i> _ϕ (ara-leu)7697 galU galK rpsL (StrR) endA1 nupG

*Invasive phenotype is based on the *in vitro* invasion potential of isolates tested using INT-407 cells. Hyperinvasive *C. jejuni* strains showed 25X invasiveness whereas the low invasive *C. jejuni* corresponded to 10X invasion potential compared to the reference *C. jejuni* 81116. ** The *E. coli* competent cells for cloning used in the pooled suppressive subtractive hybridization study.

2.2 Primers used in this study

All primers used in this study were purchased from Eurofins (MWG operon, UK) and are shown in the tables 2.2.1, 2.2.2 and 2.2.3.

Table 2.2.1: Oligonucleotides used in the Comparative Genomic Hybridization (CGH) study.

Oligonucleotide	Gene	T _m * (°C)	Sequence (5' to 3')	Expected product size (bp)
modA F	<i>modA</i>	52	TTAAAAGAATTTGAAGAAAAATT	750
modA R		52	TTCAAAGGCTAATTTTTTATC	
metF F	<i>metF</i>	52	GAGCAAATTTGGATCAATC	1005
metF R		52	TTAGCTTTAATGCTTTTTGG	
CJ_10000868 F	CJE0669	58	CACTTCCCCCTTGACTGG	490
CJ_10000868 R		58	GGAGAAGTCCACACAGAAG	
CJ_10000908 F	CJE0838	60	GTATAGTAGGGCAAATATTGC	369
CJ_10000908 R		62	CCAATAACTTTTCCAGCTTGTG	
his S F	<i>hisS</i>	48	GTAGGAGAAAGT TCTGA	1120
his S R		48	ACAAACTTTACCATTTTTG	
Pro C F	<i>proC</i>	40	ATTAATCTTGATAATAAT	575
Pro C R		44	CAGCATTAGCAATGC	
CJ_10001541 F	CJE0320	52	CTTTTAAATCAAGATGGGG	507
CJ_10001541 R		50	CACAAGATCTTTTACATC	
aat F	<i>aat</i>	52	GGAGAAAAA ATCAGTGATG	535
aat R		48	CCCATAAATTCTAAATGTT	
CCOA0033 F	CCOA0033	54	GCTAAACTTGTATAGATTGG	390
CCOA0033 R		52	CTAGCCCATGCA AGTTC	
opCcV F	CJE1112	60	GCAGGATCTTTTATTGATACAC	760
opCcV R		62	CCTATATATAAACCATCTTCCATG	
CJ_10001535 F	CJE0801	48	CGGAGTGTTTTAATGC	490
CJ_10001535 R		49	GGTGTGATAAAATTCAGG	
opCcV609 F	CJE0315	50	GAAGCT TTTGTGCATGG	750
opCcV609 R		54	CATAATCACTCTTGCTTCC	
tgt F	<i>tgt</i>	52	GCCTTTTCAAACAAAGC	950
tgt R		50	CATGATCGTTGATAAATTC	
CJE1128 F	CJE1128	56	CATGCTACCTACACATTTAG	230
CJE1128 R		58	GGAGTCTTTCCA ACACCAC	
CJE0731 F	CJE0731	56	GCGACTTGTGATGGAGTG	1700
CJE0731 R		58	GTTGCCATATTAACATTAATG	

T_m* (°C) is the melting temperature of oligonucleotides. These genes were selected for PCR screening as they were found as present (Table 4.1a) or absent/highly divergent (Table 4.1b) in the hyperinvasive *C. jejuni* strains (Chapter 4). The PCR primers were designed based on the conserved sequence of genes in all sequenced *Campylobacter*

species strains publically available of Campybd (<http://www.xbase.ac.uk/campydb/>) and NCBI (<http://www.ncbi.nlm.nih.gov/genome/campylobacter/>) unless otherwise stated.

Table 2.2.2: The primer and adaptor sequences used in Pooled Suppressive Subtractive Hybridization (PSSH) study.

Oligonucleotide	Insert/gene	Sequence (5' to 3')	T _m * (°C)	Expected product size (bp)
C1-bioF-2F	<i>bioF-2</i> (8-amino-7-oxononanoate synthase)	GGGGAATGATTATTTAAATTTAAGC	64	941
C1 bioF-2 R		GCACAGTAGGTTCTTTTATAGC	62	
C2 Res mod F	Restriction modification system	GACGCAGCAGAGTATAAAC	56	1400
C2 Res mod R		CACTAGCTCTTGCTCAG	54	
C5-ThiG F	<i>thiG</i> (thiazole synthase)	GGCAAATACGAGTTTGATTCAAG	64	700
C5-ThiG R		GTTAATGGAGAACTGCTTTAGC	62	
C6-P kinase F	Polyphosphate kinase	CGCGAACTTTCTTGGCTTCGC	66	1450
C6-P kinase R		GAAGCTAAGTGC GTTGCACGAC	68	
C8-lysC F	<i>lysC</i> (aspartokinase)	GTGGAACAAGCGTTGGAACAC	64	950
C8-lysC R		GCTCTAACTGCAAGTTCTCCG	64	
C9-mem lipo F	Membrane-associated lipoprotein	CATTAGAGGTATCCCAAGATG	60	735
C9-mem lipo R		GGTGC GGCCCTCTTCGC	60	
C10-Ferredox F	Ferredoxin	CTATTGCTTGTGGTTCTTGTATTG	64	225
C10-Ferredox R		CAAAAACGGAGTGTCCCCAC	64	
C11-Tri ester F	Tributylin esterase	CAAACCTCTAGTGTGTACACG	64	235
C11-Tri ester R		GATCGTTTCAGTATCATCGGG	62	
C13-Put reg F	Putative two component regulator	GCCTATTATGGATGGACTGG	60	445
C13-Put reg R		CTCACCCACTTTATCTCTTAC	62	
C14-MuSo1 F	Prophage MuSo1	GAATAGCAAAAAGCCCAAGCGG	64	760
C14-MuSo1 R		CACTTCTACTTTTCCTAATGTAAC	64	
C15-GDP dehyd F	Putative GDP-mannose 4,6 dehydratase	CAGGATTACAGGGCAAGTTG	62	750
C15-GDP dehyd R		CTTCAGGAAGTTTAAAAGGCTTC	62	
C20-CJE1142 F	Hypothetical protein CJE1142	GATGATGATATAGATAAACCTTATG	64	740
C20-CJE1142 R		CCACCTGCTTTAATGATAACAC	64	
C21-flaA F	<i>flaA</i> (Flagellin subunit A)	CACCAATGTTGCAGCTTTAAATGC	68	1620
C21-flaA R		CTGAACAGAATTAGCCTGTGCC	66	
C22-fkbM F	FkbM family methyltransferase	GACCTTTACCTTTTATACTTGC	60	740
C22-fkbM R		CAGGATCTGCTTTGTAATAGC	62	
C23-Esterase F	Esterase (<i>Clostridium lentocellum</i> DSM 5427)	CAATATCCAAAAGCGTATGCGC	64	445
C23-Esterase R		CACCAAACACATCTTGATAATAG	62	
C25-C8J 1243 F	Hypothetical protein (C8J_1243)	CGATTCTTGTGATGATGTAGAAC	64	1100
C25-C8J 1243 R		GATCCCTACAAGGATAAGAGATG	66	
C27-Dom prot F	Conserved domain protein	GCTTTAAACCTTAGCCTCTTTG	62	645
C27-Dom prot R		GATTAATCAAAGCATCATAATACC	62	
C28-Phage pro F	Phage uncharacterized protein	CAAATGAACAAAAGCACGAAAACAC	68	1680
C28-Phage pro R		CTTATTCTTGAAACAGAGTTAAAATG	66	
C29-unknown 1F	Unknown 1	CCTATAGGTATCCATAGAAAAG	60	235
C29-unknown 1R		GGTCTACTGGTATTTATTTAC	60	
C32-DNA dom F	Restriction modification system	GCAGGCTCGGAGAGAAAAGAAC	64	690
C32-DNA dom R	DNA specificity domain-containing	CTAATACTTTTCAGTTTCTGCTTTC	66	
C33-S trans F	Putative sugar transferase (<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902)	GTTATGATTGACTTTAGTCATCAAAG	66	1690
C33-S trans R		GAAGCTACTATATCTGGACGATG	66	

Continuous

Table 2.2.2: The primer and adaptor sequences used in Pooled Suppressive Subtractive Hybridization (PSSH) study.

Oligonucleotide	Insert/gene name	Sequence (5' to 3')	T _m [*] (°C)	Expected product size (bp)
C34-Thi syn F	Thiazole synthase <i>thiG</i> (<i>C. jejuni</i> subsp. <i>doylei</i> 269.97)	GGCAAATACGAGTTTGATTCAAGA	66	741
C34-Thi syn R		GGAGAACTTGCTTTAGCCTCG	64	
C37-50S F	50S ribosomal protein	CGCAAAATGATGAAAGGGCG	60	375
C37-50S R		CAGTATATTTTCATTTGGCTCTC	62	
C38-Unknown 2F	Unknown 2	CCTATAGGTATCCATAGAAGAG	62	229
C38-Unknown 2R		GAGAATTAATAATGAATTATGGAAC	60	
C39-Put mem F	Putative membrane protein	GAACAAAGACAAATCATAGAAGAAG	66	1699
C39-Put mem R		GTATGGTATATAGTATGTTTAGCAC	66	
C40- S kinase F	Sensor histidine kinase	CTTAGAAAGTCTTATAGAACAAAC	62	763
C40- S kinase R		CACTTTGAAAACCTTAACCTCC	60	
C41-Hypo pro F	Conserved hypothetical protein	GGAAGGGATGATGGATTTGG	60	1678
C41-Hypo pro R		CAAACCACAACATAATATATCC	58	
C42-Nic Tran F	Nicotinate phosphoribosyltransferase	CTT CTT TAG CCT TGC TTT GTG	60	1374
C42-Nic Tran R		GTT TAG GCG AGA GTT TTA CTT C	62	
C43-ISHa1675 F	ISHa1675 transposase B	GTG CAG TAA AAT ATA GAA TTT ATC C	64	574
C43-ISHa1675 R		CTT CTT GCT TCA ACG ATA GAT AG	64	
C44-Cj8486 F	Hypothetical protein Cj8486_0894c and CJ0961	CAT CTT TCA CAT ACA GAT CTT G	60	890
C44-cj8486 R		GCT TTT AAT TAA ATC AAT AAT TTG AG	62	
C45-CCO0105 F	Hypothetical protein CCO0105	GAT GCT TGA AAA TCT AAT AGC AC	62	144
C45-CCO0105 R		CAC AAA TAC CCA TAA AGA TGA G	60	
C50-flaB F	<i>flaB</i> (Flagellin subunit B)	GGA TAA ACA CCA ACA TCG GTG	62	1610
C50-flaB R		CAT TTT GCT GCA CTG CAT TAG C	64	
C51-C8J0142 F	Hypothetical protein C8J_0142	CAT CTT TGT TTG CTT TAG TAG	56	625
C51-C8J0142 R		CAA AGA AAT AGG CTC TAT CAC	58	
C52-murD F	<i>murD</i> (UDP-N-acetylmuramoylalanine--D-glutamate ligase)	CAC TTT TTG GAT ACG GAA AAA CC	64	1180
C52-murD R		CTT TAA AGA CTT TTC CAC GCT C	62	
C54-tet F	<i>tet</i> (tetracycline)	CGATCTTGTTGATAAAGATAACG	62	730
C54-tet R		GTTAAGGAATATAATTAGATTGAG	60	
C56-Met chem F	Methyl- accepting chemotaxis protein	CAAAGACTTGATAAACATAGTTC	60	1710
C56-Met chem R		GAGTTTAGAGGCACCATTTAG	60	
C57-Phage tail F	Phage tail fibre protein	GTCTTAATAGGAGGATTGGGAG	64	250
C57-Phage tail R		GGGCAGGTCTAAATACATATTTG	64	

Continuous

Table 2.2.2: The primer and adaptor sequences used in Pooled Suppressive Subtractive Hybridization (PSSH) study.

Oligonucleotide	Insert/gene name	Sequence (5' to 3')	T _m [*] (°C)	Expected product size (bp)
C62-Put S tran F	Putative sugar transferase (<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CG8486)	CGTAGTTATCCCAATCTATAATG	62	1210
C62-Put S tran R		CGAGTAAGAACATAACCTATTC	60	
M13 F(-20)	Sequencing primers for PCR @ 2.1 TOPO cloning vector (Invitrogen UK)	5'-GTAAAACGACGGCCAG-3'	Not applicable	Dependent on cloned sequence size
M13 R		5'-CAGGAAACAGCTATGAC-3'		
16S rRNA F	<i>C. jejuni</i> (MWG, Eurofins, UK)	5'-AATCTAATGGCTTAACCATTA-3'	54	852
16S rRNA R		5'-GTAAGTAGTTAGTATTCCGG-3'	58	
Adaptor 1	CLONE-TECH PCR™-Select Bacterial Genome Subtraction kit (Clontech, UK)	5'-CTAATACGACTCACTATAGGGC tcgagcggccgccccggcgaggt-3' 3'-GGCCCGTCCA-5'	Not applicable	
Adaptor 2R		5'-CTAATACGACTCACTATAGGGC agcgtggtcgcggccgaggt-3' 3'-GCCGGCTCCA-5'		
PCR primer 1		5'-CTAATACGACTCACTATAGGGC-3'	66	Not applicable
Nested primer 1		5'-tcgagcggccgccccggcgaggt-3'	68	Not applicable
Nested primer 2R	5'-agcgtggtcgcggccgaggt-3'	68	applicable	

*T_m (°C) represents the melting temperature. Where the gene was present in more than one *Campylobacter* species strain with sequences publically available on Campydb (<http://www.xbase.ac.uk/campydb/>) and NCBI (<http://www.ncbi.nlm.nih.gov/genome/campylobacter/>), the PCR primers were designed based on conserved sequence of the gene in these strains. In case of gene present in other bacteria the PCR primer pair was designed based on the sequence in respective strain only. For the unknown inserts 1 and 2, the primers were designed from the insert sequence.

Table 2.2.3: MLST PCR and sequencing primers used in this study.

Gene	PCR primers	Sequencing primers
	Sequence (5' to 3')	
<i>aspA F</i>	A1 AAAGCTGCAGCTATGGC	S3 CCAACTGCAAGATGCTGTACC
aspartate	A3 ATGAGGTTTATTATGGAGTGC	
	A9 AGTACTAATGATGCTTATCC*	
<i>aspA R</i>	A2 AAGCGCAATATCAGCCACTC	S6 TTCATTTGCGGTAATACCATC
	A4 CCTCTTTGGCTATAGAAGCTG	
	A10 ATTCATCAATTTGTTCTTTGC*	
<i>glnA F</i>	A1 TAGGAACTTGGCATCATATTACC	S1 GCTCAATTCATGGATGGC
glutamine synthetase		S3 CATGCAATCAATGAAGAAAC*
<i>glnA R</i>	A2 TTGGACGAGCTTCTACTGGC	S4 GCATACCATTGCCATTATCTCCG
		S6 TTCCATAAGCTCATATGAAC*
<i>gltA F</i>	A1 GGGCTTGACTTCTACAGCTACTTG	S1 GTGGCTATCCTATAGAGTGGC
citrate synthase		S3 CTTATATTGATGGAGAAAATGG*
<i>gltA R</i>	A2 CCAAATAAAGTTGTCTTGGACGG	S6 CCAAAGCGCACCAATACCTG*
		S8 TGCTATACAGGCATAAGGATG
<i>glyA F</i>	A1GAGTTAGAGCGTCAATGTGAAGG	S3 AGCTAATCAAGGTGTTTATGCGG
serine hydroxy methyl transferase		S5 GCTAATCAAGGTGTTTATAT**
		S7 AGCCTAATCAGGTTCTCAA**
<i>glyA R</i>	A2 AAACCTCTGGCAGTAAGGGC	S4 AGGTGATTATCCGTTCCATCGC
<i>tkt F</i>	A1 TTTAAGTGCTGATATGGTGC	S1 TGCACCTTTGGGCTTAGC
transketolase	A3 GCAAACCTCAGGACACCAGG*	S5 GCTTAGCAGATATTTTAAGTG
<i>tkt R</i>	A4 CATAGCGTGTCTCTGATACC	S4 ACTTCTTCACCCAAAGGTGCG
	A6 AAAGCATTGTTAATGGCTGC*	S6 AAGCCTGCTTGTCTTTGGC
<i>pgm F</i>	A1 TTGGAACCTGATGGAGTTCG	S3 GCTTATAAGGTAGCACCTACTG
phospho glucomutase	A3 TCAGGGCTTACTTCTATAGG	S5 GGTTTTAGATGTGGCTCATG*
	A7 TACTAATAATATCTTAGTAGG*	
<i>pgm R</i>	A2AAGAGCTTAATATCTCTGGCTTCTAG	S2 TCCAGAATAGCGAAATAAGG*
	A4 AGCTTAATATCTCTGGCTTC	
	A8 CACAACATTTTTCATTTCTTTTTC*	
<i>uncA F</i>	A3 AAAGCTGATGAGATCACTTC	S3 AAAGTACAGTGGCACAAGTGG*
ATP synthase alpha subunit	A7 ATGGACTTAAGAATATTATGGC*	S5 TGTTGCAATTGGTCAAAAGC
<i>uncA R</i>	A2 GCTAAGCGGAGAATAAGGTGG	S4 TGCCTCATCTAAATCACTAGC*
	A4 ATTCTTTGTCCACGTTCAAG	
	A8 ATAAATTCCATCTTCAAATTCC*	

The *C. jejuni* MLST primers were sourced from pubmlst.org/campylobacter/. PCR primer pairs amplify ~1kb internal region of each gene. Sequencing primers are nested inside the PCR primers and sequence ~600 bases region. Alternate primers are provided for some genes. * represent preferred primers with best results. ** are primers designed by Jonas Waldenström (Lund University, Sweden) and work best for the wild bird isolates. The melting temperature T_m (°C) for the MLST PCR primers was 50 °C.

2.3 Culture media

All culture media were obtained from Oxoid (Basingstoke, Hampshire, UK). The media were prepared in distilled water and sterilised by autoclaving at 121 °C, 15 psi for 15 minutes. The media plates were stored at 4 °C and were used within two weeks of preparation.

- mCCDA (modified *Campylobacter* Blood free selective agar base)

mCCDA was prepared in Duran bottles by weighing out 22.75 g of mCCDA agar powder dissolved in 500 ml of deionised distilled water.

Composition: nutrient broth no. 2 (25 g), amphotericin B (10 mg), bacteriological charcoal (4 g), cefoperazone (33 mg), casein hydrolysate (3 g), sodium deoxycholate (1 g), ferrous sulfate (250 mg), sodium pyruvate (250 mg), agar (12 g) added to deionized water (1 L).

- Mueller Hinton Broth* (MHB)

MHB was prepared in Duran bottles by weighing out 21 g of MHB dissolved in 1 L of deionized distilled water.

- Mueller-Hinton Agar* (MHA)

Thirty eight grammes of MHA was dissolved in 1 L of distilled water.

*composed of beef, dehydrated infusion from 300 g, starch 1.5 g, casein hydrolysate 17.5 g and agar 17.0 g per litre.

2.4 Bacterial culture maintenance and growth

All *Campylobacter* stocks were maintained at -80 °C as 1ml aliquots in 20% glycerol MHB.

The *Campylobacter* stocks from the -80 °C freezer were routinely sub-cultured on mCCDA (modified *Campylobacter* Blood free selective agar base) grown for 48 hours at 37 °C in microaerobic atmosphere (10% CO₂, 5% O₂, 85% N₂ by vol) in anaerobic jars containing a CampyGen[®] pack to generate an anaerobic atmosphere or in an anaerobic workstation (Don Whitley scientific, UK).

The *E. coli* TOP10F' competent cells were grown on Luria Bertani (LB) agar plates incubated at 37 °C for 16-20 hours.

2.5 General buffers

- 0.1 M Phosphate buffered saline (PBS) and 1 M Tris-Chloride buffer (Sigma-Aldrich UK)

PBS and 10 mM Tris-Cl buffer were purchased as a sterile ready to use product. The 1 M Tris-Cl buffer was diluted to 10 mM concentration in distilled water for use in genomic DNA extraction procedure.

- 50X Tris-acetate-EDTA buffer (TAE)

50X TAE buffer was purchased from National Diagnostics, England. It was diluted in distilled water to obtain 1X TAE buffer.

2.6 DNA extraction procedures

2.6.1 Genomic DNA extraction

The Qiagen genomic DNA extraction kit (Midi) and Qiagen genomic kit 100/G, (Qiagen, Crawley, UK) were used for the genomic DNA preparation following the manufacturer's instructions. It was critical to have high quality DNA for all molecular procedures used in this study. The following main steps were amended in the Qiagen genomic extraction procedure;

- To ensure the complete degradation of proteins and nucleases, proteinase K (3-15 units/mg) (Sigma-Aldrich, UK) was used at a concentration of 20 mg/ml and the duration of incubation was extended to 120 minutes at 37 °C.
- It was also important that the DNA was washed three times with 500 µl of 70% (v/v) ethanol to ensure complete removal of buffer salts.
- After the wash with 70% (v/v) ethanol the genomic DNA pellet was dried for 15 minutes. It was important not to over dry the DNA pellet to ensure complete solubilisation in the rehydration buffer (*i.e.*, 10 mM Tris-Cl buffer/distilled sterile water). For CGH and Illumina genome sequencing the genomic DNA was eluted in 10 mM Tris-Cl buffer whereas for the PSSH plasmid DNA sequencing and genome sequencing by pyrosequencing/454 the genomic DNA was dissolved in distilled sterile water.

2.6.2 Plasmid DNA extraction

Plasmid DNA extraction was carried out using QIAprep spin miniprep kit (Qiagen, Crawley, UK) as instructed by the manufacturer. Briefly, a single *E. coli* colony containing plasmid was picked and grown in 5ml LB broth supplemented with antibiotics (50 µg/ml kanamycin or ampicillin) for 16-18 hours with shaking at 37 °C. The bacterial pellet was re-suspended in 250 µl of buffer P1 followed by the cell lysis by addition of 250 µl of buffer P2. Buffer N3 (350 µl) was added and mixed to allow precipitation of genomic DNA and cell proteins. The suspension was applied to the silica column provided with the kit and centrifuged at 14,000×g to allow the genomic DNA to bind to the column and any precipitated proteins were removed. The column was washed with wash buffer PE and the plasmid DNA was eluted in 50 µl of water.

2.6.3 Determination of DNA concentration

The concentration and purity of plasmid and genomic DNA was determined by a NanoDrop® ND-1000 UV-Vis spectrophotometer (NanoDrop Technologies, USA) prior to use in molecular techniques. For double stranded DNA, one microliter of the sample was used to record the concentration at a wavelength of 260 nm. For purified DNA, the desired 260:280 ratio reflecting protein contamination was 1.8-1.9 and 260:230 ratio (hydrocarbon contamination) was 2.0. Both the plasmid and genomic DNA were kept frozen at -20 °C.

2.7 Polymerase Chain Reaction

2.7.1 Standard PCR

Each PCR reaction was carried out in either 50 µl, 30 µl or 25 µl volumes. Each PCR reaction mix contained 5X Green GoTaq® flexi PCR buffer (1X final concentration), MgCl₂ (25 mM) (1.0-4.0 mM final concentration), GoTaq® DNA polymerase 5 u/µl (1.25 u final concentration) (Promega, Southampton, UK), 0.7 mM dNTPs (Sigma-Aldrich, UK), 5 µM of each forward and reverse primer (Eurofins, MWG) and 15-25 ng of DNA template. Initial DNA denaturation was carried out at 94 °C for 5 minutes, followed by 30 cycles of denaturation at 94 °C for 45 sec, annealing for 45 sec based on the melting temperature (T_m°C) of the primers and extension was carried out for 1-2

minutes at 72 °C. A final extension step was performed at 72 °C for 5-10 minutes. The PCR product was visualised by agarose gel electrophoresis as stated later (section 2.7.3).

2.7.2 Colony PCR

A loopful of freshly grown bacteria was suspended in 300 µl of sterile distilled water. The suspension was boiled to 100 °C for 10 minutes to lyse the cells. The suspension was centrifuged at 15,500 rpm for 5 minutes and 1 µl of the supernatant was used in the PCR procedure as explained in section 2.7.1.

2.7.3 Agarose gel electrophoresis

DNA fragments were analysed on 1-1.5% w/v agarose gels containing SYBR[®] Safe DNA gel stain (10,000X) (1× final concentration) (Invitrogen, UK) prepared in 1×TAE buffer. The sample (10 µl) was mixed with 2 µl of 6×loading buffer (Promega, Southampton, UK) before loading on the gel. A 1 kbp or 100 bp DNA ladder (Promega, UK) was used as the DNA marker and 250 ng of the marker was loaded on the gel. The agarose gel was electrophoresed in a Mini Sub[®] Cell GT tank submerged in 1×TAE and at 90-100 V. The DNA bands were visualised under ultra-violet light using InGenius[®] gel documentation system (Syngene, UK).

2.7.4 Cloning into pCR 2.1-TOPO[®] vector

The pCR 2.1-TOPO[®] vector was used for cloning PSSH secondary PCR products which comes a part of TOPO TA Cloning[®] kit (Invitrogen, UK). This vector contains single 3'-thymidine (T) overhangs that bind with a single deoxyadenosine (A) residues added to the 3' ends of PCR products by *Taq* polymerase activity. It contains ampicillin and kanamycin resistance genes for antibiotics screening. It has M13 forward and reverse primers binding sites for sequencing and contains several restriction sites including *EcoR* I, *Hind* III and *BamH* I with endonuclease activity that can be used to determine the size of cloned PCR product.

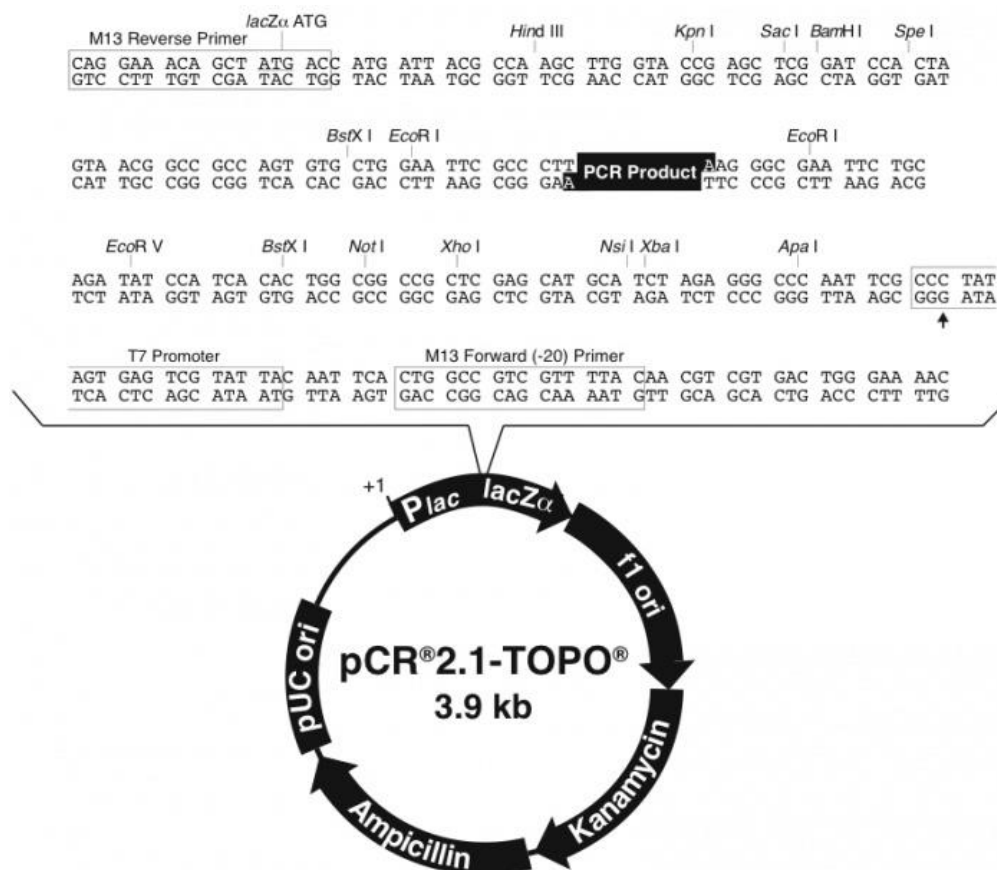


Figure 2.1: The map of pCR 2.1-TOPO[®] cloning vector (Invitrogen, UK).

Cloning was performed as described in the manufacturer's manual. Briefly, a reaction mixture was prepared containing 0.5–4 μ l of fresh PCR product, 1 μ l of salt solution made up to a total volume of 5 μ l of sterile distilled water. To this 1 μ l of pCR 2.1-TOPO[®] vector was added making up the total reaction mixture volume to 6 μ l. After gently mixing the reaction mixture, it was incubated for 30 minutes at the room temperature (22–23 $^{\circ}$ C). The ligation mixture was used to transform 10F' *E. coli* chemically competent cells. The clones with inserts were selected by blue white screening using 40 mg/ml X-gal, 100 mM IPTG (isopropyl-beta-D-thiogalactopyranoside) (Invitrogen, UK) and antibiotics 50 μ g/ml kanamycin (Sigma-Aldrich, UK) or 50 μ g/ml ampicillin (Sigma-Aldrich, UK) in LB agar plates which were incubated for 16-18 hours at 37 $^{\circ}$ C. After incubation, the light blue or white colonies were picked up and sub-cultured onto the fresh LB plates supplemented with 50 μ g/ml ampicillin or 50 μ g/ml kanamycin.

2.7.5 Heat shock transformation of 10F' *E. coli* cells

Two microliters of the ligation reaction mix was added to one 50 µl vial of One Shot® TOPO 10F' chemically competent *E. coli* cells (Invitrogen, UK) and incubated on ice for 30 minutes. The cells were heat shocked for 30 seconds at 42 °C followed by recovery in 250 µl of S.O.C medium (provided with the kit) with shaking (200 rpm) for one hour at 37 °C. The cells were plated onto antibiotic (ampicillin or kanamycin) supplemented LB plates. These plates were incubated overnight and transformed *E. coli* cells were selected.

2.7.6 Purification of PCR products

The PCR products were purified using QIAquick PCR purification kit (Qiagen, Crawley, UK) to get rid of excess nucleotides, primers and DNA polymerase and salts. The cleanup was carried out as per the manufacturer's instructions. Briefly, the PCR product was mixed with a loading buffer PB and applied to a silica spin column provided with the kit. The column was centrifuged at 14,000×g followed by two washes with wash buffer PE and the purified product was eluted in 50 µl of distilled water.

2.8 Multi locus sequence typing (MLST)

MLST was carried out as described previously by Dingle *et al.*, 2001. Briefly, seven *C. jejuni* housekeeping genes *aspA*, *glnA*, *gltA*, *glyA*, *pgm*, *tkt*, *uncA* were amplified by PCR (2.7.1) using the gene specific PCR primer listed in the table 2.2.3. Each PCR product was purified as stated in section 2.7.6. A 400-500 bp fragment of each gene was sequenced (Source Bioscience, UK) using a separate set of sequencing primers (Table 2.2.3). Based on the sequencing data each strain was assigned an allele number, sequence type (ST) and clonal complex (CC) from the *Campylobacter* MLST database (<http://pubmlst.org/campylobacter/>).

2.9 Serotyping

Six hyperinvasive and four low invasive *C. jejuni* strains were serotyped by using soluble heat-stable (HS) antigenic Penner serotyping scheme with a panel of 66 O antisera (Penner *et al.*, 1983). The serotyping was performed by Dr Judith Richardson at the *Campylobacter* reference unit, Health Protection Agency (HPA), Colindale, UK.

Chapter Three

**PHENOTYPIC
CHARACTERIZATION OF THE
HYPERINVASIVE *C. JEJUNI*
STRAINS**

PHENOTYPIC CHARACTERIZATION OF THE HYPERINVASIVE *C. JEJUNI* STRAINS

3.1 INTRODUCTION

Enteric pathogens that cause disease in humans including *C. jejuni* often have to survive in the environment as well as inside the human host. *C. jejuni* have evolved mechanisms that allow survival under both conditions.

3.1.1 *C. jejuni* virulence factors inside the host

C. jejuni have a wide host range and are present in cattle, wild birds, poultry, pigs, cattle, dogs, cats, mink, rabbit and insects (Fitzgerald and Nachamkin., 2007, Humphrey *et al.*, 2007). These bacteria are commensal in chicken and the contaminated chicken carcasses during slaughtering process are the major source of transmission of *C. jejuni* inside humans. *C. jejuni* prefer to grow under microaerobic conditions. The optimum human body temperature (37°C) and reduced oxygen conditions inside human gut offer suitable conditions for *C. jejuni* growth (Humphrey *et al.*, 2007).

During the infection process inside the human host, *C. jejuni* encounters the host immune system which provides defence against any pathogen attacks (Wooldridge and Ketley., 1997). The innate immune response to *C. jejuni* infection recruits macrophages and dendritic cells (DCs) initiating an inflammatory response at the site of infection (Hickey *et al.*, 1999, Mellits *et al.*, 2002, Zheng *et al.*, 2008). Inside the macrophages, other defence mechanisms are also active such as oxidative products, nutrient limitation and acidic pH which have detrimental effects on *C. jejuni* survival (De Melo *et al.*, 1989, Day *et al.*, 2000). These reactive oxygen molecules including hydrogen peroxide, superoxides and halogenated oxygen molecules target ribosomal RNA, nucleic acid and protein structures resulting in prompt death of bacteria (Baillon *et al.*, 1999). Research has suggested that *Campylobacters* are generally killed within 24 to 48 hours of entering the human phagocytes (Wassenaar *et al.*, 1997). On the contrary, the increasing number of enteric infections by *C. jejuni* indicates that they have developed mechanisms for survival under unfavourable environmental conditions (Mihaljevic *et al.*, 2007, Day *et al.*, 2000, Baillon *et al.*, 1999). Some of these survival mechanisms reported include transition from a viable to a viable-nonculturable state and transformation from spiral to

coccal form in which bacteria are present in a dormant state (Kelly *et al.*, 2001, Harvey and Leach., 1998). A number of stress related response systems have been related to pathogenesis in *C. jejuni*. These include global stress responses under *spoT* dependent stringent response that play a role in aerotolerance and survival in the stationary phase. The stringent response system has also been found to control some key pathogenesis mechanisms in *C. jejuni* including adhesion, invasion and intracellular survival (Gaynor *et al.*, 2005)

3.1.2 *C. jejuni* survival in environment

C. jejuni are widely dispersed in a variety of environments. *C. jejuni* are frequently isolated from food and exist as planktonic form in rivers, lakes, streams and soil (Luechtefeld *et al.*, 1982, Harvey and Leach., 1998, Baillon *et al.*, 1999, Kelly *et al.*, 2001, Martinez-Rodriguez *et al.*, 2004, Mihaljevic *et al.*, 2007, Kaakoush *et al.*, 2007). Poultry farms and slaughter houses are the major sources of *Campylobacter* species contamination (Humphrey *et al.*, 2007, Havelaar *et al.*, 2007). Also, campylobacters are frequently isolated from domestic kitchens and catering environments where they appear to stick to the work surfaces (Humphrey *et al.*, 2007). In these environments, *C. jejuni* has to withstand stresses such as atmospheric oxygen pressure, temperature and nutrition variations (Day *et al.*, 2000, Mihaljevic *et al.*, 2007). *C. jejuni* has shown to form biofilm on inert surfaces including food products (Sulaeman *et al.*, 2010). The aerobic conditions have been found to enhance biofilm formation ability of some *C. jejuni* isolates which may suggest biofilm formation as a useful adaptation for the environmental survival of these bacteria (Reuter *et al.*, 2010).

3.1.3 *In vitro* invasion assay related stresses

Adhesion to host epithelial cells is a critical step in *C. jejuni* pathogenesis that involves several adhesins present on the bacterial surface (e.g. flagella, lipooligosaccharides and capsule). These adherent bacteria may invade the host epithelial cells and develop an infection. Adhesion and invasion have been studied as important events in *C. jejuni* enteritis in a number of *in vitro* assays using human and non-human epithelial cell lines (Konkel *et al.*, 2001). These studies have used human intestinal cells (INT407) and human colon cells (Caco-2) to study adhesion and invasion of *C. jejuni* and *C. coli* (Everest *et al.*, 1992, Konkel *et al.*, 1997, Fearnley *et al.*, 2008). A clear relationship

exists between the intensity of the *Campylobacter* disease severity and the adhesion and invasion potential of strains to the epithelial cells (Russell *et al.*, 1993). *In vitro*, gentamicin protection invasion assays have been used to determine the bacterial invasion potential using these intestinal cell lines (Elsinghorst, 1994). During this assay, cells are exposed to different environmental stresses that affect the survival of bacteria while invading epithelial cells. Reactive oxygen molecules are released as the product of oxidative metabolism in eukaryotic cells that may affect the survival of invading bacteria.

In this study, the hyperinvasive and low invasive *C. jejuni* strains were tested for their growth rate, stresses encountered during the invasion assays (normal air pressure, hydrogen peroxide (H₂O₂) stress and motility) and other virulence related phenotypes such as autoagglutination activity, the ability to survive under sodium deoxycholic acid stress and biofilm formation. The aims of this study were;

- To determine if there were any other phenotypic characteristics that distinguished the hyperinvasive *C. jejuni* from the low invasive *C. jejuni* strains.
- To investigate the effects of a number of stresses that the organism would experience during *in vitro* invasion assays and to confirm that the differences seen were due to the hyperinvasion rather than the stresses.

3.2 METHODS

3.2.1 Growth curve using viable count method

The growth rate of the hyperinvasive and low invasive *C. jejuni* strains was determined. For this study, bacteria were grown on modified *Campylobacter* Blood free selective agar base (mCCDA) plates (Oxoid, UK) for 48 hours under microaerobic conditions at 37 °C. The growth from the plate was harvested using a cotton swab in 2.5 ml of sterile Phosphate Buffered Saline (PBS). A two ml volume of this suspension was used to inoculate 200 ml of pre-warmed MH broth (Oxoid, UK). A 2 ml sample was carefully removed from the above broth at time zero for viable count. The bacterial broth was then sealed in a gas jar containing a CampyGen gas pack (Oxoid, UK) and incubated with shaking (180 rpm) at 37 °C. The growth rate readings were taken during the late log phase, stationary and death phase. At time intervals 16, 20, 24, 28, 48 hours, 2 ml samples were carefully removed from the broth, serially diluted, and plated onto pre-

dried mCCDA plates. The plates were microaerobically incubated at 37 °C for 24-48 hours. After incubation *C. jejuni* colonies were identified and counted to determine the viable count.

3.2.2 Resistance to aeration stress

During the invasion assay, the bacterial cells are exposed to a variety of atmospheric air stress conditions. For this assay, the bacterial strain grown as stated above was resuspended in 2.5 ml of sterile PBS. The pre-warmed MH broth (200 ml) was inoculated with 2 ml of the bacterial suspension. The broth was microaerobically incubated at 37 °C in gas jars with a CampyGen pack with constant agitation at 180 rpm for 24 hours. After overnight incubation when the cell density would be in the order of approx 10^8 - 10^9 cfu/ml, the gas pack was removed and a time zero reading was taken and the bacterial culture was exposed to atmospheric oxygen stress by gentle shaking at 180 rpm at 37 °C. Samples were removed at hourly intervals for 6 hours. The samples were serially diluted and plated onto mCCDA plates for viable counts.

3.2.3 Sensitivity to hydrogen peroxide

The bacterial cells are exposed to the toxic effects of free oxygen radicals released by the eukaryotic cells during an invasion assay. For the assay, the bacterial growth from mCCDA plate grown under microaerobic conditions at 37 °C for 48 hours was used to prepare bacterial cell suspension in 2.5 ml of sterile PBS. The pre-warmed MH broth (200 ml) was inoculated with 2 ml of this bacterial suspension. The bacterial broth was incubated in anaerobic gas jars containing CampyGen pack with shaking at 180rpm at 37 °C for 24 hours. After overnight incubation (approx 10^8 - 10^9 cfu/ml), a time zero reading was taken for viable count after which 0.36 μ l of 30% H_2O_2 was added to the broth to give a final concentration of 0.5 mM (calculation given below). After the addition of H_2O_2 , samples were taken out at time intervals 20, 40, 60, 80, 100, 120 minutes of exposure which were serially diluted and cfu/ml calculated.

Calculation of H_2O_2 concentration

The formula used for calculating molarity of 30% H_2O_2 for use in hydrogen peroxide sensitivity assay is given below;

$$\text{Molarity} = \frac{\text{percentage} \times \text{density} \times 1000}{100 \times \text{Mol. weight}}$$

The molarity calculated for 30% H₂O₂ was 9.7 M.

For use in the hydrogen peroxide assay, a fresh stock solution of 9.7 mM H₂O₂ was prepared each time by diluting 30% H₂O₂ 1000 folds in sterile distilled water (10 µl of 30% H₂O₂ added to 10 ml of sterile distilled water). Required concentration of 0.5 mM was made by diluting 52 µl of 9.7 mM H₂O₂ stock solution in 950 µl of MH broth.

3.2.4 Motility assay

The *C. jejuni* cell suspension was prepared by harvesting bacterial growth from mCCDA plates grown for 48 hours under microaerobic conditions at 37 °C in sterile PBS. The optical density at 600 nm was adjusted to approximately 1.0 for each bacterial suspension. An aliquot of 0.1 µl of this bacterial cell suspension was used to inoculate 0.4% (w/v) MH agar plates. The plates were incubated microaerobically at 37 °C for 24 to 48 hours. After incubation the diameter of the zone of growth was measured in millimetres.

3.2.5 Autoagglutination assay

The autoagglutination assay was followed as described by Golden and Acheson (2002). The autoagglutination ability of the hyperinvasive and low invasive *C. jejuni* strains was measured by harvesting the bacterial growth from 48 hours grown mCCDA plates in sterile PBS. The absorbance at 600 nm of this bacterial suspension was adjusted to approximately 1.0. Two millilitres of this bacterial suspension was transferred to sterile glass test tubes (10 x 75 mm) and incubated undisturbed at 37 °C for 24 hours under microaerobic conditions. After incubation, 1ml of the supernatant was carefully pipetted out from the test tubes and the absorbance at 600 nm was recorded again. Autoagglutination ability was calculated as the difference between the absorbance (600 nm) measured after 24 hours and the absorbance taken at the start of incubation.

3.2.6 Resistance to sodium deoxycholic acid stress

The survival ability of hyperinvasive and low invasive *C. jejuni* strains under sodium deoxycholic acid stress was measured. A stock solution (50 mg/ml) of sodium

deoxycholic acid ($\geq 98\%$ purity) (Sigma-Aldrich, USA) was prepared in sterile distilled water. This stock solution was used to prepare a series of concentrations of sodium deoxycholic acid in mCCDA from 1-16 mg/ml. The *C. jejuni* suspension was prepared by harvesting growth from a 48 hour grown mCCDA plate into 2.5 ml of sterile PBS. This bacterial suspension was diluted 1000 folds in sterile PBS. Five microlitres of the diluted suspension was spotted onto mCCDA plates containing different concentrations of sodium deoxycholic acid. Plates were incubated for 48 hours at 37 °C under microaerobic conditions. After incubation, the presence or absence of *C. jejuni* growth on each tested concentration was observed and the Minimum Inhibitory Concentration (MIC) of sodium deoxycholic acid was recorded.

3.2.7 Biofilm study

C. jejuni grown on mCCDA plate for 48 hours under microaerobic conditions at 37 °C was used to inoculate 2.5 ml of sterile PBS. This bacterial suspension was diluted 1000 fold in MH broth. Aliquots of 150 μ l of 1000 fold diluted bacterial stock suspension and MH broth only (control) were added in the allocated columns of a 96 well micro-titre plate. The plate was placed in a plastic box containing a CampyGen pack sealed with a lid and autoclave tape to ensure microaerobic conditions. The plate was incubated without shaking at 37 °C for 2, 5 and 7 days to allow the biofilm to grow. After incubation at specified time intervals, the contents of the micro-titre plate were emptied and each well was washed twice with 200 μ l of deionised water.

All inoculated wells of the micro-titre plate were stained with 200 μ l of 0.01% (v/v) crystal violet dye (Biomérieux, Marcy l'Étoile, France) and left at room temperature for 15 minutes. The crystal violet dye was discarded and the plate was washed twice with deionised sterile water to remove excess dye. One hundred and fifty microliters of 70% ethanol was added to each well for 15 minutes at room temperature to solubilise the stain. The optical density of each well was read at 540 nm using a micro-titre plate reader.

Calculation of biofilm formed

The optical density reading for each well of 96 well plate was used to calculate the average OD_{540 nm} and amount of biofilm formed by it as follows,

Biofilm formed = OD_(540 nm) of strain investigated (test) – OD_(540 nm) of MH broth only (control)

3.3 RESULTS AND DISCUSSION

The growth profile of six hyperinvasive (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and six low invasive *C. jejuni* strains (01/30, 01/32, 01/46, 01/39, 01/44, 81116) was performed. All *C. jejuni* strains were also tested for the invasion assays related stresses (atmospheric oxygen, hydrogen peroxide, motility) and other virulence related phenotypes (autoagglutination, survival in sodium deoxycholic acid stress and biofilm formation). It was determined if the hyperinvasive *C. jejuni* strains could be distinguished from the low invasive strains based on these phenotypes. Also, to find if the invasion phenotype observed for these *C. jejuni* strains was not due to the stresses encountered by them during the invasion assays.

3.3.1 Growth rate study of the hyperinvasive and low invasive *C. jejuni* strains

The ability of *C. jejuni* strains to grow in nutrient broth (MH broth) was compared to determine if the differences observed in their invasion profile was not due to any variation in their growth rate. To achieve this, the growth profile of the hyperinvasive *C. jejuni* strains was compared with low invasive strains in MH broth overtime. A representative graph of triplicate growth experiments is shown (Figure 3.1). The readings were taken during the log phase because in invasion assays *C. jejuni* inoculums are taken from the exponential growth phase. Hu and Kopecko (1999) showed that *C. jejuni* in exponential growth phase showed maximum invasion of INT 407 cells.

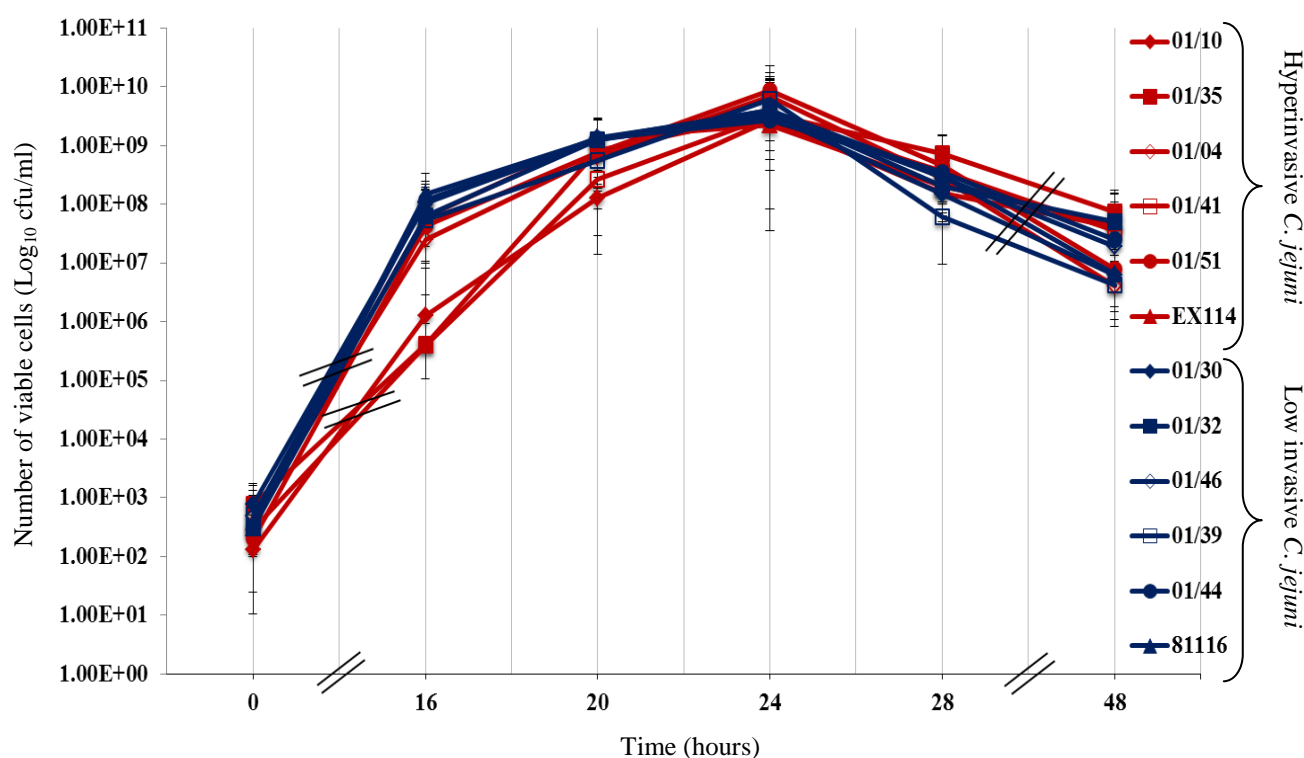


Figure 3.1: The growth curve of the hyperinvasive and the low invasive *C. jejuni* strains.

The growth rate of the hyperinvasive *C. jejuni* (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* strains (01/30, 01/32, 01/46, 01/39, 01/44, 81116) was determined in MH broth and readings were taken at different time intervals up to 48 hours and the viable count determined. The error bars represent 95% +/- CI in triplicate experiments.

It was observed from figure 3.1 that all the *C. jejuni* strains exhibited normal growth in late exponential phase under microaerobic conditions at 37 °C. The majority of *C. jejuni* strains reached 10^7 - 10^8 cfu/ml after 16 hours of growth. Interestingly, after the same growth time the hyperinvasive *C. jejuni* 01/10, 01/35 and 01/41 showed 10^5 - 10^6 cfu/ml. This suggests that these strains might be slow in adjusting to the growth medium compared to the other strains. This growth rate variation in early log phase is not an influencing factor to invasion as all the strains showed progressive growth up to 24 hours ($\sim 10^9$ cfu/ml). This was followed by a rapid decline in growth up to 48 hours. Generally, the growth profile of bacteria is divided into four phases. During the initial lag phase bacteria adjust to the new environment of the growth medium which is

followed by the exponential growth or log phase. The bacteria then enter into a prolonged stationary phase and finally the death phase. The *C. jejuni* strains studied here did not show a classic stationary phase. The lack of a stationary phase in *C. jejuni* strains has been reported in previous studies (Kelly *et al.*, 2001).

Ideally the growth study should be performed in MHA/MHB biphasic medium as it has been used as a preferred medium in invasion assays in previous studies (Hu and Kopecko., 1999, Gaynor *et al.*, 2004, Kakuda and DiRita., 2006). Recently, Javed *et al* (2010) reported that the hyperinvasive *C. jejuni* 01/51 showed same growth rate in both the MH broth and Mueller-Hinton agar/Mueller Hinton broth (MHA/MHB) biphasic medium. This suggests that the MH broth can also be used to find growth phases over time for *C. jejuni* strains for use in the invasion assay. This experiment showed that the inoculum taken from 24 hours grown *C. jejuni* cultures were suitable for use in invasion assays. Javed *et al* (2010) selected a 22 hours grown *C. jejuni* 01/51 culture in (MHA/MHB) biphasic medium for use in the invasion assay.

There was no distinct grouping of the hyperinvasive *C. jejuni* strains separate from the low invasive strains in this experiment.

3.3.2 Survival of the hyperinvasive and low invasive C. jejuni strains under atmospheric stress

C. jejuni are microaerobic bacteria that cannot survive under normal atmospheric pressure (Luechtefeld *et al.*, 1982, Ketley., 1997, Mihaljevic *et al.*, 2007). Recently, Kaakoush *et al* (2007) defined *C. jejuni* as an obligate microaerophile. Cold temperature treatment and exposure to aeration are considered as the control measures for eliminating *C. jejuni* from poultry (Kelana and Griffiths., 2003). Also, during *in vitro* assays *C. jejuni* are exposed to a number of stresses. For example, during gentamicin protection invasion assay after the epithelial cell monolayers are infected with bacteria they are incubated under 5% (v/v) CO₂ to allow bacteria to invade cells. *C. jejuni* are exposed to the atmospheric oxygen when cell monolayers are washed with PBS and later treated with gentamicin to remove any external bacteria that have not invaded the cells. Finally, the epithelial cells are treated with a detergent, Triton-X100 to lyse the cells. All these stresses could introduce bias in invasion assays. In this study, the hyperinvasive and the low invasive *C. jejuni* strains were exposed to the atmospheric air stress for a period of upto 6 hours in MH broth with constant shaking at 180 rpm at 37

°C. A representative graph of triplicate experiments showing survival of strains under atmospheric air stress is presented in figure 3.2.

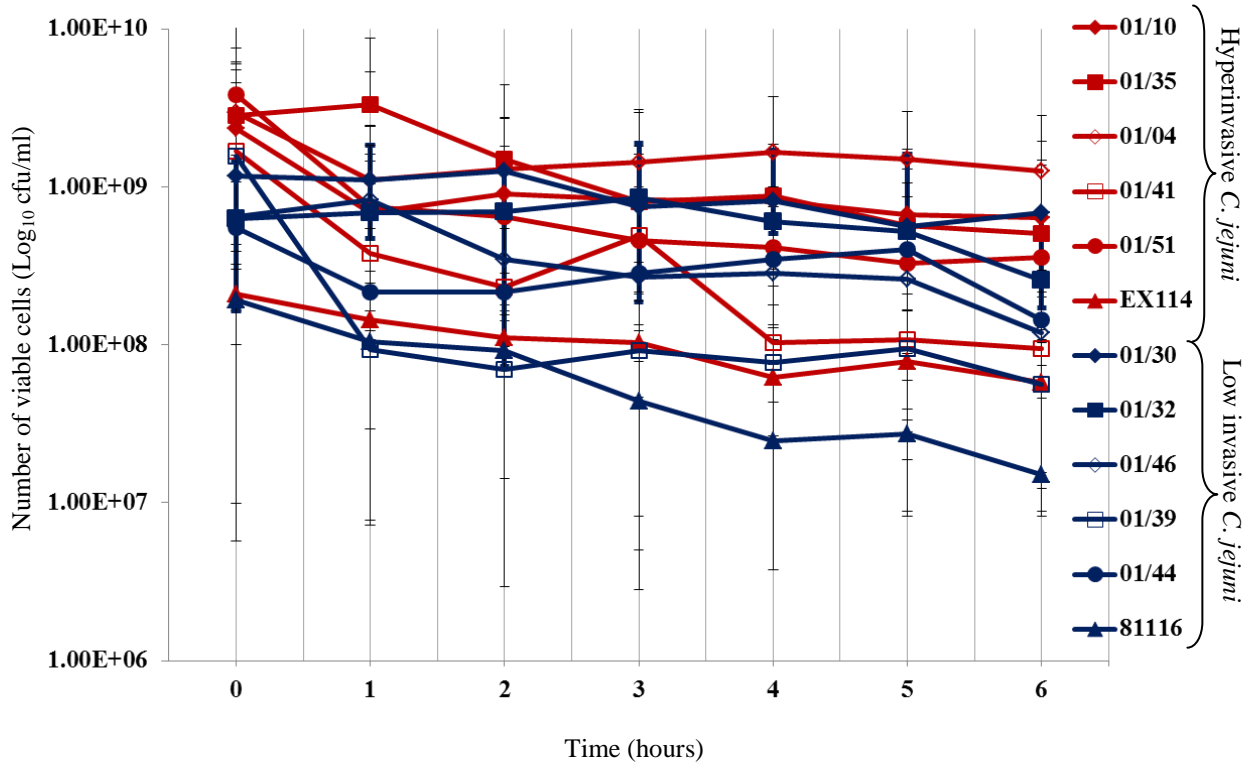


Figure 3.2: The resistance to atmospheric air stress study of hyperinvasive and low invasive *C. jejuni*.

Response of the hyperinvasive *C. jejuni* (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* strains (01/30, 01/32, 01/46, 01/39, 01/44, 81116) to aeration stress was performed by exposing the *C. jejuni*, grown cultures in MH broth, to atmospheric air and samples were taken at hourly intervals up to 6 hours and the viable cell count determined. The error bars represent 95% +/-CI in triplicate assays.

Ideally, this experiment should be performed in the tissue culture medium and 5% (v/v) CO₂ rather than full atmospheric to evaluate the actual effects of these conditions on bacteria during gentamicin protection assay but the aim of this study was not to use *C. jejuni* strains in invasion assays as the invasion phenotype of these strains was already characterized by Fearnley *et al* (2008). It would be interesting to assess if the hyperinvasive *C. jejuni* would behave differently from the low invasive *C. jejuni* strains in response to the normal atmospheric air. Figure 3.2 showed that both the hyperinvasive and the low invasive *C. jejuni* strains survived upto 6 hours of air exposure but showed variability in their response to air stress. However, no grouping of

the hyperinvasive *C. jejuni* separate from the low invasive strains was observed based on their response pattern to atmospheric stress. After one hour of atmospheric exposure there was an initial drop in the viable cell number observed for the hyperinvasive *C. jejuni* 01/10, 01/51, 01/41 and the low invasive *C. jejuni* strain 01/39. This reduction was most prominent for *C. jejuni* 01/39 where the number of viable cells decreased by one log. This initial drop in the viable cell numbers may be the time required by these strains to adjust to the air stress and turn on their atmospheric response system. The other *C. jejuni* strains where this reduction in viable cell counts was not seen may reflect that they are more stable strains and adapt better to the atmospheric stress. The *C. jejuni* cultures grown for upto 6 hours but under microaerobic conditions would be a useful control for the aeration stress study but it was not performed.

The survival of all *C. jejuni* isolates in this study under the normal atmospheric air suggests the aerobic adaptation of *C. jejuni* strains. In a previous study, *C. jejuni* were exposed to air for 2-3 days on blood agar. After this prolonged air exposure the bacteria were sub-cultured and grew normally under air. This aerobic adaptation of *C. jejuni* was accompanied by the change in cell morphology to coccid shape and changes in the expression pattern of some outer membrane proteins suggesting that *C. jejuni* adapts to aerobic metabolism when outside the host body such as in food and farm environments (Jones *et al.*, 1993). Later, Harvey and Leach (1998) also reported that *C. jejuni* avoids oxidative damage by cell surface changes as reflected in the coccal cell morphology. Another study investigated the use of aeration stress as a control measure for reducing the number of *C. jejuni* in poultry semen (Cole *et al.*, 2004b). Cole *et al.* (2004b) reported that aeration was unable to reduce *C. jejuni* from poultry semen samples suggesting that they survived in air. *C. jejuni* cultures grown in MH broth for 24 hours with the maximum cell density of $10^8 - 10^9$ cfu/ml were selected for this aeration stress study. The high bacterial cell density might facilitate microaerophilic bacteria to withstand the aeration stress. In addition, in the liquid bacteriological media the solubility of oxygen is relatively low with nutrient rich conditions that might provide favourable conditions for the bacteria to survive. *C. jejuni* strains have been reported previously to show marked resistance when exposed to air in high cell densities (Kelly., 2005, Mohammad *et al.*, 2005, Kaakoush *et al.*, 2007) and in liquid growth media (Mohammad *et al.*, 2005).

3.3.3 Sensitivity to hydrogen peroxide (H₂O₂) of hyperinvasive and low invasive C. jejuni strains

Inside the eukaryotic cells, *C. jejuni* are exposed to the reactive oxygen species released as bi-products of aerobic respiration such as superoxides and hydrogen peroxide. Hydrogen peroxide acts as a bactericidal agent reacting with reduced iron, products of the nitric oxide synthase activity, to form toxic intermediates including hypochlorous ions, hydroxyl radicals, hydroxide anions and nitrogen dioxide (Day *et al.*, 2000).

The hyperinvasive and the low invasive *C. jejuni* strains were tested to determine if the two groups of strains showed same level of response to hydrogen peroxide stress. *C. jejuni* strains were exposed to 0.5 mM hydrogen peroxide solution in MH broth and the number of surviving bacteria was determined every 20 minutes for up to 120 minutes. All the *C. jejuni* strains survived exposure to 0.5 mM hydrogen peroxide with viable cells recovered after 120 minutes exposure. A representative graph of triplicate experiments showing response of each strain to 0.5 mM hydrogen peroxide is plotted (Figure 3.3). The low invasive *C. jejuni* 01/39, however, showed a 6 log reduction in viable cell count after 120 minutes which suggest that this strain was relatively sensitive to 0.5 mM H₂O₂ stress. There was no grouping of the hyperinvasive separating them from the low invasive *C. jejuni* strains based on their response to 0.5 mM H₂O₂.

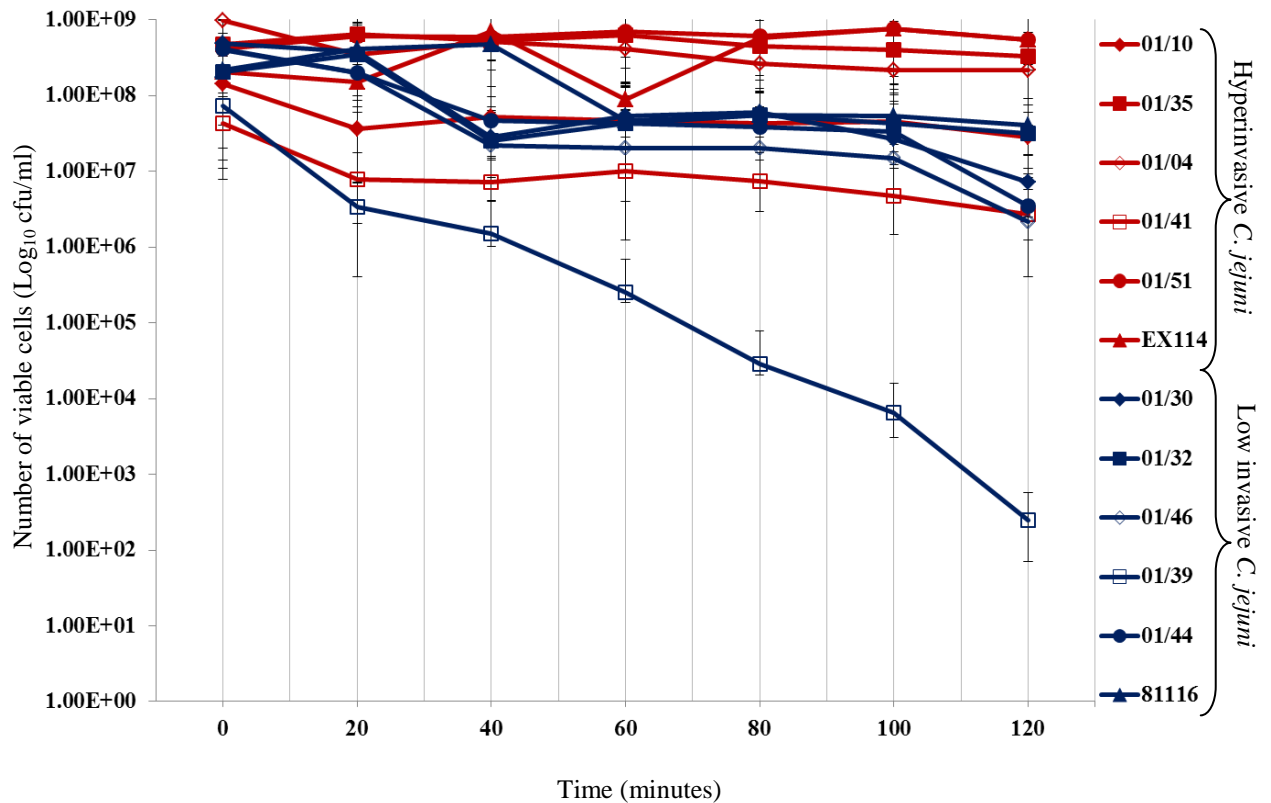


Figure 3.3: The sensitivity to hydrogen peroxide study of the hyperinvasive and the low invasive *C. jejuni* strains.

The hyperinvasive (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* (01/30, 01/32, 01/46, 01/39, 01/44, 81116) strains were grown for 24 hours in MH broth and exposed to 0.5 mM H₂O₂. The samples were taken at different time intervals up to 120 minutes for viable counts and the response of hyperinvasive and low invasive *C. jejuni* strains to 0.5 mM hydrogen peroxide was studied. The error bars represent 95% +/-CI in triplicate assays.

In a previous report, the *C. jejuni* strains were reported as resistant to 1 mM hydrogen peroxide over a sixty minutes exposure period which is in agreement with our results (Day *et al.*, 2000). This resistance was attributed to the protective activity of catalase which is encoded by the *katA* gene (van Vliet *et al.*, 1999, Day *et al.*, 2000). The *C. jejuni* strains with catalase activity degraded H₂O₂ allowing them to survive the oxygen burst inside macrophages (Day *et al.*, 2000). An ankyrin containing protein, Cj1386 (Flint *et al.*, 2012) located downstream to KatA regulon allows intracellular survival of *C. jejuni* by trafficking heme to catalase. All the hyperinvasive and low invasive *C. jejuni* strains used in this study were tested for the catalase activity. Catalase test was performed by dipping a small bacterial colony on a plastic loop into 30% H₂O₂

solution and the release of bubbles showed catalase activity. All the *C. jejuni* strains were positive for catalase activity including *C. jejuni* 01/39 which showed a dramatic reduction in viable cells when exposed to 0.5 mM H₂O₂ for 120 minutes. This suggests that the relative sensitivity of *C. jejuni* 01/39 to H₂O₂ may be related to the oxygen stress response systems in *C. jejuni* other than catalase. Iron dismutase reductases (SodB) in *C. jejuni* have been reported to provide oxygen resistance (Pesci *et al.*, 1994) that allowed intracellular survival of *C. jejuni* in INT407 cell lines. Baillon *et al* (1999) reported an iron containing hydroperoxide reductase (AhpC) that provides aerotolerance and resistance to oxidative stress in *C. jejuni*. Studies by Ishikawa *et al* (2003) demonstrated the activity of an iron based protein (Dps) that confers resistance to *C. jejuni* against the toxic effects of hydrogen peroxide by scavenging free iron particles present inside cells. In contrast, Velayudhan *et al* (2004) and Wainwright *et al* (2005) have attributed the strict microaerophilic nature of *C. jejuni* to the presence of active oxygen sensitive enzymes in *C. jejuni* such as L-serine dehydratase and rubredoxin oxidoreductase.

3.3.4 Motility profile of hyperinvasive and low invasive *C. jejuni* strains

The flagellar motility in *C. jejuni* has been proved as a key virulence factor involved *in vivo* invasion of the human intestinal cell line models (Morooka *et al.*, 1985, Grant *et al.*, 1993, Guerry., 2007). Motility is responsible for the invasion and internalisation of *C. jejuni* (Grant *et al.*, 1993) but not the only factor required for *C. jejuni* pathogenesis (Wassenaar *et al.*, 1991). The motility profile of the hyperinvasive and low invasive *C. jejuni* strains was tested to determine if the reduced invasion potential of the low invasive *C. jejuni* strains (Fearnley *et al.*, 2008) was due to the reduced motility of these strains. The *C. jejuni* strains showed a lot of variability in motility profile when observed after 48 hours of microaerobic incubation at 37 °C (Figure 3.4).

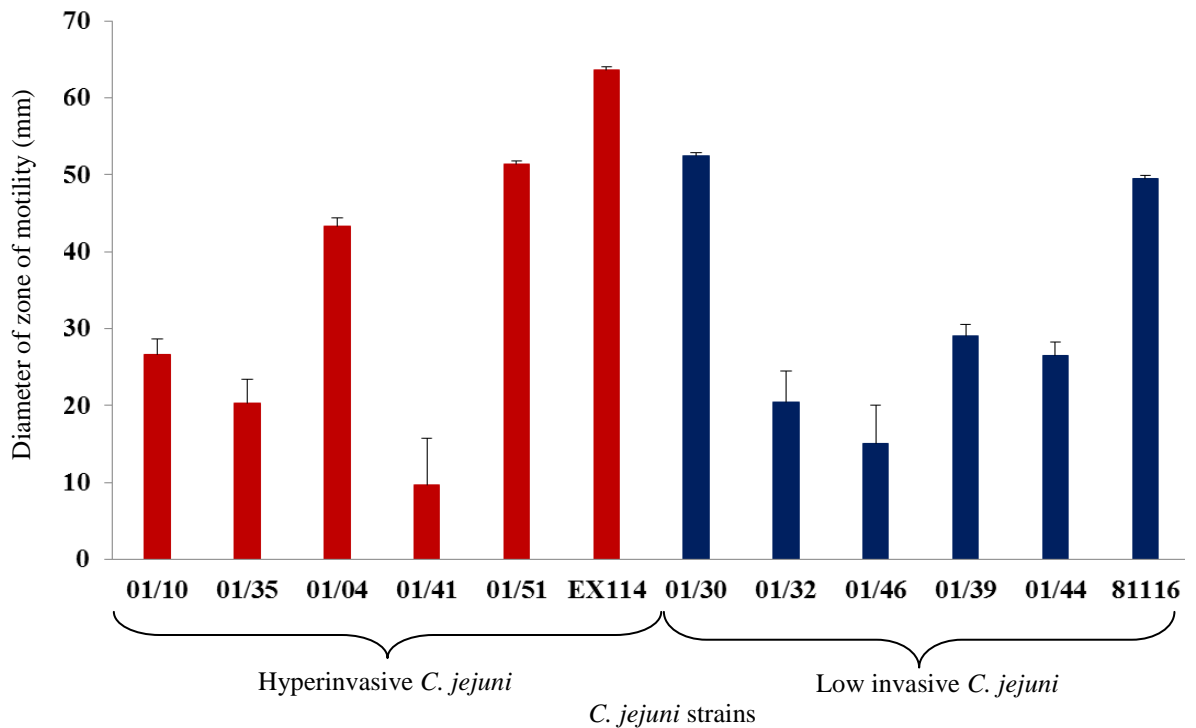


Figure 3.4: The comparison of motility zones (millimeters) for the hyperinvasive and low invasive *C. jejuni* strains measured after 48 hours of microaerobic incubation at 37°C.

The motility profile of the hyperinvasive *C. jejuni* (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* (01/30, 01/32, 01/46, 01/39, 01/44, 81116) was measured as the growth zone diameters in millimetres after 48 hours of microaerobic incubation. The error bars indicate 95% CI in triplicate experiments.

Among the hyperinvasive *C. jejuni* strains, EX114 was most motile followed by *C. jejuni* 01/51 and 01/04 whereas 01/41 showed lowest motility. Within the low invasive *C. jejuni* strains, 01/30 and 81116 displayed highest motility while 01/46 had reduced motility. In addition, there was no clear grouping of the hyperinvasive *C. jejuni* strains as a distinct group based on the motility phenotype. Fearnley *et al.*, 2008 tested motility for the hyperinvasive *C. jejuni* EX114 and the low invasive 81116 using semi-solid media. Both strains showed a diameter of growth zones varying between 50-58 mm and were classed as fully motile. The average diameter of growth zones recorded in our study for *C. jejuni* EX114 and 81116 was 64 mm and 50 mm respectively which is very similar to those reported by Fearnley *et al* (2008).

3.3.5 Autoagglutination activity of hyperinvasive and low invasive *C. jejuni* strains

The autoagglutination (AAG) activity has been recognised as a key virulence factor responsible for the host cell interaction in many Gram negative pathogenic bacteria (Misawa and Blaser., 2000) particularly in the *C. jejuni* closely related bacteria *Helicobacter pylori* (Cole *et al.*, 2004a). Autoagglutination activity in *C. jejuni* has also been reported in the literature (Golden and Acheson., 2002, Misawa and Blaser., 2000, Guerry *et al.*, 2006). In this study, most of the *C. jejuni* strains autoagglutinated (AAG) after undisturbed microaerobic incubation at 37 °C for 24 hours except for the low the invasive *C. jejuni* strains 01/39 and 81116 (Figure 3.5). The AAG activity of the low invasive *C. jejuni* 01/39 and 81116 was significantly lower ($p=0.000384$ and 0.000689 respectively) when compared against the low invasive *C. jejuni* 01/46 which showed highest autoagglutination activity in this study. However, there was no grouping of the hyperinvasive *C. jejuni* strains based on AAG phenotype.

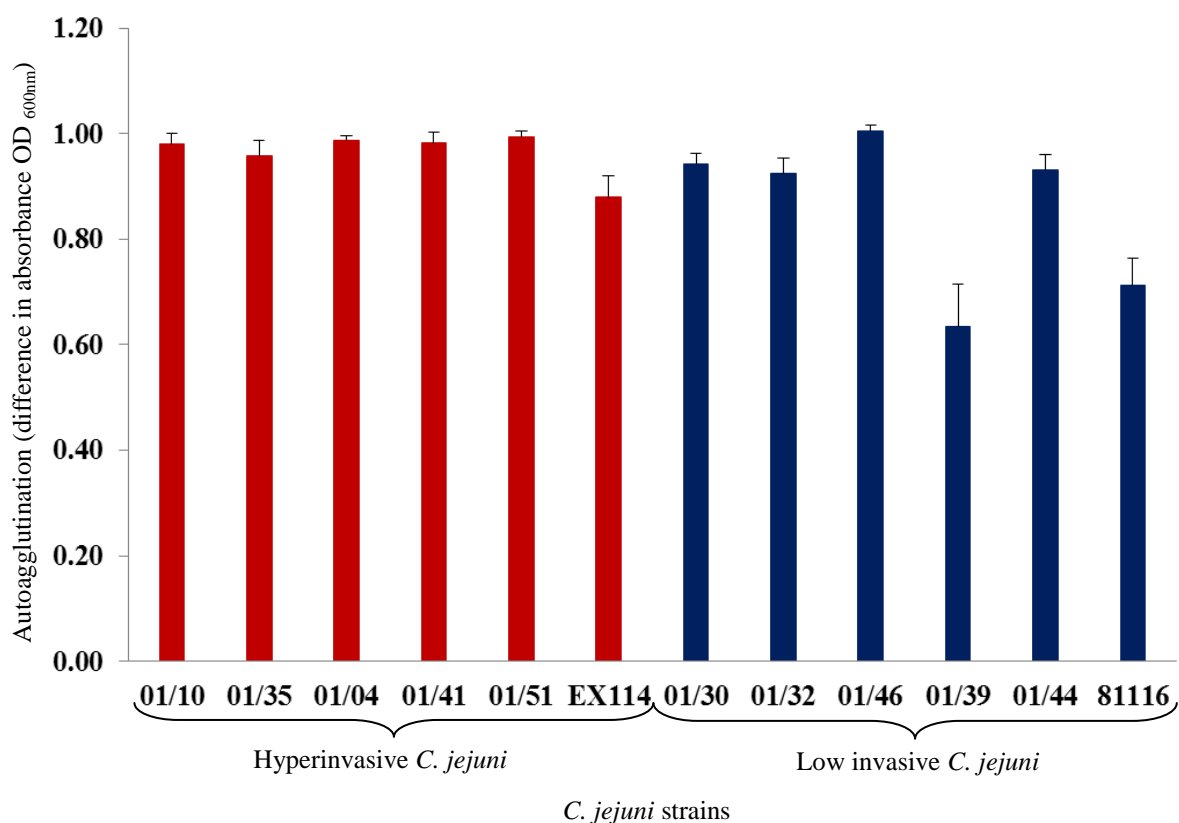


Figure 3.5: The autoagglutination ability of the hyperinvasive and the low invasive *C. jejuni* strains.

The ability to autoagglutinate in the hyperinvasive *C. jejuni* (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* strains (01/30, 01/32, 01/46, 01/39, 01/44, 81116) was determined by measuring the difference in absorbance at OD_{600nm} of *C. jejuni* cultures incubated overnight at 37 °C. The error bars show 95% CI in triplicate experiments. The statistical significance was calculated by using the Student's T-test ($p=0.001$) (Microsoft[®] Excel 2010).

The autoagglutination (AAG) characteristic of *C. jejuni* strains has been found to have an essential role in bacterial adherence to INT407 cell lines, hence linked to *C. jejuni* virulence (Misawa and Blaser., 2000). Misawa and Blaser (2000) and Guerry (2007) also reported that intact flagella were required for autoagglutination in *C. jejuni* and the aflagellate mutants of *C. jejuni* lack autoagglutination ability. In this study, the hyperinvasive *C. jejuni* 01/41 showed reduced motility (Figure 3.4) but retained high levels of autoagglutination (Figure 3.5). A similar trend was observed for the hyperinvasive *C. jejuni* 01/35, and the low invasive *C. jejuni* strains 01/32 and 01/46. By contrast, the hyperinvasive *C. jejuni* (01/51 and EX114) and the low invasive *C. jejuni* strain 01/30 showed high motility profile (figure 3.4) and high AAG activity.

The low invasive *C. jejuni* strain 81116 was fully motile but displayed lower level of AAG activity. This suggests that the findings of Misawa and Blaser (2000) and Guerry (2007) could not be truly compared with the autoagglutination and motility experiment results recorded in this study.

3.3.6 Survival of the hyperinvasive and the low invasive *C. jejuni* strains under sodium deoxycholic acid stress

Enteric pathogens are exposed to a number of antimicrobial agents in the intestine including bile salts and gastric acid. *C. jejuni* have been found to be resistant against the damaging effects of bile salts (Raphael *et al.*, 2005). The sensitivity of the hyperinvasive and low invasive *C. jejuni* strains to bile salts was tested at a range of concentrations from 16 mg/ml to 1 mg/ml of sodium deoxycholic acid. All the hyperinvasive and low invasive *C. jejuni* isolates studied exhibited similar levels of resistance to sodium deoxycholic acid with all strains showing Minimum Inhibitory Concentration (MIC) at 7.5 mg/ml except for the hyperinvasive *C. jejuni* 01/04 that showed inhibition at a lower concentration of 2 mg/ml (Table 3.1). There was no grouping of the hyperinvasive *C. jejuni* separate from the low invasive *C. jejuni* strains based on their response to sodium deoxycholic acid stress.

<i>C. jejuni</i> strain	Invasion profile	Threshold inhibitory sodium deoxycholic acid concentration (mg/ml) n=3
01_10	Hyperinvasive	7.5
01_35		7.5
01_04		2
01_41		7.5
01_51		7.5
EX114		7.5
01_30	Low invasive	7.5
01_32		7.5
01_46		7.5
01_39		7.5
01_44		7.5
81116		7.5

Table 3.1: The Minimum Inhibitory Concentration (MIC) of sodium deoxycholic acid for *C. jejuni* isolates.

The response of hyperinvasive (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* (01/30, 01/32, 01/46, 01/39, 01/44, 81116) strains to the sodium deoxycholic acid stress was investigated. The concentrations of sodium deoxycholic acid tested ranged between 16 mg/ml to 1 mg/ml. The assay was performed in triplicate.

All the *C. jejuni* strains used in our study were human clinical isolates except the hyperinvasive *C. jejuni* EX114 (Table 2.1) which is an environmental isolate. A previous study reported that the clinical *C. jejuni* isolates were more resistant to sodium deoxycholate stress than the poultry isolates. The clinical *C. jejuni* isolates survived under a much higher concentration (16 mg/ml) of sodium deoxycholate (Van Deun *et al.*, 2007). In contrast, our study showed that the human clinical *C. jejuni* isolates were more sensitive at a much lower concentration of sodium deoxycholic acid. This suggests that the response of *C. jejuni* strains to bile salts in the human intestine is not dependant on source of strain isolation. Also, the growth of majority of *C. jejuni* strains was inhibited at the same sodium deoxycholic acid concentration showing that this phenotype does not influence invasion.

3.3.7 Biofilm formation in the hyperinvasive and low invasive *C. jejuni*

A bacterial biofilm is a stable and complex structure formed on a living or non-living surface with different cohesive forces strengthened by exopolysaccharides (EPS) (Shirtiliff *et al.*, 2002, Dunne., 2002). The first step in biofilm formation is the “primary adhesion” of bacteria to any surface (biotic or abiotic). The primary adhesion is a reversible attachment of bacteria to a surface that requires the surface to be suitable for bacterial attachment and growth termed “surface conditioning”. The final step in biofilm formation called “secondary adhesion” which is the anchoring or irreversible locking phase of bacterial attachment to the surface. As the stage of secondary adhesion progresses, other planktonic microorganisms and materials in the surrounding environment can also stick to surface bound bacteria forming an aggregate on the substratum. Once the process of irreversible attachment of bacteria completes the overall density and complexity of the bacterial community may start to increase forming mature biofilm. At some point, when the biofilm reaches a critical mass, a dynamic equilibrium is reached at which the cells in closest contact with the surface closest to the

substratum begin to die due to the lack of nutrients or perfusion, pH changes, oxygen or an accumulation of a toxic by-product (Shirtliff *et al.*, 2002).

The biofilm formation in *C. jejuni* has been reported (Joshua *et al.*, 2006, Kalmokoff *et al.*, 2006, Reeser *et al.*, 2007). Joshua *et al.* (2006) reported three forms of biofilm formed by *C. jejuni* in liquid media *i.e.* attachment to the glass surfaces, unattached clumps (flocs) formation, pellicle formation at the liquid gas interface. *C. jejuni* biofilm formation on a variety of other abiotic surfaces such as polyvinyl chloride plastic, acrylonitrile butadiene etc has also been shown (Reeser *et al.*, 2007). The biofilm mode of life provides resistance to bacteria under stress conditions such as the antimicrobial agents, oxygen and nutritional pressures (Joshua *et al.*, 2006, Kalmokoff *et al.*, 2006). Motility (Joshua *et al.*, 2006, Kalmokoff *et al.*, 2006, Reeser *et al.*, 2007) and quorum sensing (Reeser *et al.*, 2007) influenced biofilm phenotype in *C. jejuni*. The molecular analysis of *C. jejuni* biofilm revealed upregulation of genes responsible for stress control, protein synthesis, catabolism and energy generation (Kalmokoff *et al.*, 2006).

In this study, the hyperinvasive and low invasive *C. jejuni* strains were allowed to form biofilm in 96 well plastic tissue culture plates undisturbed under microaerobic conditions at 37 °C and the amount of biofilm formed was studied at different time intervals (Figure 3.6). The biofilm formation potential of the hyperinvasive and the low invasive *C. jejuni* strains was very variable with no grouping of the hyperinvasive *C. jejuni* strains separate from the low invasive *C. jejuni* strains.

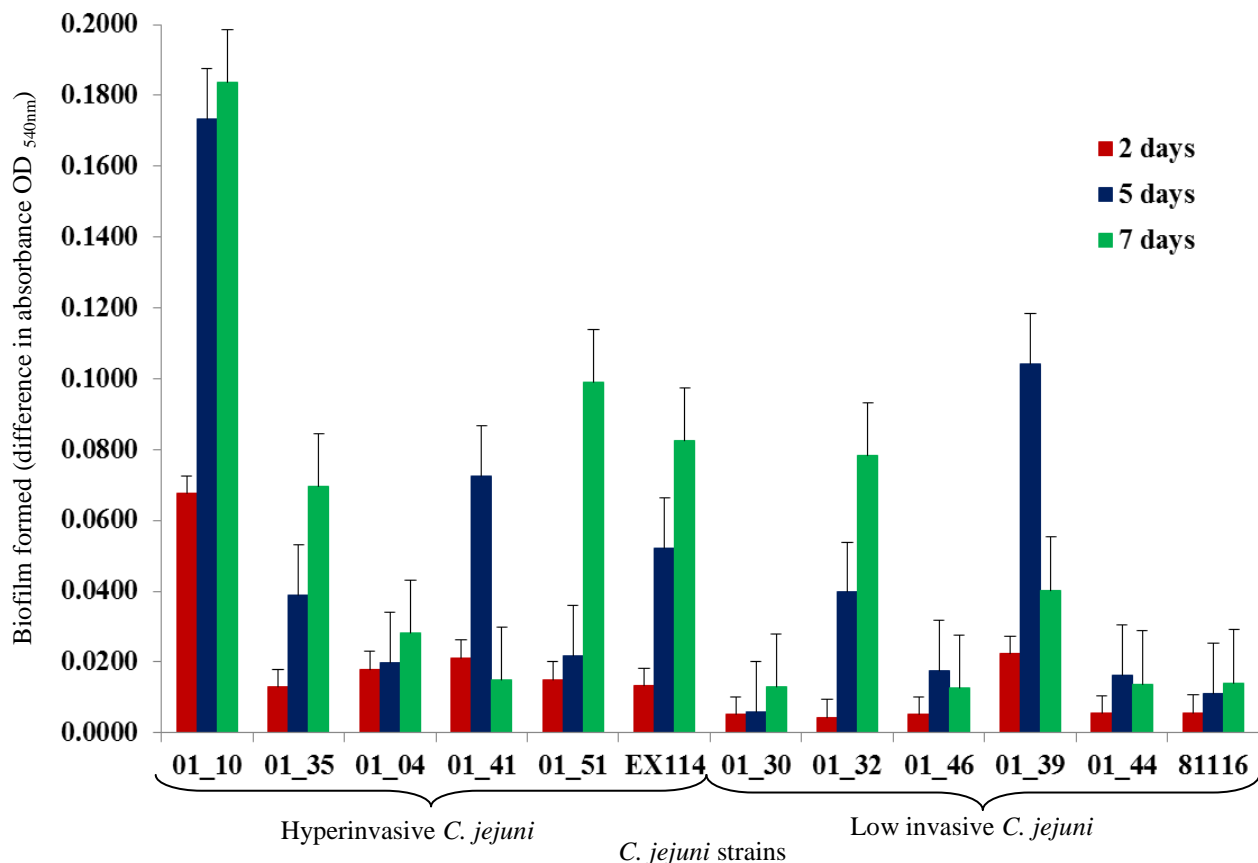


Figure 3.6: The percentage (%) mean biofilm formation of hyperinvasive and low invasive *C. jejuni* isolates at different time intervals.

The biofilm formation ability of the hyperinvasive (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and low invasive *C. jejuni* strains (01/30, 01/32, 01/46, 01/39, 01/44, 81116) was investigated. The error bars indicate 95% CI in triplicate experiments.

After 2 days of static microaerobic incubation at 37 °C the hyperinvasive *C. jejuni* 01/10 was the only strain which formed a lot of biofilm while the other strains showed only very low levels of biofilm formation. This suggests that the majority of *C. jejuni* strains were in early stages of biofilm development after 2 days. The biofilm formation progressed for all *C. jejuni* strains to 5 days of undisturbed microaerobic incubation. The low invasive *C. jejuni* 01/39 showed the highest increase in the amount of biofilm formed followed by *C. jejuni* 01/10 and *C. jejuni* 01/41 compared to other strains after 5 days period.

There was no link between the motility phenotype (Joshua *et al.*, 2006, Kalmokoff *et al.*, 2006, Reeser *et al.*, 2007) and the amount of biofilm formed in our study as *C. jejuni* 01/41 showed remarkably reduced motility (Figure 3.4) but still formed biofilm.

Similarly, *C. jejuni* 01/10 and *C. jejuni* 01/39 were only moderately motile but showed high amount of biofilm formed. After 7 days of microaerobic incubation, 4 out of 12 *C. jejuni* strains (01/41, 01/46, 01/39, 01/44) showed a decrease in the amount of biofilm whereas the rest of strains continued to form biofilm. This might suggest that the bacterial cells reached dynamic equilibrium in the mature biofilm (Shirliff *et al.*, 2002) and began to die and detached themselves which was evident by the decrease in the amount of biofilm formed for some *C. jejuni* strains. Also, this experiment suggested that the *C. jejuni* strains achieved biofilm maturity at different times.

3.4 CONCLUSIONS AND FUTURE WORK

The hyperinvasive and the selected low invasive *C. jejuni* strains were tested in the invasion assays related stress factors (response to atmospheric air, H₂O₂ and motility) and the other virulence related phenotypes (autoagglutination, sodium deoxycholic acid stress and biofilm formation). All the *C. jejuni* investigated survived the atmospheric air and H₂O₂ stress. All the *C. jejuni* strains showed high level of variability in their motility and there was no distinct grouping of the hyperinvasive *C. jejuni* separate from the low invasive *C. jejuni* strains based on these phenotypes. All the *C. jejuni* strains showed variable autoagglutination activity, sodium deoxycholic acid stress response and biofilm formation ability. In addition, the hyperinvasive *C. jejuni* could not be distinguished from the low invasive group of *C. jejuni* strains based on any of the virulence phenotypes tested.

It seems therefore that this group of strains share only the hyperinvasive phenotype and it was hypothesised that the difference in invasion potential between the hyperinvasive and the low invasive *C. jejuni* strains reported by Fearnley *et al* (2008) might be due to the differences at their genome level. Therefore, as the next step in this project the six hyperinvasive *C. jejuni* strains were compared with a selected group of the low invasive *C. jejuni* strains using comparative genomic hybridization (CGH). Some future work may also involve actually investigating the mechanism of *C. jejuni* uptake and survival within the host cells.

Chapter Four

**GENOTYPING OF THE
HYPERINVASIVE *C. JEJUNI*
STRAINS BY COMPARATIVE
GENOMIC HYBRIDIZATION
(CGH)**

GENOTYPING OF THE HYPERINVASIVE *C. JEJUNI* STRAINS BY COMPARATIVE GENOMIC HYBRIDIZATION (CGH)

4.1 INTRODUCTION

The previously defined group of six clinical hyperinvasive *C. jejuni* strains with the hyperinvasive phenotype were compared at the genome level using DNA microarray with four low invasive clinical *C. jejuni*. This comparative phylogenomics study was performed to find whether the hyperinvasive *C. jejuni* strains form a cluster separate from the low invasive strains based on their genomic content as well as to identify any genomic regions conserved or variable between the two groups.

4.1.1 Comparative Genomic Hybridization as a tool to study phylogeny and diversity

Comparative Genomic Hybridization (CGH) is a DNA microarray based technique used to compare a test genome with a control genome, affixed on a microscopic slide-sized glass support, to find genomic content present or absent in the test genome. Comparative genomics using DNA microarrays have been used to study relatedness and differences between different bacterial populations and some examples are discussed below. CGH has been used to study genomic content differences between 42 isolates of *Helicobacter pylori* causing a number of chronic conditions in humans including peptic ulcer, gastric cancer or gastritis and other complex diseases that are linked to virulence gene clusters including the *cag* pathogenicity island (PAI). This study identified genes uniquely associated with gastroduodenal diseases. One thousand and nineteen genes were found to be present in all isolates, with 341 genes being variable. The most variable genes linked with gastritis, duodenal ulcer, or gastric cancers were present in Pathogenicity Zones (PZs) and the *cag* PAI. (Romo-Gonzalez *et al.*, 2009).

In another study, CGH was used to find genomic diversity amongst *Cronobacter* species including *C. sakazakii*, *C. malonaticus* and *C. turicensis*. This study identified *Cronobacter* species specific genes and genes related to *Cronobacter* isolates from outbreaks in neonatal intensive care units. Genes associated with copper and silver resistance, those encoding for multidrug efflux pumps and many adhesins were reported

(Kucerova *et al.*, 2010). This study used the GACK algorithm, trinary cut off for determining present and divergent genes (Kim *et al.*, 2002) and phylogenomic analysis was performed by using “Cluster” software (Eisen *et al.*, 1998). Interstrain genomic polymorphisms associated with *P. gingivalis* were investigated by comparing the known invasive *P. gingivalis* strain to the non-invasive *P. gingivalis* strain by DNA microarray. Several accessory genes with functions encoding for lipoproteins, capsular biosynthesis, regulatory and immunoreactive proteins, and transport of metabolites were divergent in the non-invasive strain suggesting that gene loss was indicative of the *P. gingivalis* non-invasive phenotype (Dolgilevich *et al.*, 2011).

DNA microarrays have been used to study phylogeny and genetic diversity of *C. jejuni* strains based on host association, disease profile and virulence related phenotypes (Quiñones *et al.*, 2008, Parker *et al.*, 2006, Champion *et al.*, 2005, Pearson *et al.*, 2003, Dorrell *et al.*, 2001). *C. jejuni* strains with a variety of Penner serotypes were studied using CGH to define the structure of the *C. jejuni* genome and to find genomic similarities between them (Dorrell *et al.*, 2001). DNA microarrays separated the functional core comprising essential genes from the dispensable genes that make up the accessory genome. This study found 1,300 out of 1,654 genes were part of the core genome and at least 21% of the genes were dispensable as they were missing in one or more of the strains studied. The conserved genes mainly encoded for metabolism, cellular, regulatory and biosynthetic processes. The accessory genome was dominated by virulence related genes including flagellar modification genes, lipo-oligosaccharide and capsule encoding genes. CGH data did not cluster strains based on the Penner serotypes suggesting that the genome similarity between strains from the same serotype may not be enough to cluster them together (Dorrell *et al.*, 2001).

Another study, investigated the genomic diversity in 18 *C. jejuni* strains isolated from a variety of sources (Pearson *et al.*, 2003). This study identified 7 regions of immense variability between the strains studied which were called the plasticity regions (PRs). PR1 to PR7 made up 50% of the variable genome content of the strains. PR1 consisted of genes required for using alternative electron acceptors for respiration that may provide a selective advantage to strains in oxygen depleted conditions. PR2, PR3, and PR7 were dominated by genes encoding outer membrane and periplasmic proteins and

many hypothetical genes with putative functions that may explain the phenotypic variation and survival of strains in different environments. PR4, PR5, and PR6 contained genes involved in the synthesis part of the flagellin glycosylation locus. Pearson *et al* (2003) defined a dynamic cut off algorithm similar to the GACK matrix (Kim *et al.*, 2002) to determine present or divergent genes.

Due to the highly diverse nature of the *C. jejuni* genome and the advantage of CGH in the absence of genome sequencing the use of CGH as a genotyping tool did expand. A large scale CGH study exploited a collection of CGH data from three previous small scale *C. jejuni* microarray studies (Leonard *et al.*, 2004, Pearson *et al.*, 2003, Dorrell *et al.*, 2001) and integrated it with additional DNA microarray analysis performed on 51 *C. jejuni* strains isolated from food and clinical sources (Taboada *et al.*, 2004). This CGH meta-analysis of 97 *C. jejuni* strains showed that the newly investigated *C. jejuni* strains contain the majority of the genes which were reported as part of the accessory gene pool in previous studies, suggesting that the *C. jejuni* core genome was expanding (Taboada *et al.*, 2004).

Another study used DNA microarrays to investigate the GBS and enteritis associated *C. jejuni* isolates with an aim to identify genetic markers of GBS. However, this study could not identify any GBS specific genetic markers (Leonard *et al.*, 2004). A shot gun DNA microarray technique comparing sequenced *C. jejuni* NCTC11168 with the, at that time unsequenced *C. jejuni* 81-176 genome, identified several *C. jejuni* 81-176 specific genes that were dispersed in conserved and accessory parts of the genome (Poly *et al.*, 2005).

A comparative phylogenomics study combined with Bayesian-based statistical analysis was used to study phylogeny of *C. jejuni* strains from humans, chicken, bovines, ovines and the environment. Out of 111 *C. jejuni* isolates studied, 70 strains were isolated from patients with gastroenteritis, septicaemia, GBS and from asymptomatic carriers. The aim of this study was to find if strains clustered together based on host sources and disease severity in human hosts. The Bayesian analysis revealed two distinct clades a “livestock” clade and a “nonlivestock” clade with further clusters of environmental isolates (Champion *et al.*, 2005). Interestingly, the majority of the human isolates were part of the nonlivestock clade proposing that most *C. jejuni* infections came from nonlivestock sources. Several genes (Cj1321 to Cj1326) within the flagellin

glycosylation locus were associated with strains in the livestock cluster (Champion *et al.*, 2005).

The genomic diversity in a collection of *C. jejuni* and *C. coli* strains from clinical and veterinary sources was investigated using a *C. jejuni* RM1221 and *C. jejuni* NCTC11168 pan array (Parker *et al.*, 2006). The *C. jejuni* and *C. coli* populations investigated showed genomic diversity in four *C. jejuni* RM1221 integrated mobile genomic islands (CJIEs). An additional 18 regions of diversity were also identified containing lipooligosaccharide and capsule biosynthesis genes (Parker *et al.*, 2006). Later, Parker *et al.*, (2007) used CGH to study genomic diversity at the *C. jejuni* subspecies level. The *C. jejuni* subsp. *jejuni* and *C. jejuni* subsp. *doylei* formed separate clusters and showed immense genomic diversity between them based on the two techniques used. *C. jejuni* subsp. *doylei* unique gene clusters mainly related to metabolism, transport and pathogenesis were identified that were absent from *C. jejuni* subsp. *jejuni* (Parker *et al.*, 2007).

A 70-mer oligonucleotide array was used to find similarities and differences between closely related (based on similar PFGE profile) chicken and human clinical isolates. CGH grouped isolates in the same way as defined by PFGE. Also, the chicken and human clinical *C. jejuni* isolates did not cluster according to source by CGH (Rodin *et al.*, 2008).

C. jejuni clinical strains with different disease severities were compared using DNA microarrays to see if gene markers associated with different disease outcomes could be identified. It was shown that *C. jejuni* strains from patients with neurological disorders like Guillain Barré syndrome and Miller Fisher syndromes clustered separately from enteritis associated *C. jejuni* isolates (Quinones *et al.*, 2008). Large mobile genetic islands mainly characterized by prophage genes were dominant in strains causing neuropathies (Quinones *et al.*, 2008). Recently, DNA microarray analysis was used to study the role of genetic diversity in the disease profile of three *C. jejuni* human disease isolates (11168, 33292 and 81-176) and genetically marked variants of these strains. The data showed immense genetic variability in three gene clusters associated with the synthesis and modification of capsule, flagella and lipooligosaccharide (Wilson *et al.*, 2010).

CGH was used to study population dynamics of *C. coli* strains from poultry, environment and human clinical cases. *C. coli* isolated from diverse sources clustered based on the host species and CGH identified host associated genes showing host adaptation. The clustering of *C. coli* strains from diverse sources was attributed to origin from a common ancestor and molecular events like lateral gene transfer (Lang *et al.*, 2010).

Another CGH study was performed on 80 *C. jejuni* isolates from diverse sources representing a variety of clonal complexes to show the association of clonal complexes with isolation source. This study identified a clade of water/wildlife associated *C. jejuni* strains separate from the human food chain isolates. Nine regions of divergence were found unique to this clade of strains suggesting a trend towards niche adaptation. These gene clusters were part of the accessory genome and included the virulence related cytolethal distending toxin (*cdt*) genes (Hepworth *et al.*, 2011).

All above studies signify the importance of DNA microarray as a powerful molecular tool to study phylogeny and diversity between *C. jejuni* strains. The strains have been distinguished based on disease profile and transmission source at the sub-species level. Most studies reported that the flagellar modification genes, capsule biosynthesis loci, and LOS were the most variable regions. Here, we have used CGH to compare six hyperinvasive *C. jejuni* strains with four low invasive *C. jejuni* identified by Fearnley *et al* (2008).

The aim of this study was to:

- (i) Determine the phylogenomic relationship between hyperinvasive and low invasive *C. jejuni* strains at the whole genome level using hierarchical clustering of CGH data.
- (ii) Identify gene/gene clusters specific to the hyperinvasive *C. jejuni* that may distinguish them from low invasive *C. jejuni* and explain the hyperinvasive phenotype of this unique group of hyperinvasive *C. jejuni*.

4.2 METHODS

4.2.1 Microarray design and construction

The *C. jejuni/C. coli* pan array used here was designed based on 1884 annotated coding sequences (CDSs) from the fully genome sequenced *C. jejuni* RM1221, additional CDSs from *C. jejuni* NCTC11168 and *C. jejuni* 81-176. Genes unique to *C. coli* RM2228, ORFs from the *C. jejuni* 81-176 pTet plasmid and *C. coli* pCC31 plasmid were also represented on the array. This represented a total of 2,628 annotated CDSs on the pan array. The pan array was constructed as described elsewhere (John *et al.*, 2011).

4.2.2 Enzymatic labelling of genomic DNA with Cy3 dCTP and Cy5 dCTP and hybridization

Three micrograms of the control (*C. jejuni* RM1221) and test genomic DNA (Hyperinvasive/low invasive *C. jejuni*) was prepared as described in section 2.6.1. The reference and test genomic DNA was enzymatically labelled with fluorescent dyes Cy5-dCTP and Cy3-dCTP (Amersham, GE healthcare) respectively by following the protocol described in detail by Dorrell *et al* (2001). A brief protocol is given below;

- One microgram of the genomic DNA from test *C. jejuni* strain was mixed with the random primers (Promega, UK) at a concentration of 3 µg/µl and volume was made up to 41.5 µl with sterile distilled water in a PCR tube.
- The reaction mix was heated at 95 °C in thermocycler for 5 minutes and then snap cooled on ice.
- Five microliters of 10X buffer (part of random primers, Promega, UK), 5 µl of dNTP mix (5 mM each dATP, dGTP And dTTP, 2 mM dCTP, Sigma-Aldrich, UK), 1.5 µl of Cy3-dCTP (25 nmol) (Amersham, GE healthcare; catalogue number PA55021) and 1 µl of DNA polymerase I large klenow fragment, exonuclease minus (Promega, UK; catalogue no; M2181) were added to the above reaction mix.
- The labelling reaction mix was incubated at 37 °C for 90 minutes.
- The above procedure was carried out for the labelling of reference *C. jejuni* genomic DNA (section 2.5) replacing Cy3-dCTP with Cy5-dCTP (25 nmol) (Amersham, GE healthcare; catalogue number PA53021).

4.2.2.1 Hybridization

Each enzymatic labelling reaction was purified using MinElute PCR purification kit (Qiagen, Crawley, UK) as stated in the kit's instruction manual. Each labelled test and control genomic DNA was mixed together in a maximum reaction volume of 4.5 μ l (0.5 μ g of genomic DNA concentration). The hybridization was carried out at the University of Nottingham, post genomics facility at the Queen's Medical Centre (Nottingham, UK). The hybridization procedure was followed as detail in detail previously (John *et al.*, 2011). Each array image was obtained as a .gps file and the layout of oligonucleotides on the array was presented as .gal file.

4.2.3 Tools for CGH data analysis and hierarchical clustering

Each microarray image was analysed using Gene pix pro 6 software (Molecular Devices Corporation, Sunnyvale, CA). Any anomalous spots with low signal intensities or too high or low background noise on the array were identified. Gene pix pro 6 computed several ratio quantities each of which contained different information for each spot on the array. Among these, an important parameter called the "log₂ratio" for each spot was calculated. Log₂ratio is the base two logarithm of ratio of median intensities *i.e.* log₂ratio of median intensity at 532 nm (Cy3-test strain)/ ratio of median intensity at 635 nm (Cy5-reference strain). The ratio of medians is the ratio of the background subtracted median pixel intensity at the second wavelength (532 nm) to the background subtracted median pixel intensity at the first wavelength (635 nm). The log base two transform is a simple and quicker way of presenting differences between two conditions (for example, present and absent/divergent genes in this study). The data for each spot on the array was corrected by normalization by using the global normalization method in which the image and result were normalized together. By default, the mean of the ratio of medians used for normalization was set to 1.0. This deleted any ratio values less than 0.1 or greater than 10 as well as any spots that were flagged as bad spots. The results for each complete image were generated as Gene pix results (.gpr) file. This file contained general information about the image acquisition and analysis as well as the numerical data generated from each spot on the array (Molecular Devices Corporation, Sunnyvale, CA). The numerical data in .gps file was converted into text delimited (.dat) file and imported into the statistical software, SPSS 17.0 (IBM SPSS Statistics, IBM

Corporation). This program was used for further data rearrangement and to check the smoothness of the array data by plotting histograms. All the unnormalized bad spots were removed from data. Furthermore, any control spots on the array were checked to confirm the successful hybridization and image analysis. Finally, a file (.SPSS) which only contained unique oligo IDs, gene name and \log_2 ratios for all the normalized and perfect spots was generated. This file was converted into a text delimited file (.dat) for use in “GACK” software (Kim *et al.*, 2002) which assigned genes into present and absent/divergent categories. The GACK analysis program selected a dynamic cut off value based on shape of the signal strength for each spot to be classified as present or divergent (Kim *et al.*, 2002). This was done by calculating the Estimated Probability of Presence (EPP) value for each spot on the array based on its \log_2 ratio. GACK calculated EPP by dividing the normal (expected) distribution curve \log_2 ratio values where all the spots on the array have worked against the actual (observed) \log_2 ratio distribution values for all spots on the array.

$$\%EPP=100x \text{ (normal expected value/observed value)}$$

The trinary cut off method was used at a %EPP cut off 1 set to 0 and %EPP cut off 2 set to 100. Under these parameters, the genes with 0% or 100% EPP were assigned into highly divergent/absent and present categories respectively. Any genes with EPP between 0% and 100% were classed into an uncertain category. These uncertain genes represent the class of genes that could not be assigned into present or highly divergent/absent categories with high confidence therefore they were called slightly divergent genes. Trinary output (.cdt) file containing the cut off values for all genes as binary numbers was generated with present genes designated as 1, slightly divergent genes designated as 0 and highly divergent/absent genes designated as -1 (Kim *et al.*, 2002). Phylogenomic analysis was performed by “Cluster” software (Eisen *et al.*, 1998). The GACK trinary file was converted to text delimited (.dat) format by using SPSS statistical software and imported to Cluster. Using “Cluster” software, the Pearson Correlation coefficient was applied with correlation centered selected as the similarity metrics to establish relationship between strains. The Pearson Correlation between any two series of numbers $x = (x_1, x_2, \dots, x_n)$ and $y = (y_1, y_2, \dots, y_n)$ is defined by the equation below;

$$r = \frac{1}{n} \sum_{i=1}^n \left(\frac{x_i - \bar{x}}{\sigma_x} \right) \left(\frac{y_i - \bar{y}}{\sigma_y} \right)$$

where \bar{x} represents the average values in x and σ_x is the standard deviation of these values. If x and y values were plotted as curves then r shows how similar the shapes of the two curves are.

The Pearson correlation is always between 1 and -1 where 1 represents that the two series are identical, 0 indicates that they are completely uncorrelated and -1 means that they are perfectly opposite (Eisen *et al.*, 1998).

For hierarchical clustering, an agglomerative clustering method was used based on the average linkage clustering algorithm by Sokal and Michener (1958). This process performed repeated clustering cycles joining the true items (genes) or pseudo-items (groups of genes) with the smallest distance between them by a branch/node of a tree whereby the length of the branch represented the distance between them. The whole process was repeated for whole dataset replacing items with computed distances by new items until one item remained. The average distance between joining nodes is generated as (.atr) output file (Appendix: supplementary table 1). The clustering along with the average linkage distance between clusters was visualised by “Tree view” software (Eisen *et al.*, 1998) (Figure 4.3).

4.2.4 Classification of loci specific to the hyperinvasive *C. jejuni* and further analysis

Further mining of CGH data was performed by Professor Nadia Chuzhanova (Physics and Maths department, NTU) through personal communication. This analysis was carried out by using the Delphi 7 object-oriented programming language. The analysis run is included in appendix, supplementary file 1. The aim was to find genes present or absent/highly divergent in all *C. jejuni* strains in one group compared against the other group (*e.g.* genes present in all the six hyperinvasive *C. jejuni* strains and absent/highly divergent from the four low invasive *C. jejuni* strains studied by CGH and vice versa). If there were n number of *C. jejuni* strains in the hyperinvasive group and k number of *C. jejuni* strains in the low invasive group then the criteria was defined as follow:

- If a gene was present in all n strains and absent/highly divergent from all k strains.

Various other criteria were tried with the following being successful:

- (1) if a gene was present in all n strains and absent/highly divergent in k-1 strains.
- (2) if a gene was absent/highly divergent in all n strains and present in k-1 strains.
- (3) if a gene was present in all k strains and absent/highly divergent in n-1 strains.
- (4) if a gene was absent/highly divergent in all k strains and present in n-1 strains.
- (5) if a gene was present in n-1 strains and absent/highly divergent from k-1 strains.
- (6) if a gene was absent/highly divergent in n-1 strains and present in k-1 strains.

Artemis software from <http://www.sanger.ac.uk/resources/software/artemis/> (Rutherford *et al.*, 2000) was used to produce circular diagrams of the pan array to compare the variability between strains based on the CGH data.

4.3 RESULTS AND DISCUSSION

4.3.1 Genotyping of hyperinvasive *C. jejuni* using Comparative Genomic Hybridization

The six hyperinvasive *C. jejuni* (01/10, 01/35, 01/04, 01/41, 01/51, EX114) and four selected low invasive *C. jejuni* (01/30, 01/32, 01/46, 01/39) strains (Fearnley *et al.*, 2008) were studied by using comparative genomic hybridization (CGH). Each of the test *C. jejuni* strain and the control strain, *C. jejuni* RM1221, was hybridized on the *C. jejuni* /*C. coli* pan DNA microarray. Each hybridization was performed in triplicate. Each gene was represented by duplicate synthetic oligonucleotides on the array.

4.3.1.1 Robustness of genotyping using DNA microarray

After data processing and thorough filtration using Gene pix pro 6 software (Molecular Devices Corporation, Sunnyvale, CA) and SPSS 17.0 (IBM SPSS Statistics, IBM Corporation) the normalized \log_2 ratio values (base two logarithm of ratio of median intensities) were also used to generate histograms to check the quality of each array. A histogram comparing the \log_2 ratio values for all the normalized spots for one array

experiment of *C. jejuni* 01/41 has been presented as an example to demonstrate the robustness of our DNA microarray data (Figure 4.1). Using SPSS 17.0 (IBM SPSS Statistics, IBM Corporation), the \log_2 ratio values for each strain from three independent array experiments was averaged (Appendix: supplementary table 2) for calculating the GACK trinary cutoffs.

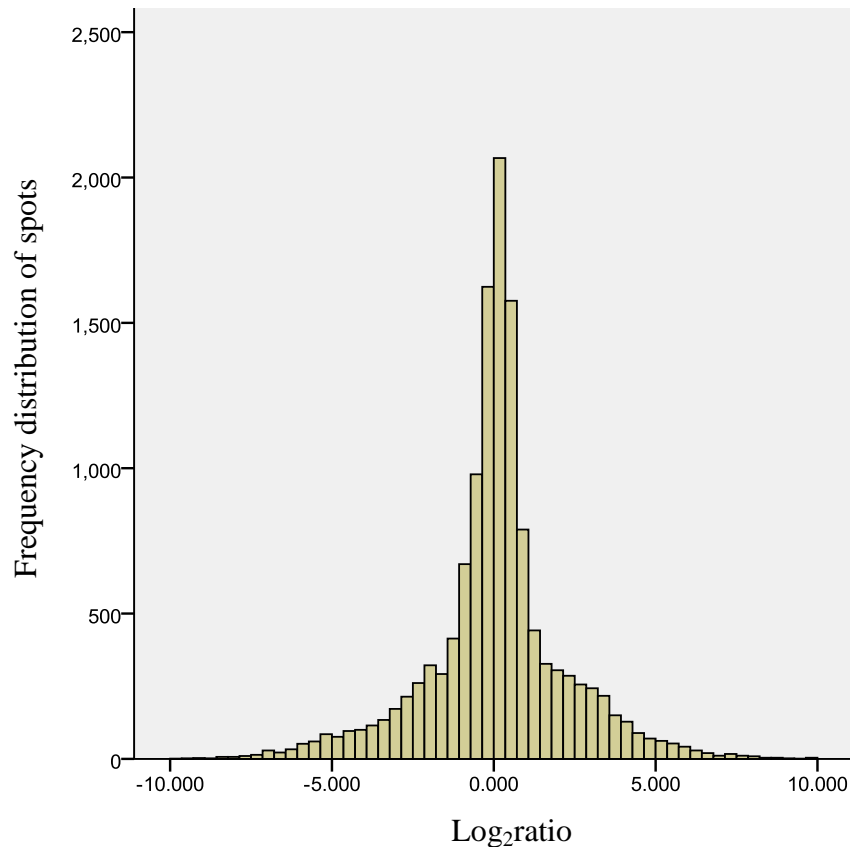


Figure 4.1: A histogram comparing the frequency distribution of spots based on their \log_2 ratios.

The data showed a smooth and tight peak representing minimal technical error associated with the microarray experiment that could occur due to inefficient samples labelling and failure of proper hybridisation. It is important to note that a histogram is only an indicative of overall array quality based on the ratios but doesnot provide information about the individual genes. The associated detailed information of signal strength for each spot is analysed to have complete confidence in data quality.

4.3.1.2 Preliminary microarray data analysis

For phylogenomic comparisons, the normalized \log_2 ratios for each spot on the array were applied to the most strict values of the trinary cut off matrix in GACK software (available at <http://falkow.stanford.edu/>) to classify genes as present or absent/highly divergent (Kim *et al.*, 2002). The GACK cut off algorithm has been developed as an improved analytical method for determining the presence or divergence of genes applied to *C. jejuni* and *H. pylori* genotyping data (Kim *et al.*, 2002). GACK assigns a gene into present or divergent category irrespective of any normalization. Unlike constant cut off values used in empirical methods, GACK generates an independent cut off for each experimental dataset. Thus GACK provides more confidence in gene category assignment. Since each gene is represented twice on the array, SPSS 17.0 (IBM SPSS Statistics, IBM Corporation) was used to determine an average presence/absence category for each gene (Appendix: supplementary table 3). The dataset from one *C. jejuni* 01/41 array experiment has been presented as an example to demonstrate trinary cut off as the best method for identification of divergent genes using “Estimated Probability of Present genes (EPP)” values (Figure 4.2).

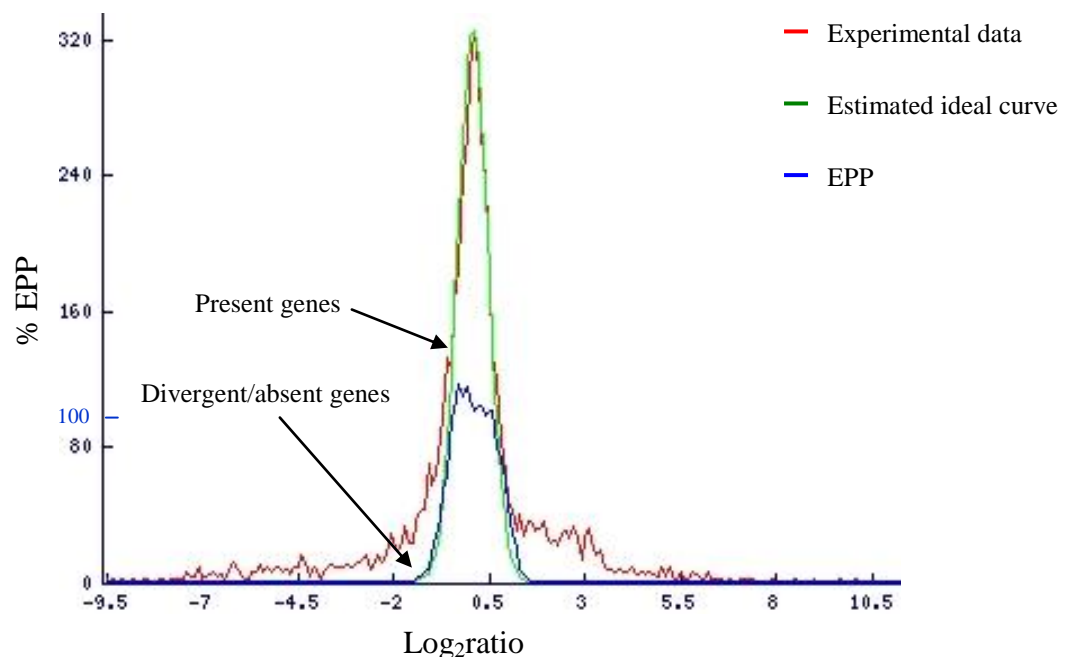


Figure 4.2: GACK trinary cut off algorithm for gene category assignment for an experimental dataset (*C. jejuni* 01/04).

The observed \log_2 ratio values from one *C. jejuni* 01/41 array experiment compared against the ideal curve \log_2 ratio values and EPP for each gene determined by GACK trinary cut-off. The EPP is 0% at \log_2 ratio of ~ -1.5 and 100% at ~ -1.4 .

The majority of the distribution of observed normalized hybridization data overlapped the normal distribution of data as expected. The observed distribution tailed off to 0% EPP (\log_2 ratio ~ -1.5) where highly divergent genes existed. The increase of EPP to 100% (\log_2 ratio ~ -1.4) subsequently increased the expected probability towards present genes. The transition region between 0% EPP and 100% EPP (*i.e.* \log_2 ratio of ~ -1.5 and ~ -1.4) contained slightly divergent genes. Other studies phylogenomically comparing the human clinical *C. jejuni* isolates derived from different sources and their disease profile have successfully validated GACK software for classifying genes into present or divergent categories (Champion *et al.*, 2005, Poly *et al.*, 2004, Pearson *et al.*, 2003). For example in the study by Champion *et al* (2005) the classification of genes into present and divergent categories was validated by an empirical cut off determination that produced a very similar list of present and divergent genes. Based on the successful application of GACK analysis, only GACK cut offs were used for assigning genes present or divergent status in this study.

Using the CGH data, the core genome was determined by calculating the total number of genes present in all test *C. jejuni* strains studied by CGH. This was calculated as follows;

$$\text{Core genome (\%)} = \frac{\text{Number of CDSs present in all test } C. \textit{jejuni} \text{ strains}}{\text{Total number of functional reference CDSs on array}} \times 100$$

There were 623 (23.7%) functional CDSs in the core genome that mainly consisted of genes involved in essential regulatory, cellular and metabolic functions. The remaining genes formed the accessory genome as they were variable between the strains. The accessory genome mainly comprised of CDSs associated with capsule, LOS, flagellar and restriction modification systems. For selection of genes specific to the hyperinvasive *C. jejuni* strains only present and highly divergent genes were considered to have absolute confidence in selection.

4.3.2 Phylogenomic clustering of hyperinvasive and low invasive *C. jejuni* by CGH

Based on CGH data, the phylogenomic relationship was established between the hyperinvasive and low invasive *C. jejuni* strains by using “Cluster” software that performed a Pearson correlation average linkage clustering by comparing the present and highly divergent/absent genes. The hyperinvasive and low invasive *C. jejuni* strains were also compared based on their MLST profiles (Figure 4.3).

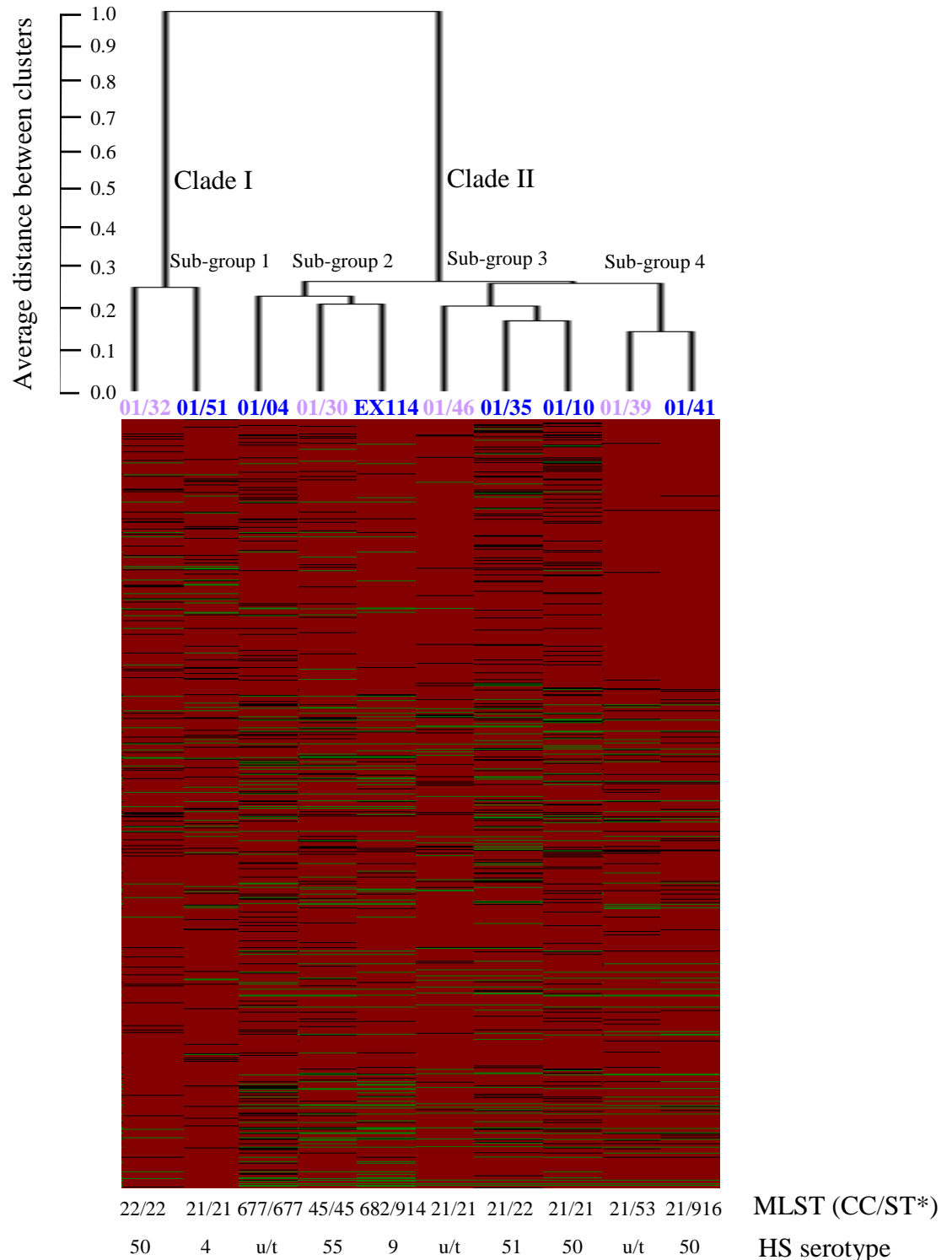


Figure 4.3: The hierarchical clustering of hyperinvasive *C. jejuni* and low invasive *C. jejuni* based on CGH along with the MLST profile and Penner (HS) serotype.

The heat map representing present and highly divergent/absent genes in all *C. jejuni* isolates across the whole genome with genes arranged in ascending order.

*CC = Clonal Complex and ST = Sequence Type; u/t = untypeable

Hyperinvasive *C. jejuni* = blue; Low invasive *C. jejuni* = purple; Genes present = red; Genes highly divergent/absent = green and Genes slightly divergent = black

The phylogenomic clustering of six hyperinvasive and four low invasive *C. jejuni* strains examined in CGH study was performed using “Cluster” software (Eisen *et al.*, 1998). *C. jejuni* isolates formed two distinct clades (I and II) based on the genome similarity between them and there was no distinct grouping of the hyperinvasive *C. jejuni* isolates as a separate cluster from the low invasive *C. jejuni*. Clade I had only one subgroup 1 and Clade II was subdivided into three sub groups 2, 3 and 4. Clade I clustered two strains in sub group 1; low invasive *C. jejuni* 01/32 and a hyperinvasive *C. jejuni* 01/51. Clade II clustered eight strains which were sub divided into three sub groups. The subgroup 2 showed the hyperinvasive *C. jejuni* EX114 linked to the low invasive *C. jejuni* 01/30 and both of these strains were related to the hyperinvasive *C. jejuni* 01/04. The third sub-group represented two hyperinvasive *C. jejuni* 01/10 and 01/35 that clustered together and were very closely related to a low invasive *C. jejuni* 01/46. Finally, the sub-group 4 was composed of the low invasive *C. jejuni* isolate 01/39 and the hyperinvasive *C. jejuni* 01/41.

All the hyperinvasive *C. jejuni* strains showed different MLST profiles. The *C. jejuni* strains in subgroups 3 and 4 in Clade II belonged to the clonal complex 21 but there was no grouping based on sequence type. All the other subgroups possessed mixed MLST types. The clonal complex 21 is one of the largest *C. jejuni* clonal complexes containing strains from human, animal and environmental sources (Manning *et al.*, 2003, Dingle *et al.*, 2001, Best *et al.*, 2004). *C. jejuni* 01/51 was the only strain in clade I which belonged to ST 21 complex therefore the association between clade II and ST 21 complex is not absolute. Similarly, there was no common Penner (HS) serotype amongst all the hyperinvasive *C. jejuni* strains. Indeed HS50 was shared between the

hyperinvasive and low invasive *C. jejuni* strains. Three strains were not successfully serotyped which may be due to the age of bacterial culture or antisera (Mckay *et al.*, 2001).

Taboada *et al* (2007) investigated the relationship between neuropathogenic and enteritis causing *C. jejuni* strains using CGH. In agreement with our findings, Taboada *et al* (2007) also could not identify distinct clustering associated with different disease profiles and reported great levels of genomic similarity between them. In a later investigation, Taboada *et al* (2008) compared 32 South African *C. jejuni* strains associated with enteritis, Guillain-Barré or Miller Fisher syndromes using CGH and MLST. This study argued that MLST is a useful but limited technique as it considers variation in few housekeeping genes and does not account for variation across the whole genome as represented by DNA microarray (Taboada *et al.*, 2008). In a phylogenomics study, Champion *et al* (2005) reported results similar to our CGH study with no clustering of the 111 clinical isolates based on the disease symptom. In contrast, the same study identified distinct clusters based on the source of transmission. A partial grouping was observed based on MLST profiles of *C. jejuni* strains studied. The phylogenetic analysis of *C. jejuni* strains from a variety of disease profiles showed that strains with asymptomatic carriage, diarrhoea, bloody diarrhoea, vomiting, septicaemia, and GBS did not cluster as distinct clonal groups. However, six environmental *C. jejuni* isolates clustered as a separate clonal group (Champion *et al.*, 2005). In another study, *C. jejuni* strains selected based on Penner serotypes were compared by CGH. The *C. jejuni* strains with similar Penner serotypes could not be clustered together by CGH suggesting that the *C. jejuni* strains with the same serotype were otherwise variable based on the whole genome content (Dorrell *et al.*, 2001).

It is also important to note here that the *C. jejuni* strains studied in this project were not selected based on MLST or HS serotype but on the invasion phenotype hence, complete association of strains based on MLST type or Penner HS serotype was not expected. The fact that only four low invasive *C. jejuni* strains have been selected for phylogenomic comparison which could be a limiting factor to emphasize on the genetic similarities and differences identified in the hyperinvasive group of *C. jejuni*.

The failure of hyperinvasive *C. jejuni* to form a distinct clade by CGH might reflect the characteristic invasion profile of these strains to be associated with subtle changes in number of genes at the nucleotide level that would not be detected using DNA microarrays (Malik-Kale *et al.*, 2007). It can be argued that the variation in a phenotype may not necessarily be reflected in the genome content of *C. jejuni*. It may be stimulated by the expression of gene (s) belonging to one or more functional pathways (Konkel *et al.*, 1990). In addition, the host factors may also significantly contribute to the *C. jejuni* virulence phenotypes (Taboada *et al.*, 2007).

4.3.3 Identification of loci specific to hyperinvasive C. jejuni and PCR validation of CGH results

The CGH data was further analysed to identify gene/s present in all six hyperinvasive *C. jejuni* strains and absent/highly divergent from the low invasive group of *C. jejuni* strains, and vice versa that might explain the hyperinvasive phenotype of these strains. The analysis performed is described in section 4.2.4. There was no such combination of conserved or variable genes found in all the hyperinvasive *C. jejuni* compared to the low invasive *C. jejuni* strains.

The selection was extended to find present or divergent genes in the majority of hyperinvasive *C. jejuni* strains (section 4.2.4), which identified 67 genes. These genes were classified into six groups based on their presence or divergence in the hyperinvasive *C. jejuni* strains. Only Groups 1 and 2 are presented here in tables 4.1a and 4.1b. The presence or variability of genes in these two groups was further validated by PCR (Figures 4.4a and 4.4b). The primers and expected amplicon sizes are listed in table 2.2.1. The rest of genes arranged in four groups (3, 4, 5 and 6) are provided in the supplementary table 4 in the appendix.

Table 4.1a: Genes selected from CGH data as present in all hyperinvasive *C. jejuni*.

Gene name/number	Hyperinvasive <i>C. jejuni</i>										Low-invasive <i>C. jejuni</i>				Role/function		COG functional grouping
	01_10	01_35	01_04	01_41	01_51	EX114	01_30	01_32	01_46	01_39	<i>C. jejuni</i> NCTC11168	<i>C. jejuni</i> RM1221	<i>C. coli</i> RM2228	<i>C. coli</i> RM2228			
GROUP 1: Genes present in all hyperinvasive <i>C. jejuni</i> and <i>C. coli</i> RM2228	01_10	01_35	01_04	01_41	01_51	EX114	01_30	01_32	01_46	01_39	<i>C. jejuni</i> NCTC11168	<i>C. jejuni</i> RM1221	<i>C. coli</i> RM2228	<i>C. coli</i> RM2228	Fouts <i>et al.</i> , 2005		
<i>Cj0738</i>	1	1	1	1	1	1	1	1	-1	-1	hypothetical protein	hypothetical protein	Hypothetical		
<i>hisS</i> (Cj0765c)	1	1	1	1	1	1	1	1	-1	-1	histidyl-tRNA synthetase	histidyl-tRNA synthetase	histidyl-tRNA synthetase	Translation		
<i>aat</i> (Cj1109)	1	1	1	1	1	1	1	1	-1	-1	putative leucylphenylalanyl-tRNA--protein	leucylphenylalanyl-tRNA--protein transferase	leucylphenylalanyl-tRNA--protein transferase	Posttranslational modification, protein turnover, chaperones		
<i>proC</i> (Cj1076)	1	1	1	1	1	1	1	1	-1	-1	putative pyrroline-5-carboxylate reductase	pyrroline-5-carboxylate reductase	pyrroline-5-carboxylate reductase	Amino acid transport and metabolism		
<i>metF</i> (Cj1202)	1	1	1	1	1	1	1	1	-1	-1	5,10-methylenetetrahydrofolate reductase	5,10-methylenetetrahydrofolate reductase	5,10-methylenetetrahydrofolate reductase	Amino acid transport and metabolism		
.....	1	1	1	1	1	1	1	1	-1	-1	Hypothetical		
<i>Cj0564</i>	1	1	1	1	1	1	1	1	-1	-1	putative integral membrane protein	hypothetical protein	Hypothetical		
<i>modA</i> (Cj0303c)	1	1	1	1	1	1	1	1	-1	-1	putative molybdate-binding lipoprotein	molybdenum ABC transporter, periplasmic	molybdenum ABC transporter, periplasmic	Transport and metabolism		
<i>Cj0271</i>	1	1	1	1	1	1	1	1	-1	-1	bacterioferritin	antioxidant, AlpC/Tsa family homolog	bacterioferritin	Posttranslational modification, protein turnover, chaperones		

1=present; -1=absent/highly divergent

Group 1 contains 9 genes present in all hyperinvasive *C. jejuni* and absent/highly divergent from three of four low invasive *C. jejuni*.

Table 4.1b: Genes selected from CGH data as absent/highly divergent in all hyperinvasive *C. jejuni*.

Gene name/number	Hyperinvasive <i>C. jejuni</i>					Low-invasive <i>C. jejuni</i>					Role/function			COG functional grouping
	01_10	01_35	01_04	01_41	01_51	EX114	01_30	01_32	01_46	01_39	<i>C. jejuni</i> RMI221	<i>C. coli</i> RM2228		
GROUP 2: Genes highly divergent/absent in all hyperinvasive <i>C. jejuni</i> and present in three of four low invasive <i>C. jejuni</i>	01_10	01_35	01_04	01_41	01_51	EX114	01_30	01_32	01_46	01_39	<i>C. jejuni</i> NCTC11168	<i>C. coli</i> RM2228		Fouts <i>et al.</i> , 2005
.....	CJE1112	CCOA0144											
Cj0266c		CJE0315												
Cj0701		CJE0801												
<i>tgt</i> (Cj1010)														
.....														
.....														
.....														

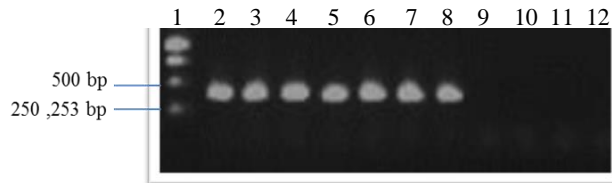
1=present;-1=absent/highly divergent

Group 2 consists of 6 genes highly divergent/absent in all hyperinvasive *C. jejuni* and present in three of four low invasive *C. jejuni*.

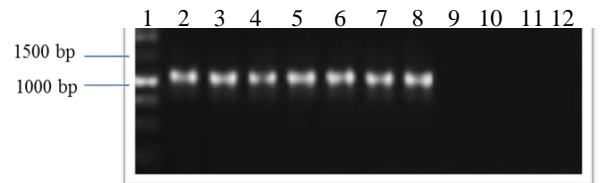
PCR screening of genes listed in group 1 and 2 (Table 4.1a and 4.1b) further validated their presence and absence in the hyperinvasive and low invasive *C. jejuni* isolates.

(a)

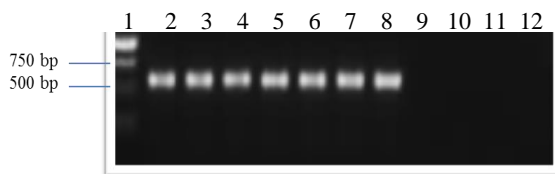
(i) CJE0838



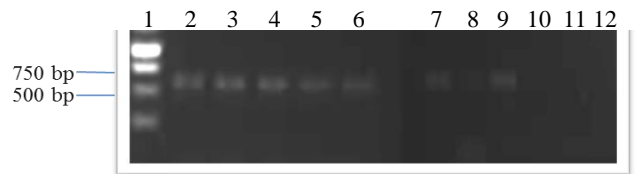
(ii) *hisS*



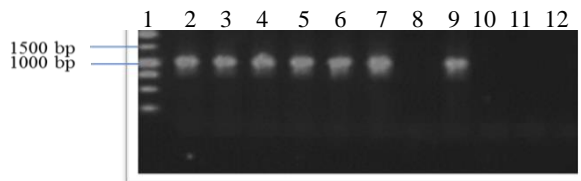
(iii) *aat*



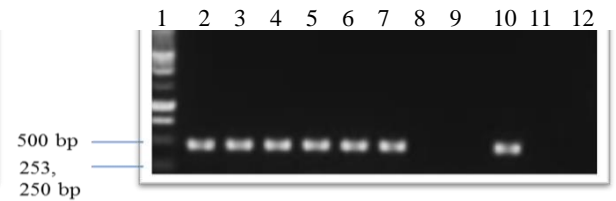
(iv) *proC*



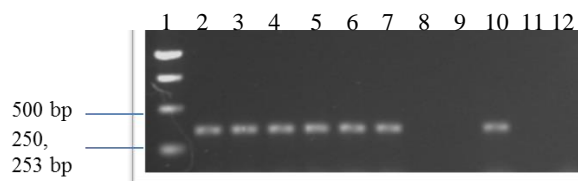
(v) *metF*



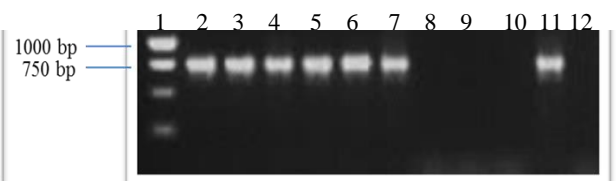
(vi) CJE0669



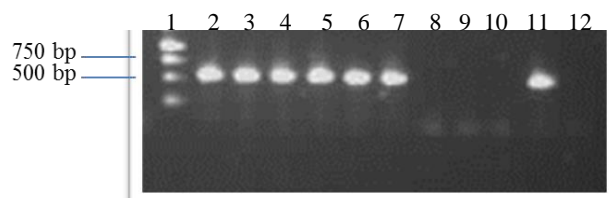
(vii) CCOA0033



(viii) *modA*



(ix) CJE0320



Continous

(b)

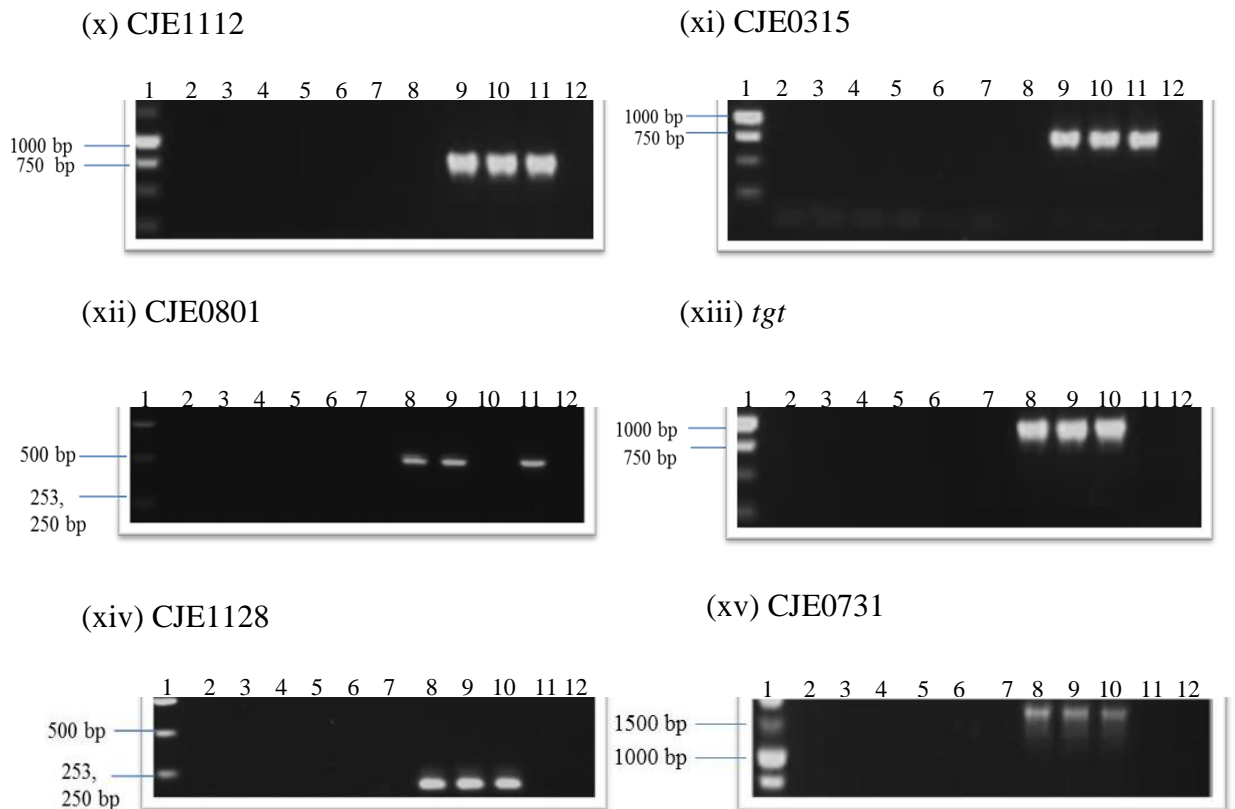


Figure: 4.4: PCR validation of CGH data.

(a) PCR verification of genes present in all hyperinvasive *C. jejuni* and highly divergent from three of four low invasive *C. jejuni* (Group 1, table 4.1a). (i) CJE0838 (ii) *hisS* (iii) *aat* (iv) *proC* (v) *metF* (vi) CJE0669 (vii) CCOA0033 (viii) *modA* (ix) CJE0320. (b) Six genes included in group 2 (Table 4.1b) were found as highly divergent in all hyperinvasive and low invasive *C. jejuni* with an exception of one low invasive *C. jejuni* strain in each case. (x) CJE1112 (xi) CJE0315 (xii) CJE0801 (xiii) *tgt* (xiv) CJE1128 (xv) CJE0731.

Each gel was loaded: Lane 1; 1 kbp DNA marker (Promega, UK); (lanes 2-7); hyperinvasive *C. jejuni* strains 01/10, 01/35, 01/04, 01/41, 01/51, EX114. Lanes (8-11); low invasive *C. jejuni* strains 01/30, 01/32, 01/46, 01/39. Lane 12; negative control.

4.3.4 Presence of hyperinvasive-associated loci in an additional group of low invasive *C. jejuni* isolates

To investigate the presence of the identified genes amongst a wider group of strains PCR screening on additional 9 low invasive *C. jejuni* strains was conducted (Table 4.2).

Table 4.2: Summary of PCR screening for the identified genes (Table 4.1a and 4.1b) in the hyperinvasive and low invasive *C. jejuni* strains.

Gene name/number in <i>C. jejuni</i> RMI221	PCR profile of hyperinvasive and low invasive <i>C. jejuni</i>																	p-value*			
	Hyperinvasive <i>C. jejuni</i>					Low invasive <i>C. jejuni</i>															
GROUP 1: Genes present in all hyperinvasive <i>C. jejuni</i>	01_10	01_35	01_04	01_41	01_51	EX114	01_30	01_32	01_46	01_39	01_05	01_08	01_11	01_36	C2/3	C12/11	C27/14	C69/2	C110/4		
CJE0838	+	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	0.000
<i>hisS</i> (CJE0856)	+	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	0.000
<i>aat</i> (CJE1252)	+	+	+	+	+	+	+	-	-	+	-	-	-	-	-	-	-	-	-	-	0.001
<i>proC</i> (CJE1219)	+	+	+	+	+	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	0.000
<i>metF</i> (CJE1336)	+	+	+	+	+	+	+	+	-	-	+	-	-	-	-	-	-	-	-	-	0.001
CCO40033	+	+	+	+	+	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	0.000
CJE0669	+	+	+	+	+	+	+	-	+	-	-	-	-	-	-	-	-	-	-	-	0.000
<i>modA</i> (CJE0348)	+	+	+	+	+	+	+	-	-	+	+	-	-	-	-	-	-	-	-	-	0.003
CJE0320	+	+	+	+	+	+	+	-	-	+	-	-	-	-	-	-	-	-	-	-	0.000
GROUP 2: Genes highly divergent/absent in all hyperinvasive <i>C. jejuni</i>																					
CJE1112	-	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	0.000
CJE0315	-	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	0.000
CJE0801	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	0.000
<i>tgt</i> (CJE1090)	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	0.000
CJE1128	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	0.011
CJE0731	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	0.517

+ = present, - = absent

*Chi-square, Fisher's exact test was performed to determine if the presence or absence of identified genes in the hyperinvasive group of *C. jejuni* strains was statistically significant. Significance level = 1% ($p=0.01$).

Table 4.2 (group 1) showed that most of the additional low invasive *C. jejuni* isolates were negative for genes present in all hyperinvasive *C. jejuni*. In group 1, six genes (CJE0838, *hisS*, *proC*, CCOA0033, CJE0669 and CJE0320) were highly divergent in 1/13 low invasive *C. jejuni* isolates. Among the additional *C. jejuni* strains PCR screened, *C. jejuni* 01/05 was the only isolate positive for three genes *i.e.* *aat*, *metF* and *modA* followed by *C. jejuni* 01/08 that was positive for *modA* only.

For the genes in group 2, the majority of the additional low invasive *C. jejuni* strains possessed these genes. PCR analysis showed that four genes (CJE1112, CJE0315, CJE0801 and *tgt*) were present in a total of 12/13 low invasive *C. jejuni*. Additionally, all the 9 additional low invasive *C. jejuni* strains were negative by PCR for CJE0731 whereas, three low invasive *C. jejuni* (C27/14, C69/2, C110/4) strains showed negative results for CJE1128 in this group.

PCR screening of 9 additional low invasive *C. jejuni* isolates for genes present or highly divergent in all hyperinvasive *C. jejuni* (group 1 and group 2, Table 4.1a and 4.1b) showed a strong statistical correlation ($p=0.01$ at 1% significance level) between the presence and absence of these genes in the hyperinvasive *C. jejuni* for 13/15 genes. CJE1128 (hypothetical protein) ($p=0.011$) and CJE0731 (type III restriction modification enzyme) (0.517) were the two exceptions. This result suggests that although the genes in group 1 and 2 are not part of the same functional group or pathway these genes could be considered as genetic markers for the hyperinvasive phenotype. Previously, a cluster of six genes (*cj1321-cj1326*) in the O-linked glycosylation locus were identified, using parsimony based algorithm, as characteristic of chicken/livestock campylobacters. This gene cluster present in 16/17 isolates, was identified in an additional six chicken isolates that were not tested in the original study confirming that these genes: *cj1321* to *cj1326*, were genetic markers for livestock/chicken isolates (Champion *et al.*, 2005).

4.3.5 Functional importance of hyperinvasive linked loci identified by CGH

The phylogenomics study identified 67 genes which belonged to different functional categories. The functional significance of the presence or absence of these genes in the hyperinvasive *C. jejuni* strains, particularly for the genes listed in group 1 and 2 (Table 4.1a and 4.1b) is discussed below;

4.3.5.1 Group 1: Genes present in the all hyperinvasive *C. jejuni*

Group 1 had nine genes present in all hyperinvasive *C. jejuni* strains but highly divergent in the majority of the low invasive *C. jejuni* (Table 4.2). Important metabolic genes including carboxylate reductase and methyltetrahydrofolate reductase, tRNA synthesis genes *i.e.* histidyl-tRNA synthase and leucyl/phenylalanyl tRNA synthase formed this group. Metabolic and biosynthetic and DNA and RNA processing genes have been reported as an integral part of the *C. jejuni* core genome (Parker *et al.*, 2006, Pearson *et al.*, 2003, Dorrell *et al.*, 2001). The conservation of these genes may confer increased survival chances for bacteria especially in the hostile environment inside the human host. A histidyl tRNA synthetase encoding gene (*hisS*-CJE0856) was present in all hyperinvasive *C. jejuni* strains. In *C. jejuni* RM1221, *hisS* is a 1,227 bp (408 amino acids) long gene. Histidyl tRNA synthetase (HisRS) is an enzyme which is composed of a small peptide chain of about 420-550 amino acids. HisRS has important catalytic properties in several regulatory mechanisms of cell metabolism. HisRS is responsible for synthesizing histidyl-transfer RNA, which is important for inserting histidine into protein structures. HisRS has been found to act as an antigen responsible for causing autoimmune diseases in humans such as rheumatic arthritis or myositis (Freist *et al.*, 1999). Using comparative phylogenomics Aklujkar and Lovely (2010) showed that the attenuation of *hisS* resulted in physiological changes in *Pelobacter carbinolicus* by altering its iron III reduction pathway.

Another gene in Group 1, which was found to be present in all the hyperinvasive *C. jejuni*, is *aat* (CJE1252 in *C. jejuni* RM1221). It is a 648 bp long gene. It encodes for a Leucyl/phenylalanyl-tRNA protein transferase and contains 415 amino acids. The role of *aat* is to transfer a Leucine or Phenylalanine to the N terminal of Arginine or Lysine containing proteins to allow their breakdown (Shrader *et al.*, 1993). The function of *aat* in *C. jejuni* pathogenesis is not yet characterised.

A metabolic gene present in all hyperinvasive *C. jejuni* strains and highly divergent in the majority of low invasive *C. jejuni* is *proC* (CJE1219 in *C. jejuni* RM1221), encoding for pyrroline-5-carboxylate synthase. CJE1219 is 732 bp (199 amino acids) in size in *C. jejuni* RM1221. This is a key enzyme of proline metabolism involved in the synthesis of L-proline and ornithine. Proline has a well-studied function related to maintaining redox balance under osmotic stress in several bacteria (Perez-Arellano *et al.*, 2010). Previously, in a transposon mutagenesis study of hyperinvasive *C. jejuni* 01/51, a transposon was inserted in a metabolic gene *putA* (Cj1503c in *C. jejuni* NCTC11168) encoding for a putative proline dehydrogenase/delta-1-pyrroline-5-carboxylate dehydrogenase, an enzyme involved in the oxidation of proline into glutamate. The transposon inserted in *putA* reduced invasion of the *C. jejuni* 01/51 mutant into INT-407 and Caco2 cells (Javed *et al.*, 2010). Another metabolic gene in Group 1, is *metF* (CJE1336 in *C. jejuni* RM1221). CJE1336 is a 849 bp long gene and encodes 282 amino acids. In *C. jejuni* RM1221, *metF* encodes for a metabolic enzyme 5, 10-methylenetetrahydrofolate reductase. This protein is a flavoprotein that catalyses the conversion of 5-methyltetrahydrofolate to 5, 10-methylenetetrahydrofolate.

The presence of genes related to metabolic and regulatory pathways in hyperinvasive *C. jejuni* might reflect that the hyperinvasive *C. jejuni* do not have well characterised pathogenicity factors as in other enteric pathogens like *E. coli* and *Salmonella enterica* but are opportunistic pathogens. In 1988, Stanley Falkow devised the molecular version of Koch's postulates by defining the "virulence factor" in a pathogen and stated that a virulence factor must be present in a pathogenic strain and absent from the non-pathogenic strain. Pallen and Wren (2007) argued that the definition of virulence factors cannot be strict as many bacteria adapt to different functions in different conditions. For example, the uropathogenic *E. coli* lives in the human intestine but acts as a pathogen in the human bladder. Similarly, the enterohaemorrhagic *E. coli* is commensal in bovine intestine but causes disease when inside the human gut (Pallen and Wren., 2007). Likewise, *C. jejuni* is commensal in chickens and a pathogen in humans. Thus, the *C. jejuni* infection in humans is a multifactorial mechanism that results from the requirement of bacteria to survive inside the human host. In a recent study, Hofreuter *et al* (2008) reported that the subtle genomic changes in *C. jejuni* metabolic pathways significantly influenced their ability to utilize nutrients and colonize specific host tissues. This suggests that the metabolic diversity is an important factor in this organism in defining host specificity.

A molybdenum ABC transporter gene, *modA* (CJE0348 in *C. jejuni* RM1221) was present in all hyperinvasive *C. jejuni* strains but highly divergent in 10/13 of the low invasive *C. jejuni* isolates studied. CJE0348 is a 750 bp (249 amino acids) in size in *C. jejuni* RM1221. The putative molybdenum ABC transporter binding protein (*modA*) is a part of molybdenum transport apparatus *modC* CJE0345 (Cj0300c), *B* CJE0346 (Cj0301c) and *A* CJE0348 (Cj0302) and it is present in the hypervariable Plasticity Region (PR) 1 (Pearson *et al.*, 2003) and Plasticity Region (PR) 2 (Dorrell *et al.*, 2001) in *C. jejuni* strains identified in comparative phylogenomics studies (Pearson *et al.*, 2003, Dorrell *et al.*, 2001). Molybdenum has been shown to have a role in nitrate metabolism by the catalytic activity of a flavoprotein enzyme, nitrate reductase, enabling nitrate to act as a terminal electron acceptor in place of oxygen under oxygen deficient conditions in *Campylobacter* (Sellars *et al.*, 2002, Pearson *et al.*, 2003). DNA microarray studies based on NCTC11168 as the reference strain (Parker *et al.*, 2006) showed that the *C. jejuni* RM1221 and other investigated *C. jejuni* strains were diverse in the molybdenum ABC transport region. However, in the current study this region was present in all hyperinvasive *C. jejuni* strains which may reflect the increased fitness of these strains compared to the low invasive *C. jejuni* to allow survival in diverse ecological niches and utilize a variety of metabolic resources.

Another gene, CJE0320 (in *C. jejuni* RM1221) is present in all hyperinvasive *C. jejuni* and highly divergent in the majority of low invasive *C. jejuni*. The size of this gene is 456 bp (151 amino acids). In *C. jejuni* RM1221, CJE0320 is annotated as peroxiredoxin enzyme in the antioxidant/AhpCTCA family. Cj0271 and CCO0340 are the homologues of this gene in *C. jejuni* NCTC11168 and *C. coli* RM2228 respectively. In *C. jejuni* NCTC11168, Cj0271 has been shown to have a role in reducing hydrogen and organic peroxides, thus conferring resistance under oxidative stress conditions (Atack *et al.*, 2008).

Interestingly, *modA* and CJE0320 have related functions and their presence in the hyperinvasive *C. jejuni* strains in this study may provide advantage under oxidative stress conditions.

In group 1, three genes (CCOA0033 in *C. coli* RM2228, CJE0838 and CJE0669 in *C. jejuni* RM1221) with a putative role were present in all the hyperinvasive *C. jejuni* strains. CCOA0033 is a 213 bp gene encoding 70 amino acids. CJE0838 and CJE0669 are 957 bp (318 amino acids) and 207 bp (68 amino acids) in size respectively. Fouts *et*

al (2005) identified several hypothetical genes in the genome sequences of different *Campylobacter* strains. These genes add to the genomic diversity of this organism.

A periplasmic membrane encoding gene *lolA* (CJE1021 in RM1221) (Group 5; supplementary table 4) was present in five hyperinvasive *C. jejuni* and three low invasive *C. jejuni* strains except and highly divergent in *C. jejuni* 01/04 and the low invasive *C. jejuni* strain 01/32. Two other genes (Cj0544, Cj0151c in NCTC11168) with a predicted role as a periplasmic protein were also included in group 5. The homologues of these genes in *C. jejuni* RM1221 (*i.e.* CJE0648 and CJE0147) have no function characterized. In *C. jejuni*, outer membrane proteins have been found to play a role in virulence in particular in the adherence and invasion of intestinal epithelial cells. The role of surface exposed proteins including, major outer membrane proteins (MOMP-PorA), JlpA, PEB1a, *Campylobacter* adhesion to fibronectin (CadF) and FlpA in adherence and internalization of cultured epithelial cells is known in *C. jejuni* (Jin *et al.*, 2001, Pei *et al.*, 1998, Monteville *et al.*, 2003, Krause-Gruszczynska *et al.*, 2007, Flanagan *et al.*, 2009). The function of these genes is not characterised yet in the *C. jejuni* strains studied here but taking into account the importance of outer membrane proteins in host cell interaction, these genes may have a potential role in virulence in particular invasion of hyperinvasive *C. jejuni* strains.

4.3.5.2 Group 2: Genes highly divergent/absent in all hyperinvasive *C. jejuni*

Based on CGH data, a second group of six genes were identified as divergent in all hyperinvasive *C. jejuni* but present in most of the low invasive *C. jejuni* strains studied (Table 4.2). It can be argued that in the case of this group the presence of genes in the low invasive *C. jejuni* strains may be responsible in reducing the invasion potential of these strains. The process of gene loss or genome decay is evident in some bacterial pathogens. The best examples where the pathogens have lost genes to acquire smaller genomes in order to adapt to the specialised host niches are *Yersinia pestis* and *S. enterica* serovar Typhi (Pallen and Wren., 2007).

A hypothetical protein (CJE0315) was found to be highly divergent in all hyperinvasive *C. jejuni* and present in 12/13 low invasive *C. jejuni* (01/32, 01/46, 01/39) but highly divergent in low invasive *C. jejuni* 01/30. In *C. jejuni* RM1221, this gene is a 510bp long and the translated protein contains 169 amino acids. In NCTC11168 and *C. coli* RM2228, this gene is annotated as a putative integral membrane protein (*i.e.* Cj0266c

and CCO0335 respectively). There are other membrane associated protein encoding genes found to be absent from the majority of the hyperinvasive *C. jejuni* strains. For example, a membrane protein encoding gene (CJE0989) annotated as SCO1/SenC family protein in *C. jejuni* RM1221 (Group 3; supplementary table 4) and a porin domain protein CJE1165 in *C. jejuni* RM1221 (Group 6; supplementary table 4) characterised as a probable periplasmic protein were highly divergent from the majority of the hyperinvasive *C. jejuni* strains but present in the most of the low invasive *C. jejuni*. This may suggest that these genes may not have a role in invasion phenotype of the hyperinvasive group of *C. jejuni* strains.

Another gene highly divergent in all hyperinvasive *C. jejuni* strains is CJE0801. This gene is a 1,254 bp (417 amino acids) long and has similarity to peptidase U32, in RM1221. The role of this peptidase in *C. jejuni* pathogenesis is not known. Periplasmic proteases are an important part of the cell envelope of Gram negative bacteria as they provide resistance against extracellular stresses including oxygen and temperature fluctuations. CJE0801 belongs to the functional category of post-translational modification, protein turn over, chaperone. In *C. jejuni*, a periplasmic protease HtrA, has been shown to provide protection against heat and oxidative stress conditions in *C. jejuni* (Baek *et al.*, 2011b). Recently, the role of a peptidoglycan peptidase (*pgp1*) in *C. jejuni* virulence has been characterised using *C. jejuni* 81-176 as the model organism (Frirdich *et al.*, 2012). In *C. jejuni* 81-176, has been shown to be involved in maintaining the helical cell shape and in other virulence phenotypes mainly altered innate immune response, motility and biofilm formation (Frirdich *et al.*, 2012). Since, CJE0801 was missing from all the hyperinvasive *C. jejuni* strains; this may suggest that this gene may be involved in suppressing invasion in the low invasive *C. jejuni* strains.

A tRNA processing and modification gene, queuine tRNA ribosyltransferase (*tgt*) (CJE1090 in *C. jejuni* RM1221) was highly divergent/absent from all hyperinvasive *C. jejuni* but present in all low invasive strains except *C. jejuni* 01/39. In *C. jejuni* RM1221, CJE1090 is 1,122 bp in size and encodes a 373 amino acids polypeptides. The function of *tgt* in the bacterial translational process is to catalyse the incorporation of queuine which is a hyper-modified base in the wobble position of the anticodon of tRNAs by an exchange reaction with guanine (Fouts *et al.*, 2005).

A type III restriction/modification enzyme encoding gene CJE0731 in *C. jejuni* RM1221 was found to be highly divergent in all hyperinvasive *C. jejuni* strains and present in

only three low invasive *C. jejuni* (01/30, 01/32, 01/46). CJE0731 in *C. jejuni* RM1221 is 2,028 bp (675 amino acids) in size. In contrast, a type I restriction enzyme M protein (Cj1553c-*hsdM*) was found to be present in majority of hyperinvasive *C. jejuni* strains except in *C. jejuni* 01/51 and highly divergent from three low invasive *C. jejuni* strains but present in low invasive *C. jejuni* 01/30 (Group 5, supplementary table 4). Restriction modification regions (RM) are variable regions in the *C. jejuni* genome. There are different types of RM systems characterised in *C. jejuni* strains referred to as type I, II and III. *C. jejuni* strains have characteristic RM regions that show strain to strain variation. The diversity seen in the RM systems in *C. jejuni* might reflect it as hotspot for horizontal gene transfer like within other variable regions in the *C. jejuni* genome including CAP, LOS and FM regions (Miller *et al.*, 2005). For example, *C. jejuni* 81-176 has its unique type I restriction modification system which is absent from *C. jejuni* RM1221 and NCTC11168 (Hofreuter *et al.*, 2006). Another *C. jejuni* strain, ATCC43431, has all restriction modification systems as present in *C. jejuni* NCTC11168 and also contains additional complements of RM systems (Poly *et al.*, 2004). Previously, RM regions have been reported as highly divergent in NCTC11168 (Cj0625–Cj0629) and RM1221 (CJE0731 and CJE0732) in a number of *C. jejuni* strains studied by DNA microarray (Parker *et al.*, 2006, Dorrell *et al.*, 2001). This reflects that RM systems are dispensable in *C. jejuni* and may not necessarily influence virulence but are required to generate diversity in genomes.

4.3.5.3 Important loci present or variable in other groups

Flagellar biosynthesis loci have been well studied virulence determinants in *C. jejuni* (Duong and Konkel., 2009, Rodin *et al.*, 2008, Parker *et al.*, 2006, Champion *et al.*, 2005, Pearson *et al.*, 2003, Dorrell *et al.*, 2001). In our study, the flagellar basal-hook body protein (*fliE*) was present in five hyperinvasive *C. jejuni* but highly divergent in the hyperinvasive *C. jejuni* 01/51 (Group 5; supplementary table 4). This flagella gene, *fliE* was highly divergent in three of four low invasive *C. jejuni*. However, *C. jejuni* 01/51 exhibited fully motile profile in our study (section 3.4.5). Based on the importance of flagella in *C. jejuni* virulence and survival inside the human host and its fully motile phenotype (Grant *et al.*, 1993, Nachamkin *et al.*, 1993, Morooka *et al.*, 1985) it may be argued that the divergence of *fliE* in *C. jejuni* 01/51 might only be the sequence variation rather than the absence of gene. It would be interesting to re-confirm the absence of *fliE* in *C. jejuni* 01/51 by PCR or to be more precise by Southern blotting.

Two genes (*cfrA*; CJE0847 and *ceuB*; CJE1541) encoding for ferric receptor and iron transport permease proteins respectively were also identified in this study (Group 5; supplementary table 4). These genes were present in five of six hyperinvasive *C. jejuni* strains and highly divergent from three of four low invasive *C. jejuni*. The iron receptor, CfrA and CeuB are important components of the ferric enterobactin transport system in many *C. jejuni* strains. These receptors have a high affinity for iron and scavenge iron molecules from inside the gastrointestinal tract by forming ferric enterobactin complex which are transported through the bacterial periplasmic membrane (Zeng *et al.*, 2009). The role of *cfrA* and *ceuB* genes in iron uptake system and in chicken colonization has been well investigated in *C. jejuni* (Stintzi *et al.*, 2008, Palyada *et al.*, 2004). A mutant in the *cfrA* gene, completely abolished enterobactin mediated transport and chicken colonization potential compared to the parent strain where the colonization was recorded to 10^7 cfu/ml of caecal content (Palyada *et al.*, 2004). Recently, another ferric enterobactin receptor, CfrB was identified in *Campylobacter* with a role in iron acquisition and chicken colonization. This study showed that inactivation of the *cfrB* gene greatly reduced colonization of the chicken intestine (Xu *et al.*, 2010).

Another important gene CCO0783 (*arsC*) in *C. coli* RM2228 encoding for arsenic resistance has been found as present in the majority of hyperinvasive *C. jejuni* and absent/highly divergent from the majority of low invasive *C. jejuni* strain (Group 5; supplementary table 4). Arsenic is a naturally present toxic metalloid which is an additive in poultry feed. Poultry being the main reservoir of *Campylobacter* species the isolates have developed marked arsenic resistance. A well characterised arsenic resistance gene *arsC* in *C. jejuni* encodes for an arsenate reductase that converts arsenate into arsenite (Wang *et al.*, 2009). A previous study used Suppressive Subtractive Hybridization to show the presence of *arsC* in *C. jejuni* NCTC11168 and absence from *C. jejuni* 81116 (Ahmed *et al.*, 2002). In the recently sequenced *C. jejuni* RM1221 four genes with three of them similar to *ars* genes have been identified (Fouts *et al.*, 2005). These *ars* genes have homologues in other sequenced *C. jejuni* strains including *C. jejuni* NCTC11168 (Parkhill *et al.*, 2000) and *C. jejuni* 81-176 (Hofreuter *et al.*, 2006). Wang *et al* (2009) has characterised *ars* operon in *C. jejuni* RM1221 and found a number of clinical *C. jejuni* isolates to show very high levels of arsenic resistance. The presence of *arsC* in most of the hyperinvasive *C. jejuni* and absence in majority of low invasive might signify increased fitness of hyperinvasive *C. jejuni* in the environment compared to the low invasive *C. jejuni*.

Another gene CJE1310 in *C. jejuni* RM1221 encoding for twin-arginine translocation protein (TatA/E) was found to be present in 5 hyperinvasive *C. jejuni* except in *C. jejuni* 01/35. This gene was found as absent/highly divergent in 3 low invasive *C. jejuni* but present in low invasive *C. jejuni* 01/30 (Group 5, supplementary table 4). In bacteria, the twin-arginine translocase (TAT) secretion system has a role in identifying partially or fully folded proteins and transports them across the cytoplasmic membrane. With an increasing number of *C. jejuni* genome sequences becoming available a number of TAT related genes are being identified (Rajashekara *et al.*, 2009). A knockout mutant in *tatC* was found to be deficient in virulence related phenotypes including biofilm formation, motility, flagella synthesis, increased antimicrobial resistance and survival under environmental stresses (Rajashekara *et al.*, 2009). Rajashekara *et al.* (2009) also showed the importance of *tatC* in chicken colonization. The presence of a twin-arginine related gene in the majority of hyperinvasive *C. jejuni* and absence in most of the low invasive *C. jejuni* might contribute to the improved stress responses and virulence related traits in the hyperinvasive *C. jejuni*. It would be interesting to characterize the role of this gene in all hyperinvasive *C. jejuni*.

A DNA methyltransferase encoding gene CJE1077 (*gidB*) was identified as highly divergent/absent in 5 hyperinvasive *C. jejuni* but present in hyperinvasive *C. jejuni* EX114. This gene was highly divergent in 3/4 low invasive *C. jejuni* (Group 6, supplementary table 4). In a recent study, a gene Cj1461 encoding for a putative DNA methyltransferase was found to have a role in the regulation of virulence in *C. jejuni* (Kim *et al.*, 2008). In that study, a mutant in Cj1461 showed reduced adhesion and motility, but increased invasion. CJE1077 (*gidB*) may have a potential role in *C. jejuni* virulence but it has not been characterised as yet.

It is important to analyse the sequence of genes which are identified as being present in the majority of hyperinvasive *C. jejuni* and in one low invasive *C. jejuni* strain or vice versa to account for any gene sequence variation which cannot be detected by the DNA microarray or PCR.

In summary, CGH identified genes of diverse functions. The majority of genes have functions related to the essential cellular and metabolic pathways. Some other genes with role in virulence are also identified. The genes identified as being present and highly divergent/absent in all the hyperinvasive *C. jejuni* strains are the markers of hyperinvasion phenotype as being statistically associated with this group of strains.

4.3.6 Distribution of hyperinvasive loci in the pan genome

The 67 genes identified as present or highly divergent in the hyperinvasive *C. jejuni* strains compared against the low invasive *C. jejuni* and vice versa were classified into six groups (Table 4.1a and 4.1b, and supplementary table 4 in appendix). The distribution of these 67 loci was plotted on the pan genome to see whether any of the selected genes were present in the same operon or functionally identical clusters (Figure 4.5).

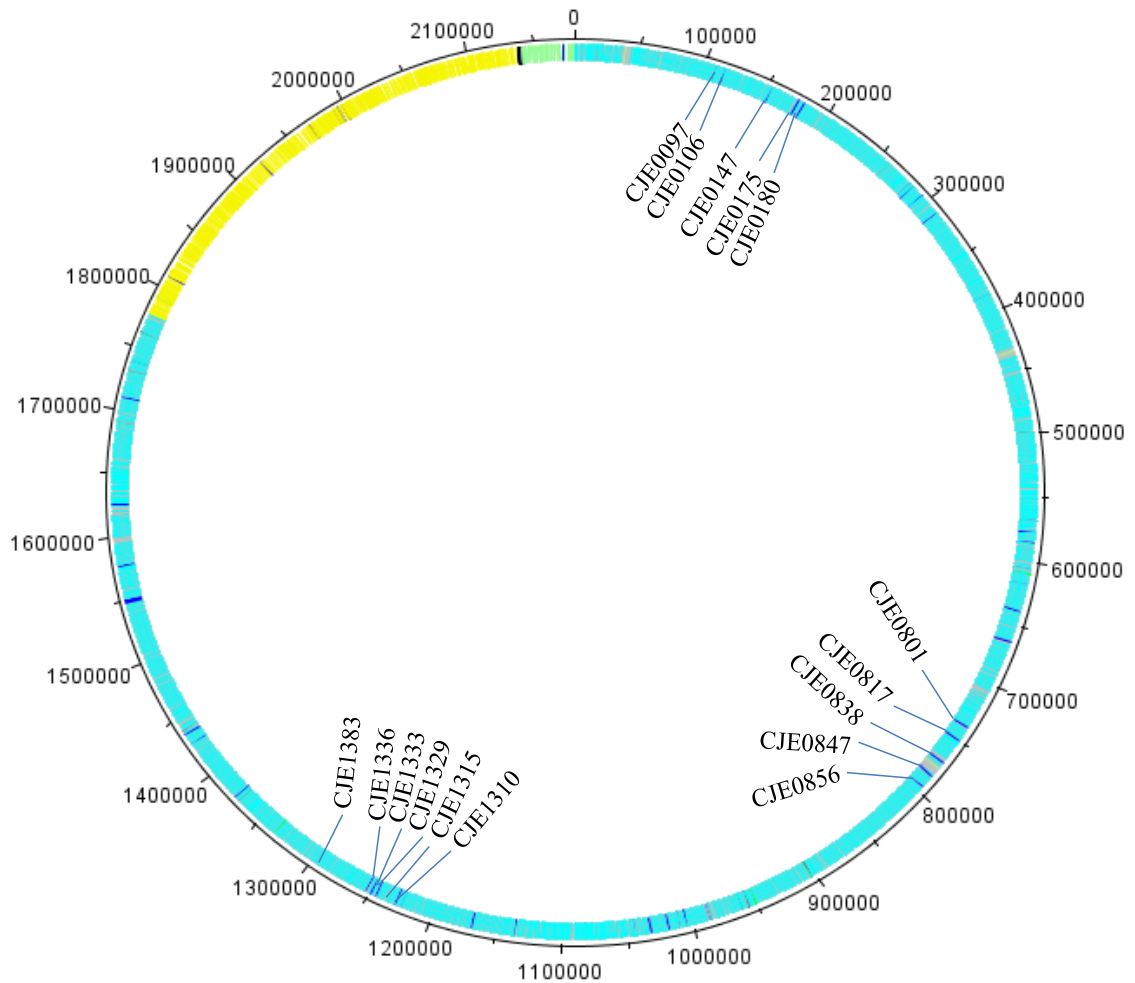


Figure 4.5: Circular diagram of the *C. jejuni/C. coli* pan genome showing the distribution of 67 hyperinvasive-associated loci. The region on the pan genome starting from mark '0' and coloured sky blue represents the *C. jejuni* RM1221 genome. The yellow region shows additional genes present in *C. coli* RM2228. The light green and black regions are the loci unique to *C. jejuni* 81-176 and *C. jejuni* 11168 respectively. The selected 67 CDSs (Table 4.1a and 4.1b, and supplementary table 4 in appendix) that were conserved or highly divergent in the hyperinvasive compared to the low invasive *C. jejuni* in this phylogenomics study are highlighted in a dark blue colour. This diagram was generated using the Artemis genome viewing tool (Rutherford *et al.*, 2000). The representative position of some loci which are present close to each other on the pan genome is also shown.

Overall, the majority of 67 genes were randomly distributed across the pan genome. None of the genes belonged to the same operon. The figure 4.5 showed that the genes, CJE0097, CJE0106, CJE0147, CJE0175, *purN* (CJE0180), are present close together

between 100,000 bp and 200,000 bp positions on the pan genome but these genes are not adjacent or functionally identical. Similarly, a group of five genes (CJE0801, CJE0817, CJE0838, *cfrA* (CJE0847) and *hisS* (CJE0856) are present between 700,000 bp and 800,000 bp locations and 6 genes (CJE1310, *tsf* (CJE1315), CJE1329, CJE1333, *metF* (CJE1336) and CJE1383) are located between 120,0000 bp and 130,0000 bp positions on the pan genome. All the genes within these clusters are not part of the same functional category. The phylogenomic analysis of hyperinvasive *C. jejuni* strains indicates that the hyperinvasive phenotype may not be defined by the presence or absence of gene clusters of one particular functional group. This unique phenotype may be a function of genes of different cellular pathways.

4.3.7 Variability in the hyperinvasive C. jejuni at the whole genome level based on CGH data

Given that no obvious clusters of the previously identified 67 loci were found it was decided to see the overall diversity in the hyperinvasive and low invasive *C. jejuni* strains and to identify genomic regions that were hotspots for diversity (Figure 4.6).

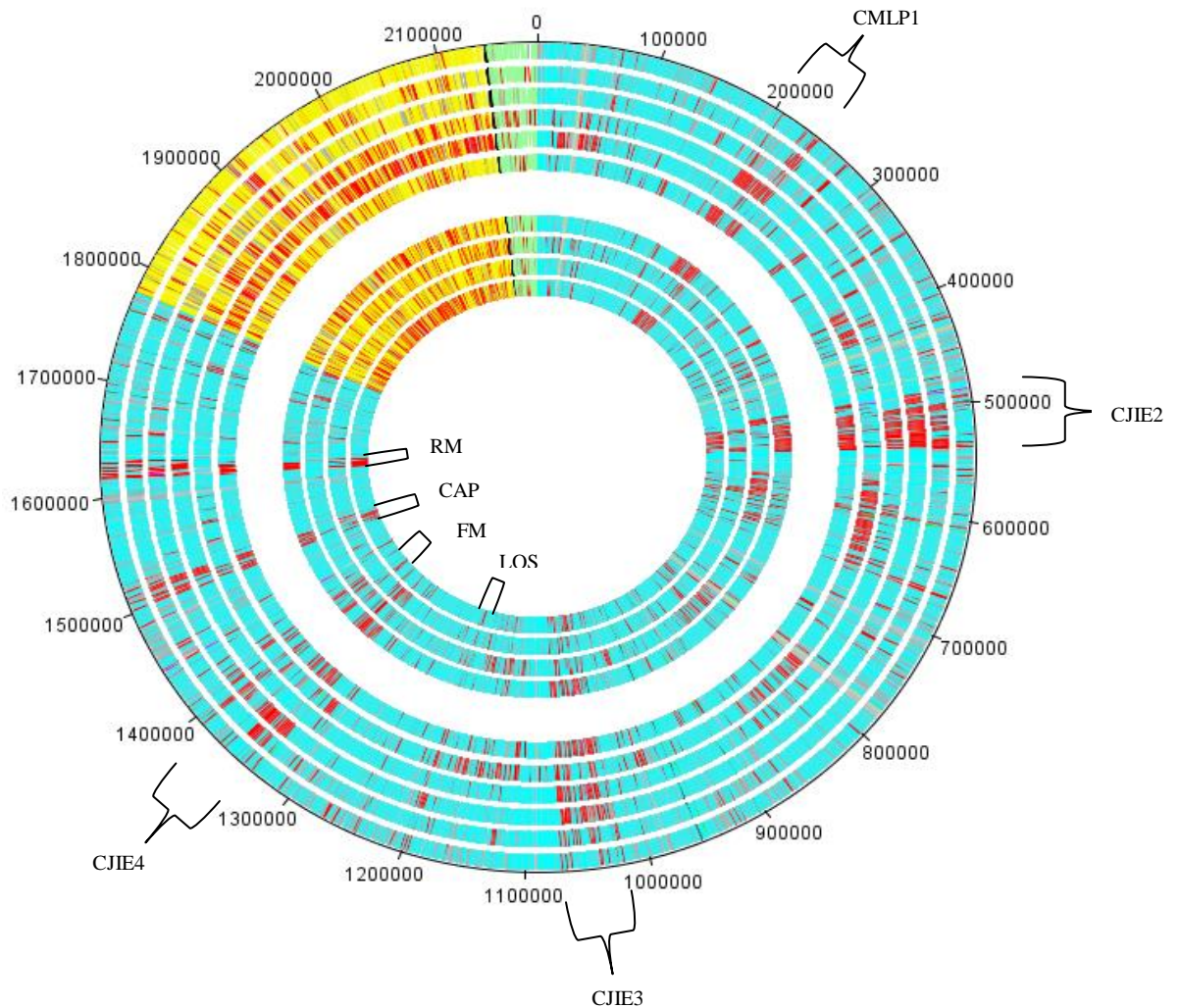


Figure 4.6: The colour coded diagrammatic representation of the CGH pan genome showing genes slightly or highly divergent/absent in the six hyperinvasive and four low invasive *C. jejuni*. The *C. jejuni* RM1221 genome on the pan array is coded sky blue. The genes unique to other *C. jejuni* sequenced strains and *C. coli* RM2228 are added to the end of *C. jejuni* RM1221 sequence and include the unique *C. coli* RM2228 CDS (Yellow), unique *C. jejuni* 11168 CDS (Black) and *C. jejuni* 81-176 CDS (Green). The outer six circles (outwards to inwards) represent hyperinvasive *C. jejuni* strains in the order of *C. jejuni* 01/10, 01/35, 01/04, 01/41, 01/51, EX114 followed by four low invasive *C. jejuni* strains, 01/30, 01/32, 01/46, 01/39. Slightly divergent genes are coloured grey and highly divergent genes are red as identified by CGH for each *C. jejuni* strain. CMLP (*Campylobacter* Mu like prophage), CJIE 2, 3, 4 (*Campylobacter jejuni* integrated element), LOS (Lipooligosaccharide), FM (Flagellar modification), CAP (Capsule polysaccharide), RM (Restriction modification) regions in *C. jejuni* RM1221 are labelled.

Our results showed that both groups of *C. jejuni* strains were very diverse irrespective of their invasion phenotype. In the majority of *C. jejuni* strains investigated by CGH, the most variability appeared in the *C. jejuni* RM1221 specific prophages and other mobile genetic integrated regions including CMLP1, CJIE2, CJIE3, CJIE4 (Fouts *et al.*, 2005). Virulence related surface structures in *C. jejuni* including capsular polysaccharides (CAP) and lipo-oligosaccharides (LOS), Restriction Modification (RM) and Flagellar Modification (FM) regions were also variable amongst most strains (Figure 4.6). The result is not surprising as these regions showed a great degree of variation in other *C. jejuni* strains with different phenotypes, originating from a variety of sources and disease profiles (Parker *et al.*, 2006, Pearson *et al.*, 2003, Dorrell *et al.*, 2001). Pearson *et al.* (2003) studied genomic diversity in 18 *C. jejuni* isolates from a variety of sources using a NCTC11168 DNA microarray. This study identified seven hypervariable genomic regions called Plasticity Regions (PR). Most *C. jejuni* strains were highly divergent in PR5 and 6 which included capsule, LOS biosynthesis and flagella modification genes (Pearson *et al.*, 2003). In the other phylogenomics study by Parker *et al.* (2006) a collection of 67 *C. jejuni* and 12 *C. coli* strains were studied using a *C. jejuni* NCTC11168 and RM1221 combined array. This study showed that the majority of the strains investigated were lacking/highly divergent in the unique *C. jejuni* RM1221 CJIEs. These tested strains were also highly variable in the capsule and LOS regions (Parker *et al.*, 2006).

C. jejuni and *C. coli* are closely related species with the majority of genes common in the core and dispensable part of their pan genome. Still the two species contain unique sets of genes that define them as separate species (Lefebure *et al.*, 2010). This variation was clearly shown in all *C. jejuni* strains (Figure 4.6) whereby there are many of the *C. coli* specific genes (yellow) that are highly divergent in the *C. jejuni* strains tested here. This also shows a clear advantage of a *C. jejuni* and *C. coli* pan array used in the comparative study over a single genome based arrays used in previous DNA microarray studies (Dorrell *et al.*, 2001, Pearson *et al.*, 2003) as it covers diversity across the *Campylobacter* genus.

The diversity seen in figure 4.6 agrees with the phylogenomic clustering of the six hyperinvasive and four low invasive *C. jejuni* strains (Figure 4.3). The hyperinvasive *C. jejuni* 01/10 and *C. jejuni* 01/35 were most similar to the reference strains on the pan array with the least number of divergent genes *i.e.* (522 and 550 respectively). The most

diverse across the *C. jejuni*/*C. coli* pan genome was *C. jejuni* 01/51 with 859 divergent genes. The hyperinvasive *C. jejuni* 01/10 possessed some of the loci in CMLP1 and the genes in other *C. jejuni* RM1221 characteristic genomic islands also appeared to be present in this strain. Similarly, most genes in the LOS and FM regions were found to be similar to those present in the reference *C. jejuni* RM1221, whereas, some genes in the CPS region and the RM locus were highly divergent from *C. jejuni* RM1221 (Figure 4.6).

Interestingly, unlike the majority of other *C. jejuni* strains investigated in this phylogenomics study it appears that the *C. jejuni* RM1221 integrated genetic elements CMLP1, CJIE2 and RM region were present in the hyperinvasive *C. jejuni* 01/51. The LOS and FM loci appeared to be highly divergent from those found in *C. jejuni* RM1221. Some genes, including the *kps* genes, in the capsule region of *C. jejuni* RM1221 were also found to be present in *C. jejuni* 01/51. In a previous study, using a *C. jejuni* NCTC11168 based DNA microarray the capsule locus of *C. jejuni* 01/51 was reported as absent (Dr Georgina Manning; personal communication).

4.4 SUMMARY AND CONCLUSIONS

The comparative phylogenomics study of six hyperinvasive compared against four low invasive *C. jejuni* strains showed that the hyperinvasive *C. jejuni* did not form a distinct cluster separate from the low invasive *C. jejuni*. There was no gene or gene cluster present or highly divergent/absent in all hyperinvasive *C. jejuni*. However, a number of genes were found to be statistically associated with the hyperinvasion phenotype either by being present or absent from the majority of these strains. These represented genes from a variety of functional categories including genes involved in cellular and metabolic functions, some pathogenesis loci and several genes encoding for hypothetical proteins. Among these, nine genes were present and six genes were absent/highly divergent in all six hyperinvasive *C. jejuni*. PCR screening showed the distribution of these 15 genes in an additional population of 9 low invasive *C. jejuni* strains. This confirmed that the presence or absence of these 15 genes as a genetic marker for the hyperinvasive *C. jejuni*. This DNA microarray study showed that all 10 *C. jejuni* strains showed great diversity at the whole genome level irrespective of their invasion phenotype. The most variable regions in the majority of *C. jejuni* strains were CMLP1, CJIE 2, 3, 4 characteristic of *C. jejuni* RM1221 and *C. jejuni* virulence associated

regions like flagellar modification, restriction modification, lipooligosaccharide and capsule. Based on this comparative phylogenomics study it is possible to conclude that the hyperinvasive phenotype is a multifactorial function involving genes from a variety of pathways. Genes involved in essential cellular process that form a part of the core genome may have key a role in the hyperinvasive phenotype in addition to the known *C. jejuni* pathogenesis related loci. Additionally, each of the hyperinvasive *C. jejuni* may use different mechanisms resulting in the hyperinvasive phenotype rather than same gene/gene clusters or pathways.

4.5 LIMITATIONS OF DNA MICROARRAY AND NEXT STEP

A major drawback of CGH is that the array is based on the genomic content of already sequenced genomes limiting the information generated to the variation of genes compared to the reference genomes for the phylogenomic comparisons. In addition, DNA microarray does not take into account point mutations, deletions and genomic rearrangements. Also, lack of representation of promoter regions and non-translated RNAs are some other limitations to this approach (Champion *et al.*, 2005, Pearson *et al.*, 2003). This suggests that the *C. jejuni* strains investigated might contain additional genomic content that could not be picked up in DNA microarray. A quantitative bias is introduced by using all genes on the array for analysis. A number of false positive or negative genes can be detected. To eliminate these will require further validation by alternate screening methods like PCR.

To address the drawbacks of CGH and investigate the additional genomic content present in the hyperinvasive *C. jejuni* a Pooled Suppressive Subtractive Hybridization (PSSH) technique was performed (Gerrish *et al.*, 2010). The aim of PSSH was to identify any genomic regions that were unique to the pool of six hyperinvasive *C. jejuni* strains compared against the four low invasive *C. jejuni*. This approach will be discussed in the next chapter.

Chapter Five

**SUPPRESSIVE SUBTRACTIVE
HYBRIDIZATION STUDY OF THE
HYPERINVASIVE *C. JEJUNI*
STRAINS**

SUPPRESSIVE SUBTRACTIVE HYBRIDIZATION STUDY OF THE HYPERINVASIVE *C. JEJUNI* STRAINS

5.1 INTRODUCTION

After the phylogenomic comparison performed using CGH discussed in the previous chapter, the hyperinvasive *C. jejuni* strains were further investigated at the genome content level by using a technique called Suppressive Subtractive Hybridization (SSH). The main objective of this study was to find genomic regions unique to the hyperinvasive *C. jejuni* strains that were absent from the low invasive *C. jejuni*. It was assumed that these genomic regions are common in all the hyperinvasive *C. jejuni* strains and would be linked to their hyperinvasive phenotype.

5.1.1 Suppressive Subtractive Hybridization as a genotyping technique

Suppressive Subtractive hybridization is a method to identify unique genomic regions present in one strain (the tester) but absent from the reference strain (the driver) (Agron *et al.*, 2002, Ahmed *et al.*, 2002). SSH complements the DNA microarray by providing additional information about the genomic structure of the test organism. Whole genome sequencing projects of a large number of strains can be costly and very time consuming. In the absence of genome sequences, SSH can be a good technique to a study closely related strains at the genome level. SSH can easily identify unique genomic regions of about 10-50kb in size and any false positive sequences can be deleted by PCR analysis. SSH has identified genomic variability in *C. jejuni* strains with different colonisation potentials (Ahmed *et al.*, 2002). In that study, *C. jejuni* strain 81116, with relatively good chicken colonization potential was compared with *C. jejuni* NCTC11168 by subtractive hybridization to find unique genomic regions in *C. jejuni* 81116 that would account for the difference in colonization potential seen between the two strains. Two hybridization cycles identified 24 clones containing sequences present in 81116 and absent from NCTC11168. Some sequences had similarities with other *C. jejuni* strains while the majority were orthologous to other bacteria. The majority of sequences were found to have functions related to the restriction modification enzymes, arsenic resistance genes, cytochrome c oxidase III, dTDP glucose 4, 6 dehydratase, gamma glutamyl transpeptidase and abortive phage resistance genes (Ahmed *et al.*, 2002). In

another study, SSH has been used to identify the virulence markers in clinical *H. pylori* isolates from children with peptic ulcers (Oleastro *et al.*, 2006). That study identified two genes with putative roles as an outer membrane protein and in lipooligosaccharide biosynthesis to be associated with peptic ulcer disease in children. Another study used SSH to explore the genomic diversity between *C. jejuni* strains from diverse sources including rabbit, cattle and wild birds representing different MLST clonal complexes (Hepworth *et al.*, 2007). Here the majority (97%) of SSH inserts identified had similarities with other *C. jejuni*. These inserts mainly encoded for metabolic genes and they were variably distributed among different clonal complexes. However there was no correlation found with the source of strain isolation suggesting that horizontal gene transfer was a rare event between the MLST types studied (Hepworth *et al.*, 2007). Using suppressive subtractive hybridization, a number of virulence associated genomic regions have been found in *Escherichia coli* (Dai *et al.*, 2010). This study compared an avian associated pathogenic *E. coli* (APEC) with the human uropathogenic *E. coli* to identify genomic regions that would differentiate the two pathotypes. Twenty eight genes were identified that were present in the APEC but absent from the UPEC strain. A genetic marker for adhesion unique to the APEC strain was found. This gene encoded for a putative autotransporter that lead to a 124 kDa adhesive protein that showed high levels of adhesion *in vitro* in chicken fibroblast cell lines (Dai *et al.*, 2010). In a recent study, enterohaemorrhagic *E. coli* (EHEC) strains of serotype O26 from a young calf and a human diarrheal patient were investigated by SSH with an aim to identify genomic markers for host preference. The genomic markers associated with host specificity were not found but a number of pathogenicity islands not associated with the EHEC strains were present in the EHEC bovine strain (Bardiau *et al.*, 2012). In all above studies, SSH was used to compare one strain with another strain in a single experiment.

5.1.2 Pooled Suppressive Subtractive Hybridization to study hyperinvasive C. jejuni

A new SSH technique called Pooled Suppressive Subtractive Hybridization (PSSH) has been developed and validated recently (Gerrish *et al.*, 2010). This method involves genomic DNA comparisons of sets of phenotypically related strains with a reference strain in a single experiment. The inserts identified can be related to the specific strains by PCR screening. Gerrish *et al* (2010) have successfully used PSSH to screen multiple clinical *Staphylococcus aureus* isolates with different levels of disease severity identifying unique genes specific to strains in each pool creating a signature profile for

the strains. In this study, the six hyperinvasive *C. jejuni* strains were pooled together as the tester and hybridized against a pool of four low invasive *C. jejuni* (driver) that were previously investigated in the comparative phylogenomic study.

5.2 METHODS

5.2.1 Pooled Suppressive Subtractive Hybridization

Suppressive Subtractive Hybridization was carried out by using the CLONE-TECH PCR™-Select Bacterial Genome Subtraction kit (Clontech, UK) according to manufacturer's instructions. Briefly, the genomic DNA from *C. jejuni* strains was prepared as stated in the genomic DNA extraction procedure in section 2.6.1. The genomic DNA (2 µg) of all six hyperinvasive *C. jejuni* strains (01/10, 01/35, 01/04, 01/41, 01/51, EX114) was mixed together as a “tester” and the genomic DNA (2 µg) of the four low invasive *C. jejuni* strains (01/30, 01/32, 01/46, 01/39) was pooled together as a “driver”. Both tester and driver genomic DNA was enzymatically fragmented with *AluI* and *DraI* to create blunt ended fragments of size 0.1 to 2.0 kbp. The fragmented tester DNA was divided into two aliquots each ligated to a different adaptor (1 or 2R). The adaptor was ligated to the 5' end of each strand of the double stranded DNA. The first hybridization was carried out by denaturing each aliquot of the adaptor ligated tester DNA and mixing separately with an excess of heat-denatured driver DNA. This hybridization step was carried out at 63 °C for 1.5 hours. In the second hybridization, the two adaptor-ligated aliquots were mixed with more freshly denatured driver DNA and incubated overnight at 63 °C. The two sets of hybridizations allowed tester DNA to hybridize with the driver DNA or if tester specific, to itself. The tester specific DNA was amplified by primary and secondary PCR cycles. The first set of subtractive hybridization was carried out at 63 °C as recommended by the manufacturer. For *C. jejuni* SSH, Ahmed *et al* (2002) repeated the second subtractive hybridization step once by using 10µl of secondary PCR product with a hybridization temperature of 61 °C. It ensured efficient subtraction by reducing the number of driver specific sequences. In this study, the secondary hybridization step was carried out twice as recommended by Ahmed *et al* (2002) since a pooled *C. jejuni* sample was used in SSH here. A flow diagram illustrates the PSSH procedure followed (Figure 5.1).

Chapter 5: Suppressive subtractive hybridization

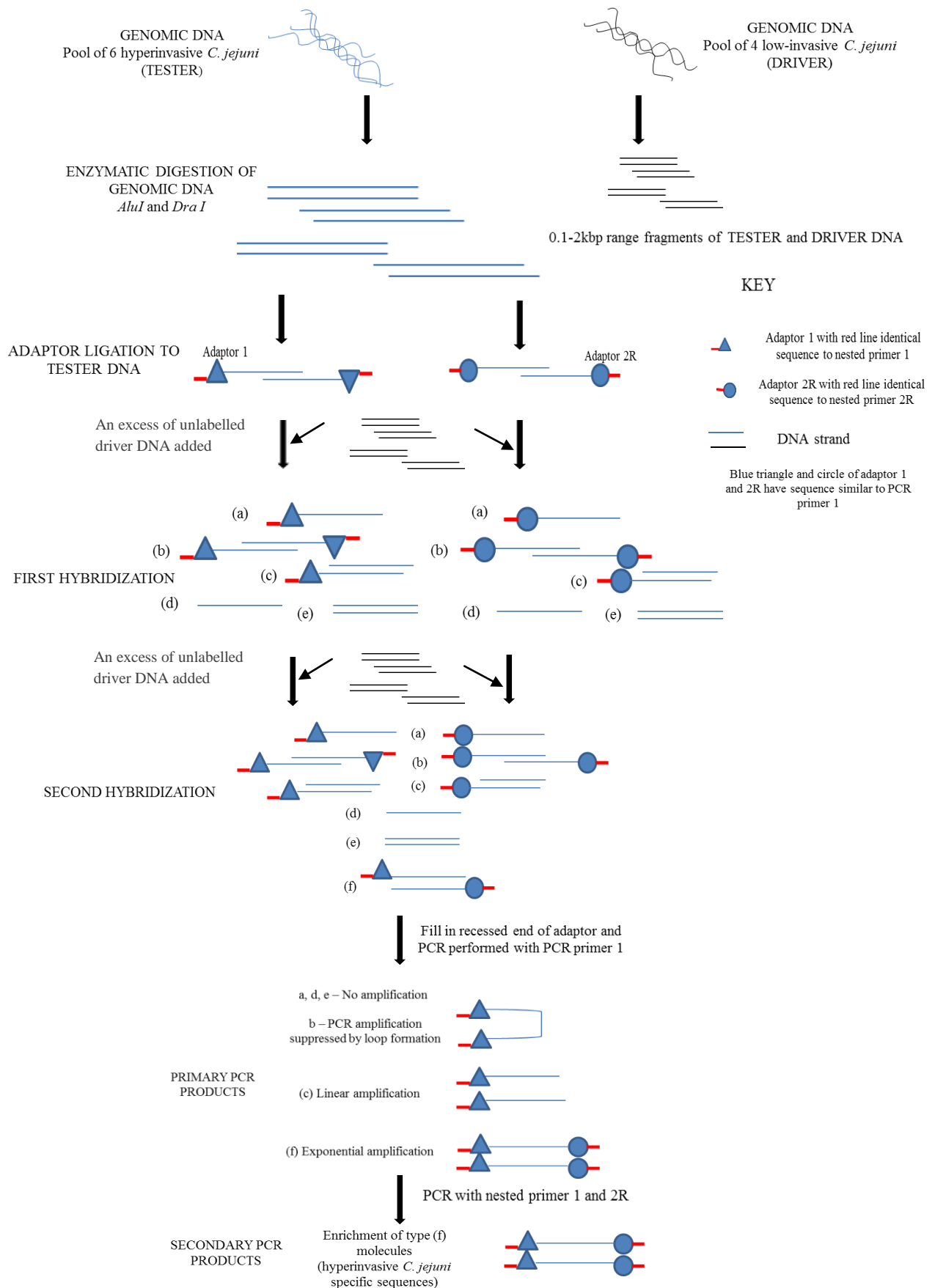


Figure 5.1: A diagram illustrating the methodology used for the pooled suppressive subtractive hybridization.

The enzymatically digested tester DNA of pool of 6 hyperinvasive *C. jejuni* strains was subdivided into two sections. Each aliquot was ligated to separate adaptor. During the first hybridization each aliquot was mixed with an excess of freshly denatured driver DNA (pool of 4 low invasive *C. jejuni*). The two aliquots were mixed in the second hybridization with the addition of more denatured driver DNA. During primer PCR, primer 1 amplified the DNA sequences specific to the tester DNA and absent from driver DNA (f molecules). The type f molecules were further enriched in the secondary PCR reaction using nested primers 1 and 2R.

The regions common between the diver and tester genomic DNA hybridized to produce type c molecules. Tester specific DNA fragments formed hybrids with the complementary tester specific DNA (molecules b & f). Similarly, homologous driver specific DNA fragments annealed with complementary driver specific DNA fragments (molecule e). In addition, single stranded tester and driver molecules were also produced (molecules a & d). Two sets of suppression PCR reactions were carried out to amplify and select tester specific sequences. During primary PCR, the adaptors (1 and 2R) at the 5' end of the dsDNA were duplicated onto the 3' end of the opposite strand. Primer 1 was used to exponentially amplify dsDNA fragments with both adaptors, 1 and 2R, at either end (molecule f). The dsDNA with adaptor attached at one end only (type c molecules) underwent linear amplification. The dsDNA molecules with only one type of adaptor (molecules b) at both ends were not amplified due to binding of complementary adaptor sequence. The driver specific sequences were not amplified as there was no adaptor attached to them (molecules a, d & e). Secondary PCR was performed using nested primers 1 and 2. These primers were specific to the adaptors 1 and 2R sequences and therefore exponentially amplified the type f molecules. The secondary PCR product was rich with the tester specific sequences.

5.2.2 Cloning of PSSH inserts

The secondary PCR amplicons were ligated into the pCR[®] 2.1 TOPO vector using TOPO TA Cloning[®] kit (Invitrogen, UK). The pCR[®] 2.1 TOPO vector with the insert was cloned in 10F' *E. coli* chemically competent cells as recommended by the manufacturer. The successfully transformed bacteria were selected by using antibiotic(s) and blue white screening. The procedures for cloning and transformation of 10F' *E. coli* cells (Table 2.1) are briefly discussed in section 2.7.4. The plasmid DNA was extracted

from each clone containing the insert by using QIAprep spin miniprep kit (Qiagen, Crawley, UK) (2.6.2). The plasmid DNA was sequenced by single read sequencing using M13 reverse primers (Eurofins, MWG, UK). The similarity of the sequences to the known bacterial genome sequences was performed by using BLASTn and BLASTx searches at the National Centre of Biotechnology Information (NCBI) website. PCR screening was carried out by standard PCR protocol (section 2.7.1) to determine the distribution of these hyperinvasive specific sequences in all six hyperinvasive *C. jejuni* strains. An additional population of 9 low invasive *C. jejuni* strains was also analysed by PCR for the presence or absence of these hyperinvasive *C. jejuni* specific sequences. Details of all primers used in the PCR screen are listed in Table 2.2.2.

5.3 RESULTS AND DISCUSSION

5.3.1 Preparation of the driver and tester genomic DNA fragments

Genomic DNA of all 6 hyperinvasive *C. jejuni* strains as a pooled sample “tester” and 4 low invasive *C. jejuni* strains mixed together as “driver” was partially digested with *AluI* and *DraI* enzymes. This enzymatic digestion successfully generated small DNA fragments in the size range of 0.1-2 kbp (Figure 5.2).

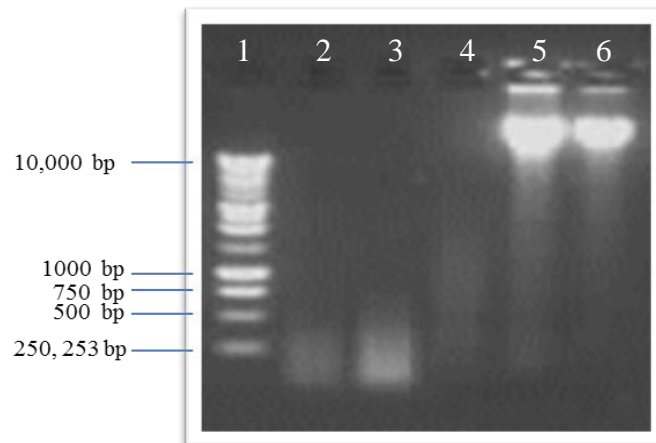


Figure 5.2: The enzymatically digested genomic DNA of the pool of hyperinvasive *C. jejuni* (tester), low invasive *C. jejuni* (driver) and *E. coli* (control) compared to the undigested genomic DNA of the hyperinvasive and low invasive *C. jejuni*.

Lane 1: 1 kbp DNA marker (Promega, UK). The *AluI* and *DraI* digested genomic DNA of “tester” *C. jejuni* (lane 2) and “driver” *C. jejuni* (lane 3). The control *E. coli* genomic DNA was digested with *RsaI* (lane 4). The undigested genomic DNA of “tester” *C. jejuni* (lane 5) and “driver” *C. jejuni* (lane 6) was run as a control to determine the success of enzymatic digestion. The PCR products are analysed on a 2% (w/v) agarose gel.

5.3.2 Ligation of adaptors 1 and 2R to the fragmented tester genomic DNA

After the enzymatic digestion of the genomic DNA, the relevant adaptor (1 or 2R) was then ligated to blunt tester fragments via a blunt ended reaction (section 5.2.1) which occurs between two ds DNA fragments. Therefore, because the 5' ends of the adaptor are unphosphorylated, the adaptor can only ligate via its 3' end to the 5' end of the tester DNA fragment and so only one strand of the adaptor anneals to generate the tester fragments below;



Figure 5.3: The diagram of the adaptor (1 or 2R) ligation to the tester genomic DNA fragment and the primers annealing in PCR reaction for adaptor efficiency testing.

The efficiency of ligation of adaptors to the genomic DNA was tested by PCR amplification (Figure 5.4).

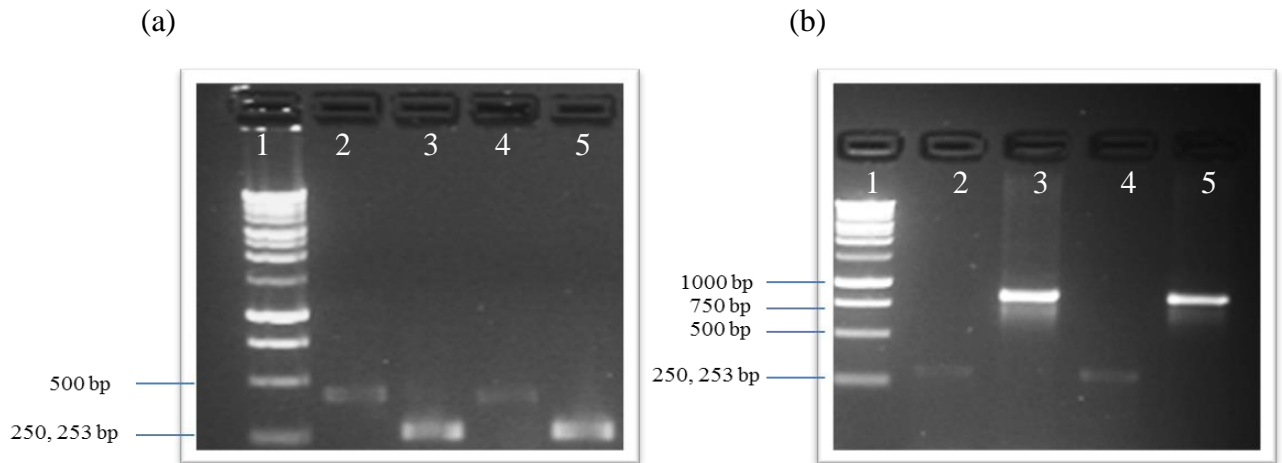


Figure 5.4 (a): The ligation efficiency testing of *E. coli* (control) and (b) the tester hyperinvasive *C. jejuni*.

(a) Lanes 2 and 4 contain the control *E. coli* (tester) genomic DNA PCR products amplified by using 23S RNA forward and reverse primers. The partially digested genomic DNA of the control *E. coli* (tester) ligated to adaptor 1 (lane 3) and 2R (lane 5) was amplified by using PCR 1 and 23S RNA forward primers. (b) Adaptor 1 and 2R ligated to the partially digested *C. jejuni* “tester” genomic DNA was amplified using PCR 1 and 16S rRNA forward primers (lanes 2 and 4 respectively). The tester genomic DNA of *C. jejuni* was amplified by using 16S rRNA forward and reverse primers (lanes 3 and 5 respectively). Lane 1: 1 kbp DNA marker (Promega, UK). The PCR products were analysed on a 1% (w/v) agarose gel.

PCR amplification using PCR primer 1 specific to the sequence of adaptor 1 and 2R, and 16S rRNA forward primer for the tester *C. jejuni* as a template generated an ~ 290 bp PCR product. A similar size PCR product was produced when amplification was performed for the adaptor ligated control *E. coli* (tester) using PCR 1 and 23S RNA forward primers. This suggests that the enzymatic digestion (5.3.1) produced the DNA fragments of the expected size range of 0.1-2 kbp however, a larger proportion of smaller sized DNA fragment were generated compared to the larger fragments. An expected band of 852 bp was produced by using 16S rRNA forward and reverse primers

for the undigested *C. jejuni* tester genomic DNA and 374 bp for the control *E. coli* tester genomic DNA with 23S RNA forward and reverse primers.

The brightness of the adaptor ligated DNA fragment band was very similar to the 16S rRNA product band (Figure 5.4b). This suggests that a high proportion of the adaptor ligated DNA fragments was generated that would ensure efficient hybridization.

5.3.3 Subtractive Hybridization I and II

The suppressive subtractive hybridization procedure was followed as previously described in detail (Ahmed *et al.*, 2002). Briefly, 1 and 2R adaptor ligated tester DNA aliquots and the driver DNA (without adaptors) were heat denatured. Two sets of hybridization reactions were carried out at 63 °C. During the first hybridization step, each aliquot of the adaptor ligated tester DNA was mixed separately with an excess of the freshly denatured driver DNA. This step allowed homologous tester and driver DNA fragments to form hybrids. During the second hybridization the 1 and 2R adaptor ligated aliquots were mixed together with an addition of fresh driver DNA. This allowed annealing of any leftover homologous tester and driver DNA fragments. In this step, adaptor 1 ligated tester specific DNA fragments formed homologues with the adaptor 2R ligated tester specific DNA fragments. During hybridization, a range of other molecules were also produced that are shown in figure 5.1.

5.3.4 Two second sets of subtractive hybridization steps for the selection of tester specific C. jejuni sequences

The subtractive hybridization was repeated twice at a lower temperature of 61 °C to enhance the efficiency of subtraction as recommended by Ahmed *et al* (2002). This ensured selection of sequences unique to the hyperinvasive *C. jejuni*. During this step, 10 µl of the secondary PCR product was used as a tester which was heat denatured and mixed with an excess of freshly denatured driver DNA. Two hybridizations were performed as before followed by primary and secondary PCRs. The products of primary and secondary PCR were checked by gel electrophoresis. The primary PCR product showed a smear whereas the secondary PCR product had more distinct bands (Figure 5.5). This showed that secondary PCR product was enriched with tester specific sequences containing products of different sizes.

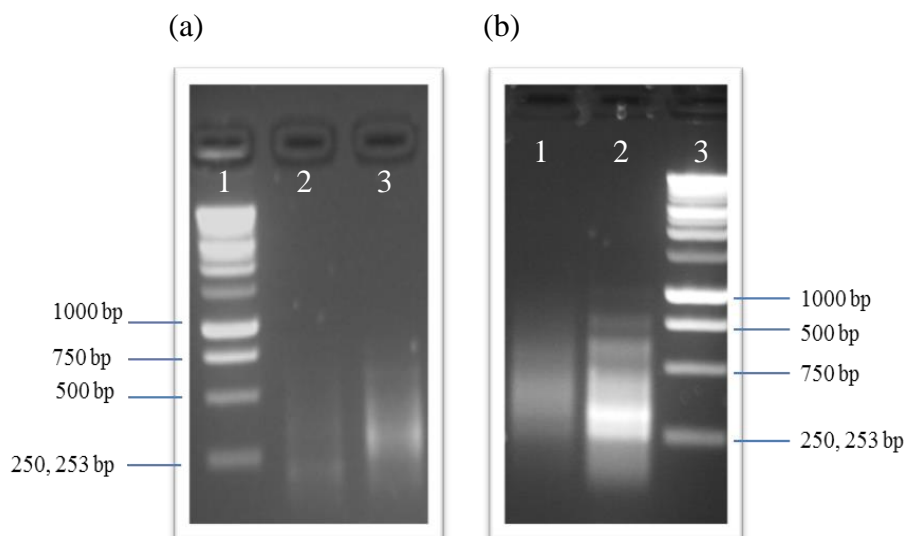


Figure 5.5: The primary and secondary PCR products (a) Control *E. coli* and (b) *C. jejuni*.

(a) Lane 1: 1 kbp DNA marker (Promega, UK), lane 2: primary PCR product of subtracted sample, lane 3: secondary PCR product of subtracted sample. (b) Lane 1: primary and lane 2 secondary PCR product of subtracted sample, lane 3: 1 kbp DNA marker (Promega, UK). The PCR products are analysed on a 2% (w/v) agarose gel.

5.3.5 Analysis of successful subtraction of the secondary PCR product

To further show that the secondary PCR product is enriched with tester *C. jejuni* sequences a validation PCR was performed comparing the loss of a housekeeping gene in the subtracted sample and conservation of this gene in the unsubtracted sample. The PCR analysis was performed with the subtracted control *E. coli* sample after subtractive hybridizations performed at 63 °C compared with the unsubtracted sample using the 23S RNA forward and reverse primers (Figure 5.6 a and b). The PCR screening of the subtracted test *C. jejuni* sample after two sets of subtractive hybridization at 61 °C was carried out using 16S rRNA primers which was compared against the unsubtracted sample (Figure 5.6 c and d).

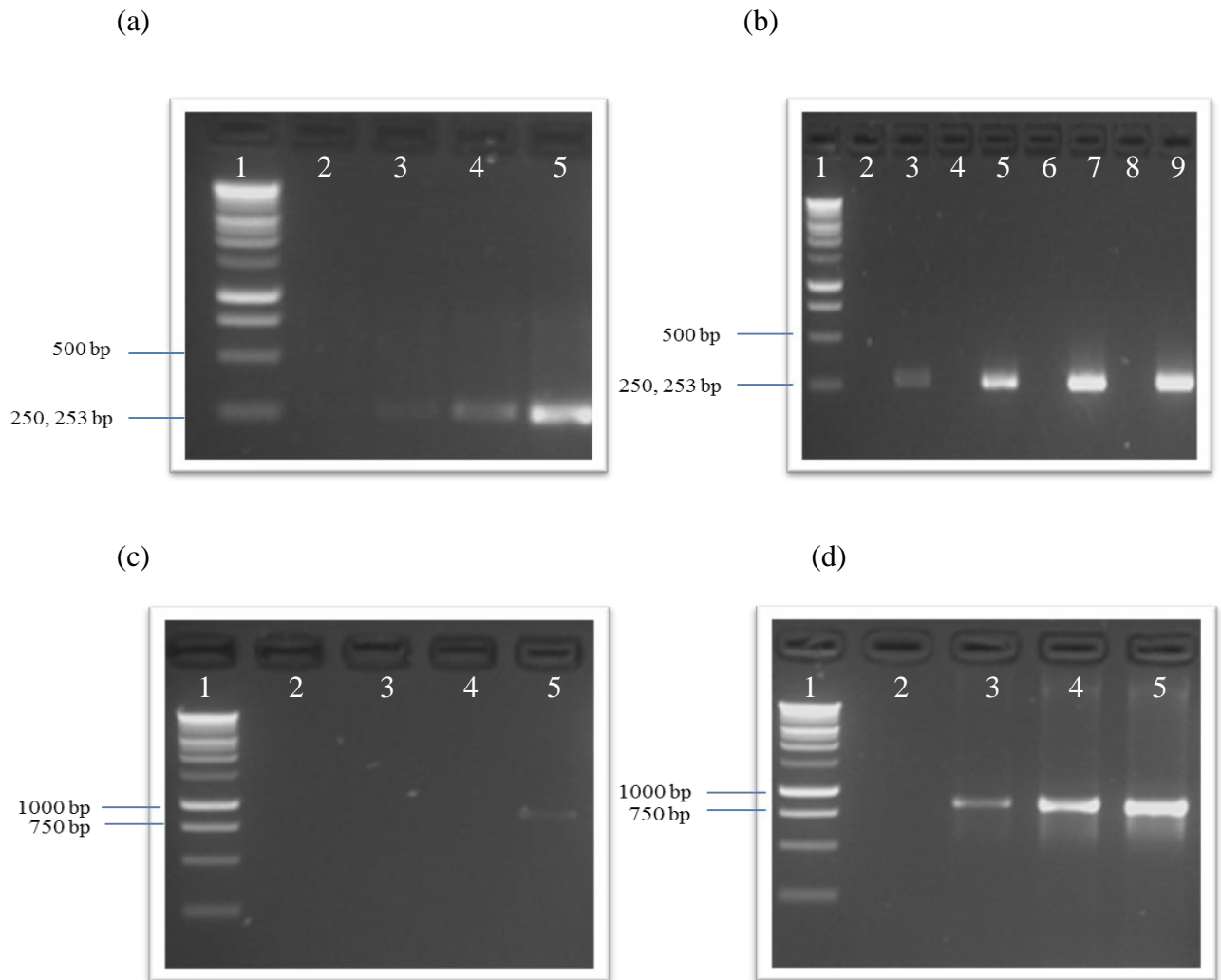


Figure 5.6: The subtraction efficiency of (a) subtracted control *E. coli* (b) unsubtracted control *E. coli* (c) subtracted test hyperinvasive *C. jejuni* and (d) unsubtracted test hyperinvasive *C. jejuni*.

Each gel was loaded: Lane 1; 1 kbp DNA marker (Promega, UK); (a) (lanes 2, 3, 4, 5); subtracted sample after 18, 21, 24, 27 PCR amplification cycles respectively; (b) (lanes 3, 5, 7 and 9); unsubtracted sample after 18, 21, 24, 27 cycles respectively; (lanes 2, 4, 6, 8); Blank; (c) (Lanes 2, 3, 4, 5); PCR products after 18, 21, 24 and 27 amplification cycles respectively (d) (Lanes 2, 3, 4, 5); PCR products after 18, 21, 24 and 27 amplification cycles respectively. The PCR products were analysed on a 2% (w/v) agarose gel.

For the subtracted test *C. jejuni* sample, there was no product seen after 18, 21 and 24 cycles for the subtracted sample whereas a faint band was seen after 27 cycles. For the unsubtracted *C. jejuni* sample on the other hand, no band was observed after 18 cycles

with a faint band observed after 21 cycles that got brighter after 24 and 27 cycles. This result showed that two cycles of subtractive hybridization at a lower annealing temperature proved to be a successful step when studying groups of strains as “tester” in a single subtractive hybridization experiment. This reduced the number of tester and driver homologous sequences thus improving the efficiency of subtractive hybridization.

5.3.6 Preparation of subtractive hybridization library

The secondary PCR products were ligated into pCR[®] 2.1 TOPO vector using TOPO TA Cloning[®] kit (Invitrogen, UK) and transformed into 10F' *E. coli* chemically competent cells. This technique produced 102 colonies of which 62 colonies had inserts. The size of inserts was determined by colony PCR using M13 forward and reverse primers followed by analysis on a 1% (w/v) agarose gel. An example gel picture is shown here (Figure 5.7).

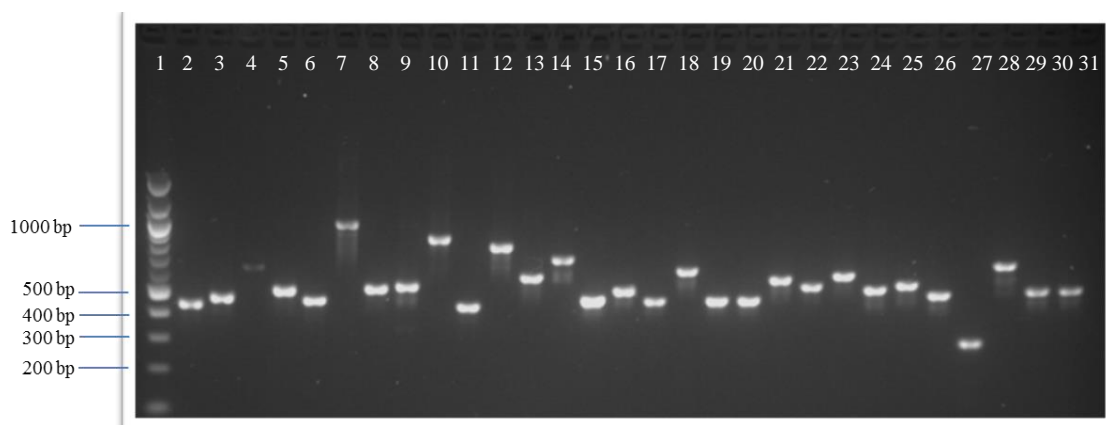


Figure 5.7: The colony PCR screening of clones to determine the size of sequences inserted into pCR[®] 2.1 TOPO vector.

The clones 1-29 (lanes 2-30) showed insert sizes in the range of 300-1000 bp. Lane 1: 1 kbp DNA marker (Promega, UK). Lane 31: negative control.

Each clone generated a PCR product more than 200 bp in size. Therefore, all inserts were sequenced.

5.3.7 Selection of the hyperinvasive *C. jejuni* specific sequences from subtractive hybridization library

Sequence similarity carried out using *Campylobacter* specific database (www.campydb) and BLAST searches at NCBI genome bank (www.blast.ncbi.nlm.nih.gov/Blast.cgi) identified 38 out of 62 inserts as duplicates. This showed a limitation of the subtractive hybridization technique where inserts that are in more abundance get amplified (Gerrish *et al.*, 2010). Only one representative of the duplicate insert was further analysed. PCR analysis of the remaining 24 inserts identified 11/24 inserts as specific to one or more hyperinvasive *C. jejuni* strains and absent from all low invasive *C. jejuni* strains used as driver. The rest of the 13 inserts were false positives (*i.e.* inserts present in one or more of the low invasive *C. jejuni* strains used as driver). PCR screening also showed the distribution of inserts in an additional 9 low invasive *C. jejuni* strains that was not used in PSSH (Figure 5.8). The PCR analysis results are summarized in table 5.1.

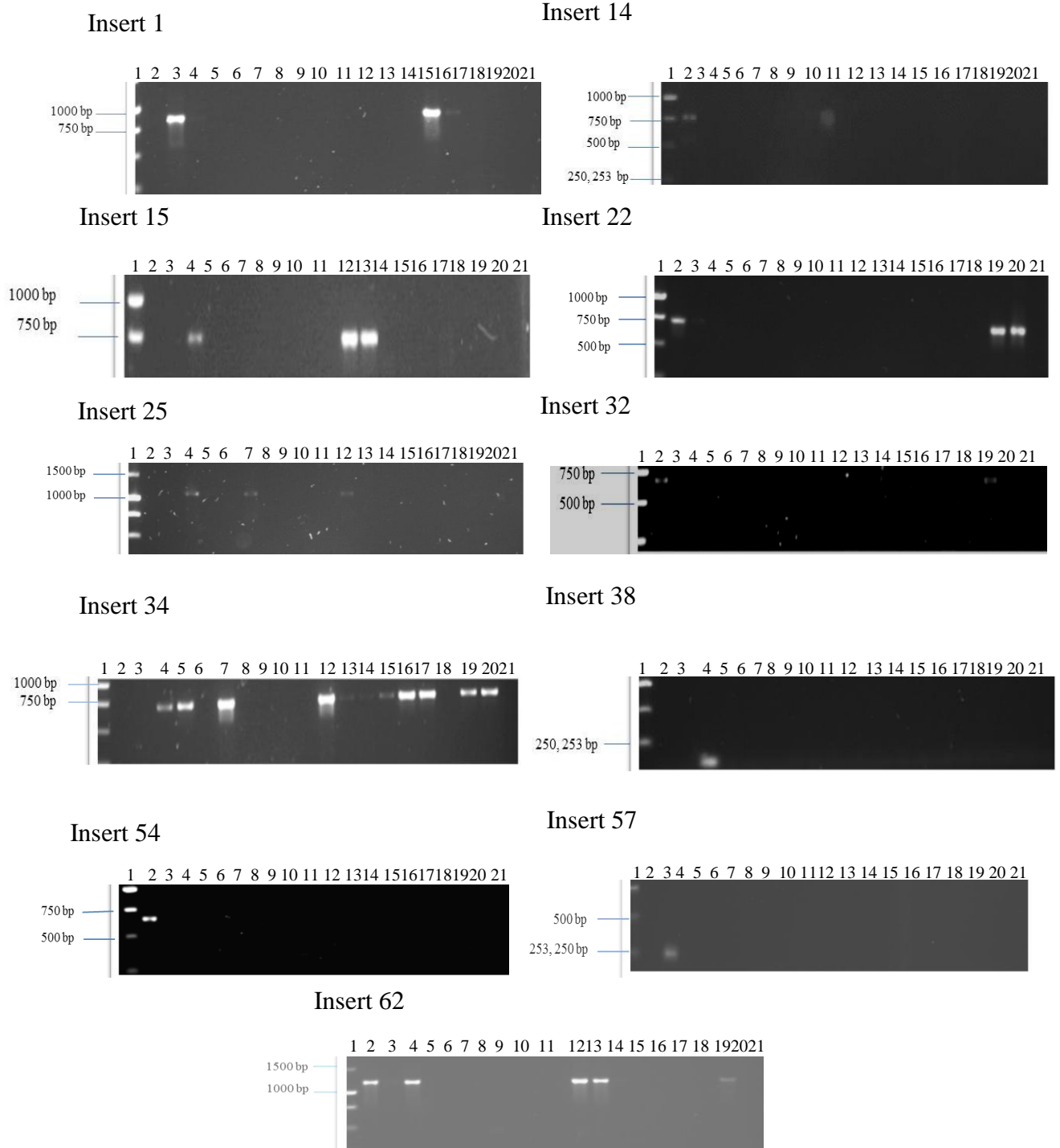


Figure 5.8: PCR screening on a 1% (w/v) agarose gel to show the distribution of 11 inserts in the hyperinvasive and low invasive *C. jejuni* strains.

Lane 1: 1 kbp DNA marker (Promega, UK). Lane 2-7; hyperinvasive *C. jejuni* strains 01/10, 01/35, 01/04, 01/41, 01/51, EX114. Lanes 8-20; low invasive *C. jejuni* strains 01/30, 01/32, 01/46, 01/39, 01/05, 01/08, 01/11, 01/36, C2/3, C12/11, C27/14, C69/2, C110/4. Lane 21: negative control. The primers used in PCR screening and the expected product band size are listed in table 2.2.2.

Table 5.1: The distribution of inserts identified by PSSH in the hyperinvasive and low invasive *C. jejuni* strains by PCR analysis.

Insert	Distribution of PSSH inserts in the hyperinvasive and low-invasive <i>C. jejuni</i> by PCR																Total-3	p-value**						
	Hyperinvasive <i>C. jejuni</i>								Low invasive <i>C. jejuni</i>															
	01_10	01_35	01_04	01_41	01_51	EX114	Total-2		01_30*	01_32*	01_46*	01_39*	01_05	01_08	01_11	01_36			C2_3	C12_11	C27_14	C69_2	C110_4	
1	-	+	-	-	-	-	1	-	-	-	-	-	-	-	-	+	(+)	-	-	-	-	-	2	1.000
14	+	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0	0.316
15	-	-	+	-	-	-	1	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	2	1.000
22	+	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	2	1.000
25	-	-	+	-	-	+	2	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	1	0.222
32	+	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-	1	0.544
34	-	-	+	-	-	+	3	-	-	-	-	-	+	(+)	(+)	(+)	+	+	-	-	+	+	8	0.506
38	-	-	+	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0	0.316
54	+	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0	0.316
57	-	+	-	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0	0.316
62	+	-	+	-	-	-	2	-	-	-	-	+	+	-	-	-	-	-	-	(+)	(+)	-	3	0.520
Total-1	5	2	5	1	0	2		0				4	3	1	2	2	2	1	0	4	2			

+ = present, - = absent; (+) = partially present (a faint band seen on the gel).

* = The low invasive *C. jejuni* strains used as “driver” in PSSH experiment.

Total-1 = Total number of inserts present in each hyperinvasive/low invasive *C. jejuni* strain. Total-2: Total number of hyperinvasive *C. jejuni* strains positive for each insert. Total-3: Total number of low invasive *C. jejuni* strains positive for each insert.

** = Chi-square, Fisher’s exact test was performed to determine if the genes detected in the hyperinvasive group of *C. jejuni* strains was statistically significant. Significance level = 1% ($p=0.01$).

The presence of inserts in the hyperinvasive *C. jejuni* strains and their absence from the 13 low invasive *C. jejuni* strains was not statistically significant (Table 5.1). There was no sequence present in all hyperinvasive *C. jejuni*. Eight inserts were specific to only one hyperinvasive *C. jejuni* strain (inserts; 1, 4, 15, 22, 32, 38, 54, 57). Inserts 25 and 62 were present in two hyperinvasive *C. jejuni* whereas insert 34 was found in 3/6 hyperinvasive *C. jejuni* strains. All the inserts were absent from the low invasive *C. jejuni* strains used in the PSSH. This suggests that the pooled suppressive subtractive hybridization technique can successfully detect the sequences specific to strains studied together as a group rather than one to one strain comparison as in a DNA microarray and single SSH experiments. In addition, PSSH can even detect sequences specific to a single strain in a tester pool of several strains. These sequences might be small in size or present in small quantities. Thus, PSSH is sensitive, cost effective and time efficient modification of subtractive hybridization (Gerrish *et al.*, 2010).

To complement the CGH data; the prevalence of these 11 inserts was screened by PCR in an additional 9 low invasive *C. jejuni* strains which were not a part of PSSH experiment (Table 5.1). The PCR screening showed that 4 inserts (14, 38, 54, and 57) were absent from all additional low-invasive strains tested, hence specific to the hyperinvasive *C. jejuni*. Six inserts were present in up to three of the low invasive *C. jejuni* strains. Insert 34 was most widely prevalent in the additionally screened low invasive *C. jejuni* strains as it was present in 8/9 of the strains. After the identification of tester specific inserts by PSSH, a standard PCR reaction or Southern hybridization (Ahmed *et al.*, 2002) can be used to determine the distribution of inserts in a larger population of strains. This prevents the need of repeating individual subtractions thus reducing chances of error due to multiple sets of experiments and more confidence of dataset.

Interestingly, 5/11 inserts were present in the hyperinvasive *C. jejuni* 01/04 and *C. jejuni* 01/10 with no insert detected in *C. jejuni* 01/51. The PCR profile of the non-PSSH tested low invasive *C. jejuni* population showed that *C. jejuni* 01/05 and C69/2 were positive for greatest number of inserts (4/11) and *C. jejuni* C27/14 did not show presence of any of the 11 inserts (Table 5.1).

5.3.8 Homology of the hyperinvasive *C. jejuni* specific sequences

The predicted amino acid similarity based on BLAST searches was between 89-100 % for all the inserts (Table 5.2).

Table 5.2: The inserts identified by PSSH as specifically present in hyperinvasive *C. jejuni* strains “tester” and absent from low invasive *C. jejuni* “driver”.

Insert/Clone	Size (bp)	Predicted protein similarity	Closest match	Query coverage (%)	Predicted amino acids similarity (%)
1	154	8-amino-7-oxononanoate	<i>bioF-2 (C. coli RM2228)</i>	72	98
14	191	Prophage MuSo1, F protein, putative	<i>C. jejuni</i> subsp. <i>jejuni</i> 260.94/ICDCCJ07001_672 (<i>C. jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001)	76	100
15	238	GDP mannose 4,6 dehydratase	<i>C. jejuni</i> subsp. <i>jejuni</i> 260.94/ICDCCJ07001_1357 (<i>C. jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001)	86	93
22	312	FkbM family methyltransferase	JJD26997_1251 <i>C. jejuni</i> subsp. <i>doylei</i> 269.97	86	96
25	214	Hypothetical protein	C8J_1243(<i>C. jejuni</i> subsp. <i>jejuni</i> 81116)	82	95
32	756	Restriction modification system DNA specificity domain containing protein	<i>Helicobacter canadensis</i> MIT 98-5491	81	89
34	213	Thiazole synthase	<i>thiG (C. jejuni</i> subsp. <i>doylei</i> 269.97)	80	100
38	229	No similarity found
54	791	Putative tetracycline resistance protein	<i>tet (O/W/32/O) Streptococcus gallolyticus</i> subsp. <i>gallolyticus</i> ATCC BAA-2069 plasmid pSGG1	56	97
57	282	Phage tail fibre protein	BAV0033 (<i>Bordetella avium</i> 197N)	63	35
62	235	Putative sugar transferase	(<i>C. jejuni</i> subsp. <i>jejuni</i> CG8486)	84	97

Predicted protein similarity and closest match show the function/role of the best match for the insert in publically available microbial genome sequences.

Query coverage percentage is the length of each insert used by BLASTx search to find similarity in the sequence database.

The percentage predicted amino acid similarity between the query sequence (insert) and its best protein match in the genome sequence database.

Out of 11 inserts, 4 inserts (14, 38, 54 and 57) were only present in the hyperinvasive *C. jejuni* and absent from the low invasive *C. jejuni* strains. Seven inserts had sequence similarities within the *Campylobacter* genus. Four of 11 inserts (14, 15, 25, and 62) were similar to other *C. jejuni* subsp. *jejuni* strains and 2 inserts (22, 34) were similar to

the *C. jejuni* subsp. *doylei* whilst one insert (insert 1) was similar in the *C. coli* genome. Three inserts (32, 54 and 57) were similar to other bacteria. One insert (insert 38) had no sequence similarity to the available sequences in the genome databases searched.

5.3.9 Genotyping based on PSSH reveals hyperinvasive *C. jejuni* specific genes

Using a pool of six hyperinvasive *C. jejuni* as tester and a group of four low invasive *C. jejuni* a suppressive hybridization study was performed. A variety of genes were identified in this study as specifically present in one or more of the six hyperinvasive *C. jejuni* in the pooled SSH study compared to the low invasive *C. jejuni*. The majority of inserts have homologues found in the *Campylobacter* genus. This may be because the number of *Campylobacter* strains that have been sequenced has rapidly increased providing more information about the genomic structure and diversity exhibited by this pathogen (Fouts *et al.*, 2005).

The genes identified in the PSSH study will be discussed below.

Insert 1

Genomic subtractive hybridization identified insert 1 to have similarity to a gene, *bioF-2* encoding for 8-amino-7-oxononanoate synthase in *C. coli* RM2228 (Table 5.2).

PCR screening showed that insert 1 was present in only one hyperinvasive *C. jejuni* 01/35 and two non-PSSH low invasive *C. jejuni* (*i.e.* 01/36, C2/3) (summary Table 5.1). The PCR product was seen as a bright band in the low invasive *C. jejuni* 01/36 after 30 amplification cycles whereas only a faint band was observed for *C. jejuni* C2/3 after the same number of cycles (Figure 5.8). This might suggest that *bioF-2* may have a variable sequence in this strain. By contrast, our CGH study (chapter 4) showed that *bioF-2* gene was present in 4/6 hyperinvasive (*C. jejuni* 01/35, 01/04, 01/41 and EX114) whilst slightly divergent in the hyperinvasive *C. jejuni* 01/10 and highly divergent/absent in the hyperinvasive *C. jejuni* 01/51. CGH data also showed that *bioF-2* was present in the low invasive *C. jejuni* 01/30 and 01/46 but highly divergent in the low invasive *C. jejuni* 01/32 and 01/39. Unlike for PSSH, where the distribution of each insert in *C. jejuni* strains was determined by PCR the CGH result for this gene was not validated by PCR. BioF-2 is an enzyme involved in biotin biosynthesis. It catalyses the decarboxylation of amino acids by condensation between amino acid and acetyl-coA producing 8-amino-7-

oxononanoate as an important substrate in biotin production (Alexeev *et al.*, 1998). BioF-2 is an important metabolic enzyme in *C. jejuni* (Fouts *et al.*, 2005). The detection of metabolic gene as a genetic marker by the genomic subtractive hybridization in a hyperinvasive *C. jejuni* may suggest the role of metabolic pathways in the hyperinvasive phenotype. The metabolic core of *C. jejuni* is generally considered stable. The horizontal transfer of metabolism related genes showed that these genes are under selective pressure and confer advantage to pathogenic bacteria in disease process inside the host cells. The similarity of insert 1 to the gene *bioF-2* in *C. jejuni* sister species, *C. coli* validates recent research findings that *C. jejuni* and *C. coli* are more closely related species to each other than to other member of the *Campylobacter* genus (Fouts *et al.*, 2005). Still there is enough genomic variation in two species allowing the transfer of genomic material between them. There is also emerging scientific evidence of convergence of *C. jejuni* and *C. coli* genomes resulting in the evolution of conserved core genomes which is attributed to several epidemiological and ecological factors including human activity (Sheppard *et al.*, 2008) which further supports our findings.

Insert 14

Insert 14, showed similarity to the prophage MuSo1, F protein, with putative similarity in *C. jejuni subsp. jejuni* ICDCCJ07001 and 260.94 (Table 5.2). This insert was present in the hyperinvasive *C. jejuni* 01/10 and absent from the PSSH and PCR screened low invasive *C. jejuni* strains (Figure 5.8 and Table 5.1).

Bacteriophage are the vehicle of horizontal gene transfer that contribute to diversity in the bacterial genome and may carry genes that contribute to the virulence in bacterial pathogens. Examples include emergence of bacterial pathogens including *E. coli* O157 and *V. cholerae* (Canchaya *et al.*, 2004). *E. coli* O157, an important human pathogen contains two lambdoid prophages that encode for Shiga-like toxins (Stx) which is a major virulence factor for this strain (Wagner *et al.*, 2002). There is an evidence of presence of prophages and insertion elements (IS) in *Campylobacter* species (Fouts *et al.*, 2005). A *Campylobacter* Mu-like phage (CMLP1) in *C. jejuni* RM1221 genome is located upstream of *argC* locus and encodes for proteins with resemblance to the bacteriophage Mu and other Mu-like prophage proteins. CMLP1 in *C. jejuni* does not contain any virulence determinants but may contribute towards pathogenicity by changing the function of other virulence related genes. Other mobile genetic elements in

C. jejuni RM1221 include *Campylobacter* Integrated Elements (CJIEs 2, 3 and 4) scattered along the genome. These integrated elements are absent from the first genome sequenced *C. jejuni* NCTC11168. *C. jejuni* RM1221 like genetic elements are conserved in several other *C. jejuni* and *C. coli* strains but show a high level of diversity (Parker *et al.*, 2006, Clark and Ng., 2008). The prophages showed high levels of recombination resulting in mosaic pattern of distribution of these prophages in *C. jejuni* strains. The detailed sequence analyses of CMLP 1 from different *C. jejuni* strains showed homology to the bacteriophage characteristic of enteric pathogens that are known to be responsible for a number of virulence phenotypes including host specificity (Clark and Ng., 2008). *C. jejuni* ICDCJ07001 is a GBS associated strain isolated from a GBS outbreak in North China in 2007. This strain contains *C. jejuni* RM1221 characteristic CMLP1 but in the reverse orientation. The CMLP1 in *C. jejuni* ICDCJ07001 is inducible but is not known to contain any virulence related genes (Zhang *et al.*, 2010). To date, there is no biological evidence to prove the role of these prophage genes in virulence of the hyperinvasive *C. jejuni* strains.

Insert 57

A phage tail fibre protein I (insert 57) in *Bordetella avium* 197N (Table 5.2) has been identified only in the hyperinvasive *C. jejuni* 01/35 and absent from all low invasive *C. jejuni* strains (Table 5.1 and Figure 5.8). *Bordetella avium* is a pathogenic bacteria of wild and domesticated birds mainly commercially raised turkeys. It is the causative agent of bordetellosis, a respiratory illness in avian species (Sebahia *et al.*, 2006). *B. avium* 197N is a well-studied sequenced strain of this avian pathogen. *B. avium* 197N contains three prophages namely prophage A (BAV0391-BAV0433), prophage B (BAV1280-BAV1342) and the third prophage (BAV1423-BAV1482). Prophage A has genes encoding for a well-known phenomenon in *Bordetella* species known as switching tropism (Sebahia *et al.*, 2006). Tropism switching activity determines the specificity of bacterial receptors to the host surface. However, in *B. avium* 197N most of the internal region responsible for tropism (BAV0416-BAV0430) is missing and the transcriptase responsible for tropism switch over is also absent (Sebahia *et al.*, 2006). The insert found in the hyperinvasive *C. jejuni* 01/35 with similarity to the phage tail fibre protein, (BAV0033) in *B. avium* 197N is a part of prophage A. Since, poultry is the common host for *B. avium* and *C. jejuni*, the transfer of genes encoded on mobile genetic elements is expected which may contribute to the diversity in genome.

Insert 15 and 62

A sequence (insert 15) identified by PSSH was only found in the hyperinvasive *C. jejuni* 01/04. In the non-PSSH low invasive *C. jejuni* strains analysed by PCR, insert 15 was detected in *C. jejuni* 01/05 and 01/08 (Figure 5.8 and Table 5.1). This insert showed similarity to GDP mannose 4, 6 dehydratase in *C. jejuni* 260.94 and ICDCJ07001 (Table 5.2). The gene ICDCJ07001_1357 is the part of *C. jejuni* ICDCJ07001 capsule locus.

The product of gene *dmhA* (GDP mannose 4, 6 dehydratase) is an enzyme responsible for the conversion of heptose to deoxyheptose. It is located in the variable region of CPS loci of many *C. jejuni* strains (Karlyshev *et al.*, 2005a, Poly *et al.*, 2011). Karlyshev *et al* (2005a) found *dmhA* in the capsule locus of many *C. jejuni* strains, fully functional in some strains but variable in other *C. jejuni* strains. *C. jejuni* ICDCJ07001 is a GBS associated clinical isolate. It is very similar to another sequenced GBS causing *C. jejuni* strain 260.94. Both strains have the Penner serotype (HS41) and share sequence similarity in their capsule region (Zhang *et al.*, 2010).

Another insert 62, showed similarity to a putative sugar transferase in *C. jejuni* 8486 (Table 5.2). PCR analysis showed that this insert was present in two hyperinvasive *C. jejuni* 01/10 and 01/04. Among the additional low invasive *C. jejuni* strains screened by PCR, this insert was present in *C. jejuni* 01/05 and 01/08. A faint band was also seen in low invasive *C. jejuni* C69/2 (Figure 5.8 and Table 5.1). Putative sugar transferase is a sugar biosynthesis gene present in the *C. jejuni* CPS region (Poly *et al.*, 2011). *C. jejuni* CG8486 is a human clinical isolate from a soldier presenting symptoms of bloody diarrhoea. The CAP locus in *C. jejuni* GC8486 is a 26kbp region and belongs to the Penner serotype (HS4). The CAP locus in *C. jejuni* CG8486 is similar in size to the CAP region in *C. jejuni* 81-176 but smaller than in *C. jejuni* NCTC11168. The *C. jejuni* CG8486 CAP locus encodes for sugar transferases already known in the capsule locus of *C. jejuni* strains (Poly *et al.*, 2007b).

The capsule locus in *C. jejuni* is composed of a highly variable central region enclosed by the conserved *kps* genes on either side. Due to the variability in middle capsular region it is responsible for the generation of diverse polysaccharide structures in different *C. jejuni* strains. These diverse capsule structures assigns strains to different

Penner serotypes (Dorrell *et al.*, 2001, Pearson *et al.*, 2003, Karlyshev *et al.*, 2005a and b, Poly *et al.*, 2011). The strains within the same Penner serotype complexes are found to have similar CPS region (Karlyshev *et al.*, 2005a and b, Poly *et al.*, 2011). In contrast to the reports of above authors the Penner serotype of the hyperinvasive *C. jejuni* 01/10 is HS50 whereas *C. jejuni* 01/04 is not known (data discussed in chapter 4). The capsule biosynthesis genes in the hyperinvasive *C. jejuni* strains identified here with the best match to other *C. jejuni* CPS genes may suggest a mosaic pattern of distribution of the variable capsular genes in *C. jejuni* (Poly *et al.*, 2011). As the CPS regions are known to be variable amongst strains which is likely the reason that by using SSH approach these kinds of strain-specific genes are detected. The surface capsular structures in *C. jejuni* are known to contribute to pathogenesis mainly, adhesion and invasion of host cells and serum resistance (Bacon *et al.*, 2001, Guerry *et al.*, 2002). Hence, the role of capsule in the hyperinvasive phenotype of *C. jejuni* strains studied here cannot be neglected.

Inserts 22 and 34

Inserts 22 and insert 34 showed maximum similarity to the FkbM family methyltransferase and thiazole synthase (*thiG*) in *C. jejuni* subsp. *doylei* respectively (Table 5.2). Insert 22 was present in the hyperinvasive *C. jejuni* 01/10. The low invasive *C. jejuni* C69/2 and C110/4 were also positive for insert 22. Insert 34, was found to be present in the hyperinvasive *C. jejuni* 01/04, 01/41 and EX114 (Table 5.1). Among the additional low invasive *C. jejuni* strains insert 34 was present in *C. jejuni* 01/05, C2/3, C12/11, C69/2, C110/4 seen as bright bands on the agarose gel (Figure 5.8). There were faint bands detected in the low invasive *C. jejuni* 01/08, 01/11, 01/36 that may represent polymorphic gene in these strains.

The similarity of inserts 22 and 34 in *C. jejuni* subsp. *doylei* suggests that the two species (*i.e.* *C. jejuni* subsp. *jejuni* and *C. jejuni* subsp. *doylei*) frequently exchange genetic material. This is in contrast to the findings by Parker *et al* (2007) who reported these two subspecies of *C. jejuni* as highly divergent from each other using MLST and a DNA microarray based comparative genomics based indexing (CGI).

DNA methyltransferases are diversely present in bacteria and usually form a part of R-M systems in bacteria (Dale and Park., 2004). They are responsible for the methylation of N⁶ position in adenine and N⁴ and C⁵ position in cytosine in bacteria (Wion and

Casadesús., 2006. Like the variable CPS genes, DNA methyltransferases encoding genes are likely to be identified in SSH screening as they are variably distributed among the *C. jejuni* strains. Recently, the role of a DNA methyltransferase encoded by the gene Cj1461 in *C. jejuni* virulence has been demonstrated. This DNA methyltransferase influenced adherence, invasion and motility in *C. jejuni* 81-176 (Kim *et al.*, 2008). Having known the role of DNA methyltransferase in virulence it would be interesting to study the role of FkbM family methyltransferase in invasion and other virulence traits in the hyperinvasive *C. jejuni* 01/10.

Thiazole synthase is an important enzyme of thiazole biosynthesis. In *E. coli* and other anaerobic bacteria the components of this enzyme; ThiH, ThiG, ThiS and ThiF are essential for thiamine biosynthesis process. During this process an intermediate product, dehydroglycine is produced which is taken up by ThiG to be used for thiazole cyclization process (Kriek *et al.*, 2007).

Insert 32

Insert 32 was similar to the Restriction modification system DNA specificity domain-containing protein in *H. canadensis* MIT 98-5491 (Table 5.2). PCR screening showed the presence of this insert in the hyperinvasive *C. jejuni* 01/10. Among the low invasive *C. jejuni* strains not a part of PSSH experiment, insert 32 was only present in the low invasive *C. jejuni* C69/2 (Figure 5.8 and Table 5.1). The restriction modification systems in bacteria are defence systems against the foreign DNA especially introduced by the lytic and lysogenic bacteriophage. Different types of RM systems are present in bacteria called type I, II, III and IV. The type I RM system in a number of *C. jejuni* strains has been studied in detail (Miller *et al.*, 2005). There are few type II and III RM systems in sequenced genomes of *C. jejuni* NCTC11168, *C. jejuni* RM1221, *C. coli* RM2228 and *C. lari* RM2100 (Fouts *et al.*, 2005). *H. hepaticus* ATCC 51449 is like other *Campylobacter* species strains in RM systems with only few RM system genes. In contrast, *C. upsaliensis* RM3195 genome has DNA sequences encoding for adenosine and cytosine DNA-methyltransferases in addition to a putative type II and III RM systems (Fouts *et al.*, 2005) hence, similar to *H. pylori* that has three to four type I R-M systems (Miller *et al.*, 2005). The restriction modification systems have shown extensive diversity based on the origin of strains (Parker *et al.*, 2007, Ahmed *et al.*, 2002, Dorrell *et al.*, 2001). A putative RM DNA sequence in the hyperinvasive *C. jejuni*

from *Helicobacter* species may contribute to enhanced resistance against the foreign DNA making this strain more stable inside the host and in outside environment.

5.3.9.7 Insert 54

Insert 54, showed match with a putative tetracycline resistance gene *tet* (O/W/32/O) in *Streptococcus gallolyticus* subsp. *gallolyticus* ATCC BAA-2069 plasmid pSGG1 (Table 5.2). This insert was only present in the hyperinvasive *C. jejuni* 01/10 (Figure 5.8 and Table 5.1). Antibiotic resistance among *C. jejuni* strains is highly prevalent and is a major threat to human health (Pratt and Korolik., 2005). There are a number of tetracycline resistance genes that are either chromosomally encoded or located on a plasmid (Roberts., 2005). In *Campylobacter* species *tetO* gene is mainly responsible for tetracycline resistance (Pratt and Korolik., 2005). This gene encodes for a Tet(O) protein that protects the ribosome from the damaging effects of tetracycline (Roberts., 2005). Natural transformation allows transfer of antibiotics resistance between *Campylobacter* species in a mixed population (Jeon *et al.*, 2008). A self-transmissible plasmid pIP1433 in *C. coli* BM2509 carries *tetO* gene and transfer of this gene is evident in the streptococcus and enterococcus species causing high levels of tetracycline resistance in these organisms (Zilhao *et al.*, 1988). A recent study has shown the transfer of antibiotic resistance between *C. jejuni* and *H. pylori* strains by conjugation (Oyarzabal *et al.*, 2007). This suggests that *C. jejuni* is capable of uptaking antibiotics resistance genes from other organisms. The identification of insert in the hyperinvasive *C. jejuni* 01/10 with similarity to *tetO* from *S. gallolyticus* subsp. *gallolyticus* ATCC BAA-2069 plasmid pSGG1 may suggest it to be a stable strain with added tetracycline resistance. The increased antibiotic resistance would allow improved survival inside the human host. It would be interesting to know how antibiotic resistance would influence the invasion phenotype in this hyperinvasive *C. jejuni* strain.

Insert 25

Insert 25 was found in the hyperinvasive *C. jejuni* 01/04 and EX114. PCR analysis showed that this insert was present in the low invasive *C. jejuni* 01/05 (Figure 5.8 and Table 5.1). The similarity of insert 25 to the hypothetical gene in the *C. jejuni* 81116

(Table 5.2) highlights the fact that a large proportion of *C. jejuni* genome is functionally uncharacterized and these regions of unknown function add to the genome diversity.

Insert 38

One sequence detected in only *C. jejuni* 01/04 (Figure 5.8 and Table 5.1) has no similarity found in the genome databases searches (Table 5.2). The identification of such functionally uncharacterised sequences may suggest that the hyperinvasive *C. jejuni* strains have sufficient variation in their genome.

Such broad range of inserts recovered from PSSH with similarity to within and between species genomic regions and to other organisms suggests that hyperinvasive *C. jejuni* strains have a striking ability to diversify by accepting genomic materials from other sources to evolve as versatile human pathogens.

5.4 CONCLUSIONS AND NEXT STEP

The pooled suppressive subtracted hybridization study compared 6 hyperinvasive *C. jejuni* as a tester with a group of 4 low invasive *C. jejuni* strains as a driver. This technique identified 11 sequences specific to the hyperinvasive *C. jejuni* that were absent from the low invasive *C. jejuni* strains. There was no insert common in all the hyperinvasive *C. jejuni* strains. All inserts were present in one or more hyperinvasive *C. jejuni* strains and were variably distributed in the additional low invasive *C. jejuni* strains only screened by PCR. This suggests that the sequences identified may just represent the strain specific genes rather than the hyperinvasive *C. jejuni* specific genes. The detected sequences had a range of functions with inter and intra species homology and similarity with other bacteria. This proved that the hyperinvasive *C. jejuni* are a diverse group of strains. Both the DNA microarray and subtractive hybridization showed that the hyperinvasive *C. jejuni* 01/10 was the most diverse strain and *C. jejuni* 01/51 was least variable. These two hyperinvasive strains were selected for further analysis by the next generation genome sequencing.

Chapter Six

**WHOLE GENOME SEQUENCING
OF *C. JEJUNI* 01/10 AND 01/51**

WHOLE GENOME SEQUENCING OF *C. JEJUNI* 01/10 AND 01/51

6.1 INTRODUCTION

Genome sequencing has become a tool in providing detailed insight into the genomic diversity and evolution of bacteria. The genomic structure of two hyperinvasive *C. jejuni* strains (01/10 and 01/51) was studied in greater depth by using whole genome sequencing.

6.1.1 *Campylobacter* genome

The first *Campylobacter* genome to be sequenced was *C. jejuni* NCTC11168 was sequenced in 2000. The sequencing data revealed that *C. jejuni* 11168 has a 1.6 Mb genome containing 1,641,481 base pairs with 30.6% G+C content and represented an AT rich genome. The genes in the genome encoded for 1,654 proteins and 54 stable types of RNA (Parkhill *et al.*, 2000). Later, re-annotation and reanalysis of NCTC11168 genome sequencing data reduced the predicted protein sequences from 1,654 to 1,643. The functional categories were revised and new information for several coding sequences was added which was not reported before (Gundogdu *et al.*, 2007). The *C. jejuni* 11168 genome is unique as it does not contain large inserts or prophage sequences and very few sequence repeats. There are hypervariable regions present marked by homopolymeric repeats and low G+C content compared to the whole genome (Parkhill *et al.*, 2000). Most of the hypervariable sequences encode for surface structures such as LOS, capsule, flagellar biosynthesis and the glycosylation locus (Miller, 2008). Later another *C. jejuni* isolate was sequenced and compared with non-*C. jejuni* isolates. *C. jejuni* RM1221 has a large size genome (1.8 Mb) compared to *C. jejuni* 11168. It contains one *Campylobacter* Mu-like prophage (CMLP1) and three insertion elements and some additional capsule biosynthesis genes that are absent from *C. jejuni* NCTC11168. *C. jejuni* NCTC11168 and *C. jejuni* RM1221 were more closely related to *C. coli* than to *C. lari* and *C. upsaliensis* (Fouts *et al.*, 2005). Other *C. jejuni* strains fully genome sequenced and characterised include *C. jejuni* 81-176 (Hofreuter *et al.*, 2006), 81116 (Pearson *et al.*, 2007), CG8486 (Poly *et al.*, 2007b), M1 (Friis *et al.*, 2010), ICDCCJ07001 (Zhang *et al.*, 2010) and S3 (Cooper *et al.*, 2011). The genome of *C. jejuni* subsp. *doylei* has been sequenced and contains 2,037 genes with 251

pseudogenes (Parker *et al.*, 2007). Strain to strain variation is observed based on the genome sequence data. The genome sequence of human clinical isolates NCTC11168, 81-176 and CG8486 have 1,474 conserved genes. *C. jejuni* 81-176 contains 35 unique genes, CG8486 shows 38 unique genes and *C. jejuni* NCTC11168 has 8 unique genes (Champion *et al.*, 2008). A DNA microarray comparison showed that several *C. jejuni* strains were highly variable compared to the reference strain NCTC11168. Seven hypervariable plasticity (PR1-PR7) regions were identified in the genome of *C. jejuni* strains examined that consisted of 50% of the variable gene pool (Pearson *et al.*, 2003). Some *Campylobacter* strains contain plasmids. For example, pVir and pTet plasmids are present in *C. jejuni* 81-176 (Batchelor *et al.*, 2004, Bacon *et al.*, 2000) and the pVir plasmid has a role in pathogenesis (Champion *et al.*, 2008). The number of *Campylobacter* strains sequenced is ever increasing. To date, 15 *Campylobacter* strains have been full genome sequenced consisting of 11 *C. jejuni* strains (including *C. jejuni* subsp. *doylei* 269.97) and 4 non-*C. jejuni* strains. There are also a number of on-going *Campylobacter* genome projects. Whole genome sequencing has become a crucial technique providing useful information about the differences in their virulence potential and host specificity of campylobacters.

6.1.2 Study aims

The CGH study of the hyperinvasive *C. jejuni* showed that *C. jejuni* 01/10 and *C. jejuni* 01/51 were the most variable strains when compared to the reference strains. Similarly, the PSSH comparison of the hyperinvasive *C. jejuni* strains identified most additional sequences in *C. jejuni* 01/10 whereas none of the inserts were present in *C. jejuni* 01/51. Additionally, *C. jejuni* 01/51 has been studied in detail by insertional mutagenesis which identified several genes with potential roles in hyperinvasion of this strain (Javed *et al.*, 2010). It was observed using DNA microarray technology that *C. jejuni* 01/51 lacked some capsule biosynthesis genes which are present in many other *C. jejuni* strains (Dr Georgina Manning; personal communication) and the CGH data in this project showed that this strain possesses the *kps* and some other capsule genes similar to those present in *C. jejuni* RM1221. Based on these studies and available resources *C. jejuni* 01/10 and *C. jejuni* 01/51 were selected for sequencing using next generation sequencing techniques. *C. jejuni* 01/51 was genome sequenced using two methods; pyrosequencing/454 and Illumina sequencing. *C. jejuni* 01/10 was genome sequenced

only by Illumina sequencing. The genome sequence data mining was performed with the following key aims;

- Identify additional genomic content unique to the hyperinvasive *C. jejuni* 01/51 and *C. jejuni* 01/10
- Annotate potential CDS to classify gene function and associate any relationship to the hyperinvasive phenotype exhibited by these strains.
- Phylogenomic comparison of the hyperinvasive *C. jejuni* compared with other available genome sequenced *C. jejuni* strains.

6.2 METHODS

The genomic DNA of *C. jejuni* 01/51 and 01/10 was prepared by using the Qiagen genomic DNA extraction kit. The method followed is discussed in material and methods chapter 2 (section 2.6.1).

6.2.1 Illumina sequencing

The Illumina sequencing was performed by the genome sequence facility at the University of Exeter, Exeter, UK (Figure 6.1).

Genomic DNA library

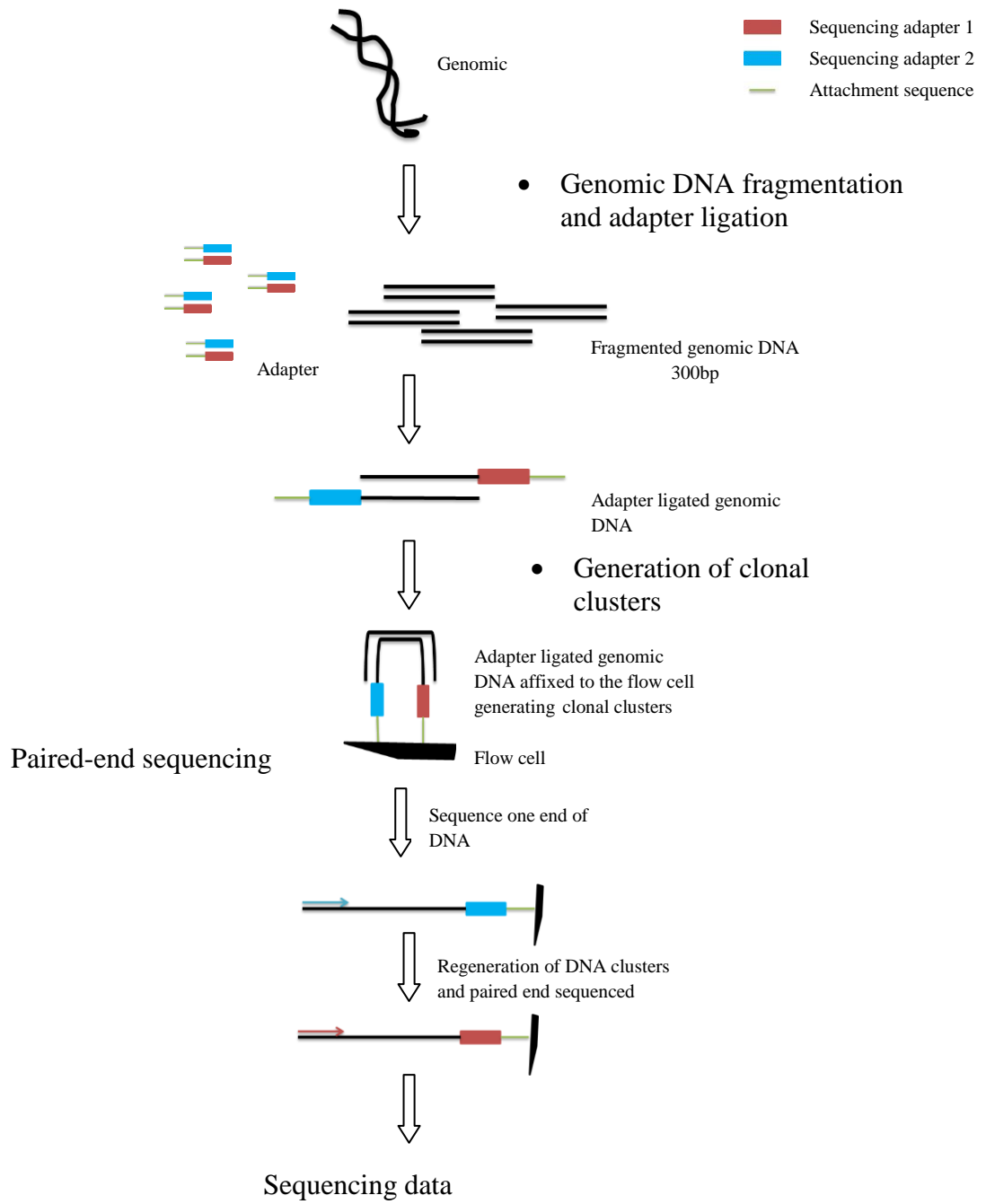


Figure 6.1: A diagrammatic representation of Illumina sequencing technique.

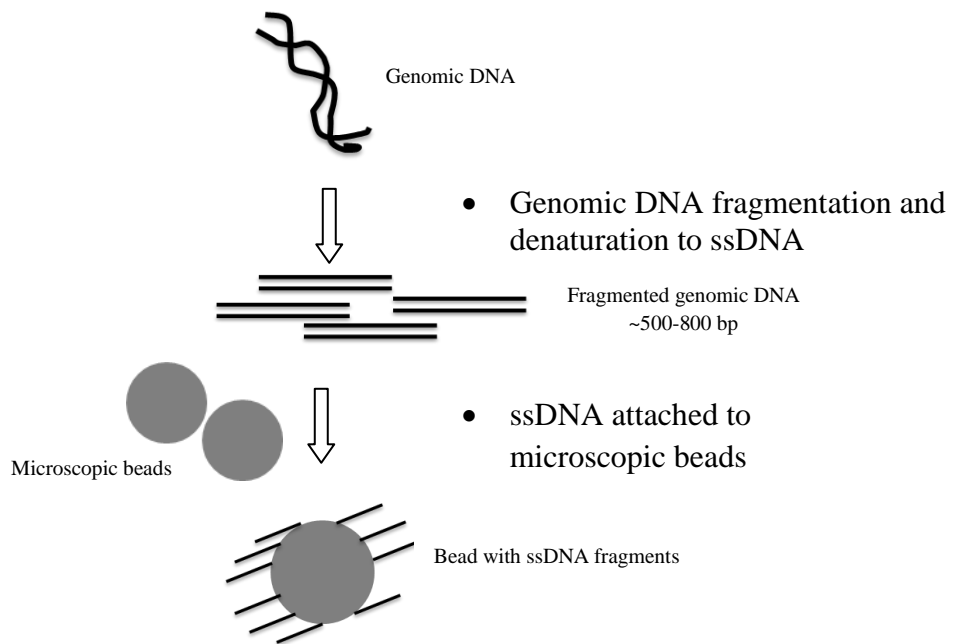
In this sequencing method, the genomic DNA was fragmented by shearing generating 300bp size fragments using the Tru-Seq Genomic library preparation kit. Multiplex PCR was then performed using 6 bp index sequences into a single lane. The genomic DNA fragments were sequenced using paired end 72 bp read lengths on an Illumina GAIIx sequencer using SCS software (v 2.8) (Figure 6.1).

The raw sequence data for *C. jejuni* 01/51 and 01/10 was provided in 72 and 75 contigs respectively. *De novo* assembly was performed using the velvet assembly program (v 1.0.18) (Zerbino and Birney., 2008). The annotation of the reference strain *C. jejuni* RM1221 was transferred onto the un-annotated query sequence using the Rapid Annotation Transfer Tool (RATT) (Otto *et al.*, 2011). GLIMMER (v 3.02) (Salzberg *et al.*, 1998) prediction was used to identify ORFs which were unique to the sequenced genome. These ORFs were annotated by Campydb and the NCBI BLASTx tools.

6.2.2 Pyrosequencing/454

Pyrosequencing of *C. jejuni* 01/51 and genome assembly was performed by Dr Chrystala Constantinidou in the genome sequence facility at the University of Birmingham (Figure 6.2).

Genomic DNA general library preparation



Sequencing reaction

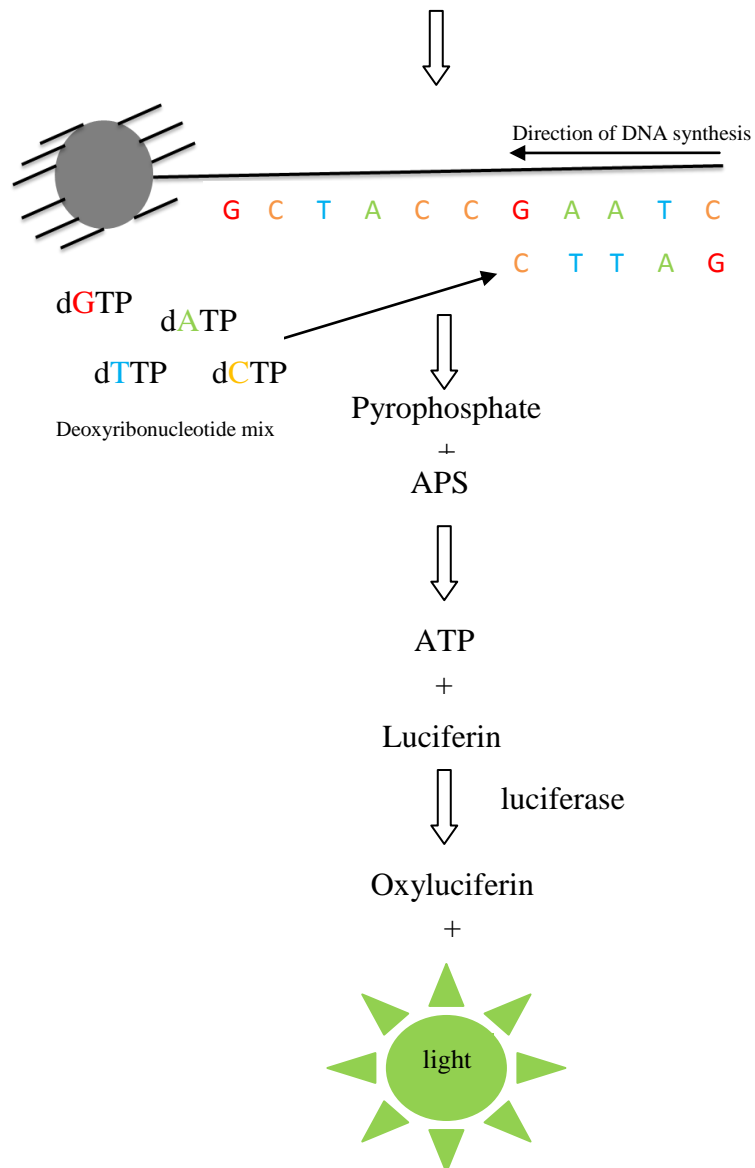


Figure 6.2: A schematic illustration of 454/pyrosequencing method.

C. jejuni 01/51 genomic DNA was sheared into single stranded DNA fragments of 500-800 bp in length. A single end DNA library was prepared following the general library preparation method. The GS FLX Titanium general DNA library kit was used to generate DNA template with amplification and sequencing adaptors. Emulsion PCR was performed for the enrichment of DNA template and the resulting library was sequenced by using the GS FLX Titanium platform (Figure 6.2).

The 454 sequencing performed 12x coverage of the *C. jejuni* 01/51 genome. The contigs were assembled using Newbler (v 2.5) (<http://454.com/products/analysis-software/index.asp>) mapped against the reference *C. jejuni* RM1221. The assembled genome sequence (.embl) of *C. jejuni* 01/51 was provided. An ACT (Carver *et al.*, 2005) comparison sequence file of *C. jejuni* 01/51 and RM1221 was also provided.

The Illumina and 454 sequence data for *C. jejuni* 01/51 was combined using MIRA (v 3.4) (Chevreux *et al.*, 1999). Any un-annotated genomic regions identified at this stage were annotated using Campydb and NCBI BLASTx searches. Features of the sequenced data were analysed using Artemis (Rutherford *et al.*, 2000). The combined *C. jejuni* 01/51 assembly was compared with *C. jejuni* 01/10 Illumina sequence using the Annotation Comparison Tool (ACT) (Carver *et al.*, 2005).

Dr Alan McNally (NTU) assembled *C. jejuni* 01/10 Illumina sequencing data, and performed the combined assembly on *C. jejuni* 01/51 and 01/10 sequences.

6.2.3 Phylogeny

Whole genome phylogeny was performed using the assembled 01/51 and 01/10 genomes and 11 publicly available *C. jejuni* genome sequences (including *C. jejuni* subsp. *doylei* 269.97). Whole genome alignments were performed using Mugsy (Angiuoli., 2011), and the core genome extracted from the resulting alignment using a pipeline developed by Jason Sahl (Sahl., 2012) and adapted by Alan McNally. The concatenated core genome alignment was used to create a maximum likelihood phylogeny with RaxML (Stamatakis., 2005) implementing 100 bootstraps. The resulting tree was visualised and edited using Figtree.

6.3 RESULTS AND DISCUSSION

Two hyperinvasive *C. jejuni* strains (*C. jejuni* 01/10 and 01/51) were genome sequenced and analysed to determine if the two strains contain any additional genomic content that may be the signature for their hyperinvasive phenotype.

6.3.1 Genome sequence facts

The combined assembly of pyrosequencing and Illumina sequencing data for *C. jejuni* 01/51 showed that the genome of *C. jejuni* 01/51 is a single circular chromosome of 1,617,079 bp in length with an average G+C content of 30.45% (Table 6.1). Illumina sequencing of *C. jejuni* 01/10 genome identified it as a single circular chromosome with a genome size of 1,677,053 bp and G+C ratio of 30.49%. Other key characteristics of genomes are in table 6.1 below;

Table 6.1: The genome features of *C. jejuni* 01/51 and 01/10.

Genome features of two <i>C. jejuni</i> genomes	<i>C. jejuni</i> 01/51*	<i>C. jejuni</i> 01/10**
Chromosome size (bp)	1,617,079	1,677,053
Number of contigs	72	75
N50 contig length	103,524	120,479
G+C content	30.45%	30.49%
Number of genes (without pseudo)	1,739	1,724
Genes with function	1,378	1,385
Hypothetical genes (including conserved)	361	339
Genes with unknown function	0	0
Phage/genomic islands	1	2
Plasmids	0	0
Restriction/Modification system	8	8
Virulence related genes		
<i>cadF</i>	1	1
<i>jlpA</i>	1	1
43-kDa MOMP	1	1
Fibronectin binding proteins	3	3
Two component regulator systems	19	17
Response regulator	11	10
Sensor histidine kinase	8	7
Membrane associated proteins	77	72

*features based on Illumina/454 combined assembly

** features based on Illumina sequencing

6.3.2 Phylogeny

A phylogenetic comparison of the whole genome sequence of the hyperinvasive *C. jejuni* 01/51 and 01/10 was made with other published complete *C. jejuni* and *C. jejuni* subsp. *doylei* 269.97 genome sequences (Figure 6.3).

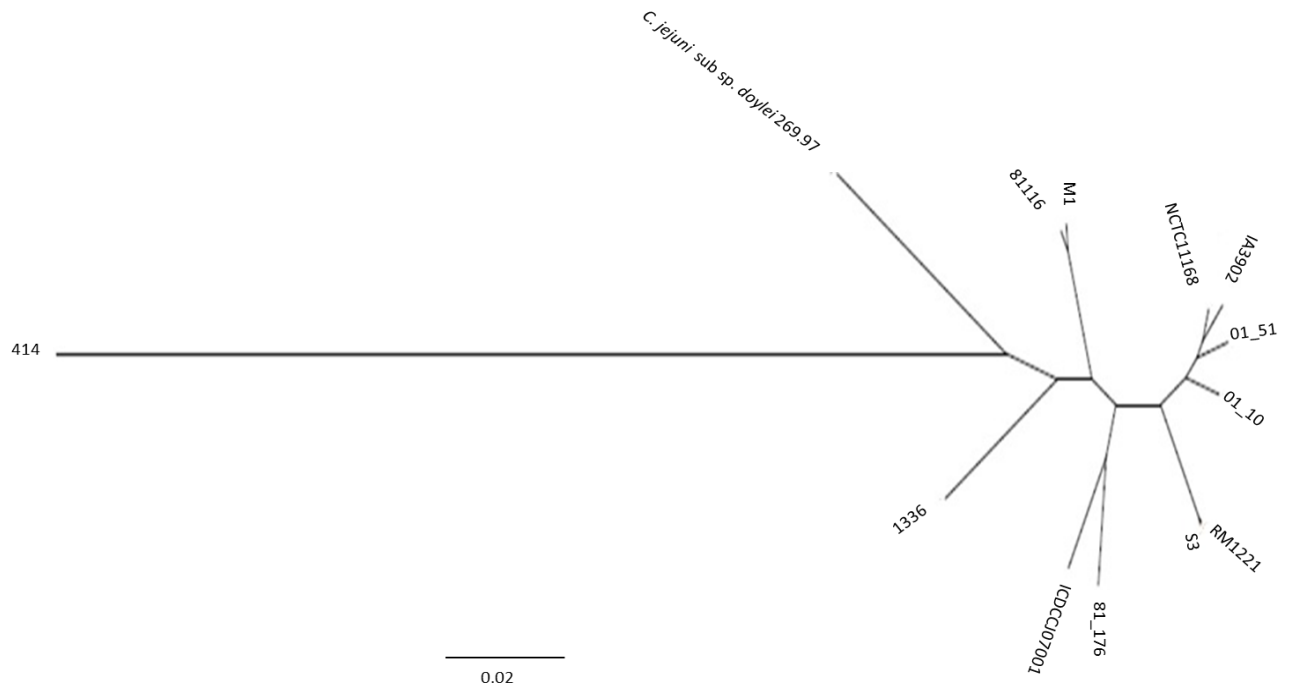


Figure 6.3: Phylogeny of *C. jejuni* 01/10 and 01/51 compared with other *C. jejuni* strains and *C. jejuni* subsp. *doylei* 269.97 based on genome sequence data.

The bar scale at the bottom represents the number of nucleotide substitutions.

Based on the genome sequence data, *C. jejuni* 01/10 and *C. jejuni* 01/51 formed a cluster with *C. jejuni* NCTC11168 and *C. jejuni* IA3902 suggesting that the genome composition of these strains is very similar. The human clinical isolate *C. jejuni* NCTC11168 has a chromosome size of 1,641,481 bp (Parkhill *et al.*, 2000) which is smaller in size compared to the *C. jejuni* chicken isolate RM1221 which is interrupted by four prophages/insertion sequences and some additional capsule loci in its genome (Fouts *et al.*, 2005). *C. jejuni* IA3902 is a pathogenic strain responsible for causing ovine abortion cases in sheep in USA with a chromosome size of 1,672,219 bp (Burrough *et al.*, 2009). The chromosome size of these strains is comparable to that of *C. jejuni* 01/51 and 01/10 respectively. The phylogenomic clustering of *C. jejuni* 01/10

and 01/51 close together reflected that these two hyperinvasive *C. jejuni* strains have a similar genomic content.

6.3.3 Prophages/Genomic regions

When compared against the reference *C. jejuni* RM1221 genome both hyperinvasive *C. jejuni* strains lacked the four *C. jejuni* RM1221 characteristic prophages /genomic islands. Prophages are the vehicle of horizontal or lateral gene transfer. These may carry genes that provide a selective advantage to bacteria for survival in diverse environments and may have a role in virulence (Fouts *et al.*, 2005). *C. jejuni* 01/51 contains only one small prophage of 5,031 bp containing 6 ORFs with similarity to a replication domain protein in *Vibrio cholerae* species and genes with hypothetical function in *Halomonas* strain GFAJ-1 (Phung *et al.*, 2012) (Appendix: supplementary table 5).

In *C. jejuni* 01/10, two prophages were inserted in the genome (Appendix: supplementary table 6). Prophage 1 is 10,297 bp in size containing 7 ORFs with functional homology to phage related integrases and nucleases. Prophage 2 is large 28,602 bp in size and it is composed of 33 ORFs. The majority of these ORFs shared identity with phage structural proteins including prophage basal plate and tail assembly proteins. Additionally, ORFs with homology to nucleases and several hypothetical proteins were also present within this prophage. A few ORFs showed similarity to the genes from *C. jejuni* RM1221 Mu-like prophage (CMLP1) which may suggest that the role of these prophages in these strains is similar. The role of *C. jejuni* RM1221 prophage in *C. jejuni* pathogenesis is not known (Fouts *et al.*, 2005) however recently it was showed that the presence of *dns* (extracellular deoxyribonuclease) in the *C. jejuni* RM1221 Mu-like prophage inhibits natural transformation in *C. jejuni* strains (Gaasbeek *et al.*, 2010). The activation of these prophages and their role in the hyperinvasive virulence trait of these strains cannot be demonstrated at this stage.

C. jejuni 01/51 and 01/10 genomes do not contain any unique genes that are not present in other campylobacters. Some ORFs which showed best matches to members of the *Campylobacter* genus other than *C. jejuni* RM1221 mainly *C. jejuni* subspecies are listed in the supplementary tables 4 (*C. jejuni* 01/51) and 5 (*C. jejuni* 01/10) in the appendix. The LOS biosynthesis region for *C. jejuni* 01/51 and 01/10 are also listed (Appendix: supplementary tables 4 and 5 respectively). Other genes with membrane associated

functions, restriction modification systems and hypothetical roles were mainly identified.

6.3.3.1 Validation of CGH and PSSH study results in *C. jejuni* 01/51 and 01/10 whole genome sequences

Based on the results of CGH study, 522 and 859 divergent genes were identified in *C. jejuni* 01/10 and 01/51 respectively compared with the reference *C. jejuni/C.coli* genes on the pan array. This indicated that *C. jejuni* 01/10 was most similar and *C. jejuni* 01/51 was most divergent from the reference genes on the array compared with other *C. jejuni* strains investigated in the CGH study (section 4.3.7). The whole genome sequencing showed 337 genes to be absent from *C. jejuni* 01/10 and 313 genes were indicated as absent from *C. jejuni* 01/51 when mapped against the reference *C. jejuni* RM1221.

In agreement with the CGH findings, the whole genome sequence of *C. jejuni* 01/10 validated that some capsule and LOS genes homologous to those present in *C. jejuni* RM1221 were identified in the capsule and LOS regions of this strain. The capsule region of *C. jejuni* 01/10 is discussed later (Table 6.3; section 6.3.4) and the LOS region is listed in the supplementary table 6 in appendix. Unlike CGH data, however, the whole genome sequencing showed that *C. jejuni* RM1221 characteristic CJIE 2, 3 and 4 were absent from *C. jejuni* 01/10.

Both the CGH and whole genome sequencing showed that CJIE3 and 4 which are present in *C. jejuni* RM1221 were absent from *C. jejuni* 01/51 genome. In contrast to CGH data, the whole genome sequencing showed that *C. jejuni* RM1221 characteristic CMLP1, CJIE2 were missing from *C. jejuni* 01/51 genome. Interestingly, the whole genome sequencing did not identify any *C. jejuni* RM1221 homologous capsular genes in this strain. The capsule region of *C. jejuni* 01/51 (Table 6.2) is discussed in detail in section 6.3.4. The genome sequence of *C. jejuni* 01/51 showed that the LOS region possessed several LOS genes similar to those present in the LOS locus of *C. jejuni* RM1221 (Appendix: supplementary table 5). Based on the whole genome sequencing, mainly the hypothetical genes present in *C. jejuni* RM1221 were absent from *C. jejuni* 01/10 and 01/51.

The PSSH study identified five sequences (14, 22, 32, 54 and 62) to be present in the hyperinvasive *C. jejuni* strain 01/10 (Table 5.1; section 5.3.7). The whole genome sequencing verified 3/5 of these sequences (14, 22 and 54) in *C. jejuni* 01/10 (Appendix: supplementary table 6). It can be argued that the two sequences (32 and 62) which could not be detected in the genome sequence may not be sequenced as the Illumina sequence represents the draft genome sequence of this strain. In *C. jejuni* 01/51, none of the PSSH identified sequences were present (Table 5.1; section 5.3.7).

6.3.4 *C. jejuni* capsule region

Capsule polysaccharides regions are known to be hypervariable in *Campylobacter* species (Fouts *et al.*, 2005, Dorrell *et al.*, 2001) and so the CPS regions of both 01/10 and 01/51 were studied in some detail.

6.3.4.1 *C. jejuni* 01/51 capsule

Based on 454 and Illumina genome sequencing, *C. jejuni* 01/51 was found to have a diverse capsule region (Figure 6.4).

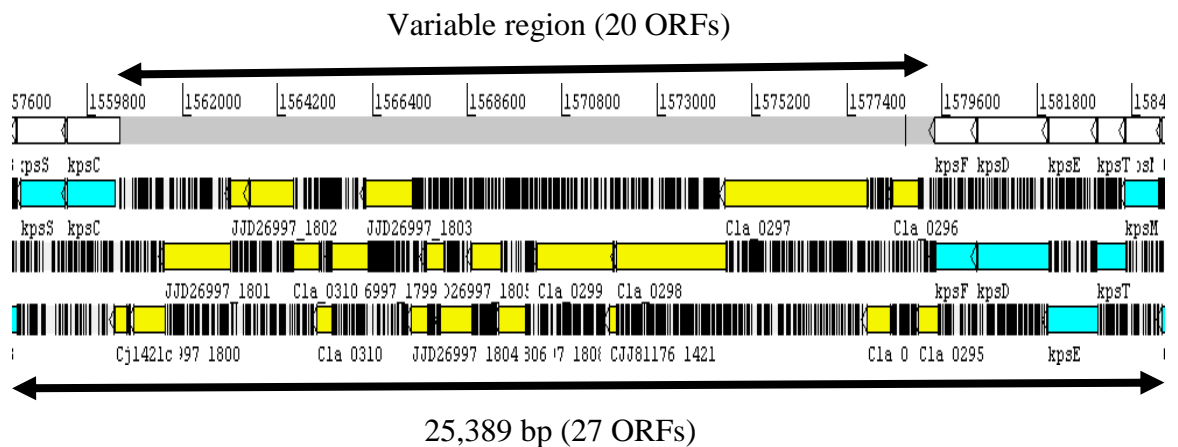


Figure 6.4: The capsule region in *C. jejuni* 01/51.

The ORFs in the capsular central variable region are yellow and the bordering *kps* genes are skyblue.

The capsule in *C. jejuni* 01/51 is a 25,389 bp region containing 27 ORFs. It is composed of a unique central variable region (20 genes) bordered by conserved *kps* genes encoding for the capsule polysaccharide export proteins.

The majority of genes in the variable region share similarity with *C. jejuni* subsp. *doylei* (9/27) and *C. lari* (9/27) capsule genes (Table 6.2) annotated here as JJD26997 and Cla respectively.

Table 6.2: CDS in the capsule region of *C. jejuni* 01/51.

Locus tag	Gene homolog/ortholog	Function
<i>kpsS</i>	CJE1600	capsule polysaccharide export protein KpsS [<i>Campylobacter jejuni</i> RM1221]
<i>kpsC</i>	CJE1601	capsule polysaccharide export protein KpsC [<i>Campylobacter jejuni</i> RM1221]
	CJ1421c	capsular polysaccharide biosynthesis heptosyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC11168]
	JJD26997_1800	nucleoside-diphosphate-sugar pyrophosphorylase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1801	capsular polysaccharide biosynthesis protein [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1802	capsular polysaccharide biosynthesis protein, putative [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	Cla_0310	capsular polysaccharide biosynthesis protein, putative [<i>Campylobacter lari</i> RM2100]
	Cla_0310	capsular polysaccharide biosynthesis protein, putative [<i>Campylobacter lari</i> RM2100]
	Cla_0310	capsular polysaccharide biosynthesis protein, putative [<i>Campylobacter lari</i> RM2100]
	JJD26997_1799	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1803	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1804	conserved domain protein [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1806	conserved hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1807	HAD-superfamily hydrolase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	JJD26997_1808	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	Cla_0299	putative sugar transferase [<i>Campylobacter lari</i> RM2100]
	CJJ81176_1421	putative sugar transferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
	Cla_0298	hypothetical protein [<i>Campylobacter lari</i> RM2100]
	Cla_0297	putative glycosyltransferase [<i>Campylobacter lari</i> RM2100]
	Cla_0296	conserved hypothetical protein [<i>Campylobacter lari</i> RM2100]
	Cla_0296	conserved hypothetical protein [<i>Campylobacter lari</i> RM2100]
	Cla_0295	putative glycerol-3-phosphate cytidyltransferase [<i>Campylobacter lari</i> RM2100]
<i>kpsF</i>	CJE1617	rabinose-5-phosphate isomerase [<i>Campylobacter jejuni</i> RM1221]
<i>kpsD</i>	CJE1618	capsular polysaccharide ABC transporter, periplasmic polysaccharide-binding protein [<i>Campylobacter jejuni</i> RM1221]
<i>kpsE</i>	CJE1619	capsular polysaccharide ABC transporter [<i>Campylobacter jejuni</i> RM1221]
<i>kpsT</i>	CJE1620	capsular polysaccharide ABC transporter, ATP-binding protein [<i>Campylobacter jejuni</i> RM1221]
<i>kpsM</i>	CJE1621	capsular polysaccharide ABC transporter, permease protein [<i>Campylobacter jejuni</i> RM1221]

6.3.4.2 *C. jejuni* 01/10 capsule

The complete capsule region in *C. jejuni* 01/10 is 35,448 bp in size and contains 29 ORFs (Figure 6.5). Of these 29 ORFs, seven are conserved *kps* genes enclosing the central variable region consisting of 22 ORFs.

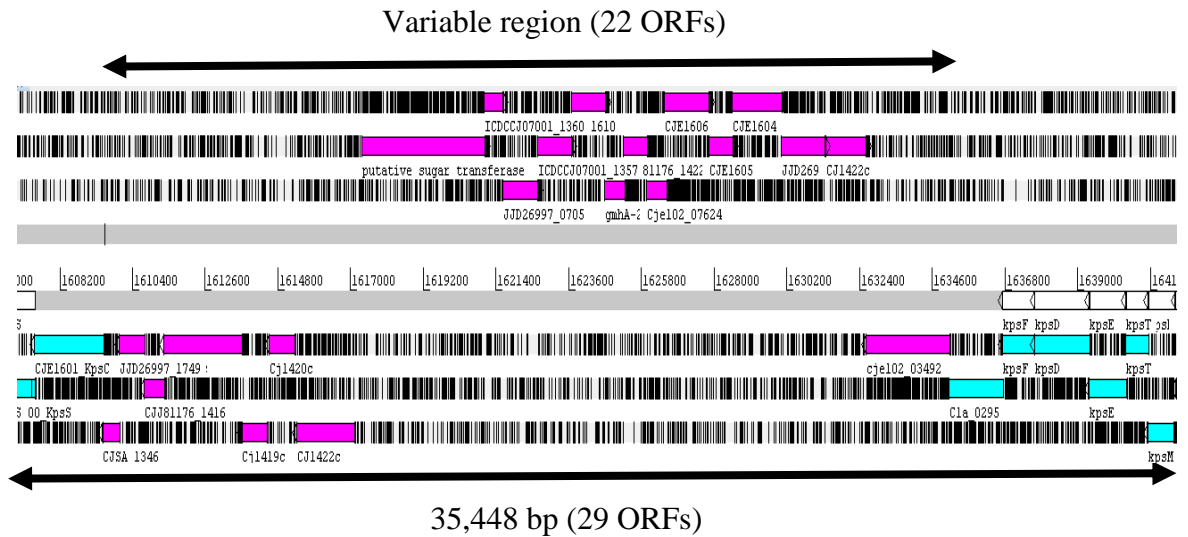


Figure 6.5: The capsule locus in *C. jejuni* 01/10.

The CDS in the central variable capsular region are pink and the surrounding *kps* genes are skyblue.

The CDS in the capsule region of *C. jejuni* 01/10 are listed in table 6.3.

Table 6.3: Genes in the capsule region of *C. jejuni* 01/10.

Locus tag	Gene homolog/ortholog	Function
<i>kpsS</i>	CJE1600	capsule polysaccharide export protein KpsS [<i>Campylobacter jejuni</i> RM1221]
<i>kpsC</i>	CJE1601	capsule polysaccharide export protein KpsC [<i>Campylobacter jejuni</i> RM1221]
	CJSA_1346	<i>cysC</i> adenylsulfate kinase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]
	JJD26997_1749	putative sugar-1-phosphate nucleotidyltransferase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	CJH81176_1416	class I glutamine amidotransferase, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
	CJ1418c	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC11168]
	CJ1419c	methyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC11168]
	CJ1420c	methyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC11168]
	CJ1422c	sugar transferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC11168]
	putative sugar transferase	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218
	ICDCCJ07001_1360	dTDP-6-deoxy-D-xylo-4-hexulose-3,5-epimerase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
	JJD26997_0705	GDP-fucose synthetase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	ICDCCJ07001_1357	GDP-mannose 4,6-dehydratase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
	CJE1610	capsular biosynthesis sugar kinase [<i>Campylobacter jejuni</i> RM1221]
	<i>gmhA-2</i>	phosphoheptose isomerase [<i>Campylobacter jejuni</i> RM1221]
	CJH81176_1422	capsular biosynthesis nucleotidyltransferase, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
	Cje102_07624	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
	CJE1606	haloacid dehalogenase-like hydrolase [<i>Campylobacter jejuni</i> RM1221]
	CJE1605	capsular polysaccharide biosynthesis protein [<i>Campylobacter jejuni</i> RM1221]
	CJE1604	capsular polysaccharide biosynthesis protein [<i>Campylobacter jejuni</i> RM1221]
	JJD26997_1797	alpha-2,3-sialyltransferase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
	CJ1422c	sugar transferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC11168]
	Cje102_03492	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
	Cla_0295	putative glycerol-3-phosphate cytidyltransferase [<i>Campylobacter lari</i> RM2100]
<i>kpsF</i>	CJE1617	rabinose-5-phosphate isomerase [<i>Campylobacter jejuni</i> RM1221]
<i>kpsD</i>	CJE1618	capsular polysaccharide ABC transporter, periplasmic polysaccharide-binding protein [<i>Campylobacter jejuni</i> RM1221]
<i>kpsE</i>	CJE1619	capsular polysaccharide ABC transporter [<i>Campylobacter jejuni</i> RM1221]
<i>kpsT</i>	CJE1620	capsular polysaccharide ABC transporter, ATP-binding protein [<i>Campylobacter jejuni</i> RM1221]
<i>kpsM</i>	CJE1621	capsular polysaccharide ABC transporter, permease protein [<i>Campylobacter jejuni</i> RM1221]

The majority of genes (25/29) in the capsule region were homologous to other *C. jejuni* capsule genes however, the CPS region in *C. jejuni* 01/10 was also found to contain three genes with similarity to *C. jejuni* subsp. *doylei* capsule genes (JJD26997_1749, JJD26997_0705 and JJD26997_1797) and one gene showed homology to *C. lari* CPS gene (Cla_0295). This *C. lari* homologous capsule gene was also present in the *C. jejuni* 01/51 capsule region. However, the level of interspecies similarity of genes in *C. jejuni* 01/10 CPS region was not as seen in the case of *C. jejuni* 01/51.

C. lari is found in wild birds, particularly seagulls (Glunder and Petermann., 1989). It is frequently isolated from freshwater, seawater and shellfish (Rosef *et al.*, 2008). Unlike *C. jejuni* and *C. coli* which are isolated from human gastroenteritis cases only a limited number of *C. lari* isolates are associated with human illness (Miller *et al.*, 2008b), however there are reports of *C. lari* causing severe bacteraemia in humans (Werno *et al.*, 2002, Godreuil *et al.*, 2000). *C. lari* RM2100 is a human clinical isolate (Fouts *et al.*,

2005, Miller *et al.*, 2008b). Based on its genome sequence, ~ 90% of the genome content of this strain is similar to other *Campylobacter* species (Fouts *et al.*, 2005, Miller *et al.*, 2008b). Similarly, *C. jejuni* subsp. *doylei* is more frequently isolated from blood cultures than stool samples (Parker *et al.*, 2007).

The homology of genes within the capsule region of the hyperinvasive *C. jejuni* 01/51 and *C. jejuni* 01/10 to *C. lari* and *C. jejuni* subsp. *doylei* capsule genes makes it a novel region as this interspecies mosaicism in the capsule locus has not been reported in other sequenced *C. jejuni* strains. This mosaic pattern of genes in the capsule region suggests that homologous recombination is an active phenomenon in this region which has resulted in these two hyperinvasive *C. jejuni* strains acquiring genes from other pathogenic *Campylobacter* subspecies. The presence of capsule genes from *C. lari* and *C. jejuni* subsp. *doylei* in hyperinvasive *C. jejuni* 01/10 and 01/51 strains is a common genetic signature of these two CPS regions which has not been observed in other campylobacters and perhaps this trait is the determinant of hyperinvasiveness in these strains. Furthermore, JJD26997_1801 in the *C. jejuni* 01/51 CPS region with similarity to the *C. jejuni* subsp. *doylei* capsule gene (Table 6.2) was also previously identified by transposon mutagenesis study of *C. jejuni* 01/51 and a mutant in this gene resulted in reduced invasion in INT-407 and Caco-2 epithelial cells (Javed *et al.*, 2010) confirming the role of capsule in the hyperinvasive phenotype of *C. jejuni* 01/51.

It is important to validate the role of capsule in the hyperinvasive profile of *C. jejuni* 01/10 by mutagenesis. The Cla_0295 homologous gene encoding for a putative glycerol-3-phosphate cytidyltransferase in *C. lari* RM2100 is the best candidate for mutagenesis study as a homologue of this gene was also identified in *C. jejuni* 01/51 CPS region.

Interestingly, some genes were identified in multiple copies. For example, three copies of the Cla_0310 orthologous genes encoding for a putative polysaccharide biosynthesis protein and two copies of genes encoding for the conserved hypothetical proteins (Cla_0296) were identified in the CPS region of *C. jejuni* 01/51. Similarly, the capsule locus in *C. jejuni* 01/10 possesses two copies of a sugar transferase encoding gene, Cj1422c. Previous studies on *C. jejuni* capsule regions have reported that gene duplication is commonly observed in the CPS region of *C. jejuni* strains which adds to the variability in this region (Karlyshev *et al.*, 2005a, Guerry *et al.*, 2012, Parker and

Huynh., 2012). These multiple copies of genes may provide selective advantage to bacteria in survival inside the host and disease profile (Fouts *et al.*, 2005). An additional 11 capsule polysaccharide genes were identified in *C. jejuni* 01/51 which were not a part of the capsule region but were randomly distributed in the chromosome (supplementary table 5). Of these eight genes (*kpsS*, *kpsC*, CJSA_1346, JJD26997_1749, CJJ81176_1416, Cj1418c, Cj1419c, Cj1420c) were present clustered together. This also shows that *C. jejuni* 01/51 has two copies of some of the *kps* gene homologues. The other three genes (CJSA_1363 and CJSA_1352) and CJSA_1357 were located separately. This random distribution of capsule genes has also been observed in CPS regions for *C. jejuni* strains of different Penner serotypes (Parker and Huynh., 2012) and was reported in the *C. upsaliensis* capsule locus (Fouts *et al.*, 2005). Unlike in *C. jejuni* 01/51, no additional CPS genes outside the capsule region could be identified in *C. jejuni* 01/10.

It have been reported that despite the variability in the central CPS region some capsular genes are conserved in many *C. jejuni* strains. These include the heptose biosynthesis genes (*hddC*, *gmhA*, *hddA*) and the genes encoding for O-methyl phosphoramidate (MeOPN) capsular modifications (Guerry *et al.*, 2012). In *C. jejuni* NCTC11168, four genes (Cj1415c-Cj1418c) have been reported to be involved in MeOPN synthesis and these genes are highly conserved in other *C. jejuni* strains. Also, two genes Cj1419c and Cj1420c with a role in methyl transferases were always found adjacent to the MeOPN synthesis genes (McNally *et al.*, 2007). Two transferases encoding genes Cj1421c and Cj1422c present in *C. jejuni* NCTC11168 are responsible for adding MeOPN to two different sugars in a HS2 serotype CPS (McNally *et al.*, 2007) (Figure 6.6).

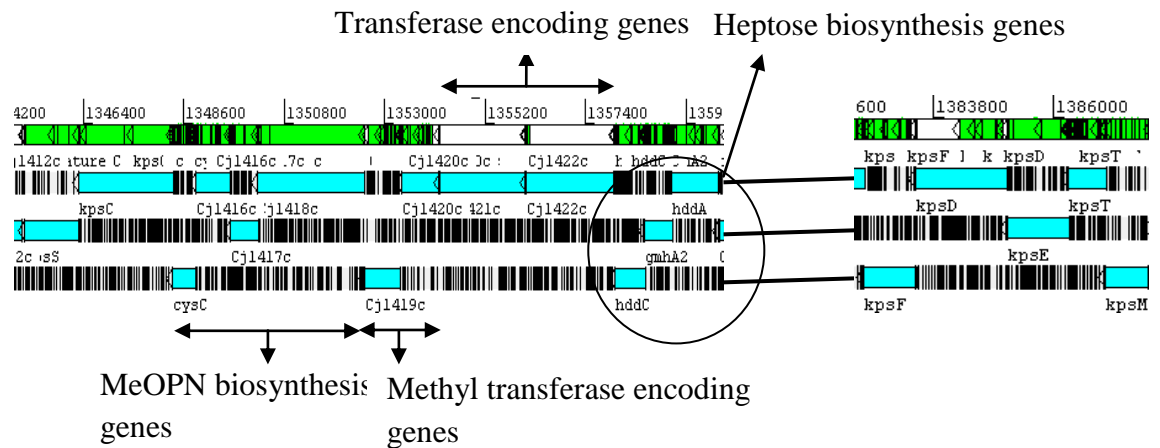


Figure 6.6: A representation of the partial capsule locus in *C. jejuni* NCTC111168 showing heptose and MeOPN biosynthesis genes and transferase encoding genes.

In *C. jejuni* 01/51 CPS region, the *hddC*, *gmhA*, *hddA* homologous gene cluster was not identified. *C. jejuni* 01/10 contains a *gmhA-2* homologous gene in its CPS locus. Parker and Huynh (2012) have also observed that these heptose synthesis genes were missing from *C. jejuni* subsp. *jejuni* IA3902. Additionally, one MeOPN synthesis gene homolog (Cj1418c) was identified within the *C. jejuni* 01/10 CPS region and as a part of the *C. jejuni* 01/51 additional capsule genes with the methyl transferases encoding genes (Cj1419c-Cj1420c) present adjacent to it. In *C. jejuni* 01/51, the ORFs with similarity to the two transferases (Cj1421c and Cj1422c) were not present and *C. jejuni* 01/10 contains two copies of Cj1422c homologous gene.

Further analysis of the capsule locus does need to be done.

6.3.5 Is capsule or LOS the serodeterminant in *C. jejuni* 01/51?

Capsule is considered as the serodeterminant in Penner HS serotyping of *C. jejuni* strains (Karlyshev *et al.*, 2000) and in this study *C. jejuni* 01/51 was identified as serotype HS4 (section 4.3.2 and Figure 4.3 in chapter 4).

The nucleotide sequence of the capsule locus of other HS4 serotype *C. jejuni* strains (Poly *et al.*, 2011) was compared with the capsule sequence of *C. jejuni* 01/51 but no similarity was observed. Interestingly a mutant in capsule gene, JJD26997_1801 (Javed *et al.*, 2010) and a mutant in a LOS gene (*cj1136*) (Javed *et al.*, 2012) in *C. jejuni* 01/51

were also serotyped as part of this study. The capsule gene (JJD26997_1801) mutant retained the serotype of the wild type strain whereas the mutant in the LOS gene (*cj1136*) gene altered the serotype of the mutant to HS50. This may suggest that LOS and not the capsule is responsible for serotype specificity in this strain.

It would be useful to confirm the above observation in *C. jejuni* 01/10 generating mutants in LOS and capsule genes.

6.4 SUMMARY

In summary, the phylogeny based on the whole genome sequence of the hyperinvasive *C. jejuni* 01/10 and 01/51 grouped them together. There was no unique genomic content present in these strains except for a prophage in *C. jejuni* 01/51. Two prophages were found in *C. jejuni* 01/10. The capsule region was the most diverse region but was different between the two strains. The capsule loci in *C. jejuni* 01/51 and 01/10 CPS region showed homology with the *C. jejuni* subsp. *doylei* and *C. lari* capsule genes. This mosaicism in the capsule region containing genes with homology to other *Campylobacter* species has not been reported in other *C. jejuni* strains. This shows that the capsule region in hyperinvasive *C. jejuni* 01/51 and 01/10 is a hotspot for homologous recombination and that this mosaicism in the capsule region may be a marker for hyperinvasion in *C. jejuni*. Some capsular genes were present in multiple copies in the capsule region of *C. jejuni* 01/51 and 01/10 with a few capsule genes also identified outside the CPS region in *C. jejuni* 01/51. *C. jejuni* 01/51 is serotype HS4 but its capsule sequence is different from other HS4 serotype *C. jejuni* strains. A capsule gene, JJD26997_1801, mutant in *C. jejuni* 01/51 did not change the serotype whereas a LOS gene, *cj1136*, mutant changed the serotype of the mutant suggesting that LOS is the serodeterminant in *C. jejuni* 01/51.

Chapter Seven

**THESIS DISCUSSION AND
FUTURE WORK**

THESIS DISCUSSION AND FUTURE WORK

C. jejuni is an enteric pathogen and it is the major cause of campylobacteriosis worldwide. The molecular mechanisms underlying *C. jejuni* pathogenesis are still being investigated. Some key factors that have been studied to be associated with the *C. jejuni* pathogenesis include motility, chemotaxis, toxin production and invasion (Young *et al.*, 2007, Dasti *et al.*, 2010). *C. jejuni* has been shown to successfully invade the epithelial cells in *in vitro* assays in several studies (Fauchere *et al.*, 1986, Konkel and Joens., 1989, DeMelo *et al.*, 1989, Everest *et al.*, 1992). Usually the severity of clinical symptoms is related to the ability of *C. jejuni* isolates to invade the intestinal epithelial cells (Konkel *et al.*, 2001). Recently, an invasion study of *C. jejuni* isolated from different sources grouped together six strains based on their hyperinvasive phenotype (Fearnley *et al.*, 2008). One of these hyperinvasive *C. jejuni* strains, 01/51, was investigated further by transposon mutagenesis and a number of genes were identified with potential roles in invasion (Javed *et al.*, 2010). In this research project, all the hyperinvasive *C. jejuni* strains identified by Fearnley *et al.* (2008) were studied at the genome content level to identify any common basis of hyperinvasiveness in this group of strains. For the genome based study, CGH, PSSH and next generation genome sequencing methods have been used.

In the preliminary part of this research project all the hyperinvasive *C. jejuni* strains were tested in *in vitro* assays of environmental stress that the bacteria are exposed to during invasion assays (*i.e.* growth rate, survival in atmospheric air and hydrogen peroxide stress, and motility) and a few other virulence related factors (*i.e.* autoagglutination, sodium deoxycholic acid stress and biofilm formation). Motility is an important virulence phenotype in *C. jejuni* and loss of flagella has been shown to result in reduced invasion of intestinal epithelial cells in several studies (Grant *et al.*, 1993, Wassenaar *et al.*, 1991). Reactive oxygen species (ROS) are produced as toxic molecules inside macrophages to kill intracellular pathogens by damaging the essential cellular components including nucleic acids, lipids and proteins (Jamieson, 1998). To circumvent the harmful effects of ROS, *C. jejuni* has evolved several protective enzymes *e.g.* catalase and superoxide dismutase (De Melo *et al.*, 1989, Day *et al.*, 2000). During the invasion assays, bacteria are exposed to a number of stress factors including 5% (v/v) CO₂, atmospheric air and Triton X-100. Also, the radical oxygen molecules are

produced as a byproduct of oxidative metabolism in the actively growing tissue culture cells. In this study, all the *C. jejuni* strains showed a similar pattern of growth in the exponential phase (section 3.3.1). In addition, all the *C. jejuni* strains survived the atmospheric air and hydrogen peroxide stresses and displayed high levels of variation in motility (sections 3.3.2 to 3.3.4). For the other virulence related phenotypes studied, strain to strain variation was evident with no grouping of the hyperinvasive *C. jejuni* strains (sections 3.3.5 to 3.3.7) and so at this point it seemed that the only phenotype relating these strains to each other was their hyperinvasiveness.

Since the six hyperinvasive strains appeared to be significantly more invasive than the other strains tested at the time (Fearnley *et al.*, 2008) it was hypothesized that they possessed extra/novel DNA that was related to this phenotype. Several studies have successfully used DNA microarrays to highlight similarities and differences at the gene content level that may account for the observed phenotypic variation between *C. jejuni* strains (Quiñones *et al.*, 2008, Parker *et al.*, 2006, Champion *et al.*, 2005, Pearson *et al.*, 2003, Dorrell *et al.*, 2001). In this study, the hyperinvasive *C. jejuni* strains were compared with a group of four low invasive *C. jejuni* strains by using CGH. The objective was to determine if the hyperinvasive *C. jejuni* strains would group together as a distinct group based on their genomic content and also to identify similar regions of variability between these strains. The hierarchical clustering based on CGH did not group the hyperinvasive *C. jejuni* strains together. The hyperinvasive *C. jejuni* strains exhibited different MLST and HS serotype profiles (section 4.3.2). Further analysis of the CGH data identified 67 genes which were present or variable in the majority of the hyperinvasive *C. jejuni* compared with the low invasive *C. jejuni* strains (section 4.3.3 and appendix: supplementary table 4). Group 1 consisted of 9 genes that were present in all the hyperinvasive *C. jejuni* strains and highly divergent/absent from the majority of low invasive strains. Similarly, group 2 possessed 6 genes that were highly divergent/absent from the all the hyperinvasive *C. jejuni* and present in most of the low invasive *C. jejuni* strains (section 4.3.3). Since only 4 low invasive *C. jejuni* strains were studied by using CGH an additional 9 low invasive *C. jejuni* strains were screened by PCR for the genes identified in groups 1 and 2 (section 4.3.4). The PCR analysis further validated the CGH data and showed a statistically significant association between presence or absence of 13/15 genes (except for CJE1128 and CJE0731) in the

hyperinvasion phenotype. Hence, these genes may be considered as putative markers of hyperinvasiveness.

Further characterisation of these genes is required.

The majority of genes identified in the CGH study belonged to different functional categories (section 4.3.5). These genes were not located in the same operon and were randomly distributed in the pan genome (section 4.3.6). Group 1 was dominated by the metabolic genes (*proC* and *metF* and *modA*) as well as posttranslational modification, protein turnover, and chaperone encoding genes (*aat* and CJE0320). In group 2, genes encoding for proteins involved in translation and posttranslational functions (*tgt* and CJE0801), cell membrane biogenesis (CJE0315) and nucleic acid replication and repair (CJE0731) were identified. In addition both groups (1 and 2) contained genes with hypothetical roles. Overall the other groups (Appendix: supplementary table 4) also represented functional groups including the iron transport system (CfrA; CJE0347 and CeuB; CJE1541), arsenic resistance encoding gene (*arsC*) and the twin-arginine translocase (TAT) secretion system (CJE1310). This suggests that the genes involved in essential cellular and metabolic functions are important in defining the hyperinvasive phenotype. Additionally, this unique phenotype is characterised by genes from different cellular networks rather than by gene(s) belonging to a single functional category. Subtle genetic changes in the nucleotide sequence of a gene introduced by point mutation, addition/deletion or substitution of a single nucleotide may account for the observed variation in phenotype (Malik-Kale *et al.*, 2007). These changes cannot be identified by DNA microarrays and whole genome sequencing will be required to observe them. However, genome sequencing for a large number of strains can be expensive; hence, the importance of DNA microarrays cannot be neglected. It is also important to consider the host factors that contribute to an infection process which further complicates the strain to strain phenotype variations (Taboada *et al.*, 2007). The increased invasiveness observed in *in vitro* cultured cell lines may not be due to differences in genetic content but due to expression or regulation differences in single or multiple genes (Konkel *et al.*, 1990). Therefore, it would be interesting to compare the full sequences of genes identified in the CGH study as present in both hyperinvasive and low invasive strains (Appendix: supplementary table 4) to identify more subtle differences contributed by single nucleotide changes in the gene sequence. Also, it is possible that those genes identified as being present in all the hyperinvasive *C. jejuni* and only one of the low invasive *C. jejuni* strains (Group 1: section 4.3.3) are not

expressed in that low invasive *C. jejuni* strain which can be tested by transcriptional analysis of these genes and this work is currently being undertaken by others in the laboratory.

The majority of *C. jejuni* strains showed genomic diversity, mainly in surface related structures, including the LOS, CPS and flagella biosynthesis loci as well as in the restriction modification (RM) systems (section 4.3.7). This strain to strain variation has been observed in other studies (Dorrell *et al.*, 2001, Pearson *et al.*, 2003, Parker *et al.*, 2006). It was also interesting to note that *C. jejuni* 01/10 was most similar to the reference genes on the array whereas *C. jejuni* 01/51 was the most diverse hyperinvasive *C. jejuni* strain.

Since DNA microarray analysis only provides comparison with the reference genes on the array, the hyperinvasive *C. jejuni* strains were also studied by using pooled suppressive subtractive hybridization (PSSH) (Gerrish *et al.*, 2010) to identify any additional genomic content that the hyperinvasive *C. jejuni* strains commonly share which may account for their hyperinvasive phenotype. In this technique, the genomic DNA of all the hyperinvasive *C. jejuni* strains was pooled together as “tester” and hybridized against a pool of four low invasive *C. jejuni* strains “driver”. Eleven sequences were identified in total and validated by PCR analysis as being absent from all the four low invasive driver *C. jejuni* strains (section 5.3.7). PCR screening also showed the distribution of the 11 identified sequences in the hyperinvasive *C. jejuni* strains and in an additional 9 low invasive *C. jejuni* strains that were not used in the PSSH experiment. Each sequence was found to be present in one or more hyperinvasive *C. jejuni* strains but none of them were present in all hyperinvasive strains. These sequences were variably distributed in the non-PSSH tested low invasive strains. Four sequences (14, 38, 54, and 57) were only identified in the hyperinvasive *C. jejuni* strains as these sequences were also absent from the PCR analysed 9 low invasive *C. jejuni* strains. However, these sequences cannot be associated with the hyperinvasive phenotype and likely only represent strain specific sequences. None of the identified sequences were present in *C. jejuni* 01/51 whereas *C. jejuni* 01/10 was positive for most (5/11) inserts.

The 11 identified inserts showed homology with genes in other *Campylobacter* strains and from other bacteria (section 5.3.8). Insert 14 showed similarity to a prophage

MuSo1, F protein in *C. jejuni* subsp. *jejuni* ICDCJ07001 and 260.94. Inserts 54 and 57 were homologous to a putative tetracycline resistance gene *tet* (O/W/32/O) in *Streptococcus gallolyticus* subsp. *gallolyticus* ATCC BAA-2069 plasmid pSGG1 and the phage tail fibre protein I in *Bordetella avium* 197N respectively. Most genome sequenced *Campylobacter* strains contain characteristic prophages which contribute to the genetic diversity and stability of bacterial genomes (Fouts *et al.*, 2005). Insert 38 had no known sequence match in the genome sequence databases searched and many represent strain specific DNA. Therefore, the identification of prophage-related sequences is expected in the hyperinvasive *C. jejuni* strains.

The role of these genes in invasiveness can be studied by mutagenesis. The PSSH method can successfully identify additional genomic content in the tester strains but it is a complicated and time-consuming method (Ahmed *et al.*, 2002, Hepworth *et al.*, 2007).

Based on the results of CGH and PSSH studies, the genomic structure of two hyperinvasive *C. jejuni* strains was studied further by whole genome sequencing. High through-put genome sequencing is the most reliable method to study genetic content variations and to identify pathogenesis associated in *C. jejuni* strains (Fouts *et al.*, 2005, Hofreuter *et al.*, 2006, Pearson *et al.*, 2007, Poly *et al.*, 2007b, Friis *et al.*, 2010, Zhang *et al.*, 2010). The hyperinvasive *C. jejuni* 01/51 strain was sequenced by using 454/pyrosequencing and Illumina sequencing methods and the genome sequences by the two methods were combined whereas for the hyperinvasive *C. jejuni* 01/10 the sequencing was performed just by using the Illumina sequencing. The genomes of *C. jejuni* 01/51 and *C. jejuni* 01/10 are 1,617,079 bp and 1,677,053 bp in sizes respectively (section 6.3.1). The phylogenomic clustering compared with all eleven published complete *C. jejuni* and *C. jejuni* subsp. *doylei* 269.97 genome sequences clustered the two hyperinvasive *C. jejuni* strains together suggesting that their genomic content is more similar to each other than to the other strains in the database (section 6.3.2). Genome sequencing of the other 4 hyperinvasive *C. jejuni* strains (Fearnley *et al.*, 2008) would be helpful in studying the genomic diversity and evolution of these strains. The genome sequences of both strains identified strain specific sequences with gene similarities within the *Campylobacter* genus suggesting that these strains do not contain unique sequences. The only exception was of a prophage identified in the *C. jejuni* 01/51 that showed similarity with genes from *V. chloreae* and *Halomonas* species. Two prophages were identified in *C. jejuni* 01/10 which showed similarity with

the other *Campylobacter* prophage related genes (section 6.3.3). The capsule region of the two genome sequenced hyperinvasive *C. jejuni* strains was highly diverse (section 6.3.4). In the hyperinvasive *C. jejuni* strain 01/51, the CPS region was found to be 25,389 bp in size and the hyperinvasive *C. jejuni* 01/10 possessed a larger (35,448 bp) CPS region. The majority of genes in *C. jejuni* 01/51 capsule region showed similarity with the *C. lari* and *C. jejuni* subsp. *doylei* capsule genes. The capsule region in *C. jejuni* 01/10 contains most genes showing homology with other *C. jejuni* strains, however three genes with similarity to the *C. jejuni* subsp. *doylei* capsule genes and one *C. lari* capsule gene homologue were also identified in the *C. jejuni* 01/10 CPS region. This mosaic pattern of gene distribution in the capsule region sharing homology with other *Campylobacter* species capsule genes has not been reported previously in *C. jejuni*. It can be suggested that the capsule region in the two genome sequenced hyperinvasive *C. jejuni* strains is highly recombinogenic and may be the signature for the hyperinvasive phenotype in these strains. Furthermore, a mutant in a capsule gene (JJD26997_1801) in *C. jejuni* 01/51 resulted in reduced invasion in INT-407 and Caco-2 (Javed *et al.*, 2010) proposing the role of this gene in the hyperinvasive phenotype of *C. jejuni* 01/51. The homologue of the Cla_0295 capsule gene in *C. lari* RM2100 was present in both the *C. jejuni* 01/10 and 01/51 CPS regions and mutants in this gene should be tested in invasion studies to confirm the role of capsule in hyperinvasiveness. The structural analysis of the *C. jejuni* 01/51 and 01/10 capsules will further support the genome sequence data.

There were additional capsule genes identified as randomly scattered in the genome of *C. jejuni* 01/51 that were not a part of the capsular region. This was also observed by Parker and Huynh (2012) in capsules of *C. jejuni* strains of different Penner serotypes. Also, gene duplication was noted in the capsule region of *C. jejuni* 01/51 and 01/10 which has been reported in other *Campylobacter* strains (Karlyshev *et al.*, 2005a, Fouts *et al.*, 2005, Parker and Huynh., 2012).

The capsule has been considered as the serodeterminant in the Penner serotyping scheme (Karlyshev *et al.*, 2000). *C. jejuni* 01/51 and 01/10 were serotyped as HS4 and HS50 respectively (section 4.3.2 and Figure 4.3; chapter 4). Poly *et al* (2011) recently suggested that the capsule region of *C. jejuni* strains within the same serotype is conserved. Contrary to this, a sequence comparison of the *C. jejuni* 01/51 CPS region with a HS4 *C. jejuni* strain did not show any homology between them. This suggests

that in *C. jejuni* 01/51 other outer surface structures (LOS or flagella) are being recognised in HS serotyping instead of the capsule. It would also be useful to compare the capsule sequence of *C. jejuni* 01/10 with other HS50 *C. jejuni* strains to validate the association between serotype and the capsule in this hyperinvasive strain.

To further investigate if the LOS is responsible for Penner serotype specificity mutants in a LOS gene (*cj1136*) and a capsule gene (JJD26997_1801) in *C. jejuni* 01/51 (Javed *et al.*, 2012) were serotyped (section 6.3.5). The LOS gene (*cj1136*) mutant changed the serotype to HS50 whereas the mutant in the capsule gene, with homology to JJD26997_1801, in *C. jejuni* 01/51 (Javed *et al.*, 2010) remained the same as the wild type strain which supports the fact that the capsule is not being recognised in serotyping and the LOS is the serodeterminant instead.

In summary, the phenotypic characterization of the *C. jejuni* strains in virulence related assays showed variation among strains with no grouping of the hyperinvasive *C. jejuni* as a distinct group. It therefore indicates that these strains share only the hyperinvasive phenotype which strengthens the approach to look for the underlying genetic components responsible for this uniquely shared phenotype. It also confirmed that the differences in the invasion potential of the *C. jejuni* strains was not due to the reduced growth and motility or inability to survive under atmospheric air and reactive oxygen stresses. The phylogenomic clustering based on the CGH data could not cluster the hyperinvasive *C. jejuni* strains as a separate group away from low invasive strains. In total, 67 genes were identified as present or missing from the hyperinvasive *C. jejuni* strains. Genes related to essential cellular functions were mainly identified along with some characterized virulence related loci. All the *C. jejuni* strains showed genome diversity in the LOS, capsule, RM and flagella biosynthesis regions. Two groups of genes were identified; with group 1 containing 9 genes present in all the hyperinvasive *C. jejuni* strains and group 2 with 6 genes which were highly divergent/absent from all the hyperinvasive *C. jejuni* strains. The genes were markers for hyperinvasion as a strong association was seen between their presence and absence and the hyperinvasive phenotype. The PSSH, identified 11 sequences which were variably distributed in the hyperinvasive *C. jejuni* and 9 non-PSSH tested low invasive *C. jejuni* strains. From these, 4/11 inserts were hyperinvasive *C. jejuni* specific but these were not present in all the hyperinvasive strains. The pyrosequencing/Illumina genome sequences of the hyperinvasive *C. jejuni* 01/10 and 01/51 did not identify any sequences unique to these

strains that were not present in the other sequenced *Campylobacter* strains except for a prophage in *C. jejuni* 01/51. A novel CPS region was characterised in *C. jejuni* 01/51 and 01/10 possessing loci with similarity to the *C. jejuni* subsp. *doylei* and *C. lari* capsule genes which is evident of interspecies homologous gene transfer in the capsule confirming the role of capsule in the hyperinvasive phenotype of these strains. It would be interesting to characterize the CPS regions in other four hyperinvasive *C. jejuni* strains to further validate the association between capsule and hyperinvasiveness. *C. jejuni* 01/51 was serotyped as HS4 but its capsule sequence was different from HS4 serotyped *C. jejuni* strains and the capsule gene mutant in *C. jejuni* 01/51 did not change its serotype suggesting that the capsule is not recognised during serotyping. However, the changed serotype of a LOS mutant in *C. jejuni* 01/51 suggests that LOS is the serodeterminant. This study represents a complex analysis of the genome of a phenotypically identical group of *C. jejuni* strains and has provided insight into the genetic basis for hyperinvasion.

What next in this project?

The transcriptional study of genes identified by CGH as present in all hyperinvasive *C. jejuni* and highly divergent from the majority of low invasive *C. jejuni* strains will show whether these genes are only expressed in the hyperinvasive group of strains and may account for the hyperinvasive phenotype of these strains. Since genes of different functional categories have been identified in the CGH and PSSH studies, it would be interesting to study the link/communication between different cellular networks and their role in hyperinvasion. The CGH and PSSH identified loci should be investigated further by mutagenesis and *in vitro* invasion assays. These genes are also the potential markers for studying the mechanism of *C. jejuni* uptake and survival into host cells.

The whole genome sequences of the hyperinvasive *C. jejuni* 01/10 and 01/51 should be compared in more detail with the sequenced *C. jejuni* strains other than *C. jejuni* RM1221. The remaining four hyperinvasive *C. jejuni* strains (01/35, 01/04, 01/41 and EX114) should be whole genome sequenced and the genome sequences of all six hyperinvasive *C. jejuni* strains should be compared. It would be interesting to observe if all the hyperinvasive *C. jejuni* strains share the mosaic pattern of interspecies gene distribution in their CPS regions. The comparison with CPS regions of some of the low invasive *C. jejuni* strains would also be a useful control as this will validate the role of capsule in hyperinvasiveness. The invasion profile of the majority of sequenced

C. jejuni strains is unknown. It would be useful to screen these strains in invasion assays to see if more strains possess the hyperinvasive phenotype.

Chapter Eight

REFERENCES

REFERENCES

- Abeyta, C., Trost, P.A., Bark, D.H., Hunt, J.M., Kaysnet, C.A., and Wekell, M.M. (1997). The use of bacterial membrane fractions for the detection of *Campylobacter* species in shellfish. *J.Rapid.Methods.Autom.Microbiol.* **5**:223-247.
- Agron, P.G., Macht, M., Radnedge, L., Skowronski, E.W., Miller, W., and Andersen, G.L. (2002). Use of subtractive hybridization for comprehensive surveys of prokaryotic genome differences. *FEMS Microbiol.Lett.* **211**:175-182.
- Ahmed, I.H., Manning, G., Wassenaar, T.M., Cawthraw, S., and Newell, D.G. (2002). Identification of genetic differences between two *Campylobacter jejuni* strains with different colonization potentials. *Microbiol.* **148**:1203-1212.
- Ahmed, M.U., Dunn, L., and Ivanova, E.P. (2012). Evaluation of current molecular approaches for genotyping of *Campylobacter jejuni* strains. *Foodborne Pathog.Dis.* **9**:375-385.
- Aklujkar, M., and Lovley, D.R. (2010). Interference with histidyl-tRNA synthetase by a CRISPR spacer sequence as a factor in the evolution of *Pelobacter carbinolicus*. *BMC Evol.Biol.* **10**(1):230.
- Alexeev, D., Alexeeva, M., Baxter, R.L., Campopiano, D.J., Webster, S.P., and Sawyer, L. (1998). The crystal structure of 8-amino-7-oxononanoate synthase: a bacterial PLP-dependent, acyl-CoA-condensing enzyme. *J.Mol.Biol.* **284**:401-419.
- Alm, R.A., Guerry, P., Power, M.E., and Trust, T.J. (1992). Variation in antigenicity and molecular weight of *Campylobacter coli* VC167 flagellin in different genetic backgrounds. *J.Bacteriol.* **174**:4230-4238.
- Al-Sayeqh, A.F., Loughlin, M.F., Dillon, E., Mellits, K.H., and Connerton, I.F. (2010). *Campylobacter jejuni* activates NF-kappaB independently of TLR2, TLR4, Nod1 and Nod2 receptors. *Microb.Pathog.* **49**:294-304.
- Altekruse, S.F., Swerdlow, D.L., and Stern, N.J. (1998). Microbial food borne pathogens. *Campylobacter jejuni*. *Vet.Clin.North.Am.Food.Animal.Prac.* **14**:31-40.
- Andersen-Nissen, E., Smith, K.D., Strobe, K.L., Barrett, S.L., Cookson, B.T., Logan, S.M., and Aderem, A. (2005). Evasion of Toll-like receptor 5 by flagellated bacteria. *Proc.Natl.Acad.Sci.U.S.A.* **102**:9247-9252.
- Anderson, J.B., Tanner, A.H., and Brodribb, A.J. (1986). Toxic megacolon due to *Campylobacter colitis*. *Int.J.Colorectal Dis.* **1**:58-59.
- Angiuoli, S.V., Dunning, H.J.C., Salzberg, S.L., and Tettelin, H. (2011). Improving pan-genome annotation using whole genome multiple alignment. *BMC Bioinformatics.* **30**:12:272.

- Anonymous. (2010). Preliminary FoodNet Data on the incidence of infection with pathogens transmitted commonly through food in 10 states (2009). *Weekly MMWR*. April 16, 2010/59(14); 418–422. Available at: http://www.foodconsumer.org/newsite/Nutrition/foodborne_illnesses_on_the_decline_15041008_53.html.
- Asadullah, K., Sterry, W., and Volk, H.D. (2003). Interleukin-10 therapy--review of a new approach. *Pharmacol.Rev.* **55**:241-269.
- Asakura, H., Yamasaki, M., Yamamoto, S., and Igimi, S. (2007). Deletion of *peb4* gene impairs cell adhesion and biofilm formation in *Campylobacter jejuni*. *FEMS Microbiol.Lett.* **275**:278-285.
- Asbury, A.K., and Cornblath, D.R. (1990). Assessment of current diagnostic criteria for Guillain-Barre syndrome. *Ann.Neurol.* **27 Suppl**: S21-4.
- Ashgar, S.S., Oldfield, N.J., Wooldridge, K.G., Jones, M.A., Irving, G.J., Turner, D.P., and Ala'Aldeen, D.A. (2007). CapA, an autotransporter protein of *Campylobacter jejuni*, mediates association with human epithelial cells and colonization of the chicken gut. *J.Bacteriol.* **189**:1856-1865.
- Aspinall, G.O., Fujimoto, S., McDonald, A.G., Pang, H., Kurjanczyk, L.A., and Penner, J.L. (1994). Lipopolysaccharides from *Campylobacter jejuni* associated with Guillain-Barre syndrome patients mimic human gangliosides in structure. *Infect.Immun.* **62**:2122-2125.
- Aspinall, G.O., McDonald, A.G., Pang, H., Kurjanczyk, L. A., Penner, J.L. (1993). An antigenic polysaccharide from *Campylobacter coli* serotype O:30. Structure of a teichoic acid-like antigenic polysaccharide with the lipopolysaccharide. *J.Biol.Chem.* **268**:18321-18329.
- Atabay, H.I., and Corry, J.E. (1998). The isolation and prevalence of campylobacters from dairy cattle using a variety of methods. *J.Appl.Microbiol.* **84**:733-740.
- Atack, J.M., Harvey, P., Jones, M.A., and Kelly, D.J. (2008). The *Campylobacter jejuni* thiol peroxidases Tpx and Bcp both contribute to aerotolerance and peroxide-mediated stress resistance but have distinct substrate specificities. *J.Bacteriol.* **190**:5279-5290.
- Babakhani, F.K., and Joens, L.A. (1993). Primary swine intestinal cells as a model for studying *Campylobacter jejuni* invasiveness. *Infect.Immun.* **61**:2723-2726.
- Bachtiar, B.M., Coloe, P.J., and Fry, B.N. (2007). Knockout mutagenesis of the *kpsE* gene of *Campylobacter jejuni* 81116 and its involvement in bacterium-host interactions. *FEMS Immunol. Med.Microbiol.* **49**:149-154.
- Bacon, D.J., Alm, R.A., Burr, D.H., Hu, L., Kopecko, D.J., Ewing, C.P., Trust, T.J., and Guerry, P. (2000). Involvement of a plasmid in virulence of *Campylobacter jejuni* 81-176. *Infect.Immun.* **68**:4384-4390.

- Bacon, D.J., Szymanski, C.M., Burr, D.H., Silver, R.P., Alm, R.A., and Guerry, P. (2001). A phase-variable capsule is involved in virulence of *Campylobacter jejuni* 81-176. *Mol.Microbiol.* **40**:769-777.
- Baek, K.T., Vegge, C.S., and Brondsted, L. (2011a). HtrA chaperone activity contributes to host cell binding in *Campylobacter jejuni*. *Gut Pathog.* **3**:13.
- Baek, K.T., Vegge, C.S., Skorko-Glonek, J., and Brondsted, L. (2011b). Different contributions of HtrA protease and chaperone activities to *Campylobacter jejuni* stress tolerance and physiology. *Appl.Environ.Microbiol.* **77**:57-66.
- Bahrani, B., Macfarlane, S., and Macfarlane, G.T. (2011). Induction of cytokine formation by human intestinal bacteria in gut epithelial cell lines. *J.Appl.Microbiol.* **110**:353-363.
- Baig, B.H., Wachsmuth, I.K., and Morris, G.K. (1986). Utilization of exogenous siderophores by *Campylobacter* species. *J.Clin.Microbiol.* **23**:431-433.
- Baillon, M.L., van Vliet, A.H., Ketley, J.M., Constantinidou, C., and Penn, C.W. (1999). An iron-regulated alkyl hydroperoxide reductase (AhpC) confers aerotolerance and oxidative stress resistance to the microaerophilic pathogen *Campylobacter jejuni*. *J.Bacteriol.* **181**:4798-4804.
- Baker, N.T. and Graham, L.L. (2010). *Campylobacter* fetus translocation across Caco-2 cell monolayers. *Microb.Pathog.* **49**:260-272.
- Balbontin, R., Rowley, G., Pucciarelli, M.G., Lopez-Garrido, J., Wormstone, Y., Lucchini, S., Garcia-Del Portillo, F., Hinton, J.C., and Casadesus, J. (2006). DNA adenine methylation regulates virulence gene expression in *Salmonella enterica* serovar Typhimurium. *J.Bacteriol.* **188**:8160-8168.
- Baqar, S., Applebee, L.A., Gilliland, T.C., Jr., Lee, L.H., Porter, C.K., and Guerry, P. (2008). Immunogenicity and protective efficacy of recombinant *Campylobacter jejuni* flagellum-secreted proteins in mice. *Infect.Immun.* **76**:3170-3175.
- Bardiau, M., Taminiau, B., Duprez, J.N., Labruzzo, S., and Mainil, J.G. (2012). Comparison between a bovine and a human enterohaemorrhagic *Escherichia coli* strain of serogroup O26 by suppressive subtractive hybridization reveals the presence of atypical factors in EHEC and EPEC strains. *FEMS Microbiol.Lett.* **330**:132-139.
- Barrero-Tobon, A.M., and Hendrixson, D.R. (2012). Identification and analysis of flagellar coexpressed determinants (Feds) of *Campylobacter jejuni* involved in colonization. *Mol.Microbiol.* **84**:352-369.
- Baserisalehi, M., and Bahador, N. (2011). Chemotactic behavior of *Campylobacter* spp. in function of different temperatures (37 degrees C and 42 degrees C). *Anaerobe.* **17**:459-462.

- Batchelor, R.A., Pearson, B.M., Friis, L.M., Guerry, P., and Wells, J.M. (2004). Nucleotide sequences and comparison of two large conjugative plasmids from different *Campylobacter* species. *Microbiology*. **150**:3507-3517.
- Bax, M., Kuijff, M.L., Heikema, A.P., van Rijs, W., Bruijns, S.C., Garcia-Vallejo, J.J., Crocker, P.R., Jacobs, B.C., van Vliet, S.J., and Van Kooyk, Y. (2011). *Campylobacter jejuni* lipooligosaccharides modulate dendritic cell-mediated T cell polarization in a sialic acid linkage-dependent manner. *Infect.Immun.* **79**:2681-2689.
- Baylis, C. L., MacPhee, S. A., Martin, K. W., Humphrey, T. J, and Betts, R. P. (2000). Comparison of three enrichment media for the isolation of *Campylobacter* spp. from foods. *J. Appl.Microbiol.* **89**:884-891.
- Beltinger, J., del Buono, J., Skelly, M.M., Thornley, J., Spiller, R.C., Stack, W.A., and Hawkey, C.J. (2008). Disruption of colonic barrier function and induction of mediator release by strains of *Campylobacter jejuni* that invade epithelial cells. *World J.Gastroenterol.* **14**:7345-7352.
- Berrang, M.E., Buhr, R.J., Cason, J.A., and Dickens, J.A. (2001). Broiler carcass contamination with *Campylobacter* from feces during de-feathering. *J.Food.Prot.* **64**:2063-2066.
- Best, E.L., Fox, A.J., Frost, J.A., and Bolton, F.J. (2004). Identification of *Campylobacter jejuni* Multilocus Sequence Type ST-21 Clonal Complex by Single-Nucleotide Polymorphism Analysis. *J.Clin.Micro.* **42**:2836-2839
- Biswas, D., Itoh, K., and Sasakawa, C. (2000). Uptake pathways of clinical and healthy animal isolates of *Campylobacter jejuni* into INT-407 cells. *FEMS Immunol.Med.Microbiol.* **29**:203-211.
- Blaser, M.J., Sazie, E., and Williams, L.P., Jr. (1987). The influence of immunity on raw milk--associated *Campylobacter* infection. *JAMA.* **257**:43-46.
- Bogdan, C., and Nathan, C. (1993). Modulation of macrophage function by transforming growth factor beta, interleukin-4, and interleukin-10. *Ann.N.Y.Acad.Sci.* **685**:713-739.
- Borrmann, E., Berndt, A., Hanel, I., and Kohler, H. (2007). *Campylobacter*-induced interleukin-8 responses in human intestinal epithelial cells and primary intestinal chick cells. *Vet.Microbiol.* **124**:115-124.
- Bou-Abdallah, F., Lewin, A.C., Le Brun, N.E., Moore, G.R., and Chasteen, N.D. (2002). Iron detoxification properties of Escherichia coli bacterioferritin. Attenuation of oxyradical chemistry. *J.Biol.Chem.* **277**:37064-37069.
- Bras, A.M., and Ketley, J.M. (1999). Transcellular translocation of *Campylobacter jejuni* across human polarised epithelial monolayers. *FEMS Microbiol.Lett.* **179**:209-215.

- Buelow, D.R., Christensen, J.E., Neal-McKinney, J.M., and Konkel, M.E. (2011). *Campylobacter jejuni* survival within human epithelial cells is enhanced by the secreted protein CiaI. *Mol.Microbiol.* **80**:1296-1312.
- Burrough, E.R., Sahin, O., Plummer, P.J., Zhang, Q., and Yaeger, M.J. (2009). Pathogenicity of an emergent, ovine abortifacient *Campylobacter jejuni* clone orally inoculated into pregnant guinea pigs. *Am.J.Vet.Res.* **70**:1269-1276.
- Canchaya, C., Fournous, G., and Brussow, H. (2004). The impact of prophages on bacterial chromosomes. *Mol.Microbiol.* **53**:9-18.
- Carrillo, C.D., Taboada, E., Nash, J.H., Lanthier, P., Kelly, J., Lau, P.C., Verhulp, R., Mykytczuk, O., Sy, J., Findlay, W.A., Amoako, K., Gomis, S., Willson, P., Austin, J.W., Potter, A., Babiuk, L., Allan, B., and Szymanski, C.M. (2004). Genome-wide expression analyses of *Campylobacter jejuni* NCTC11168 reveals coordinate regulation of motility and virulence by flhA. *J.Biol.Chem.* **279**:20327-20338.
- Carver, T.J., Rutherford, K.M., Berriman, M., Rajand Ream, M.A., Barrell, B.G., and Parkhill, J. (2005). ACT: the Artemis Comparison Tool. *Bioinformatics.* **21**:3422-3423.
- CAST. (1994). Foodborne Pathogens: Risk and Consequences. Task Force Report No.122. The Council for Agricultural Science and Technology, Iowa State University, Ames, IA.
- Cavalier-Smith, T. (2002). The neomuran origin of archaeobacteria, the negibacterial root of the universal tree and bacterial megaclassification. *Int .J.Sys.Evol.Microbiol.* **52**:7-76.
- Cawthraw, S.A., Lind, L., Kaijser, B., and Newell, D.G. (2000). Antibodies, directed towards *Campylobacter jejuni* antigens, in sera from poultry abattoir workers. *Clin.Exp.Immunol.* **122**:55-60.
- Cawthraw, S.A., and Newell, D.G. (2010). Investigation of the presence and protective effects of maternal antibodies against *Campylobacter jejuni* in chickens. *Avian Dis.* **54**:86-93.
- Champion, O.L., Gaunt, M.W., Gundogdu, O., Elmi, A., Witney, A.A., Hinds, J., Dorrell, N., and Wren, B.W. (2005). Comparative phylogenomics of the food-borne pathogen *Campylobacter jejuni* reveals genetic markers predictive of infection source. *Proc.Natl.Acad.Sci.U.S.A.* **102**:16043-16048.
- Champion, O.L., Al-Jaberi, S., Stabler, R.A., and Wren, B.W. (2008). Comparative genomic of *Campylobacter jejuni*. In *Campylobacter*. Nachamkin, I., Szymanski, C.M. and Blaser, M.J. (eds). Washington, DC: ASM Press, pp.63-95.
- Chantarapanont, W., Berrang, M., and Frank, J.F. (2003). Direct microscopic observation and viability determination of *Campylobacter jejuni* on chicken skin. *J.Food.Prot.* **66**:2222-2230.

- Chevreux, B., Wetter, T., and Suhai, S. (1999). Genome sequence assembly using trace signals and additional sequence information. In German Conference on Bioinformatics, pp. 45-56.
- Christensen, J.E., Pacheco, S.A., and Konkel, M.E. (2009). Identification of a *Campylobacter jejuni*-secreted protein required for maximal invasion of host cells. *Mol.Microbiol.* **73**:650-662.
- Clark, G.C., and Ng, Lai-King. (2008). Sequence variability of *Campylobacter* temperate bacteriophages. *BMC Microbiol.* **8**:49
- Cody, A.J., Maiden, M.J., and Dingle, K.E. (2009). Genetic diversity and stability of the *porA* allele as a genetic marker in human *Campylobacter* infection. *Microbiology.* **155**:4145-4154.
- Coker, A.O., Isokpehi, R.D., Thomas, B.N., Amisu, K.O., and Obi, C.L. (2002). Human *Campylobacteriosis* in developing countries. *Emerg.Infect.Dis.* **8**:237-244.
- Cole, S.P., Harwood, J., Lee, R., She, R., and Guiney, D.G. (2004a). Characterization of monospecies biofilm formation by *Helicobacter pylori*. *J.Bacteriol.* **186**:3124-3132.
- Cole, K., Donoghue, A.M., Blore, P.J., Holliman, J.S., Cox, N.A., Musgrove, M.T., and Donoghue, D.J. (2004b). Effects of aeration and storage temperature on *Campylobacter* concentrations in poultry semen. *Poult.Sci.* **83**:1734-1738.
- Cooper, K.K., Cooper, M.A., Zuccolo, A., Law, B., and Joens, L.A. (2011). Complete genome sequence of *Campylobacter jejuni* strain S3. *J.Bacteriol.* **193**:1491-1492.
- Corcoran, A.T., and Moran, A.P. (2007). Influence of growth conditions on diverse polysaccharide production by *Campylobacter jejuni*. *FEMS Immunol.Med.Microbiol.* **49**:124-132.
- Cornelius, A.J., Gilpin, B., Carter, P., Nicol, C., and On, S.L. (2010). Comparison of PCR binary typing (P-BIT), a new approach to epidemiological subtyping of *Campylobacter jejuni*, with serotyping, pulsed-field gel electrophoresis, and multilocus sequence typing methods. *Appl.Environ.Microbiol.* **76**:1533-1544.
- Corry, J.E.L., Post, D.E., Colin, P., and Laisney, M.J. (1995). Culture media for the isolation of campylobacters. *Int.J.FoodMicrobiol.* **26**:43-76.
- Corry, J.E.L., and Atabay, H.I. (2001). Poultry as a source of *Campylobacter* and related organisms. *J.Appl.Microbiol.* **90**:96S-114S.
- Dai, J., Wang, S., Guerlebeck, D., Laternus, C., Guenther, S., Shi, Z., Lu, C., and Ewers, C. (2010). Suppression subtractive hybridization identifies an autotransporter adhesin gene of *E. coli* IMT5155 specifically associated with avian pathogenic *Escherichia coli* (APEC). *BMC Microbiol.* **10**:236.

- Dale, J.W., and Park, S. (2004). *Molecular genetics of bacteria*. 4th ed. John Wiley & Sons Inc., Chichester, UK.
- Dasti, J.I., Tareen, A.M., Lugert, R., Zautner, A.E., and Gross, U. (2010). *Campylobacter jejuni*: a brief overview on pathogenicity-associated factors and disease-mediating mechanisms. *Int.J.Med.Microbiol.* **300**:205-211.
- Davis, L., and DiRita, V. (2008). Growth and laboratory maintenance of *Campylobacter jejuni*. *Curr.Protoc.Microbiol.* **Chapter 8**:Unit 8A.1.1-8A.1.7.
- Day, W.A., Jr., Sajecki, J.L., Pitts, T.M., and Joens, L.A. (2000). Role of catalase in *Campylobacter jejuni* intracellular survival. *Infect.Immun.* **68**:6337-6345.
- Debruyne, L., Gevers, D, and Vandamme, P. (2005). Taxonomy of the family Campylobacteraceae. In *Campylobacter*. Nachamkin, I and Blaser, M.J. (eds). 3rd Edn. Washington, DC: ASM, pp.3–27
- Debruyne, L., Samyn, E., De Brand T, E., Vand Enberg, O., Heyndrickx, M., and Vand Amme, P. (2008). Comparative performance of different PCR assays for the identification of *Campylobacter jejuni* and *Campylobacter coli*. *Res.Microbiol.* **159**:88-93.
- Dekeyser, P., Gossuin-Detrain, M., Butzler, J.P., and Sternon, J. (1972). Acute enteritis due to related vibrio: first positive stool cultures. *J.Infect.Dis.* **125**:390-392.
- De Melo, M.A., Gabbiani, G., and Pechere, J.C. (1989). Cellular events and intracellular survival of *Campylobacter jejuni* during infection of HEp-2 cells. *Infect.Immun.* **57**:2214-2222.
- de Zoete, M.R., Kestra, A.M., Roszczenko, P., and Van Putten, J.P. (2010). Activation of human and chicken toll-like receptors by *Campylobacter* spp. *Infect.Immun.* **78**:1229-1238.
- Dinant, S., Schurink, C.A., Deckers, J.W., and Severin, J.A. (2011). Aortic homograft endocarditis caused by *Campylobacter jejuni*. *J.Clin.Microbiol.* **49**:4016-4017.
- Dingle, K.E., Colles, F.M., Ure, R., Wagenaar, J.A., Duim, B., Bolton, F.J., Fox, A.J., Wareing, D.R., and Maiden, M.C. (2002). Molecular characterization of *Campylobacter jejuni* clones: a basis for epidemiologic investigation. *Emerg.Infect.Dis.* **8**:949-955.
- Dingle, K.E., Colles, F.M., Wareing, D.R., Ure, R., Fox, A.J., Bolton, F.E., Bootsma, H.J., Willems, R.J., Urwin, R., and Maiden, M.C. (2001). Multilocus sequence typing system for *Campylobacter jejuni*. *J.Clin.Microbiol.* **39**:14-23.
- Doig, P., Kinsella, N., Guerry, P., and Trust, T.J. (1996). Characterization of a post-translational modification of *Campylobacter* flagellin: identification of a sero-specific glycosyl moiety. *Mol.Microbiol.* **19**:379-387.

- Dolgilevich, S., Rafferty, B., Luchinskaya, D., and Kozarov, E. (2011). Genomic comparison of invasive and rare non-invasive strains reveals *Porphyromonas gingivalis* genetic polymorphisms. *J.Oral Microbiol.* **3**:10.3402/jom.v3i0.5764.
- Dorrell, N., Champion, O.L., and Wren, B.W. (2002). Microarray analysis of *Campylobacter jejuni*: to the guts of the problem! *Comp.Funct.Genomics.* **3**:338-341.
- Dorrell, N., Mangan, J.A., Laing, K.G., Hinds, J., Linton, D., Al-Ghusein, H., Barrell, B.G., Parkhill, J., Stoker, N.G., Karlyshev, A.V., Butcher, P.D., and Wren, B.W. (2001). Whole genome comparison of *Campylobacter jejuni* human isolates using a low-cost microarray reveals extensive genetic diversity. *Genome Res.* **11**:1706-1715.
- Doyle, L. (1944). A vibrio associated with swine dysentery. *Am.J.Vet.Res.* **5**:3-5
- Du, L.F., Li, Z.J., Tang, X.Y., Huang, J.Q., and Sun, W.B. (2008). Immunogenicity and immunoprotection of recombinant PEB1 in *Campylobacter-jejuni*-infected mice. *World J.Gastroenterol.* **14**:6244-6248.
- Dunne, A., and O'Neill, L.A. (2005). Adaptor usage and Toll-like receptor signaling specificity. *FEBS Lett.* **579**:3330-3335.
- Duong, T., and Konkel, M.E. (2009). Comparative studies of *Campylobacter jejuni* genomic diversity reveal the importance of core and dispensable genes in the biology of this enigmatic food-borne pathogen. *Curr.Opin.Biotechnol.* **20**:158-165.
- Eckmann, L. (2004). Innate immunity and mucosal bacterial interactions in the intestine. *Curr.Opin.Gastroenterol.* **20**:82-88.
- EFSA. (2007). The community summary report on trends and sources of zoonoses, zoonotic agents, antimicrobial resistance and foodborne outbreaks in the European Union in 2006. *EFSA J.* **130**:130-155.
- EFSA. (2009). The community summary report on trends and sources of zoonoses and zoonotic agents in the European Union in 2007. *EFSA J.* **223**:223-440.
- EFSA. (2010a). The community summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in the European Union in 2008. *EFSA J.* **8**:1496-1906.
- EFSA. (2010b). Scientific opinion on quantification of the risk posed by broiler meat to human campylobacteriosis in the EU. *EFSA J.* **8**:1437-1526.
- EFSA, (2012). The European Union Summary Report on antimicrobial resistance in zoonotic and indicator bacteria from humans, animals and food in 2010. *EFSA J.* **10** (3):2598.
- Eisen, M.B., Spellman, P.T., Brown, P.O., and Botstein, D. (1998). Cluster analysis and display of genome-wide expression patterns. *Proc.Natl.Acad.Sci.U.S.A.* **95**:14863-14868.

- El-Shibiny, A., Connerton, P.L., and Connerton, I.F. (2005). Enumeration and diversity of campylobacters and bacteriophages isolated during the rearing cycles of free-range and organic chickens. *Appl.Environ.Microbiol.* **71**:1259-1266.
- Elsinghorst, E.A. (1994). Measurement of invasion by gentamicin resistance. *Methods Enzymol.* **236**:405-420.
- Escherich, T. (1886). Beitrage zur Kenntniss der Darmbakterien. III. Ueber das Vorkommen von Vibrionen in Darmcanal und den Stuhlgangen der Sauglinge. (Articles adding to the knowledge of intestinal bacteria. III. On the existence of vibrios in the intestines and faeces of babies). *Münchener.Med.Wochenschrift.* **33**:815-817.
- Escobar, L., Perez-Martin, J., and De Lorenzo, V. (1999). Opening the iron box: transcriptional metalloregulation by the Fur protein. *J.Bacteriol.* **181**:6223-6229.
- Eucker, T.P., and Konkel, M.E. (2012). The cooperative action of bacterial fibronectin-binding proteins and secreted proteins promote maximal *Campylobacter jejuni* invasion of host cells by stimulating membrane ruffling. *Cell.Microbiol.* **14**:226-238.
- Everest, P.H., Goossens, H., Butzler, J.P., Lloyd, D., Knutton, S., Ketley, J.M., and Williams, P.H. (1992). Differentiated Caco-2 cells as a model for enteric invasion by *Campylobacter jejuni* and *C. coli*. *J.Med.Microbiol.* **37**:319-325.
- Ewing, C.P., Reishcheva, E., and Guerry, P. (2009). Functional characterization of flagellin glycosylation in *Campylobacter jejuni* 81-176. *J.Bacteriol.* **191**:7086-7093.
- Falkow, S. (1988). Molecular Koch's postulates applied to microbial pathogenicity. *Rev. Infect.Dis.* **10 (suppl. 2)**: S274-S276.
- Fauchere, J.L., Rosenau, A., Veron, M., Moyen, E.N., Richard, S., and Pfister, A. (1986). Association with HeLa cells of *Campylobacter jejuni* and *Campylobacter coli* isolated from human feces. *Infect.Immun.* **54**:283-287.
- Fearnley, C., Manning, G., Bagnall, M., Javed, M.A., Wassenaar, T.M., and Newell, D.G. (2008). Identification of hyperinvasive *Campylobacter jejuni* strains isolated from poultry and human clinical sources. *J.Med.Microbiol.* **57**:570-580.
- Fernandez, H., Giusti, G., and Bertoglio, J.C. (1995). Effect of the complement system on the sensitivity of *Campylobacter jejuni* and *Campylobacter coli* to human blood serum. *Braz.J.Med.Biol.Res.* **28**:227-229.
- Fernández, V., Villanueva, MP and García, A. (2008). Occurrence of campylobacter species in healthy well-nourished and malnourished children. *Braz.J.Med.Biol.* **39**:56-58.
- Fernando, U., Biswas, D., Allan, B., Willson, P., and Potter, A.A. (2007). Influence of *Campylobacter jejuni* fliA, rpoN and flgK genes on colonization of the chicken gut. *Int.J.Food Microbiol.* **118**:194-200.

- Ferrero, R. L., and Lee, A. (1988). Motility of *Campylobacter jejuni* in a viscous environment: comparison with conventional rod shaped bacteria. *Gen Microbiol.* **134**:53-59.
- Fitzgerald, C., and Nachamkin, I. (2007). *Campylobacter* and *Arcobacter*. In Manual of Clinical Microbiology. Murray, P.R. (eds). 9th ed. Washington DC: ASM Press, pp.933-946.
- Flanagan, R.C., Neal-McKinney, J.M., Dhillon, A.S., Miller, W.G., and Konkel, M.E. (2009). Examination of *Campylobacter jejuni* putative adhesins leads to the identification of a new protein, designated FlpA, required for chicken colonization. *Infect.Immun.* **77**:2399-2407.
- Flint, A., Sun, Y.Q., and Stintzi, A. (2012). Cj1386 is an ankyrin-containing protein involved in heme trafficking to catalase in *Campylobacter jejuni*. *J.Bacteriol.* **194**:334-345.
- Fouts, D.E., Mongodin, E.F., Mand Rell, R.E., Miller, W.G., Rasko, D.A., Ravel, J., Brinkac, L.M., DeBoy, R.T., Parker, C.T., Daugherty, S.C., Dodson, R.J., Durkin, A.S., Madupu, R., Sullivan, S.A., Shetty, J.U., Ayodeji, M.A., Shvartsbeyn, A., Schatz, M.C., Badger, J.H., Fraser, C.M., and Nelson, K.E. (2005). Major structural differences and novel potential virulence mechanisms from the genomes of multiple *Campylobacter* species. *PLoS Biol.* **3**:e15.
- Freist, W., Verhey, J.F., Ruhlmann, A., Gauss, D.H., and Arnez, J.G. (1999). Histidyl-tRNA synthetase. *Biol.Chem.* **380**:623-646.
- Friedman, C.R., Neimann, J., Wegener, H. C., and Tauxe, R.V. (2000). Epidemiology of *Campylobacter jejuni* infections in the United States and other industrialized nations. In *Campylobacter*. Nachamkin, I. and Blaser, M.J. (eds). 2nd edn. Washington, DC: ASM press, pp.121-138.
- Friis, C., Wassenaar, T.M., Javed, M.A., Snipen, L., Lagesen, K., Hallin, P.F., Newell, D.G., Toszeghy, M., Ridley, A., Manning, G., and Ussery, D.W. (2010). Genomic characterization of *Campylobacter jejuni* strain M1. *PLoS One.* **5**:e12253.
- Friidich, E., Biboy, J., Adams, C., Lee, J., Ellermeier, J., Giolda, L.D., Dirita, V.J., Girardin, S.E., Vollmer, W., and Gaynor, E.C. (2012). Peptidoglycan-Modifying Enzyme Pgp1 Is Required for Helical Cell Shape and Pathogenicity Traits in *Campylobacter jejuni*. *PLoS Pathog.* **8**:e1002602.
- Frost, J.A., Kramer, J.M., and Gilland Ers, S.A. (1999). Phage typing of *Campylobacter jejuni* and *Campylobacter coli* and its use as an adjunct to serotyping. *Epidemiol.Infect.* **123**:47-55.
- Gaasbeek, E.J., Wagenaar, J.A., Guilhabert, M.R., van Putten, J.P., Parker, C.T., and Van der Wal, F.J. (2010). Nucleases encoded by the integrated elements CJIE2

and CJIE4 inhibit natural transformation of *Campylobacter jejuni*. *J.Bacteriol.* **192**:936-941.

Gangaiah, D., Liu, Z., Arcos, J., Kassem, I.I., Sanad, Y., Torrelles, J.B., and Rajashekara, G. (2010). Polyphosphate kinase 2: a novel determinant of stress responses and pathogenesis in *Campylobacter jejuni*. *PLoS One.* **5**:e12142.

Gaynor, E.C., Cawthraw, S., Manning, G., MacKichan, J.K., Falkow, S., and Newell, D.G. (2004). The genome-sequenced variant of *Campylobacter jejuni* NCTC11168 and the original clonal clinical isolate differ markedly in colonization, gene expression, and virulence-associated phenotypes. *J.Bacteriol.* **186**:503-517.

Gaynor, E.C., Wells, D.H., MacKichan, J.K., and Falkow, S. (2005). The *Campylobacter jejuni* stringent response controls specific stress survival and virulence-associated phenotypes. *Mol.Microbiol.* **56**:8-27.

Ge, B., Girard, W., Zhao, S., Friedman, S., Gaines, S.A., and Meng, J. (2006). Genotyping of *Campylobacter* spp. from retail meats by pulsed-field gel electrophoresis and ribotyping. *J.Appl.Microbiol.* **100**:175-184.

Gerrish, R.S., Gill, A.L., Fowler, V.G., and Gill, S.R. (2010). Development of pooled suppression subtractive hybridization to analyze the pangenome of *Staphylococcus aureus*. *J.Microbiol.Methods.* **81**:56-60.

Gilpin, B., Cornelius, A., Robson, B., Boxall, N., Ferguson, A., Nicol, C., and Henderson, T. (2006). Application of pulsed-field gel electrophoresis to identify potential outbreaks of *Campylobacteriosis* in New Zealand. *J.Clin.Microbiol.* **44**:406-412.

Gilpin, B.J., Scholes, P., Robson, B., and Savill, M.G. (2008). The transmission of thermotolerant *Campylobacter* spp. to people living or working on dairy farms in New Zealand. *Zoonoses Public.Health.* **55**:352-360.

Glunder, G and Petermann, S. (1989). The occurrence and characterization of *Campylobacter* spp. in silver gulls (*Larus argentatus*), three-toed gulls (*Rissa tridactyla*) and house sparrows (*Passer domesticus*). *Zentralbl.Veterinarmed.B.* **36**:123-130.

Godreuil, S., Maslin, J., Morillon, M., Sagui, E., De Pina, J.J., and Martet, G. (2000). *Campylobacter lari* bacteremia (letter). *Presse Med.* **29**:1603.

Golden, N.J.and Acheson, D.W. (2002). Identification of motility and autoagglutination *Campylobacter jejuni* mutants by rand Om transposon mutagenesis. *Infect.Immun.* **70**:1761-1771.

Goodwin, C.S., McConnell, W., McCulloch, R.K., McCullough, C., Hill, R., Bronsdon, M.A., and Kasper, G. (1989). Cellular fatty acid composition of *Campylobacter pylori* from primates and ferrets compared with those of other *Campylobacters*. *J.Clin.Microbiol.* **27**:938-943.

- Goon, S., Ewing, C.P., Lorenzo, M., Pattarini, D., Majam, G., and Guerry, P. (2006). A sigma28-regulated nonflagella gene contributes to virulence of *Campylobacter jejuni* 81-176. *Infect.Immun.* **74**:769-772.
- Goon, S., Kelly, J.F., Logan, S.M., Ewing, C.P., and Guerry, P. (2003). Pseudaminic acid, the major modification on *Campylobacter* flagellin, is synthesized via the Cj1293 gene. *Mol.Microbiol.* **50**:659-671.
- Govoni, V., and Granieri, E. (2001). Epidemiology of the Guillain-Barre syndrome. *Curr.Opin.Neurol.* **14**:605-613.
- Grant, C.C., Konkel, M.E., Cieplak, W., Jr., and Tompkins, L.S. (1993). Role of flagella in adherence, internalization, and translocation of *Campylobacter jejuni* in nonpolarized and polarized epithelial cell cultures. *Infect.Immun.* **61**:1764-1771.
- Guerry, P. (2007). *Campylobacter* flagella: not just for motility. *Trends Microbiol.***15**:456-461.
- Guerry, P., Ewing, C.P., Hickey, T.E., Prendergast, M.M., and Moran, A.P. (2000). Sialylation of lipooligosaccharide cores affects immunogenicity and serum resistance of *Campylobacter jejuni*. *Infect.Immun.* **68**:6656-6662.
- Guerry, P., Ewing, C.P., Schirm, M., Lorenzo, M., Kelly, J., Pattarini, D., Majam, G., Thibault, P., and Logan, S. (2006). Changes in flagellin glycosylation affect *Campylobacter* autoagglutination and virulence. *Mol.Microbiol.* **60**:299-311.
- Guerry, P., Perez-Casal, J., Yao, R., McVeigh, A., and Trust, T.J. (1997). A genetic locus involved in iron utilization unique to some *Campylobacter* strains. *J.Bacteriol.* **179**:3997-4002.
- Guerry, P., Poly, F., Riddle, M., Maue, A.C., Chen, Y.H., and Monteiro, M.A. (2012). *Campylobacter* polysaccharide capsules: virulence and vaccines. *Front.Cell.Infect.Microbiol.* **2**:7.
- Guerry, P., and Szymanski, C.M. (2008). *Campylobacter* sugars sticking out. *Trends Microbiol.* **16**:428-435.
- Guerry, P., Szymanski, C.M., Prendergast, M.M., Hickey, T.E., Ewing, C.P., Pattarini, D.L., and Moran, A.P. (2002). Phase variation of *Campylobacter jejuni* 81-176 lipooligosaccharide affects ganglioside mimicry and invasiveness in vitro. *Infect.Immun.* **70**:787-793.
- Gundogdu, O., Bentley, S.D., Holden, M.T., Parkhill, J., Dorrell, N., and Wren, B.W. (2007). Re-annotation and re-analysis of the *Campylobacter jejuni* NCTC11168 genome sequence. *BMC Genomics.* **8**:162.
- Habib, I., Louwen, R., Uyttendaele, M., Houf, K., Vand Enberg, O., Nieuwenhuis, E.E., Miller, W.G., van Belkum, A., and De Zutter, L. (2009). Correlation between genotypic diversity, lipooligosaccharide gene locus class variation, and caco-2 cell

- invasion potential of *Campylobacter jejuni* isolates from chicken meat and humans: contribution to virulotyping. *Appl.Environ.Microbiol.* **75**:4277-4288.
- Hadden, R.D., and Gregson, N.A. (2001). Guillain--Barre syndrome and *Campylobacter jejuni* infection. *Symp.Ser.Soc.Appl.Microbiol.* **30**:145S-54S.
- Hänninen, M.L., Perko-Mäkelä, P., Pitkälä, and Rautelin, H. (2000). A three-year study of *Campylobacter jejuni* genotypes in humans with domestically acquired infections and in chicken samples from the Helsinki area. *J.Clini.Microbiol.* **38**:1998-2000.
- Hannu, T., Kauppi, M., Tuomala, M., Laaksonen, I., Klemets, P., and Kuusi, M. (2004). Reactive arthritis following an outbreak of *Campylobacter jejuni* infection. *J.Rheumatol.* **31**:528-530.
- Hantke, K. (1981). Regulation of ferric iron transport in *Escherichia coli* K12: isolation of a constitutive mutant. *Mol.Gen.Genet.* **182**:288-292.
- Harvey, P., and Leach, S. (1998). Analysis of coccal cell formation by *Campylobacter jejuni* using continuous culture techniques, and the importance of oxidative stress. *J.Appl.Microbiol.* **85**:398-404.
- Havelaar, A.H., Manges, M.J., de Koeijer, A.A., Bogaardt, M.J., Evers, E.G., Jacobs-Reitsma, W.F., van Pelt, W., Wagenaar, J.A., de Wit, G.A., van der Zee, H., and Nauta, M.J. (2007). Effectiveness and efficiency of controlling *Campylobacter* on broiler chicken meat. *Risk Anal.* **27**:831-844.
- Havelaar, A.H., Nauta, M.J., Manges, M. J.J., de Koeijer, A.G., Bogaardt, M. J., Evers, E.G., Jacobs-Reitsma, W.F., van Pelt, W., Wagenaar, J.A., de Wit, G.A., and van der Zee, H. (2005). Costs and Benefits of Controlling *Campylobacter* in the Netherlands; Integrating Risk Analysis, Epidemiology and Economics. RIVM report250911009/2005. Available at: <http://www.rivm.nl/bibliotheek/rapporten/250911009.pdf>.
- Heikema, A.P., Bergman, M.P., Richards, H., Crocker, P.R., Gilbert, M., Samsom, J.N., van Wamel, W.J., Endtz, H.P., and Van Belkum, A. (2010). Characterization of the specific interaction between sialoadhesin and sialylated *Campylobacter jejuni* lipooligosaccharides. *Infect.Immun.* **78**:3237-3246.
- Hendrixson, D.R. and DiRita, V.J. (2004). Identification of *Campylobacter jejuni* genes involved in commensal colonization of the chick gastrointestinal tract. *Mol.Microbiol.* **52**:471-484.
- Hepworth, P.J., Leatherbarrow, H., Hart, C.A., and Winstanley, C. (2007). Use of suppression subtractive hybridisation to extend our knowledge of genome diversity in *Campylobacter jejuni*. *BMC Genomics.* **8**:110.
- Hepworth, P.J., Ashelford, K.E., Hinds, J., Gould, K.A., Witney, A.A., Williams, N.J., Leatherbarrow, H., French, N.P., Birtles, R.J., Mendonca, C., Dorrell, N, Wren B.W.,

- Wigley, P., Hall, N., and Winstanley, C. (2011). Genomic variations define divergence of water/wildlife-associated *Campylobacter jejuni* niche specialists from common clonal complexes. *Environ Microbiol.* **6**:1549-60.
- Hickey, T.E., Baqar, S., Bourgeois, A.L., Ewing, C.P., and Guerry, P. (1999). *Campylobacter jejuni*-stimulated secretion of interleukin-8 by INT407 cells. *Infect.Immun.* **67**:88-93.
- Hickey, T.E., Majam, G., and Guerry, P. (2005). Intracellular survival of *Campylobacter jejuni* in human monocytic cells and induction of apoptotic death by cytolethal distending toxin. *Infect.Immun.* **73**:5194-5197.
- Hickey, T.E., McVeigh, A.L., Scott, D.A., Michielutti, R.E., Bixby, A., Carroll, S.A., Bourgeois, A.L., and Guerry, P. (2000). *Campylobacter jejuni* cytolethal distending toxin mediates release of interleukin-8 from intestinal epithelial cells. *Infect.Immun.* **68**:6535-6541.
- Hofreuter, D., Novik, V., and Galan, J.E. (2008). Metabolic diversity in *Campylobacter jejuni* enhances specific tissue colonization. *Cell.Host Microbe.* **4**:425-433.
- Hofreuter, D., Tsai, J., Watson, R.O., Novik, V., Altman, B., Benitez, M., Clark, C., Perbost, C., Jarvie, T., Du, L., and Galan, J.E. (2006). Unique features of a highly pathogenic *Campylobacter jejuni* strain. *Infect.Immun.* **74**:4694-4707.
- Holden, K.M., Gilbert, M., Coloe, P.J., Li, J., and Fry, B.N. (2012). The role of WlaRG, WlaTB and WlaTC in lipooligosaccharide synthesis by *Campylobacter jejuni* strain 81116. *Microb.Pathog.* **52**:344-352.
- Holmes, K., Mulholland, F., Pearson, B.M., Pin, C., McNicholl-Kennedy, J., Ketley, J.M., and Wells, J.M. (2005). *Campylobacter jejuni* gene expression in response to iron limitation and the role of Fur. *Microbiology.* **151**:243-257.
- Hoosain, N. and Lastovica, A.J. (2009). An evaluation of the Oxoid Biochemical Identification System Campy rapid screening test for *Campylobacteraceae* and *Helicobacter* spp. *Lett.Appl.Microbiol.* **48**:675-679.
- Hopkins, K.L., Desai, M., Frost, J.A., Stanley, J., and Logan, J.M. (2004). Fluorescent amplified fragment length polymorphism genotyping of *Campylobacter jejuni* and *Campylobacter coli* strains and its relationship with host specificity, serotyping, and phage typing. *J.Clin.Microbiol.* **42**:229-235.
- Howard, S.L., Jagannathan, A., Soo, E.C., Hui, J.P., Aubry, A.J., Ahmed, I., Karlyshev, A., Kelly, J.F., Jones, M.A., Stevens, M.P., Logan, S.M., and Wren, B.W. (2009). *Campylobacter jejuni* glycosylation island important in cell charge, legionaminic acid biosynthesis, and colonization of chickens. *Infect.Immun.* **77**:2544-2556.

- Hu, L., Bray, M.D., Osorio, M., and Kopecko, D.J. (2006a). *Campylobacter jejuni* induces maturation and cytokine production in human dendritic cells. *Infect.Immun.* **74**:2697-2705.
- Hu, L., McDaniel, J.P., and Kopecko, D.J. (2006b). Signal transduction events involved in human epithelial cell invasion by *Campylobacter jejuni* 81-176. *Microb.Pathog.* **40**:91-100.
- Hu, L., Tall, B.D., Curtis, S.K., and Kopecko, D.J. (2008). Enhanced microscopic definition of *Campylobacter jejuni* 81-176 adherence to, invasion of, translocation across, and exocytosis from polarized human intestinal Caco-2 cells. *Infect.Immun.* **76**:5294-5304.
- Hu, L., and Kopecko, D.J. (2008). Cell biology of human host cell entry by *Campylobacter jejuni*. In *Campylobacter*. Nachamkin, I., Szymanski, C.M. and Blaser, M.J. (eds). Washington, DC: ASM Press, pp.297-313.
- Hu, L., and Kopecko, D.J. (1999). *Campylobacter jejuni* 81-176 associates with microtubules and dynein during invasion of human intestinal cells. *Infect.Immun.* **67**:4171-4182.
- Hugdahl, M.B., Beery, J.T., and Doyle, M.P. (1988). Chemotactic behavior of *Campylobacter jejuni*. *Infect.Immun.* **56**:1560-1566.
- Hughes, R.A. and Cornblath, D.R. (2005). Guillain-Barre syndrome. *Lancet.* **366**:1653-1666.
- Huizinga, R., Easton, A.S., Donachie, A.M., Guthrie, J., van Rijs, W., Heikema, A., Boon, L., Samsom, J.N., Jacobs, B.C., Willison, H.J., and Goodyear, C.S. (2012). Sialylation of *Campylobacter jejuni* lipo-oligosaccharides: impact on phagocytosis and cytokine production in mice. *PLoS One.* **7**:e34416.
- Humphrey, T., O'Brien, S., and Madsen, M. (2007). *Campylobacters* as zoonotic pathogens: a food production perspective. *Int.J.Food Microbiol.* **117**:237-257.
- Iimura, M., Gallo, R.L., Hase, K., Miyamoto, Y., Eckmann, L., and Kagnoff, M.F. (2005). Cathelicidin mediates innate intestinal defense against colonization with epithelial adherent bacterial pathogens. *J.Immunol.* **174**:4901-4907.
- Ishikawa, T., Mizunoe, Y., Kawabata, S., Takade, A., Harada, M., Wai, S.N., and Yoshida, S. (2003). The iron-binding protein Dps confers hydrogen peroxide stress resistance to *Campylobacter jejuni*. *J.Bacteriol.* **185**:1010-1017.
- ISO. (2006a). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for Detection and Enumeration of *Campylobacter* spp. Part 1: Detection Method. Geneva: International Organization for Standardization. [ISO 10272-1:2006].

- ISO. (2006b). Microbiology of Food and Animal Feeding Stuffs - Horizontal Method for Detection and Enumeration of *Campylobacter* spp. Part 2: Colony Count Technique. Geneva: International Organization for Standardization. [ISO/TS 10272- 2:2006].
- Jacobsen, A., Hendriksen, R.S., Aaresturp, F.M., Ussery, D.W., and Friis, C. (2011). The *Salmonella enterica* pan-genome. *Microb.Ecol.* **62**:487-504.
- Jagannathan, A., and Penn, C. (2005). Motility. In *Campylobacter*. *Mol.Cell.Biol.* Ketley J.M. and Konkel, M.E. (eds). Norfolk, Horizon Bioscience, pp.331-347.
- Jain, D., Prasad, K.N., Sinha, S., and Husain, N. (2008). Differences in virulence attributes between cytolethal distending toxin positive and negative *Campylobacter jejuni* strains. *J. Med. Microbiology.* **57**: 267-272
- Jamieson, D.J. (1998). Oxidative stress responses of the yeast *Saccharomyces cerevisiae*. *Yeast.* **14**:1511-1527.
- Javed, M.A. (2009). Hyperinvasiveness in the major food-borne pathogen *Campylobacter jejuni*. PhD thesis. Nottingham Trent University
- Javed, M.A., Cawthraw, S.A., Baig, A., Li, J., McNally, A., Oldfield, N.J., Newell, D.G., and Manning, G. (2012). Cj1136 is required for lipooligosaccharide biosynthesis, hyperinvasion, and chick colonization by *Campylobacter jejuni*. *Infect.Immun.* **80**:2361-2370.
- Javed, M.A., Grant, A.J., Bagnall, M.C., Maskell, D.J., Newell, D.G., and Manning, G. (2010). Transposon mutagenesis in a hyper-invasive clinical isolate of *Campylobacter jejuni* reveals a number of genes with potential roles in invasion. *Microbiol.* **156**:1134-1143.
- Jeon, B., Muraoka, W., Sahin, O., and Zhang, Q. (2008). Role of Cj1211 in natural transformation and transfer of antibiotic resistance determinants in *Campylobacter jejuni*. *Antimicrob. Agents Chemother.* **52**:2699-2708.
- Jervis, A.J., Butler, J.A., Lawson, A.J., Langdon, R., Wren, B.W., and Linton, D. (2012). Characterization of the structurally diverse N-linked glycans of *Campylobacter* species. *J.Bacteriol.* **194**:2355-2362.
- Jin, S., Joe, A., Lynett, J., Hani, E.K., Sherman, P., and Chan, V.L. (2001). JlpA, a novel surface-exposed lipoprotein specific to *Campylobacter jejuni*, mediates adherence to host epithelial cells. *Mol.Microbiol.* **39**:1225-1236.
- John, A., Connerton, P.L., Cummings, N., and Connerton, I.F. (2011). Profound differences in the transcriptome of *Campylobacter jejuni* grown in two different, widely used, microaerobic atmospheres. *Res.Microbiol.* **162**:410-418.
- Johnsen, G., Kruse, H., and Hofshagen, M. (2007). Genotyping of thermotolerant *Campylobacter* from poultry slaughterhouse by amplified fragment length polymorphism. *J.Appl.Microbiol.* **103**:271-279.

- Johnson, D.C., Dean, D.R., Smith, A.D., and Johnson, M.K. (2005). Structure, function, and formation of biological iron-sulfur clusters. *Annu.Rev.Biochem.* **74**:247-281.
- Jones, D.M., Sutcliffe, E.M., Rios, R., Fox, A.J., and Curry, A. (1993). *Campylobacter jejuni* adapts to aerobic metabolism in the environment. *J.Med.Microbiol.* **38**:145-150.
- Joshua, G.W., Guthrie-Irons, C., Karlyshev, A.V., and Wren, B.W. (2006). Biofilm formation in *Campylobacter jejuni*. *Microbiology.* **152**:387-396.
- Kaakoush, N.O., Miller, W.G., De Reuse, H., and Mendz, G.L. (2007). Oxygen requirement and tolerance of *Campylobacter jejuni*. *Res.Microbiol.* **158**:644-650.
- Kakuda, T. and DiRita, V.J. (2006). Cj1496c encodes a *Campylobacter jejuni* glycoprotein that influences invasion of human epithelial cells and colonization of the chick gastrointestinal tract. *Infect.Immun.* **74**:4715-4723.
- Kale, A., Phansopa, C., Suwannachart, C., Craven, C.J., Rafferty, J.B., and Kelly, D.J. (2011). The virulence factor PEB4 (Cj0596) and the periplasmic protein Cj1289 are two structurally related SurA-like chaperones in the human pathogen *Campylobacter jejuni*. *J.Biol.Chem.* **286**:21254-21265.
- Kalmokoff, M., Lanthier, P., Tremblay, T.L., Foss, M., Lau, P.C., Sand Ers, G., Austin, J., Kelly, J., and Szymanski, C.M. (2006). Proteomic analysis of *Campylobacter jejuni* 11168 biofilms reveals a role for the motility complex in biofilm formation. *J.Bacteriol.* **188**:4312-4320.
- Kanipes, M.I., Holder, L.C., Corcoran, A.T., Moran, A.P., and Guerry, P. (2004). A deep-rough mutant of *Campylobacter jejuni* 81-176 is noninvasive for intestinal epithelial cells. *Infect.Immun.* **72**:2452-2455.
- Kanipes, M.I., Akelatis, A., Guerry, P., Monteiro, M.A. (2006). Mutation of waaC encoding heptosyl transferase I in *Campylobacter jejuni* 81-176 affects the structure of both lipooligosaccharide and capsular carbohydrate. *J.Bacteriol.* **188**:3273-3279. doi: 10.1128/JB.188.9.3273-3279.2006.
- Kanungpean, D., Kakuda, T., and Takai, S. (2011). Participation of CheR and CheB in the chemosensory response of *Campylobacter jejuni*. *Microbiology.* **157**:1279-1289.
- Karlyshev, A.V., Everest, P., Linton, D., Cawthraw, S., Newell, D.G., and Wren, B.W. (2004). The *Campylobacter jejuni* general glycosylation system is important for attachment to human epithelial cells and in the colonization of chicks. *Microbiology.* **150**:1957-1964.
- Karlyshev, A.V., Champion, O.L., Churcher, C., Brisson, J.R., Jarrell, H.C., Gilbert, M., Brochu, D., St Michael, F., Li, J., Wakarchuk, W.W., Goodhead, I., Sand Ers, M., Stevens, K., White, B., Parkhill, J., Wren, B.W., and Szymanski, C.M. (2005a). Analysis of *Campylobacter jejuni* capsular loci reveals multiple

- mechanisms for the generation of structural diversity and the ability to form complex heptoses. *Mol.Microbiol.* **55**:90-103.
- Karlyshev, A.V., Ketley, J.M., and Wren, B.W. (2005b). The *Campylobacter jejuni* glycome. *FEMS Microbiol.Rev.* **29**:377-390.
- Karlyshev, A.V., Linton, D., Gregson, N.A., Lastovica, A.J., and Wren, B.W. (2000). Genetic and biochemical evidence of a *Campylobacter jejuni* capsular polysaccharide that accounts for Penner serotype specificity. *Mol.Microbiol.* **35**:529-541.
- Karlyshev, A.V., McCrossan, M.V, and Wren, B.W. (2001). Demonstration of polysaccharide capsule in *Campylobacter jejuni* using electron microscopy. *Infection and Immunity.* **69**:5921-5924.
- Kelana, L.C.and Griffiths, M.W. (2003). Growth of autbioluminescent *Campylobacter jejuni* in response to various environmental conditions. *J.Food Prot.* **66**:1190-1197.
- Kelly, A.F., Park, S.F., Bovill, R., and Mackey, B.M. (2001). Survival of *Campylobacter jejuni* during stationary phase: evidence for the absence of a phenotypic stationary-phase response. *Appl.Environ.Microbiol.* **67**:2248-2254.
- Kelly, J., Jarrell, H., Millar, L., Tessier, L., Fiori, L.M., Lau, P.C., Allan, B., and Szymanski, C.M. (2006). Biosynthesis of the N-linked glycan in *Campylobacter jejuni* and addition onto protein through block transfer. *J.Bacteriol.* **188**:2427-2434.
- Kelly, D.J. (2005). Metabolism, electron transport and bioenergetics of *Campylobacter jejuni*: implications of understanding life in gut and survival in the environment. In *Campylobacter: molecular and cellular biology*. Ketley, J.M. and Konkel, M.E. (eds). Norfolk, UK: Horizon Biosciences, pp.275-292.
- Keo, T., Collins, J., Kunwar, P., Blaser, M.J., and Iovine, N.M. (2011). *Campylobacter* capsule and lipooligosaccharide confer resistance to serum and cationic antimicrobials. *Virulence.* **2**:30-40.
- Kervella, M., Pages, J.M., Pei, Z., Grollier, G., Blaser, M.J., and Fauchere, J.L. (1993). Isolation and characterization of two *Campylobacter* glycine-extracted proteins that bind to HeLa cell membranes. *Infect.Immun.* **61**:3440-3448.
- Ketley, J.M. (1997). Pathogenesis of enteric infection by *Campylobacter*. *Microbiology.* **143 (Pt 1)**:5-21.
- Kim, C.C., Joyce, E.A., Chan, K., and Falkow, S. (2002). Improved analytical methods for microarray-based genome-composition analysis. *Genome Biol.* **3**:RESEARCH0065.
- Kim, J.S., Li, J., Barnes, I.H., Baltzegar, D.A., Pajaniappan, M., Cullen, T.W., Trent, M.S., Burns, C.M., and Thompson, S.A. (2008). Role of the *Campylobacter jejuni* Cj1461 DNA methyltransferase in regulating virulence characteristics. *J.Bacteriol.* **190**:6524-6529.

- King, E. (1957). Human infections with *Vibrio fetus* and a closely related vibrio. *J.Infect.Dis.* **101**:119-128.
- Ko, M., and Park, C. (2000). H-NS-Dependent regulation of flagellar synthesis is mediated by a LysR family protein. *J.Bacteriol.* **182**:4670-4672.
- Koenraad, P.M., Jacobs-Reitsma, W.F., van der Laan, T., Beumer, R.R., and Rombouts, F.M. (1995). Antibiotic susceptibility of *Campylobacter* isolates from sewage and poultry abattoir drain water. *Epidemiol.Infect.* **115**:475-483.
- Konkel, M.E., Babakhani, F., and Joens, L.A. (1990). Invasion-related antigens of *Campylobacter jejuni*. *J.Infect.Dis.* **162**:888-895.
- Konkel, M.E., Garvis, S.G., Tipton, S.L., And Erson, D.E., Jr., and Cieplak, W., Jr. (1997). Identification and molecular cloning of a gene encoding a fibronectin-binding protein (CadF) from *Campylobacter jejuni*. *Mol.Microbiol.* **24**:953-963.
- Konkel, M.E., Mead, D.J., Hayes, S.F., and Cieplak, W., Jr. (1992a). Translocation of *Campylobacter jejuni* across human polarized epithelial cell monolayer cultures. *J.Infect.Dis.* **166**:308-315.
- Konkel, M.E., Hayes, S.F., Joens, L.A., and Cieplak, W., Jr. (1992b). Characteristics of the internalization and intracellular survival of *Campylobacter jejuni* in human epithelial cell cultures. *Microb.Pathog.* **13**:357-370.
- Konkel, M.E., and Joens, L.A. (1989). Adhesion to and invasion of HEp-2 cells by *Campylobacter* spp. *Infect.Immun.* **57**:2984-2990.
- Konkel, M.E., Klena, J.D., Rivera-Amill, V., Monteville, M.R., Biswas, D., Raphael, B., and Mickelson, J. (2004). Secretion of virulence proteins from *Campylobacter jejuni* is dependent on a functional flagellar export apparatus. *J.Bacteriol.* **186**:3296-3303.
- Konkel, M.E., Monteville, M.R., Rivera-Amill, V., and Joens, L.A. (2001). The pathogenesis of *Campylobacter jejuni*-mediated enteritis. *Curr.Issues Intest Microbiol.* **2**:55-71.
- Korinthenberg, R.and Monting, J.S. (1996). Natural history and treatment effects in Guillain-Barre syndrome: a multicentre study. *Arch.Dis.Child.* **74**:281-287.
- Kowarik, M., Young, N.M., Numao, S., Schulz, B.L., Hug, I., Callewaert, N., Mills, D.C., Watson, D.C., Hernand Ez, M., Kelly, J.F., Wacker, M., and Aebi, M. (2006). Definition of the bacterial N-glycosylation site consensus sequence. *EMBO J.* **25**:1957-1966.
- Krause-Gruszczynska, M., van Alphen, L.B., Oyarzabal, O.A., Alter, T., Hanel, I., Schliephake, A., Konig, W., van Putten, J.P., Konkel, M.E., and Backert, S. (2007). Expression patterns and role of the CadF protein in *Campylobacter jejuni* and *Campylobacter coli*. *FEMS Microbiol.Lett.* **274**:9-16.

- Kriek, M., Martins, F., Leonardi, R., Fairhurst, S.A., Lowe, D.J., and Roach, P.L. (2007). Thiazole synthase from *Escherichia coli*: an investigation of the substrates and purified proteins required for activity in vitro. *J.Biol.Chem.* **282**:17413-17423.
- Kucerova, E., Clifton, S.W., Xia, X.Q., Long, F., Porwollik, S., Fulton, L., Fronick, C., Minx, P., Kyung, K., Warren, W., Fulton, R., Feng, D., Wollam, A., Shah, N., Bhonagiri, V., Nash, W.E., Hallsworth-Pepin, K., Wilson, R.K., McClelland, M., and Forsythe, S.J. (2010). Genome sequence of *Cronobacter sakazakii* BAA-894 and comparative genomic hybridization analysis with other *Cronobacter* species. *PLoS One.* **5**:e9556.
- Kuijf, M.L., Samsom, J.N., van Rijs, W., Bax, M., Huizinga, R., Heikema, A.P., van Doorn, P.A., van Belkum, A., van Kooyk, Y., Burgers, P.C., Luider, T.M., Endtz, H.P., Nieuwenhuis, E.E., and Jacobs, B.C. (2010). TLR4-mediated sensing of *Campylobacter jejuni* by dendritic cells is determined by sialylation. *J.Immunol.* **185**:748-755.
- Lang, P., Lefebure, T., Wang, W., Pavinski Bitar, P., Meinersmann, R.J., Kaya, K., and Stanhope, M.J. (2010). Expand Ed multilocus sequence typing and comparative genomic hybridization of *Campylobacter coli* isolates from multiple hosts. *Appl.EnvIRON.Microbiol.* **76**:1913-1925.
- Lara-Tejero, M., and Galan, J.E. (2000). A bacterial toxin that controls cell cycle progression as a deoxyribonuclease I-like protein. *Science.* **290**:354-357.
- Larsen, J.C., Szymanski, C., and Guerry, P. (2004). N-linked protein glycosylation is required for full competence in *Campylobacter jejuni* 81-176. *J.Bacteriol.* **186**:6508-6514.
- Lastovica, A.J. (2006). Isolation and clinical relevance of emergent *Campylobacter* spp. Proceedings of “*Emerging Campylobacter spp. in the food chain, CAMPYCHECK*” held at Croke Park Conference Centre, Dublin. February 8th 2006.
- Lee, A., O'Rourke, J.L., Barrington, P.J., and Trust, T.J. (1986). Mucus colonization as a determinant of pathogenicity in intestinal infection by *Campylobacter jejuni*: a mouse cecal model. *Infect.Immun.* **51**:536-546.
- Lee, R.B., Hassane, D.C., Cottle, D.L., and Pickett, C.L. (2003). Interactions of *Campylobacter jejuni* cytolethal distending toxin subunits CdtA and CdtC with HeLa cells. *Infect.Immun.* **71**:4883-4890.
- Lefebure, T., Bitar, P.D., Suzuki, H., and Stanhope, M.J. (2010). Evolutionary dynamics of complete *Campylobacter* pan-genomes and the bacterial species concept. *Genome Biol.Evol.* **2**:646-655.
- Lehrer, R.I. (2004). Primate defensins. *Nat.Rev.Microbiol.* **2**:727-738.

- Lehtola, M.J., Pitkanen, T., Miebach, L., and Miettinen, I.T. (2006). Survival of *Campylobacter jejuni* in potable water biofilms: a comparative study with different detection methods. *WaterSci.Technol.* **54**:57-61.
- Leonard, E.E., 2nd., Tompkins, L.S., Falkow, S., and Nachamkin, I. (2004). Comparison of *Campylobacter jejuni* isolates implicated in Guillain-Barre syndrome and strains that cause enteritis by a DNA microarray. *Infect.Immun.* **72**:1199-1203.
- Leon-Kempis Mdel, R., Guccione, E., Mulholland, F., Williamson, M.P., and Kelly, D.J. (2006). The *Campylobacter jejuni* PEB1a adhesin is an aspartate/glutamate-binding protein of an ABC transporter essential for microaerobic growth on dicarboxylic amino acids. *Mol.Microbiol.* **60**:1262-1275.
- Levin, R.E. (2007). *Campylobacter jejuni*: a review of its characteristics, pathogenicity, ecology, distribution, subspecies characterization and molecular methods of detection. *Food Biotechnol.* **21**:271-347.
- Lindsay, J.O., and Hodgson, H.J. (2001). Review article: the immunoregulatory cytokine interleukin-10--a therapy for Crohn's disease? *Aliment.Pharmacol.Ther.* **15**:1709-1716.
- Linton, D., Allan, E., Karlyshev, A.V., Cronshaw, A.D., and Wren, B.W. (2002). Identification of N-acetylgalactosamine-containing glycoproteins PEB3 and CgpA in *Campylobacter jejuni*. *Mol.Microbiol.* **43**:497-508.
- Lior, H., Woodward, D.L., Edgar, J.A., Laroche, L.J., and Gill, P. (1982). Serotyping of *Campylobacter jejuni* by slide agglutination based on heat-labile antigenic factors. *J.Clin.Microbiol.* **15**:761-768.
- Lippert, E., Karrasch, T., Sun, X., Allard, B., Herfarth, H.H., Threadgill, D., and Jobin, C. (2009). Gnotobiotic IL-10; NF-kappaB mice develop rapid and severe colitis following *Campylobacter jejuni* infection. *PLoS One.* **4**:e7413.
- Logan, S.M., Trust, T.J., and Guerry, P. (1989). Evidence for posttranslational modification and gene duplication of *Campylobacter* flagellin. *J.Bacteriol.* **171**:3031-3038.
- Louwen, R., Heikema, A., van Belkum, A., Ott, A., Gilbert, M., Ang, W., Endtz, H.P., Bergman, M.P., and Nieuwenhuis, E.E. (2008). The sialylated lipooligosaccharide outer core in *Campylobacter jejuni* is an important determinant for epithelial cell invasion. *Infect.Immun.* **76**:4431-4438.
- Luechtefeld, N.W., Reller, L.B., Blaser, M.J., and Wang, W.L. (1982). Comparison of atmospheres of incubation for primary isolation of *Campylobacter fetus* subsp. *jejuni* from animal specimens: 5% oxygen versus cand Le jar. *J.Clin.Microbiol.* **15**:53-57.
- Lux, R., and Shi, W. (2004). Chemotaxis-guided movements in bacteria. *Crit.Rev.Oral Biol.Med.* **15**:207-220.

- MacCallum, A.J., Harris, D., Haddock, G., and Everest, P.H. (2006). *Campylobacter jejuni*-infected human epithelial cell lines vary in their ability to secrete interleukin-8 compared to in vitro-infected primary human intestinal tissue. *Microbiology*. **152**:3661-3665.
- Malik-Kale, P., Raphael, B.H., Parker, C.T., Joens, L.A., Klena, J.D., Quinones, B., Keech, A.M., and Konkel, M.E. (2007). Characterization of genetically matched isolates of *Campylobacter jejuni* reveals that mutations in genes involved in flagellar biosynthesis alter the organism's virulence potential. *Appl.Environ.Microbiol.* **73**:3123-3136.
- Man, S.M., Kaakoush, N.O., Octavia, S., and Mitchell, H. (2010). The internal transcribed spacer region, a new tool for use in species differentiation and delineation of systematic relationships within the *Campylobacter* genus. *Appl.Environ.Microbiol.* **76**:3071-3081.
- Manning, G., Dowson, C.G., Bagnall, M.C., Ahmed, I.H., West, M., and Newell, D.G. (2003). Multilocus sequence typing for comparison of veterinary and human isolates of *Campylobacter jejuni*. *Appl.Environ.Microbiol.* **69**:6370-6379.
- Mansfield, L.S., Bell, J.A., Wilson, D.L., Murphy, A.J., Elsheikha, H.M., Rathinam, V.A., Fierro, B.R., Linz, J.E., and Young, V.B. (2007). C57BL/6 and congenic interleukin-10-deficient mice can serve as models of *Campylobacter jejuni* colonization and enteritis. *Infect.Immun.* **75**:1099-1115.
- Marotta, F., Zilli, K., Tonelli, A., Sacchini, L., Alessiani, A., Migliorati, G., and Di Giannatale, E. (2012). Detection and Genotyping of *Campylobacter jejuni* and *Campylobacter coli* by Use of DNA Oligonucleotide Arrays. *Mol.Biotechnol.*
- Marsden, G.L., Li, J., Everest, P.H., Lawson, A.J., and Ketley, J.M. (2009). Creation of a large deletion mutant of *Campylobacter jejuni* reveals that the lipooligosaccharide gene cluster is not required for viability. *J.Bacteriol.* **191**:2392-2399.
- Martinez-Rodriguez, A., Kelly, A.F., Park, S.F., and Mackey, B.M. (2004). Emergence of variants with altered survival properties in stationary phase cultures of *Campylobacter jejuni*. *Int.J.Food Microbiol.* **90**:321-329.
- Mason, J.R., and Cammack, R. (1992). The electron-transport proteins of hydroxylating bacterial dioxygenases. *Annu.Rev.Microbiol.* **46**:277-305.
- McDermott, P.F., Bodeis-Jones, S.M., Fritsche, T.R., Jones, R.N., Walker, R. D., and The Campylobacter Susceptibility Testing Group. (2005). Broth microdilution susceptibility testing of *Campylobacter jejuni* and the determination of quality control ranges for fourteen antimicrobial agents. *J.Clin.Microbiol.* **43**:6136-6138.
- McFadyean, J., and Stockman, S. (1913). Report of the Departmental Committee appointed by the Board of Agriculture and Fisheries to inquire into Epizootic Abortion.III. Abortion in Sheep. London: HMSO.

- McGrogan, A., Madle, G.C., Seaman, H.E., and De Vries, C.S. (2009). The epidemiology of Guillain-Barre syndrome worldwide. A systematic literature review. *Neuroepidemiology*. **32**:150-163.
- McHugh, J.P., Rodriguez-Quinones, F., Abdul-Tehrani, H., Svistunenko, D.A., Poole, R.K., Cooper, C.E., and And Rews, S.C. (2003). Global iron-dependent gene regulation in *Escherichia coli*. A new mechanism for iron homeostasis. *J.Biol.Chem.* **278**:29478-29486.
- McKay, D., Fletcher, J., Cooper, P., and Thomson-Carter, F.M. (2001). Comparison of two methods for serotyping *Campylobacter* spp. *J.Clin.Microbiol.* **39**:1917-1921.
- McNally, D.J., Hui, J.P., Aubry, A.J., Mui, K.K., Guerry, P., Brisson, J.R., Logan, S.M., and Soo, E.C. (2006a). Functional characterization of the flagellar glycosylation locus in *Campylobacter jejuni* 81-176 using a focused metabolomics approach. *J.Biol.Chem.* **281**:18489-18498.
- McNally, D.J., Jarrell, H.C., Khieu, N.H., Li, J., Vinogradov, E., Whitfield, D.M., Szymanski, C.M., and Brisson, J.R. (2006b). The HS:19 serostrain of *Campylobacter jejuni* has a hyaluronic acid-type capsular polysaccharide with a nonstoichiometric sorbose branch and O-methyl phosphoramidate group. *FEBS J.* **273**:3975-3989.
- McNally, D.J., Lamoureux, M.P., Karlyshev, A.V., Fiori, L.M., Li, J., Thacker, G., Coleman, R.A., Khieu, N.H., Wren, B.W., Brisson, J.R., Jarrell, H.C., and Szymanski, C.M. (2007). Commonality and biosynthesis of the O-methyl phosphoramidate capsule modification in *Campylobacter jejuni*. *J.Biol.Chem.* **282**:28566-28576.
- McSweegan, E., and Walker, R.I. (1986). Identification and characterization of two *Campylobacter jejuni* adhesins for cellular and mucous substrates. *Infect.Immun.* **53**:141-148.
- Medzhitov, R. (2001). Toll-like receptors and innate immunity. *Nat.Rev.Immunol.* **1**:135-145.
- Mellits, K.H., Mullen, J., Wand, M., Armbruster, G., Patel, A., Connerton, P.L., Skelly, M., and Connerton, I.F. (2002). Activation of the transcription factor NF-kappaB by *Campylobacter jejuni*. *Microbiology*. **148**:2753-2763.
- Melmed, G., Thomas, L.S., Lee, N., Tesfay, S.Y., Lukasek, K., Michelsen, K.S., Zhou, Y., Hu, B., Arditi, M., and Abreu, M.T. (2003). Human intestinal epithelial cells are broadly unresponsive to Toll-like receptor 2-dependent bacterial ligand S: implications for host-microbial interactions in the gut. *J.Immunol.* **170**:1406-1415.
- Mihaljevic, R.R., Sikic, M., Klancnik, A., Brumini, G., Mozina, S.S., and Abram, M. (2007). Environmental stress factors affecting survival and virulence of *Campylobacter jejuni*. *Microb.Pathog.* **43**:120-125.

- Miller, W.G., Parker, C.T., Heath, S., and Lastovica, A.J. (2007). Identification of genomic differences between *Campylobacter jejuni* subsp. *jejuni* and *C. jejuni* subsp. *doylei* at the nap locus leads to the development of a *C. jejuni* subspeciation multiplex PCR method. *BMC Microbiol.* **7**:11.
- Miller, W.G., Pearson, B.M., Wells, J.M., Parker, C.T., Kapitonov, V.V., and Mand Rell, R.E. (2005). Diversity within the *Campylobacter jejuni* type I restriction-modification loci. *Microbiol.* **151**:337-351.
- Miller, C.E., Rock, J.D., Ridley, K.A., Williams, P.H., and Ketley, J.M. (2008a). Utilization of lactoferrin-bound and transferrin-bound iron by *Campylobacter jejuni*. *J.Bacteriol.* **190**:1900-1911.
- Miller, W.G., Wang, G., Binnewies, T.T., and Parker, C.T. (2008b). The complete genome sequence and analysis of the human pathogen *Campylobacter lari*. *Foodborne Pathog.Dis.* **5**:371-386.
- Miller, W.G. (2008). Comparative genomics of *Campylobacter* species other than *Campylobacter jejuni*. In *Campylobacter*. Nachamkin, I., Szymanski, C.M. and Blaser, M.J. (eds). Washington, DC: ASM Press, pp.73-95.
- Misawa, N., and Blaser, M.J. (2000). Detection and characterization of autoagglutination activity by *Campylobacter jejuni*. *Infect.Immun.* **68**:6168-6175.
- Mohammed, K.A., Miles, R.J., and Halablab, M.A. (2005). Simple method to grow enteric *Campylobacters* in unsupplemented liquid medium without the need for microaerophilic kits. *J.Microbiol.Methods.* **61**:273-276.
- Monteville, M.R., Yoon, J.E., and Konkel, M.E. (2003). Maximal adherence and invasion of INT 407 cells by *Campylobacter jejuni* requires the CadF outer-membrane protein and microfilament reorganization. *Microbiology.* **149**:153-165.
- Moore, J.E., Corcoran, D., Dooley, J. S. G., Fanning, S., Lucey, B., Matsuda, M., McDowell, D.A., Mégraud, F., Millar, B.C., O'Mahony, R., O'Riordan, L., O'Rourke, M., Rao, J. R., Rooney, J., Sails, A., and Whyte, P. (2005). *Campylobacter* – Article review. *Vet. Res.* **36**: 351-382.
- Moore, J., Barton, M., Blair, I., Corcoran, D., Dooley, J., Fanning, S., Kempf, I., Lastovica, A., Lowery, C., and Seal, B. (2006). The epidemiology of antibiotic resistance in *Campylobacter* spp. *Microbes.Infect.* **8**:1955–1966.
- Moran, A.P. (1997). Structure and conserved characteristics of *Campylobacter jejuni* lipopolysaccharides. *J.Infect.Dis.* **176 Suppl 2**:S115-21.
- Moran, A.P., and Prendergast, M.M. (2001). Molecular mimicry in *Campylobacter jejuni* and *Helicobacter pylori* lipopolysaccharides: contribution of gastrointestinal infections to autoimmunity. *J.Autoimmun.* **16**:241-256.

- Morooka, T., Umeda, A., and Amako, K. (1985). Motility as an intestinal colonization factor for *Campylobacter jejuni*. *J.Gen.Microbiol.* **131**:1973-1980.
- Murphy, H., Cogan, T., and Humphrey, T. (2011). Direction of neutrophil movements by *Campylobacter*-infected intestinal epithelium. *Microbes Infect.* **13**:42-48.
- Nachamkin, I., Yang, X.H., and Stern, N.J. (1993). Role of *Campylobacter jejuni* flagella as colonization factors for three-day-old chicks: analysis with flagellar mutants. *Appl.Environ.Microbiol.* **59**:1269-1273.
- Naikare, H., Palyada, K., Panciera, R., Marlow, D., and Stintzi, A. (2006). Major role for FeoB in *Campylobacter jejuni* ferrous iron acquisition, gut colonization, and intracellular survival. *Infect.Immun.* **74**:5433-5444.
- Naito, M., Frirdich, E., Fields, J.A., Pryjma, M., Li, J., Cameron, A., Gilbert, M., Thompson, S.A., and Gaynor, E.C. (2010). Effects of sequential *Campylobacter jejuni* 81-176 lipooligosaccharide core truncations on biofilm formation, stress survival, and pathogenesis. *J.Bacteriol.* **192**:2182-2192.
- Nesbakken, T., Eckner, K., Hoidal, H. K., and Rotterud, O. (2003). Occurrence of *Yersinia enterocolitica* and *Campylobacter* spp. in slaughter pigs and consequences for meat inspection, slaughtering and dressing procedures. *Int.J.FoodMicrobiol.* **80**:231-240.
- Newell, D.G., McBride, H., and Dolby, J.M. (1985). Investigations on the role of flagella in the colonization of infant mice with *Campylobacter jejuni* and attachment of *Campylobacter jejuni* to human epithelial cell lines. *J.Hyg.(Lond).* **95**:217-227.
- Nielsen, E.M., Engberg, J., Fussing, V., Petersen, L., Brogren, C.H., and On, S.L. (2000). Evaluation of phenotypic and genotypic methods for subtyping *Campylobacter jejuni* isolates from humans, poultry, and cattle. *J.Clin.Microbiol.* **38**:3800-3810.
- Nielsen, L.N., Luijckx, T.A., Vegge, C.S., Johnsen, C.K., Nuijten, P., Wren, B.W., Ingmer, H., and Kroghelt, K.A. (2012). Identification of immunogenic and virulence-associated *Campylobacter jejuni* proteins. *Clin. Vaccine Immunol.* **19**:113-119.
- Nothaft, H., Liu, X., McNally, D.J., Li, J., and Szymanski, C.M. (2009). Study of free oligosaccharides derived from the bacterial N-glycosylation pathway. *Proc.Natl.Acad.Sci.U.S.A.* **106**:15019-15024.
- Novik, V., Hofreuter, D., and Galan, J.E. (2010). Identification of *Campylobacter jejuni* genes involved in its interaction with epithelial cells. *Infect.Immun.* **78**:3540-3553.
- Oelschlaeger, T.A., Guerry, P., and Kopecko, D.J. (1993). Unusual microtubule-dependent endocytosis mechanisms triggered by *Campylobacter jejuni* and *Citrobacter freundii*. *Proc.Natl.Acad.Sci.U.S.A.* **90**:6884-6888.
- Oldfield, N.J., Moran, A.P., Millar, L.A., Prendergast, M.M., and Ketley, J.M. (2002). Characterization of the *Campylobacter jejuni* heptosyltransferase II gene,

- waaF, provides genetic evidence that extracellular polysaccharide is lipid A core independent. *J.Bacteriol.* **184**:2100-2107.
- O'Leary, A.M., Whyte, P., Madden, R.H., Cormican, M., Moore, J.E., Mc Namara, E., Mc Gill, K., Kelly, L., Cowley, D., Moran, L., Scates, P., Collins, J.D., and Carroll, C.V. (2011). Pulsed field gel electrophoresis typing of human and retail foodstuff *Campylobacters*: an Irish perspective. *Food Microbiol.* **28**:426-433.
- Oleastro, M., Monteiro, L., Lehours, P., Megraud, F., and Menard, A. (2006). Identification of markers for *Helicobacter pylori* strains isolated from children with peptic ulcer disease by suppressive subtractive hybridization. *Infect.Immun.* **74**:4064-4074.
- On, S.L.W. (2001). Taxonomy of *Campylobacter*, *Arcobacter*, *Helicobacter* and related bacteria: current status, future prospects and immediate concerns. *J.Appl.Microbiol.* **90**:1S-15S.
- On, S.L. (1996). Identification methods for *Campylobacters*, helicobacters, and related organisms. *Clin.Microbiol.Rev.* **9**:405-422.
- On, S.L.W., McCarthy, N., Miller, W.G., and Gilpin, B. J. (2008). Molecular epidemiology of *Campylobacter* species. Nachamkin, I., Szymanski, C.M. and Blaser, M.J. (eds). Washington, DC: ASM Press, pp.191-211.
- Otto, T.D., Dillon, G.P., Degrave, W.S., and Berriman, M. (2011). RATT: Rapid Annotation Transfer Tool. *Nucleic Acids Res.* **39**:e57.
- Oyarzabal, O.A., Rad, R., and Backert, S. (2007). Conjugative transfer of chromosomally encoded antibiotic resistance from *Helicobacter pylori* to *Campylobacter jejuni*. *J.Clin.Microbiol.* **45**:402-408.
- Pallen, M.J and Wren, B.W. (2007). Bacterial pathogenomics. Nature reviews. **449**: doi:10.1038/nature06248.
- Palmer, S.R., Gully, P.R., White, J.M., Pearson, A.D., Sucking, W.G., Jones, D.M., Rawes, J.C.L., and Penner, J.L. (1983). Water-borne outbreak of *Campylobacter* gastroenteritis. *Lancet.* **i**: 287-290.
- Palyada, K., Sun, Y.Q., Flint, A., Butcher, J., Naikare, H., and Stintzi, A. (2009). Characterization of the oxidative stress stimulon and PerR regulon of *Campylobacter jejuni*. *BMC Genomics.* **10**:481.
- Palyada, K., Threadgill, D., and Stintzi, A. (2004). Iron acquisition and regulation in *Campylobacter jejuni*. *J.Bacteriol.* **186**:4714-4729.
- Parker, C.T., Gilbert, M., Yuki, N., Endtz, H.P., and Mand Rell, R.E. (2008). Characterization of lipooligosaccharide-biosynthetic loci of *Campylobacter jejuni* reveals new lipooligosaccharide classes: evidence of mosaic organizations. *J.Bacteriol.* **190**:5681-5689.

- Parker, C.T., Miller, W.G., Horn, S.T., and Lastovica, A.J. (2007). Common genomic features of *Campylobacter jejuni* subsp. *doylei* strains distinguish them from *C. jejuni* subsp. *jejuni*. *BMC Microbiol.* **7**:50.
- Parker, C.T., Quinones, B., Miller, W.G., Horn, S.T., and Mand Rell, R.E. (2006). Comparative genomic analysis of *Campylobacter jejuni* strains reveals diversity due to genomic elements similar to those present in *C. jejuni* strain RM1221. *J.Clin.Microbiol.* **44**:4125-4135.
- Parker, C.T., and Huynh, S. (2012). The draft genome sequences of 56 *C. jejuni* and *C. coli* strains. *USDA ARS, Produce Safety and Microbiology Unit, 800 Buchanan St. Albany, CA 94170 USA.*
- Parkhill, J., Wren, B.W., Mungall, K., Ketley, J.M., Churcher, C., Basham, D., Chillingworth, T., Davies, R.M., Feltwell, T., Holroyd, S., Jagels, K., Karlyshev, A.V., Moule, S., Pallen, M.J., Penn, C.W., Quail, M.A., Rajand Ream, M.A., Rutherford, K.M., van Vliet, A.H., Whitehead, S., and Barrell, B.G. (2000). The genome sequence of the food-borne pathogen *Campylobacter jejuni* reveals hypervariable sequences. *Nature.* **403**:665-668.
- Parthasarathy, G., and Mansfield, L.S. (2009). Recombinant interleukin-4 enhances *Campylobacter jejuni* invasion of intestinal pig epithelial cells (IPEC-1). *Microb.Pathog.* **47**:38-46.
- Pavlic, M., and Griffiths, M.W. (2009). Principles, applications, and limitations of automated ribotyping as a rapid method in food safety. *Foodborne Pathog.Dis.* **6**:1047-1055.
- Pearson, B.M., Gaskin, D.J., Segers, R.P., Wells, J.M., Nuijten, P.J., and Van Vliet, A.H. (2007). The complete genome sequence of *Campylobacter jejuni* strain 81116 (NCTC11828). *J.Bacteriol.* **189**:8402-8403.
- Pearson, B.M., Pin, C., Wright, J., I'Anson, K., Humphrey, T., and Wells, J.M. (2003). Comparative genome analysis of *Campylobacter jejuni* using whole genome DNA microarrays. *FEBS Lett.* **554**:224-230.
- Pei, Z. and Blaser, M.J. (1993). PEB1, the major cell-binding factor of *Campylobacter jejuni*, is a homolog of the binding component in gram-negative nutrient transport systems. *J.Biol.Chem.* **268**:18717-18725.
- Pei, Z., Burucoa, C., Grignon, B., Baqar, S., Huang, X.Z., Kopecko, D.J., Bourgeois, A.L., Fauchere, J.L., and Blaser, M.J. (1998). Mutation in the *peb1A* locus of *Campylobacter jejuni* reduces interactions with epithelial cells and intestinal colonization of mice. *Infect.Immun.* **66**:938-943.
- Pei, Z.H., Ellison, R.T., 3rd., and Blaser, M.J. (1991). Identification, purification, and characterization of major antigenic proteins of *Campylobacter jejuni*. *J.Biol.Chem.* **266**:16363-16369.

- Penner, J.L., Hennessy, J.N., and Congi, R.V. (1983). Serotyping of *Campylobacter jejuni* and *Campylobacter coli* on the basis of thermostable antigens. *Eur.J.Clin.Microbiol.* **2**:378-383.
- Penner, J.L., and Hennessy, J.N. (1980). Passive hemagglutination technique for serotyping *Campylobacter fetus* subsp. *jejuni* on the basis of soluble heat-stable antigens. *J. Clin.Microbiol.* **12**:732-737.
- Perez-Arellano, I., Carmona-Alvarez, F., Martinez, A.I., Rodriguez-Diaz, J., and Cervera, J. (2010). Pyrroline-5-carboxylate synthase and proline biosynthesis: from osmotolerance to rare metabolic disease. *Protein Sci.* **19**:372-382.
- Pesci, E.C., Cottle, D.L., and Pickett, C.L. (1994). Genetic, enzymatic, and pathogenic studies of the iron superoxide dismutase of *Campylobacter jejuni*. *Infect.Immun.* **62**:2687-2694.
- Peters, T.M. (2009). Pulsed-field gel electrophoresis for molecular epidemiology of food pathogens. *Methods Mol.Biol.* **551**:59-70.
- Phung le, T., Silver, S., Trimble, W.L., and Gilbert, J.A. (2012). Draft genome of halomonas species strain GFAJ-1 (ATCC BAA-2256). *J.Bacteriol.* **194**:1835-1836.
- Pickett, C.L., Pesci, E.C., Cottle, D.L., Russell, G., Erdem, A.N., and Zeytin, H. (1996). Prevalence of cytolethal distending toxin production in *Campylobacter jejuni* and relatedness of *Campylobacter* sp. *cdtB* gene. *Infect.Immun.* **64**:2070-2078.
- Pittenger, L.G., Englen, M.D., Parker, C.T., Frye, J.G., Quinones, B., Horn, S.T., Son, I., Fedorka-Cray, P.J., and Harrison, M.A. (2009). Genotyping *Campylobacter jejuni* by comparative genome indexing: an evaluation with pulsed-field gel electrophoresis and *flaA* SVR sequencing. *Foodborne Pathog.Dis.* **6**:337-349.
- Pittenger, L.G., Frye, J.G., McNerney, V., Reeves, J., Haro, J., Fedorka-Cray, P.J., Harrison, M.A., and Englen, M.D. (2012). Analysis of *Campylobacter jejuni* whole-genome DNA microarrays: significance of prophage and hypervariable regions for discriminating isolates. *Foodborne Pathog.Dis.* **9**:473-479.
- Plummer, P.J. (2012). LuxS and quorum-sensing in *Campylobacter*. *Front.Cell.Infect.Microbiol.* **2**:22.
- Pogacar, M.S., Klančnik, A., Mozina, S.S., and Cencic, A. (2010). Attachment, invasion, and translocation of *Campylobacter jejuni* in pig small-intestinal epithelial cells. *Foodborne Pathog.Dis.* **7**:589-595.
- Poly, F., Ewing, C., Goon, S., Hickey, T.E., Rockabrand, D., Majam, G., Lee, L., Phan, J., Savarino, N.J., and Guerry, P. (2007a). Heterogeneity of a *Campylobacter jejuni* protein that is secreted through the flagellar filament. *Infect.Immun.* **75**:3859-3867.

- Poly, F., Read, T., Tribble, D.R., Baqar, S., Lorenzo, M., and Guerry, P. (2007b). Genome sequence of a clinical isolate of *Campylobacter jejuni* from Thailand. *Infect.Immun.* **75**:3425-3433.
- Poly, F., Serichatalergs, O., Schulman, M., Ju, J., Cates, C.N., Kanipes, M., Mason, C., and Guerry, P. (2011). Discrimination of major capsular types of *Campylobacter jejuni* by multiplex PCR. *J.Clin.Microbiol.* **49**:1750-1757.
- Poly, F., Threadgill, D., and Stintzi, A. (2005). Genomic diversity in *Campylobacter jejuni*: identification of *C. jejuni* 81-176-specific genes. *J.Clin.Microbiol.* **43**:2330-2338.
- Poly, F., Threadgill, D., and Stintzi, A. (2004). Identification of *Campylobacter jejuni* ATCC 43431-specific genes by whole microbial genome comparisons. *J.Bacteriol.* **186**:4781-4795.
- Poly, F., and Guerry, P. (2008). Pathogenesis of *Campylobacter*. *Curr Opin.Gastroenterol.* **24**:27-31.
- Pope, C., Wilson, J., Taboada, E.N., Mackinnon, J., Felipe Alves, C.A., Nash, J.H., Rahn, K., and Tannock, G.W. (2007). Epidemiology, relative invasive ability, molecular characterization, and competitive performance of *Campylobacter jejuni* strains in the chicken gut. *Appl.Environ.Microbiol.* **73**:7959-7966.
- Poropatich, K.O., Walker, C.L., and Black, R.E. (2010). Quantifying the association between *Campylobacter* infection and Guillain-Barre syndrome: a systematic review. *J.Health Popul.Nutr.* **28**:545-552.
- Prasad, K.N., Dhole, T.N., and Ayyagari, A. (1996). Adherence, invasion and cytotoxin assay of *Campylobacter jejuni* in HeLa and HEp-2 cells. *J.Diarrhoeal Dis.Res.* **14**:255-259.
- Pratt, A., and Korolik, V. (2005). Tetracycline resistance of Australian *Campylobacter jejuni* and *Campylobacter coli* isolates. *J.Antimicrob.Chemother.* **55**:452-460.
- Pryjma, M., Apel, D., Huynh, S., Parker, C.T., and Gaynor, E.C. (2012). FdhTU-Modulated Formate Dehydrogenase Expression and Electron Donor Availability Enhance Recovery of *Campylobacter jejuni* following Host Cell Infection. *J.Bacteriol.* **194**:3803-3813.
- Quinones, B., Guilhabert, M.R., Miller, W.G., Mand Rell, R.E., Lastovica, A.J., and Parker, C.T. (2008). Comparative genomic analysis of clinical strains of *Campylobacter jejuni* from South Africa. *PLoS One.* **3**:e2015.
- Quinones, B., Miller, W.G., Bates, A.H., and Mand Rell, R.E. (2009). Autoinducer-2 production in *Campylobacter jejuni* contributes to chicken colonization. *Appl.Environ.Microbiol.* **75**:281-285.

- Rajashekara, G., Drozd, M., Gangaiah, D., Jeon, B., Liu, Z., and Zhang, Q. (2009). Functional characterization of the twin-arginine translocation system in *Campylobacter jejuni*. *Foodborne Pathog.Dis.* **6**:935-945.
- Raphael, B.H., and Joens, L.A. (2003). FeoB is not required for ferrous iron uptake in *Campylobacter jejuni*. *Can.J.Microbiol.* **49**:727-731.
- Raphael, B.H., Pereira, S., Flom, G.A., Zhang, Q., Ketley, J.M., and Konkel, M.E. (2005). The *Campylobacter jejuni* response regulator, CbrR, modulates sodium deoxycholate resistance and chicken colonization. *J.Bacteriol.* **187**:3662-3670.
- Reeser, R.J., Medler, R.T., Billington, S.J., Jost, B.H., and Joens, L.A. (2007). Characterization of *Campylobacter jejuni* biofilms under defined growth conditions. *Appl.Environ.Microbiol.* **73**:1908-1913.
- Reina, J., Ros, M.J., and Serra, A. (1995). Evaluation of the API-campy system in the biochemical identification of hippurate negative *Campylobacter* strains isolated from faeces. *J.Clin.Pathol.* **48**:683-685.
- Reuter, M., Mallett, A., Pearson, B.M., and Van Vliet, A.H. (2010). Biofilm formation by *Campylobacter jejuni* is increased under aerobic conditions. *Appl.Environ.Microbiol.* **76**:2122-2128.
- Rhodes, K.M., and Tattersfield, A.E. (1982). Guillain-Barre syndrome associated with *Campylobacter* infection. *Br.Med.J.(Clin.Res.Ed)*. **285**:173-174.
- Richardson, P.T., and Park, S.F. (1995). Enterochelin acquisition in *Campylobacter coli*: characterization of components of a binding-protein-dependent transport system. *Microbiology*. **141 (Pt 12)**:3181-3191.
- Ridley, A.M., Allen, V.M., Sharma, M., Harris, J.A., and Newell, D.G. (2008). Real-time PCR approach for detection of environmental sources of *Campylobacter* strains colonizing broiler flocks. *Appl.Environ.Microbiol.* **74**:2492-2504.
- Roberts, M.C. (2005). Update on acquired tetracycline resistance genes. *FEMS Microbiol.Lett.* **245**:195-203.
- Rodin, S., And Ersson, A.F., Wirta, V., Eriksson, L., Ljungstrom, M., Bjorkholm, B., Lindmark, H., and Engstrand, L. (2008). Performance of a 70-mer oligonucleotide microarray for genotyping of *Campylobacter jejuni*. *BMC Microbiol.* **8**:73.
- Romo-Gonzalez, C., Salama, N.R., Burgeno-Ferreira, J., Ponce-Castaneda, V., Lazcano-Ponce, E., Camorlinga-Ponce, M., and Torres, J. (2009). Differences in genome content among *Helicobacter pylori* isolates from patients with gastritis, duodenal ulcer, or gastric cancer reveal novel disease-associated genes. *Infect.Immun.* **77**:2201-2211.
- Rose, A., Kay, E., Wren, B.W., and Dallman, M.J. (2012). The *Campylobacter jejuni* NCTC 11168 capsule prevents excessive cytokine production by dendritic cells. *Med.Microbiol. Immunol.* **201**:137-144.

- Rosef, O., Rettedal, G., and Lageide, L. (2001). Thermophilic *Campylobacters* in surface water: a potential risk of *Campylobacteriosis*. *Int.J.Environ.Health Res.* **11**:321-327.
- Russell, R.G., O'Donnoghue, M., Blake, D.C., Jr., Zulty, J., and DeTolla, L.J. (1993). Early colonic damage and invasion of *Campylobacter jejuni* in experimentally challenged infant *Macaca mulatta*. *J.Infect.Dis.* **168**:210-215.
- Rutherford, K., Parkhill, J., Crook, J., Horsnell, T., Rice, P., Rajand Ream, M.A., and Barrell, B. (2000). Artemis: sequence visualization and annotation. *Bioinformatics.* **16**:944-945.
- Sahl, J.W., Matalaka, M.N., and Rasko, D.A. (2012). Phylomark, a tool to identify conserved phylogenetic markers from whole-genome alignments. *Appl.Environ.Microbiol.* **78**(14):4884-92.
- Salzberg, S.L., Delcher, A.L., Kasif, S., and White, O. (1998). Microbial gene identification using interpolated Markov models. *Nucleic Acids Res.* **26**:544-548.
- Scherer, K., Bartelta, E., Sommerfelda, C., and Hildebrandt, G. (2006). Comparison of different sampling techniques and enumeration methods for the isolation and quantification of *Campylobacter* spp. in raw retail chicken legs. *Int.J.FoodMicrobiol.* **108**:115-119.
- Schirm, M., Schoenhofen, I.C., Logan, S.M., Waldron, K.C., and Thibault, P. (2005). Identification of unusual bacterial glycosylation by tandem mass spectrometry analyses of intact proteins. *Anal.Chem.* **77**:7774-7782.
- Schirm, M., Soo, E.C., Aubry, A.J., Austin, J., Thibault, P., and Logan, S.M. (2003). Structural, genetic and functional characterization of the flagellin glycosylation process in *Helicobacter pylori*. *Mol.Microbiol.* **48**:1579-1592.
- Schnare, M., Barton, G.M., Holt, A.C., Takeda, K., Akira, S., and Medzhitov, R. (2001). Toll-like receptors control activation of adaptive immune responses. *Nat.Immunol.* **2**:947-950.
- Sebahia, M., Preston, A., Maskell, D.J., Kuzmiak, H., Connell, T.D., King, N.D., Orndorff, P.E., Miyamoto, D.M., Thomson, N.R., Harris, D., Goble, A., Lord, A., Murphy, L., Quail, M.A., Rutter, S., Squares, R., Squares, S., Woodward, J., Parkhill, J., and Temple, L.M. (2006). Comparison of the genome sequence of the poultry pathogen *Bordetella avium* with those of *B. bronchiseptica*, *B. pertussis*, and *B. parapertussis* reveals extensive diversity in surface structures associated with host interaction. *J.Bacteriol.* **188**:6002-6015.
- Sebald, M., and Véron, M. (1963). Teneur en bases de l'AND et classification des vibrions. *Ann.Inst.Pasteur.* **105**:897-910.

- Sellars, M.J., Hall, S.J., and Kelly, D.J. (2002). Growth of *Campylobacter jejuni* supported by respiration of fumarate, nitrate, nitrite, trimethylamine-N-oxide, or dimethyl sulfoxide requires oxygen. *J.Bacteriol.* **184**:4187-4196.
- Senior, N.J., Bagnall, M.C., Champion, O.L., Reynolds, S.E., La Ragione, R.M., Woodward, M.J., Salguero, F.J., and Titball, R.W. (2011). Galleria mellonella as an infection model for *Campylobacter jejuni* virulence. *J.Med.Microbiol.* **60**:661-669.
- Shane, S.M. (2000). *Campylobacter* infection of commercial poultry. *Rev.Sci.Tech.* **19**:376-395.
- Sheppard, S.K., McCarthy, N.D., Falush, D., and Maiden, M.C. (2008). Convergence of *Campylobacter* species: implications for bacterial evolution. *Science.* **320**:237-239.
- Shirtliff, M.E., Mader, J.T., and Camper, A.K. (2002). Molecular interactions in biofilms. *Chem.Biol.* **9**:859-871.
- Shoaf-Sweeney, K.D., Larson, C.L., Tang, X., and Konkel, M.E. (2008). Identification of *Campylobacter jejuni* proteins recognized by maternal antibodies of chickens. *Appl.Environ.Microbiol.* **74**:6867-6875.
- Shrader, T.E., Tobias, J.W., and Varshavsky, A. (1993). The N-end rule in Escherichia coli: cloning and analysis of the leucyl, phenylalanyl-tRNA-protein transferase gene *aat*. *J.Bacteriol.* **175**:4364-4374.
- Siemer, B.L., Nielsen, E.M., and On, S.L. (2005). Identification and molecular epidemiology of *Campylobacter coli* isolates from human gastroenteritis, food, and animal sources by amplified fragment length polymorphism analysis and Penner serotyping. *Appl.Environ.Microbiol.* **71**:1953-1958.
- Silverman, N. and Maniatis, T. (2001). NF-kappaB signaling pathways in mammalian and insect innate immunity. *Genes Dev.* **15**:2321-2342.
- Skirrow, M.B. (1977). *Campylobacter* enteritis: a "new" disease. *Br.Med.J.* **2**:9-11.
- Smith, T., and Taylor, M.S. (1919). Some morphological and biological characters of *Spirilla* (*Vibrio foetus* n.sp.) associated with disease of fetal membranes in cattle. *J.Exp.Med.* **30**:299-311.
- Sokal, R.R., and Michener, C.D. (1958). A statistical method for evaluating systemic relationships. *University of Kansas science Bulletin.* **38**. 1409-1438.
- Song, Y.C., Jin, S., Louie, H., Ng, D., Lau, R., Zhang, Y., Weerasekera, R., Al Rashid, S., Ward, L.A., Der, S.D., and Chan, V.L. (2004). FlaC, a protein of *Campylobacter jejuni* TGH9011 (ATCC43431) secreted through the flagellar apparatus, binds epithelial cells and influences cell invasion. *Mol.Microbiol.* **53**:541-553.

- Stahl, M., Friis, L.M., Nothaft, H., Liu, X., Li, J., Szymanski, C.M., and Stintzi, A. (2011). L-fucose utilization provides *Campylobacter jejuni* with a competitive advantage. *Proc.Natl.Acad.Sci.U.S.A.* **108**:7194-7199.
- Stamatakis, A., Ludwig, T., and Meier, H. (2005). RAxML-II: a program for sequential, parallel and distributed inference of large phylogenetic trees. *Concurr.Comput.Prac.Exp.* **17**: 1705-1723.
- Stanley, F. (1988). Molecular Koch postulates applied to microbial pathogenicity. *Microbial Surfaces: Determinants of Virulence and Host Responsiveness*. In *Reviews of Infectious Diseases*. (Vol. 10, Supplement. 2). Oxford Uni Press, pp.S274:S276.
- Stintzi, A., V. Vliet, A.H.M., and Ketley, J.M. (2008). Iron Metabolism, Transport, and Regulation. In *Campylobacter*. Nachamkin, I., Szymanski, C.M. and Blaser, M.J. (eds). Washington, DC: ASM Press, pp.591-610.
- Sulaeman, S., Le Bihan, G., Rossero, A., Federighi, M., De, E., and Tresse, O. (2010). Comparison between the biofilm initiation of *Campylobacter jejuni* and *Campylobacter coli* strains to an inert surface using BioFilm Ring Test. *J.Appl.Microbiol.* **108**:1303-1312.
- Szymanski, C.M., Burr, D.H., and Guerry, P. (2002). *Campylobacter* protein glycosylation affects host cell interactions. *Infect.Immun.* **70**:2242-2244.
- Szymanski, C.M., Michael, F.S., Jarrell, H.C., Li, J., Gilbert, M., Larocque, S., Vinogradov, E., and Brisson, J.R. (2003). Detection of conserved N-linked glycans and phase-variable lipooligosaccharides and capsules from *Campylobacter* cells by mass spectrometry and high resolution magic angle spinning NMR spectroscopy. *J.Biol.Chem.* **278**:24509-24520.
- Szymanski, C.M., and Wren, B.W. (2005). Protein glycosylation in bacterial mucosal pathogens. *Nat.Rev.Microbiol.* **3**:225-237.
- Szymanski, C.M., Yao, R., Ewing, C.P., Trust, T.J., and Guerry, P. (1999). Evidence for a system of general protein glycosylation in *Campylobacter jejuni*. *Mol.Microbiol.* **32**:1022-1030.
- Taboada, E.N., Acedillo, R.R., Carrillo, C.D., Findlay, W.A., Medeiros, D.T., Mykytczuk, O.L., Roberts, M.J., Valencia, C.A., Farber, J.M., and Nash, J.H. (2004). Large-scale comparative genomics meta-analysis of *Campylobacter jejuni* isolates reveals low level of genome plasticity. *J.Clin.Microbiol.* **42**:4566-4576.
- Taboada, E.N., Mackinnon, J.M., Luebbert, C.C., Gannon, V.P., Nash, J.H., and Rahn, K. (2008). Comparative genomic assessment of Multi-Locus Sequence Typing: rapid accumulation of genomic heterogeneity among clonal isolates of *Campylobacter jejuni*. *BMC Evol.Biol.* **8**:229.

- Taboada, E.N., Ross, S.L., Mutschall, S.K., Mackinnon, J.M., Roberts, M.J., Buchanan, C.J., Kruczkiewicz, P., Jokinen, C.C., Thomas, J.E., Nash, J.H., Gannon, V.P., Marshall, B., Pollari, F., and Clark, C.G. (2012). Development and validation of a comparative genomic fingerprinting method for high-resolution genotyping of *Campylobacter jejuni*. *J.Clin.Microbiol.* **50**:788-797.
- Taboada, E.N., van Belkum, A., Yuki, N., Acedillo, R.R., Godschalk, P.C., Koga, M., Endtz, H.P., Gilbert, M., and Nash, J.H. (2007). Comparative genomic analysis of *Campylobacter jejuni* associated with Guillain-Barre and Miller Fisher syndromes: neuropathogenic and enteritis-associated isolates can share high levels of genomic similarity. *BMC Genomics.* **8**:359.
- Tareen, A.M., Dasti, J.I., Zautner, A.E., Gross, U., and Lugert, R. (2011). Sulphite : cytochrome c oxidoreductase deficiency in *Campylobacter jejuni* reduces motility, host cell adherence and invasion. *Microbiology.* **157**:1776-1785.
- Tareen, A.M., Dasti, J.I., Zautner, A.E., Gross, U., and Lugert, R. (2010). *Campylobacter jejuni* proteins Cj0952c and Cj0951c affect chemotactic behaviour towards formic acid and are important for invasion of host cells. *Microbiology.* **156**:3123-3135.
- Theoret, J.R., Cooper, K.K., Glock, R.D., and Joens, L.A. (2011). A *Campylobacter jejuni* Dps homolog has a role in intracellular survival and in the development of *Campylobacteriosis* in neonate piglets. *Foodborne Pathog.Dis.* **8**:1263-1268.
- Thibault, P., Logan, S.M., Kelly, J.F., Brisson, J.R., Ewing, C.P., Trust, T.J., and Guerry, P. (2001). Identification of the carbohydrate moieties and glycosylation motifs in *Campylobacter jejuni* flagellin. *J.Biol.Chem.* **276**:34862-34870.
- Thomas, M.T., Shepherd, M., Poole, R.K., van Vliet, A.H., Kelly, D.J., and Pearson, B.M. (2011). Two respiratory enzyme systems in *Campylobacter jejuni* NCTC11168 contribute to growth on L-lactate. *Environ.Microbiol.* **13**:48-61.
- Tribble, D.R., Sand Ers, J.W., Pang, L.W., Mason, C., Pitarangsi, C., Baqar, S., Armstrong, A., Hshieh, P., Fox, A., Maley, E.A., Lebron, C., Faix, D.J., Lawler, J.V., Nayak, G., Lewis, M., Bodhidatta, L., and Scott, D.A. (2007). Traveler's diarrhea in Thailand: rand Omized, double-blind trial comparing single-dose and 3-day azithromycin-based regimens with a 3-day levofloxacin regimen. *Clin.Infect.Dis.* **44**:338-346.
- Tribble, D.R., Baqar, S., and Thompson, S.A. (2008). Development of a human vaccine. In *Campylobacter*. Nachamkin, I., Szymanski, C.M. and Blaser, M.J. (eds). Washington, DC: ASM Press, pp.429-444.
- Tu, Q.V., McGuckin, M.A., and Mendz, G.L. (2008). *Campylobacter jejuni* response to human mucin MUC2: modulation of colonization and pathogenicity determinants. *J.Med.Microbiol.* **57**:795-802.

- van Alphen, L.B., Bleumink-Pluym, N.M., Rochat, K.D., van Balkom, B.W., Wosten, M.M., and Van Putten, J.P. (2008). Active migration into the subcellular space precedes *Campylobacter jejuni* invasion of epithelial cells. *Cell.Microbiol.* **10**:53-66.
- Van Deun, K., Haesebrouck, F., Heyndrickx, M., Favoreel, H., Dewulf, J., Ceelen, L., Dumez, L., Messens, W., Leleu, S., Van Immerseel, F., Ducatelle, R., and Pasmans, F. (2007). Virulence properties of *Campylobacter jejuni* isolates of poultry and human origin. *J.Med.Microbiol.* **56**:1284-1289.
- van Sorge, N.M., Bleumink, N.M., van Vliet, S.J., Saeland, E., van der Pol, W.L., van Kooyk, Y., and Van Putten, J.P. (2009). N-glycosylated proteins and distinct lipooligosaccharide glycoforms of *Campylobacter jejuni* target the human C-type lectin receptor MGL. *Cell.Microbiol.* **11**:1768-1781.
- van Vliet, A.H., Baillon, M.L., Penn, C.W., and Ketley, J.M. (1999). *Campylobacter jejuni* contains two fur homologs: characterization of iron-responsive regulation of peroxide stress defense genes by the PerR repressor. *J.Bacteriol.* **181**:6371-6376.
- van Vliet, A.H., Ketley, J.M., Park, S.F., and Penn, C.W. (2002). The role of iron in *Campylobacter* gene regulation, metabolism and oxidative stress defense. *FEMS Microbiol.Rev.* **26**:173-186.
- Vandamme, P., Debruyne, L., De Brand T, E., and Falsen, E. (2010). Reclassification of *Bacteroides ureolyticus* as *Campylobacter ureolyticus* comb. nov., and emended description of the genus *Campylobacter*. *Int.J.Syst.Evol.Microbiol.* **60**:2016-2022.
- Vandamme, P. (2000). Taxonomy of the family Campylobacteraceae. In: *Campylobacter*. Nachamkin, I. and Blaser, M.J. (2nd eds). Washington, DC: ASM Press, pp.3-26.
- Vandamme, P., Falsen, E., Rossau, R., Hoste, B., Segers, P., Tytgat, R., and De Ley, J. (1991). Revision of *Campylobacter*, *Helicobacter*, and *Wolinella* taxonomy: emendation of generic descriptions and proposal of *Arcobacter* gen. nov. *Int.J.Syst.Bacteriol.* **41**:88-103.
- Vandamme, P., Vancanneyt, M., Pot, B., Mels, L., Hoste, B., Dewettinck, D., Vlaes, L., van den Borre, C., Higgins, R., and Hommez, J. (1992). Polyphasic taxonomic study of the emended genus *Arcobacter* with *Arcobacter butzleri* comb. nov. and *Arcobacter skirrowii* sp. nov., an aerotolerant bacterium isolated from veterinary specimens. *Int.J.Syst.Bacteriol.* **42**:344-356.
- Vegge, C.S., Brondsted, L., Li, Y.P., Bang, D.D., and Ingmer, H. (2009). Energy taxis drives *Campylobacter jejuni* toward the most favorable conditions for growth. *Appl.Environ.Microbiol.* **75**:5308-5314.
- Velayudhan, J., Jones, M.A., Barrow, P.A., and Kelly, D.J. (2004). L-serine catabolism via an oxygen-labile L-serine dehydratase is essential for colonization of the avian gut by *Campylobacter jejuni*. *Infect.Immun.* **72**:260-268.

- Vijayakumar, S., Merkx-Jacques, A., Ratnayake, D.B., Gryski, I., Obhi, R.K., Houle, S., Dozois, C.M., and Creuzenet, C. (2006). Cj1121c, a novel UDP-4-keto-6-deoxy-GlcNAc C-4 aminotransferase essential for protein glycosylation and virulence in *Campylobacter jejuni*. *J.Biol.Chem.* **281**:27733-27743.
- Wagner, P.L., Livny, J., Neely, M.N., Acheson, D.W., Friedman, D.I., and Waldor, M.K. (2002). Bacteriophage control of Shiga toxin 1 production and release by *Escherichia coli*. *Mol.Microbiol.* **44**:957-970.
- Wai, S.N., Takata, T., Takade, A., Hamasaki, N., and Amako, K. (1995). Purification and characterization of ferritin from *Campylobacter jejuni*. *Arch.Microbiol.* **164**:1-6.
- Wainwright, L.M., Elvers, K.T., Park, S.F., and Poole, R.K. (2005). A truncated haemoglobin implicated in oxygen metabolism by the microaerophilic food-borne pathogen *Campylobacter jejuni*. *Microbiol.* **151**:4079-4091.
- Walker, R.I., Schmauder-Chock, E.A., Parker, J.L., and Burr, D. (1988). Selective association and transport of *Campylobacter jejuni* through M cells of rabbit Peyer's patches. *Can.J.Microbiol.* **34**:1142-1147.
- Wang, L., Jeon, B., Sahin, O., and Zhang, Q. (2009). Identification of an arsenic resistance and arsenic-sensing system in *Campylobacter jejuni*. *Appl.Environ.Microbiol.* **75**:5064-5073.
- Wassenaar, T.M., and Blaser, M.J. (1999). Pathophysiology of *Campylobacter jejuni* infections of humans. *Microbes Infect.* **1**:1023-1033.
- Wassenaar, T.M., Bleumink-Pluym, N.M., and Van der Zeijst, B.A. (1991). Inactivation of *Campylobacter jejuni* flagellin genes by homologous recombination demonstrates that flaA but not flaB is required for invasion. *EMBO J.* **10**:2055-2061.
- Wassenaar, T.M., Engelskirchen, M., Park, S., and Lastovica, A. (1997). Differential uptake and killing potential of *Campylobacter jejuni* by human peripheral monocytes/macrophages. *Med.Microbiol.Immunol.* **186**:139-144.
- Weiss, J. (2003). Bactericidal/permeability-increasing protein (BPI) and lipopolysaccharide-binding protein (LBP): structure, function and regulation in host defence against Gram-negative bacteria. *Biochem.Soc.Trans.* **31**:785-790.
- Werno, A.M., Klena, J.D., Shaw, G.M., and Murdoch, D.R. (2002). Fatal case of *Campylobacter lari* prosthetic joint infection and bacteremia in an immunocompetent patient. *J.Clin.Microbiol.* **40**:1053-1055.
- Whitehouse, C.A., Balbo, P.B., Pesci, E.C., Cottle, D.L., Mirabito, P.M., and Pickett, C.L. (1998). *Campylobacter jejuni* cytolethal distending toxin causes a G2-phase cell cycle block. *Infect.Immun.* **66**:1934-1940.

- Wilma, C., Hazeleger, R.R., Beumer, F.D, and Rombouts, F.M. (1992). The use of latex agglutination tests for determining *Campylobacter* species. *Lett.Appl.Microbiol.* **14**:181-184.
- Wills, W.L., and Murray, C. (1997). *Campylobacter jejuni* seasonal recovery observations of retail market broilers. *Poult.Sci.* **76**: 314–317.
- Wilson, D.L., Rathinam, V.A., Qi, W., Wick, L.M., Land Graf, J., Bell, J.A., Plovanich-Jones, A., Parrish, J., Finley, R.L., Mansfield, L.S., and Linz, J.E. (2010). Genetic diversity in *Campylobacter jejuni* is associated with differential colonization of broiler chickens and C57BL/6J IL10-deficient mice. *Microbiol.* **156**:2046-2057.
- Wilson, I.G., and Moore, J.E. (1996). Presence of *Salmonella* spp. and *Campylobacter* spp. in shellfish. *Epidemol.Infect.* **116**:147-153.
- Wion, D. and Casadesus, J. (2006). N6-methyl-adenine: an epigenetic signal for DNA-protein interactions. *Nat.Rev.Microbiol.* **4**:183-192.
- Wooldridge, K.G., and Ketley, J.M. (1997). *Campylobacter*-host cell interactions. *Trends Microbiol.* **5**:96-102.
- Wooldridge, K.G., Williams, P.H., and Ketley, J.M. (1996). Host signal transduction and endocytosis of *Campylobacter jejuni*. *Microb.Pathog.* **21**:299-305.
- Wren, B.W., Linton, D., Dorrell, N., and Karlyshev, A.V. (2001). Post genome analysis of *Campylobacter jejuni*. *Symp.Ser.Soc.Appl.Microbiol.* **30**:36S-44S.
- Xu, F., Zeng, X., Haigh, R.D., Ketley, J.M., and Lin, J. (2010). Identification and characterization of a new ferric enterobactin receptor, CfrB, in *Campylobacter*. *J.Bacteriol.* **192**:4425-4435.
- Yao, R., Burr, D.H., and Guerry, P. (1997). CheY-mediated modulation of *Campylobacter jejuni* virulence. *Mol.Microbiol.* **23**:1021-1031.
- Young, K.T., Davis, L.M., and Dirita, V.J. (2007). *Campylobacter jejuni*: molecular biology and pathogenesis. *Nat.Rev.Microbiol.* **5**:665-679.
- Young, N.M., Brisson, J.R., Kelly, J., Watson, D.C., Tessier, L., Lanthier, P.H., Jarrell, H.C., Cadotte, N., St Michael, F., Aberg, E., and Szymanski, C.M. (2002). Structure of the N-linked glycan present on multiple glycoproteins in the Gram-negative bacterium, *Campylobacter jejuni*. *J.Biol.Chem.* **277**:42530-42539.
- Yu, R.K., Ariga, T., Usuki, S., and Kaida, K. (2011). Pathological roles of ganglioside mimicry in Guillain-Barre syndrome and related neuropathies. *Adv.Exp.Med.Biol.* **705**:349-365.
- Yuki, N., Manda, S., Tai, T., Takahashi, M., Saito, K., Tsujino, Y, and Taki, T. (1995). Ganglioside-like epitopes of lipopolysaccharides from *Campylobacter jejuni* (PEN19) in

- three isolates from patients with Guillain-Barré syndrome. *J.Neuro.Sciences*. **130**:112-116.
- Yuki, N. (2010). Human gangliosides and bacterial lipo-oligosaccharides in the development of autoimmune neuropathies. *Methods Mol.Biol.* **600**:51-65.
- Zeng, X., Xu, F., and Lin, J. (2009). Molecular, antigenic, and functional characteristics of ferric enterobactin receptor CfrA in *Campylobacter jejuni*. *Infect.Immun.* **77**:5437-5448.
- Zerbino, D.R., and Birney, E. (2008). Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res.* **18**:821-829.
- Zhang, M., He, L., Li, Q., Sun, H., Gu, Y., You, Y., Meng, F., and Zhang, J. (2010). Genomic characterization of the Guillain-Barre syndrome-associated *Campylobacter jejuni* ICDCJ07001 Isolate. *PLoS One*. **5**:e15060.
- Zheng, J., Meng, J., Zhao, S., Singh, R., and Song, W. (2008). *Campylobacter*-induced interleukin-8 secretion in polarized human intestinal epithelial cells requires *Campylobacter*-secreted cytolethal distending toxin- and Toll-like receptor-mediated activation of NF-kappaB. *Infect.Immun.* **76**:4498-4508.
- Zilbauer, M., Dorrell, N., Boughan, P.K., Harris, A., Wren, B.W., Klein, N.J., and Bajaj-Elliott, M. (2005). Intestinal innate immunity to *Campylobacter jejuni* results in induction of bactericidal human beta-defensins 2 and 3. *Infect.Immun.* **73**:7281-7289.
- Zilbauer, M., Dorrell, N., Wren, B.W., and Bajaj-Elliott, M. (2008). *Campylobacter jejuni*-mediated disease pathogenesis: an update. *Trans.R.Soc.Trop.Med.Hyg.* **102**:123-129.
- Zilhao, R., Papadopoulou, B., and Courvalin, P. (1988). Occurrence of the *Campylobacter* resistance gene tetO in *Enterococcus* and *Streptococcus* spp. *Antimicrob.Agents Chemother.* **32**:1793-1796.
- Zweifel, M.A.Z., and Stephan, R. (2004). Prevalence and characteristics of Shiga toxin-producing *Escherichia coli*, *Salmonella* spp. and *Campylobacter* spp. isolated from slaughtered sheep in Switzerland. *Int.J.Food.Microbiol.* **92**:45-53.

APPENDIX

APPENDIX

Supplementary table 1: The average linkage distance between clusters as determined by the “Cluster” software.

NODE1X	ARRY7X	ARRY3X	0.95513916015625
NODE2X	ARRY6X	NODE1X	0.90911865234375
NODE3X	ARRY0X	NODE2X	0.8590087890625
NODE4X	ARRY1X	NODE3X	0.8590087890625
NODE5X	ARRY8X	ARRY5X	0.8580322265625
NODE6X	NODE4X	NODE5X	0.8580322265625
NODE7X	ARRY2X	NODE6X	0.85394287109375
NODE8X	ARRY9X	ARRY4X	0.836669921875
NODE9X	NODE7X	NODE8X	0.836669921875

Note: The Pearson correlation (centered) was used as the similarity metrics for performing clustering (Eisen *et al.*, 1998).

Supplementary table 2: The log 2 ratios *for all genes on the array for six hyperinvasive and four low invasive *C. jejuni* strains.

Oligo_ID	Hyperinvasive <i>C. jejuni</i>						Low invasive <i>C. jejuni</i>			
	<i>C. jejuni</i> 01_10	<i>C. jejuni</i> 01_35	<i>C. jejuni</i> 01/04	<i>C. jejuni</i> 01/41	<i>C. jejuni</i> 01/51	<i>C. jejuni</i> EX114	<i>C. jejuni</i> 01/30	<i>C. jejuni</i> 01/32	<i>C. jejuni</i> 01/46	<i>C. jejuni</i> 01/39
CJ_10000002	-0.1	0.3	0.5	-0.1	0	0	0	-0.1	0.1	0
CJ_10000005	-1.1	1.2	0.2	0.3	-0.2	0.1	0.1	-0.4	0.3	0.2
CJ_10000007	-0.3	0.2	0	0.1	0.2	0	0	-0.1	-0.3	0
CJ_10000008	-0.1	-0.2	0.6	0.1	0.1	0.1	-0.1	0	-0.2	0
CJ_10000009	-0.2	-0.6	-0.8	0.1	-0.1	-2	-0.1	0.1	0.9	-0.9
CJ_10000010	-0.2	-0.1	0.4	0.2	0	0.1	0.2	-0.2	-0.2	0.2
CJ_10000011	-0.5	-0.1	-0.2	0	0	-1.6	-0.1	0	0.2	0
CJ_10000013	-0.4	0	0.4	0	-0.1	0.1	-0.1	0.1	0	0
CJ_10000014	0.1	0.5	-0.2	0.1	0	0.1	0	-0.2	0.2	0.1
CJ_10000015	2.2	2.5	3.1	0.8	-1.9	1.4	2.4	-0.5	1.1	1
CJ_10000016	0.2	0.2	0.7	0.4	0.1	0	0.1	0	-0.1	0.1
CJ_10000017	2.8	3.8	0.5	1.1	-1.8	0.3	1	-1	1.5	1.8
CJ_10000018	-1.6	-0.4	-0.9	-0.2	-0.4	-0.1	-0.1	0.1	0.1	0.2

CJ_10000020	3.3	4.3	1.9	1.1	-1.6	5.6	1.6	-1.8	1.2	2
CJ_10000022	-0.6	0	-0.1	0.1	0	0	-0.1	0	-0.1	0
CJ_10000023	0.6	-0.4	0	0.1	-1	0	1.5	0.8	0.2	0
CJ_10000024	-0.7	1.6	1.6	0	-0.1	0.1	-0.2	0	-0.8	-0.1
CJ_10000026	0.1	0.2	1	0.1	0.3	0.1	0.1	-0.1	-0.4	-0.1
CJ_10000028	-0.3	-2	0.2	0.2	0.1	0.1	0	-0.4	-0.2	-0.3
CJ_10000031	0.3	-0.2	0.7	0.1	0.1	0.1	0.1	0	-0.1	0.1
CJ_10000033	-2.2	-2.3	0.6	-0.1	-0.3	0.2	-0.1	-0.1	0.8	0
CJ_10000035	2.4	3	1.2	0.7	-1.1	1.1	0.8	-0.8	1	1
CJ_10000036	1.2	0.7	-0.2	0.5	-0.8	0.5	0.5	0.1	0.1	0.3
CJ_10000043	-0.5	-0.1	0.4	0	-0.1	0.2	0	0	0	0.1
CJ_10000044	2.2	2.1	1.4	0.6	-0.7	0.2	-0.1	-1.3	0.3	0.7
CJ_10000045	0.1	-1.1	0.5	0.3	0.1	0.1	0.2	0.2	0	0
CJ_10000046	-0.2	-0.1	0.7	0.2	0	0.1	0.2	0.1	-0.3	0
CJ_10000047	-1.7	-0.1	-0.1	0.2	-0.3	0.5	0.1	0	0.1	0.3
CJ_10000050	-0.7	-0.5	0.3	0.1	0	0.3	0.1	0	0	0.1
CJ_10000052	-1.4	-0.8	-0.9	0	0	0.3	0	-0.2	-0.3	0

CJ_1000053	-2	-0.1	-0.9	0	0	0.2	-0.2	0	-0.1	0
CJ_1000054	-0.5	-0.4	0.7	0	-0.1	0.3	0	0	-1.2	0.1
CJ_1000055	-0.5	-0.3	-1.2	0	0.1	0	-0.1	-0.2	-0.1	0.1
CJ_1000056	-2	1.1	0	0.5	-0.3	0	0.2	-0.2	-0.3	0.3
CJ_1000058	-0.5	0	-0.2	0.2	0.1	-0.1	-0.1	-0.2	-0.2	0.1
CJ_1000059	-0.6	0.5	0.5	0.1	-0.1	0.4	0.1	-0.2	0.4	0.2
CJ_1000061	-1.1	1.6	0.6	0.2	-0.3	0.4	-0.1	-0.4	0.1	0.3
CJ_1000062	-1.1	-0.3	0.6	0.2	0	0.2	0	-0.2	0.2	0.1
CJ_1000063	0.1	0.1	0	0.1	0	0.1	0.1	0.3	0.5	0
CJ_1000064	0.5	0.4	0.5	0.1	-0.3	0.4	0	-0.3	0	0.2
CJ_1000065	0.1	0.2	0.3	0.2	0.1	0.2	-0.2	-0.1	-0.1	0.1
CJ_1000066	-0.6	-0.4	-0.2	0.2	-0.2	0.1	0	0	0	0
CJ_1000067	0	0	0.2	0.2	-0.1	0.3	0	-0.1	0	0.2
CJ_1000068	0.5	1.1	-0.8	0.3	-0.5	-0.4	-0.4	-0.3	0.4	0.4
CJ_1000069	-0.8	-0.3	0.4	0.2	0	0.2	0.1	0	-0.1	0.1
CJ_1000070	2.6	1.1	0.9	0.4	-0.9	0.4	0.4	-0.2	0.3	0.6
CJ_1000071	0.1	0.2	0.5	0.2	0.3	0	0.1	0	-0.1	-0.1

CJ_10000073	-1.3	-1.3	-0.2	0.2	0.3	0.1	0.1	0.1	-0.8	-0.2
CJ_10000074	-0.2	0.1	0.4	0.2	0.3	0.1	0	0	0	-0.1
CJ_10000075	0.2	-0.9	0.5	0.1	0.3	-0.1	0.1	0.2	-0.5	-0.2
CJ_10000077	-0.3	-0.2	0.6	0.2	0.1	0.2	0.2	-0.1	-0.1	0.1
CJ_10000078	-0.2	-1.4	1.3	0.1	0.1	-0.1	0.2	0	-0.1	-0.1
CJ_10000080	0.3	0.3	0.8	0.2	0.2	0.1	0.2	0	-0.1	0.1
CJ_10000082	0.6	0.3	0.3	0.3	0.2	0.2	0.2	-0.1	0	0.1
CJ_10000083	0	0	0.5	0.2	0.1	0.1	0	-0.1	-0.1	0
CJ_10000084	0.4	-0.6	0.4	0.3	0	0.1	0.1	0.1	0.2	-0.1
CJ_10000085	-0.1	0.2	0.7	0.2	0.1	0.1	0.1	0	0	0
CJ_10000086	-0.5	-0.1	0.4	0.3	0.1	0.3	0.2	0.1	0.1	0.1
CJ_10000087	-0.9	-1.4	1	0.2	-0.1	0	0.2	0	0	0
CJ_10000088	0.1	0	0.8	0.2	0.1	-0.1	0.1	0	-0.1	0
CJ_10000089	0.1	0	0.4	0.2	0.2	0.1	0.1	0	-0.2	0
CJ_10000091	-0.4	-0.7	0.5	0.2	0.1	0.2	0.2	0.1	-0.1	0
CJ_10000092	-0.2	0.1	0.1	0.2	0.1	0	0	0.1	-0.1	0
CJ_10000095	0.1	0.2	-1	0.2	0	-0.7	-0.9	-1.2	-0.1	0.1

CJ_10000100	-0.9	0.1	-4.4	0.1	-0.9	-3.2	-3.4	-1.8	-0.2	0
CJ_10000102	0.5	0.3	-0.3	0.2	-0.3	0.2	0.1	-0.7	0.3	0.4
CJ_10000103	-0.3	0.2	0.4	0.2	-1.4	0.1	0.1	0	-0.1	0
CJ_10000104	0.2	-0.2	0.6	0.1	0	0.3	0.1	-0.1	0.5	0
CJ_10000105	0.4	-0.3	1.3	0.2	0.2	0.3	0.3	0	-0.2	0
CJ_10000106	-0.7	-0.3	0.3	0.2	0.1	0.2	-0.1	0.2	-0.3	0
CJ_10000107	0.4	-0.3	0.4	0.1	-0.1	0.2	0.2	-0.1	0.3	-0.1
CJ_10000110	-1.9	-1.2	-1.1	0	0	0	-0.4	-0.4	0.1	-0.1
CJ_10000112	0.9	0.5	0.7	0.3	0	0.4	0.3	0	0.2	0.3
CJ_10000113	-0.1	-0.3	0.5	0.1	-0.1	0.1	0.2	0.1	0	0
CJ_10000114	0.1	0.2	0	0.2	-0.1	0.2	0.1	-0.1	-0.1	0.1
CJ_10000115	0.2	0.4	0.7	0.4	0.2	0.1	0.2	0.1	-0.1	0.1
CJ_10000116	0	-0.1	0.6	0.2	0	0.1	0.3	0.1	0	0
CJ_10000117	1	1	0.2	0.3	-0.3	0.3	0.4	-0.1	0.1	0.4
CJ_10000118	-0.7	-0.4	0.1	0.1	0.1	0.2	0.3	0.2	-0.3	0
CJ_10000120	-0.4	-0.9	0.7	0.2	0.1	0.1	0.1	-0.1	0	0
CJ_10000121	0.2	0.2	0.7	0.2	-0.1	0.1	0.1	-0.1	-0.1	-0.1

CJ_10000123	-0.5	-1.6	-0.1	0.1	0	-0.2	0.2	0	-0.3	0.1
CJ_10000125	0.8	-0.4	1.3	0.2	0.1	0.1	0.2	0.2	-0.1	0.2
CJ_10000126	-0.2	0.2	0.3	0.1	-0.1	0.2	0.1	-0.1	0.1	0.1
CJ_10000128	0.3	-1	0.9	0.2	0.1	-0.4	0.1	0.1	0	0.1
CJ_10000129	-0.1	0	0.1	0.2	0.2	0	-0.2	-0.3	-0.1	0
CJ_10000131	0.1	-0.1	0.2	0.1	-0.2	0.1	0.1	-0.1	0.1	-0.1
CJ_10000132	-0.6	-0.3	0.4	0.1	0	0.2	-0.4	-0.1	-0.2	0
CJ_10000135	1.2	1.2	0.5	0.1	-0.4	0.4	0.3	-0.1	0.3	0.5
CJ_10000136	-0.9	0	0	0.2	-0.1	0.3	0.1	0	0.1	0.2
CJ_10000137	-0.9	0.1	-0.3	0	-0.3	0	0	-0.1	0.2	0.1
CJ_10000138	-0.4	0.1	0	0.3	0.1	0.2	0.1	-0.1	0.1	0.1
CJ_10000139	-0.8	-0.1	0.5	0.2	0	0.4	0.2	0	0	0.2
CJ_10000140	-0.1	0	0.5	0.2	0	0.1	0.1	0	-0.2	-0.1
CJ_10000141	0.4	0.3	0.6	0.1	0.2	0	0.2	0.1	-0.1	0
CJ_10000142	0.6	0.2	0.5	0.1	0.1	0.3	0.2	0	-0.1	0.1
CJ_10000143	-0.4	-0.6	0	0	0	0.3	0	0.1	0.3	0
CJ_10000144	-2.5	-1.7	-2.5	0	0	0.1	-0.1	-0.2	-0.1	0.1

CJ_10000145	0.4	0.8	-1.4	0.1	0.9	0.5	0.1	0	0.2	0.1
CJ_10000146	-0.4	-1.3	-2.4	-0.1	-0.3	0.2	0.1	0	0.5	0
CJ_10000147	-1	0.4	-0.1	0.2	-0.1	0.4	-0.3	-0.1	0	0.2
CJ_10000148	-1.1	-0.5	-0.3	0.1	1.2	0.1	-0.2	0.1	-0.3	-0.1
CJ_10000149	-2	-0.2	0.7	0.1	0.1	0.1	0.1	-0.2	0.1	0
CJ_10000150	-0.4	-0.2	0.4	0	0	0.2	0	-0.2	0	0
CJ_10000151	0.2	0.3	0.6	0.2	-0.4	0.1	0	-0.2	-0.1	0.1
CJ_10000152	-0.5	-0.3	0.3	0	0	0	0	-0.1	0.2	0
CJ_10000153	-0.3	-0.1	0.4	0.1	0	0.1	0.1	0	0	-0.1
CJ_10000154	-0.2	0.1	0.4	0.2	0.5	0.3	-0.1	0	0.5	0.1
CJ_10000155	0	-0.7	0.8	0.1	-0.1	0.2	0.2	0.2	0	0
CJ_10000156	-0.2	-0.6	0.2	0.1	-0.4	0.3	0	0	-0.1	0.1
CJ_10000157	-0.5	-0.2	0.7	0.1	0.1	0.1	0.1	-0.1	0.2	0
CJ_10000158	-2.1	-0.4	0	0.1	-0.1	0.1	-0.1	-0.3	-0.1	0
CJ_10000159	0	-0.1	0.4	0.2	0.1	0.2	0.1	-0.1	-0.1	0.1
CJ_10000160	-0.2	0.2	0.6	0.3	0	0.2	0.1	0	-0.1	0.2
CJ_10000162	-0.2	-0.2	0.2	0.2	0.2	0.2	0.1	0	0	0

CJ_10000163	-0.2	-0.1	0.2	0.2	0.1	0.2	0.1	0	-0.1	0
CJ_10000164	1.5	1.3	1.1	0.2	-0.2	0	0.3	0.2	0.2	0
CJ_10000165	-0.1	-0.4	0.6	0.2	0	0.3	0	0.2	0	0.1
CJ_10000166	-0.2	0.2	0.5	0.2	0.2	0.1	0	0	-0.1	0.1
CJ_10000168	0.7	0.7	0.5	0.3	0.1	0.3	0.2	0	0.1	0
CJ_10000169	0.1	0.6	0	0.2	0.4	0.2	0	-0.1	0.2	0.1
CJ_10000170	1.4	0.1	1.3	0.5	-0.3	0.4	0.3	-0.3	0.7	0.6
CJ_10000171	-0.6	0.3	0.1	0.1	0	0.4	0	-0.1	-0.1	0.1
CJ_10000172	-0.9	0.1	-0.2	0.1	-0.5	0.2	0	-0.1	0.1	0
CJ_10000173	-0.5	-0.9	0.1	0	-0.2	0.2	0	-0.3	-0.1	0.1
CJ_10000174	-0.5	-0.5	0	0.1	0.1	0.2	-0.1	0	-0.1	0
CJ_10000175	-0.7	-0.6	0.4	0.2	0.5	0.3	0	-0.1	0	0.1
CJ_10000176	-0.6	-0.1	-0.1	-0.1	-0.3	-0.1	-0.1	0	0.3	0.1
CJ_10000177	0.1	-0.3	-0.2	0.1	0	-0.2	-0.5	-0.2	0.1	0.1
CJ_10000179	-0.9	-0.5	0	0.1	-0.1	0.2	0.1	-0.1	0.1	0
CJ_10000180	0.6	0.1	-0.1	0.1	-0.4	0.4	0.1	-0.1	-0.2	0.1
CJ_10000181	-0.6	-0.1	-0.4	0.1	0	0.2	-0.1	-0.1	0	0.1

CJ_10000183	1	0.4	0.6	0.2	-0.3	0.4	0.5	0	0.2	0.4
CJ_10000184	-0.2	-0.1	0.3	0.3	0.1	0.1	0.1	-0.2	-0.3	0.2
CJ_10000185	-1.6	-0.7	-1	0.1	0.1	0.2	0	0	0.1	0
CJ_10000186	0	-1.5	-0.9	-0.3	0	-0.2	-0.5	-0.1	-0.3	-0.4
CJ_10000187	0.1	0.1	0.4	0.2	0.2	0.2	0.1	0	0	0.1
CJ_10000188	0.3	0.1	-5.8	0.2	-0.4	-1.5	-0.2	-1.2	0.4	0.1
CJ_10000189	0	0	-0.2	0.1	-0.1	-0.1	-0.5	-0.6	0	0.1
CJ_10000190	-0.6	-0.1	-0.2	0.2	0.1	0	-0.2	0.1	0	0
CJ_10000191	0	-0.3	0.8	0.2	0.1	0	0	0.1	0	0
CJ_10000193	-0.8	-1.2	1	0.2	-0.2	0.4	0.3	0.1	0.3	0.1
CJ_10000196	0.1	-0.4	0.3	0	0.1	0.1	0	0.1	-0.1	-0.2
CJ_10000200	0.4	0.1	0.7	0.1	0	0.1	0.1	-0.2	0	0.1
CJ_10000204	0.2	0	-0.9	0	-0.2	0	-0.3	0.1	0.1	0.1
CJ_10000210	0	0.1	0.3	0.1	0.2	0.2	0.1	0.2	-0.1	0
CJ_10000213	0.7	0.2	0.8	0.3	0.1	0.2	0.3	0.2	-0.2	0
CJ_10000214	0.5	-0.6	0.6	0.1	-0.1	0.1	0	-0.5	0.2	-0.1
CJ_10000215	-0.1	0	0.9	0.1	0	0.3	0	0	0	0

CJ_10000216	-0.6	-0.2	0.1	0.2	0.1	0.2	-0.2	0	-0.1	0.1
CJ_10000217	-1.1	-0.3	-0.1	0	0	0.3	0	0	0.4	-0.1
CJ_10000218	-1	-0.2	-0.4	0.1	-0.1	0.2	-0.1	-0.2	0	0.1
CJ_10000219	-0.4	0	0.3	0.1	0.1	0.2	-0.1	-0.1	0	0
CJ_10000220	-0.2	-0.6	0.2	0.1	-0.1	0	0	-0.2	0	0
CJ_10000221	-0.1	0.1	0.5	0.1	0	0.2	0.1	0	0	0.1
CJ_10000222	-1.8	0	-0.1	0.1	0	0.3	-0.1	0	0	0.1
CJ_10000223	0	0.3	0	0	-0.2	0.2	0	0	0.2	0.1
CJ_10000224	-0.4	-0.1	0.4	0.1	0.1	0.2	0	-0.1	0	0
CJ_10000225	-1.1	0.9	-0.3	0.3	-0.1	0.4	0.2	-0.1	-0.1	0.2
CJ_10000228	-0.4	-0.1	-0.7	0.1	0	0.3	0	0.1	0.1	0.1
CJ_10000230	-0.2	0.1	0.2	0.8	-0.2	-0.1	0	0.5	0.1	0.9
CJ_10000231	-1.1	-0.2	0	0.2	0.2	0.1	-0.1	0.1	0	0
CJ_10000232	-0.1	0.1	0.5	0.3	0.1	0	-0.3	0.2	-0.4	-0.3
CJ_10000234	0.2	0.2	0.3	0.2	0	0.1	0	0	0.2	0.1
CJ_10000235	0	-0.1	0	0.2	0	0	-0.2	0.1	0	0
CJ_10000236	0.2	0	-0.3	0.2	0.1	-1.7	-0.2	0.1	-0.3	-0.1

CJ_10000237	-0.8	-0.4	-0.3	0.1	0	0.2	0	0	-0.1	0.1
CJ_10000238	-0.7	-1.3	-0.2	0	-0.2	0.4	0	-0.1	0.3	0.2
CJ_10000239	-1.3	-1	-0.4	0.2	-0.1	0.3	-0.1	-0.2	-0.1	0.1
CJ_10000240	-1.3	-0.3	-0.7	0.1	0	0.3	-0.2	0	0.3	0.2
CJ_10000241	-0.2	-0.1	0.6	0	0	0.1	0.1	0.1	0.1	-0.1
CJ_10000242	-2.9	-0.2	0	0.1	0.1	0.1	-0.1	-0.3	-0.1	0.1
CJ_10000243	-1	-0.2	-0.2	0.1	0	0.2	0	-0.1	0	0.1
CJ_10000244	-0.3	-0.1	0.4	0	-0.1	0.2	0	0	-0.2	0.1
CJ_10000245	-0.7	0.3	0	0.1	0.2	0.1	-0.2	0	0.2	0.1
CJ_10000246	-0.6	-0.2	0.2	0.2	0.2	0.2	0	0	-0.1	0.1
CJ_10000247	0.4	0.4	0.5	0.1	0	0	0.2	0	0	0.1
CJ_10000248	0.3	0.2	0.6	0.2	-0.1	0.1	0.2	0	0	0.1
CJ_10000249	-0.6	-0.2	-0.2	0.1	0	0.1	-0.1	0	0	0
CJ_10000252	-0.6	-0.2	0.3	0	0	0.3	0.1	0.1	-0.2	0
CJ_10000254	0.4	-1	0.5	0.4	0.1	0	-0.1	-0.2	-0.1	0.1
CJ_10000255	0.2	0.3	0.3	0.2	0	0.1	0.2	0	0.1	0
CJ_10000256	0.3	-0.2	0.3	0.2	0.3	0	0	0.2	-0.3	-0.1

CJ_10000257	0	0.2	0.2	0.1	-0.1	0	-0.7	0	0.1	0
CJ_10000258	-0.1	0	0.1	0.2	0	0.3	0	0	-0.3	0
CJ_10000259	0.8	0.3	0.8	0	0.1	0	0.3	0.1	0.1	0
CJ_10000260	0.6	-0.2	-0.2	0.2	0	0.1	0.1	0.1	0.1	0.1
CJ_10000261	-0.4	0.2	0.2	0.1	0.1	0.1	0	0.1	-0.1	0
CJ_10000262	0.1	-0.7	0.9	-0.2	0.1	-0.1	0.1	0.1	0	-0.2
CJ_10000263	0	0.2	0.3	0.1	-0.1	0.3	0.1	0	0	0.1
CJ_10000264	0.7	0.2	0.5	0.3	0.2	0	0	-0.1	-0.2	0.1
CJ_10000265	-0.1	0.4	1	0.1	-0.2	0.1	0.2	-0.1	0	0.2
CJ_10000267	-0.1	0.1	0.5	0.2	0.1	0.1	0	-0.2	0	0
CJ_10000268	0.7	0.8	0.2	0.1	-0.2	0.1	0.1	0	0.1	0.1
CJ_10000269	0	0	0.7	0.2	0.1	0.2	0.1	-0.3	0	0.1
CJ_10000270	0.7	0.3	0.9	0.2	0	0.2	0.1	0	-0.1	0.1
CJ_10000271	-0.8	-0.2	0.4	0.3	-0.1	0.1	0.2	0.1	0.1	0
CJ_10000272	-0.1	-0.2	0.5	0.2	0.1	0.1	0.2	-0.3	-0.1	0.1
CJ_10000273	0.5	0.2	0.6	0.1	0	0.2	0.2	0	-0.1	-0.1
CJ_10000276	-0.5	-0.2	-0.2	0	0	0.3	0.1	0	0	0.1

CJ_10000278	-0.1	0.4	0	1	0.2	0.1	0.2	-0.1	0	-0.1
CJ_10000279	-0.2	-0.1	0	0.1	0.1	0.1	-0.1	0.1	-0.1	0
CJ_10000280	-1	-0.7	0.1	0.2	0.2	-0.1	-0.3	0.3	-0.6	-0.1
CJ_10000281	5.6	1.3	-0.1	1.2	0.1	-0.9	1.9	1.7	2.7	-1.3
CJ_10000282	-0.2	-0.8	0.4	0.1	0.4	0.2	0	0.3	0	0
CJ_10000284	0.5	-0.6	0	0.1	0.4	-0.4	0	0.1	-0.2	-0.1
CJ_10000285	-0.6	-0.1	-0.2	0	0	0.1	-0.1	0.1	0	0
CJ_10000286	-0.3	0.6	0.9	0.2	0.1	0.1	0.1	-0.1	-0.2	0
CJ_10000287	-0.5	-0.2	0.1	0.1	0	0.1	0	-0.1	-0.1	0
CJ_10000288	0.2	-0.6	1.4	0.1	0	0	0	0.1	0.2	-0.1
CJ_10000289	-0.1	0.1	0.7	0.3	0.2	0.1	0	0	0	0.1
CJ_10000290	1	0.5	0.5	0.4	-0.1	0.3	0.2	-0.2	0	0.2
CJ_10000291	-0.4	-0.7	0.6	0.2	-0.1	0.1	0.1	0.1	0	-0.1
CJ_10000292	-1.7	-0.9	-0.5	0.1	-0.1	0.5	0.1	0.1	0.4	0.1
CJ_10000293	3	2.4	3.1	0.9	-0.7	0.8	1.3	-0.4	0.7	0.8
CJ_10000294	-0.7	-0.6	0.2	0.1	-0.1	-0.1	0	0.1	-0.4	0.1
CJ_10000295	0.1	-0.1	0.6	0.1	-0.1	0.1	0	0	0.1	0

CJ_10000296	-2.1	-1.2	-0.1	0.1	0.2	0.3	0	0.1	-0.2	0
CJ_10000297	-0.7	-0.5	0.2	0.1	-0.1	0.1	0	0	-0.1	-0.1
CJ_10000298	-0.6	0.2	0.2	-0.2	-0.3	0.2	0.1	0	0.3	0
CJ_10000299	0.5	-0.1	-0.8	0	-0.2	0.1	-0.2	0	0.4	0.1
CJ_10000302	-0.8	-0.2	-0.8	-1.7	0	0.1	-0.4	1.2	-0.1	-1.5
CJ_10000304	0.4	0.5	-0.1	0.2	0.1	0.2	-0.1	-0.2	-0.1	0
CJ_10000305	-0.6	0	-0.6	-1.3	0	0.1	-0.1	1.4	0	-1.3
CJ_10000307	0.3	-0.3	0.4	0.2	0.2	0.1	0.2	0.3	-0.3	-0.1
CJ_10000308	-0.8	-0.6	-0.1	0.1	0.2	0	-0.7	-0.1	-0.2	0
CJ_10000310	-0.2	-0.2	1.2	0.2	0.1	0.2	-0.1	-0.2	-0.2	0.1
CJ_10000311	-0.4	-0.3	0	0.2	0.1	0.3	-0.2	0	0	0.1
CJ_10000312	-0.1	-0.3	1	0.2	0.1	0.1	0	0	-0.1	-0.1
CJ_10000313	-3.9	-2.5	0.3	0.1	-0.1	0.2	0.1	-0.2	-0.1	0
CJ_10000314	0.1	0.1	0.3	0.2	-0.1	0.2	0	-0.2	-0.1	0.1
CJ_10000316	-0.1	-0.1	0.6	0.1	-0.1	0.1	0	-0.1	-0.2	-0.1
CJ_10000317	-0.7	0.5	0.3	0.3	-0.2	0.1	0	-0.6	-0.1	0.1
CJ_10000318	1.2	0.4	1.2	0.2	-0.2	0.2	0.3	-0.1	0.2	0.1

CJ_10000319	0.1	0.3	0.2	0.1	-0.1	0.2	0.1	0	0.1	0
CJ_10000320	-0.6	0.1	-2	0	-0.2	-1.2	0	0	0	0.1
CJ_10000322	-0.4	-0.2	0.1	0.1	-0.1	0.1	-0.1	-0.4	0	0
CJ_10000323	-1.8	-0.7	0.2	0	-0.1	0.2	-0.2	0.2	-0.4	0
CJ_10000325	-0.1	0	0.3	0.1	0.1	0.1	0.2	0.1	0	0
CJ_10000326	-1.6	-0.3	-0.2	0	-0.1	0.2	0	0	0.1	0.1
CJ_10000328	0.4	0.6	-0.2	0.1	-0.1	0.2	0.1	0.3	-0.1	-0.1
CJ_10000329	-0.3	0.3	0.5	0.2	0.2	0.1	0.1	0.2	-0.3	0
CJ_10000331	0.9	-0.1	-0.2	0.2	0	-0.9	-0.1	0.1	-0.3	-0.1
CJ_10000332	-0.2	-0.3	-0.2	0.1	0	0.5	-0.1	-0.1	0.2	0.1
CJ_10000334	-0.8	-0.4	0.9	0	-0.2	0.2	0.1	0	-0.2	0
CJ_10000335	-1.3	-0.4	-0.7	0.2	0.1	0.3	-0.2	0	0.4	0.1
CJ_10000336	-0.1	-1.9	1	0.1	0	0.1	0	-0.1	0.3	-0.1
CJ_10000337	-0.1	-0.2	0.6	0.2	0.1	0.1	0.1	0	-0.1	0
CJ_10000340	-3.2	-0.1	0.1	0.1	-0.3	0.2	-0.1	0	0.1	0.1
CJ_10000341	0	-0.2	0.6	0.1	0	0.2	0.1	0	0	0
CJ_10000342	-0.7	-0.5	-0.1	0	-0.2	0.2	0	0	0	0

CJ_10000343	-0.2	-0.2	0.4	0.2	0.1	0.2	-0.1	-0.1	0	0.1
CJ_10000344	-0.2	-0.1	0.3	0.1	0.1	0.1	0	0	-0.1	0.1
CJ_10000347	-0.4	-0.2	0.3	0.1	0	0.2	0.1	0.1	-0.2	0
CJ_10000349	-0.8	-0.3	0.4	0.1	0	0.2	0.1	0.1	0.1	0
CJ_10000350	-1.6	-1	-0.7	0.1	0	0.1	-0.2	-0.8	0.1	0.1
CJ_10000352	-0.7	0.1	0.6	0.1	0.1	0.2	0.3	0.2	-0.2	0
CJ_10000353	-0.4	-0.1	-0.3	0.1	-0.1	0.2	0	0.1	0.1	0.1
CJ_10000354	-0.7	-0.1	0.2	0.3	0.1	0.1	0	0.2	-0.3	-0.1
CJ_10000355	0.3	-0.4	0.4	0.1	0	0.1	0.1	0	0	-0.1
CJ_10000356	0.1	0	-0.2	0.1	0.1	0.1	-0.1	0	-0.1	-0.1
CJ_10000357	-0.4	-0.9	0.4	-0.1	0.1	-0.2	0	0	-0.1	-0.2
CJ_10000358	-1.6	-0.2	0.3	0.1	0.4	0.1	-0.3	0	-0.2	0
CJ_10000359	-0.1	-0.1	0.3	0.1	0	0.3	0.1	0	0	0.2
CJ_10000361	-0.4	0.1	0.6	0.2	0	0.2	0	-0.1	-0.1	0
CJ_10000362	-0.4	0	-0.8	0.2	0.1	0.2	0.1	0	0.1	0.1
CJ_10000364	-0.1	0.2	-0.2	0.1	-0.4	0.4	0.2	-0.1	0.1	0.2
CJ_10000365	0.5	0	0.5	0.3	0.1	0.2	0.2	0	0	0

CJ_10000367	0.2	0.3	0.3	0.2	-0.1	0.2	0.2	0	-0.1	0.1
CJ_10000368	0.7	0.1	0.4	0.1	0	0.1	0.2	-0.2	0.1	0
CJ_10000370	0	-0.3	0.3	0.3	0	0.1	0.1	0.1	-0.1	-0.1
CJ_10000373	0.3	0.3	0	0.2	1	-0.5	-0.2	0.1	-0.1	-0.1
CJ_10000374	2.3	1.6	0.9	0.4	-0.4	0.4	0.5	-0.1	0.2	0.2
CJ_10000376	-0.9	-0.4	-0.3	0.2	0.1	0.2	0	0.3	-0.1	0
CJ_10000377	2	2.5	2.5	0.7	-1	1.7	1.6	-0.6	0.7	0.9
CJ_10000379	-0.5	-1.3	0.3	0.1	0.1	0.2	0	0.2	-0.3	0
CJ_10000381	-1	-1	-0.1	0.2	0	0.3	-0.2	-0.2	0	0
CJ_10000382	-0.5	0.3	0.2	0.1	-0.2	0.3	0.2	0	0	0.2
CJ_10000383	-0.3	-0.2	0.1	-0.1	-0.1	0.2	0	-0.3	-0.2	-0.1
CJ_10000384	-1.2	0.7	-0.1	0.1	-0.3	0.2	0.1	-0.3	0.1	0.1
CJ_10000385	-0.2	0	0.4	0.2	0	0.1	-0.2	-0.3	0	0
CJ_10000386	-0.7	-0.7	0.5	0.1	0	0.1	0	0	-1.5	0
CJ_10000387	1	1.8	0.7	0.3	-1	0.5	0.6	-0.1	0.3	0.3
CJ_10000388	-0.3	0	0.1	0.2	0.1	0.3	-0.1	-0.1	-0.1	0
CJ_10000389	-0.4	-1.6	0	0	0.2	0.1	-0.1	0.2	0.4	0

CJ_10000390	0.1	0.2	1.9	-0.1	0	0.1	-0.1	-0.2	-0.3	0.4
CJ_10000391	0	0	0.6	0	0	0.2	0	-0.1	-0.1	0.1
CJ_10000393	-0.5	-0.1	0.2	0.2	-0.2	0.2	0	0	0	0
CJ_10000394	-0.6	0	-0.2	0.1	-0.1	0.3	0.1	0	0	0.1
CJ_10000395	-1.4	0	-0.4	-0.2	-0.3	0.3	-0.1	0.1	0.4	0
CJ_10000396	-0.5	0	0.3	0.1	-0.3	0.3	0.2	0	0	0.2
CJ_10000397	-0.1	0.1	0.4	0.1	-0.1	0.2	0.1	0	-0.1	0.1
CJ_10000398	-0.9	-0.3	-0.2	-0.1	-0.1	0.1	-0.2	0.1	0.1	-0.1
CJ_10000399	0	0.1	0.8	0	-0.2	0	0	-0.1	0.3	0.1
CJ_10000400	-0.8	-0.6	-1.8	0.2	-0.2	0.3	-1.1	-0.6	0	0.2
CJ_10000401	0.2	0.2	0.7	0.1	-0.2	0.1	0.1	0.1	-0.1	0
CJ_10000402	0	0.7	-0.1	0.1	-0.2	0.3	0.1	0	0.2	0.2
CJ_10000403	-0.3	0.4	-0.1	0.2	-0.1	0.3	0	0	0.1	0
CJ_10000404	0.2	-0.1	-0.6	0	-0.4	0.3	0	0.2	-0.1	0
CJ_10000405	0.3	0.5	0.3	0.2	0.6	0	0.1	0	-0.2	0.1
CJ_10000406	-0.9	-0.3	-0.6	0	0	0.2	0	0.1	0	0.1
CJ_10000407	-0.2	-0.3	0.2	0.1	-0.1	0.2	0	0	0.1	-0.1

CJ_10000408	-0.7	0.3	1	0.1	0.2	0	0.1	0	-0.1	-0.1
CJ_10000409	0.2	0.5	0.4	0.2	0.3	0	-0.1	-0.1	0.1	-0.1
CJ_10000410	0	0.6	0.4	0.1	0	0.2	0	0	0.1	0
CJ_10000411	0.3	-0.1	0.5	0	-0.1	0	0.1	0	-0.2	0
CJ_10000412	-0.1	-0.1	0.4	0.1	0.1	0.1	0	0	-0.1	0
CJ_10000413	-0.1	-0.5	-0.2	0	0	0.1	0	0.1	0.5	0.1
CJ_10000414	-0.8	0	-0.5	-0.1	0	0.3	0	0	0.1	0
CJ_10000415	-0.7	0	0.1	0.2	0.1	0.2	0	0	0.1	0.2
CJ_10000417	-1.1	-0.5	0	0.2	-0.2	0.4	0	0.1	-0.2	0.2
CJ_10000418	-0.5	-0.4	-0.5	0.1	0	0.2	0	-0.1	-0.2	0.1
CJ_10000419	-1	-0.6	-2	0.1	-0.4	-0.3	-0.3	-1.2	-0.1	0.1
CJ_10000420	-0.3	-0.1	0.1	0.2	-0.2	0.1	0	0	0	0
CJ_10000421	-0.4	-0.2	0	0.2	0.1	0.2	0.2	0	-0.1	0.1
CJ_10000422	-0.3	0.1	-0.1	0	-0.3	0.1	-0.1	0.1	0	0
CJ_10000423	-0.1	0.3	0.1	0.1	-0.1	0.3	0.1	-0.1	-0.1	0.1
CJ_10000424	-0.4	-0.1	-0.4	0	-0.1	0.1	-0.1	0	0	0
CJ_10000425	-0.9	0.1	0.1	0.2	-0.3	0	0	-0.1	0.6	0.3

CJ_10000426	0	0	0.1	0.1	0	0.1	0.1	0	-0.1	0.1
CJ_10000427	-0.3	0.2	-0.4	0	-0.2	0.3	-0.2	-0.1	0	0.1
CJ_10000428	0	-2	0.4	0.1	0.1	0.2	0	0	-0.9	0
CJ_10000429	-0.1	-0.7	-0.2	0.2	0.1	0.2	0.1	-0.1	-0.3	0.1
CJ_10000430	-0.4	-0.1	0.2	0.1	-0.1	0.3	0	-0.1	0.2	0.2
CJ_10000433	-0.2	0.1	0.2	0.1	0.1	0.2	-0.1	-0.1	0	0
CJ_10000434	0.6	-0.5	0.8	0.2	0.2	0	0	0	0.3	0
CJ_10000435	-0.3	-0.1	-0.1	0.1	0.1	0.2	-0.2	-0.2	0	0
CJ_10000436	-0.1	0.7	0.9	0.1	0	0	0.1	-0.1	0	0.2
CJ_10000437	0.5	0.9	1.1	0.3	0	0.1	0.1	0.1	0.2	-0.1
CJ_10000438	-0.8	0.2	-1.2	0	0.1	-0.1	-0.1	-0.3	-0.9	-0.2
CJ_10000439	0.1	0.5	0.8	0.2	0	0.2	0.1	0	0.2	0.2
CJ_10000441	0.2	0.6	0.6	0.3	-0.4	0.3	0.3	0	0.2	0.1
CJ_10000442	-0.5	0	-0.1	0.1	0	0.2	0	0	0	0.1
CJ_10000444	0	0.6	0.7	0.2	-0.3	0.2	0.3	-0.2	0.1	0.1
CJ_10000445	-0.1	0	-0.2	0.1	0	0.2	-0.1	-0.2	0.1	0.1
CJ_10000446	-2.6	-1.2	0.2	0.1	0	0.1	0	-0.2	0.5	0

CJ_10000447	0.1	0.8	0.6	0.3	-0.1	0.1	0.2	-0.1	0	0
CJ_10000448	0.7	0.8	0.6	0.3	-0.1	0.1	0.1	0	0.2	0
CJ_10000450	-0.1	1.1	1.2	0.4	-0.3	0.3	0.3	0	0.1	0.2
CJ_10000451	-0.3	0.1	-0.1	0	0	0.1	-0.2	-0.3	0.1	0.1
CJ_10000452	0.2	-0.2	-0.1	0.1	0	0.1	0.1	0.1	-2.3	-0.1
CJ_10000453	-0.4	-0.2	-0.1	0.1	0	0	-0.1	-0.1	0	0
CJ_10000454	-0.5	0.1	0.3	0.2	0.1	0.2	0.1	0	-0.4	0
CJ_10000455	-0.4	-0.6	0.4	-0.1	-0.2	0.2	-0.1	0	-0.2	0
CJ_10000456	-1.2	-0.3	-0.7	0.1	0	0.3	0	0	0.2	0
CJ_10000457	0	-0.1	0.2	0.2	0.1	0.2	-0.1	-0.1	0	0
CJ_10000458	0.7	0.5	1.3	0.3	0	0.3	0	-0.1	-0.1	0.1
CJ_10000459	0.1	0.3	0	0.1	0.1	0.2	-0.1	0.1	0	0
CJ_10000465	3.2	4.2	0.7	-0.1	-0.8	0.9	1.5	-1.9	1.1	-0.2
CJ_10000466	-0.4	-0.3	-0.1	0.2	0.1	0.2	-0.1	-0.2	0	0
CJ_10000467	3.9	3	0.1	-0.2	-1.4	1.2	3.7	0.5	0.2	-0.1
CJ_10000468	0.4	0.3	0.3	0.2	-0.2	0.2	0.3	0	0	0.2
CJ_10000470	-0.5	-0.1	0.2	0.2	-0.2	0.1	0.2	0	0.2	0.1

CJ_10000471	-0.1	0.2	0.4	0.2	0	0.1	0.1	0	0	0
CJ_10000472	-0.8	-0.1	0	0	-0.3	0.2	0.1	0	0	0.1
CJ_10000473	1.1	0.4	0.5	0.4	0.1	-0.4	0	0.1	0	-0.1
CJ_10000474	0.7	-0.9	-0.7	0.2	0.1	-0.7	0	0.1	0.1	0
CJ_10000475	0.2	0	0.3	0.2	0.1	0.1	-0.1	-0.1	0.1	0.2
CJ_10000476	-0.6	-1.4	0.8	0.1	0	0.2	0.4	0	0	0.1
CJ_10000477	0.3	0.3	0.6	0.2	1.2	0.1	0	0.1	0	0
CJ_10000478	0	-0.7	-0.2	-0.1	0	0.1	-0.2	-0.1	-0.3	0
CJ_10000479	-0.5	-0.3	-0.4	0	0	0.2	0	0	0	0
CJ_10000480	-1.2	-0.5	-0.7	0.1	0.5	0.3	-0.1	0	0	0
CJ_10000481	0.2	-0.4	0.6	0.1	0.2	0.1	0.1	0.1	-0.1	0
CJ_10000482	1.4	1.2	0	0.2	0	0.1	1.7	1.5	1.5	0
CJ_10000484	-0.1	-0.7	0.1	0	0.2	0.3	0.1	0	-0.2	0
CJ_10000485	0.1	-0.1	0.5	0	0.3	0.1	0	0.1	-0.1	0
CJ_10000486	-0.2	-0.3	0.2	0.2	0	0.2	0.1	0	-0.1	0.1
CJ_10000487	-0.2	-1	0.5	0.3	0	0	0.1	-0.1	-0.1	0.1
CJ_10000488	0.7	-0.3	1.1	0.2	0	0.2	0.1	0	-0.2	0.2

CJ_10000489	-0.4	-0.2	-0.2	0.2	0	0.2	0	-0.1	-0.1	0
CJ_10000490	-0.5	-0.4	-4.2	0.1	0.1	-1.3	-1.7	-1.1	0	0
CJ_10000491	0.1	-0.1	0.5	0.1	0	0.2	0	0	-0.1	0.1
CJ_10000492	-0.5	-0.3	0.5	0	-0.1	0.3	-0.1	-0.2	0	0.1
CJ_10000494	-0.8	-0.3	-0.5	0	-0.2	0.3	0	-0.1	0	0.2
CJ_10000495	-0.2	-0.1	0.1	0.1	0	0.2	0	0	-0.2	0.1
CJ_10000496	-0.1	0.1	0	-0.1	-0.2	0.3	0	-0.1	0.3	0.1
CJ_10000497	-0.4	0	0.1	0.1	-0.1	0.2	0.1	0	-0.1	0.2
CJ_10000498	0.1	-0.1	0.1	0.1	0.2	0.1	-0.1	0	0.1	0.1
CJ_10000507	-0.2	-0.5	-0.1	0.1	-1.7	0.2	-0.1	0	-0.2	0
CJ_10000508	0	-1.1	0.4	0	0	0.3	0	-0.1	-0.1	0
CJ_10000509	-0.1	0	0.4	0.1	0.2	0	0.1	-0.2	-0.1	0
CJ_10000510	0.2	0	0.3	0.2	0.2	0.1	0.1	0.1	0	0.2
CJ_10000511	-0.5	-0.1	0.3	0.2	-0.2	0.3	0.2	-0.1	-0.2	0
CJ_10000512	-0.4	-0.1	-0.1	0.2	0	0.2	0.1	0.1	-0.2	0.1
CJ_10000513	0.1	0.1	0.1	0.2	0	0.1	0	-0.1	-0.1	0
CJ_10000514	0	-0.1	0.2	0.3	0.1	0	0.2	0	0	-0.1

CJ_10000515	-2.5	0.2	0.2	0.2	0	0.2	0.1	-0.2	0	-0.1
CJ_10000516	0.1	0	0.4	0.1	0.1	0.1	-0.4	-0.1	-0.1	0
CJ_10000517	0.2	0	0.8	0.3	0.2	0	0.2	0	-0.2	0
CJ_10000518	0.9	0.4	0.2	0.2	-0.2	0.2	0.1	0	0	0
CJ_10000519	-1	-0.1	-0.4	0.2	0	0.2	0	0	0	0.1
CJ_10000520	-0.7	0.1	0.5	0	0	0.3	0.2	0.2	-0.1	-0.1
CJ_10000522	-0.4	0.1	0.1	0.1	-0.1	0.1	-0.1	-0.2	0	0.1
CJ_10000526	-0.4	-0.1	0.1	0	-0.1	0.1	-0.1	-0.1	0	-0.1
CJ_10000528	-0.5	0	-0.1	0.2	0.8	0.2	-0.2	0	-0.1	-0.1
CJ_10000534	2.6	2.9	1.4	2	0.6	-0.8	-1.4	-1.4	2.9	4.1
CJ_10000538	-0.8	-0.6	0.1	0.1	0	0.2	-0.1	-0.1	-0.1	-0.1
CJ_10000539	-0.5	0.1	0	0.1	0	0.1	0	0.1	0.1	0.1
CJ_10000540	2.5	-2.2	2	0.5	0	0.6	0.6	-0.1	0.4	0.5
CJ_10000541	0.1	-0.2	-0.1	0	-0.1	0	-0.1	0	0	0.2
CJ_10000543	0	0.2	-0.3	-0.4	1	0.3	0.1	1.2	0.2	-0.5
CJ_10000544	-2.1	-1	-1.2	0.1	0	-1	0.2	-0.2	0	0
CJ_10000545	0.3	0	0.1	0.1	-0.1	0.2	-0.1	0	0	0.2

CJ_10000546	-0.1	0	-0.3	0.2	0.2	0.2	0.1	0	0.1	0.1
CJ_10000547	-0.6	-0.7	-0.5	0.1	0.2	0.2	0.2	-0.1	-0.4	0.1
CJ_10000548	-0.4	0	0.4	0.2	0.1	0.2	0	-0.3	0.2	0.1
CJ_10000549	-0.6	-0.1	-0.1	0.2	-0.3	0.1	-0.3	-0.4	0.1	-0.2
CJ_10000550	0	0	0.3	0.1	0.1	0	0	-0.1	0.1	0
CJ_10000551	-0.6	-0.3	-0.1	0.1	-0.1	0.3	-0.1	-0.1	0.3	0.1
CJ_10000552	1.7	0	0.4	0.1	0.9	0	-0.7	0.1	0	-0.1
CJ_10000555	0.2	1.3	0.2	0.4	-0.6	0	0.1	1.1	1.9	0
CJ_10000557	-1.1	0.2	-0.5	0.7	-0.1	0.2	-0.1	-0.8	0.8	0.7
CJ_10000564	0.2	-0.3	-0.1	0.1	0.1	0.2	0.1	-0.1	-0.1	0.1
CJ_10000565	0.3	0	-0.1	0.1	-0.1	0.2	0.1	0	0.3	0.1
CJ_10000566	-0.2	0	0.2	0.2	0	0.1	0.1	-0.1	0	0.2
CJ_10000567	-0.2	-0.2	-0.4	0.1	0.1	0.2	0.1	0.1	0.1	0.2
CJ_10000568	-0.1	-0.2	0.3	0.1	0	0.1	0.1	0	0	0.1
CJ_10000569	0	0.3	0.4	-0.1	-0.2	0.2	0	-0.2	0.2	0.2
CJ_10000570	-0.2	0.1	0.2	0.1	0.1	0.2	0.1	0	-0.1	0.2
CJ_10000571	-0.5	-0.3	0.1	0	0	0	0	-0.2	-0.1	0

CJ_10000572	-0.1	0	0	0.1	0	0.3	0	0.1	-0.1	0
CJ_10000573	0.2	0.2	0.2	0.1	-0.2	0	-0.3	-0.1	0	0
CJ_10000574	-1.5	-2.5	-0.2	0	0	0.1	-0.1	0	0	0.1
CJ_10000575	0.2	0.2	0	0.3	0	0	-0.2	-0.1	-0.1	0.1
CJ_10000576	0.1	-0.1	0.2	0	-0.1	0.1	0.1	0.2	0.1	0.1
CJ_10000578	0.2	0.2	0.2	0.1	0.2	0.1	-0.2	0	-0.1	0.1
CJ_10000579	-0.1	0.1	0.7	0.2	0.1	0.2	0	-0.1	-0.1	0.1
CJ_10000580	0.1	0.2	0.6	0.1	0.1	0.1	0.1	-0.1	-0.2	0.1
CJ_10000581	-0.4	0.3	0.2	0.1	0	0.2	-0.1	-0.1	-0.1	0.1
CJ_10000582	-0.4	-0.4	0.3	0.1	0.4	-0.2	0.2	0.3	-0.9	0
CJ_10000583	-0.7	-0.4	-0.2	0	0.1	-0.1	0.1	0.4	-0.6	0
CJ_10000584	-0.1	-0.1	-0.8	0.1	0.1	-0.5	-0.5	-0.3	-0.2	0
CJ_10000586	0.4	-0.2	0.4	0.3	0.3	-0.1	0	-0.4	-0.4	0
CJ_10000587	0.9	-0.2	0.2	0.4	0	0.3	0.2	0.1	0	0.1
CJ_10000589	0.5	-0.1	-1.3	-0.1	-0.2	0.1	-0.1	0.5	-0.1	0
CJ_10000590	-0.6	-0.1	-0.5	0.1	0	0.1	-0.1	-0.1	0.2	0
CJ_10000591	-0.1	-0.5	-0.1	0.2	-0.4	0.1	0	-0.8	-0.4	0

CJ_10000592	-0.1	0	-0.2	0.2	-0.1	0.4	1.3	0.4	0.7	0.5
CJ_10000593	-0.1	0	-0.3	0	0.1	0.3	0.1	-0.1	0	0.1
CJ_10000594	-0.3	-0.2	0.3	0.1	0	0.2	0.1	0	-0.2	0
CJ_10000595	-1	-0.5	0.2	0	0	0.2	0	-0.3	0.1	0
CJ_10000596	-0.4	-0.2	0.2	0.1	0	0.2	0.1	0	0	0.2
CJ_10000597	0.3	0.4	0.7	0.1	0	0	0	0	0.1	0
CJ_10000598	-0.6	-0.2	-0.6	0.2	-0.1	0.2	-0.2	-0.3	0.2	0
CJ_10000599	-0.4	-0.1	-0.4	0.1	0.1	0.2	0	0	-0.2	0.2
CJ_10000600	-0.2	-0.3	-0.1	0.1	-0.1	0.2	0.1	-0.1	0.1	0
CJ_10000601	-0.7	-0.3	-0.3	0	0	0.3	0	-0.1	0.1	0.1
CJ_10000602	0.1	0	0	0.2	0	0	-0.1	-0.1	0	0.1
CJ_10000603	-0.1	0.1	-0.8	0.1	-0.1	0.2	0	0	0.2	0.1
CJ_10000604	-0.3	-0.1	-0.2	0	-0.1	0.2	-0.1	-0.1	0.2	0.1
CJ_10000605	-0.2	0.1	-0.2	0.1	0	0.1	-0.1	-0.1	0	0.1
CJ_10000606	0.1	-0.4	-0.3	0.2	0	0.1	-1	-0.2	-0.4	0
CJ_10000607	-0.2	-0.1	0.4	0.3	0.1	0.1	-0.1	0.2	0	0
CJ_10000608	-0.4	-0.2	-0.2	0.1	0.1	0.2	0	0.1	-0.1	0.1

CJ_10000609	-0.2	0	-0.4	0	-0.1	0	0	-0.1	0	0
CJ_10000610	0.1	-0.3	0.4	0.2	0.2	0.1	0	-0.1	-0.2	0
CJ_10000611	0.1	-0.1	0.1	0.2	0.1	0.1	0	0.2	-0.4	0
CJ_10000614	0	5.2	-0.2	0.2	0	0.1	0	0.2	0.1	0
CJ_10000615	-0.2	-0.3	0.2	0	-0.3	0.3	0	0.1	-0.1	0.1
CJ_10000616	0.3	-0.3	0	0.3	-0.3	0.5	0.4	0.2	0.4	0
CJ_10000617	0	0.1	0.2	0.1	0	0.2	0.1	0.1	0	0.1
CJ_10000618	-0.2	-0.2	-0.5	0.2	-0.2	-0.1	0.1	-0.2	-0.3	0.1
CJ_10000619	-0.1	0.2	-0.2	0.2	0.1	0.1	0.2	-0.1	0.2	0.1
CJ_10000620	-0.1	0	0	0.1	0	0.3	0	0.1	-0.2	0
CJ_10000621	-0.4	-0.2	-0.3	0	-0.3	0.4	0.1	0.1	0.3	0.2
CJ_10000622	0.1	0.2	0	0.1	-0.1	0.2	0.1	0	0	0.1
CJ_10000625	3.4	3.8	0.8	1.2	-1.6	1.3	-0.1	-0.9	1.1	1.4
CJ_10000626	1.9	2.4	0.3	0.8	-0.6	0.8	0.1	-0.5	0.8	0.7
CJ_10000627	-0.4	-0.3	0.2	0	0	0.1	0	0	0.1	0
CJ_10000628	-0.1	-0.2	0	0.1	0	-0.1	-0.1	0	0	0.1
CJ_10000629	0.2	0.4	0.4	0.1	0.1	0.2	0.1	-0.2	-0.1	0.1

CJ_10000630	0	0.3	1.2	0.3	0.4	-0.2	0	0.3	-0.3	0.2
CJ_10000631	0.1	0.1	0.3	0.2	0.2	-0.1	0.1	0.1	-0.3	-0.1
CJ_10000632	0.5	0.1	0.1	0	0	0.2	-0.3	0	-0.2	0.1
CJ_10000633	-1.1	-0.2	-2.2	0.4	0.1	0	-0.2	-0.3	0.1	-0.2
CJ_10000634	-0.5	-0.3	-0.8	0.1	0	-0.1	-0.2	0.2	0.1	0
CJ_10000635	0.1	-0.2	-0.3	0.2	0.1	0.2	-0.1	0.1	-0.2	0
CJ_10000637	0.1	-0.4	0.2	0	0.1	0.3	0.3	0	0	0.1
CJ_10000639	-0.3	-0.5	-1.1	0	-0.2	-0.2	-0.1	0.3	-0.7	0.1
CJ_10000640	0.1	0.2	-2.2	0.5	0.1	-0.3	-1.2	-2.2	0.1	0
CJ_10000641	0.2	0.1	0.2	0.1	-0.1	0.2	0	0	0	0.1
CJ_10000642	-0.2	0	0.2	0.2	0.1	0.3	0.2	0	0	0.3
CJ_10000643	0.4	0.3	-0.4	0.3	-0.2	0.5	0.2	-0.2	0	0.3
CJ_10000644	-0.4	-0.1	-0.7	0.1	0	0.2	-0.3	0	-0.1	0
CJ_10000645	0.2	0.4	0.2	0	-0.3	0.2	0	0.2	0.3	0.2
CJ_10000646	0.1	0.2	-0.3	0.3	-0.2	0.3	0	-0.3	0	0.3
CJ_10000647	-0.4	0.5	-0.8	0.2	-0.1	0.2	-0.1	-0.3	0.2	0.2
CJ_10000648	0.4	0.2	0.7	0.1	0.1	0.2	0.1	0	-0.1	0.1

CJ_10000649	-0.4	0.3	-0.1	0.2	-0.2	0.3	0.1	0	0	0.3
CJ_10000650	-0.7	0.3	0	0.2	0.2	0	-0.2	-0.2	0	0
CJ_10000651	-0.3	-0.3	-0.2	0.1	0	0.1	0	-0.2	0.2	0
CJ_10000652	-0.4	0.3	-0.2	0.1	-0.2	0.2	0	-0.2	0.3	0.1
CJ_10000653	0	0.2	0.2	0.1	0	0.2	0.1	-0.1	0.1	0.1
CJ_10000655	-0.1	-0.4	-0.7	0	0.1	0.3	0.1	-0.2	0	-0.1
CJ_10000656	-0.1	-0.2	-2.9	0.2	0.2	-1.4	-1.2	-1.1	-0.2	-0.1
CJ_10000657	0.2	-0.2	0.5	0.1	0.1	-0.3	-1.2	0.4	-2	-0.5
CJ_10000658	0.1	0.1	0.2	0	0.1	0	0	-0.6	-0.2	-0.2
CJ_10000659	0.1	0	0.1	0.2	0	0.2	0	0.1	0	0
CJ_10000660	0.7	-1.3	-0.5	-0.3	-0.1	0	0.8	0.3	-0.7	-0.4
CJ_10000661	-0.2	0.2	0	0.2	0	0.3	0.6	0.5	-0.1	0
CJ_10000662	1	0.8	0.2	0.3	-0.2	0.1	0.1	-0.3	0.1	0.2
CJ_10000663	-0.3	0.5	0	0.1	-0.3	0	0.2	0.2	0	0
CJ_10000664	0.1	0.4	0.4	0.3	0.1	0.1	0.1	0	0	0.1
CJ_10000665	-0.2	-0.3	0	0.1	0.1	0.2	0.1	0	-0.1	0
CJ_10000666	0.1	-0.1	0.5	0.3	0	0.2	0.2	0	-0.1	0.1

CJ_10000667	-0.4	-0.1	0.1	0.1	0.1	0.2	0.1	-0.1	0.1	0.2
CJ_10000668	-1.1	-0.5	0.4	0.1	-0.1	0.1	0.1	0	0.1	0
CJ_10000669	-0.1	-0.1	0.3	0.1	-0.1	0.3	0.1	0	0.1	0.1
CJ_10000670	-0.4	-0.2	-0.2	0.2	0.1	0.2	-0.1	-0.1	0	0.1
CJ_10000671	-2.3	-1.3	-0.8	0.1	-0.2	0.3	-0.1	0.1	0.3	0
CJ_10000672	-0.8	-0.1	-0.7	0.1	-0.1	0.2	-0.1	0	0.3	0.1
CJ_10000673	-0.1	0	0	0.1	0	0.2	0.1	0	-0.1	0.2
CJ_10000675	-0.2	0.3	-0.2	0.1	-0.2	0.5	0	-0.1	0.1	0.2
CJ_10000676	0	0	0.4	0	0.2	0.1	0.1	0.2	-0.1	0
CJ_10000677	0.2	0	0.5	0.2	0.2	0	0.1	0	-0.1	0.1
CJ_10000678	-0.5	-0.5	-0.1	0.1	0	0.1	0.1	0	0	0
CJ_10000679	-0.5	-0.6	0.1	0.1	0.2	0.2	0.1	0.4	-0.1	0
CJ_10000680	-1.1	-0.4	0.3	0.1	0.2	0.1	0.2	0.2	-0.3	-0.1
CJ_10000681	0	-0.1	0.4	0	0	0.1	0.1	0.1	-0.3	0
CJ_10000682	0.3	-0.5	0.3	0	0.1	0.1	0.1	0.2	-0.1	0
CJ_10000683	-0.5	0	-0.2	0.1	0	0.2	0.1	0	-0.1	0
CJ_10000684	-0.2	-0.1	-0.2	0	-0.2	0.2	0	0.1	0	0

CJ_10000685	-0.1	0.1	-0.1	0	-0.1	0.2	-0.1	0.1	-0.1	0
CJ_10000686	-0.7	-0.4	0.2	0	0	0.2	0.1	0.1	-0.1	0.1
CJ_10000687	0.2	-0.1	0.3	0.1	0	0.1	0.1	0	-0.1	0
CJ_10000688	-0.3	-0.3	-0.1	0	-0.1	0.2	-0.1	0	0	0.1
CJ_10000689	0.3	0	0	0.1	0.2	0	0	0.1	0	0
CJ_10000690	1.9	1.1	1.3	0.4	-0.4	0.3	0.3	-0.2	0.2	0.4
CJ_10000691	0.9	0.5	0.1	0.2	0	-0.1	0.1	-0.3	0.1	0.2
CJ_10000692	-0.8	-0.9	0.2	0.1	0.1	0.1	-0.1	0.2	-0.3	-0.1
CJ_10000693	0.3	0.1	0.5	0.3	0.2	0.1	0.1	0	0	0.1
CJ_10000694	0.3	0.4	0.1	0.3	0.1	0.2	0.1	-0.1	0.2	0.1
CJ_10000695	0.1	0.1	0.3	0.2	0	0	-0.2	0.1	0.1	0
CJ_10000696	0.3	0.3	0.6	0.2	0.1	0	0.1	0	-0.1	0
CJ_10000697	-0.3	0	-0.2	0	0	0.2	0	0	0	0.1
CJ_10000698	0	-0.6	0.3	0.1	0.1	0	-0.1	0	0	0
CJ_10000699	1.1	1.2	0.2	0.3	-0.7	0.5	0.4	-0.7	0.6	0.5
CJ_10000700	-0.4	0.1	-0.1	0.2	-0.1	0.4	0.2	-0.2	0.2	0.4
CJ_10000701	-1.2	-0.5	-0.4	0.1	-0.1	0.2	-0.1	0.1	-0.2	0.1

CJ_10000702	0	0	0.5	0.3	0.1	0.1	-0.1	0.1	0	0
CJ_10000703	-0.4	0.1	0.4	0.1	0.2	0.2	0	0	-0.1	0
CJ_10000704	-1.4	-0.4	-0.3	0.2	-0.1	0.1	0	0	0	0
CJ_10000705	-1.4	-0.4	-0.1	-0.1	0	0.1	0	0	-0.1	0
CJ_10000706	-1.6	0	-0.6	0.2	0.1	-0.1	-0.1	0	0	0.1
CJ_10000708	0.3	-0.2	0	0.2	0.3	0.2	0.2	-0.2	-0.1	-0.1
CJ_10000709	-0.9	-0.1	-0.4	0.3	0.2	0	-0.2	0	-0.1	-0.1
CJ_10000710	-0.4	-0.2	-0.2	0.2	0	0.2	-0.2	-0.2	0	0.2
CJ_10000711	-0.4	0	-0.1	0.2	-0.2	0.2	0	-0.1	0	0.2
CJ_10000712	-0.1	0.1	-0.8	0.1	-0.1	-0.1	-0.2	-0.5	0.1	0.1
CJ_10000713	-1.6	-0.4	-4.2	0.1	-0.1	-1	0	0.1	0.4	0.1
CJ_10000714	-0.8	-0.1	-0.5	0.1	-0.2	0.4	0	0	0.1	0.3
CJ_10000715	-0.3	0	-2.9	0.1	-0.2	-1	0.1	-0.2	0.1	0.2
CJ_10000716	-0.4	-0.4	0.5	0.3	-0.2	0.2	0.2	0	0	0
CJ_10000717	-0.5	0	0	0.1	0.1	0	0	0.1	0	0
CJ_10000718	-0.1	0.5	0.3	0.1	-0.1	0.2	0.2	-0.1	0.2	0.2
CJ_10000719	0.1	-0.8	0.9	0	0	0	0.1	0	-0.1	-0.1

CJ_10000720	-0.1	0.2	0	0	-0.1	0.3	-0.1	-0.1	-0.1	0.2
CJ_10000721	0.4	0.5	0.7	0.3	0.1	0.1	0.2	0	0.1	0.1
CJ_10000722	-0.2	0.5	0.3	0.1	-0.2	0.2	-0.1	0	0.5	0.1
CJ_10000723	0.3	-2	0.5	0	0	0.1	0	0	-0.1	0.1
CJ_10000724	0.1	-0.1	0.7	0.1	0.3	0.1	-0.1	0.1	-0.2	0
CJ_10000725	1.7	2	2.3	0.2	-0.5	0.5	1.1	-0.5	0.7	0.6
CJ_10000726	-0.1	-0.1	-0.3	0.1	-0.1	0.2	0	0.1	-0.2	0
CJ_10000727	0.8	0.2	-1.8	0.2	0	-2.9	-0.4	-0.2	-0.2	0.2
CJ_10000729	-0.5	-0.3	-0.3	-0.1	0.1	0.1	-0.1	0.1	-0.2	-0.1
CJ_10000730	-0.8	-0.3	0	0.2	0.3	0.1	-0.1	0.1	-0.2	0
CJ_10000731	0.4	0.4	0.6	0.4	-0.3	0.2	0.2	0.1	0	0.1
CJ_10000732	-0.1	0.2	-0.1	0.2	0	-0.2	-0.9	-0.7	0.1	0
CJ_10000733	-0.9	0.1	-0.2	0.1	0	0.3	0	-0.2	-0.1	0.1
CJ_10000734	-1	-1	-0.1	0	-0.1	0.2	0.1	0.1	-0.1	0
CJ_10000735	-0.2	0.1	0.6	0.3	0.1	0.2	0.1	0.1	-0.1	0.1
CJ_10000736	-0.1	-0.2	0.1	0.3	0.1	0.2	0	0	-0.1	0
CJ_10000737	0.2	-0.2	0.7	0.2	0	0.1	0.2	0.1	0.8	0.1

CJ_10000738	0.1	0.1	-0.1	0.2	0	0	0.1	0.1	0	0.1
CJ_10000739	-0.6	-0.1	0	0.1	0	0.3	0.2	-0.1	0	0.2
CJ_10000740	-0.3	0.1	0.8	0.1	-0.1	0.3	0.2	0	0	0.1
CJ_10000741	-0.1	0.6	0.2	0.1	-0.2	0	0.2	-0.1	0.2	0.2
CJ_10000742	-0.8	0.3	0.4	0	0	-0.5	-0.8	-0.9	0	0.2
CJ_10000743	-0.3	0.2	0.1	0	0.1	0	-0.3	-0.2	-0.3	0
CJ_10000744	0.4	0.8	0.5	0.1	0	0.1	0.1	0	0	0.1
CJ_10000745	-0.1	-0.1	0.2	0.1	0.2	0.1	-0.1	0.1	-0.3	0.1
CJ_10000746	0	-0.1	0.2	0.1	0	0.1	-0.1	-0.1	0	0
CJ_10000747	-0.2	-0.1	-0.3	0.1	-0.1	0.2	-0.2	-0.2	0.1	0.1
CJ_10000748	0.2	0.4	0	0.2	-0.1	0.3	0.1	-0.1	0.1	0.2
CJ_10000749	0	0.1	0.2	0.1	-0.1	0.1	-0.2	-0.1	-0.2	0.1
CJ_10000750	-0.6	-0.2	-0.3	0	0	0.1	0	0.2	-0.1	0
CJ_10000751	-0.2	-0.2	-0.2	0.1	0	0.2	0	0.1	0	0.1
CJ_10000753	2.1	1.4	0	-0.1	-0.1	1.1	0.3	-1.4	1.1	-0.4
CJ_10000757	0.4	0.6	-0.4	0.2	-0.4	0.3	-0.1	-0.1	0.2	0.3
CJ_10000758	-1	-0.8	0.3	0.2	0	0.2	0.1	0.1	-0.1	0.1

CJ_10000759	-0.1	0	0.1	0.1	-0.1	0.2	0	0	0	0.1
CJ_10000760	0	0.1	0.2	0.2	-0.1	0.3	0.1	0	-0.1	0.1
CJ_10000765	-0.8	-0.1	0.1	0.4	0.3	0.5	-0.1	0.1	0.1	0.2
CJ_10000786	-0.1	-5.2	-0.4	0.3	1	-0.2	-0.5	0.4	-0.3	0.1
CJ_10000790	0	0.2	0.1	0.3	0.1	0	-0.5	-0.1	-0.2	0.2
CJ_10000791	0.6	-0.4	0.4	0.2	0.1	0.1	-0.3	0.2	0	0
CJ_10000794	-0.2	0.6	0.8	0.4	-0.1	0.3	0.3	0	-0.1	0.2
CJ_10000806	-0.1	-0.1	0	0.1	-0.1	0.4	0.1	0.1	0.2	0
CJ_10000816	-0.7	1.3	1.5	0.5	-0.1	0.1	0.1	0.8	0	0.4
CJ_10000819	1.5	1	2	0.5	0	0.3	0	1.1	-0.2	0.1
CJ_10000822	0.3	0.3	0	0.2	0	0.3	-0.1	0	0	0.1
CJ_10000825	2.8	2.5	1.6	0.8	-0.8	0.1	0.7	-0.7	0.6	0.8
CJ_10000826	3	2.7	0.8	0.1	-1.5	-0.2	0.5	0.6	0.9	0
CJ_10000828	-0.1	1.2	1.1	0.1	0.2	0.1	0	0.7	0.2	-0.1
CJ_10000842	0	-0.4	0	0.2	0.3	0.2	-0.2	0	-0.2	0.1
CJ_10000846	1.2	0.7	1.6	1	0.5	0.7	0.1	0.4	1.2	-0.1
CJ_10000847	2.8	0.3	-0.9	0.1	0	-1.1	-0.3	1.5	1.8	-1.5

CJ_10000875	3.3	4	2.9	0.9	-1.5	1.1	1.3	-0.9	1	1.4
CJ_10000880	1.4	1.3	3.4	1.6	-3	2.8	1.5	0.5	2.2	0.9
CJ_10000885	-0.7	0.1	0	0.3	-0.2	-0.1	-0.5	-0.8	0.7	0.5
CJ_10000897	-0.6	0.9	2.6	0.5	-1	0.1	0.3	0.2	1.5	1
CJ_10000914	0.1	0.1	-0.8	0.2	0.2	-0.2	-0.3	0.3	0.1	0.1
CJ_10000933	0	-1.8	1.4	0.3	-0.3	0.2	0	0	0.5	0.3
CJ_10000937	0.5	1.1	0.9	0.4	-0.3	0.2	0	-0.5	0.3	0.4
CJ_10000938	0.3	0.2	0.2	0.2	-1.5	0.3	0	-0.1	-0.1	0.1
CJ_10000960	-0.1	0.2	-0.1	0.1	-0.2	0.4	0.1	0	0.1	0.2
CJ_10000970	-2.5	-0.6	-1.1	0.2	0.1	-0.2	-0.6	0.2	0.2	0.2
CJ_10000981	0.5	0.2	0.6	0.4	0.1	0.2	0	0	0	0
CJ_10000982	0.3	0	-0.2	0.1	0.1	-0.1	-0.3	-0.3	0	0
CJ_10000983	1.5	-0.1	1.1	0.5	0	0.2	0.7	0.2	0.3	0.3
CJ_10000991	-0.3	-0.2	0.3	0.4	0.3	0.2	0	0.6	0	-0.1
CJ_10000992	-0.2	0.3	0.3	0.2	0.2	0	0.1	0	-0.2	-0.1
CJ_10000995	0.6	0.3	0.7	0.1	0.2	-0.1	0.3	0.1	-0.2	-0.2
CJ_10000996	0.3	0.2	0.4	0.1	0.1	0.2	0	0.1	-0.1	-0.1

CJ_1000997	0.3	0.5	0.2	0.3	0	0.1	-0.1	0.3	0	0.2
CJ_10001000	-0.1	-0.1	-0.1	0.2	0	0.1	0	-0.1	0.2	0.1
CJ_10001006	-0.2	0.3	-0.3	0.3	0.1	0.1	-0.1	-0.4	-0.1	0.1
CJ_10001008	0.4	0.3	0.5	0.1	-0.1	0.2	0.1	0	0	0.1
CJ_10001009	1.1	1.4	1.7	0.4	-0.3	0.4	0.3	0.5	0.3	0.6
CJ_10001011	0.4	0.7	0.3	0.4	0	0.2	-0.2	0	0.1	0.3
CJ_10001014	0.2	0.5	0.8	0.2	0.1	0.2	0.1	0.5	0	0.1
CJ_10001017	-1.2	-0.7	0.4	0.2	0.3	0.2	-0.1	0.4	-0.3	0
CJ_10001018	-0.1	0.1	0.1	0	0	0.2	0	0.1	-0.1	0.1
CJ_10001019	-0.2	-0.4	-0.8	0.2	-0.1	0.1	0	0.6	0	0.3
CJ_10001029	-0.7	0	0.6	0.3	0	0.2	0.1	0.3	0.1	0
CJ_10001030	0.5	0.5	0.2	0	0	0	0	0	-0.1	0
CJ_10001032	0	-0.1	0.1	0.2	-0.1	0.1	0	-0.1	0.1	0.2
CJ_10001033	0.1	-0.4	0.4	0.1	0	0.2	0	-0.1	0	0
CJ_10001045	-0.6	-0.5	0	0.2	0.2	0.1	-0.2	0.6	-0.1	0.1
CJ_10001047	-1.3	-0.3	0.1	0.1	0.1	0.3	0	0.1	-0.3	0.1
CJ_10001055	-0.4	-0.1	-0.2	0.2	-0.1	0.4	-0.1	-0.2	0.2	0.1

CJ_10001061	-0.7	-0.1	-0.5	0.3	0.1	0.3	0	0.1	0.1	-0.1
CJ_10001062	1.7	1.2	1.1	0.6	0	0.4	0.4	0	0	0.2
CJ_10001066	3.9	0.9	0.8	0	-0.1	0.4	0.6	0.6	0.1	-0.1
CJ_10001074	0.5	0.5	0.1	0.2	0.2	0	0.1	0	-0.1	-0.1
CJ_10001075	0.2	0.2	0.8	0.2	0	0.1	0.1	0.1	0.1	0
CJ_10001082	-0.1	0.5	0.7	0.5	0	0.1	-0.1	0.3	0.3	0.1
CJ_10001101	-0.2	0.4	0	0.3	-0.1	0.3	0.1	-0.1	0.1	0.1
CJ_10001104	2.4	1.6	1.3	0.2	-0.3	0.9	1.3	1	0.7	0.9
CJ_10001107	2.7	2.4	1.7	1	-1.3	1	0.9	-0.8	0.8	1
CJ_10001109	0.2	0	0.3	0.1	0.1	0.1	0.1	0	-0.2	-0.1
CJ_10001112	0	-0.1	0.1	0.1	0.1	0.2	0.1	0.2	-0.1	-0.2
CJ_10001122	2.1	1.4	0.8	0.3	-0.4	-0.7	0	0.3	0.2	0.2
CJ_10001128	-0.5	-0.2	-0.9	0.1	0	0	0	0	0.1	0.1
CJ_10001136	-0.9	-0.1	0.3	0.1	0	0.1	0.4	0.1	0.1	-0.1
CJ_10001141	-0.4	-0.5	-0.2	0.1	0.1	0.1	0	-0.1	-0.1	0.1
CJ_10001142	0.5	0.4	0.7	0.1	0.1	0.1	0.1	0	-0.1	-0.1
CJ_10001144	0	-0.1	0.5	0	0.2	0.1	0.1	-0.1	-0.2	0.1

CJ_10001145	-0.1	0.1	0.2	0.1	0.2	0.2	0	-0.1	0	0
CJ_10001146	-0.2	0.1	0	-0.1	-0.1	0.1	-0.2	0	0.4	-0.1
CJ_10001148	0.2	0.3	0.4	0.1	0.1	0.2	-0.1	-0.1	-0.2	-0.1
CJ_10001149	0.5	0.4	0.6	0.2	0.1	0	0.1	-0.2	1.5	0
CJ_10001150	0.1	0.1	0.4	0	0	0.2	0	0	0	0
CJ_10001151	0.2	0.2	0.5	0.1	0.1	0	0	0	-0.1	0
CJ_10001152	1.5	1.8	0	0.2	-0.8	0.3	0.2	-0.2	0.5	0.2
CJ_10001153	-0.5	0.1	-0.4	0.1	0	0.1	0	0	0.1	0
CJ_10001154	0	0	0.1	0	0	0.1	0	-0.1	0	-0.1
CJ_10001155	1.9	2.1	1.2	0.3	-0.7	0.3	0.4	-0.3	0.6	0.2
CJ_10001157	-0.3	0.2	-0.1	0	-0.1	0.2	0.1	0	0	0.1
CJ_10001158	0.6	0.9	-2	0.1	-0.4	-1.8	-2.9	-0.4	-0.5	0
CJ_10001161	-0.2	0.2	0	0.1	0	0.3	-0.1	-0.3	0	0
CJ_10001162	0.3	0	0.4	0	-0.1	0.2	0	0	0	0.1
CJ_10001164	-0.3	0	-0.2	0	-0.1	0.2	0	-0.1	0.2	0.1
CJ_10001166	-0.2	0.1	-0.4	0.1	0	0.1	-0.3	-0.4	0	0
CJ_10001168	-0.4	-0.6	-0.5	-0.1	0.1	0.2	-0.1	0	0.2	0.2

CJ_10001169	-0.3	0.6	-0.1	0.1	-0.2	0.3	0	-0.2	0.3	0.1
CJ_10001170	-0.4	-0.7	0.4	0.2	-0.2	0.5	-0.1	-0.1	0.7	0.3
CJ_10001171	-0.5	0.5	0.3	0	0	-0.2	0.1	0	-0.1	0.1
CJ_10001172	0.1	-0.2	0.1	0.1	0.2	0	0	0	-0.1	0.1
CJ_10001173	-0.3	-0.3	-0.2	0	0.1	-0.2	-0.2	-0.2	1.7	0
CJ_10001174	0.2	0.2	0.7	0.2	0.2	0.1	0.1	0	0	0.1
CJ_10001175	0	0.2	0.4	0.1	0	0.2	0	0	0	0
CJ_10001176	0.4	0	-0.2	0.1	0.1	-1.4	1.4	1.5	0.8	0
CJ_10001177	0	0.1	0.2	0.1	-0.2	0.2	0.1	0	0	0
CJ_10001178	0.3	-0.1	0.5	0.1	0.1	0.1	0	-0.1	0.1	0
CJ_10001179	-0.5	0	-0.7	0.1	0	0.1	-0.1	0.1	-0.1	0
CJ_10001180	-0.1	-0.1	-0.4	0.1	0	0.1	0	0	-0.1	0
CJ_10001181	-0.2	0.2	-0.1	0.1	-0.1	0.2	0	0	0.1	0.1
CJ_10001182	-0.3	0.4	0.6	0.1	-0.2	0.2	0.1	0	0.1	0
CJ_10001183	0	-0.2	0.1	0.1	0	0.1	0	0.1	0	0.1
CJ_10001186	0	0	0.1	0.1	-0.1	0.2	0.1	0	0	0
CJ_10001188	-0.7	-0.9	-0.1	0	-0.1	0.2	0	0	0.2	0.1

CJ_10001189	0.3	0.1	0.1	-0.1	-0.2	0.2	0.1	-0.1	0	0.2
CJ_10001191	-0.1	0.2	-1.2	0	-0.1	0.2	0	-0.2	0	-0.1
CJ_10001192	-1.5	-0.2	0.5	0	0	0.3	0	0.1	0	0.1
CJ_10001193	-0.7	-0.3	0.4	0.1	0	0.3	0	0	0	0
CJ_10001195	-0.5	-0.1	0.1	-0.1	-0.1	0.2	-0.2	-0.1	0.2	0
CJ_10001196	-0.5	0.4	-0.5	0.3	-0.2	-0.5	0.2	-0.2	0.3	0.2
CJ_10001198	-0.9	2.8	0.3	0.8	-1.1	0.1	0	-1.7	-0.2	0.8
CJ_10001199	0	-0.2	0.5	0.3	0.2	0.2	-0.1	0.2	0	0.2
CJ_10001201	-0.7	-0.2	0	0.1	-0.5	0.3	0	0.1	0.2	0.1
CJ_10001206	0.1	0.2	0	0.1	-0.1	-0.1	-0.1	0.2	0.1	-0.1
CJ_10001207	-0.7	0.4	-0.8	0	-0.3	0.2	0	-0.3	0.2	0.1
CJ_10001208	2.9	2.6	1.5	0.8	-0.8	0.8	1	-0.6	0.6	0.9
CJ_10001210	0.3	0.2	0.4	0.1	-0.2	0.2	0	-0.2	0.1	0.1
CJ_10001211	-0.4	1.1	3.6	0.3	-0.7	0.5	1.1	0.5	0	0.4
CJ_10001213	0.2	0.5	0.4	0.1	0	0.1	0.1	0	0.1	0.1
CJ_10001214	-0.4	0.1	0.3	0.1	0	0.3	0.1	-0.2	0	0
CJ_10001216	0.2	0	0.3	0.1	0.3	0.1	0.1	0	-0.1	0.1

CJ_10001217	0.4	0.4	0.5	0.2	0.3	0	0.1	0	0	0.1
CJ_10001218	-0.3	0.2	0.4	0	0	0.2	0	0	1.3	0
CJ_10001220	-0.7	-0.3	0.1	0.2	0.2	0.4	0.1	0	0	0.1
CJ_10001222	0.3	0	0.6	0	0	0.2	-0.1	-0.2	0	0
CJ_10001223	-0.5	-0.5	0.1	-0.1	0	0.2	0.2	-0.1	0.1	0.1
CJ_10001224	0	-0.3	0.1	0.2	0	0.1	0.1	0	0.2	0.1
CJ_10001225	-0.4	-0.2	-0.3	0.4	0.3	0.1	0	0.1	0	0.1
CJ_10001226	0	1	0.2	0.3	-0.6	0.2	0.1	-0.5	0.4	0.3
CJ_10001227	0	-0.2	-1.7	0.2	-0.1	0	0.1	-0.1	0.1	0
CJ_10001228	0.1	0.1	0.3	0.1	-0.1	0.2	0	0.1	0	0
CJ_10001229	1.6	1.4	0.6	0.3	-0.8	0.4	-0.2	-0.7	0.2	0.3
CJ_10001230	1.1	0.6	0	0	-0.3	0.2	0.1	0	0.3	0.1
CJ_10001231	-0.2	0.1	-0.1	0.1	-0.1	-0.1	0	0	0.1	0
CJ_10001232	0.4	0.3	0.3	0.1	0	0.1	0.2	-0.1	0	0.1
CJ_10001233	1.3	1.3	0.2	0.2	-0.5	0.3	0.3	-0.3	0.3	0.1
CJ_10001234	0	-0.3	0.8	0.3	0.1	0.1	-0.1	0.7	0	0.1
CJ_10001235	0	0.1	-0.3	0.1	0	0	-0.1	-0.1	0.1	0

CJ_10001236	0.5	0.5	-1.2	0	-0.3	-0.2	0.2	0	0.1	0
CJ_10001239	-0.3	-0.2	0.4	0	0	0.2	0.1	-0.3	0.2	0.1
CJ_10001241	0.1	0.9	0.1	0.3	0	-0.1	-0.1	-0.3	-0.4	0.1
CJ_10001243	3.1	2.9	3.1	1	0.2	1.5	0.8	-0.7	1.1	1.1
CJ_10001244	-2.4	0	-0.9	0	0	-0.8	-0.2	-0.1	0	0.1
CJ_10001246	-0.1	-0.2	0.4	0	-0.9	0	-0.1	0.1	-0.2	0
CJ_10001249	-0.5	-0.1	-0.5	0	-0.1	0.2	-0.1	-0.1	0	0
CJ_10001250	-0.1	0	-0.2	0	-0.1	0.1	0	0.1	-0.2	-0.1
CJ_10001251	-1.4	0.2	-0.9	0.1	-0.1	0.1	-0.2	-0.1	0	0
CJ_10001252	-0.3	0.3	-0.5	0.1	-0.1	0.1	-0.1	-0.2	0.1	0.1
CJ_10001254	-0.1	0.2	-1.4	-1.1	0	-0.6	0	1.2	0	-1.4
CJ_10001256	0.3	0.3	0.4	0	-0.1	0.1	0	0	0.1	0
CJ_10001257	-0.8	0	-0.8	-0.2	-0.1	0.2	0	-0.1	0.4	-0.1
CJ_10001258	0.8	0.6	-0.2	-0.2	0.4	-0.3	-0.5	0.7	-0.1	-0.5
CJ_10001259	-0.2	-0.1	-0.8	-0.1	0	0.2	-0.1	0	0.1	0
CJ_10001260	0	0.1	0.5	0	-0.2	0.4	0.1	-0.1	0	0.2
CJ_10001265	-0.6	-0.1	-1.2	-0.4	0	0	-0.5	0	0.4	-0.2

CJ_10001266	-0.5	-0.2	-0.2	0.1	0.1	0.2	-0.1	-0.1	0.2	0.1
CJ_10001268	0.3	-0.3	0.8	0.1	0.2	0.1	0.1	-0.1	0	-0.1
CJ_10001269	0.9	0.7	0.3	0.4	-0.1	0.2	0.1	-0.5	0.2	0.2
CJ_10001271	0.9	0.1	0.4	-0.1	-0.1	0.1	0	0.3	0	0
CJ_10001272	0.2	0	-0.1	0.1	-0.2	0.3	0.2	-0.2	0	0.2
CJ_10001273	0	0.1	0.2	0	-0.1	0.2	0.2	0	-0.2	0.1
CJ_10001274	0.1	0.3	-5.1	0.1	-0.2	-1.7	0.1	-0.1	0.2	0
CJ_10001275	-0.1	0	-0.4	0.1	0	0.1	-0.1	-0.2	0	0.1
CJ_10001279	2.5	2.2	0.9	0.7	-1	0.7	0.9	-0.6	1	0.8
CJ_10001280	0.1	0.3	-4.1	0.1	-0.1	-0.9	-1.7	-1.1	0.1	0
CJ_10001281	-0.5	-0.1	-0.7	0	-0.1	0.2	-2.4	-0.1	0.1	-0.1
CJ_10001283	4.7	4.5	-0.8	1.6	-1.5	1.7	0.2	-1.2	2.3	1.5
CJ_10001284	0.2	0.8	0.7	0.1	-0.1	0.3	0.1	-0.3	0.2	0.1
CJ_10001285	0.4	0.2	0.4	0.1	0.2	0.2	0.1	0	0	0.1
CJ_10001286	-0.1	0	0.1	-0.1	0.1	0.2	0.1	-0.2	0.2	0
CJ_10001287	-0.3	-0.1	0.4	0	0.1	0.1	0	-0.2	0	0.1
CJ_10001288	-0.4	0	-0.6	-0.1	0.2	0.1	0	0.2	0.3	0.1

CJ_10001289	-0.6	0.6	0.4	-0.1	-0.2	0.3	0.2	0.1	0.5	0.1
CJ_10001290	0.3	0.3	0.3	0.2	0.1	0.2	0.1	-0.1	-0.1	0.1
CJ_10001292	-0.7	0.4	0.2	0.1	-0.1	0.2	0.1	-0.3	0.2	0.1
CJ_10001293	0.2	0.2	0.4	0.1	0.1	0.1	0.1	-0.1	0	0
CJ_10001294	-0.1	-0.2	0.1	0	-0.8	-1.3	0	0	-0.1	0
CJ_10001295	0.9	0.1	0.5	0.1	0	0.3	0.2	0	-0.2	0
CJ_10001296	0	0.1	-0.1	0	-0.1	0.1	0.1	0	0	0
CJ_10001297	2.9	2.8	1.2	0.9	-1.2	1.1	1.4	-0.1	0.9	1.2
CJ_10001298	0.8	0.6	0.8	0.2	0	0	0.2	0	0	-0.1
CJ_10001299	0.4	0.2	0.4	0.1	-0.1	0.1	0.2	0	0	0
CJ_10001300	0.4	0.4	0.4	0.1	0.1	0	0.1	0	0.1	-0.1
CJ_10001301	0.3	0.2	0.1	0.1	0	0	-0.1	0	0.1	0
CJ_10001302	1.6	1.3	1	0.1	-0.6	0.3	0.5	-0.3	0	0.4
CJ_10001303	0	0	-0.3	0	-0.1	-0.1	0	-0.1	0.1	0.1
CJ_10001304	-0.7	-0.1	-0.8	0	-0.3	0.1	-0.9	0	-1.2	0
CJ_10001305	0.3	0.5	-0.4	-0.1	-0.2	0.2	-0.1	-0.1	0.1	-0.1
CJ_10001308	-0.1	-0.2	0.1	0.1	0.1	0.1	0	0	0.1	0

CJ_10001309	0.1	0.2	-5.2	0.2	0.2	0.1	0	-1.6	0	0.1
CJ_10001310	-0.2	0.1	0.1	0.1	0.2	0.1	0	-0.1	0.1	0
CJ_10001311	0.3	0.3	0.5	0.2	0.1	0.2	0.1	0	0.1	0.1
CJ_10001312	0	-0.4	0	0	0.2	0	-0.1	0	-0.1	-0.1
CJ_10001314	0.8	1.5	0.5	0.5	-0.5	0.4	0.3	-0.8	0.4	0.6
CJ_10001315	0	0.3	0.4	0.2	-1	0.2	-0.1	-0.3	0.1	0.1
CJ_10001316	0.2	-0.1	0.9	0	0.2	0.1	-0.3	-0.2	-0.1	0
CJ_10001317	0	-0.1	0	0.1	0.1	0.1	0	-0.1	0.1	0.1
CJ_10001318	0.1	0.1	0.2	0.1	-0.4	0	-0.1	0	0	0
CJ_10001319	0.3	0.1	0.3	0.1	0	0.1	0.1	-0.2	0	0
CJ_10001320	-0.8	-0.1	-0.7	-0.1	-0.4	0.2	-0.1	0	0.3	0.1
CJ_10001322	-0.7	-0.4	0.1	0.1	0	0	0	0.1	0	0.1
CJ_10001323	-0.2	0	-0.2	0	-0.1	0.1	-0.1	-0.2	-0.1	0
CJ_10001324	0.6	0.5	0.3	0.2	-0.1	0.2	0.2	-0.1	0.1	0.1
CJ_10001325	-0.1	0.1	-1.8	0.1	-0.3	0.2	0	0	0.1	0.1
CJ_10001327	0.5	0	0.2	0.1	0	0.1	0.2	-0.1	-0.1	0.1
CJ_10001328	0	-0.1	0.2	0.1	0	0.2	-0.1	0	-0.2	-0.1

CJ_10001329	0.5	0.1	0.4	0	0	0	0	0	0	-0.1
CJ_10001330	-0.2	-0.1	-0.1	0	0	0	0.1	-0.2	-0.1	-0.1
CJ_10001332	-0.4	-0.1	-6.7	0	-0.1	0	0	-2.1	-0.1	0.1
CJ_10001333	1.2	1	1.1	0.2	-0.2	0.2	0.3	-0.1	0.1	0.1
CJ_10001334	0.2	0.2	0.3	0.1	0.1	0.1	0.1	-0.1	0.1	0
CJ_10001335	0.3	0.1	0.5	0.1	-0.1	0.1	0.1	0	0	0.1
CJ_10001336	6	7.2	-0.1	1.3	-2.6	-0.4	1.9	1.1	7.1	0.4
CJ_10001337	0	-0.1	0.1	0	0	0.1	0.1	0	0	0
CJ_10001338	0.1	0	0.4	0	0	0	0.1	0	-0.1	-0.1
CJ_10001340	0.4	0.1	0.4	0.2	0.2	-0.1	0.1	0	-0.2	0
CJ_10001341	-0.1	0.2	0.1	0.1	0	0.2	0	0	0.1	0
CJ_10001347	-0.2	0	-7.1	0.1	0	0.2	0	-2.3	0	0
CJ_10001350	0.8	0.5	0.5	0.1	0.1	0	0.2	0	0	0
CJ_10001352	0.2	-0.3	0	0	0.1	0.1	0.2	0.2	-0.2	0
CJ_10001353	0	0	-0.1	0.1	-0.1	0.2	0.1	0	0.1	0
CJ_10001354	0.6	0.4	0.6	0.2	0	0.1	0.2	0.1	0	0
CJ_10001355	0	0	0.2	0.2	0.1	0.2	0	0.1	0	0.1

CJ_10001356	-0.5	-0.3	-0.2	0.1	0	0.2	-0.1	-0.1	0.1	0
CJ_10001357	0.3	0.3	0.2	0	-0.1	0	-0.1	-0.1	0.1	0
CJ_10001358	-0.3	0	-0.2	0	-0.1	0.2	-0.1	-0.1	0	0
CJ_10001359	0.3	0.1	0.2	0.2	0.2	0	0.1	0	-0.2	0
CJ_10001360	0	0.1	0.3	-0.1	-0.2	0.2	0	0	0.3	0
CJ_10001361	0.1	0.1	0	0	-0.1	0.2	0.1	-0.1	0.1	0
CJ_10001362	0.3	0.3	0.5	0.1	0	0.2	0.1	0	0	0
CJ_10001363	0.2	0.1	0.4	0	-0.1	0.2	0	-0.1	0.1	0
CJ_10001365	0	0.1	0.1	0.1	0	0.1	0	0	0	-0.1
CJ_10001367	-0.1	-0.1	0	0	0	0.2	-0.1	0	0	0.1
CJ_10001370	0.1	-0.5	0.1	-0.1	-0.2	0.1	0.1	0	0	-0.1
CJ_10001371	-1.1	1.2	-0.4	0.6	-0.5	0.7	-0.2	-1	0.5	0.6
CJ_10001372	0.2	-0.1	0.2	0.1	0.1	-0.1	-0.1	0.4	-0.2	-0.2
CJ_10001374	-0.1	0.1	-3.4	0.1	-0.2	0.1	0	-1.3	0	0.1
CJ_10001375	0.4	0.2	0.3	0	-0.1	0.1	0.1	0.1	-0.1	-0.1
CJ_10001376	0.3	0.3	0.4	0	0	0.2	0	0.1	-0.1	-0.1
CJ_10001377	-0.2	0.1	-0.4	0.2	-0.2	0.1	0	0	0.2	0.2

CJ_10001378	0.2	0	0.1	0.1	0.1	-0.1	0.1	0.1	0	-0.1
CJ_10001379	-0.1	0.1	0.2	0.1	0	0.1	0.1	-0.5	0.1	0
CJ_10001380	0	0.2	0.1	0.1	0.1	-1.4	0	-0.1	0.1	0
CJ_10001381	-0.2	0	-0.2	0	0	0	-0.1	0	0.1	-0.1
CJ_10001382	0.2	0.3	0.3	0.2	0	-1.4	0	-0.1	0	0
CJ_10001383	-0.4	-0.2	-0.2	0.2	0	0.2	-0.1	-0.2	0.2	0.2
CJ_10001384	0.5	0.3	0.6	0	0	0.1	0	0	0	0
CJ_10001385	0.4	0.3	0.1	0	0	0	0	0	0	0
CJ_10001386	0.6	0.5	0.6	0.2	0	0.3	0.2	-0.1	0	0
CJ_10001389	0	0	-0.2	0.2	0.2	0	-0.2	-0.1	-0.1	-0.1
CJ_10001390	-0.3	0.2	0.3	0.1	0.1	0.3	0.1	0.9	-0.1	-0.1
CJ_10001391	-1.9	0.5	-0.7	0.8	0.2	0.8	-0.1	0.5	0.6	0.7
CJ_10001393	0.3	-0.1	-0.3	0.4	0.1	0	0.2	0.3	0	0
CJ_10001395	-0.1	-0.1	0.1	0	0.1	0.2	0	0	-0.2	0
CJ_10001396	0.4	0.4	0.4	0	0	-0.2	0.2	-0.1	0	-0.2
CJ_10001398	0.3	0.4	-0.4	0.2	-0.1	-0.1	-0.1	-0.2	0.1	-0.1
CJ_10001399	0.3	0.1	0.5	0.1	0.1	-0.6	-0.1	0.1	-0.3	-0.1

CJ_10001401	-0.4	0.2	0.3	0.1	0.2	0	-0.1	0.2	-0.1	-0.2
CJ_10001402	-0.2	0.2	0.2	0.1	0	0.1	0.1	-0.2	0	-0.1
CJ_10001403	0.1	0.1	0.2	0.2	-0.1	0.1	0.1	0.3	0	0.2
CJ_10001404	-0.4	-0.4	-0.5	0	0.1	-0.1	0	-0.2	0	0
CJ_10001405	0	-0.4	0.3	0	0	0.1	0.1	0	-0.2	0
CJ_10001406	0.1	0	0.2	0.1	0.1	0	0.1	0	-0.1	0
CJ_10001407	0	0	0	0.1	0.1	0.1	0	0	-0.1	0
CJ_10001409	0.2	0.3	0.5	0.1	-0.1	0.1	0.2	-0.1	-0.1	0.1
CJ_10001411	0.1	0.1	-0.1	0	-0.1	0	0	0	-0.1	0
CJ_10001412	-1.1	0.2	0.1	0.1	-0.1	0.2	0.1	-0.1	0.1	0.1
CJ_10001413	0.4	0.2	0.3	0.1	0.1	-0.2	0	-0.1	-0.1	0
CJ_10001415	-0.1	-0.2	0	0	-0.1	0.1	-0.1	0.1	-0.1	0
CJ_10001416	-0.3	-0.1	0.2	0.2	0	0.1	-0.1	0.1	-0.1	-0.1
CJ_10001417	-0.4	-0.1	-0.1	0.1	0.1	0.1	-0.2	0.2	-0.2	0
CJ_10001418	0.4	0.2	0.5	0.2	0.1	0.1	0.1	0.2	-0.1	-0.1
CJ_10001420	-0.3	0	0	0	0.1	0.1	0.2	0.2	-0.2	0
CJ_10001421	0.3	0	0.2	0.1	0.1	0	0.2	0.2	-0.2	-0.1

CJ_10001423	1.2	0.7	0.3	0.1	-0.1	0.1	0.1	0.1	-0.1	-0.1
CJ_10001424	3.3	2.8	2.4	0.9	-1.3	0.5	1.5	-0.4	0.8	0.8
CJ_10001425	0.9	0.6	0.3	0.1	-0.2	-0.2	-0.1	-0.2	0.1	0.1
CJ_10001426	0	0.1	0.2	0.1	-0.1	0.2	0.1	0	-0.1	0.1
CJ_10001427	0.5	0.3	0.2	0.1	0	0.1	0	0	0.1	0.1
CJ_10001428	-1	-0.7	-0.2	-0.1	-0.1	0.1	-0.2	0	-0.3	-0.2
CJ_10001429	-0.2	0.1	-0.2	0.1	0	0.1	0	0	0.1	-0.1
CJ_10001430	-0.1	-0.2	-0.1	0	0	0	-0.1	-0.1	-0.1	0
CJ_10001431	0.5	-0.1	-0.1	0	0.1	0.1	-0.3	0	0	-0.2
CJ_10001432	-0.1	-0.1	0.1	0	-0.1	0.1	-0.1	0.1	0	0.1
CJ_10001433	-0.2	-0.1	-0.3	0.1	0	0.3	-0.1	-0.1	0.2	0.1
CJ_10001434	0.1	0.3	0.3	0.1	0	0.1	0	0.1	0.2	0
CJ_10001435	-0.2	0	-0.3	0.3	0.1	0.3	-0.1	-0.1	0	0.1
CJ_10001436	0.1	0.3	0.3	0.1	0	0.2	0.2	-0.1	0	0.1
CJ_10001438	0.2	0	0.1	-0.1	-0.1	0.1	0.1	0.2	-0.1	-0.1
CJ_10001439	0.2	0	0.4	0.1	0.1	0.1	0.2	0.1	-0.1	-0.1
CJ_10001440	0	-0.1	0.5	0.2	0	0	0.1	0.1	0	-0.1

CJ_10001441	0.6	0.3	0.5	0.3	0	0	0.3	0.1	-0.2	0
CJ_10001442	0.3	0.1	0.2	0.2	0.2	-0.1	0	0.1	-0.1	-0.1
CJ_10001443	-0.1	-0.4	0.6	0.2	0.1	0	0.1	0	-0.2	-0.1
CJ_10001444	0.5	0.3	0.3	0.1	0	0	0.1	0	0	-0.1
CJ_10001445	0.6	0.5	0.5	0.3	0.1	0.1	0.1	0.1	-0.1	-0.1
CJ_10001446	0.5	0.1	0.5	0.2	0	0	0.2	0.1	-0.3	0
CJ_10001448	0.7	0.3	0.6	0.3	-0.1	0.4	0.2	-0.3	0.1	0.2
CJ_10001449	-0.6	-0.1	0	0	-0.3	0.1	0.1	0.1	-0.1	0
CJ_10001450	-0.4	0.1	-0.2	0.1	-0.3	0.3	0	0	0.1	0.1
CJ_10001451	-0.4	0	-0.1	0.2	0	0.3	0.2	0	0.3	0.2
CJ_10001452	0.6	0.4	0.7	0.1	0.1	-0.1	0.1	0	-0.2	-0.1
CJ_10001453	0	0.1	0.3	0.1	0.1	0.2	0.1	0	-0.1	0.1
CJ_10001454	0	0	0.1	0.1	0.1	0.1	0.1	0	0.1	0.1
CJ_10001455	0.4	0.1	0.7	-0.1	0	0.1	0	0	0.1	0
CJ_10001456	-1.2	0	-0.1	-0.1	-0.1	0.1	0	0	0.2	0
CJ_10001457	-1.1	0.1	-0.9	0.1	-0.1	0.3	-0.3	0	0.2	0.1
CJ_10001458	-0.3	0	0.1	0	-0.2	0.2	0	0	0.3	0.1

CJ_10001459	-1.2	0.1	-0.2	0.1	0.2	0	-0.5	0.4	0	0.1
CJ_10001460	-0.4	0.3	0	0	0	0.1	-0.1	0	-0.1	0
CJ_10001461	-0.1	0	0.4	0.1	0	0	-0.1	0.1	-0.1	-0.1
CJ_10001462	-1.2	-0.1	-1.8	-0.3	-0.1	0.4	-0.4	0.6	-0.7	0.5
CJ_10001463	0	0	0.1	0	0	0.2	0	0.1	-0.1	0
CJ_10001464	0.3	-0.6	-1.3	0.1	0.1	0.1	-0.1	0.1	-0.3	-0.1
CJ_10001465	-0.9	-0.3	-1	0	-0.1	0.1	-0.3	0.1	-0.1	-0.1
CJ_10001466	0	-0.1	-0.1	0	0	0.1	-0.1	0	0	0
CJ_10001467	-1.5	-0.5	0.1	0	0.1	0.2	0.1	0.2	0.1	0
CJ_10001469	0.2	-0.1	0	0.1	0	0.4	0.2	0	-0.1	0
CJ_10001471	-1	0	-1.2	0.2	0	0	-0.6	0	0.2	-0.2
CJ_10001472	-0.3	-0.1	-0.3	0	0	0	0	0	0	0
CJ_10001473	-0.2	0.1	0.2	0	-0.1	0	0	0	0.3	0
CJ_10001474	-0.1	0.2	-0.2	0.1	-0.1	0.1	0	0	0.1	0.1
CJ_10001475	1.9	0	0.6	0	-0.7	0.3	-0.2	-0.3	0.6	0.1
CJ_10001476	-0.6	-0.6	-1.2	0	0	0	-0.3	-0.2	0.2	0
CJ_10001477	-0.2	0.9	0.9	0.4	-0.3	0.5	0.4	-0.2	0.3	0.2

CJ_10001479	0.1	0.2	-0.1	0	-0.1	-0.1	-0.3	-0.2	0.3	-0.1
CJ_10001480	0	0	0.6	0.1	0.1	0.1	0	-0.1	0	0
CJ_10001481	3.8	3.6	1.5	0.1	-1.2	1.1	1.1	-0.8	0.9	0.2
CJ_10001482	-0.1	0	0.1	0	-0.1	0	-0.1	-0.2	0.3	0
CJ_10001483	1	0.7	0.3	0.3	-0.6	0.5	0.3	0.2	0.3	0.3
CJ_10001484	0	0.2	0.1	0.2	0.1	0.1	-0.1	-0.1	0	0
CJ_10001486	-0.6	-0.1	-0.5	0.2	0	0.3	0.1	0	0	0
CJ_10001487	-0.2	-0.1	-0.2	0.1	0.1	0.1	-0.3	0	-0.1	-0.1
CJ_10001488	-0.4	-0.6	0.1	0	0	0.3	0.1	0.1	-0.3	0.1
CJ_10001489	-0.2	-0.3	-0.3	0.1	0.1	0	0	0	-0.1	0
CJ_10001490	-0.3	-1.5	-0.7	0.2	0.5	0.2	-0.6	0.3	0	0.1
CJ_10001491	-0.1	0.1	0.3	0	-0.1	0.1	-0.1	0.2	-0.1	-0.1
CJ_10001492	0	0.1	0	0.1	0	0.2	-0.1	0	0	0
CJ_10001493	0.2	0.3	0	0.2	-0.1	0.2	0	-0.1	0	0.1
CJ_10001494	0.6	0.3	0.5	0.3	0.2	-0.1	0.2	0.2	-0.1	-0.1
CJ_10001495	0.4	0.2	0	0.1	-0.1	0	0	-0.2	0.1	0
CJ_10001496	0.2	0.2	-0.2	0.1	-0.1	0.1	0	0	0	0.1

CJ_10001497	0.4	0.1	0.2	0.1	-0.2	0	0	0.1	0.1	0
CJ_10001498	-0.4	-0.1	-0.5	0.1	0	0.3	0	-0.1	0.1	0.1
CJ_10001499	0	0.1	0.1	0.1	-0.2	0.3	0.1	-0.2	0	0
CJ_10001500	-2.1	0	-0.2	0	0	0	0	-0.3	-0.2	0.1
CJ_10001501	0.5	0.9	0.3	0.2	-0.4	0.3	0.3	-0.2	0.3	0.3
CJ_10001502	0.7	0	-0.1	0.2	0.1	-0.3	0.3	0.1	0.1	0
CJ_10001505	-0.6	-0.2	-0.4	0	0	0.3	0	0	0.2	0
CJ_10001506	-0.1	0	-0.4	-0.1	-0.2	-0.1	0.2	0	0.1	0.1
CJ_10001507	0	0.1	-0.6	0.1	-0.1	0.1	-0.2	-0.2	0	0.1
CJ_10001508	2.8	-0.4	0.8	0.1	0.2	0.1	1.1	0	0.7	0
CJ_10001509	-0.8	-0.2	-0.1	0.2	0.1	0.1	-0.1	0	-0.1	-0.2
CJ_10001510	0.9	0.4	-0.1	0.2	-0.2	0.1	0.2	-0.3	0.1	-1.3
CJ_10001512	0.3	0.2	0.1	0.1	0.1	0	0.1	-0.3	-0.1	-0.1
CJ_10001513	-0.3	0	0	0.1	0.1	0.1	0	0	-0.1	0
CJ_10001514	0.3	0	0.1	0.2	0	0.1	0.1	0	-0.1	0
CJ_10001515	-0.8	-0.3	-0.5	0.1	0	0.1	-0.2	0.1	0.1	-0.1
CJ_10001516	0.4	0.1	0.2	0.2	0.1	0.1	0.1	0	0.1	0

CJ_10001517	-0.3	-0.6	-0.3	0.1	0.2	0.1	-0.2	0.1	-0.2	0
CJ_10001518	0.1	-0.1	0.1	0.1	0	0.1	-0.1	0	-0.1	-0.1
CJ_10001519	-0.5	-0.2	0.7	0.1	0.1	0	0.1	0.4	-0.1	-0.2
CJ_10001520	0.5	0.5	-0.1	0.2	-0.2	-0.9	-0.1	0	0.2	0.2
CJ_10001521	-3.8	0	0.5	0.1	0.1	0.3	0	0	0.1	0.1
CJ_10001522	-1.4	-0.2	-0.2	0.1	0.1	0.2	-0.2	-0.6	-0.1	0
CJ_10001523	3.3	5.5	-0.5	1.9	-1.4	5.4	-0.1	-1.3	1.1	1.8
CJ_10001524	-0.8	-0.3	-0.6	0	0	0.2	-0.1	-0.2	0.2	0.1
CJ_10001526	-0.6	-0.3	0	-0.1	0	0.1	0.1	0.1	1.7	0.1
CJ_10001527	-0.5	-0.6	-0.1	0	0	0.2	0.1	0	-0.1	0.1
CJ_10001528	-0.7	-0.4	-0.9	0.1	0	0.3	-0.1	0	0.1	0
CJ_10001529	0.2	0.1	0.5	0.1	0.1	-0.1	0	-0.1	-0.3	0
CJ_10001530	-0.2	0	0.2	0	0	0.1	0	-0.1	0.1	0
CJ_10001531	0.3	-0.1	0.1	0.1	0.1	0.1	-0.1	0	0	0
CJ_10001533	0.9	0.4	0.6	0.3	0.2	0	0.1	0	0	0
CJ_10001535	-0.2	0	-1.3	0.1	-0.3	-0.2	0.1	0.1	0.3	0
CJ_10001536	-0.5	0	-0.5	0	-0.6	0.2	0	0	0.2	0.2

CJ_10001537	0.3	0.3	0	0.1	-0.1	0.1	0.2	0	0.2	0.1
CJ_10001538	0.2	0	-0.1	0	-0.2	0.2	0.1	0.1	-0.1	0.1
CJ_10001539	1.7	1.2	0.6	0.1	-0.4	0.3	0.5	0.2	0.1	-0.1
CJ_10001540	0.3	0	0.1	0.1	0.2	0.1	0	-0.1	-0.1	-0.1
CJ_10001541	-0.1	-0.2	0.2	0.1	0.1	0.1	-1	-0.5	-0.3	-0.1
CJ_10001542	0.5	0	0.3	0.1	0	0.1	0.1	0.1	-0.1	-0.1
CJ_10001543	-0.9	0	0.1	0	-0.1	0	0	-0.1	-0.1	-0.1
CJ_10001544	0	-0.5	0.5	-0.1	0	0	0	0	1.2	-0.1
CJ_10001545	-0.4	0.3	0.1	0.1	0.1	0.3	0.1	-0.1	-0.3	0.1
CJ_10001546	0.1	0	0.2	0.1	0.1	0	0	0	0	0
CJ_10001547	1.4	1.1	0.6	0.2	0	0.1	0.3	-0.3	2.4	0
CJ_10001548	0.4	0.2	0.3	0	0	0	-0.1	-0.3	0	0
CJ_10001550	0.2	0	0.5	-0.1	0.1	0	-0.1	0.1	1.3	-0.2
CJ_10001552	0	0.1	0	0.1	0.2	-0.1	-0.1	0	0	-0.2
CJ_10001553	-0.4	-0.2	-0.1	0.1	0.1	-0.1	-0.1	0.1	0.6	-0.1
CJ_10001555	-3.8	0.6	0.4	0.3	-0.2	-1.2	-1.2	-1.6	0.2	0.2
CJ_10001556	0	0.1	-0.1	0.1	-0.1	0.1	-0.1	-0.2	-0.1	0

CJ_10001558	-2.3	0	-0.4	0.1	-0.1	-0.1	-0.3	-0.3	0.1	-0.1
CJ_10001560	-6.6	0.3	0.3	0.1	-0.4	0.2	-0.1	-0.2	-0.1	0.1
CJ_10001562	-0.3	0.5	0	-0.1	-0.5	0.3	0.1	0	-0.2	0
CJ_10001563	0.8	0.4	0.7	0.2	0.1	0	0.2	0.1	-0.1	-0.1
CJ_10001564	0.4	0.3	0.2	0.2	-0.1	0.1	0.2	-0.1	-0.1	-0.1
CJ_10001565	-0.1	-0.2	0.1	0	-0.1	0.1	0.1	0.1	0	0
CJ_10001566	0.3	0	0	0	0.1	-0.1	0	0.1	-0.1	-0.2
CJ_10001567	-0.4	-0.8	-0.7	0	0	0.1	-0.1	0	0.1	0
CJ_10001568	0	-0.2	0.3	0	0.1	0.1	-0.1	-0.1	1.4	0
CJ_10001569	-0.6	-0.1	-0.2	-0.1	-0.1	0.2	0	-0.1	-0.1	0
CJ_10001570	-0.1	0	0	0.2	0.1	0	0.1	-0.1	0	0
CJ_10001571	-0.3	0	-0.4	0	0	0.2	-0.1	0	1.9	0.1
CJ_10001572	-0.1	-0.3	-0.4	0	0	0.3	0.1	0	-0.1	0.2
CJ_10001573	0.2	-0.3	0.3	0.1	0.1	0.1	0.1	-0.1	-0.1	0.1
CJ_10001574	0.4	0.3	0.6	0.1	0	0	0	-0.2	0.9	0
CJ_10001575	0.3	0.3	0.5	0.1	0.1	0.1	0.1	-0.2	0	0
CJ_10001576	0.4	0.2	0.5	0.1	0.1	0.1	0	0	0	0

CJ_10001577	-0.3	-0.3	0	-0.1	0	0	-0.1	-0.2	1.3	0
CJ_10001578	-0.7	-0.3	-0.6	0.2	-0.2	-1	-0.1	-0.6	0.1	0
CJ_10001579	0.2	-0.2	-0.1	0.1	0.2	0.2	-0.1	-0.2	-0.2	0.1
CJ_10001580	-0.2	-0.3	-0.2	0	-0.3	0.3	0.2	0	0.1	0.1
CJ_10001581	0	0.1	0	0	-0.2	0	0	-0.2	-0.2	0.1
CJ_10001582	-0.1	0.1	0.1	0.1	0.1	0.1	0	0	0	-0.1
CJ_10001583	0.3	0.4	0.1	0.2	-0.3	0.1	-1.8	0	0.2	0
CJ_10001584	-0.7	-0.3	-0.1	0	-0.1	0	-0.3	0.1	-0.1	-0.2
CJ_10001585	-0.2	0	-0.3	-0.1	0	0.2	-0.1	-0.1	0.2	0
CJ_10001586	-0.5	0	-0.5	0.1	0.1	0.1	-0.1	0.1	0.2	-0.2
CJ_10001587	0.3	0.5	-0.2	0.1	0	-0.1	-0.4	-0.6	0.2	-0.1
CJ_10001588	0.3	0.4	0.3	0.1	0	-0.1	-0.4	-0.1	0	-0.2
CJ_10001590	0	0.2	0.4	0.2	0.1	-0.1	-0.1	-0.1	0	-0.1
CJ_10001591	-0.2	0.2	0.3	0.1	0.2	0	-0.1	-0.1	-0.2	-0.2
CJ_10001593	-0.3	-0.1	-0.5	0.1	0	0.2	-0.1	-0.3	0	0.1
CJ_10001596	0.7	0.1	0.4	0	-0.1	0.3	0.1	-0.1	0.1	0.1
CJ_10001597	0	0.1	-0.1	-0.1	-0.1	0.2	0	-0.1	0	0

CJ_10001598	-0.1	0	0	-0.1	0	0.1	0	-0.3	0.2	0
CJ_10001599	0	-0.1	-0.1	0	0	0.1	0	0	0.2	0
CJ_10001600	-0.2	0	-0.1	0.1	0	0	0	0	0	-0.1
CJ_10001601	0.5	0.4	0.5	0	-0.1	0.2	0.1	-0.1	0.8	0
opCcV0100000001	-0.1	-0.2	0.5	-0.2	0.2	0.1	0	0.1	-0.2	-0.1
opCcV0100000002	0	0.1	0.1	-0.1	0	0	-0.2	-0.1	0.2	-0.2
opCcV0100000004	-0.4	-0.4	0	-0.1	0.1	0.1	-0.1	-0.2	-0.2	0
opCcV0100000008	-0.1	-0.2	-0.1	0.1	0	0	0	-0.1	0.1	-0.2
opCcV0100000009	0.6	0.6	0.4	0.1	0.2	-0.1	-0.2	0	-0.6	-0.1
opCcV0100000014	0.5	0.2	0.2	-0.1	0	0	0.2	0.1	0	-0.2
opCcV0100000017	0.7	-0.7	0.7	-0.1	0	0.3	0.4	0	-0.1	0
opCcV0100000019	-0.1	0.1	0	0	-0.1	0.2	0	-0.1	0.1	0.1
opCcV0100000023	-2.2	-1.8	0	-0.5	0.8	0	-0.8	1.2	-0.6	-0.7
opCcV0100000025	0.1	0	0.3	-0.2	0.1	0.2	0	0	-0.1	0
opCcV0100000033	-1.1	-1.5	-0.3	-0.1	0	-0.3	0.3	0.4	-0.4	-0.1
opCcV0100000035	-0.6	-0.2	-0.9	-0.1	-0.1	-0.1	0	0.1	0	0
opCcV0100000036	0.6	0.4	0.2	-0.1	-0.2	0.2	-0.2	0	0.1	0

opCcV0100000037	0.1	-0.1	-0.1	0.1	0.1	0	0	0	-0.1	-0.1
opCcV0100000038	-0.6	-0.4	-0.5	-0.1	-0.3	0.2	-0.2	-0.9	0.2	0.1
opCcV0100000039	-0.9	-0.2	0.3	0	0	0.1	0.2	-0.1	0.1	0
opCcV0100000040	-0.5	-0.7	-2.1	-0.3	0.2	-1.4	-1.5	-0.3	-0.1	-0.1
opCcV0100000041	-0.9	1.3	-1.9	-0.1	0.1	-1.9	-1.3	-0.1	0	-0.2
opCcV0100000046	-1.4	-0.9	1	-0.1	0.6	0	0.2	0.5	-0.5	-0.4
opCcV0100000047	-1	-0.2	-1	-0.2	0	0.1	-0.2	-0.1	0.3	0
opCcV0100000049	-1.6	-1.9	-0.8	0	0.4	-0.3	-0.2	0.9	-1	-0.2
opCcV0100000050	-1.9	-1.4	0	-0.2	-0.1	0.1	0.1	0	-0.1	-0.4
opCcV0100000053	0.2	-0.2	0.7	0	0	0.2	0.1	-0.2	0.6	0
opCcV0100000055	0.1	0.2	0	0.1	0.1	0	0	0	0	-0.1
opCcV0100000057	1.1	-0.6	0.6	0	0	0.2	0.5	0.1	0.1	0
opCcV0100000060	1.6	0.4	-0.1	0.1	-0.1	0.3	0.1	-0.1	0.1	-0.1
opCcV0100000068	0	0.2	0.2	-0.1	-0.1	0	-0.2	0	-0.1	0
opCcV0100000069	2.5	1.4	1.8	0.6	-0.7	0.5	0.5	-0.6	0.7	0.6
opCcV0100000072	-0.9	-1.9	0.1	-0.3	-0.1	0.3	0.2	0.4	-0.3	0.1
opCcV0100000075	0.2	0.3	0.4	0	0	0	0	-0.2	0	-0.1

opCcV0100000078	0.4	0.3	0	0	0	0.3	0	0	0.2	0
opCcV0100000079	0.3	0	0.2	0	0	0	-0.2	0.1	0.2	0.1
opCcV0100000085	-0.3	0	-0.2	0	0.1	0	-0.1	-0.1	0	-0.2
opCcV0100000087	0.2	0.2	0.1	-0.1	-0.1	0.1	0	0.1	0	-0.1
opCcV0100000091	0.1	0	-0.1	0.1	-0.1	0.1	0	-0.1	0	0
opCcV0100000095	0	-0.1	-0.3	0.1	0.3	-1.7	-2.4	0.1	0	0
opCcV0100000098	1.8	1.3	1.4	0.5	-0.4	0.6	0.4	-0.6	0.2	0.6
opCcV0100000101	0.4	0.1	0.3	0.1	0	0.1	0	-0.1	0.1	0.1
opCcV0100000104	0.6	0.3	0.6	0.1	0.1	0	-0.1	-0.1	0	0
opCcV0100000105	0.9	0.7	0.4	0.1	-0.2	0.2	-0.3	-0.5	0.3	0.2
opCcV0100000109	-0.9	0.1	0.4	-0.1	-0.2	-0.4	-0.2	0.2	-0.2	0
opCcV0100000111	-0.2	-0.3	0.7	0.1	0.2	0.3	0.5	0	-0.2	0
opCcV0100000118	0.2	0.1	0.3	0	0.1	-0.1	-0.3	-0.1	-0.1	0
opCcV0100000123	-1.1	-0.9	-0.6	0	0.2	-0.6	-0.2	-0.2	0	-0.1
opCcV0100000126	0.2	-0.1	0.1	0.1	0.1	0.1	0.1	-0.1	0.1	0
opCcV0100000129	-0.6	-0.2	-0.6	0	-0.1	0.2	-0.5	-0.1	-0.1	0
opCcV0100000132	0.7	0.4	-1.8	0.1	0.1	-1.6	-1.9	-1.7	-0.1	0

opCcV0100000135	-0.3	1	1.3	0.1	-0.3	0.5	0.4	-0.4	-0.3	-0.1
opCcV0100000137	0	0	-4.6	-0.2	-0.2	-1.4	-3	-1.4	0	0.1
opCcV0100000143	1.2	0.9	0.2	0.3	0	0	0.7	-0.1	0.3	0.2
opCcV0100000148	0.3	-0.2	0.1	0	0	-0.1	-0.1	0	0	-0.1
opCcV0100000149	0.4	0.6	1	0.3	0	0.5	0.2	0.1	-0.1	0.4
opCcV0100000151	0.4	0.5	0.3	0	0	0	-0.1	0	0.1	-0.1
opCcV0100000153	0.1	0.1	0.6	0.2	0	0.3	0	-0.1	0	0
opCcV0100000155	0.8	1.1	-0.5	0.4	-0.4	-0.2	0.3	-0.6	0.1	0.4
opCcV0100000158	0	0.2	0.2	0	0	0.2	0	0	-0.1	-0.1
opCcV0100000161	0.7	1.1	0.7	0.1	-0.2	-0.1	0.4	-0.1	0.1	-0.1
opCcV0100000162	0.2	0.1	-4.6	0	0.1	-0.1	0.2	-1.6	0	0
opCcV0100000164	-0.2	-0.1	-0.1	-0.2	-0.1	0.3	-0.1	0	0	-0.1
opCcV0100000167	-1.2	-0.5	-1	-0.1	0	0.2	-0.2	0	0.1	0
opCcV0100000169	0.3	0	0.1	0.1	0.1	0	0.1	0.1	0	-0.1
opCcV0100000171	-0.1	-0.2	0.1	0	0.1	-0.7	0	0	0.2	0.1
opCcV0100000172	-0.5	-0.2	1.3	-0.4	0	-0.1	-0.2	0.2	-0.3	-0.2
opCcV0100000174	0.2	0	0.6	0.2	0.1	0.3	0.2	-0.1	0.1	-0.1

opCcV0100000182	-0.4	0	-0.6	0.1	0.1	0.1	0	0	0	-0.1
opCcV0100000184	0	-0.3	-0.8	0	0.1	-0.2	0.2	0.2	-0.1	0.1
opCcV0100000186	0.2	-0.6	0.6	-0.1	0.4	0	-0.1	0.2	-0.1	-0.1
opCcV0100000192	0.2	0.1	0.3	0	0.1	0.1	0.1	0.1	0	0
opCcV0100000195	-0.4	-0.1	-0.3	0	0	0.2	0	0	0	0
opCcV0100000198	1.8	0	0.9	0.4	-0.2	0.1	0.4	-0.4	0.3	0.3
opCcV0100000200	1.5	1.2	0.7	-0.5	-0.7	0.9	0.7	0.4	0.8	0.1
opCcV0100000201	-0.6	2.2	2.2	0.8	-0.9	0.6	0.8	-0.5	0.5	0.5
opCcV0100000205	0.1	-0.2	0.1	-0.2	-0.1	0.2	-0.1	0	-0.1	-0.1
opCcV0100000216	0.2	0	-0.4	0.1	0	0.1	0.1	0	-0.1	0
opCcV0100000220	-0.1	-0.7	1.2	0.4	-0.2	0.4	0.6	-0.2	0.3	0.5
opCcV0100000222	-0.4	0	-0.3	-0.1	-0.3	0.2	0	-0.1	0.1	0.1
opCcV0100000225	-2	-1.5	-0.9	-0.8	0.4	0	-0.4	0.9	-0.4	-0.6
opCcV0100000232	0.3	0	0	0	0.1	0	0.1	0.1	-0.1	-0.1
opCcV0100000236	-3.2	-1.4	0.3	0	-0.1	-0.3	0.1	0.9	0.5	0.1
opCcV0100000237	-0.3	0	-0.5	0	0.1	0	-0.1	0	0.1	-0.2
opCcV0100000239	-2.6	-1.3	0.8	-0.2	-0.2	-0.2	0	0.2	0.1	0.1

opCcV0100000240	1.4	0.1	0.8	0	-0.1	0.2	0	0.1	-0.1	0
opCcV0100000241	-0.1	0.3	0.6	0	0	0.4	0.2	0	0	0
opCcV0100000242	-1.2	-1.1	0	-0.2	0.1	-0.2	0.3	0.4	0.1	0
opCcV0100000243.80	3.7	1.7	0.4	-0.7	-1.7	-0.2	0.6	1.9	1.8	-0.9
opCcV0100000243.90	6	1.2	-0.2	0.2	-2.7	-0.2	-1.9	1.1	2.8	-1.9
opCcV0100000243.C	1.5	1.2	-0.2	-1.8	-0.5	0.8	-0.7	-1	1.9	-0.6
opCcV0100000244	0.8	0	0.2	0	-0.1	0	0.3	-0.3	0	0.1
opCcV0100000245	-0.1	0.1	-6.1	0	0	-2.4	-2.1	0.1	0.1	0
opCcV0100000250	0.1	-0.9	-0.5	-0.1	0.2	-0.5	0.1	-0.6	-0.1	-0.1
opCcV0100000256	0.5	0	0.1	0.1	0.2	0	0.1	0.1	-0.1	0
opCcV0100000260	1.3	-0.4	0.6	-0.2	-0.1	0.3	-0.1	0.5	-0.1	0
opCcV0100000264	0.1	-0.3	-0.1	-0.1	0	0	-0.1	0	0.1	0
opCcV0100000268	0.7	0	0.5	0	0.1	0.1	0.1	0	-0.1	0
opCcV0100000274	0.6	0.3	0.1	0	0	0.1	0.2	0	-0.2	-0.2
opCcV0100000278	-0.4	-1	-0.1	-0.1	0.1	-0.3	-0.2	0.1	-0.2	-0.1
opCcV0100000279	-0.5	-1.7	-0.1	-0.6	0.5	0.1	-0.1	1.3	-0.4	-0.4
opCcV0100000280	4	0.5	1.1	-0.3	-1.9	-0.1	0.6	-1.9	1	-0.4

opCcV0100000281	0.7	-0.1	0.2	0	-0.1	0.2	-0.6	-0.1	0.3	0
opCcV0100000282	1.5	0.1	0.5	0.2	0.1	0.1	0.3	0.1	-0.2	0
opCcV0100000286	0	-0.3	0.5	0.1	0	0.1	-0.1	0	-0.1	0
opCcV0100000291	-0.1	0.2	-0.3	0	-0.2	0	0	-0.4	0.4	0
opCcV0100000293	0.3	0.3	0.1	0	0	-2.4	-2.3	0.1	0	-0.1
opCcV0100000299	0.6	0.2	0.9	0	0.1	0	0.2	0	-0.1	-0.2
opCcV0100000300	0.8	0.4	0.7	0	0	-0.1	0.1	0.1	0	-0.2
opCcV0100000304	0.6	0.4	-2	0	-0.1	-2	-1.7	-1.6	0	-0.1
opCcV0100000307	-1.9	-0.6	-0.2	0	0.1	-0.7	-1	-0.3	0	0
opCcV0100000311	0.6	0.2	0.5	0.2	0	-0.1	0.1	0	0	-0.1
opCcV0100000312	5.1	4.5	-1	0.9	-2.1	-1.2	-0.1	-1.1	2.4	1.6
opCcV0100000315	-1.1	-1.2	-0.2	0.1	0.1	0.6	-0.1	0.5	-0.7	0
opCcV0100000317	0.2	0.1	0	0.1	-0.1	0	0.2	0.1	-0.1	-0.1
opCcV0100000320	0.5	0.1	0.4	-0.1	0	0.1	0	0	-0.2	-0.1
opCcV0100000321	-0.1	-0.2	-0.1	0	0	-0.1	-0.1	-0.1	-0.2	-0.1
opCcV0100000324	0.3	-0.1	0.2	0	-0.1	0.2	-0.1	0.1	-0.2	0
opCcV0100000326	-0.5	-0.3	-0.1	0	0	0	0.1	0.1	-0.1	-0.2

opCcV0100000327	-0.1	-0.2	0.2	0	-0.1	0.2	0	0.1	-0.1	-0.1
opCcV0100000330	-0.8	-0.4	0.6	-0.2	-0.1	-0.2	0.2	0.9	-0.5	-0.2
opCcV0100000333	-0.2	0	-0.2	0	-0.2	0	-0.2	-0.4	0	0
opCcV0100000336	0	-0.1	0.1	-0.1	-0.2	0.1	0	-0.1	0	0.1
opCcV0100000337	0.5	0.2	-3.1	0.1	0.1	-1.2	0	-0.1	-0.1	-0.1
opCcV0100000338	0.4	-0.4	-1.4	0.2	0.1	-0.5	-0.9	0.1	-0.1	-0.1
opCcV0100000339	0.8	-0.9	0.8	0.1	-0.3	0.6	0.5	0.2	0.5	0.3
opCcV0100000340	0.7	-0.5	0.3	0.1	-0.4	0.3	-0.3	0.1	-0.8	-0.2
opCcV0100000343	1.7	1.2	0.8	0.3	-0.7	0.5	0.5	-0.2	0.4	0.4
opCcV0100000344	-3.1	-0.5	0.3	-0.1	0	-0.5	0.3	0	-0.5	-0.1
opCcV0100000348	1.5	1.2	-0.2	0.1	-0.6	-0.1	-0.4	-0.6	0.1	0.2
opCcV0100000349	0	0	-0.1	-0.1	-0.1	0.1	0	0.1	0.1	0
opCcV0100000351	-0.4	0	-0.5	-0.1	-0.3	-0.2	-0.8	0.1	0.1	0
opCcV0100000355	-1.8	-1.4	-0.5	-0.2	0.3	-0.1	-0.4	0.4	-0.2	-0.3
opCcV0100000358	-0.7	-0.7	0.5	-0.1	0.2	0.2	0.2	0.2	-0.2	-0.2
opCcV0100000360	-0.1	-1.2	-0.6	0	0.1	-0.2	-0.2	-0.1	-0.2	-0.1
opCcV0100000361	-5.5	0.3	-4.3	-0.7	1.3	-1.3	-2	0.9	-0.6	-1

opCcV0100000364	-1.4	-2.1	-0.5	-0.4	0.4	-0.1	-0.3	0.5	-0.5	-0.4
opCcV0100000367	-1.5	-0.4	1.4	-0.2	-0.3	0.4	0.5	0.2	0	-0.1
opCcV0100000368	-0.9	-0.2	0.4	0.1	0.1	0.3	0.2	0.3	-0.6	-0.1
opCcV0100000370	0.3	-0.2	-0.2	-0.1	0	0.1	0	0	-0.1	0.1
opCcV0100000376	1.8	1.6	1.9	1.3	-1.3	0.3	1.1	0.2	1.4	-0.5
opCcV0100000377	-0.7	0.1	0.6	-0.1	0.1	0.1	0	0.3	-0.9	-0.1
opCcV0100000384	0.2	0.1	0.2	0.1	0.1	-0.1	-0.1	0	0	-0.1
opCcV0100000385	-0.4	-0.9	0	-0.1	0	-0.1	-0.4	0	-0.1	0
opCcV0100000387	-2	-1.1	-0.2	0	0.3	0.1	-0.2	0.7	-0.6	0
opCcV0100000391	-0.4	-0.3	0.3	0	-0.4	0.3	0.1	0.2	0.1	0.1
opCcV0100000396	-0.2	0.3	0.1	-0.3	-0.2	0.3	0.1	0.2	0.1	0.1
opCcV0100000397	0.4	-0.8	0.1	-0.1	0.2	0	0.1	-0.2	0.1	-0.1
opCcV0100000400	0.2	-1	-0.2	0	0	-0.3	-0.2	-0.1	0	-0.1
opCcV0100000408	-3.2	-1.1	-1.4	-0.2	0	-0.9	0	0.2	-0.3	0.1
opCcV0100000411	-0.4	-0.1	-0.1	0	0.1	0.1	-0.1	-0.1	-0.1	0
opCcV0100000412	0.8	-0.2	0.1	0	0.1	-0.1	0.1	0.2	-0.2	-0.1
opCcV0100000414	-0.6	-0.1	-0.5	0.1	-0.2	0.2	0.1	-0.2	0.2	0.1

opCcV0100000416	0.9	-0.6	-0.3	0.3	0.5	0	-0.1	0	0	0.3
opCcV0100000422	0.3	0.2	-2.3	0	0	-1.7	-1.7	-0.8	0.2	-0.1
opCcV0100000425	1	0.7	0.8	0.2	0.1	-0.2	0.2	-0.1	0	0
opCcV0100000427	0.8	0.5	0.7	0.3	0.2	-0.1	0.2	0.1	-0.1	-0.1
opCcV0100000428	0.1	0.6	0.1	0.1	0	0.1	0	0	1	0.2
opCcV0100000429	0	0	-0.2	-0.1	0.1	0.2	-0.1	-0.1	0.1	0.1
opCcV0100000434	0.3	-0.2	0	0.1	0.1	-0.1	-0.2	-0.1	1.1	0
opCcV0100000437	-0.3	-0.6	0.3	0	-0.1	0.2	0.1	-0.3	-0.2	0
opCcV0100000443	-0.1	0.4	0	-0.1	-0.2	0.1	-0.1	0.1	0.2	-0.1
opCcV0100000444	0	0	-1.6	-0.1	-0.2	-0.8	-1.1	-1.1	0	0.1
opCcV0100000446	-0.1	-0.5	0	0	0.1	0.2	0.2	0	-0.1	-0.1
opCcV0100000447	0.4	-0.2	0.4	0	-0.1	-0.1	0.2	0.1	-0.2	0
opCcV0100000448	0.2	-1.6	-0.2	-0.1	0	0.1	-0.2	-0.1	0.2	-0.1
opCcV0100000451	0.9	1	0.8	0.2	0.1	0.2	0.3	-0.1	-0.1	0.2
opCcV0100000452	-1.4	-0.1	-0.4	-0.1	0	0.2	-0.2	-0.2	0	0.1
opCcV0100000455	1.3	1.2	0.8	0	0	0.2	0.2	-0.1	0.3	0
opCcV0100000458	0.1	0.2	0.4	0.1	0.1	0.1	0	0	0	0

opCcV0100000463	0.1	0.1	-0.2	-0.1	-0.1	0	-0.2	-0.1	0.1	0
opCcV0100000467	0.9	0	0.2	0.1	-0.3	0	0.3	0.3	-0.2	-0.1
opCcV0100000469	-1.5	-1.6	-0.5	-0.1	0.3	-0.1	-0.2	0.3	-0.4	-0.3
opCcV0100000470	1.1	-1.6	0.2	0.6	0	0.5	1	0.5	-0.9	-0.4
opCcV0100000472	-3.4	-3.5	-6.1	-1.1	1.3	-1.7	-1.5	1.3	-1.5	-1.4
opCcV0100000476	0.9	-0.3	0.2	0.1	0	-0.3	-0.5	-0.1	-0.1	0
opCcV0100000478	0.8	0.7	0.8	0.1	0.1	0.1	0.1	-0.1	0	0.1
opCcV0100000479	-2.1	-1.3	-0.8	-0.4	0.7	-0.6	-0.3	0	-1.8	0
opCcV0100000481	-0.4	0.2	0.5	0.1	0	0.3	-0.1	0	0.2	0.1
opCcV0100000484	0.2	0.1	0.4	0.3	-0.1	0.4	0.5	-0.3	0.4	0.2
opCcV0100000492	0.3	0.4	0.3	-0.1	-0.2	0	-0.1	0.1	0.1	-0.1
opCcV0100000495	-1.8	-0.7	0.1	0	0.8	-0.1	-0.1	0.6	-1.3	-0.1
opCcV0100000497	0	-0.2	0	0.1	0.1	-0.1	-0.1	0.1	0	0
opCcV0100000498	0.4	0.6	0.2	0	0.2	0	0	-0.1	-0.2	-0.1
opCcV0100000503	3.2	0.5	-0.5	0	1.2	1.1	-1.9	1.2	1.3	0.2
opCcV0100000504	0.2	0	0.5	-0.1	0.1	0.2	-0.1	0	0.1	-0.1
opCcV0100000505	-1.2	0	-0.7	0	0.4	-0.2	-0.4	-0.1	-0.5	0

opCcV0100000506	3	3.1	3.2	0.9	-0.8	1	1.2	-0.9	1.3	1
opCcV0100000510	0.4	-0.3	0.5	0	-0.1	0	0.1	0.3	-0.4	-0.3
opCcV0100000512	-0.2	-0.2	-0.6	0	-0.3	0.2	0.1	-0.1	0	0.1
opCcV0100000513	-1.4	-0.6	-0.2	-0.2	0	-0.1	0.2	0.3	-0.3	0
opCcV0100000517	-0.9	-0.8	0	0	0.1	-0.1	0	0.2	-0.3	-0.1
opCcV0100000520	-0.3	-0.2	-0.4	-0.1	0	0.2	-0.1	-0.1	0.2	0
opCcV0100000521	1.2	0.6	0.6	0.1	-0.2	0.3	0.1	0	0.1	0
opCcV0100000530	-0.1	0.1	-0.1	-0.1	-0.1	0.4	0	-0.1	0.2	0.2
opCcV0100000532	-0.4	-0.2	0.5	-0.2	-0.4	0.5	0.1	-0.3	0.1	0
opCcV0100000534	-0.1	-0.2	-0.1	-0.1	-0.1	0.1	0	-0.2	0	0
opCcV0100000540	0.5	0.1	0.2	0.1	-0.1	0.1	0	-0.1	0	0
opCcV0100000548	-1.3	0.6	0	0	0.2	-0.1	-0.2	0.2	-0.3	-0.1
opCcV0100000549	-1.5	-2.1	-0.9	-0.1	0	-0.3	0.3	1.7	-2.8	-0.3
opCcV0100000551	0.9	0	0.7	0.4	0.7	-0.9	0.2	0.7	0.5	0
opCcV0100000555	-2.7	-2.1	-0.7	0	0.2	0.1	-0.3	0.4	-0.2	-0.1
opCcV0100000557	-0.3	-0.1	0	0	-0.1	0.2	0	0.1	0	-0.1
opCcV0100000559	0	0	0.2	-0.1	-0.3	0.2	0.1	0	0.1	0

opCcV0100000560.60	2.2	0.8	-0.3	0.1	-1.3	-0.1	0.1	1.6	1.4	-0.8
opCcV0100000560.80	2.1	-0.1	-2.1	-0.5	0.4	-0.6	-1	1.7	0	-2.6
opCcV0100000560.90	0.1	1.2	-0.4	-1.2	-1	-0.4	-1.4	0	-0.3	-0.3
opCcV0100000561	-0.4	0.2	-0.3	-0.1	-0.1	0.2	0	-0.1	0	0
opCcV0100000562	-0.2	-0.2	-0.2	-0.3	-0.1	0.1	-0.1	0.1	0	-0.1
opCcV0100000563	0.3	0	-0.1	0.1	0.1	0	0	-0.3	-0.1	-0.1
opCcV0100000564	0.5	0.2	0.3	0.1	0.1	0	0.1	0	-0.1	-0.2
opCcV0100000567	1.1	0.7	0.8	0.3	-0.2	0.4	0.2	-0.2	0.3	0.2
opCcV0100000568	0.2	-0.1	0.2	-0.1	0	0.1	0	0	-0.1	0
opCcV0100000570	-0.8	-0.3	-0.1	0.2	0.3	0	-0.1	0.6	-0.1	-0.2
opCcV0100000573	0.1	0.1	0.1	-0.1	0.2	-0.1	-0.1	0	-0.1	-0.2
opCcV0100000574	0.8	0.9	-0.6	0.2	-0.5	0.1	0	-0.6	-0.2	0.2
opCcV0100000578	-0.1	-0.5	-0.2	-0.1	0	0	-0.1	0.2	-0.1	-0.2
opCcV0100000581	4.4	1.4	1.3	-0.5	-0.6	-0.2	0.4	-0.5	1.1	-0.8
opCcV0100000583	0.4	-0.1	-4.6	0	0.2	-2	-2.1	-1.8	-0.1	-0.1
opCcV0100000584	1.6	2.1	-2.1	0.9	-1.8	-0.8	-1	-0.2	0.9	-2
opCcV0100000585	0	0	-5.4	0	-0.2	-1.7	-1.9	-1.8	-0.1	0

opCcV0100000588	0.2	-0.2	0.5	0.1	-0.1	0.1	0.3	-0.1	-0.1	0.1
opCcV0100000589	-0.4	0	-5.1	0	-0.1	0.2	0.1	-1.7	0.1	0
opCcV0100000590	-0.4	-0.1	-0.1	0	0	0.2	0.1	0	0	0
opCcV0100000593	-1.2	-0.7	0.4	-0.2	0.2	0.5	-0.1	0.6	-0.3	-0.3
opCcV0100000595	-0.2	0.1	0.2	-0.1	0.1	0.1	0	-0.2	0.1	0.1
opCcV0100000596	-2.7	-1.1	1.2	-0.1	0	0.8	0.4	0.2	-0.1	0.1
opCcV0100000602	-0.8	-0.4	-0.2	-0.3	-0.1	0.1	0	0.2	0	0
opCcV0100000606	0.7	0.7	0.5	0.1	-0.5	0.5	0.3	-0.3	-0.2	0.4
opCcV0100000615	0.7	0.3	-0.7	-0.1	-0.2	0.1	0.2	0	0.3	0.2
opCcV0100000623	-4.8	-4.8	0.5	-1.3	1.9	-1.2	-1.5	1.7	-1.6	-1.5
opCcV0100000625	0.2	0.1	-1	0.1	0.1	-0.7	-0.9	-0.6	-0.1	-0.1
opCcV0100000627	-2.4	0.9	-0.9	0.2	-0.3	0.3	0.5	-0.2	0.1	0.2
opCcV0100000628	0.3	0	0.3	0	0	0.1	0	0.2	0	0
opCcV0100000630	0.1	-0.2	0.4	0.1	-0.1	0.2	0	0	0.2	0
opCcV0100000631	0.1	-0.1	0.3	0.1	0	0.1	-0.1	0.2	0.1	0
opCcV0100000633	2.4	2.1	-1.2	-0.3	-2.2	2.6	0.6	0.4	1.3	0.1
opCcV0100000637	-0.3	0	0	0	-0.1	0.2	-0.1	0	0	0

opCcV0100000640	-4.5	-3.6	0.2	-0.6	1.6	0.1	0	1.6	-1.3	-1.5
opCcV0100000643	0.1	0.1	-0.1	-0.1	0	0.1	0.1	0	0.1	0
opCcV0100000645	-1.8	-0.8	-0.3	0.1	0.2	0.3	0	0	0.1	-0.1
opCcV0100000651	1.3	2	1.6	0.1	0.3	0.9	0.6	-1.1	1.4	-0.4
opCcV0100000654	0.6	0.3	0.4	-0.1	0.1	0	0	0.2	0	-0.2
opCcV0100000655	0.5	-0.1	0.3	0.2	0.2	-0.7	0.1	-0.1	0.2	-0.1
opCcV0100000657	0.8	0.3	0.4	0.2	0.1	-1.4	0.4	0	-0.1	0.1
opCcV0100000658	-0.3	-0.4	-0.2	-0.1	0.1	0.2	-0.1	0.1	0	-0.1
opCcV0100000661	-0.2	0	-0.3	-0.1	-0.2	0	0	0	0.1	-0.1
opCcV0100000662	0.5	0.3	0.9	-0.1	-0.1	0	0.1	-0.2	-0.1	-0.1
opCcV0100000664	0.5	-0.3	-0.1	0	0	0.2	0.1	0	0	0
opCcV0100000666	0	-0.2	-0.1	-0.1	0.2	-0.2	0.1	-0.1	-0.1	-0.2
opCcV0100000672	0.9	0.6	0.7	0.2	0	0	0	-0.3	0.3	0.1
opCcV0100000674	-3.4	-0.7	-0.3	-0.2	0.1	0.1	-0.2	0.6	-0.2	-0.3
opCcV0100000681	0	-0.1	-0.8	0	0	-0.1	-2.4	0	0.1	0
opCcV0100000683	-0.2	-0.3	-0.2	0	0.1	0	0.1	0.1	-0.3	-0.1
opCcV0100000684	0	0	1	-0.1	-0.1	0.2	0	0.1	-0.1	-0.1

opCcV0100000685	-0.8	-1.9	2.8	0.2	-0.1	-0.2	0.3	0.1	-0.3	0
opCcV0100000691	-3.3	-3.5	0	-0.9	1	0	-0.6	1	-1.4	-0.9
opCcV0100000693	0.8	0.3	0.2	0.1	0.3	-0.1	0	-0.1	-0.1	-0.2
opCcV0100000696	2.7	1.6	1	0.3	-0.8	0.4	0.7	0	0.5	0.3
opCcV0100000698	-0.4	-0.4	1.2	-0.2	0	0.4	0.5	0.1	-0.1	-0.3
opCcV0100000699	0.6	0.2	0.4	0	-0.2	0.3	0.2	0	0.3	0
opCcV0100000700	0	-0.5	-2	-0.1	0	0.2	-0.1	-0.5	-0.1	-0.1
opCcV0100000702	0.3	-0.3	-0.3	0	0.1	-0.4	-0.5	0	0.1	-0.1
opCcV0100000703	-0.1	0	0.4	0	0	0.1	0	0	-0.1	-0.1
opCcV0100000704	-0.8	-0.7	-0.5	-0.1	0.3	-0.2	-0.1	0.1	-0.1	-0.2
opCcV0100000707	5.1	0.5	-0.3	-0.3	1.9	-0.6	-1.4	1.8	-0.8	-0.2
opCcV0100000708	0.7	-0.2	0.4	0.1	0.2	0.2	0.1	0.3	-0.3	-0.1
opCcV0100000710	-0.7	-0.5	0.2	-0.1	0.4	-0.3	0	0.5	-0.3	-0.4
opCcV0100000711	0.1	0	-3.6	-0.1	0.1	-1.9	-2	-1	0.1	-0.1
opCcV0100000714	0	0.2	0.2	-0.2	0	0	-0.1	-0.1	0.2	-0.1
opCcV0100000717	0.1	-0.1	-5.3	0	0	-2.2	-2.4	-1.7	0.3	0
opCcV0100000718	3.5	3.1	-0.2	0.9	-1.2	-0.6	-0.5	-0.7	1.2	1

opCcV0100000719	0.6	0.4	-0.5	0	-0.3	-0.1	0	-0.4	0.3	0.1
opCcV0100000721	4.9	3.9	-0.7	1	-1.9	0.5	0.9	-0.7	1.2	1.1
opCcV0100000723	-0.8	-0.2	-0.2	0	-0.3	0.2	-0.1	0.2	0.3	0
opCcV0100000724	0.5	-0.5	1.8	0	-0.1	0.1	0.1	0.3	0.1	0
opCcV0100000725	0.1	0.1	0.6	0	-0.1	0.3	0.1	0.2	0.2	0.1
opCcV0100000727	1.7	1.6	-0.8	-0.1	0.2	0	-0.1	0.6	-0.8	-0.3
opCcV0100000730	0	0.2	-0.2	0	-0.1	-0.3	-0.2	-0.4	0	-0.1
opCcV0100000733	0.2	0	-0.1	0	-0.1	0	0	-0.2	0	-0.1
opCcV0100000734	0	0.1	0	0	0.1	0	0.1	0.1	-0.1	0
opCcV0100000736	0.2	0.2	-0.3	-0.1	-0.1	-0.1	0	0	-0.1	0
opCcV0100000737	-1.2	-0.7	-0.6	-0.2	0.2	0	-0.3	0	-0.1	-0.2
opCcV0100000739	-3.2	-0.6	-2	-0.5	-0.1	-0.4	-1	0.3	0	-0.7
opCcV0100000743	0.3	0	0	0.1	0	0.1	0	0	0	0
opCcV0100000750	0	-0.4	0.1	0.1	0	0.3	0.1	0.4	-0.4	0
opCcV0100000751	-0.1	0.2	-1.2	0	-0.2	0.2	-0.1	-0.1	0.2	0
opCcV0100000756	2.5	1.6	1.2	0.2	-1.3	0.8	0.8	-0.2	1.4	-1.5
opCcV0100000757	-0.6	-0.5	-0.6	-0.2	-0.1	-0.1	0	0.2	0	-0.1

opCcV0100000765	-0.7	0.1	-1.8	-0.1	-0.1	0.1	-0.1	-0.1	-0.1	0.1
opCcV0100000767	-0.6	0.2	-0.6	-0.2	-0.3	0.2	-0.1	0	0.3	-0.1
opCcV0100000769	0.4	0.7	-0.3	0	-0.3	0.4	0.1	-0.1	0.2	0.1
opCcV0100000770	-0.4	-0.4	0	-0.1	-0.2	0.1	0	0.1	-0.1	-0.1
opCcV0100000778	2.7	2.2	2.5	0.6	-1.3	0.9	1	-0.5	1	0.8
opCcV0100000779	1.7	1.2	1.8	0.3	-0.6	0.7	0.7	-0.5	0.2	0.6
opCcV0100000786	-0.5	-0.4	-0.4	0	0	0.1	-0.3	-0.1	0	0
opCcV0100000787	0.4	0.5	0.4	0.1	-0.1	0	-0.1	0	0.3	0
opCcV0100000788	0.5	-0.1	0.7	0	-0.1	0.1	0.2	-0.1	0	0
opCcV0100000795	-0.2	-0.1	-4.3	0	0	-1.4	-1.6	-1.4	0	0
opCcV0100000797	0.5	0.5	0.3	0.3	-0.1	0.1	0.2	0	0	-0.1
opCcV0100000801	-1.5	-1.4	-0.4	1.6	0.7	-0.5	-1.2	0.8	-0.4	-0.5
opCcV0100000806	0	0.1	0	-0.1	-0.2	0.3	0	-0.1	0.2	0
opCcV0100000813	0.3	0.1	0.2	0	0.1	0.1	-0.1	-0.1	-0.1	-0.1
opCcV0100000814	-0.9	-1.4	0.5	0.1	0	-0.1	0	0.3	-0.3	-0.1
opCcV0100000816	0	0.1	0.1	0	-0.1	0.2	0	-0.1	0.2	0.1
opCcV0100000821	1.2	0.8	0.4	0	-0.6	0.3	0.2	-0.2	0.3	0.2

opCcV0100000826	-0.4	-0.7	0.5	-0.1	0.3	0.1	0	0.2	0.5	-0.1
opCcV0100000828	-2.7	-2.9	-0.2	-0.2	0.1	-0.2	-0.3	0.3	-0.6	-0.4
opCcV0100000829	0.4	0.8	0.7	0	-0.1	0.1	0	-0.2	0	0
opCcV0100000830	-0.9	-0.2	-3.9	-0.1	0.1	-1	-1.3	-1.4	0	0
opCcV0100000834	-0.1	0	-0.4	0	0	-0.4	-0.1	-0.1	0	-0.1
opCcV0100000835	0.8	0.6	-5.4	0	-0.2	-2.1	-2.3	0	-0.5	0.1
opCcV0100000840	-0.9	-0.7	-0.4	0	0.3	0.2	0	0	-0.1	0
opCcV0100000845	-2.1	-1.3	-0.6	-0.2	0.2	0	-0.3	0.4	-0.3	0.1
opCcV0100000852	-0.1	-0.2	0	0	0.1	0.2	0	-0.3	0	0
opCcV0100000853	0.1	-1.2	-0.3	-0.4	0.6	0	0.1	0.5	-0.2	-0.1
opCcV0100000856	-0.6	-0.4	0.4	-0.1	-0.1	0.1	-0.7	0	0	0.1
opCcV0100000857	0.5	0.2	0.3	0.1	0	0.2	0.1	0	0.1	-0.1
opCcV0100000860	1.1	-0.6	0.8	0.1	-0.4	0.5	0.5	0.4	0.1	0
opCcV0100000861	0.4	0.3	-0.4	0.2	-0.3	-0.2	0.1	-0.3	0.2	0.1
opCcV0100000864	0.2	0.2	0	0.1	-0.1	-0.1	0	0	0.1	-0.1
opCcV0100000865	0.2	0.1	-4	0	-0.2	-1.3	-1.9	0.1	0.1	0
opCcV0100000867	-1.6	0.2	0.2	-0.1	0.4	0.2	0.4	0.4	-0.9	-0.3

opCcV0100000868	0	0.1	0.7	0.1	0.6	0.2	0.3	0.5	0	0.1
opCcV0100000873	0.3	0.1	0.5	0	0.2	0	-0.1	-0.1	0.1	0
opCcV0100000875	-0.7	-0.7	-0.4	-0.1	-0.1	0.1	-0.3	-0.1	-0.2	-0.2
opCcV0100000877	-2.2	-2	-0.5	0	0	0	-0.5	0.4	-0.1	-0.1
opCcV0100000878	0.6	0.3	0.8	0.1	0.1	0.1	0.2	-0.2	0	0
opCcV0100000879	-0.8	-0.6	-0.4	-0.1	0.1	-0.4	-0.7	0.3	-0.7	-0.2
opCcV0100000882	-1	-0.3	-1.5	-0.5	0	0.2	-0.4	0	0	-0.3
opCcV0100000884	0	-0.1	-0.1	0.1	-1.6	-0.1	-0.1	-0.1	0.1	-0.2
opCcV0100000885	0.3	0.1	0.2	0.2	0.1	0.1	0.1	-0.5	0	0
opCcV0100000887	0.3	-0.6	0.2	0.3	0.3	-0.6	0	0	-0.1	-0.2
opCcV0100000888	0.3	0	-0.2	0	-0.2	-1.5	0.1	0	0	-0.9
opCcV0100000890	-0.1	-4.4	0.1	-1.1	2.5	0.2	-1	1.8	-1.5	-2
opCcV0100000894	0.3	0	0.2	-0.1	-0.1	0.1	0.1	0	0.1	0
opCcV0100000895	0.4	0	0.2	0.1	0.2	0	0	0	-0.3	-0.1
opCcV0100000896	0.3	-0.2	0.5	0	0.1	0.2	0.5	0	-0.3	0
opCcV0100000898	1.6	1.4	1.6	0.3	-0.6	0.4	0.4	0	0.3	0.1
opCcV0100000900	0.4	0.1	0	0.1	0	0.1	0.1	-0.1	0	0

opCcV0100000903	0.2	0.6	0.2	0.2	0	-0.1	0	-0.2	0	0
opCcV0100000904	0	0.2	-0.1	0	0.9	-2.1	-2.7	0	0.2	-0.1
opCcV0100000905	-0.5	-0.3	0.1	-0.1	-0.3	0.3	0.1	-0.1	0.1	0.1
opCcV0100000908	0.2	0.1	0.2	-0.1	-0.1	0	0.1	-0.1	0	-0.1
opCcV0100000909	0.5	0.1	-0.7	0.1	0	-0.7	-0.6	-0.4	-0.1	-0.1
opCcV0100000910	-0.6	-0.7	-1	-0.2	-0.1	0.1	-0.3	0.1	0.3	-0.1
opCcV0100000911	-0.3	-0.2	0	0.1	0.2	0	0.1	0.2	-0.2	-0.2
opCcV0100000912	0.2	0	0.2	-0.1	-0.1	0.2	0.1	0	0	0
opCcV0100000915	0.2	0.1	-4.3	-0.1	-0.1	-1.4	-2.2	0.1	0	0
opCcV0100000917	0.7	0.2	0.3	0	0	0.3	0.4	0.2	-0.2	-0.2
opCcV0100000918	0	0	0.1	0	-0.1	0.2	0.1	0.1	-0.1	0
opCcV0100000921	-1	-0.5	-3.2	-0.3	0.2	-2.9	-3.2	-0.9	0.3	0
opCcV0100000922	0.1	0	0	0.1	0.1	0.1	-0.4	0	0	0.1
opCcV0100000923	-2.8	-3.3	-3.7	-1.1	0.6	-1	-1.3	0.4	-0.4	-1.1
opCcV0100000924	0.4	0	0.3	-0.1	0.1	0	0	0.1	0.1	-0.1
opCcV0100000928	0.2	-0.8	-1.7	0.1	-0.5	0.1	-0.6	0.5	0.4	-0.1
opCcV0100000931	-1.4	-1.3	-0.1	-0.4	2.3	0	0	0.6	-0.2	-0.4

opCcV0100000932	0.5	0.3	-0.3	0	-0.4	0.1	0.2	-0.2	0.1	0.1
opCcV0100000934	-0.3	-0.3	-0.3	0	0	0.2	0	-0.1	-0.1	-0.1
opCcV0100000937	0.3	0	0.1	0	0	0.1	0	0	-0.1	0
opCcV0100000943	0.5	0.2	0.2	0	-0.1	0	0.1	0	0	-0.1
opCcV0100000951	0.2	-0.3	0.2	-0.1	-0.1	0.1	-0.5	0.4	0	0
opCcV0100000954	-0.1	-1.4	0	-0.3	0	0.1	0	0	-0.3	-0.4
opCcV0100000958	-0.6	0.1	0	-0.1	0	0.1	-0.1	-0.3	0.4	0
opCcV0100000962	0.7	0.1	0.6	0	0.2	0	0.2	0	-0.2	-0.1
opCcV0100000965	-2.4	0.2	-2	0.9	0.7	-1	1.6	0.7	-0.2	0.1
opCcV0100000966	-0.9	-0.7	-1.2	-0.1	0.3	-0.4	-0.7	0	-0.3	0.1
opCcV0100000970	-2.3	-1.9	-1.1	-0.5	0.5	-0.1	-0.6	1	-0.4	-0.5
opCcV0100000973	-0.3	0.4	0.6	0	0	0.1	0	-0.1	0.3	0
opCcV0100000975	0.3	0.1	-4.3	-0.1	-0.2	-1.3	0.1	0	0.3	0
opCcV0100000976	-0.3	0.2	-0.1	0	-0.1	0.2	0.1	-0.1	0.1	0.2
opCcV0100000981	0.6	-0.2	0.6	0	0.1	-0.1	-0.1	0.1	-0.3	-0.1
opCcV0100000987	-0.9	-0.4	0.1	0.2	0.6	-0.4	0	0.7	-0.5	-0.3
opCcV0100000989	-1.2	-0.8	-0.3	-0.2	0.1	-0.2	-0.1	1	-0.3	-0.1

opCcV0100000996	-0.1	0.4	0.8	0.4	-0.7	0.3	0	-0.2	-0.2	0.6
opCcV0100000998	-0.2	-0.2	-0.2	0.1	0	0.3	0	-0.1	0	0.1
opCcV0100000999	-0.5	-0.4	0.2	0	0.1	0.1	0.1	0.1	-0.1	0
opCcV0100001000	0.3	0.1	0.4	0	-0.2	-0.1	0	0.2	-0.3	0
opCcV0100001001	1	0.5	0.3	0.2	-0.2	0.2	0.2	-0.3	0	0.1
opCcV0100001004	0.2	0	0.5	0	-0.1	0.3	0.3	-0.1	0	-0.1
opCcV0100001007	-0.2	-0.3	-2.5	-0.1	0	-0.6	-0.1	0	0	0
opCcV0100001009	-0.1	-0.3	-2.8	-0.1	0.1	-1.5	-0.1	0.1	0.1	-0.1
opCcV0100001010	0	-0.2	0	0	0	0.1	0.1	0.1	0	0
opCcV0100001011	0.3	-0.2	0.1	0.2	0.2	-0.1	0	0.3	-0.4	-0.3
opCcV0100001012	-0.1	-0.1	0.1	0	0	0.2	0.1	0	-0.1	0
opCcV0100001013	-5.7	-3.8	-0.9	-2.7	3.9	-1.9	-1.9	1	-3.7	-1.8
opCcV0100001015	-0.1	0.1	0.8	0	0	0.3	0.2	-0.2	-0.1	0.1
opCcV0100001017	1.3	-0.9	0.8	0.1	0.2	0.3	0.6	0.6	-0.7	0
opCcV0100001019	-0.1	-0.1	0	0.1	0	0.1	0	-0.2	0.2	0.1
opCcV0100001021	-0.5	0.3	-0.4	0	-0.1	-0.1	-0.1	0.2	0	-0.1
opCcV0100001023	0	-1.9	-0.6	-0.3	0.5	-0.3	-0.3	0.2	-0.5	-0.4

opCcV0100001025	-0.1	-0.5	0.1	0	0.3	-0.2	0	0.1	-0.2	-0.1
opCcV0100001026	1.6	0.6	0.8	-0.1	-1.2	0.3	0.6	-1.1	1	-0.4
opCcV0100001029	-0.5	-0.8	-0.5	-0.1	0	0.2	0	-0.1	-0.2	-0.1
opCcV0100001030	0.8	0	0.4	0.1	0.1	0.2	0.3	0.1	-0.2	-0.1
opCcV0100001032	0.4	0	0.7	-0.1	0.2	-0.1	0	-1	-0.2	-0.2
opCcV0100001034	-0.1	0.1	-0.5	-0.2	-0.1	-0.1	-0.1	0	0.1	-0.1
opCcV0100001037	-0.8	-0.3	-1.7	-0.2	0	0.1	-0.2	0.7	0	-0.5
opCcV0100001038	0.2	0	0.2	0.1	0.1	0	0.1	0.1	-0.2	-0.1
opCcV0100001047	0.4	0.3	-5.3	0.1	-0.2	-1.8	-1.9	-0.1	0.2	0.1
opCcV0100001049	-2.3	-1.8	-0.2	-0.1	0.3	-0.2	-0.2	0.4	-0.3	0.1
opCcV0100001051	0.3	0	-0.1	0	0	0	0	0	0	-0.1
opCcV0100001058	0.9	0	-0.1	0	0.1	-0.1	0.1	0.2	-0.2	-0.1
opCcV0100001061	-0.8	-0.5	-0.4	-0.1	-0.1	0.1	-0.1	0	0	-0.1
opCcV0100001062	0.2	-0.1	0	0.1	0.3	-0.1	0	0.1	-0.2	-0.2
opCcV0100001064	-0.1	-0.2	0	-0.1	0	0.2	0	0	0.2	0
opCcV0100001066	-3.1	-2.6	-0.5	-0.1	0	-0.1	-0.3	0.3	1	-0.1
opCcV0100001071	-2.6	-1	-1.6	-0.2	0	-0.5	-1.3	0.2	-0.3	-0.1

opCcV0100001072	1	0.9	0.5	0.2	-0.2	0.1	0.2	-0.1	0.5	-0.1
opCcV0100001073	0.7	0.6	0.4	0.1	-0.2	0.1	0.1	-0.2	0.1	0
opCcV0100001075	0	0.1	0.2	-0.1	-0.1	0.2	0	0.1	0.1	0
opCcV0100001078	0.3	0	0.1	-0.1	0	0.2	0.1	0	-0.1	0
opCcV0100001079	0.3	0.1	0.3	0.1	0	0	0.1	0.1	-0.1	-0.1
opCcV0100001080	-0.4	-0.4	-0.6	0	0	0.1	0	0.1	0	-0.1
opCcV0100001081	1.2	-0.1	0.1	0	-0.2	0	-0.1	0.3	0.3	0
opCcV0100001082	2.8	2.6	1.8	0.7	-0.7	0.2	0.4	-0.4	0.4	0.4
opCcV0100001086	0.1	0.1	-2.7	-0.2	0	-1	-1.2	0.2	0	-0.1
opCcV0100001089	0.7	0.3	0.6	0.1	0.1	-0.1	0	-0.1	0.1	0
opCcV0100001090	-3.1	-0.5	1.1	-0.6	-0.1	0.5	0.3	1.1	0	-0.5
opCcV0100001098	1.4	1	-1.2	-0.1	-0.1	-1.6	-2.2	-1.8	0.1	-0.3
opCcV0100001100	-0.3	-0.1	-0.1	0	0	0	-0.2	0.1	-0.1	0
opCcV0100001102	-0.7	0.2	-0.1	-0.2	-0.4	0.3	0.1	-0.1	0.3	0.1
opCcV0100001107	0.5	0.5	0.9	0.2	0.2	-0.2	0.5	0.2	0.1	0.1
opCcV0100001110	-4.7	-3.4	-0.7	-1.3	1.1	-1.1	-1	0.8	-1.3	-1.2
opCcV0100001111.50	2.1	-0.5	-0.3	-1.7	-0.7	-1.3	0.3	0.3	1.9	0

opCcV0100001111.70	1.6	0.7	-1.2	0	-1.1	-0.1	1.8	0.4	1.9	-0.5
opCcV0100001111.90	1.3	0.5	-0.6	0.2	-1	-0.3	-0.2	0	1.6	-0.3
opCcV0100001111.A	-0.5	0.1	-0.7	-0.2	0.5	0.2	-0.1	0	0.3	0
opCcV0100001111	0.7	0.5	0.7	0	-0.1	0.1	0.1	0.1	0.3	-0.1
opCcV0100001112	0.2	0.2	0.3	0	0.1	0	0	0	0	-0.1
opCcV0100001113	-0.4	-0.4	-0.1	0	0.1	0.1	-0.6	0	-0.1	0
opCcV0100001117	0.5	0.4	0.5	0	0	-1.4	0	0	0.1	-0.1
opCcV0100001120	-0.1	-0.1	-0.3	0	-0.1	0.2	0.1	-0.1	0.2	0
opCcV0100001125	0	-0.3	-0.1	-0.3	-0.1	0.1	-0.1	0	0.1	-0.3
opCcV0100001126	-0.4	-1	-1	0.2	0.1	0	-0.1	0.2	-0.4	0.1
opCcV0100001127	-0.9	-0.6	-0.9	-0.1	0	0.5	0.1	0.1	0.2	0
opCcV0100001129	-1.9	-0.3	-0.5	-0.2	0.3	-0.2	-0.4	0.1	-0.4	-0.4
opCcV0100001135	-0.8	-2.3	0	0	0.3	-1.5	0.4	0.3	-1.2	0.1
opCcV0100001137	-1.3	-1.3	0.3	-0.2	0.1	0.1	0	0	-0.2	0
opCcV0100001138	-1.6	-0.5	-0.2	0	-0.1	0	-0.8	-0.2	-0.2	-0.1
opCcV0100001143	-1.2	-2.6	-0.2	-0.2	0.2	-0.1	-0.2	0.1	-0.4	-0.2
opCcV0100001146	-0.1	0.2	0.1	0	-0.1	0.1	0	0	0.1	0

opCcV0100001149	-0.8	-0.8	0.6	0	0.2	0.1	0.2	0	-0.3	0
opCcV0100001150	0.2	0	-0.2	-0.1	0	0	0.1	0	0	-0.1
opCcV0100001151	0	0.2	-4.3	-0.1	-0.1	-1.3	-1.5	-1.1	0.1	-0.1
opCcV0100001159	-0.1	-0.1	0.1	-0.1	0.1	0.1	-0.1	0.1	0	0
opCcV0100001161	0.7	0.2	0.6	-0.1	0	0	0.1	-0.1	0.1	0
opCcV0100001164	-0.5	-0.4	-0.3	-0.1	0.1	0	-0.1	-0.1	0	0
opCcV0100001167	1.8	1.4	-0.4	-1.4	0.5	0	0.8	0.6	1	-1.3
opCcV0100001169	0.3	-0.2	0.2	0	-0.2	-0.1	0.2	0.1	-0.3	-0.1
opCcV0100001170	0.6	0.3	-2.4	-0.1	-0.3	-1.6	-2.1	0	0.3	0.1
opCcV0100001173	1	0.5	0.6	0.1	-0.1	0	0.1	0.1	0	0
opCcV0100001174	-0.9	-0.7	-0.7	-0.1	0.2	0.1	0	0.2	-0.1	-0.1
opCcV0100001175	1.3	-1.2	1.3	-0.3	-0.4	1	0	0.5	-0.2	0.3
opCcV0100001190	2.1	-1.2	1.8	-0.2	0.5	0	0.4	0.5	-0.4	-0.4
opCcV0100001191	0.6	-0.3	1.1	-0.2	-0.1	0.3	0.2	0.3	0.6	0
opCcV0100001194	0.9	0.4	0.5	0	0.1	-0.2	0.1	0	-0.2	-0.2
opCcV0100001196	0.7	0.2	0.3	0.1	-0.1	0.2	0	0	0.1	0.1
opCcV0100001198	-1.4	-0.3	-0.9	-0.1	-0.1	-0.3	-0.1	0.7	-0.3	0

opCcV0100001199	0.4	0.7	0.2	0	-0.2	0.1	0	0.1	0.1	0
opCcV0100001200.70	2	1.5	-0.4	-0.9	-2.5	-1.1	0.1	0.7	1.6	0.5
opCcV0100001200.A	0.9	1.3	0.2	0	-0.8	0.4	0.5	0	1	0.4
opCcV0100001200.C	3.5	0.5	-0.2	-0.6	-1.3	-0.5	-0.5	0.5	2	-0.9
opCcV0100001200	2.2	0.3	1	-0.2	-0.3	-0.1	0.3	0.3	0.2	-0.1
opCcV0100001203	-1.8	-0.2	-0.3	-0.1	-0.2	0.1	0	0.3	0.1	0.3
opCcV0100001204	-0.8	0.1	-0.2	0	0	0.1	0	-0.3	0	0
opCcV0100001205	0	0.1	0	0	0	0.2	0.1	0	0.1	-0.1
opCcV0100001210	-0.4	-0.1	-0.4	-0.1	0	-0.2	-0.1	0	0.2	0.1
opCcV0100001212	-0.3	-0.2	0.4	-0.1	0.1	-0.2	0	-0.3	0.5	0
opCcV0100001213	0.3	0.3	0.2	0	0.3	0	0	0	0	-0.2
opCcV0100001214	-0.6	-0.3	1.5	-0.1	0.1	-0.3	-0.2	0.2	-0.1	-0.1
opCcV0100001216	-0.6	-1	-1.1	0	0.2	-0.7	-0.4	-0.3	0	-0.1
opCcV0100001219	-0.5	-0.2	-0.3	-0.1	0	0.3	-0.1	0.1	0	0
opCcV0100001221	-2.5	-1.8	-0.9	-0.3	0.4	0.3	0	0.8	-0.2	-0.4
opCcV0100001222	0.7	0.3	0.2	0.1	0	-0.1	0.1	0	0	-0.1
opCcV0100001223	0.2	-0.1	0.7	-0.1	-0.2	0	0	0	-0.1	0

opCcV0100001225	-0.7	0.1	-1.2	0	-0.4	0.2	-0.1	-0.1	0.3	0.1
opCcV0100001231	-2	0.4	-0.5	0	0	-0.3	-0.5	-0.5	0.1	0.1
opCcV0100001232	-5.2	-5	-1.1	-0.8	1.6	-1.3	-1.5	0.5	-1.7	-0.8
opCcV0100001233	-0.7	-0.7	-0.3	0.1	0.3	-0.2	-0.1	-0.3	0	0.1
opCcV0100001236	-0.8	-0.8	-3.4	-0.4	0.2	0.2	-0.2	-0.5	-0.5	-0.3
opCcV0100001238	0.8	0.9	0.9	0	-0.3	0.2	0.2	-0.2	0.4	0
opCcV0100001240	0.4	0.9	0.4	0.3	-0.1	0.2	0.3	-0.1	0	0.2
opCcV0100001242	0.3	0.2	0	0	-0.1	0	0.2	0	0	0
opCcV0100001244	0.2	0.1	0.4	-0.1	-0.1	0.2	0.1	-0.1	-0.2	0
opCcV0100001246	0.6	0.4	0.2	0	0.1	-0.1	0	-0.1	-0.2	-0.1
opCcV0100001248	-2.6	-1.9	-1.5	-0.6	0.9	-0.1	-0.2	0.2	-0.5	-0.6
opCcV0100001251	-0.8	-0.7	0.3	-0.4	0.4	-0.2	0	0.4	-0.2	-0.3
opCcV0100001252	0.3	0.1	-0.1	0	0	0	0.1	0.1	0	-0.1
opCcV0100001255	-0.2	0	-3.5	-0.1	0.1	-0.9	0	-0.1	0.2	0.1
opCcV0100001260	1	0	1.3	0.2	0.7	0.4	0.3	-0.1	0	0.1
opCcV0100001261	-0.7	-0.6	0.1	-0.3	0.2	0.2	-0.1	0.1	-0.1	-0.1
opCcV0100001263	1.1	-0.8	0.7	0.4	0	0.3	0.3	0.4	0.4	0.2

opCcV0100001264.60	3.3	1.7	0.6	0.4	-1.7	0.1	0.4	0.9	0.9	-0.4
opCcV0100001264.70	1.9	1.2	-0.9	-0.9	-1.6	0.2	0.5	1.6	2.5	-1.2
opCcV0100001264.80	2.3	6.5	-1	0.8	-0.4	-0.8	4	0.6	0.1	0.6
opCcV0100001264.90	-1	-1.2	-1.3	-0.1	0.3	0.1	0.3	-0.2	-0.3	-0.3
opCcV0100001264.A	-2.7	-2.2	-4.5	-0.5	1	0.2	0.2	0.2	-0.1	-0.4
opCcV0100001264	-1.5	-2.2	-3.2	-0.2	0.9	-0.1	0.5	0	-0.4	-0.2
opCcV0100001265	1	-3	0	-0.7	1	0.2	-0.1	1.9	0.2	-0.9
opCcV0100001266	0.3	0.2	0.9	0	0.1	0.3	0.4	0.2	-0.1	-0.1
opCcV0100001268	1.1	0.7	0.6	0.1	-0.2	0.2	0.3	-0.1	0.1	0.1
opCcV0100001270	-1.9	-0.8	-0.3	-0.1	-0.1	0.1	-0.1	0.1	-0.1	0
opCcV0100001271	-0.5	0	0.3	-0.1	-0.1	0	0.1	0.1	0.2	0
opCcV0100001272	-1.2	-0.7	-0.2	0.2	0.4	-0.3	0.2	0.4	-0.7	0
opCcV0100001273	-0.1	0.2	0.1	-0.1	-0.4	0.2	0	-0.1	0.1	0.1
opCcV0100001275	0.7	1.4	0.5	-0.1	0.7	0.7	-0.4	1.4	-0.6	0.3
opCcV0100001276	0.3	0	0.1	-0.1	0	0.2	0	0	0	0
opCcV0100001278	1.5	-0.3	1.8	0.1	0.2	0.7	0.2	0.5	0.8	0.3
opCcV0100001279	-1	-0.6	-2.7	0	0.1	0	0	-1.2	0	-0.1

opCcV0100001281	-1.9	-2.1	-1	0	1.2	-0.1	-0.3	0.3	-0.6	-0.2
opCcV0100001286	0.2	0	0.3	0.1	0.1	0.1	-0.1	0.2	0.1	0
opCcV0100001288	-0.1	0.2	-0.2	-0.1	-0.2	0.2	-0.2	0	0.1	0.2
opCcV0100001290	1.5	-0.5	-0.1	0.3	0.1	0	0	0	-0.3	0.3
opCcV0100001291	2.8	1.5	1.5	0.2	-0.9	0.5	0.6	-0.2	0.6	0.3
opCcV0100001292.50	3.1	1.3	-0.9	-2.1	0.2	-0.1	0.5	0.7	-0.4	-1.4
opCcV0100001292.90	0	-0.1	-0.1	-0.2	0.1	-0.3	-0.9	-1	0.1	-0.1
opCcV0100001292.A	-0.1	0	-0.1	-0.1	0	0.1	0	-0.8	0.1	0
opCcV0100001292.C	3.1	1.1	0	-0.7	0.4	0.1	0.4	1.2	2.2	-1.9
opCcV0100001292	0.1	0	0.1	-0.1	0	0	-0.1	-0.6	0	-0.1
opCcV0100001294	0.6	0.7	0.2	0.1	-0.4	0.2	0.1	-0.1	0.1	0.1
opCcV0100001295	-0.9	-0.5	-0.3	-0.2	0.1	0.1	-0.1	0	-0.2	-0.1
opCcV0100001297	0.4	-0.5	0.2	0.1	0.2	-0.1	-0.1	0.2	-0.2	-0.1
opCcV0100001299	0.4	0.1	0.4	0.3	-0.1	0.2	0.1	0	-0.1	0
opCcV0100001302	-0.2	-0.3	-0.1	0	0	0.1	-0.1	-0.1	0.1	0
opCcV0100001305	-0.7	-0.9	-0.7	0.2	-0.2	-0.2	-0.2	0.7	-1.2	-0.1
opCcV0100001307	-0.2	-0.2	0.2	0	0	0.2	-0.1	-0.1	0.2	0.1

opCcV0100001308	-0.1	-0.3	0.1	0	0.3	0	0	0	-0.1	-0.1
opCcV0100001309	-3.1	-3.3	-1.1	-1.2	1.1	-0.6	-1	1.7	-1.5	-1.3
opCcV0100001310	-0.1	-0.3	0.3	-0.1	0.1	0	0	0.1	-0.2	-0.1
opCcV0100001311	-0.4	-0.2	-0.7	-0.1	-0.2	-0.2	-0.2	0	0.2	-0.1
opCcV0100001316	0.5	0.3	-2.9	0.1	-0.2	-1	-1.3	-0.1	0	0
opCcV0100001319	-4.3	-1.7	-4.5	-0.3	0.5	-0.6	-1.1	-0.9	-0.3	-0.4
opCcV0100001320	0.2	-0.2	0.5	0	0.2	0.3	0.3	0.1	0.1	-0.1
opCcV0100001322	1.8	0.7	1.8	0.3	-1	0.7	0.8	-0.4	0.7	0.6
opCcV0100001324	1.7	1.3	1.4	0.3	-0.6	0.5	0.6	-0.4	0.7	0.5
opCcV0100001325	-0.1	-0.3	0.2	-0.1	0.1	0.1	0	0.1	-0.2	-0.2
opCcV0100001326	0.1	0.4	0.2	-0.2	-0.4	0.3	0	0	-0.1	-0.1
opCcV0100001327	0.5	-0.2	0.5	0.1	0	-0.1	-0.1	0	0	-0.1
opCcV0100001332	-0.6	-0.4	0.2	-0.1	0	-0.1	0.3	0.1	-0.1	0
opCcV0100001340	-0.1	0	0.1	-0.1	-0.1	-0.1	-0.1	0.1	-0.3	-0.1
opCcV0100001341	-0.4	-0.8	2.5	0.2	1.2	1	1.4	0.3	-0.2	0.2
opCcV0100001344	-0.2	-0.6	0.5	0.1	0.3	0.1	0.3	0.3	-0.3	-0.1
opCcV0100001345	-0.3	0.3	0.1	0	-0.3	0	-0.4	0.2	-0.2	0

opCcV0100001347	0.7	0.3	0.1	0	-0.1	0	0	-0.1	-0.1	0
opCcV0100001353	0	0	-6.5	0	0	0.1	-0.1	-1.9	0.1	0
opCcV0100001356	0.1	-1.3	-0.5	0.1	0.3	-0.1	-0.1	0.4	-0.7	-0.1
opCcV0100001357	0.4	0.4	0.1	0	0	0	-0.1	0	0.1	0
opCcV0100001358	0.4	0.4	0.5	0.1	-0.2	-0.3	0.2	-0.1	-0.1	-0.1
opCcV0100001361	0.7	-0.9	0.3	0	0.2	-0.1	0.2	0.7	-0.3	-0.2
opCcV0100001367	1.7	1.6	0.4	-2	-1	0.2	-0.5	0.1	1.2	-1.6
opCcV0100001370	0.5	0.3	0.5	0	0	0.1	0.1	0.1	-0.1	-0.1
opCcV0100001371	-0.2	-0.4	0.2	0.1	0.2	-0.2	0.2	-0.1	-0.2	-0.1
opCcV0100001377	0.2	0.1	0.4	0.1	0.1	0	0	0.1	-0.1	-0.1
opCcV0100001378	0.5	0.1	0.3	0	0	0.1	0.1	-0.1	-0.1	0
opCcV0100001381	0.7	0.4	0.4	0.2	0	0	0	-0.1	0	0.1
opCcV0100001383	-0.5	-0.2	0.5	-0.1	0.2	0	0	0.2	-0.1	-0.1
opCcV0100001389	0	0.1	0.3	0	0.1	0.2	0	0.2	-0.1	0
opCcV0100001390	-0.6	-0.5	-0.5	0	0.1	0.1	0	-0.1	0	0
opCcV0100001394	-0.2	-0.3	0.1	-0.2	-0.1	0.2	-0.2	0.1	0	-0.1
opCcV0100001395	-0.5	-0.2	-7.4	-0.1	-0.1	0.2	0	-1.7	0.1	0

opCcV0100001399	-1.1	0	0.6	0.1	0.1	0.3	0.2	0.2	-0.6	0
opCcV0100001402	0.3	-0.1	0.3	0	0.2	0.1	-0.4	-0.4	0.1	0
opCcV0100001404	-0.2	-0.5	0.7	0	0.2	-0.1	0.1	0.2	-0.2	-0.2
opCcV0100001406	0.9	0.2	0.2	0.1	0.1	0	0	-0.1	0	0
opCcV0100001411	-5.4	-4.1	-4.3	-0.3	2.7	-1.4	-2.1	2	-1.5	-1.6
opCcV0100001414	-0.1	-0.4	0.5	0.1	0.3	0.3	0.3	0.7	-0.1	-0.2
opCcV0100001415.80	-0.8	-0.5	0.2	-0.1	-0.3	-1	-1.1	0.2	-0.4	-0.1
opCcV0100001415.90	0.1	-0.2	-0.2	-0.1	-0.1	-0.2	-0.2	0	-0.1	-0.1
opCcV0100001415.A	-0.8	-0.4	-0.8	-0.1	0	0	-0.3	0.1	0.1	-0.1
opCcV0100001415.C	1.3	1	-0.2	0	-1.1	-0.7	-0.2	-0.3	1	-0.1
opCcV0100001415	0.3	0	0	0	0	-0.1	-0.1	0	-0.1	-0.2
opCcV0100001418	0.2	0	0.4	0	-0.1	0.2	0	-0.1	-0.1	0
opCcV0100001421	-0.8	0	0	-0.1	-0.1	0	-0.3	-0.1	0.1	0
opCcV0100001423	0.3	0.2	0.3	0.2	0	0.1	0	0	0.1	-0.1
opCcV0100001424	1.5	0.5	2.5	0.5	-0.7	0.2	0.6	-0.4	0.5	0.5
opCcV0100001425	-0.3	-0.2	0.9	-0.1	-0.1	0.3	0.3	0.4	0	-0.1
opCcV0100001431	2.1	-1.5	1.3	-0.1	0.2	0.5	0.1	0.6	0.3	0

opCcV0100001432	-5.2	0.4	-2.6	-0.8	0.7	-1.2	-1.1	0.6	-1.2	-0.5
opCcV0100001435	0.6	0.4	0.3	0	-0.3	0.3	0.2	-0.1	0.2	0
opCcV0100001437	-0.6	-0.5	-1.2	-0.2	0.2	-0.1	0	0.1	-0.2	-0.2
opCcV0100001440	0.3	0	0	0	0.2	-0.1	0	0.1	-0.2	-0.2
opCcV0100001442	1.1	-1.2	0.5	-0.1	0.2	-0.2	-0.5	-0.2	-0.9	-0.2
opCcV0100001444	-0.8	-0.8	-0.2	-0.2	0.1	0.1	-0.4	0.6	-0.1	-0.2
opCcV0100001446	0.5	0.2	0.2	0.1	0.1	0	-0.1	-0.2	0	-0.1
opCcV0100001448	-1.1	0.4	0.5	0	0.1	0	0	0.1	0.2	0
opCcV0100001449	0.3	0.1	0.5	0.1	0.1	0.1	0.1	0	-0.1	-0.1
opCcV0100001451	2.2	-0.2	0.4	0.4	0.3	0	-0.5	0.9	-1.5	-0.2
opCcV0100001452	0	-0.2	-0.6	-0.1	0.1	-0.5	0.2	-0.3	-0.1	0
opCcV0100001453	0.2	0	0.3	0	0.1	0.2	0.2	0	-0.1	0
opCcV0100001454	-0.6	-1.3	-0.1	-0.1	0.3	0.1	-0.3	-0.2	0.2	0
opCcV0100001455	0.5	0.3	0.2	0.1	0	-0.1	0.2	-0.1	0	-0.1
opCcV0100001456	0	-0.1	0	-0.1	-0.1	0	0	0	0	-0.2
opCcV0100001459	2.2	0.9	-0.1	0.2	-0.4	-0.4	-0.5	-0.2	0.2	0.4
opCcV0100001461	-4.1	-0.4	-0.3	0	0.2	-0.2	-0.3	0.5	-0.3	-0.2

opCcV0100001462	-0.1	-0.7	-0.3	-0.2	0.1	0	-0.5	0.3	-0.2	-0.2
opCcV0100001465	-0.2	-0.1	0.1	-0.1	0	-0.1	0	-0.5	1	-0.1
opCcV0100001466	-0.8	-0.5	-1.8	0	0	0.1	-0.1	0.1	0	-0.1
opCcV0100001467	0.2	0	0.1	0	-0.1	0	0	-0.1	0	-0.1
opCcV0100001471	-1.8	-1.3	0.2	-0.3	0.1	-0.2	0.4	0.3	-0.6	-0.4
opCcV0100001476	-0.9	-0.6	0.3	-0.1	0	-0.2	0	0.3	-0.1	-0.1
opCcV0100001479	-0.8	-0.3	0.1	0	0	0	0.1	0.1	-0.2	-0.2
opCcV0100001480	-2.3	-0.2	-0.1	-0.4	-0.1	0.1	-0.1	0.6	-0.5	-0.4
opCcV0100001482	-0.6	-0.3	0.5	0	0.1	0.2	0	-0.4	0.1	0
opCcV0100001483	0.3	0	0.2	0	0	0	0.1	-0.1	0	-0.1
opCcV0100001488	-0.8	-1.5	-0.8	-0.2	0.6	-0.3	-0.5	0.6	-0.3	-0.2
opCcV0100001489	0.9	-0.2	1.9	0.5	0	0.2	0.8	-0.1	0.2	0.5
opCcV0100001493	-0.6	-1.6	-0.3	0	0	0	0	0.1	0.1	0.1
opCcV0100001498	2.1	1.7	0.2	0.6	-0.8	0	-0.1	-0.5	0.7	0.5
opCcV0100001501	-0.7	-0.3	-1	-0.1	0	-0.6	-0.3	0	0.1	-0.1
opCcV0100001504	-0.2	0	-0.5	-0.1	0	-0.9	-0.3	-0.5	-0.3	0.2
opCcV0100001513	-1.3	-1.1	0.9	0	0.4	0.1	-0.2	0.1	-0.5	-0.3

opCcV0100001515	-1.4	-0.9	-0.5	0	0.3	0.1	-0.6	0.5	-1.4	-0.1
opCcV0100001519	-1.7	-1.5	-1	-0.4	0.3	-0.1	-0.2	0	-0.4	-0.1
opCcV0100001521	-0.4	-0.9	-2.3	-0.2	-0.4	-0.2	-0.4	-0.1	-0.3	-0.1
opCcV0100001522	-0.1	-0.6	0	-0.3	-0.1	0.2	-0.1	0.3	-0.2	-0.3
opCcV0100001526	0.7	-0.4	0.6	0	-0.3	0.2	-1	0.1	0.1	0
opCcV0100001528	-0.8	0.2	0	0	0	-0.1	-0.1	0	-0.1	-0.1
opCcV0100001535	0.1	-1.1	-1.1	0	0.3	0	-0.4	0.2	-0.3	0
opCcV0100001536	0	-1.2	0	0	0.3	0	0	0.3	-1.3	0
opCcV0100001537	1.2	1.1	-0.1	0.2	-0.2	-0.1	0.1	-0.4	0.3	0.2
opCcV0100001538	0	-0.2	-0.3	-0.1	-0.1	0.2	-0.1	-0.1	-0.2	-0.1
opCcV0100001540	-0.7	-0.7	-0.3	0	0.2	-0.1	-0.4	-0.3	-0.1	0
opCcV0100001541	-1.4	1.5	0.9	0.1	0	0	0.3	0.6	-0.3	0
opCcV0100001543	0	0.3	0	0.1	-0.1	0.1	-0.1	-0.1	0.1	0.1
opCcV0100001549	-0.5	-0.5	-0.3	0	0.4	-0.2	-0.3	0.3	0.8	-0.1
opCcV0100001553	0.6	0.2	0.4	0.1	-0.1	0	0	0	0	0
opCcV0100001554	0.5	0.2	0.2	0	0	0	0.1	0	-0.1	0
opCcV0100001559	-1.6	-3.3	-0.3	0.1	0.3	0.4	0.1	0.1	0.2	-0.1

opCcV0100001561	0.5	0	0	0.1	0	-0.1	-0.1	0.1	0	-0.1
opCcV0100001563	-0.2	0	-0.5	0	0	0	-0.1	0	0.1	-0.3
opCcV0100001566	0.2	-0.6	1.3	0.4	0.4	0.1	-1	0.2	-0.2	-0.1
opCcV0100001567	0.1	-0.2	-0.2	-0.1	-0.1	0.1	-0.1	-0.2	-0.7	0
opCcV0100001569	-1.7	-0.1	-0.4	-0.2	-0.1	-0.4	-0.2	0.1	-0.3	0
opCcV0100001570	0.1	0	0	-0.1	0	0.1	-0.2	-0.3	1.2	0
opCcV0100001572	0	0	0.2	0	0.2	0.1	-0.1	0	-0.1	-0.1
opCcV0100001578	0.1	0.2	0.1	0.1	0	0.1	0.2	0	0	0
opCcV0100001583	-0.3	-0.4	0	-0.2	-0.2	0.1	-0.1	0.2	0	-0.1
opCcV0100001584	0.3	-0.1	0	0	0	0	0.1	0	0	0
opCcV0100001587	-0.5	-0.9	-0.3	0	0.1	0.3	-0.2	0.2	-0.1	0
opCcV0100001592	0.1	-0.2	0.8	0	0.1	-0.1	0.1	0.1	0.3	-0.1
opCcV0100001594	0.7	0.2	0.4	0.1	0.1	0	0	-0.1	0	-0.1
opCcV0100001598	-0.8	0.2	0.4	-0.2	-0.1	0.2	0	0.2	0.4	0.1
opCcV0100001599	-0.3	-0.3	-0.4	0.2	0.1	-0.1	-0.2	0	0.1	-0.1
opCcV0100001602	0.4	0	-0.4	-0.1	-0.2	-0.1	0.1	0.1	0	0
opCcV0100001606	1.7	0.9	0.4	0.2	0.4	0.1	-0.1	-0.1	0.1	0.1

opCcV0100001607	-2.9	-3.6	-0.2	0	0.3	-0.5	0	0.8	-1	-0.2
opCcV0100001610	0.2	-0.6	-0.9	0.1	0.2	-0.4	-0.9	0.3	-0.1	-0.2
opCcV0100001611	0.4	0	-0.2	0	0	0.1	0	0	-0.1	-0.1
opCcV0100001614	0.2	-0.5	0	-0.2	0.2	0	-0.2	0.1	-0.2	-0.2
opCcV0100001619	0.1	0.1	0	-0.1	0.7	0.2	-0.1	-0.1	0	0
opCcV0100001620	0.2	0	1.2	0	0.1	0.2	0.1	-0.1	0	0
opCcV0100001622	-2.8	-2.8	-2	-0.4	0.3	-0.2	-0.7	1	-1.3	-0.5
opCcV0100001623	0.7	0.4	0.9	0.2	0.1	0	0.1	0.2	0	0
opCcV0100001624	-0.8	-1.3	0.5	0.1	-0.5	0.1	-0.2	0.2	-0.4	0
opCcV0100001625	-0.4	-0.1	0.6	0	0	-0.2	-0.3	-0.1	-0.2	-0.1
opCcV0100001629	-0.3	-0.4	-0.1	-0.2	-0.2	0.1	-0.2	0.2	0.2	0.1
opCcV0100001630	0.4	0	0.2	-0.1	0	-0.1	-0.1	0.2	0.1	0.1
opCcV0100001631	0.2	0.5	0.8	0	-0.6	0.2	0.2	-0.2	0.2	0.2
opCcV0100001634	-0.8	-1.6	2	0.2	0.3	0.3	0.2	0.4	0.2	0.6
opCcV0100001635	0.3	0.2	0.3	0	0	0	-0.1	-0.2	0.1	0.1
opCcV0100001636	-0.2	0	-0.2	0	-0.1	0.2	-0.1	-0.1	0.1	0
opCcV0100001639	0.1	0.1	0.2	0	0	0.1	0	0	-0.2	-0.1

opCcV0100001640	-0.5	-1.9	0	0	0.2	-0.1	-0.5	0.8	0.2	0
opCcV0100001641	0.1	-0.2	0.6	0	0	-0.7	0	0	0	-0.1
opCcV0100001644	-1.8	-1.4	-1.4	-0.1	-0.2	0.2	0.1	0.1	-0.3	0
opCcV0100001647	0.1	0.1	-0.1	0	-0.2	0.1	0	0.1	0.1	0
opCcV0100001649	-0.9	-0.4	-0.3	-0.2	-0.1	0.1	-0.1	0	0	0
opCcV0100001653	0.4	0.3	0.7	0	-0.2	0.1	0	0.2	-0.1	0
opCcV0100001654	0.5	0	0.5	-0.1	-0.2	0.2	0.3	0	-0.2	0
opCcV0100001655	1.4	-1.2	-0.1	0.1	-0.2	0.1	0.6	-0.1	0.6	0.5
opCcV0100001656	0.4	-0.1	0.5	0	0	-0.1	0.1	0	0.1	-0.1
opCcV0100001657	-0.3	0	0.4	0	-0.1	0	-0.1	0.1	-0.2	-0.2
opCcV0100001658	0	0	0	0.1	0	0	-0.2	-0.2	0.2	-0.1
opCcV0100001659	0.7	0.4	1.2	0	0	0.5	0.6	0.6	-0.2	0
opCcV0100001660	0.2	0	0	0.1	0.1	-1.6	0	0.1	-0.1	-0.1
opCcV0100001664	-0.3	-0.3	0	-0.1	0.1	0.5	-0.2	0.3	-0.1	-0.1
opCcV0100001676	0.5	0.4	0.1	-0.1	-0.1	0	0	0.1	0.2	0
opCcV0100001678	-0.5	-0.4	0.7	0	0	0.5	0.4	0.1	0	0
opCcV0100001679.70	2.4	1.5	0.7	0	-0.3	0.5	-0.7	0.3	0.2	-0.2

opCcV0100001679.80	1.5	0.8	0.2	0	0.1	0.9	0.4	0.6	1	0
opCcV0100001679.90	2.7	0.5	-0.2	0.9	0.6	-0.6	0.5	1.1	0	-0.9
opCcV0100001679.A	-0.1	0.3	-0.6	0	-0.1	0	0	-0.2	0.2	0.1
opCcV0100001679	0.9	0.6	0	0.2	0.1	-0.5	-0.3	-0.5	-0.2	0
opCcV0100001680	0.4	0.3	0.7	0	-0.2	-0.1	0.2	0.1	0.1	-0.1
opCcV0100001682	0	-0.3	-0.6	0	0.1	-0.3	-0.3	0	0.2	0
opCcV0100001684	-1.7	-1.8	-0.2	0	0	-0.1	-0.3	0.5	-0.8	-0.2
opCcV0100001686	0.5	0.3	0.5	-0.1	0	0	-0.2	0	0.3	0.1
opCcV0100001688	0.8	0.5	0.6	0.2	0	0.4	0.3	-0.1	0	0
opCcV0100001689	0	-0.1	-0.6	0	0	-0.2	-0.7	0	0.1	0.1
opCcV0100001691	0	0	0.3	-0.1	0	0.1	-0.1	-0.1	-0.2	0
opCcV0100001692	0.3	-0.1	0	0.1	0.1	0	0.1	0.1	0.1	0
opCcV0100001694	1.7	-0.6	0.2	-1.2	0.6	-0.5	0.9	1	0.2	0
opCcV0100001695	-0.5	-0.4	-0.2	0	0	0	-0.1	0	-0.1	-0.1
opCcV0100001698	-0.1	0	-0.1	0	-0.1	0.2	0.2	0	0	0
opCcV0100001702	0.1	-0.3	0	0	0	0	-0.2	-0.1	-0.1	-0.1
opCcV0100001703	-0.5	-0.9	-1	-0.1	-0.1	-0.4	-0.6	-0.1	0	-0.1

opCcV0100001709	-1	0	-0.1	0.1	-0.1	-0.1	0.1	0.2	0.1	-0.1
opCcV0100001710	1.3	1	-2.3	0.4	-0.4	0.2	0	-1	0.5	0.3
opCcV0100001711	0.1	0	0.2	-0.1	-0.1	-0.1	-0.1	0	-0.1	0
opCcV0100001712	-0.7	-1.2	0	0	0.3	-0.1	-0.1	-0.1	-0.4	-0.2
opCcV0100001724	-0.2	-0.2	0	-0.1	0.1	0	0	0.1	-0.1	-0.2
opCcV0100001727	0.7	-0.2	0.1	0	0.1	-0.2	0.1	0.7	-0.1	-0.2
opCcV0100001728	0.1	-0.1	-0.2	0	-0.1	0.2	0	0	0.2	0
opCcV0100001729	0	0	-0.3	0	-0.2	0.1	0	-0.1	-0.1	-0.1
opCcV0100001730	0.3	0.1	0.1	0	-0.1	0.2	0	-0.1	0	0.1
opCcV0100001733	1.4	0.3	0.4	0.1	-0.3	0	0.1	-0.3	0.3	0.1
opCcV0100001737	0	-0.1	0.1	0	0.1	0	-0.2	0	-0.1	0
opCcV0100001738	0	0	0	0	0.1	0	0.1	0.1	0	0
opCcV0100001739	-0.4	0	-0.4	0	-0.1	0	-0.3	0	0.3	0
opCcV0100001741	0.2	0.3	-4.4	0	-0.1	-1.8	-2.1	0	0.2	0
opCcV0100001742	-0.2	-0.6	0	-0.3	0	0.2	-0.5	0.4	-0.2	0
opCcV0100001743	-0.6	-0.3	-0.8	-0.1	1.9	0.2	-0.1	-0.1	0.1	-0.1
opCcV0100001745	-4.7	-3.7	-3.6	-1	1.5	-1.1	-1.3	1.2	-1	-1.3

opCcV0100001748	-0.3	-0.2	0.1	-0.1	0.1	0	0	0.2	-0.1	-0.1
opCcV0100001750	0.2	-0.1	0.3	0	0.1	0	-0.1	-0.2	0	-0.1
opCcV0100001753	0.1	-0.1	0	-0.1	0	0.1	0.1	0.1	-0.1	-0.1
opCcV0100001754	-0.4	-4	-3.9	-1.4	1.8	0.2	-0.2	1.2	-1.6	-1.5
opCcV0100001758	-1.5	-1.8	-0.3	0	0.4	-0.1	-0.8	-0.2	-2	0.2
opCcV0100001762	-0.2	0	-0.5	0	0.1	0	-0.1	0	0.1	-0.2
opCcV0100001763	-0.9	-0.7	-4.9	-0.1	0.1	0.1	-0.2	-1.2	0.3	-0.2
opCcV0100001764	-0.5	-0.2	-1.1	0.1	-0.1	-0.3	-0.5	0.2	-0.7	-0.1
opCcV0100001765	-1.5	0	0.2	-0.1	0.5	0.1	-0.5	0.4	-0.2	0
opCcV0100001767	0.1	-0.4	1.5	0.2	0.3	0.3	0.7	0.4	-0.4	-0.1
opCcV0100001768	-1	-0.8	-1.1	-0.1	0.2	0	-0.1	0.2	0	-0.2
opCcV0100001769	-1.1	-0.6	-1.1	-0.1	0.2	-0.1	-1.1	0.1	-0.3	-0.3
opCcV0100001770	-0.4	-0.4	0.1	-0.3	-0.1	0.3	0.1	0.2	-0.2	0
opCcV0100001771	0.4	-0.3	-0.1	0.1	0	0	0	0.3	-0.1	-0.1
opCcV0100001774	-2.8	4.4	2.7	-0.1	-1.3	-0.3	0.7	0.5	1.2	-0.5
opCcV0100001776	0	1.4	1.2	-0.1	-0.7	0	0.5	0.1	-0.2	-0.1
opCcV0100001778	0.4	-3.3	-3.3	0.1	1.5	-0.1	-1.2	0.1	0	-0.2

opCcV0100001780	0.1	0.2	0.2	0	0	-0.3	-0.1	0	0.1	0.1
opCcV0100001781	0.1	0	-3.7	0	0	0	-1.7	0	0.2	0
opCcV0100001782	0.8	0.4	0.6	0.4	0	-0.3	0.1	0.7	-0.5	0
opCcV0100001783	-0.6	-0.2	-0.1	-0.2	-0.1	0.1	0	0	0.2	0
opCcV0100001784	0.7	0.5	0.9	0	0	0.1	0.3	0	-0.1	0
opCcV0100001786	-0.1	-0.2	-0.2	0.1	0	-0.1	-0.1	-0.1	0	-0.2
opCcV0100001787	-5.4	-4.9	-3.9	-1.4	1.6	-1.2	-1.7	1.4	-1.8	-1.4
opCcV0100001788	0.2	-0.1	-0.1	0	0	-1.6	-1.6	-0.1	0	0
opCcV0100001793	-0.6	-0.6	-1.7	0	0.1	-0.2	-0.5	-0.3	-0.1	-0.1
opCcV0100001795	-0.3	-0.4	0	-0.1	0.2	-0.5	-0.1	0.3	-0.3	-0.1
opCcV0100001796	0.4	0.4	0.3	-0.4	-0.3	-0.8	0.2	0.3	-0.3	-0.2
opCcV0100001797	1	1	0.2	-0.4	-0.6	1	0.8	-0.4	0.9	0.1
opCcV0100001800	-0.3	-0.1	-0.3	-0.2	-0.2	0.1	0	0	-0.1	-0.2
opCcV0100001801	-1.5	-1.1	0.1	0.1	0.3	-0.1	-0.1	-0.1	-0.4	-0.5
opCcV0100001802	-0.5	-0.5	-0.4	-0.2	-0.1	0.1	-0.4	0.1	0.2	-0.1
opCcV0100001806.50	4.1	1	-0.6	-1.3	-0.1	0.1	-0.9	0.6	0.8	1
opCcV0100001806.60	1.7	1.8	-0.3	0.5	-1.7	-1	0.8	0	0	0.4

opCcV0100001806.90	0.3	-0.1	0.5	0	0	-0.1	0.3	0.1	0.3	-0.2
opCcV0100001806.A	-0.4	0.1	-0.6	-0.1	0	0	0	0.1	0.2	-0.2
opCcV0100001806	0.6	0.1	0.4	0.1	0.1	0	0	0.1	-0.1	-0.1
opCcV0100001808	-2.4	-0.9	-1.1	-0.5	0	0.4	-0.1	0.6	-0.3	0
opCcV0100001809	0.5	0.2	0.2	-0.1	-0.2	0.2	0	-0.2	-0.1	0
opCcV0100001810	-1.7	1.5	-0.1	-0.1	-0.2	0	0.1	0.5	-1	0.3
opCcV0100001812	0.7	0.2	0.4	0.1	0	0.2	0	-0.1	0	-0.1
opCcV0100001813	-0.8	-0.1	0.7	-0.3	0	0.2	0.3	0.6	0.2	-0.1
opCcV0100001815	2.8	0.5	-0.9	-0.3	2	-0.4	-0.9	0.9	-0.8	-0.1
opCcV0100001822	-0.1	0	-0.1	0	0	0.1	0	0.1	0.1	-0.1
opCcV0100001823	-0.7	-0.5	0	0	-0.1	0	-0.2	-0.1	-0.1	-0.1
opCcV0100001824	0.7	0.2	0.7	0	0	0.1	0.1	0.1	0	0
opCcV0100001825	-0.3	-0.6	-1.5	-0.3	-0.1	0.1	0	0.1	0	0
opCcV0100001830	-0.7	-0.6	-0.4	-0.1	0.1	0.2	-0.2	0	-0.2	-0.2
opCcV0100001833	1.2	-0.8	-2.5	0.1	0.7	-0.4	-0.1	0.8	-1.4	0.3
opCcV0100001835	0	-0.1	-0.6	0	0.2	0.1	-0.2	-0.2	-0.1	-0.1
opCcV0100001836	-0.3	-0.2	1	0.2	0.1	0.4	0.2	0.1	-0.1	0.3

opCcV0100001837	0.4	0.5	0.5	0.1	0.1	-0.1	0.1	0	-0.1	-0.2
opCcV0100001838	-0.2	-0.1	0	0	0.1	-0.1	-0.1	0.1	-0.1	-0.1
opCcV0100001839	-0.6	-0.6	-0.7	-0.1	0	0.1	-0.2	0.1	-0.1	-0.1
opCcV0100001840	1.4	2.1	0.3	0.2	-0.9	0.5	0.7	-0.3	0.3	0.2
opCcV0100001842	0	-0.2	-5.8	-0.1	0.1	-0.1	-0.1	-1.9	-0.2	-0.1
opCcV0100001843	0.9	-1.4	-0.1	-0.2	-0.2	0	-0.2	0.4	-0.3	0
opCcV0100001847	-0.9	-0.6	0	-0.1	-0.1	0.3	-0.1	0	0	-0.1
opCcV0100001848	0.1	0	-0.4	-0.1	0.1	-0.2	-0.3	0.1	-0.1	-0.1
opCcV0100001850	0.5	0.4	0.1	0	-0.1	0	0.1	0	-0.1	-0.1
opCcV0100001857	-1	-1	-4.3	0.4	0.4	-0.4	-1.3	-1.4	0.3	0.1
opCcV0100001858	-0.7	-0.1	-0.8	-0.3	-0.2	0.1	-0.2	0	0.3	-0.1
opCcV0100001859	0.4	0.8	0.6	-0.2	-0.2	0.1	0	0.1	-0.1	-0.1
opCcV0100001862	-1.1	0	-0.1	0	-0.1	0.1	0	0.1	0	0
opCcV0100001863	0.3	0.2	-0.4	0	0	-1.7	-2.4	0.1	0.1	-0.1
opCcV0100001865	-0.1	-0.1	-0.5	0	0.1	0	-0.3	-0.1	-0.1	-0.1
opCcV0100001868	1.1	0.4	1.1	0.1	0.3	-0.2	0	0.1	-0.1	-0.1
opCcV0100001871	0.9	0.3	0.6	-0.2	-0.2	0.2	0.1	0	0	0

opCcV0100001874	0.4	-0.2	-1	-0.3	0.1	-0.3	-0.4	0.5	0.2	-0.3
opCcV0100001875	0.5	-0.1	0.5	0	0.1	0	0.1	0.1	0	-0.2
opCcV0100001877	1.3	0.6	0.9	0	-0.1	0.3	0.3	-0.1	0	0
opCcV0100001879	1.1	1.3	0.7	0.2	-0.5	-0.1	0	-0.8	0.7	0.2
opCcV0100001883	2	0.6	4.5	-0.4	-1	0.2	0.2	-1.1	1.1	0.2
opCcV0100001885	-0.7	-0.2	-0.1	-0.3	0.1	0	-0.1	0.1	-0.1	-0.1
opCcV0100001886	0	-0.1	-0.5	-0.1	0	0	0	0.1	-0.1	-0.2
opCcV0100001889	0.3	0	0.2	0	0.2	0	0	0.2	-0.2	-0.2
opCcV0100001890	-0.3	-0.2	-3.6	-0.1	-0.1	-0.9	-1.6	0	0	-0.1
opCcV0100001891	0.1	-0.1	0	-0.3	-0.3	-0.3	0	0	0	0
opCcV0100001893	1.2	0.7	0.7	0	-0.3	0.1	0.2	0.2	0	-0.1
opCcV0100001939	0.7	-2.6	0.1	-0.8	0.9	-0.7	-1	3.8	1.5	-1
opCcV0100001985	0.1	-0.1	-0.2	-0.1	-0.1	0.1	-0.8	-0.1	0.1	-0.1
opCcV0100001986	0.4	-0.4	0.4	0	-0.1	0.1	0.3	0	-0.1	0.1
opCcV0100001989	1.1	-0.4	1.4	-0.2	-0.4	0.2	0.2	0.3	0	0
opCcV0100001990	-1	-0.8	-0.6	0	0.6	-0.1	0.4	0	-0.6	0
opCcV0100002002	0.2	-0.3	0.4	0.1	0.2	0.3	0.3	0.1	-0.1	0

opCcV0100002009	0.7	-1.4	1.1	-0.1	-1	-0.1	0.2	1.6	1.2	-1.3
opCcV0100002077	0.1	-1.1	0.2	-0.8	-1.1	-0.2	-1.6	-0.2	0	-3.1
opCcV0100002088	0	-0.1	-0.2	0.1	0	-0.6	0	0.1	0.1	-0.1
opCcV0100002089	0.3	0.2	0	0	0	0	0	0.2	0	-0.2
opCcV0100002090	-0.9	-0.9	-0.6	-0.2	0	0	-0.4	0.3	-0.3	-0.1
opCcV0100002091	-0.3	0	-4.6	-0.1	-0.1	-0.1	-0.1	-1.4	0.2	0
opCcV0100002092.50	1.1	-0.7	0	-1.9	-1.4	0.3	-0.4	0.1	1.9	-1.5
opCcV0100002092.70	7.6	1.3	-0.1	-0.7	-1.5	0.1	0.3	0.5	0.1	-0.1
opCcV0100002092.80	2.4	1.6	-0.5	-1.1	-2	0.3	-0.4	0.6	0.6	-0.5
opCcV0100002092	2.1	0.5	-0.4	1.1	0.2	-0.3	0.3	1.2	0.3	-1.4
opCjjV010000001	0.4	0.3	0	0.1	0	0	-0.2	-0.1	0.1	0
opCjjV010000003	0.1	0.2	0.4	0	0.2	0.1	-0.2	-0.2	-0.1	-0.1
opCjjV010000004	0.4	0.2	0.5	0.2	0.2	0	-0.1	0	0.1	-0.1
opCjjV010000005	-0.3	0.2	0.2	0.1	-0.2	0.1	0.1	0	0.1	0.1
opCjjV010000006	0.2	0.1	0.2	0	0	0	0	0.2	-0.1	-0.1
opCjjV010000007	0.3	0.1	0.4	0	0	0.2	0.2	-0.1	-0.1	0
opCjjV010000008	0.2	0.1	0.1	0.1	-0.2	-0.1	0	-0.1	0.1	-0.2

opCjjV010000009	0.5	0.2	0.4	0.2	0	0.1	0.1	0.1	0	0
opCjjV010000010	0.3	0	0.2	0	0	0	-0.1	-0.1	0	0
opCjjV010000011	0	0.1	0.3	0.2	0.2	-0.2	-0.1	0	-0.1	-0.1
opCjjV010000013	0	-0.1	0	0	0	0.1	0	-0.1	0	0
opCjjV010000014	-1.5	0	-0.4	-0.1	-0.2	-0.1	-0.2	0.1	0.1	0.1
opCjjV010000016	0.4	0.4	0.3	0	-0.1	0.1	0.1	0	0.1	0
opCjjV010000017	0.4	0.6	0.4	0	-0.1	0	-0.1	0	0	0
opCjjV010000021	0.1	0.1	0.4	-0.1	0	0.1	0.1	-0.1	0.1	0
opCjjV010000022	-0.4	0	-0.2	-0.1	-0.2	0.2	-0.1	0	0.1	0
opCjjV010000023	0.3	-0.4	0.7	0	0	-0.1	-0.2	0.2	-0.5	0
opCjjV010000024	0.2	0.2	0.1	-0.1	0.1	0	-0.1	0.1	0.1	0.1
opCjjV010000025	0.5	0.2	0	0	0	0.1	0	-0.1	0	0
opCjjV010000026	0.4	0.6	0.5	0	0.1	-0.1	0.1	0	0.1	0
opCjjV010000027	0.4	0.3	0.4	0	0.1	0.1	0.1	-0.1	0.1	0.1
opCjjV010000028	-0.3	-0.3	-3.1	0	0.1	0.4	-0.1	-1.4	0.2	0
opCjjV010000030	0	0.1	-0.2	0.1	0	0.1	0	0.1	0.1	0
opCjjV010000031	-0.5	0.1	0.2	-0.1	-0.2	0.2	0	-0.1	0	0

opCjjV010000032	0.7	0.5	0.7	0.2	0	-0.1	0	0	0	0
opCjjV010000034	-0.1	0.1	-1.3	0	0	-0.6	-0.5	-0.5	0	0.1
opCjjV010000035	0.1	-0.2	-0.1	0.1	0.1	0.2	0	0	-0.3	0
opCjjV010000036	0	0.1	0.1	-0.1	-0.2	0	0	0	-0.1	0.1
opCjjV010000038	-0.2	0.3	-0.3	0.1	-0.1	0	-0.2	0	0.3	0
opCjjV010000039	-0.4	-0.1	-1.2	0.4	0.2	-0.7	0	-0.6	0.6	-0.3
opCjjV010000041	-0.8	-0.5	0.1	0.1	0	0.3	0.1	0.1	0	0.1
opCjjV010000046	0.3	0.3	0.5	0	0.1	-0.2	-0.1	0	0.1	-0.1
opCjjV010000047	0.1	0.1	0.2	0	0	0.1	0	0	-0.1	0
opCjjV010000049	0.5	0.3	0.6	0.1	0	0.1	0.1	-0.1	1.2	0
opCjjV010000050	-0.6	-0.2	-0.5	0	0.1	0.1	0	-0.1	0.1	0
opCjjV010000051	0.2	0.3	0.3	0.2	0.1	0.1	0	0	0	0
opCjjV010000052	0.7	0.6	0.8	0.1	0.1	-0.1	0.1	0.1	0.1	0
opCjjV010000055	-0.1	0	-0.2	0.1	-0.1	0.2	0	0	0	0.1
opCjjV010000058	1.1	1.1	1	0.1	-0.4	0.2	0.2	-0.1	0.2	0
opCjjV010000060	0.8	0.4	0.5	0.2	0	0.1	0.1	-0.1	-0.1	-0.1
opCjjV010000061	0.2	0.1	0.2	0	-0.1	-0.1	0	0.2	0.2	-0.1

opCjjV010000062	0.1	0	0	0	0.1	0	0.1	0.1	-0.1	0
opCjjV010000064	0.8	0.3	0.5	0.2	-0.1	0	0.1	0	0.1	0.2
opCjjV010000065	-0.5	0.1	-0.2	-0.1	-0.3	0.3	0	0	0.2	0.1
opCjjV010000066	0.4	0.4	0.2	0.1	0	-0.1	0.1	-0.1	0	-0.1
opCjjV010000067	0.9	1.4	0.9	0.1	0	-0.2	0	0	0.2	0
opCjjV010000068	0.5	0.2	0.5	0	0.1	0	0	-0.1	-0.2	-0.1
opCjjV010000070	0.1	0.4	0.2	0.1	-0.1	0	0	0	0.3	0.1
opCjjV010000071	-0.3	0	-0.4	-0.1	-0.1	0.2	-0.1	-0.1	-0.1	-0.1
opCjjV010000073	1	1.1	0.7	0.3	0	0	0.1	-0.2	0.1	0.1
opCjjV010000074	-0.4	-0.1	0.2	0	0	0.2	0	-0.1	0	0.2
opCjjV010000075	-0.1	-0.2	-0.3	-0.1	0	0	-0.3	0.1	0	-0.2
opCjjV010000076	0.4	-0.1	0.4	0	0	0.3	0	-0.1	-0.1	0.1
opCjjV010000078	0.2	0.2	0.5	0	0.1	0	0.1	-0.1	0.1	-0.1
opCjjV010000080	0.6	0.4	0.5	0	-0.2	0.1	-0.2	0	0.3	0.1
opCjjV010000081	-0.1	0.1	-0.1	-0.1	-0.1	0.2	0	0	0.2	0
opCjjV010000082	5.1	4.8	3	1.4	-2.1	0	1.7	-0.3	1.3	1.4
opCjjV010000083	0.5	0.4	0.1	0.1	-0.1	-0.1	0	0	0	-0.1

opCjjV010000084	0.3	0	0.1	0	0	0	0	-0.1	-0.1	0
opCjjV010000085	0.7	0.3	0.4	0.1	0.1	-0.1	0	0	-0.1	-0.2
opCjjV010000087	-0.6	-0.2	-0.8	-0.2	0	0.3	-0.1	0	0.2	-0.1
opCjjV010000088	0.4	0.6	0.3	0.1	-0.3	0	0.1	0.1	0.2	-0.1
opCjjV010000089	1.1	0.8	0.9	0	-0.2	0.1	0.2	0.1	0	0
opCjjV010000091	0	-1.6	0.5	0	0.7	0.1	0.1	0.2	0	0
opCjjV010000092	0.3	0.3	0.4	0.1	0	0	0.1	-0.1	0.2	0
opCjjV010000093	-0.1	0.3	-0.7	-0.1	-0.2	0	-0.1	-0.1	0.2	0.1
opCjjV010000094	0.7	0.4	0.6	0.1	0.2	0	0.1	0	0.1	-0.1
opCjjV010000095	0.2	0.3	0.3	0.1	0.1	0.1	0.1	-0.1	0.1	-0.1
opCjjV010000098	0.3	0.3	0.3	0.1	0.1	0.1	0	-0.1	0	-0.1
opCjjV010000099	0.3	0.3	0.3	0.1	0.1	-0.1	-0.1	-0.1	0.1	0
opCjjV010000100	0.9	0.4	-2.9	0.2	0	0.1	0.1	-1.1	0.2	0.1
opCjjV010000101	-0.2	0	0.1	0	0.2	0.2	-0.1	0	0.2	0.1
opCjjV010000102	0.1	0.1	0.2	-0.1	-0.3	0.2	0.1	-0.1	0.2	0.1
opCjjV010000103	-0.1	0.2	-0.1	0	-0.2	0.2	0	-0.1	0.1	0.1
opCjjV010000104	0	-0.1	0.1	0	0.1	0.1	-0.1	-0.3	0	-0.1

opCjjV010000105	-0.1	0.2	-0.2	-0.2	-0.3	0.2	-0.1	0.2	0.3	0.1
opCjjV010000108	0.3	0.6	0.1	-0.1	-0.3	0.2	0.1	-0.1	0.1	0
opCjjV010000109	-0.3	0	-0.4	-0.2	-0.4	0	-0.1	0	0	-0.1
opCjjV010000110	0	0.3	0	-0.1	-0.1	0.1	0	0.1	0.2	-0.2
opCjjV010000115	0.2	0.1	0.4	-0.1	-0.1	0.1	0	0	0	0
opCjjV010000117	-0.3	0	-0.2	-0.1	-0.1	0.3	-0.1	-0.1	0.1	0.1
opCjjV010000119	0	0	-0.5	0	0.1	-0.1	-0.1	-0.1	0.1	-0.3
opCjjV010000120	-0.2	0	-0.2	0	0	0.1	-0.1	-0.1	0.1	0
opCjjV010000121	0	-0.1	0	-0.1	0	0	0	0	0.1	-0.1
opCjjV010000122	0.2	-0.2	0.3	0	2.5	0.1	0.1	0	0	-0.1
opCjjV010000123	0.5	0.4	0.6	0.1	0.1	0	0.1	-0.1	0	0
opCjjV010000124	-0.1	-0.1	0	-0.1	-0.1	0.3	0	-0.2	0.2	0.2
opCjjV010000125	0.4	0.2	0.4	0.1	0	0	0	-0.1	0.2	0.1
opCjjV010000126	1.1	0.6	1	0	-0.1	-0.1	0.2	-0.2	0	0
opCjjV010000127	0.2	0.2	0.1	-0.1	-0.2	0	0.2	0.1	0	0
opCjjV010000128	-0.5	0.2	0.3	0	0	0	0.1	-0.1	0	0
opCjjV010000130	0.2	0.2	-0.1	-0.1	-0.2	-0.1	0	0	0.1	-0.1

opCjjV010000131	0	0.1	-0.1	0	0	0	0	-0.1	0.2	0
opCjjV010000132	0.1	0	0.1	0.2	0.1	-0.1	0.1	0.2	0	-0.3
opCjjV010000133	-0.7	-0.8	-1	-0.1	0	-0.8	-0.1	-1.2	0	-0.1
opCjjV010000134	0.7	0.3	0.2	0.1	0.1	0	0.1	-0.1	-0.1	-0.1
opCjjV010000137	0.5	0.3	0.4	0	0	0.1	0.2	0	-0.1	0
opCjjV010000138	0.3	0.3	0.2	0.2	0.2	-0.1	0.1	-0.1	-0.1	-0.1
opCjjV010000141	0.3	0.4	0.2	0.1	0	0	-0.1	0	0.2	-0.1
opCjjV010000143	0.5	0.2	0.5	0.1	0.1	0.1	0	-0.1	0	0
opCjjV010000144	0.6	0.3	0.6	0.1	0.1	-1.2	0.1	0	0	0
opCjjV010000146	0.6	0.2	0.6	0	0	0.1	0.1	-0.1	0.1	0
opCjjV010000147	0.4	0.4	0.5	0.1	2.9	0.1	0.1	0	0	0
opCjjV010000151	0.4	0.1	0.5	0	0	0	0.1	0.1	-0.1	-0.2
opCjjV010000152	0.5	0.3	0.4	0.1	-0.1	0.1	0.1	-0.1	0	-0.1
opCjjV010000154	0.2	-0.1	0	0	-0.1	0	0	0	-0.1	-0.1
opCjjV010000155	0.5	0.3	0.2	0.1	-0.1	0	0.2	-0.1	0	0.1
opCjjV010000156	-0.5	0	-0.1	0	-0.1	0.3	-0.3	0	0.2	0
opCjjV010000157	0	-0.2	0.2	-0.1	-0.2	0.1	-0.1	0.1	0	0

opCjjV010000158	-0.8	-0.5	-0.3	-0.3	-0.1	0.2	0.1	0	0.2	0
opCjjV010000159	0	-0.1	-0.1	-0.1	0	0.1	0.1	0	0	-0.1
opCjjV010000160	0.7	0.2	0.6	0	-0.1	0.1	0.1	0	0	0
opCjjV010000161	0.1	0	0.2	0	-0.2	0.3	0	0	0.1	0
opCjjV010000163	-0.4	0	-7.7	-0.1	-0.1	-2	-2.3	-2.2	0.2	0
opCjjV010000164	-0.3	0.1	-0.5	0	0.1	0	-0.1	-0.1	0.1	-0.2
opCjjV010000165	0.1	0.1	0	0	0	0.1	-0.1	0	0.1	0
opCjjV010000166	0.2	0	-0.2	0	3.7	0	-0.2	0	0	0
opCjjV010000167	0.3	0.2	0.2	0.1	-0.2	0.1	-0.1	0.1	0.3	0.1
opCjjV010000169	0	-0.2	0.4	-0.1	0.1	0.1	-0.1	0	-0.1	-0.1
opCjjV010000170	0.4	-0.4	-0.2	0.1	0.3	-0.1	-0.3	-0.1	-0.2	0
opCjjV010000171	0.1	0.1	-0.1	-0.1	0.1	0	-0.3	0	0.2	0.1
opCjjV010000172	3.6	-0.1	3.2	0.9	0.1	0.8	1	-0.1	-0.5	0.8
opCjjV010000174	0.2	0.1	0.6	0.1	0.2	0	-0.3	0	0	-0.1
opCjjV010000175	-0.6	-0.1	-0.2	-0.1	0.1	0.2	0	0.1	0.2	0.3
opCjjV010000176	0.3	0	0.1	0	-0.3	0	-0.4	-0.1	0.2	0.1
opCjjV010000177	0.1	0.2	-0.1	0	-0.1	0.1	-0.3	-0.1	0.1	-0.1

opCjjV010000179	0.2	0.4	0.2	0	0	-0.1	0.1	-0.1	0	-0.1
opCjjV010000180	-0.2	0.1	-0.5	0.1	0	0	0	0	0.1	-0.2
opCjjV010000181	1.3	1	0.8	0.1	-0.2	0.2	0.2	-0.2	0.2	0.2
opCjjV010000182	0.5	0.2	0.3	0.1	0	-0.1	0.1	0	-0.1	-0.1
opCjjV010000184	0.2	0	0.6	0.1	-0.1	0.3	0.2	-0.2	0	0
opCjjV010000187	0.4	0.2	0.6	0	0	0.1	0.1	-0.1	0	-0.3
opCjjV010000188	0.5	0.4	0.5	0	-0.2	0	-0.1	-0.1	0.1	0
opCjjV010000189	0.5	0.4	0.7	0.1	0	0	0.1	0	-0.1	-0.1
opCjjV010000192	-0.1	-0.1	-0.1	0	0.1	0	-0.4	-0.2	-0.1	-0.2
opCjjV010000193	-0.5	-0.2	-0.6	0	-0.1	0.2	-0.3	-0.1	0.2	0
opCjjV010000195	0.8	0.3	0.9	0.1	0	0.1	0.3	0	-0.1	0.1
opCjjV010000196	0.6	0.3	0.4	0.1	-0.1	0.1	0.1	0	0.1	0
opCjjV010000197	-0.1	-0.2	0	0	0	0	0	0.2	0.1	0
opCjjV010000198	-0.5	-0.1	-1	0	0	0.1	-0.3	-0.2	0.4	0
opCjjV010000201	0.7	0.3	0.3	0.1	0.2	-0.1	0.1	0.1	-0.1	-0.1
opCjjV010000205	-1.4	-0.2	-0.7	-0.1	0.2	-0.1	-0.4	0.2	-0.2	-0.1
opCjjV010000206	-0.3	-0.2	-0.3	-0.1	-0.1	0	0	0.2	0	0

opCjjV010000207	-0.5	-0.1	0.3	-0.1	0.2	0	0	0.1	-0.1	-0.2
opCjjV010000208	0.7	0.2	0.3	0.2	0.2	-0.2	0	-0.2	-0.1	-0.1
opCjV0100000001	0	0	0.3	-0.1	0	0.2	0.1	-0.2	-0.1	0.1
opCjV0100000002	0.2	-0.1	0.3	0	0.1	0.1	0.1	-0.1	-0.1	0
opCjV0100000005	-0.7	-0.2	-4	0	0	-2.2	-1.1	0.1	0.4	0
opCjV0100000006	-0.2	-0.4	0.6	-0.1	0.1	0.1	0	-0.1	-0.3	-0.1
opCjV0100000007	-0.7	-0.3	-0.2	-0.2	0.1	0.2	0	0.2	0.1	0.3
opCjV0100000010	-0.2	-0.2	-0.3	0	0.1	0.2	0	-0.1	0	0.1
opCjV0100000011	-7.1	-6.3	-2.8	-0.3	2.8	-1	-2.3	0.5	-3	-0.3
opCjV0100000012	0.5	0.3	0.8	0.1	0.1	-0.1	0.1	0.2	-0.2	-0.1
opCjV0100000013	0.2	0	0.3	-0.1	0	0.1	-0.1	0	0.1	0
opCjV0100000014	-0.6	-0.1	-0.5	0	-0.1	0	-0.1	0	0.3	0
opCjV0100000015	0.2	0.2	0.5	0	-0.1	0	-0.1	0.1	0.1	-0.1
opCjV0100000017	-1.6	0	0.2	-0.3	-0.2	0	-0.2	0	-0.2	-0.2
opCjV0100000020	0.1	0.1	0	0.1	-0.1	0.2	0.1	-0.1	0	0.1
opCjV0100000021	-0.4	0	0	-0.2	-0.2	0.1	0	0	0.1	0
opCjV0100000022	-2	-0.2	-1.4	-0.5	-0.1	-1.2	-0.9	-0.2	0	-0.5

opCjV0100000023	0.3	0.3	0.2	0	-0.1	0.2	0.2	0.1	0	0
opCjV0100000025	0.2	0	-0.1	0.1	0.1	0.2	0	-0.2	0	0
opCjV0100000026	-6.7	-5.3	-4.2	-1.8	2	-1.3	-1.9	2.2	-2.1	-1.7
opCjV0100000027	-0.8	-0.3	-6.2	0	0.1	-1.8	-2	-0.2	0.4	0
opCjV0100000028	0.2	0.5	0.8	0.1	0.1	0.3	0.1	-0.1	0	0.1
opCjV0100000029	-5.2	-4.2	-4.9	-1.4	1.6	-1.2	-2	1.6	-1.6	-1.5
opCjV0100000030	0	0	0.2	-0.1	0.1	0	-0.2	0	0.1	0
opCjV0100000036	1	0.5	0.8	0.3	0	-0.1	0.1	0	0.2	-0.1
opCjV0100000039	-0.1	-0.3	0.6	0	-0.1	0.1	0.1	0.1	-0.2	0
opCjV0100000040	-0.2	0	-0.1	-0.1	-0.2	0.1	-0.1	0.1	0	0
opCjV0100000041	-1.4	-0.2	0.9	0	0.2	0.1	0.3	0.3	-0.1	-0.2
opCjV0100000042	-0.2	0	0.1	0	-0.2	0.2	-0.1	-0.2	0.2	0
opCjV0100000043	-0.1	0.1	0	-0.1	-0.1	0.2	0	-0.2	0	0.1
opCjV0100000044	0.1	0.1	0.3	0	0	-0.1	0.1	0	0.1	0
opCjV0100000045	-0.3	0	0	-0.2	-0.3	0.2	0	0.1	0.3	-0.1
opCjV0100000047	0.4	-0.1	0.2	0	0	0.1	-0.1	-0.1	-0.1	0
opCjV0100000048	-0.4	0.2	-1	-0.1	-0.2	0.2	-0.1	-0.2	0.4	0.1

opCjV0100000049	-0.1	0	0	0	-0.1	0.3	-0.1	0	0.1	0.1
opCjV0100000050	0.1	-0.1	-0.2	0.1	0.1	0	0	-0.1	-0.2	0
opCjV0100000051	0	0.2	0.3	0.1	0.1	0.1	-0.1	-0.1	0.3	0
opCjV0100000052	-0.3	-0.4	-1.6	0	0.2	0.1	-0.1	0	0	0
opCjV0100000053	-0.4	-0.1	-0.2	-0.1	-0.2	0.2	-0.1	-0.2	0.2	0
opCjV0100000055	-0.1	-0.1	0.1	0	0.1	0	0	0	0.1	0
opCjV0100000056	-0.2	-0.2	0	-0.1	0	0.1	-0.1	0	0.1	-0.1
opCjV0100000058	0.4	0.1	0.5	0	0.1	0	0	0	0.1	0.1
opCjV0100000059	-5.3	-0.2	-4.1	0	1.7	-1.4	-1.7	0.3	-2.3	0
opCjV0100000060	0.4	0.2	0.5	0	-0.1	0.1	-0.2	0	0.1	0
opCjV0100000061	1	0.6	0.9	0.2	-0.1	0	0.2	0	0	-0.1
opCjV0100000062	-0.1	0.1	-0.1	0	0	-1.8	0	-0.1	-0.1	0
opCjV0100000064	-5.7	-0.9	-4.2	-0.2	1.9	-1.3	-1.9	0.2	-1.7	-0.2
opCjV0100000065	0.9	0.4	0.5	0.1	0	0	0.1	0	0	-0.1
opCjV0100000066	0.1	0.1	0.5	-0.1	-0.1	0.2	0.1	0	0.1	0.1
opCjV0100000067	0.4	0.4	0.2	0.1	-0.1	0	0.1	0	0.1	0
opCjV0100000068	0.2	-0.1	0	0	0.2	-0.1	0	0.1	0	-0.1

opCjV010000069	0	0.1	0.3	0	-0.1	0.1	0	0.1	-0.1	0
opCjV010000070	-0.3	0	0	-0.1	-0.1	0.3	0	-0.1	0.1	0
opCjV010000071	0	-1.2	-1.2	-2.2	4	-2.2	-2.9	2.8	-0.4	-2.3
opCjV010000072	-0.4	0.1	-0.2	-0.1	-0.1	0.2	-0.1	0.1	0.2	0
opCjV010000073	1	0.8	0.8	0.2	-0.1	0.1	0.2	-0.2	0.1	0.1
opCjV010000074	-0.1	0.1	0	0	0	0.2	-0.1	-0.1	0	0
opCjV010000075	0.5	0.6	0.7	-0.1	0	0	0.2	0	-0.2	-0.1
opCjV010000077	-0.2	0.1	-0.1	0	-0.2	0.2	-0.1	0	0	0.1
opCjV010000078	0	0.2	0.7	0	0.2	-0.1	-0.1	0	-0.4	-0.1
opCjV010000080	0.2	0.2	0.4	0.1	0.2	0.1	-0.2	0	0.1	0
opCjV010000082	-0.5	0	0.2	-0.1	0	0.2	0	-0.1	0	0.1
opCjV010000084	0.3	0.1	0.7	0.1	0.1	0	-0.4	0.1	0	-0.1
opCjV010000085	0.6	0.3	0.5	0.1	-0.1	0	0	0.1	0	-0.1
opCjV010000087	-0.4	0.2	-0.2	-0.1	-0.1	0	0	0.1	0.1	-0.1
opCjV010000088	0.2	0	0.2	0.1	0.1	0.1	0.2	-0.3	-0.1	-0.1
opCjV010000089	0.3	0	0.1	0	-0.1	0.2	0.1	0	-0.1	0
opCjV010000092	-0.3	0.2	-2	-0.1	-0.3	-1.3	-1.8	-1.1	0.1	0.1

opCjV010000093	0.7	0.5	0.2	0.2	0	0.1	0	0	0	-0.1
opCjV010000094	-0.1	0	0.1	0	0.1	0.1	-0.1	0	-0.2	0
opCjV010000095	0.3	0.2	1	0	0.2	0	0	-1	-0.1	-0.1
opCjV010000096	0.1	-0.1	0.3	0.2	0.2	0	-0.2	-0.2	0	0
opCjV010000098	0.1	0.1	0.1	0	-0.1	0.2	0.2	-0.2	0.1	0.1
opCjV010000099	-0.1	0.1	0.2	0	-2.4	0.2	0	0	0	0.1
opCjV010000100	0.1	0.1	0.4	-0.1	0.1	0.3	0.1	0	0	0.1
opCjV010000101	0.2	0.3	0.5	0.1	0	0.1	0	-0.2	0.1	0.1
opCjV010000102	0.3	0.3	0.5	0.1	-1.3	0.1	0.1	-0.1	0	0
opCjV010000103	1.1	0.6	1.4	0.2	0.3	-0.2	-0.1	-0.1	0	0.1
opCjV010000106	-0.5	-0.3	-0.5	-0.1	-0.3	0.2	-0.2	-0.1	0.1	-0.1
opCjV010000108	0.2	0.2	0	0	0	0.1	0	-0.1	-0.1	-0.1
opCjV010000109	-0.1	0.1	-0.3	-0.2	-0.3	0.2	0	0.1	0.1	0
opCjV010000110	0	0.1	0.1	0	-0.1	0.1	0	-0.1	0.2	0
opCjV010000111	0.8	0.3	0.5	0.1	0.1	0.2	0.3	0	0	0.1
opCjV010000112	0.6	0.5	-0.1	0.1	-0.1	0	0.1	0	0.1	0
opCjV010000114	-5.3	-4.1	-3.7	-1.3	1.9	-1.4	-0.1	1.6	-1.3	-1.3

opCjV0100000115	0.1	0.2	0	0.1	0	0	0	0	0.2	0
opCjV0100000116	0.1	0.4	0	0	-0.1	0.1	-0.1	0	0.3	0
opCjV0100000117	-0.2	0.1	-0.5	0.1	0.1	0	-0.1	0	0.1	-0.2
opCjV0100000118	0.4	0.1	0.4	0.1	0.2	0	-0.1	-0.1	0.1	-0.1
opCjV0100000120	0	-0.1	-0.5	0	-1.6	0.2	-0.2	-0.2	0	0
opCjV0100000121	0.1	0.4	0.3	0	0	-0.1	0	0	0	-0.1
opCjV0100000122	0.3	0.2	0.2	0.1	0	0	-0.3	0	0	0.1
opCjV0100000124	0.4	0.2	0.6	0	0.3	-0.1	0	-0.1	-0.2	-0.1
opCjV0100000125	0.7	0.4	0.8	0.2	0.2	-0.1	-0.2	-0.3	0	0.2
opCjV0100000126	0.2	0.3	0.2	0.2	-0.7	0.3	0	-0.2	0.3	0.3
opCjV0100000127	0.5	0.3	0.7	0	0.2	0	0	-0.1	-0.2	0
opCjV0100000130	0.6	0.2	0.7	-0.1	-0.1	0.3	0.1	0	0.2	-0.1
opCjV0100000131	-0.2	0.1	-0.1	0	-0.2	0.2	0.1	-0.1	0.1	0
opCjV0100000132	0.3	0	0.2	0.1	0.1	0.1	0.1	-0.1	-0.1	0
opCjV0100000133	0.2	0	0.2	0	-0.1	0.1	0	0	0.1	0
opCjV0100000134	0.2	0.2	0.2	0	0	-0.1	0.1	0.1	0	0
opCjV0100000135	0.1	0.1	0.2	-0.1	-0.1	0.2	0	0	0	0

opCjV0100000136	0.2	0.2	0	0	-0.2	0.1	0.1	0	0.1	0.1
opCjV0100000137	-0.4	0.1	-0.1	-0.1	0	0	-0.4	0.2	-0.1	-0.1
opCjV0100000138	-0.5	0.2	-0.3	0	-0.1	0	0.1	-0.3	-0.1	0
opCjV0100000140	-1.9	-4.4	-0.3	-1.3	1.9	0	-0.9	1.6	-1.5	-1.5
opCjV0100000141	0	0.1	0.1	-0.1	-0.1	0.1	0.1	-0.1	0.1	0.1
opCjV0100000142	0.6	0.2	0.7	0.1	0.1	0.1	0	0	0	0.1
opCjV0100000143	0.1	-0.1	0	0.1	-0.9	0	-0.3	-0.1	0.2	0
opCjV0100000144	0.7	-0.1	1	0	0	0.2	0	0.1	0	0
opCjV0100000145	-0.1	-0.2	0.1	0	0.1	0.1	0.1	-0.1	-0.1	0.1
opCjV0100000146	0	0	0	0	-1.2	0.1	0.1	-0.1	0	0
opCjV0100000147	0.2	0.2	0.5	0.1	0.1	0.2	0	0	0.1	0
opCjV0100000150	-0.4	-0.2	-0.6	0.1	0.1	0.1	0	-0.2	0.2	0.1
opCjV0100000151	-0.2	0.1	0	0.1	0.1	0.2	0.1	0	0.2	0.1
opCjV0100000152	0.1	0.3	0.5	0.2	-0.8	0	0	0	0	-0.1
opCjV0100000153	-0.4	-0.8	-0.4	0	0.1	0	-0.1	0	-0.1	-0.1
opCjV0100000154	0.4	0.1	0.2	0	-0.1	0.2	0	-0.1	0	0.1
opCjV0100000155	-0.2	0.1	-0.1	-0.1	-0.2	0.1	0	-0.1	0.2	0.1

opCjV0100000156	-0.2	0	-0.4	-0.1	-0.3	0.2	-0.1	0	0	-0.1
opCjV0100000158	-0.6	-2.2	-2.1	-0.7	1.2	-0.5	-0.1	1.1	-0.7	-0.1
opCjV0100000159	-0.1	0.1	0.1	-0.1	-0.4	0.3	0	-0.1	0	0.1
opCjV0100000160	0	0.3	0.3	-0.1	-0.1	0.2	0.1	0.1	0.3	0
opCjV0100000161	0.8	0	0	0.2	0.1	-0.1	0	-0.1	-0.1	0
opCjV0100000164	-2.1	-1.5	-0.9	-0.4	0.6	-0.8	-1.1	-0.1	-0.5	-0.4
opCjV0100000165	-0.2	-4.7	-2.1	-1.5	1.3	-1.2	-0.1	5.5	-2	-1.4
opCjV0100000166	-1	-0.4	-2.7	0.1	-0.3	0.4	0.1	-0.3	0.1	0.3
opCjV0100000169	0	0	-1.6	0	-0.1	-0.9	-2.7	-0.1	0.1	0.1
opCjV0100000170	0.6	0.4	-0.1	0.3	-1.8	-0.1	0	-0.4	0	0
opCjV0100000172	0.4	0.2	0.4	0.1	0.1	0.1	0	-0.1	0	0.1
opCjV0100000173	0.3	0.3	0	-1.4	-0.4	-1.4	-1.7	0.9	0	-1.5
opCjV0100000174	-1.3	-0.8	0.1	-0.3	0.2	0	-0.1	0.2	-0.1	-0.3
opCjV0100000175	0.6	0.5	0.1	0.1	0.2	0	-0.2	-0.4	0	0
opCjV0100000176	0.3	0.4	0.3	0.1	0.2	0	-0.1	0	0.1	0
opCjV0100000178	0.3	0	0.3	-0.1	-0.1	0	0.2	0	-0.1	0
opCjV0100000179	0.7	0.3	0.5	0.1	0.2	0	0	0	-0.1	-0.1

opCjV0100000180	0	0.1	-4.5	0	-0.1	-1.3	-1.4	0.2	0	-0.1
opCjV0100000181	-0.6	-0.5	0	-0.1	0.2	0	0.1	0.1	-0.2	-0.2
opCjV0100000183	0.4	0.3	0.6	0	-0.1	0.1	0.2	0	0.1	0
opCjV0100000187	-0.4	0	-0.4	0	-0.2	0.3	0	-0.1	0.2	0
opCjV0100000188	-4.8	-4	-3.7	-1.3	2.2	-1.5	-1.6	1.5	-1.6	-1.7
opCjV0100000189	0.4	0.1	0.5	0.2	0.1	0.1	0.1	-0.2	0	0
opCjV0100000190	-2.5	-2.2	-0.1	-1.7	2.3	-2.5	-0.2	2.8	0	-1.8
opCjV0100000191	0.4	0.4	0.4	0	-0.1	0.1	0.1	0.1	0.1	0
opCjV0100000192	-4	-3.5	-3.7	-1	1.6	-1.2	-1.3	1.3	-1.3	-1.2
opCjV0100000193	0.3	0.1	-0.1	0.1	0.1	-1.8	0.1	0	-0.1	0
opCjV0100000194	0.6	0.5	0.5	0.1	-0.1	0	0	0	0	-0.1
opCjV0100000195	0.1	0.2	0.4	0.2	0	0.3	0.1	-0.2	0	0.2
opCjV0100000196	-0.1	-0.2	-2	0.1	0.1	-1.5	-1.8	-1	0.2	0
opCjV0100000197	0	0.2	0.2	0.1	0.1	0	0	0	0	0
opCjV0100000198	0.4	0.5	0.4	0	0	0.1	0.1	0	0.1	0
opCjV0100000199	0.3	0.5	0.4	0	0	-0.1	0.2	-0.2	0	0
opCjV0100000200	0	-0.1	0.2	0	0	0	0.2	0.1	-0.2	-0.1

opCjV0100000201	0.1	0	0.3	0	0.2	-0.1	0.2	0.2	0	-0.1
opCjV0100000202	0.1	-0.1	0	0	0.1	0.1	0	0	0	-0.2
opCjV0100000203	-0.7	-0.1	-0.2	-0.3	-0.3	0.2	0	0.1	0	0
opCjV0100000205	-0.3	-0.5	-0.3	0.1	0.1	0.1	-0.2	0.1	-0.1	-0.1
opCjV0100000206	0	0.2	0.5	-0.1	-0.2	0.2	0	-0.1	-0.1	0
opCjV0100000207	-0.9	-0.3	-2.2	-0.1	-0.2	-0.7	-1.3	0	0.3	0
opCjV0100000208	-1.2	-0.6	-0.2	-0.1	-0.2	0.3	-0.1	0	0.2	0.1
opCjV0100000211	-0.3	-0.2	-0.8	-2.4	2.9	-2.4	-2.9	2.1	0.2	-1.9
opCjV0100000212	0.8	0.6	0.9	0	-0.1	-0.1	0.1	0	0	-0.1
opCjV0100000213	0.2	0.2	0.5	-0.1	-0.1	0.2	0	-0.1	0	0
opCjV0100000214	0.5	0.3	0.5	0	-0.1	0	0.1	-0.2	0	0
opCjV0100000215	0.4	0.4	-5.4	-1.8	5.3	-2.7	-2.9	5.5	0.1	-1.7
opCjV0100000216	-3.7	-3	-3.1	-0.9	1.3	-0.9	-1.5	2.4	-1.1	-1.3
opCjV0100000218	-0.1	0.1	0.2	0	-0.1	0.2	0	-0.1	0	0
opCjV0100000219	-0.1	0.1	0	0	-0.1	0.2	0	0	0.1	0
opCjV0100000220	-3.9	-3.5	-1.7	-1	1.5	0	-1.2	1.5	-0.1	-1.2
opCjV0100000222	0.1	0.4	0.2	0	-0.2	0.1	0	-0.2	-0.1	0

opCjV0100000223	0.2	0.1	-0.3	0.1	0.1	-1.5	0	-0.1	-0.1	0
opCjV0100000224	0.3	0.2	0.5	0.2	0	0.1	0.3	0.1	-0.3	-0.1
opCjV0100000225	0.4	0	0	0.1	0.2	-0.1	0.1	0.1	-0.1	-0.1
opCjV0100000228	0.6	0.3	-0.1	0.3	0.3	-0.1	-0.3	0.1	-0.2	-0.1
opCjV0100000232	-0.7	-0.4	-0.7	-0.1	-0.1	0.3	-0.1	0.1	0.1	0
opCjV0100000233	-0.1	0	0.2	0	-0.2	0.1	-0.1	0.1	-0.1	0
opCjV0100000235	-0.5	-2.4	-4.8	-1.7	2.2	0.1	-0.1	1.8	-3.1	-1.5
opCjV0100000238	-0.1	-0.1	0	0	-0.2	0.4	-0.1	0	0.2	0
opCjV0100000239	-0.5	-0.3	0	-0.1	0	0.1	-0.1	0	0.1	0
opCjV0100000240	-0.7	-0.4	0.1	0.1	0.1	0	0	0	-0.2	-0.1
opCjV0100000241	0.1	0.3	0.2	-0.1	0	0	0	0	-0.1	0
opCjV0100000242	-0.1	0	0	-0.1	-0.1	0.2	0	-0.1	0.1	0
opCjV0100000243	-0.8	-0.2	-0.5	0	-0.1	0.3	-0.1	-0.1	0.1	0
opCjV0100000244	0.3	0.3	0.3	0.1	0.1	0.1	0.1	-0.1	0	0
opCjV0100000245	-0.1	0.1	0.1	0	-0.1	0.2	-0.1	0	0.2	0.1
opCjV0100000246	0.5	0.5	0.3	0.2	0	0.2	0.1	-0.2	-0.1	0.1
opCjV0100000250	0.1	0	0.2	0	-0.1	0.1	0.2	0.1	0.2	-0.1

opCjV0100000252	0.1	0	-1.4	0	0	0.1	0	0.1	0.1	0
opCjV0100000253	0.7	0.5	0.6	0.1	0.1	0	0.2	0.2	0	-0.1
opCjV0100000255	0.1	0.1	0.1	0.1	0.2	0.1	0	0	-0.2	-0.2
opCjV0100000256	-0.1	0.1	0.4	0	0	0.1	0.2	0	0	-0.1
opCjV0100000257	-0.3	-0.1	-2.1	0	-0.2	-1	0.1	0	-0.1	0
opCjV0100000259	0.1	0	0.2	0	0.1	-0.1	0.1	0.1	-0.2	-0.1
opCjV0100000260	0.5	0.2	0.4	0.1	-0.1	0	0.1	-0.5	0	0
opCjV0100000262	0.5	0.5	0.3	0	-0.1	0	-0.1	0.1	0.2	0.1
opCjV0100000263	0.2	0.4	0.2	0	-0.2	0.1	-0.2	-0.1	0.1	0.1
opCjV0100000264	0.2	0.3	0.4	0.1	0	0.2	0.1	-0.1	0.1	0
opCjV0100000265	0.2	0.5	0.1	0	-0.1	-2.1	-1.7	-0.1	0.1	0
opCjV0100000266	0.4	0.3	0.5	0.1	0.1	0.2	0.2	0	0.1	0
opCjV0100000267	0.5	0.2	0.5	0	0.1	-0.1	0.1	0.1	0	0
opCjV0100000268	0.4	0.2	0.5	0	0.1	0	0	-0.1	0	0
opCjV0100000269	0.4	-1	-0.7	-1.9	3.3	-0.7	-2.8	2.8	-0.1	-2.1
opCjV0100000270	-0.5	0	-0.4	0	-0.2	0.2	0	0	0.2	0.1
opCjV0100000271	-0.4	-0.1	0.6	0	0	0.2	0.2	0.2	-0.2	0

opCjV0100000272	-0.2	-0.1	-0.2	0	-0.1	0.2	0	0.1	0	0
opCjV0100000273	0.2	0.3	0.1	0.1	0.1	0	0	0.1	0	-0.1
opCjV0100000274	0.6	0.2	0.4	0	0	0	0.2	0	-0.1	-0.1
opCjV0100000276	-0.4	-0.2	-0.4	0.1	-0.1	0.1	-0.2	0.2	0.2	0
opCjV0100000279	-5.3	-4.4	-6.1	-1.7	2.4	-2.2	-1.8	1.7	-1.6	-1.6
opCjV0100000280	0.3	0.6	0.5	0.1	0.1	0	0	0.1	-0.1	-0.2
opCjV0100000281	0.3	0.2	0.3	-0.1	-0.1	-0.1	0.1	0.1	0	0
opCjV0100000282	-0.2	0	0.1	0	0	0.2	0	0	-0.1	-0.1
opCjV0100000283	0.3	0.1	0.5	0	0.1	0	0.1	0.1	-0.1	-0.1
opCjV0100000284	0.2	0.1	0.5	-0.1	-0.1	0.1	0.1	0	0	0.1
opCjV0100000285	-0.8	0	-0.3	0	-0.2	0.2	0.1	0	0.2	0.1
opCjV0100000286	-0.2	0.1	-0.4	0.1	0.1	0	-0.1	0	0.2	-0.1
opCjV0100000287	-0.1	-0.1	-0.3	0	-0.1	0.2	0	-0.1	0	0
opCjV0100000288	0.2	0.4	0.2	0.1	0	0	0.1	-0.1	0.1	-0.1
opCjV0100000290	-0.3	-0.2	0.1	0	0	0.2	0	-0.1	0	0
opCjV0100000292	0.6	0.2	0.3	0.2	0.3	0	-0.1	-0.1	-0.1	0
opCjV0100000293	0.1	0.2	0.2	-0.3	0	0	0	0.1	-0.1	-0.2

opCjV0100000295	0.6	0.1	0.4	0	-0.1	0.2	0.1	0	0	-0.1
opCjV0100000296	0.8	0.3	0.4	0.2	0	0	0.2	0	-0.1	-0.1
opCjV0100000298	0.2	0	0.2	0	0	0	0	0	0	-0.1
opCjV0100000299	-1.2	-0.6	-1.4	-0.1	1.3	-0.6	-0.4	0.6	-0.3	-0.2
opCjV0100000300	-0.1	-0.2	-5	0	0	-1.6	-0.1	0.1	0.2	0
opCjV0100000304	-1.5	-1.2	-0.9	-0.2	0.3	0	-0.4	0.1	-0.1	-0.2
opCjV0100000305	-0.7	-0.1	-5	-0.2	-0.3	-1.5	-1.1	-1.5	0.3	0.1
opCjV0100000306	0.1	-0.1	0.6	0	-0.1	0.1	-0.1	0.1	0.1	0
opCjV0100000307	0	0.4	0.1	0	-0.2	0.1	0	0	0.2	0.1
opCjV0100000308	-0.6	0.2	-0.2	0.1	-0.1	0.1	0.1	0.1	0.2	0.1
opCjV0100000309	0.5	0.6	0.5	0	-0.1	-0.1	0.1	0.1	0.1	-0.2
opCjV0100000310	0.3	0.2	0.1	0	-0.1	0.1	0.1	0	0	-0.1
opCjV0100000311	0.2	0	0.3	0	-0.1	0	0.1	-0.2	0	0.1
opCjV0100000312	0.4	0.5	0.4	-0.2	-0.1	0	-0.1	0.1	0.3	0
opCjV0100000313	0.5	-0.1	0.4	0.1	0.1	0	0	-0.1	-0.1	0
opCjV0100000317	0.4	0.3	0.4	0	0	0.1	0.2	0	-0.1	0
opCjV0100000320	-0.6	0.2	0	0	0	0	-0.1	0.1	0	-0.2

opCjV010000321	-0.4	0	-0.1	-0.1	-0.2	0.2	0	0.1	0	0
opCjV010000322	1	0.5	0.9	0	0	-0.1	0.2	-0.1	-0.1	-0.1
opCjV010000323	0.7	0.2	0.3	0	0	0.1	0.1	-0.1	0	0
opCjV010000324	0.4	0	0.5	0	-0.1	-0.1	-0.1	0	0	0
opCjV010000326	-0.4	-0.2	-0.4	0.1	-0.1	0.1	-0.2	-0.1	0	-0.1
opCjV010000329	0.2	0.1	-3.9	0	-0.1	-1.4	-1.5	-1.5	0	0
opCjV010000330	0.2	0.4	0.4	0	-0.1	0.2	0.1	-0.1	0.3	-0.1
opCjV010000331	0.2	0	-0.2	0	0	-0.1	0	0	0.1	0
opCjV010000332	-1.9	-1.1	-4.7	-0.3	0.3	-0.7	-0.2	-1.1	0.1	-0.3
opCjV010000336	-0.7	-0.4	-0.3	-0.2	-0.1	0.2	0	-0.1	0.2	0.1
opCjV010000337	0.1	0.2	-0.4	-0.2	-0.1	0.1	-0.1	0	-0.4	-0.1
opCjV010000338	-0.8	-0.1	0	-0.1	-0.2	0.2	-0.2	0	0.4	0
opCjV010000339	-0.5	-0.1	-0.3	-0.1	-0.1	0	-0.3	0	0.1	-0.1
opCjV010000340	0	0.2	0	-0.1	-0.1	0	-0.1	-0.1	0.1	0
opCjV010000341	-0.1	-0.1	0.1	0	-0.1	0.1	-0.1	0.1	-0.1	-0.1
opCjV010000342	0.1	0	0.1	0	0.1	0	-0.1	0	-0.2	-0.2
opCjV010000345	-2.1	-1.1	-0.2	0.1	0.1	0	-0.3	-0.1	-0.2	-0.2

opCjV0100000346	0.2	0.1	0.2	0	0.1	0	0	0	0	-0.1
opCjV0100000348	0.2	0.1	0	0.2	0	0	0	0.1	0.2	-0.1
opCjV0100000349	-0.5	-0.1	-0.4	-0.4	-0.1	0.1	-0.1	0	-0.1	-0.2
opCjV0100000350	-0.5	0	0	-0.1	-0.2	0.1	0	0.1	0.1	-0.1
opCjV0100000351	-1.3	-0.1	-0.6	0.1	0.2	-0.3	-0.3	-0.2	-0.1	-0.2
opCjV0100000354	0.8	0.6	0.7	0.1	0	0	0.2	0.1	0	-0.1
opCjV0100000355	-1.4	-0.1	-0.7	-0.2	0	0.2	-0.3	0	0.1	-0.1
opCjV0100000356	0.6	0.8	1	0.1	-0.1	-0.2	0.2	-0.1	0.2	-0.1
opCjV0100000357	-5	-4.6	-1.6	-1.3	2.2	0	-0.8	3.2	-2	-1.8
opCjV0100000358	0.1	0.2	0.3	0	0	0.1	-0.1	-0.1	0	-0.1
opCjV0100000361	-0.5	0	-1.1	-0.1	-0.1	0.2	-0.1	-0.1	-0.1	0.1
opCjV0100000363	0	0.1	-0.1	0	0	0.1	-0.1	-0.1	0.1	-0.1
opCjV0100000364	0.8	-0.6	0.2	-2.3	2.8	-2.6	-2.7	2.2	-0.2	-2.2
opCjV0100000365	-0.6	-1	-1.9	0	0.2	-1.9	-0.8	0.2	-0.1	-0.1
opCjV0100000366	0.6	0.2	0.1	0	-0.1	-0.2	0	0.2	0	-0.2
opCjV0100000367	0.3	0.2	0.8	0	0	0.1	0.1	0.1	0	-0.1
opCjV0100000370	0.2	0.1	0.4	0	0.1	0	0.1	0.1	-0.1	-0.1

opCjV0100000371	1	0.4	0.8	0.1	0	-0.2	0.2	0.2	-0.1	-0.1
opCjV0100000372	0.7	0.7	0.6	0.3	0	0.1	0.1	-0.1	0.1	-0.1
opCjV0100000373	0.3	0.1	-5.4	0	0	-2.1	-1.4	-0.2	0	-0.1
opCjV0100000378	0.2	0	0.2	0	0.1	0.2	0.1	-0.3	0.2	0.1
opCjV0100000381	0.3	0.1	0.2	0.1	0.1	-0.2	0.1	0	-0.1	0
opCjV0100000382	1.1	0.5	1	0	-0.2	0	0.2	0	-0.7	0.1
opCjV0100000383	-0.1	0.3	0.3	-0.1	0	0.1	-0.4	-0.1	0.2	0
opCjV0100000384	-0.6	-0.5	-2.8	-0.1	0.1	-1.4	-1.8	0	0.2	0
opCjV0100000385	-0.2	0.1	0	0	0.1	0	0	0	0.2	0
opCjV0100000386	-0.1	0.1	0	-0.1	-0.1	0.2	0	0.1	0	0
opCjV0100000390	0.1	0.5	0.4	0.1	0	-0.1	-0.1	-0.1	0	-0.1
opCjV0100000391	-0.2	0.3	-1.1	-0.6	-0.1	-0.8	-1.2	-0.4	-0.8	0
opCjV0100000392	-0.1	0.1	0	0.1	-0.1	0	-0.1	-0.1	0	-0.1
opCjV0100000393	-0.2	0.2	-0.3	0.1	0	0.1	0	-0.2	0.2	0
opCjV0100000394	0.5	0.3	0.4	0.1	0	0	0.1	0	0	-0.1
opCjV0100000395	-0.4	0	0.7	0	0.1	0.2	0	0	-0.3	0
opCjV0100000396	-4.9	-4.1	-3.3	-1.2	1.8	-0.9	-2.5	1.5	-1.5	-1.4

opCjV0100000397	-0.7	0	-0.2	-0.1	-0.4	0.2	-0.1	0	0.2	0
opCjV0100000398	-1.3	-0.2	-0.5	-0.1	-0.2	0	0	-0.3	0.1	0.1
opCjV0100000399	-3.2	-2.6	-2.2	-0.9	1.6	-0.8	-1	1.1	-1	-1
opCjV0100000401	0	-0.2	0.4	-0.2	0.1	0.1	0	-0.2	0.1	0
opCjV0100000402	0.7	0.5	0.5	0.2	0.1	0	0.1	0	0	0
opCjV0100000403	-0.1	0.2	0.3	-0.2	-0.1	0	-0.1	-0.1	-0.6	0
opCjV0100000404	-0.3	0	-1	0	0	0.1	-1	-0.6	0.3	0.1
opCjV0100000406	-0.3	-0.6	-0.5	-1.1	0.1	0	-0.1	0.4	0.2	-2
opCjV0100000407	0.3	0.1	0.4	0	0	0.1	0	-0.4	-0.1	-0.1
opCjV0100000408	-0.5	0	-0.5	-0.1	-0.1	0.2	-0.1	-0.1	0.2	0.1
opCjV0100000409	-0.1	0.1	0.1	0	0	0.1	-0.1	-0.5	0	0.1
opCjV0100000410	0.3	0.2	0	0	0.1	0	-0.1	-0.2	0	0
opCjV0100000412	0.2	0.3	0.4	-0.1	-0.2	0.1	0.1	0.1	0	0
opCjV0100000413	0.6	0.3	0.5	0.1	-0.1	0	0	0	0.1	0.1
opCjV0100000414	-4.1	-3.2	-6.5	-1	1.6	-1.2	-1.3	0.8	-1.3	-1.3
opCjV0100000416	-0.6	0	0.3	0	0	-0.2	0.4	0.3	0.2	0
opCjV0100000417	0.2	-0.1	-0.4	-2	3.3	-2	-2.5	3.2	-0.2	-0.1

opCjV0100000418	0.1	0.1	0.1	-0.1	0	0.1	0.1	0.1	-0.1	-0.1
opCjV0100000419	0.2	0	0.4	0	-0.2	0.2	0	-0.1	0	0
opCjV0100000423	-4	-2.7	-0.3	-0.7	1.2	0	-0.2	1.1	-0.7	-0.8
opCjV0100000425	0.3	0.3	0.5	0.1	0.3	0	0.1	0.1	-0.1	-0.1
opCjV0100000426	-0.1	-0.5	0	-1.6	3.3	-1.6	-1.2	3.2	0	-3.2
opCjV0100000428	0.1	-0.1	0.3	0	0.1	0.2	0.1	0	-0.1	0
opCjV0100000430	0.6	0.4	0.7	0	0	0.1	0.1	-0.1	-0.1	0
opCjV0100000433	0.3	0.1	0.4	0	0	0	-0.1	-0.1	0	0
opCjV0100000434	0.5	0.2	0.4	0.3	0.3	0	-0.1	0.1	-0.3	0
opCjV0100000435	0.5	0.6	0.4	0	-0.3	-0.1	0.1	-0.1	0	-0.1
opCjV0100000436	0.6	0.3	0.5	-0.1	-0.2	0	-0.1	0	0.1	0
opCjV0100000437	0.8	0.6	0.9	0	0.1	0.1	0.2	0	0	0
opCjV0100000438	0.6	0.1	0.6	0	-0.1	0	0.1	0.2	0.1	0
opCjV0100000439	-0.4	0.2	-0.2	-0.1	-0.3	0.2	0	0	0.2	0.1
opCjV0100000442	0.1	-0.1	-0.1	0.1	0	0.1	-0.1	-0.1	0	0
opCjV0100000444	-0.3	-0.1	-0.5	-0.1	-0.4	-0.8	-1.3	0.1	0.3	0
opCjV0100000445	0.5	0.3	0.3	0.1	0	0	-0.1	0	0	-0.1

opCjV0100000448	-0.1	-0.2	0.2	0	0.1	0.1	0	-0.1	0.1	0.1
opCjV0100000449	-0.2	0	0	0	0.2	0.1	-0.1	0	0	0
opCjV0100000450	-0.2	0	0.5	-0.3	0	-0.1	-0.1	0	0.1	-0.1
opCjV0100000451	0.3	0.5	0.3	0	-0.1	0.1	0	-0.1	0	0
opCjV0100000454	0.3	0.1	0.6	0.1	0.2	0	0.1	0	-0.2	0
opCjV0100000455	0	0	0.1	-0.1	-0.1	0.1	0	-0.1	0.1	0.1
opCjV0100000456	-0.2	-1.5	0.3	-0.4	0.5	-0.1	-0.1	0.2	0	-0.4
opCjV0100000457	0.9	0.8	0.6	0.2	0.1	0	0.3	-0.1	-0.1	0
opCjV0100000458	0.4	-0.3	0.3	0	0.1	0.2	-0.1	0	-0.2	0
opCjV0100000459	0.4	0.4	0.3	0.1	-0.1	0	0.2	0	0.1	0
opCjV0100000460	-0.2	0.1	-0.4	0	-0.2	0.1	-0.2	-0.1	0.1	0
opCjV0100000461	-0.5	0	-0.4	-0.1	0	0.2	0	0.2	0.1	0.1
opCjV0100000462	0.3	0.4	0.5	0	-0.2	0	0.1	0.1	0	0
opCjV0100000464	0.6	0.2	-0.1	0.1	0	0.1	-0.2	-0.1	0	0
opCjV0100000467	-5.2	-3.7	-3.7	-1.4	2.1	-1.5	-1.4	1.7	-1.5	-1.4
opCjV0100000468	0.2	0.1	0.4	0	-0.1	0	-0.2	0.1	0	-0.1
opCjV0100000470	-0.5	-0.2	-0.5	0	-0.1	0.2	-0.2	0	0.2	0

opCjV0100000471	-0.4	0	0	0	0	0	-0.2	0	0.1	0.1
opCjV0100000472	0.3	0.2	0.3	0.1	-0.3	0	0	0	0	0
opCjV0100000474	0.5	0.4	0.8	0.1	0.1	0	0	-0.1	0	0
opCjV0100000475	0.1	0	0.2	0.1	0.1	0.1	-0.1	-0.1	-0.1	-0.1
opCjV0100000476	-0.2	0.1	-0.1	0.1	0.1	0.2	0	0	0.1	0
opCjV0100000478	0	0.1	0.1	0	0.1	0.2	0	0	0.2	0
opCjV0100000479	0.8	0.4	1.1	0.1	0	0.1	0.2	-0.1	0	0
opCjV0100000480	-0.1	0.1	0.1	0	0.1	0.2	0	0	0	0.1
opCjV0100000481	-0.1	0.2	0	0.1	0	0.1	0	0	0.1	0
opCjV0100000483	-0.3	-5.2	-0.3	-1.4	3	0	-0.1	2	-1.9	-1.7
opCjV0100000484	-0.1	-0.2	0	0	0.2	-0.1	0	-0.2	-0.1	0
opCjV0100000485	-0.2	0.2	-0.1	0	-0.2	0.2	-0.1	0	0.3	0
opCjV0100000486	0.7	0.3	0.5	0	0	0.1	0.2	0	-0.1	-0.1
opCjV0100000487	0	-0.1	0.4	0	0	0.1	0	0	0.1	-0.1
opCjV0100000488	0.2	0	-0.4	-0.1	0.1	-0.2	-0.5	0.2	0.1	0
opCjV0100000489	0.1	0	0	0.1	-0.1	-0.1	0	-0.1	0.1	-0.1
opCjV0100000490	0.2	0.1	0.5	0	0	0.1	0	0	0	0

opCjV010000492	0	0	-0.2	-0.1	-0.2	0.1	-0.2	-0.2	0.1	0.1
opCjV010000493	0.3	0.1	0.1	0.1	0.1	0	0.1	0	-0.1	-0.1
opCjV010000494	-0.9	-0.5	-1.1	0	0	0.4	-0.1	-0.1	0.5	0.1
opCjV010000495	-0.1	-0.2	-0.1	-0.1	0.1	0.2	-0.1	0	0	-0.1
opCjV010000496	0.1	0.1	0.2	0	0.4	0.2	0.1	0	-0.1	-0.1
opCjV010000497	0.1	0	0.8	-0.1	-0.1	0.3	0	0	0.1	0
opCjV010000500	0.3	0.1	0.3	0	0.3	-0.1	-0.1	0.1	-0.1	-0.1
opCjV010000504	-0.6	-0.5	-0.3	-0.1	0.2	-0.4	-0.3	0.1	-0.2	-0.4
opCjV010000505	-0.1	-0.3	0.3	0.1	2.9	0.1	0	0	-0.1	-0.1
opCjV010000507	0.3	0.3	0.3	0	-0.2	0.2	-0.1	-0.2	0	0.1
opCjV010000509	-0.8	0.4	0.2	-0.1	-0.1	-0.1	0	0.2	0	-0.1
opCjV010000510	-5.3	-4.7	-4.8	-1.5	2.6	-1.4	-2.1	2	-1.6	-1.5
opCjV010000512	1	1	0.6	0.2	0.1	-0.1	0.1	-0.1	-0.2	-0.2
opCjV010000513	0.4	0.3	0.1	0	0.1	0.1	0	0	0	0
opCjV010000514	1	0.5	0.7	0.1	0	0	0.3	0.1	-0.1	0
opCjV010000517	0.3	0.2	-1.3	-0.1	-0.1	-0.3	-0.8	0	0	0
opCjV010000519	0.1	-0.1	0.2	0.1	0.1	0.1	0	-0.1	-0.1	0

opCjV0100000520	0.3	0.2	0.3	0.2	0.2	0.1	0.1	-0.1	0	0.1
opCjV0100000521	-0.1	0	-0.1	0.2	-0.4	0.1	-0.1	0.1	0.1	-0.1
opCjV0100000522	0.9	0.5	1.3	0.1	0.1	-0.1	0.2	0.1	-0.2	-0.1
opCjV0100000523	-0.2	-0.1	-0.3	0.1	0	0	-0.1	-0.2	0.2	0.1
opCjV0100000524	-0.3	0.1	0.1	0	-1	0.3	0.2	-0.1	0.2	0
opCjV0100000525	-0.2	0.1	-0.3	-0.1	-0.1	0	-0.1	-0.1	0.3	0
opCjV0100000527	0.2	0.2	0.2	0.1	0.1	-0.1	-0.1	-0.1	0.1	0
opCjV0100000528	-0.5	-0.1	-0.6	0	-0.2	0.3	-0.1	0	0.3	0.2
opCjV0100000529	0.1	0.1	0.1	0	-0.1	-0.1	-0.1	-0.2	0.1	0
opCjV0100000530	0	0	-3.5	0.1	0	0.1	-1.3	-0.1	0.1	0
opCjV0100000532	0.2	0	0.3	0	0	0.1	0.1	0	-0.2	-0.1
opCjV0100000533	-0.6	0	-0.7	0	0	0	0	-0.1	0	0
opCjV0100000534	0.5	0.5	0.4	0.1	-0.2	0.1	0.2	0	0	0.1
opCjV0100000535	0.5	0.3	0.5	0	-0.1	0.2	0.1	0	0	-0.1
opCjV0100000536	1.1	1	0.6	0.3	-0.2	0.1	0.3	-0.1	0.1	0.1
opCjV0100000538	0.1	0.1	-0.3	-0.2	-0.1	0.1	0	0	-0.1	-0.1
opCjV0100000539	0.3	0.4	0.4	0	-0.2	0.1	0.1	0	0	0

opCjV0100000540	0.7	0.4	0.2	0.1	-0.1	0	0	0.1	0.1	0.2
opCjV0100000542	0	0.1	0.1	0.1	0.1	0.1	0	-0.1	0	0.1
opCjV0100000543	0.8	0.4	0.3	0.2	0.2	-0.1	0	-0.1	0	0
opCjV0100000545	-0.1	-0.2	-0.6	0	0.1	0.1	-0.2	-0.1	0.1	0
opCjV0100000546	0.8	0.6	0.7	0	0.3	-0.1	0.1	0	-0.1	-0.1
opCjV0100000547	0.2	0.1	0.3	0.1	-0.3	0.1	0	-0.1	0	0
opCjV0100000549	-4.4	-4.3	-2.5	-1.5	1.7	-0.9	-1.2	1.4	-1.5	-1.6
opCjV0100000550	0.1	0	0.2	0.2	0	0	0	-0.1	-0.1	0
opCjV0100000552	-0.1	0.3	0	-0.1	-0.1	0.1	-0.1	0	0.2	-0.1
opCjV0100000553	-8.6	-5.7	-2.9	-1.7	3.6	-1.8	-2.5	2.6	-2.9	-1.9
opCjV0100000555	-0.6	-0.4	-1.5	0.1	0.4	-1.2	-1.9	-1.4	-0.2	-0.1
opCjV0100000557	0.3	0.4	0.1	0.1	-0.2	0.1	0	-0.2	0	-0.1
opCjV0100000558	0	0.3	0.3	0.2	0.1	0.2	0.1	-0.1	-0.1	0.1
opCjV0100000559	0.4	0.2	0.5	0	0	0.1	0.1	-0.1	0.1	0
opCjV0100000560	0.9	0.5	0.9	0.1	0	0.1	0.2	0	0	-0.1
opCjV0100000562	-3.9	-3.2	-5.3	-1.1	1.5	-1.8	-1.3	1.1	-1	-1.2
opCjV0100000563	0.8	0.3	0.3	-0.1	-0.1	0.2	0.2	-0.1	0.1	0

opCjV0100000564	0.5	0.4	0	0.1	-0.1	0	-0.1	0	0	0
opCjV0100000565	-1.5	-1.3	-1.7	0	0.5	0	-0.4	0.2	-0.4	0.1
opCjV0100000566	0.1	0	0	0.1	0.1	0	0.1	0	0	0
opCjV0100000567	0.2	0.3	0.2	0.1	0.1	0	0	-0.1	0	-0.2
opCjV0100000568	0.3	0.4	0.4	0	-0.1	0	0.1	0	0	0
opCjV0100000569	0.6	0.3	0.3	0.1	-0.1	0.2	0	-0.5	-0.1	0
opCjV0100000570	0.6	0.1	0.6	0.1	0.1	0	0.2	0	-0.2	-0.2
opCjV0100000571	-0.7	-0.1	-0.6	0	-0.1	0.3	-0.2	0	0.3	-0.1
opCjV0100000572	0.4	0.3	0.1	0	0	0	0.1	0	-0.1	-0.1
opCjV0100000573	0	0.4	0.2	0.1	0	0.1	0	-0.1	0.1	0
opCjV0100000574	0.3	0.1	0.2	-0.1	-0.1	0	0	-0.1	0.1	0
opCjV0100000576	-1.4	0	-0.5	0	0.2	-0.1	-0.4	0	-0.1	0
opCjV0100000577	-0.4	0	-0.1	0	0.1	0.1	0	0.2	0.1	0
opCjV0100000578	0.8	0.5	0.4	0.3	0.3	0	0.1	0	-0.1	-0.1
opCjV0100000579	0.3	0	0	0.1	0.1	0	0	0	-0.1	0
opCjV0100000581	-0.2	-0.5	-0.3	-0.2	0.3	-0.3	0	0.4	-0.2	-0.3
opCjV0100000582	-0.1	-0.2	-0.1	0	0	0.2	0.1	0.1	-0.1	0

opCjV010000587	-0.2	-0.2	0	-0.1	-0.1	0.3	0	0.2	0	0
opCjV010000588	0.4	0.2	0.4	0	0	0.1	0.1	0.1	0	0
opCjV010000589	0.6	-0.4	0.1	0.1	0.2	0	0	0.3	-0.4	-0.2
opCjV010000590	0.7	0.2	0.9	0.1	0.2	-0.1	0.1	-0.1	-0.1	0
opCjV010000591	-1	-0.8	0.5	-0.2	0.5	0.1	-0.3	0.5	-0.2	-0.2
opCjV010000594	0.7	0.1	0.6	0.1	0.1	0	0.1	0	-0.1	0
opCjV010000595	-2.9	0.1	-0.9	0	0	-0.4	-0.7	-0.1	-0.1	-0.1
opCjV010000596	-0.8	0	0.2	0	0	0	0.1	-0.1	0	0
opCjV010000598	-2.7	-2.2	-2.4	-2.3	2.8	-1.9	-2.2	2.5	-0.6	-1.9
opCjV010000600	0.3	0.4	-0.1	0.1	0	-2.2	-3	0	0.1	0
opCjV010000601	-2.5	0	0	0.2	0.3	0.2	0.3	0.9	-1.1	0.2
opCjV010000604	-2	-1	-0.9	-1.9	1.7	-2	-1.9	1	-0.1	-1.7
opCjV010000605	-1	-0.3	-0.6	-0.1	-0.1	0.3	-0.2	-1.7	0.2	0.1
opCjV010000606	-0.4	-0.1	0	0	0	0.2	0	-0.1	-0.2	-0.1
opCjV010000609	0.3	-0.2	0	0.1	0.1	0.1	-0.1	0	0	0
opCjV010000611	-1.9	-0.1	-1.4	0	0.1	-1.6	-0.3	0.2	-0.2	-0.1
opCjV010000612	-0.4	-0.1	0.1	0.1	-0.1	0.3	0	0.2	0.2	0

opCjV0100000613	0.1	0.4	-0.9	0.1	0	0	0.2	-0.1	-0.1	0
opCjV0100000614	-0.5	-0.3	-2.7	0	0	0.2	0	0.1	0.2	0.1
opCjV0100000615	-1.4	0.1	-0.1	-0.1	0.5	0	-1.1	0.2	-0.3	-0.1
opCjV0100000616	0.5	0.4	0.6	0.1	-0.1	0.2	0.2	0	0	-0.1
opCjV0100000617	0.4	-0.2	0.4	0	0.2	0.1	-0.1	-0.1	-0.1	0.1
opCjV0100000618	0.2	0.2	-0.1	0	0	0	-0.2	-0.2	0	-0.1
opCjV0100000620	0.2	-3.4	0.1	-0.7	1.4	-0.3	-0.5	1.7	-0.1	-1.1
opCjV0100000622	-0.1	-0.2	0.7	-0.1	-0.1	0.2	0.1	0	0	0
opCjV0100000623	0	0.2	0.1	0.1	0.2	0.1	0	0	-0.2	0
opCjV0100000624	-3.8	-2.2	-1.9	-0.7	0.9	-1.1	-1.7	-0.1	-1.3	-0.7
opCjV0100000625	0.1	0.3	0.2	0.1	0	0.1	0.1	0.1	0.1	-0.1
opCjV0100000626	-5.6	-5.5	0.2	-1.5	2.4	0.3	0.1	2.9	-1.6	-1.6
opCjV0100000627	0.3	-0.1	0.2	0	0.2	0.2	-0.1	0	0.2	0
opCjV0100000628	0.3	0.1	0.5	0	0	0	0.3	0.3	-0.1	-0.1
opCjV0100000629	-0.3	-0.1	-0.3	0	0	0.2	0.1	0.1	0.1	0
opCjV0100000630	-0.2	-0.1	-0.3	-0.2	0	0	-0.2	0.1	0	-0.1
opCjV0100000631	-0.3	-0.2	-0.4	-0.1	0	0.1	0.1	0.1	0.1	0

opCjV0100000634	0.1	0	-0.1	0	0.1	0.2	0.1	0.1	0	-0.1
opCjV0100000635	0.5	0	0.3	0	0.1	0.1	0.1	0.1	-0.1	-0.1
opCjV0100000636	-1.1	-0.4	-0.5	0	0	0.3	0.1	0.2	0.5	0.1
opCjV0100000637	0.2	0	0	0	-0.1	0.2	0.2	-0.1	0	0
opCjV0100000638	-0.1	-0.3	-0.2	0	0	-0.5	0.1	0	0	0.1
opCjV0100000639	-0.1	0.2	-0.7	0	-0.2	0.1	0	0	0.1	0
opCjV0100000640	0.4	0.2	-0.4	0.1	0	-2.7	0	0	-0.1	0
opCjV0100000642	0.1	-0.1	0.2	0	0	0.1	-0.1	0	-0.1	0
opCjV0100000644	-0.2	-1.6	-0.8	0	0.1	0	-1.3	-1.3	0	0
opCjV0100000645	-6	-4.7	-1.5	-1.3	2.2	-1.7	-1.8	1.8	-1.6	-1.6
opCjV0100000646	-0.3	0.1	-0.3	-0.1	-0.2	0.2	-0.5	-0.2	0.2	0
opCjV0100000647	-0.2	0	-0.4	0	-0.2	0.3	0	-0.2	0.1	0.2
opCjV0100000649	0.1	-0.3	0.4	0	0.1	0.1	0	0.2	-0.2	0
opCjV0100000650	0.4	0.2	0.1	0	0.1	-0.1	0	0	-0.1	-0.1
opCjV0100000651	0.4	0.1	0.1	0.2	0.2	0.2	0.1	0.1	-0.1	0.1
opCjV0100000652	-0.7	-0.3	-0.9	-0.1	0	0	-0.1	0.1	0.2	0
opCjV0100000654	-1.3	-0.3	-0.7	-2.1	3.1	-2.6	-2.7	3.3	0	-0.4

opCjV0100000656	-0.2	-0.2	-2.7	0	0	-0.6	0	0	0	0
opCjV0100000658	0.3	0	0	0	0	0.1	0.1	0.2	-0.3	-0.1
opCjV0100000659	0.4	-0.1	0.4	0	0	0	0.1	0.1	-0.2	-0.2
opCjV0100000660	0.1	0.1	0.1	0.1	0.1	0	0.2	0.1	0	-0.1
opCjV0100000663	0.3	0.2	0.5	0	0	0.2	0	0	-0.1	-0.1
opCjV0100000664	0.1	-0.2	0.7	-0.1	0	0	0	0.2	-0.3	-0.1
opCjV0100000665	-0.8	-0.6	-0.6	0	-0.1	0.2	0	0.1	0.1	0.1
opCjV0100000667	0	0.1	0.8	-1.9	2.9	-2.7	-2.5	3	-0.1	-1.5
opCjV0100000668	0.4	0.3	-4.7	0	-0.1	-2	0.1	0	0	0
opCjV0100000669	0	0	-0.1	-0.1	-0.1	0.2	0.1	-0.1	-0.1	0.1
opCjV0100000670	-0.8	0	-1.6	-0.3	-0.4	-0.1	-0.1	-0.3	0.5	0.1
opCjV0100000671	0.4	0.3	-0.1	0	-0.1	-1.6	0	-0.1	0.3	0
opCjV0100000672	0	0	0.2	0	0	0.1	0.1	0	-0.3	-0.1
opCjV0100000673	0.3	0	0.3	0	0.1	0	0.1	0.2	0	-0.1
opCjV0100000674	-0.6	0	-0.6	0	-0.2	0.1	0	0.1	0.2	0
opCjV0100000676	0.3	0.1	0.8	-0.1	-0.1	0.1	0.1	0.1	0	0
opCjV0100000677	0.7	0.5	0.5	0.1	-0.2	0.2	0.1	0	0.2	-0.1

opCjV0100000678	-0.6	-0.3	-0.6	-0.1	0	0.2	0	0.2	0.1	0.1
opCjV0100000679	-0.2	-0.2	0.1	-0.1	0.1	-0.1	-0.1	0.4	-0.1	-0.3
opCjV0100000680	0.9	0.8	0.9	0.1	-0.3	0.2	0.1	-0.1	0.2	-0.1
opCjV0100000681	-0.5	0	-0.6	-0.1	-0.2	0.2	0	0	0.2	-0.1
opCjV0100000683	0.4	0.3	0.3	0.1	0	0	0.1	0.1	0	-0.1
opCjV0100000684	0.2	0.1	0.6	-0.1	-0.1	0	0	0	0.1	-0.1
opCjV0100000685	-5.2	-5.9	-0.5	-1.8	2.1	-1.6	-2.1	2.1	-3.2	-1.8
opCjV0100000687	-0.3	-0.2	0.1	-0.1	0	0.1	-0.1	-0.1	0.1	0
opCjV0100000688	0.2	0	0.2	0	0	0	0	0	-0.1	0
opCjV0100000689	0	0	-2.4	0	0.1	-2.4	-1	0	-0.1	0
opCjV0100000690	-0.1	-2.1	0.6	-0.1	0.7	-0.1	0.2	0.2	0.1	0
opCjV0100000691	0.6	0.4	0.8	0.1	0.1	0	0	0	0	-0.1
opCjV0100000692	0	0.3	0.1	0	-0.1	0.1	-0.1	-0.1	0	-0.1
opCjV0100000693	0.6	0.5	0.7	-0.1	-0.2	0	-0.1	-0.2	0.4	-0.1
opCjV0100000695	0.2	-0.7	-0.6	0	0.3	0	0	0.1	0	0
opCjV0100000696	0.7	0.4	0.6	0.1	0	-0.1	0.1	0.1	-0.1	-0.1
opCjV0100000697	-0.4	-0.2	-0.2	0	-0.1	0	-0.2	0.1	0.1	0

opCjV0100000698	-0.3	-0.1	-7.7	-0.1	-0.2	-2.4	0	0	0.1	0
opCjV0100000699	0	0.1	-5.1	-0.1	-0.2	1.7	0	0.1	0.2	0
opCjV0100000700	0.5	0.1	0.5	0.1	0	0	0.2	0.1	-0.2	-0.1
opCjV0100000701	-0.8	-0.8	0.2	-0.2	0	0.2	0.1	0.5	0	-0.2
opCjV0100000702	-0.1	0	-0.8	0	0	-2.3	0.1	0.1	-0.1	-0.2
opCjV0100000703	0.3	0.1	0.2	0.1	-0.1	0.1	0	0.1	0	0
opCjV0100000704	0.3	-0.1	0.3	-0.1	-0.1	0.1	-0.1	0.1	0	-0.1
opCjV0100000705	0.3	0	0.5	0	0.1	0.1	0	0.2	-0.1	-0.2
opCjV0100000706	0.7	0	0.5	0.1	0	0	0.1	0.2	-0.1	-0.1
opCjV0100000707	0.6	0.4	0.6	0.1	0	0	0.1	0.1	0	-0.1
opCjV0100000709	0.6	0.4	0.8	0	0	0.1	0.3	-0.1	0	0
opCjV0100000712	0.3	0	0.2	0.1	0.1	0	0	0	-0.1	0
opCjV0100000713	-0.5	-0.1	-0.4	0	-0.1	0.2	-0.1	-0.1	0.2	0
opCjV0100000714	-0.5	-0.5	-0.4	-0.1	-0.2	0	-0.1	-0.4	0.2	0
opCjV0100000715	0.3	0.3	0.4	0.1	-0.1	0.3	0.1	-0.1	0	0
opCjV0100000716	0.4	0.2	0.4	0	0	0.2	0.1	-0.1	-0.1	0.1
opCjV0100000717	1.1	0.8	0.7	0.2	0.2	-0.3	0.2	-0.1	0.1	-0.1

opCjV0100000718	-1.7	0	0.2	0	-0.1	0.1	-0.1	-0.1	0	0
opCjV0100000720	0	0	0	0	0.1	-0.1	-0.3	-0.1	-0.1	-0.2
opCjV0100000722	-1.5	-0.8	-1.6	-1.7	2	-3	-0.1	2	-0.1	-1.6
opCjV0100000723	-0.2	0.1	-0.1	-0.1	-0.2	0.1	0	0.1	0.2	-0.1
opCjV0100000725	0.1	-0.1	-0.2	0	-0.1	0.1	-0.1	0.1	0	0
opCjV0100000726	-0.1	-0.3	-0.2	-0.1	0	-0.1	0.2	0.4	-0.1	-0.2
opCjV0100000727	-0.2	-0.1	0	-0.1	-0.1	0.1	0	0.1	-0.1	-0.1
opCjV0100000728	0.2	-0.1	0.1	0	0.2	0	0.1	0	-0.2	-0.1
opCjV0100000729	-0.7	-0.4	-0.4	-0.2	0	0	-0.1	0.3	0.1	-0.1
opCjV0100000733	-0.5	-0.1	-0.6	0	-0.1	0.1	0	-0.1	0.2	0
opCjV0100000734	0.4	0.3	-5.9	0	0	-2	0.1	0	0	0
opCjV0100000735	0.3	0.2	0	-0.1	-0.1	-0.2	-0.2	0	0.1	0
opCjV0100000736	0.7	0.4	0.7	0.1	0	0.2	0.2	0	0.1	0
opCjV0100000737	0.7	0.5	0.7	0.2	0	0.1	0.1	-0.1	-0.1	0
opCjV0100000738	0.6	-0.2	-0.1	0	-0.1	0.1	0.1	-0.1	-0.1	-0.1
opCjV0100000739	0.1	-0.3	0	0	0	0.1	0	-0.1	0.1	0
opCjV0100000740	0.8	0.5	0.6	0.1	0	0	0.2	0	0	-0.1

opCjV0100000741	0	0.1	0.2	-0.1	-0.1	0.1	0.1	0	0.2	0
opCjV0100000743	-0.8	-0.3	-0.8	-1.7	2.9	-2.1	-0.3	3.7	-0.3	-1.8
opCjV0100000744	-0.6	-0.1	-0.8	0	0	-0.1	0	0.1	0.1	0
opCjV0100000747	0.1	0	0.1	0.1	0	0	-0.1	0.1	0	-0.1
opCjV0100000748	0.2	0.3	0.2	0.1	-0.1	0.2	0.1	0	0.1	0
opCjV0100000749	0.4	0.2	-0.2	0	0.1	-0.2	0	-0.1	-0.1	0
opCjV0100000750	0.1	0	0.2	-0.1	0	0.1	0.1	0	0.1	0
opCjV0100000751	0.6	0.1	0.2	0.1	0	-0.1	0.1	0	-0.1	-0.2
opCjV0100000752	-0.9	-0.6	0.4	-0.1	0.4	0	0	0.3	-0.2	-0.4
opCjV0100000753	-0.1	-0.2	0.1	0	0	0	0	0.1	-0.1	-0.3
opCjV0100000754	-0.2	-0.1	-0.2	0.1	0.1	0.1	0	0.2	-0.1	-0.1
opCjV0100000755	0.7	0.6	0.2	0	0	0.2	0.2	0.1	0.1	-0.1
opCjV0100000759	0.1	-0.3	0.4	-0.1	0.3	-0.1	-0.1	0	-0.2	-0.1
opCjV0100000760	-7.3	-4.7	-2.2	-0.1	2.3	-1.8	-2	0.1	-2.5	-0.2
opCjV0100000761	-0.5	-0.4	-0.3	0	0	0.3	-0.2	-0.3	0.1	0
opCjV0100000762	0.2	0.2	0.2	0	0.1	0.2	0.1	-0.2	0	-0.1
opCjV0100000765	0.2	0.3	-0.2	-2.3	2.3	-2.2	-2.7	2.3	0.1	-2

opCjV0100000766	-0.7	-0.2	-0.5	-0.1	-0.2	0.2	-0.1	0	0	0.1
opCjV0100000767	0.9	0.5	0.5	0.1	-0.1	0	0.1	-0.1	0	0
opCjV0100000768	-0.5	0.2	-0.5	-0.2	-0.4	0.2	0	-0.1	0.3	0.1
opCjV0100000769	-0.5	0.1	-0.7	-0.1	-0.3	-0.1	-0.4	-0.4	0.1	0
opCjV0100000770	0.6	0.4	0.5	0	-0.1	0	0	-0.1	-0.1	0
opCjV0100000771	0.6	0	0.6	0.1	0.1	0.1	0.2	0	-0.1	0
opCjV0100000774	0	0.1	0	0	0.1	0	0.1	0	-0.1	-0.1
opCjV0100000775	-1.8	-1.5	-6.1	-0.3	0.6	-2.2	-2.1	0.4	-0.4	-0.6
opCjV0100000777	-5	-3.8	-3	-1.1	1.8	-1.5	-2.3	1.8	-1.8	-1.6
opCjV0100000778	0.3	0.3	0.4	0	-0.1	-0.2	0	0	-0.1	-0.1
opCjV0100000779	-0.2	-0.1	-0.1	0	0	0	0	-0.1	0.1	0
opCjV0100000780	0.4	0.2	0.1	0	0.1	-0.1	0	0	-0.1	-0.1
opCjV0100000781	-0.3	0	0.1	-0.1	-0.1	0.2	-0.4	-0.2	0.2	0.1
opCjV0100000782	0.3	0.4	0.5	-0.1	0.1	0.1	0	-0.2	-0.1	0
opCjV0100000783	0.1	0.2	0.2	0	0.1	0.1	0	-0.1	0	0
opCjV0100000786	0.7	0.7	0.8	0.1	0	-0.2	0	0	0.2	0
opCjV0100000787	-1.1	-1.5	-0.8	-0.3	0.3	-0.1	-0.2	0.1	0.3	-0.2

opCjV0100000788	-0.2	-1.5	0.4	-0.4	0.1	0.1	0.1	0.2	-0.3	-0.1
opCjV0100000789	0.5	0.4	0.3	0.2	0	0.1	0.1	-0.3	0.1	0.2
opCjV0100000792	0.5	0.3	0.3	0.2	0	0	0	0	0.1	-0.1
opCjV0100000793	-1	-0.5	-0.2	0	-0.1	0.2	-0.1	0.1	0.1	-0.1
opCjV0100000794	-3.2	-2.6	-3	-0.6	1.2	-0.7	-1.3	2	-0.9	-1
opCjV0100000795	0.1	0.1	0.3	-0.1	-0.1	0.2	0	0.1	0	0
opCjV0100000796	0.2	0.4	0.4	0.1	0	0.2	0	0.1	0	0
opCjV0100000797	-0.2	0.2	-0.1	0.1	-0.1	0.1	0	0.1	0.1	0
opCjV0100000798	0.6	0.2	0.1	-1.2	3	-2.2	-2.1	2.3	0.1	-2.1
opCjV0100000799	-0.2	0.4	-0.7	0	-0.1	0	-0.4	0	0	0
opCjV0100000800	0.5	0.2	0.3	0	0	0.2	0	0	0	-0.1
opCjV0100000802	0.9	1	1.2	0.3	0.1	0	0.1	-0.1	-0.3	0
opCjV0100000804	-4.3	-3.1	-1	-1.1	1.2	-1.2	-1.2	1	-1.2	-1.3
opCjV0100000807	-0.1	-0.1	0.1	0	0.1	0.1	-0.1	0	0.1	0
opCjV0100000809	0.5	0.2	0.7	0	0.3	-0.1	0.1	-0.1	-0.4	0
opCjV0100000810	0.5	0.5	1	0	0	0.1	0.5	0.2	0	0.1
opCjV0100000812	0	0	-0.1	0.1	0.1	0.1	0	-0.2	0.2	0

opCjV0100000813	-0.2	-0.2	-0.3	0	0	0.1	0	0	0	0
opCjV0100000815	0.5	0.4	-0.5	0.2	0.1	-0.1	-0.5	-0.4	0	0
opCjV0100000816	0.3	0	-0.2	0	-0.1	0	0.1	-0.3	0	0
opCjV0100000817	-0.1	0.2	-1.6	0	-0.4	0.3	-0.1	0	0.1	0.1
opCjV0100000818	0	0.1	0.1	0	-0.1	0.1	0.1	0	0.1	0.1
opCjV0100000819	-0.2	0.1	0.4	0	0.2	0	0	0	-0.1	0
opCjV0100000820	0.2	0	0.3	0.1	-0.4	0.3	0.2	0.1	0.4	0.1
opCjV0100000821	0.3	0.3	0.3	0.1	0	0.1	0	0	0.1	0
opCjV0100000822	0	0	-0.4	0	0	-0.1	-0.1	-0.3	0.1	0
opCjV0100000823	-0.1	0	-0.3	-0.1	-0.3	0.2	0	0	0.2	0.1
opCjV0100000824	0.2	0.3	0.1	-0.1	-0.3	0	-0.1	-0.1	0.1	0.1
opCjV0100000828	-3.4	-2.9	-3.4	-1	1.1	-0.8	-1	1	-1.5	0.1
opCjV0100000829	0	0.1	0.3	0.1	0.2	-0.1	0	-0.1	0.2	0
opCjV0100000830	0.5	0.3	0.6	0	0.1	0.1	0.1	-0.1	0	0.1
opCjV0100000832	-5.8	-0.2	-2.1	-1.4	1.9	-1.8	-0.1	1.7	-2.4	-1.4
opCjV0100000834	0.8	0.5	1.1	0.1	0.3	0	0.1	0	0	0
opCjV0100000835	-0.7	-0.2	-0.8	-0.2	0.1	0.3	-0.2	-0.1	0.3	0

opCjV0100000836	0.4	0.3	0.8	-0.1	0	0	0	0	0.2	-0.1
opCjV0100000837	-0.2	-0.3	-0.3	-0.1	0	0.3	-0.1	-0.1	0.2	0
opCjV0100000839	0.8	0.5	0.3	0.2	0.1	-2	0	0.2	0.1	-0.1
opCjV0100000840	-0.4	0.2	0.2	0	-0.1	0.1	0	0	0	0
opCjV0100000841	-0.6	-0.1	-0.5	-0.2	-0.2	0.1	-0.1	-0.1	0.1	0.1
opCjV0100000842	0.4	0.4	0.3	0.1	-0.1	0	-0.1	0	0.2	0
opCjV0100000843	-0.9	-1	0	-0.2	0.5	0	0.1	0.7	-0.4	-0.4
opCjV0100000844	-2.2	0.1	-1.2	0	0	-1	-0.2	0	0	-0.1
opCjV0100000845	-1.8	-1	-3	0	-0.1	-0.5	-1	-0.2	0	-0.1
opCjV0100000846	0.7	0.4	0.5	0.1	0	0.1	0.1	-0.1	0	0
opCjV0100000847	-0.3	0	-0.2	-0.1	-0.2	0.2	0	0	0.1	0
opCjV0100000849	0.2	0.2	0.3	0	0	0.1	-0.1	0	-0.2	-0.1
opCjV0100000851	0.1	0	0.3	0	0.1	0	-0.2	-0.1	0	-0.1
opCjV0100000853	0.1	0.1	0.2	0	0.1	0.1	0	0	0.2	0
opCjV0100000855	0.1	0.1	0.2	0.1	2.5	0.2	0	-0.1	0.2	0.1
opCjV0100000856	-0.6	-0.6	-0.1	0	0.2	0.2	-0.2	-0.1	-0.1	0
opCjV0100000857	-0.1	-0.7	0.8	-0.1	0.3	0.1	0.1	0.5	-0.3	-0.2

opCjV0100000858	-0.7	-0.7	-0.2	-0.1	0	0.2	0	-0.2	0.2	0.1
opCjV0100000859	-0.2	0.2	0.5	0.1	0.3	0	0	0	0	0
opCjV0100000860	-0.4	-0.5	-0.2	-0.1	0.3	-0.2	-0.3	0	-0.1	-0.1
opCjV0100000861	-6.2	-0.1	-6.2	0	0.1	-1.8	-2	-0.3	-1.2	0
opCjV0100000862	0.5	0.3	0.5	0	-0.2	0.1	0.1	-0.1	0.1	0
opCjV0100000863	0.5	0.1	0.4	0.2	-0.1	0.1	0.3	-0.2	0.1	-0.1
opCjV0100000864	0.3	0.3	0	0	-0.2	0	0.1	-0.1	0	0
opCjV0100000865	0.4	0.1	0.2	0.1	0	0	-0.1	0.1	0.1	0
opCjV0100000866	-0.8	0	-0.1	-0.1	-0.2	0.1	-0.2	-0.1	0	0.1
opCjV0100000867	0	-1.7	-0.7	-0.2	2.2	-0.4	-2.2	0.1	0	-0.2
opCjV0100000868	-0.5	-0.2	-0.7	-0.1	0	-1.4	-0.3	0.1	-0.1	-0.1
opCjV0100000869	-0.3	0	-0.2	-0.1	-0.1	0	-0.1	-0.1	0	0
opCjV0100000870	0.1	0.2	-0.1	0	0	0	-0.1	0	0.1	-0.1
opCjV0100000872	-5.4	-4.5	-5.5	-1.1	1.8	-1.6	-1.6	1.7	-1.4	-1.4
opCjV0100000873	0.2	-0.1	0	0	0.1	0	0	0.1	-0.1	-0.2
opCjV0100000874	-0.1	-0.4	-0.8	-0.2	0.1	-0.2	-0.2	-0.1	-0.3	-0.2
opCjV0100000875	-0.7	-0.4	-0.9	-0.1	0	-0.8	-0.3	-0.9	0.1	-0.1

opCjV0100000876	0.1	-0.2	0.3	0.1	0.1	0	0	0.1	-0.1	0
opCjV0100000877	0.2	-0.2	-1.8	0.1	0.3	0	0	-0.2	0.1	0.1
opCjV0100000878	-0.4	-0.5	0.2	0.1	0.2	0.1	-0.1	-0.1	-0.1	0
opCjV0100000879	0.1	0	0.1	0.1	1.3	0.1	0	0	0	0
opCjV0100000880	-0.2	-0.1	0	-0.1	0	0.2	-0.1	0	0.4	0.1
opCjV0100000881	-0.6	0.1	-0.1	0.1	0.2	0.1	-0.2	0.1	0	0
opCjV0100000883	-0.6	0	-1	-0.2	0	-1.3	-0.3	-1.1	0.4	0
opCjV0100000884	-0.4	0	-0.1	0.1	0.2	0	-0.2	0.1	0.1	-0.1
opCjV0100000887	0.5	0.2	0.3	0	-0.2	0.2	0.1	0	0.1	0
opCjV0100000888	-0.4	-0.2	-0.2	-0.1	0	0.1	-0.1	-0.1	0	-0.1
opCjV0100000890	0	0.1	-0.1	0	0	0.2	-0.4	-0.1	0.1	-0.2
opCjV0100000891	0.3	0	-1.7	-1.6	1.9	-2.1	-2.1	1.6	-0.1	-1.8
opCjV0100000892	-0.1	-0.3	0.3	-0.3	0.2	-0.1	-0.1	0.3	-0.3	-0.3
opCjV0100000894	-0.4	-0.1	-0.6	0.1	0	0.2	-0.1	0	0	0
opCjV0100000895	-2.8	-1.4	-0.8	-0.3	0.4	-1.4	-1.1	0	-0.3	-0.5
opCjV0100000896	-7	-0.2	-1.7	0	2.2	-1.6	-2.3	-0.1	-1.4	-0.1
opCjV0100000897	-2.5	-1.9	-1.6	-0.6	0.6	-0.7	-0.9	0.2	-0.6	-0.6

opCjV010000898	-0.2	-0.3	-0.2	-0.2	0	0.3	-0.2	-0.2	-0.1	0.1
opCjV010000899	0.2	0.1	0.4	0.2	0.3	0	0	-0.1	-0.1	0
opCjV010000900	-0.5	-0.5	-0.3	-0.1	0.2	0.1	-0.2	0	-0.2	-0.1
opCjV010000901	-0.4	0	-0.3	-0.4	-0.1	0.1	0	0	0.1	0.1
opCjV010000902	-7.4	-0.2	-3.4	0	0.6	-1.1	-0.7	-0.1	-0.4	0.1
opCjV010000904	-0.2	-0.1	-0.6	0	0	0	0	-0.1	0	0.1
opCjV010000905	-0.3	-0.1	-0.1	-0.1	-0.1	0.2	0	-0.2	0.2	0.2
opCjV010000906	-0.1	-0.1	-0.1	0	0	0	-0.1	-0.2	-0.1	0
opCjV010000907	0	0.1	0.4	-0.1	0	0.1	-0.1	0.1	0	0
opCjV010000908	-0.1	-0.2	0.3	0	0	0.1	0	-0.1	0.1	0.1
opCjV010000909	0.3	0	-0.1	-0.5	0.3	-0.5	-0.6	0.2	-0.1	-0.7
opCjV010000910	-3	-2.6	-1.4	-0.5	0.5	-0.3	-0.7	0.4	-0.6	-0.5
opCjV010000911	-0.4	-0.2	-3	-0.2	-0.1	-1.2	-1.7	-1.1	-0.1	0
opCjV010000912	-0.7	-0.3	-1.1	-0.2	-0.2	0.1	-0.1	0	0.1	-0.1
opCjV010000913	-0.2	0.3	0.2	0	-0.2	0.2	0	0	0.2	0.1
opCjV010000914	0.5	0.2	-0.7	0.1	0	-0.2	0.1	-0.1	0.1	0
opCjV010000915	0	-1.6	-0.4	-0.5	0.6	-1.6	-0.7	0.5	-0.1	-0.6

opCjV0100000916	-0.1	0.4	0.5	0.1	-0.1	0	0	-0.1	0.1	-0.1
opCjV0100000917	-3.6	-3	-1.4	-0.8	1.3	-1	-1.3	1.1	-0.9	-0.9
opCjV0100000918	0.4	0.2	0.3	0	0	0	-0.1	0	0	0
opCjV0100000919	-0.8	-0.2	-0.7	-0.2	-0.2	0.2	-0.1	-0.1	0.1	0
opCjV0100000920	-0.2	-0.3	0.2	-0.1	0.6	-0.1	-0.6	-0.1	-0.2	-0.3
opCjV0100000921	-3.9	-2.5	-1.3	-0.8	0.8	-0.4	-0.8	0.6	-0.6	-0.6
opCjV0100000922	-0.3	-0.3	-2.2	-0.1	0.1	0.2	-0.4	0	0	0
opCjV0100000923	0.5	0.2	0.8	0.1	0.4	0.1	0.1	0	-0.1	0
opCjV0100000924	-2.5	-0.3	-1	0	0	0.2	-0.6	-0.1	-0.6	0.1
opCjV0100000925	0.4	0.2	-0.8	0	0	-0.8	-0.6	-0.6	0.1	0
opCjV0100000927	0.4	0.2	0.7	0.1	0.2	0.1	0	-0.2	0.1	0.1
opCjV0100000929	0	-0.2	0.1	0	0.1	-0.1	0	-0.1	-0.1	0
opCjV0100000930	-0.4	-0.1	-0.1	0	0	0.1	0	0	0.1	0
opCjV0100000931	-0.3	-0.3	0	-0.1	0	0	0	0.1	0.1	0
opCjV0100000933	0.3	0.1	0.5	0.1	0.2	-0.2	0	0	-0.1	-0.1
opCjV0100000935	0.6	0.5	0.1	0.1	-0.1	0	0	-0.2	0.1	0
opCjV0100000936	-1.3	-1.3	-0.6	-0.4	0.3	-0.2	-0.6	0.4	-0.3	-0.2

opCjV010000937	0.2	0.2	0.2	-0.1	-0.1	0	0	-0.2	0.1	0
opCjV010000938	0.1	0.2	-0.4	0	-0.2	-0.1	0.1	0	0.2	0.1
opCjV010000940	0.2	0.3	0.2	0	-0.2	0	0	-0.1	0	-0.1
opCjV010000941	-0.1	-0.2	0.1	-0.1	0	0.2	0	0.2	-0.1	0
opCjV010000942	0	0	0.1	0.1	0.1	-0.1	0	0	-0.1	-0.2
opCjV010000943	-0.1	-0.1	0.1	-0.1	-0.2	0.1	-0.1	0	0.1	-0.1
opCjV010000944	-2.6	-2.4	-2.7	-0.8	1.3	-2	-1.3	1.9	-0.6	-1
opCjV010000945	-1	-0.4	-0.3	-0.1	0	0.2	0	-0.1	0.1	0
opCjV010000946	0.4	0.2	0.4	0	-0.1	0.2	0.1	0	-0.2	0.1
opCjV010000947	-0.2	-0.1	-0.6	-0.2	0	0	-0.3	-0.1	-0.1	-0.1
opCjV010000948	0.4	-0.2	0.5	0.1	0.1	0.1	0	0.1	-0.1	0
opCjV010000951	-5.3	-5.6	-4.7	-1.6	1.7	-1.3	-1.9	2	-2	-1.6
opCjV010000952	0.1	0.2	-0.2	0	-0.1	-0.1	-0.1	-0.1	0.1	-0.1
opCjV010000953	0.5	0.1	0.5	0.1	0	0.1	0.1	0	0	0
opCjV010000954	-1.2	0	-0.6	-0.1	-0.3	0.2	-0.1	0	0.3	0
opCjV010000956	0.4	0.2	0.3	0.1	-0.1	0.2	0	-0.2	0.1	0
opCjV010000958	0.4	0.1	0.3	0	0.1	-0.1	0.1	0.2	-0.1	-0.2

opCjV0100000960	-0.2	0.1	-0.4	0.1	0	0	0	0	0.1	-0.3
opCjV0100000962	-0.7	-0.3	-0.9	-0.1	-0.2	0.3	0	0	0.3	0.2
opCjV0100000963	0.8	0.5	0.8	0	0	0.1	-0.9	-0.5	0	0
opCjV0100000965	0.1	-0.2	-0.5	0	0.1	0.1	0	0.2	0	-0.1
opCjV0100000966	-0.5	-0.1	-0.6	-0.1	-0.2	0.1	-0.1	0	0.1	-0.1
opCjV0100000967	-0.2	-0.1	-0.1	-0.1	-0.1	0.2	0	-0.2	0.1	0
opCjV0100000969	0.1	0.1	0.2	0	-0.1	0.2	0.1	0.1	0.1	0
opCjV0100000970	-0.1	0	-0.2	0	-0.1	0.3	0.1	-0.1	0	0.1
opCjV0100000971	-1.1	0.2	-0.5	-0.1	0	-0.1	-0.2	0	0	0.1
opCjV0100000972	0.7	0.6	0.5	0	0.1	-0.1	0.1	0	-0.1	-0.1
opCjV0100000974	0.2	0.1	0.2	0	-0.1	-0.1	-0.1	0	0.1	0
opCjV0100000975	0.6	0.2	0.6	0	0	0	0	0	-0.1	0
opCjV0100000977	0.3	0.3	0.4	0	-0.1	0	0.1	0	-0.1	-0.1
opCjV0100000979	0.7	0.3	1	0	0	0.3	0.3	0.2	0	0
opCjV0100000981	0	-0.1	0	-0.1	0.2	0	-0.1	0.1	-0.2	-0.1
opCjV0100000982	-0.4	-0.8	-0.3	-0.1	0.2	0.1	-0.1	0.1	-0.2	-0.1
opCjV0100000983	-0.8	-0.6	-0.8	-1.3	1.8	0	-0.4	2.2	0.1	-1.2

opCjV010000984	-0.1	-0.1	-0.1	-0.1	-0.3	0.2	0	0	0.2	0
opCjV010000986	-0.3	-0.1	-0.5	0.1	0	0.2	0	0.1	0.3	0
opCjV010000988	0.2	0	0.1	0	0.1	0.1	0.2	0.1	-0.1	-0.1
opCjV010000989	0.2	0	-5.6	0.1	0	0.3	0	-1.9	-0.2	0
opCjV010000991	-0.6	-0.3	-0.8	-0.1	-0.1	0.2	0	-0.1	0.1	0
opCjV010000992	0.7	0.5	0.2	0.2	0.1	-0.1	0.1	-0.1	-0.3	0
opCjV010000993	-0.5	-0.4	-5	-0.1	0	0.1	-0.2	-1.4	-0.1	-0.1
opCjV010000994	0.3	-0.1	0.3	0	0.1	0.1	0	0	-0.1	-0.1
opCjV010000995	0.2	0	0.3	0.1	0.1	0.1	0.1	-0.1	-0.1	0
opCjV010000996	0	0.2	0	-0.1	-0.2	0.1	-0.1	-0.2	0.4	0
opCjV010000997	0.4	0.1	0.3	0.1	0.1	0	0.1	0	0.1	0
opCjV010000998	0.2	0.1	0.3	0.1	0.2	0.1	-0.1	0	-0.1	0
opCjV010000999	-5.7	-5.9	-4.9	-1.5	1.9	-1.2	-1.8	1.8	-2	-1.6
opCjV0100001000	0.4	0.3	0.4	0	0	0.1	0.1	-0.1	-0.1	0
opCjV0100001001	-3.8	-1.2	-2.1	-0.4	0.5	-0.5	-0.1	-0.4	-0.1	-0.4
opCjV0100001002	0.7	0.6	0.8	0.1	0	0.1	0.1	0	0	0
opCjV0100001003	0.2	0.4	0.2	0.1	0	0.1	-0.2	-0.1	0.1	0

opCjV0100001004	-0.1	0	0.1	0	0.1	0.1	-0.2	-0.2	0	-0.1
opCjV0100001005	0	0	0.4	0	0	0.2	0.1	0	-0.1	-0.1
opCjV0100001006	-0.6	-0.5	-3.6	-0.1	0	0.3	-0.3	-1.3	0.3	-0.1
opCjV0100001007	0.4	0.4	0.4	0	-0.1	-0.1	-0.1	0.1	0.1	-0.1
opCjV0100001008	-0.3	0.1	-0.9	-0.1	-0.3	0.1	-0.3	-0.2	0.3	0.1
opCjV0100001010	0	0.1	0.2	-0.1	-0.2	0.3	0.2	0	0	0
opCjV0100001011	0.4	0	0.2	-0.1	0	0	0	0	-0.1	-0.1
opCjV0100001012	0.4	0.1	-0.4	0	0	0	-0.1	0.2	0	-0.2
opCjV0100001013	0.2	0.1	0.1	-0.1	0	0.1	0.1	0.2	0	-0.1
opCjV0100001014	0.3	-0.2	0.3	0.1	0	0.1	0.1	0	-0.1	-0.1
opCjV0100001016	-0.1	0	-0.2	-0.1	0	0.1	0.1	-0.1	-0.1	-0.1
opCjV0100001017	1.3	1	0.7	0.1	-0.1	0.2	0.2	-0.2	0.2	0.1
opCjV0100001018	0.4	0.2	-4.2	0	0	-1.4	0	0	-0.1	-0.1
opCjV0100001019	0.3	0.1	0.4	0.1	0	0.3	0	-0.2	0	0.1
opCjV0100001020	0	-0.3	-0.8	-1.7	3.2	-2.5	-2.5	3.9	-0.3	-1.8
opCjV0100001021	0.4	0.4	0.4	-0.1	-0.2	0.1	0.1	0	0.2	0
opCjV0100001022	0.2	0	0	0	0	0	-0.2	-0.1	0	0

opCjV0100001023	0.1	0.3	0.1	0.1	0	0.2	-0.2	-0.2	0	0
opCjV0100001024	-0.3	0.2	-1.1	-0.1	-0.1	-2.1	-0.1	-0.1	0.1	0
opCjV0100001025	-6.5	-6	-3	-0.2	3	-2.4	-2.6	0.6	-2.2	-0.3
opCjV0100001026	-5.3	-4.4	-6.7	-1.2	1.6	-2.2	-0.2	2.9	-1.2	-1.4
opCjV0100001028	-0.3	0	-0.1	0	-0.1	0.1	0	0	0.1	0
opCjV0100001029	0.2	-0.4	0.3	-0.1	0.1	0.2	0	-0.9	0	0
opCjV0100001030	-0.2	-0.1	-0.4	0	-0.1	0.1	-0.1	0.1	0	0
opCjV0100001031	0.2	0.1	0.2	0	0	0	0.1	0.3	-0.1	-0.1
opCjV0100001032	0.2	0.1	0.3	0	-0.1	0.1	-0.1	-0.2	-0.1	-0.1
opCjV0100001034	0.4	0.3	0.2	0	0	0	0	0.2	-0.3	-0.1
opCjV0100001035	0.3	0.2	0.1	0	0	0	-0.1	0.2	0	-0.1
opCjV0100001036	0.3	0.3	0.3	0	0	0.2	0.1	0.1	-0.1	0
opCjV0100001038	0.5	0.2	0.3	0	0.1	-0.1	0.1	0.1	-0.1	-0.2
opCjV0100001039	0.8	0.2	0.3	0.2	0.2	0	0.1	0.1	-0.1	-0.1
opCjV0100001040	-0.4	-0.1	0.3	0	0.2	0.2	0.2	0.5	-0.4	-0.2
opCjV0100001041	0.4	0.2	-0.2	0	0.1	-1.4	0	0	-0.1	-0.1
opCjV0100001042	0.2	0.1	0.1	0	0	0	0	0.1	-0.1	-0.2

opCjV0100001044	-3	0.5	0.3	-0.2	-0.2	0.2	0.4	0.3	0.2	-1.2
opCjV0100001045	-0.5	-0.1	-0.3	0	0	0.2	-0.1	0	0.3	0
opCjV0100001046	0.3	0.5	0.6	0	-0.1	0.1	0.2	0	0	0
opCjV0100001047	-0.1	0.2	-0.2	0	-0.1	0.2	0	-0.1	0.1	0
opCjV0100001048	0	0.2	0	0.1	-0.2	0.3	0	0	0.3	0
opCjV0100001049	0.4	0.3	0.2	0.2	0	0	0.1	-0.1	0	0
opCjV0100001050	-0.3	0	0.4	0	-0.1	0.3	0	0	0	-0.1
opCjV0100001051	0.2	0.1	0	0	0	0	-0.1	-0.1	0.1	0
opCjV0100001052	-0.5	-0.2	-0.8	-0.1	-0.1	0.1	-0.1	0.1	0.1	0
opCjV0100001054	-0.2	-0.3	-0.1	0	0	0.1	0	0.1	-0.1	0
opCjV0100001055	0	0	-0.2	0	0	0	0	0.1	0	0
opCjV0100001056	0.2	0.1	0.1	0.1	-0.1	0.1	0.1	0.2	-0.1	-0.1
opCjV0100001057	-4.5	-0.7	-0.6	-0.1	0	-1.1	-1.8	-0.1	-0.1	-0.1
opCjV0100001058	-3.8	-3.2	-3.4	-1.2	2	-1.3	-1.5	1.3	-1.2	-1.4
opCjV0100001061	-4.5	0.1	-2.6	0	-0.2	-0.9	-0.8	-0.9	0.1	0
opCjV0100001063	-0.8	-0.4	-0.8	-0.1	-0.1	0.3	-0.1	0.1	0.4	0
opCjV0100001066	-0.3	0.1	-0.4	0	-0.1	-0.3	-0.2	-0.2	0.1	0

opCjV0100001067	0.3	0.1	0.5	0.1	0.2	0	0.1	0	-0.2	0
opCjV0100001068	-0.5	-0.4	-2	-0.1	0.1	0.2	-0.1	-1	0.2	0
opCjV0100001069	0.3	0.2	0.3	0	0	-0.1	-0.2	0.1	0.2	0
opCjV0100001070	-0.5	-0.1	-0.4	0	0	0.2	-0.1	0.1	0.1	0
opCjV0100001071	-0.7	-0.2	-0.5	-0.1	-0.1	0.1	-0.1	0	0.1	-0.1
opCjV0100001072	-0.3	-0.1	-0.1	0	0	0.2	0	0	-0.1	-0.1
opCjV0100001073	-0.4	-0.2	-0.8	0	0	-1	0	-0.1	0.1	0
opCjV0100001074	-3	-0.7	-2.7	0	-0.2	-0.7	-0.8	0.4	0.9	0
opCjV0100001075	0.6	0.4	0.5	0	0	0	0	-0.1	0	0
opCjV0100001076	-0.1	-0.1	-0.1	-0.2	0.2	-0.2	-0.1	0.2	0	-0.2
opCjV0100001077	0.1	0.1	0.2	0	0	0	0.1	0.2	-0.1	-0.1
opCjV0100001078	0.2	0.1	0.1	0	0	-0.1	-0.1	0.2	0.1	0
opCjV0100001079	-0.5	-0.5	-0.9	0	0.2	0.1	-0.1	0	-0.1	-0.1
opCjV0100001081	0.5	-0.5	0.4	0.1	0	0.1	0.2	0	0	-0.1
opCjV0100001085	-6.4	-4.1	-3.5	0	1.3	-2	-1.8	-1	-0.8	-1.3
opCjV0100001087	-5.1	-3.7	-0.1	-0.9	1.8	-1.1	-1.5	1.7	-1.2	-1.2
opCjV0100001088	0.2	0	0	-0.1	-0.2	0	0	-0.1	0	0

opCjV0100001089	0.1	0.3	0.2	0.1	0	0	0.1	0.1	0.1	0
opCjV0100001090	0	0	-0.1	0	0.1	0.1	0	-0.2	0	0
opCjV0100001093	-4.1	0	-2.8	0	0	-1	0.1	0.1	0	0
opCjV0100001094	-0.8	0.1	0.3	0	0	0.2	0.1	0	0	-0.1
opCjV0100001095	0.4	0.3	0.6	0	0	0	-0.2	0.1	1.3	0.1
opCjV0100001096	0.7	0.4	-3.6	0	0	-1.5	-1.8	-1.6	0	-0.1
opCjV0100001097	0.1	-0.1	0.1	0	0	0.1	0.1	-0.1	-0.1	0.1
opCjV0100001098	-1.5	-0.7	0.9	-1	1.2	-0.1	-0.7	1.7	-0.8	-0.9
opCjV0100001099	-0.3	0	-0.3	0	-0.1	0.2	0	0	0.1	0.1
opCjV0100001100	0.6	0.4	0.4	0	0	0	0.2	0	0	-0.1
opCjV0100001101	-0.4	0	-0.3	0.1	-0.2	0.2	0	0	0.1	-0.1
opCjV0100001102	-3.4	-3	-0.9	-0.9	2	-0.7	-1.4	1.6	-1	-1.1
opCjV0100001103	0.5	0.3	0.3	-0.1	-0.1	0.1	0	0	0.1	-0.1
opCjV0100001104	0.1	0.2	0.4	0	-0.2	0.1	0	-0.1	0.1	-0.1
opCjV0100001105	-0.3	-0.1	-4.7	0.1	-0.1	-1.4	0	-0.1	0.3	0
opCjV0100001106	0.2	0	0.3	-0.1	0	-1	0	0.1	-0.1	-0.1
opCjV0100001107	-0.3	0.2	0.7	0.1	-0.1	0	-0.1	0	0	-0.1

opCjV0100001108	0.7	0.3	1	0.1	0	0	0.1	0.1	0	-0.1
opCjV0100001109	0.6	0.1	0.4	0.1	0.2	0	0	-0.1	-0.2	-0.2
opCjV0100001110	-0.9	-0.5	-0.6	0	-0.1	0.2	-0.2	0.1	-0.3	-0.1
opCjV0100001111	0.7	0.3	0.6	0.2	0.1	-0.1	0.3	0	-0.1	-0.2
opCjV0100001112	-1.4	-0.3	-0.9	-0.1	0	0.1	-0.1	0.3	0	0
opCjV0100001113	0.2	0.2	0.4	-0.1	0	0.1	0	-0.1	-0.1	0
opCjV0100001114	-6	-6.7	-2.1	-1.1	3.3	-0.9	-3.5	2.9	-2.6	-1.2
opCjV0100001115	0.2	0.1	0.3	0	0	0	0	-0.1	-0.1	0
opCjV0100001116	1	0.7	0.7	0.1	0.1	-0.1	0.1	0	0.1	0
opCjV0100001117	0.3	0.1	0.4	0.1	0.1	0.1	0	-0.1	-0.1	0
opCjV0100001118	0.6	0	-0.1	0.1	0	0.2	0	0	0.1	0
opCjV0100001120	0.6	0.1	0.2	0.1	0	-0.1	0.1	-0.1	0.1	0
opCjV0100001121	-3.3	-3.1	-3.1	-2	2.9	-1.7	-1.7	1.7	-1.1	-2.1
opCjV0100001122	-0.3	0.1	-0.1	-0.1	-0.2	0.2	0	0	0.1	-0.1
opCjV0100001123	0.4	0.2	0.4	0	-0.1	0	0.1	0	0	-0.1
opCjV0100001124	-0.5	-0.1	-2.1	0	-0.1	-1	0	0	0.1	0
opCjV0100001125	-0.2	-0.2	-2.7	0.1	0	0.1	-0.1	-0.6	0	0

opCjV0100001126	0.7	0.4	0.5	0	-0.2	0.2	0	0.1	0	0
opCjV0100001127	0	0.1	0.1	-0.1	-0.2	0	-0.1	-0.3	0	-0.1
opCjV0100001128	0.5	0.7	0.6	0	-0.2	0.2	0.1	0.1	0.3	0.2
opCjV0100001129	0.1	-0.2	0.1	0	-0.1	0.1	0	0.1	-0.1	0
opCjV0100001130	0	-0.3	-0.1	0	0.1	0.2	0.1	0	-0.1	0
opCjV0100001131	0.8	0.4	0.5	0.2	0	0	0.1	0.1	-0.1	-0.2

*The log 2 ratio values for each strain is calculated by taking average of the log 2 ratios for three independent array experiments.

Supplementary table 3: GACK trinary cutoffs for all genes on the array for six hyperinvasive and four low invasive *C. jejuni* strains.

Oligo_ID	Gene						Hyperinvasive <i>C. jejuni</i>						Low invasive <i>C. jejuni</i>				Role_11168	Role_RM1221	Role_RM2228
	Number_R M1221	Name_R M1221	Number_1 11168	Name_1 1168	Number_R M2228	Name_R M2228	01_10	01_35	01_04	01_41	01_51	EX 114	01_30	01_32	01_46	01_39			
"CJ_10001524"	CJE0001	dnaA	Cj0001	dnaA	CCO0032	DnaA	0	1	0	1	1	1	1	1	1	1	chromosomal replication initiator protein	chromosomal replication initiator protein	chromosomal replication initiator protein DnaA
"CJ_10001526"	CJE0002	dnaN	Cj0002	dnaN	CCO0033	DnaN	0	1	1	1	1	1	1	1	1	1	DNA polymerase III, beta chain	DNA polymerase III subunit beta	DNA polymerase III, beta subunit
"CJ_10001527"	CJE0003	gyrB	Cj0003	gyrB	CCO0034	GyrB	0	0	1	1	1	1	1	1	1	1	DNA gyrase subunit B	DNA gyrase subunit B	DNA gyrase, B subunit
"CJ_10001528"	CJE0004	-	Cj0004c	-	-	-	0	0	0	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10001529"	CJE0005	-	Cj0005c	-	-	-	1	1	1	1	1	1	1	1	1	1	putative molybdenum containing oxidoreductase	molybdopterin oxidoreductase family protein	-
"CJ_10001530"	CJE0006	-	Cj0006	-	CCO0041	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	Na ⁺ /H ⁺ antiporter family protein	probable integral membrane protein Cj0006
"CJ_10001531"	CJE0007	gltB	Cj0007	gltB	CCO0042	gltB	1	1	1	1	1	1	1	1	1	1	glutamate synthase (NADPH) large subunit	glutamate synthase, large subunit	glutamate synthase, large subunit
"CJ_10001533"	CJE0008	gltD	Cj0009	gltD	CCO0043	-	1	1	1	1	1	1	1	1	1	1	glutamate synthase (NADPH) small subunit	glutamate synthase, small subunit	glutamate synthase (NADPH) small chain Cj0009
"CJ_10001023"	CJE0009	rnhB	Cj0010c	rnhB	CCO0044	rnhB	1	1	1	1	-1	-1	1	-1	1	1	ribonuclease HII	ribonuclease HII	ribonuclease HII
"CJ_10001024"	CJE0010	comEA	Cj0011c	-	-	-	1	1	1	1	1	0	-1	-1	1	1	putative non-specific DNA binding protein	competence protein ComEA	-
"CJ_10001025"	CJE0011	rbr	Cj0012c	-	CCO0045	rr2	1	1	1	1	1	1	1	1	1	1	non-haem iron protein	rubrerythrin	rubrerythrin
"CJ_10001026"	CJE0012	ilvD	Cj0013	ilvD	-	-	1	1	1	-1	-1	1	1	0	1	1	dihydroxy-acid dehydratase	dihydroxy-acid dehydratase	-
"CJ_10001028"	CJE0013	-	Cj0014c	-	-	-	0	1	1	-1	-1	1	1	-1	1	1	putative integral membrane protein	pseudogene	-
"CJ_10001030"	CJE0015	-	Cj0015c	-	CCO0047	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001032"	CJE0016	-	Cj0016	-	CCO0049	-	1	1	1	1	1	1	1	1	1	1	putative transcriptional regulatory protein	ExsB	exsB protein
"opCcV010000 1629"	CJE0017	-	Cj0017c	-	CCO0050	-	1	0	1	1	1	1	1	1	1	1	putative ATP/GTP binding protein	disulfide bond formation protein, DsbB family	probable ATP/GTP binding protein Cj0017c
"CJ_10001036"	CJE0018	-	Cj0018c	-	CCO0051	-	1	1	1	1	-1	0	1	1	0	1	small hydrophobic protein	hypothetical protein	small hydrophobic protein Cj0018c -related
"CJ_10001038"	CJE0019	-	Cj0019c	-	-	-	1	1	1	1	-1	-1	1	1	1	-1	MCP-domain signal transduction protein	methyl-accepting chemotaxis protein	-
"CJ_10001051"	CJE0020	-	Cj0020c	-	-	-	0	1	1	-1	-1	1	1	0	1	1	cytochrome C551 peroxidase	cytochrome c551 peroxidase	-
"CJ_10001052"	CJE0021	-	Cj0021c	-	CCO0052	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	fumarylacetoacetate hydrolase family protein	fumarylacetoacetate hydrolase family protein
"CJ_10001053"	CJE0022	-	Cj0022c	-	CCO0053	-	1	1	1	0	-1	1	0	-1	1	-1	putative ribosomal pseudouridine synthase	RNA pseudouridylate synthase family protein	RNA pseudouridylate synthase family
"CJ_10001054"	CJE0023	-	Cj0023	purB	CCO0054	purB	1	1	1	1	-1	1	1	-1	1	1	adenylosuccinate lyase	adenylosuccinate lyase	adenylosuccinate lyase
"CJ_10001056"	CJE0024	nrdA	Cj0024	nrdA	CCO0055	-	1	1	1	1	-1	1	1	1	1	1	ribonucleoside-diphosphate reductase alpha	ribonucleotide-diphosphate reductase alpha	ribonucleoside reductase, alpha subunit
"CJ_10001058"	CJE0025	-	Cj0025c	-	CCO0056	gltP	1	1	1	1	1	1	1	1	1	1	putative transmembrane symporter	sodium/dicarboxylate symporter	sodium/dicarboxylate symporter family protein
"CJ_10001060"	CJE0026	thyX	Cj0026c	-	CCO0057	-	1	1	1	1	0	1	1	-1	1	-1	hypothetical protein	thymidylate synthase	Thymidylate synthase complementing protein

"CJ_10001063"	CJE0027	pyrG	Cj0027	pyrG	CCO0058	pyrG	1	1	1	1	-1	1	1	1	1	1	CTP synthase	CTP synthetase	CTP synthase
"CJ_10001065"	CJE0028	recJ	Cj0028	recJ	CCO0059	recJ	1	1	1	1	0	1	1	1	1	1	putative single-stranded-DNA-specific	single-stranded-DNA-specific exonuclease RecJ	single-stranded-DNA-specific exonuclease RecJ
"CJ_10001067"	CJE0029	ansA	Cj0029	ansA	CCO0060	-	1	1	1	1	0	1	1	-1	1	1	cytoplasmic L-asparaginase	L-asparaginase	L-asparaginase II
"CJ_10001084"	CJE0030	-	Cj0030	-	-	-	1	1	1	-1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10001086"	CJE0031	-	Cj0031	-	CCO0070	-	0	1	1	1	-1	1	1	1	0	1	putative type IIS restriction/modification	type II restriction-modification enzyme	type II restriction-modification enzyme
"CJ_10001089"	CJE0032	-	Cj0033	-	CCO0073	-	1	1	1	-1	0	1	1	1	1	-1	Putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0033
"CJ_10001092"	CJE0033	-	Cj0034c	-	CCO0075	-	1	1	1	0	-1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0034c
"CJ_10001095"	CJE0034	-	Cj0035c	-	CCO0076	-	1	1	1	-1	-1	1	1	1	1	0	putative efflux protein	drug resistance transporter, Bcr/CflA family	probable efflux protein Cj0035c
"CJ_10001100"	CJE0035	-	Cj0036	-	CCO0077	-	1	1	1	1	-1	1	1	-1	1	0	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001103"	CJE0036	-	Cj0037c	-	-	-	1	1	1	1	1	1	1	1	1	1	putative cytochrome c	cytochrome c family protein	-
"CJ_10001106"	CJE0037	-	Cj0038c	-	CCO0078	-	1	1	1	1	-1	1	1	-1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0038c
"CJ_10001109"	CJE0038	typA	Cj0039c	typA	CCO0079	typA	1	1	1	1	1	1	1	1	1	1	GTP-binding protein typA homolog	GTP-binding protein TypA	GTP-binding protein TypA
"CJ_10001119"	CJE0039	-	Cj0040	-	CCO0080	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001121"	CJE0040	-	Cj0041	-	CCO0081	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001123"	CJE0041	flgD	Cj0042	flgD	CCO0082	-	1	1	1	1	-1	1	1	1	1	1	putative flagellar hook assembly protein	flagellar hook assembly protein	probable flagellar hook assembly protein Cj0042
"CJ_10001125"	CJE0042	-	Cj0043	flgE	CCO0083	-	0	1	1	-1	-1	1	1	0	-1	0	flagellar hook protein flgE	flagellar hook protein	flagellar hook protein flgE Cj0043
"CJ_10001128"	CJE0043	-	Cj0044c	-	CCO0084	-	1	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001134"	CJE0044	-	Cj0045c	-	CCO0085	-	1	1	1	1	-1	1	-1	-1	1	1	putative iron-binding protein	hypothetical protein	conserved hypothetical protein
"opCjV0100001066"	CJE0045	-	-	-	-	-	1	1	1	1	1	0	1	0	1	1	-	hypothetical protein	-
"CJ_10001138"	CJE0046	-	Cj0046	-	CCO0087	-	1	1	1	1	-1	-1	1	-1	1	0	pseudogene	pseudogene	C4-dicarboxylate transporter
"CJ_10001141"	CJE0048	trmU	Cj0053c	trmU	CCO0088	trmU	1	0	1	1	1	1	1	1	1	1	tRNA	tRNA	tRNA
"CJ_10001144"	CJE0049	-	Cj0054c	-	CCO0089	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	lysine decarboxylase family protein	Predicted Rossmann fold nucleotide-binding
"opCjV0100000022"	CJE0050	-	-	-	-	-	-1	1	0	0	1	-1	-1	1	1	-1	-	hypothetical protein	-
"opCjV0100000192"	CJE0051	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV0100001058"	CJE0052	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV0100000772"	CJE0053	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"CJ_10001165"	CJE0054	-	Cj0057	-	CCO0113	-	1	1	-1	1	1	1	1	1	1	1	putative periplasmic protein	hmcD domain protein	probable periplasmic protein Cj0057
"CJ_10001167"	CJE0055	-	Cj0058	-	CCO0114	-	1	1	1	1	-1	1	1	0	1	0	putative periplasmic protein	peptidase family protein	probable periplasmic protein Cj0058
"CJ_10001168"	CJE0056	fliY	Cj0059c	fliY	CCO0115	fliY	1	0	0	1	1	1	1	1	1	1	putative flagellar motor switch protein	flagellar motor switch protein	fliY protein (fliY)
"CJ_10001172"	CJE0057	fliM	Cj0060c	fliM	CCO0116	fliM	1	1	1	1	1	1	1	1	1	1	flagellar motor switch protein	flagellar motor switch protein	flagellar motor switch protein FliM

"CJ_10001176"	CJE0058	fliA	Cj0061c	fliA	CCO0117	-	1	1	1	1	1	-1	1	1	1	1	putative RNA polymerase sigma factor for	flagellar biosynthesis sigma factor FliA	probable RNA polymerase sigma factor for
"CJ_10001180"	CJE0059	-	Cj0062c	-	CCO0118	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0062c
"CJ_10001183"	CJE0060	-	Cj0063c	-	CCO0119	-	1	1	1	1	1	1	1	1	1	1	putative ATP-binding protein	ATPase, ParA family	ATP-binding protein (yIxB)
"CJ_10001186"	CJE0061	flhF	Cj0064c	flhF	CCO0120	-	1	1	1	1	1	1	1	1	1	1	flagellar biosynthesis protein	flagellar biosynthesis protein	flagellar biosynthetic protein FlhF, putative
"CJ_10001188"	CJE0062	folK	Cj0065c	folK	CCO0121	folK	0	0	1	1	1	1	1	1	1	1	putative	2-amino-4-hydroxy-6-	2-amino-4-hydroxy-6-hydroxymethylidihydropteridine pyrophosphokinase
"CJ_10001210"	CJE0063	aroQ	Cj0066c	aroQ	CCO0122	aroQ	1	1	1	1	1	1	1	1	1	1	3-dehydroquininate dehydratase	3-dehydroquininate dehydratase	3-dehydroquininate dehydratase, type II
"opCcV010000682"	CJE0064	-	Cj0067	-	CCO0123	-	1	1	1	-1	1	1	1	1	1	1	hypothetical protein	chlorohydrolase	chlorohydrolase
"opCcV010000642"	CJE0065	sppA	Cj0068	pspA	CCO0124	sppA	1	0	1	1	1	1	1	1	-1	1	protease	signal peptide peptidase SppA, 36K type	protease IV (PspA)
"CJ_10001216"	CJE0066	-	Cj0069	-	CCO0125	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001219"	CJE0067	-	Cj0070c	-	CCO0126	-	-1	1	1	1	-1	1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001223"	CJE0068	-	Cj0072c	-	-	-	1	0	1	1	1	1	1	1	1	1	pseudogene	pseudogene	-
"CJ_10001227"	CJE0069	-	Cj0073c	-	CCO0129	-	1	1	-1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001230"	CJE0070	-	Cj0074c	-	CCO0130	-	1	1	1	1	0	1	1	1	1	1	putative iron-sulfur protein	iron-sulfur cluster binding protein	iron-sulfur cluster binding protein
"CJ_10001232"	CJE0071	-	Cj0075c	-	CCO0131	glcF	1	1	1	1	1	1	1	1	1	1	putative oxidoreductase iron-sulfur subunit	cysteine-rich domain protein	conserved hypothetical secreted protein
"CJ_10001235"	CJE0072	lctP	Cj0076c	lctP	CCO0132	-	1	1	1	1	1	1	1	1	1	1	L-lactate permease	L-lactate permease	L-lactate permease (lctP)
"CJ_10001257"	CJE0073	cdtC	Cj0077c	cdtC	-	-	0	1	0	1	1	1	1	0	1	1	cytolethal distending toxin	cytolethal distending toxin, subunit C	-
"CJ_10001259"	CJE0074	cdtB	Cj0078c	cdtB	CCO0369	cdtB	1	1	0	1	1	1	1	1	1	1	cytolethal distending toxin	cytolethal distending toxin, subunit B	cytolethal distending toxin A
"CJ_10001261"	CJE0075	cdtA	Cj0079c	cdtA	CCO0370	cdtA	0	1	1	-1	0	1	1	-1	1	1	cytolethal distending toxin	cytolethal distending toxin, subunit A	cytolethal distending toxin A
"CJ_10001264"	CJE0076	-	Cj0080	-	CCO0133	-	1	1	1	1	1	-1	1	-1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0080
"CJ_10001266"	CJE0077	cydA	Cj0081	cydA	CCO0134	cydA	1	1	1	1	1	1	1	1	1	1	cytochrome bd oxidase subunit I	cytochrome d ubiquinol oxidase, subunit I	cytochrome d ubiquinol oxidase, subunit I
"CJ_10001269"	CJE0078	cydB	Cj0082	cydB	CCO0135	cydB	1	1	1	1	1	1	1	0	1	1	cytochrome bd oxidase subunit II	cytochrome d ubiquinol oxidase, subunit II	cytochrome d ubiquinol oxidase, subunit II
"opCjV010000945"	CJE0079	-	-	-	-	-	0	0	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_10001272"	CJE0080	-	Cj0085c	-	CCO0139	-	1	1	1	1	1	1	1	1	1	1	putative amino acid recemase	aspartate racemase, putative	aspartate racemase
"CJ_10001273"	CJE0081	ung	Cj0086c	ung	CCO0140	ung	1	1	1	1	1	1	1	1	1	1	uracil-DNA glycosylase	uracil-DNA glycosylase	uracil-DNA glycosylase
"CJ_10001275"	CJE0082	aspA	Cj0087	aspA	CCO0141	aspA	1	1	1	1	1	1	1	1	1	1	aspartate ammonia-lyase	aspartate ammonia-lyase	aspartate ammonia-lyase
"CJ_10001277"	CJE0083	dcuA	Cj0088	dcuA	CCO0142	-	1	1	1	-1	-1	1	1	1	1	1	putative anaerobic C4-dicarboxylate transporter	anaerobic C4-dicarboxylate transporter	Dcu family anaerobic dicarboxylate transport
"CJ_10001287"	CJE0084	-	Cj0089	-	CCO0143	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0089
"CJ_10001290"	CJE0085	-	Cj0090	-	-	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	-
"CJ_10001292"	CJE0086	-	Cj0091	-	CCO0144	-	0	1	1	1	1	1	1	0	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0091
"CJ_10001294"	CJE0087	-	Cj0092	-	-	-	1	1	1	1	-1	-1	1	1	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10001296"	CJE0088	-	Cj0093	-	CCO0145	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0093 , putative

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"CJ_10001299"	CJE0089	rplU	Cj0094	rplU	CCO0146	rplU	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L21	50S ribosomal protein L21	ribosomal protein L21
"CJ_10001301"	CJE0090	rpmA	Cj0095	rpmA	CCO0147	rpmA	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L27	50S ribosomal protein L27	ribosomal protein L27
"CJ_10001303"	CJE0091	-	Cj0096	-	CCO0148	obg	1	1	1	1	1	1	1	1	1	1	putative GTP-binding protein	GTP-binding protein, GTP1/Obg family	GTP-binding protein Obg
"CJ_10001305"	CJE0092	proB	Cj0097	proB	CCO1787	proB	1	1	1	1	1	1	0	1	1	1	putative glutamate 5-kinase	gamma-glutamyl kinase	glutamate 5-kinase
"CJ_10001307"	CJE0093	fmt	Cj0098	fmt	CCO1783	fmt	1	-1			-1	-1	1	-1	-1	-1	methionyl-tRNA formyltransferase	methionyl-tRNA formyltransferase	methionyl-tRNA formyltransferase
"CJ_10001316"	CJE0094	-	Cj0099	birA	CCO1782	-	1	1	1	1	1	0	1	1	1	1	putative biotin--[acetyl-CoA-carboxylase]	biotin--protein ligase	biotin--acetyl-CoA-carboxylase ligase
"CJ_10001319"	CJE0095	-	Cj0100	-	CCO1781	-	1	1	1	1	1	1	1	1	1	1	parA family protein	chromosome partitioning protein, ParA family	parA family protein Cj0100
"CJ_10001322"	CJE0096	-	Cj0101	-	CCO1780	spo0J	0	0	1	1	1	1	1	1	1	1	parB family protein	chromosome partitioning protein, ParB family	chromosome partitioning protein, ParB family
"CJ_10001324"	CJE0097	-	Cj0102	-	CCO1779	-	1	-1	-1	-1	-1	-1	1	1	-1	1	ATP synthase F0 sector B' subunit	ATP synthase subunit B	ATP synthase F0, subunit b' (atpF), putative
"CJ_10001327"	CJE0098	-	Cj0103	atpF	CCO1778	-	1	1	1	1	1	1	1	1	1	1	ATP synthase F0 sector B subunit	ATP synthase subunit B	ATP synthase B/B' CF(0) superfamily
"CJ_10001330"	CJE0099	atpH	Cj0104	atpH	CCO1777	atpH	1	1	1	1	1	1	1	1	1	1	ATP synthase F1 sector delta subunit	ATP synthase subunit D	ATP synthase F1, delta subunit
"CJ_10001334"	CJE0100	atpA	Cj0105	atpA	CCO1776	atpA	1	1	1	1	1	1	1	1	1	1	ATP synthase F1 sector alpha subunit	ATP synthase subunit A	ATP synthase F1, alpha subunit
"CJ_10001337"	CJE0101	atpG	Cj0106	atpG	CCO1775	atpG	1	1	1	1	1	1	1	1	1	1	ATP synthase F1 sector gamma subunit	ATP synthase subunit C	ATP synthase F1, gamma subunit
"CJ_10001340"	CJE0102	atpD	Cj0107	atpD	CCO1774	atpD	1	1	1	1	1	1	1	1	1	1	ATP synthase F1 sector beta subunit	ATP synthase subunit B	ATP synthase F1, beta subunit
"CJ_10001343"	CJE0103	atpC	Cj0108	atpC	CCO1773	atpC	1	1		1	-1	1	-1	-1	1	-1	ATP synthase F1 sector epsilon subunit	ATP synthase subunit epsilon	ATP synthase F1, epsilon subunit
"CJ_10000466"	CJE0104	-	Cj0109	exbB3	CCO1772	-	1	1	1	1	1	1	1	1	1	1	exbB\toIQ family transport protein	MotA/TolQ/ExbB proton channel family protein	exbB\toIQ family transport protein Cj0109
"CJ_10000468"	CJE0105	-	Cj0110	exbD3	CCO1771	-	1	1	1	1	1	1	1	1	1	1	exbD\toIR family transport protein	TonB system transport protein ExbD, putative	exbD\toIR family transport protein Cj0110
"CJ_10000470"	CJE0106	-	Cj0111	-	CCO1770	-	1	1	1	1	1	-1	1	-1	-1	-1	periplasmic protein	TonB domain protein	periplasmic protein Cj0111
"CJ_10000471"	CJE0107	tolB	Cj0112	-	CCO1769	tolB	1	1	1	1	1	1	1	1	1	1	periplasmic protein	translocation protein TolB precursor	colicin tolerance-like protein (tolB)
"opCcV0100001578"	CJE0108	-	Cj0113	pal	CCO1768	-	1	1	1	1	1	1	1	1	1	1	peptidoglycan associated lipoprotein (omp18)	peptidoglycan-associated lipoprotein Omp18	omp18
"CJ_10000472"	CJE0109	-	Cj0114	-	CCO1767	-	0	1	1	1	0	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0114
"CJ_10000473"	CJE0110	slyD	Cj0115	slyD	CCO1766	slyD	1	1	1	1	1	0	1	1	1	1	peptidyl-prolyl cis-trans isomerase	FKBP-type peptidyl-prolyl cis-trans isomerase	FKBP-type peptidyl prolyl cis-trans isomerase;
"CJ_10000474"	CJE0111	fabD	Cj0116	fabD	CCO1765	fabD	1	1	0	1	1	-1	1	1	1	1	malonyl CoA-acyl carrier protein transacylase	acyl-carrier-protein S-malonyltransferase	malonyl CoA-acyl carrier protein transacylase
"CJ_10000475"	CJE0112	mtnA	Cj0117	pfs	CCO1764	-	1	1	1	1	1	1	1	1	1	1	5'-methylthioadenosine/S-adenosylhomocysteine	5'-methylthioadenosine/S-adenosylhomocysteine	MTA/SAH nucleosidase
"CJ_10000476"	CJE0113	-	Cj0118	-	CCO1763	-	0	-1	1	1	1	1	1	1	1	1	hypothetical protein	PP-loop family protein	conserved hypothetical protein
"CJ_10000487"	CJE0114	-	Cj0119	-	CCO1762	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	pyrazinamidase/nicotinamidase, putative	conserved hypothetical protein
"CJ_10000488"	CJE0115	-	Cj0120	-	CCO1761	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000489"	CJE0116	-	Cj0121	-	CCO1760	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR00043
"CJ_10000490"	CJE0117	-	Cj0122	-	CCO1759	-	0	0	-1	1	1	-1	-1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000491"	CJE0118	dusB	Cj0123c	-	CCO1756	-	1	1	1	1	1	1	1	1	1	1	putative transcriptional	tRNA-dihydrouridine	probable transcription

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"CJ_10000492"	CJE0119	-	Cj0124c	-	CCO1755	-	0	1	1	1	1	1	1	1	1	1	regulator	synthase B	regulator Cj0123c
"CJ_10000493"	CJE0120	-	Cj0125c	-	CCO1754	-	1	0	0	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0124c
"CJ_10000494"	CJE0121	-	Cj0126c	-	CCO1753	-	0	1	0	1	1	1	1	1	1	1	dksA-like protein	dnaK suppressor protein, putative	dnaK suppressor, putative
"CJ_10000495"	CJE0122	accD	Cj0127c	accD	CCO1752	accD	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical secreted protein
"CJ_10000496"	CJE0123	-	Cj0128c	-	CCO1751	-	1	1	1	1	1	1	1	1	1	1	acetyl-coenzyme A carboxylase carboxyl	acetyl-CoA carboxylase beta subunit	acetyl-CoA carboxylase, carboxyl transferase,
"CJ_10000507"	CJE0124	-	Cj0129c	-	CCO1750	-	1	0	1	1	-1	1	1	1	1	1	subB-like protein	Inositol monophosphatase family protein	conserved hypothetical protein
"CJ_10000508"	CJE0125	tyrA	Cj0130	tyrA	CCO1749	-	1	0	1	1	1	1	1	1	1	1	outer membrane protein	outer membrane protein, OMP85 family	outer membrane protein Cj0129c
"CJ_10000509"	CJE0126	-	Cj0131	-	CCO1748	-	1	1	1	1	1	1	1	1	1	1	putative prephenate dehydrogenase	prephenate dehydrogenase	Prephenate dehydrogenase
"CJ_10000510"	CJE0127	-	Cj0132	-	CCO1747	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	peptidase, M23/M37 family	probable periplasmic protein Cj0131
"CJ_10000512"	CJE0128	-	Cj0133	-	CCO1746	-	1	1	1	1	1	1	1	1	1	1	UDP-3-O-[3-hydroxymyristoyl] n-acetylglucosamine	UDP-3-O-[3-hydroxymyristoyl] N-acetylglucosamine	UDP-3-0-acyl N-acetylglucosamine deacetylase
"CJ_10000514"	CJE0129	thrB	Cj0134	thrB	CCO1745	thrB	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000516"	CJE0130	-	Cj0135	-	CCO1744	-	1	1	1	1	1	1	0	1	1	1	homoserine kinase	homoserine kinase	homoserine kinase
"CJ_10000517"	CJE0131	-	Cj0136	-	CCO1743	-	1	1	1	-1	1	1	-1	-1	-1	-1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF448)
"CJ_10000519"	CJE0132	infB	Cj0137	infB	CCO1743	infB	1	1	1	-1	1	1	-1	-1	-1	-1	translation initiation factor IF-2	translation initiation factor IF-2	translation initiation factor IF-2
"CJ_10000520"	CJE0133	-	Cj0138	-	CCO1742	rbfA	0	1	0	1	1	1	1	1	1	1	hypothetical protein	ribosome-binding factor A	ribosome-binding factor A
"CJ_10000536"	CJE0134	-	Cj0139	-	CCO1741	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000537"	CJE0135	-	Cj0140	-	-	-	1	1	1	1	1	1	-1	1	1	1	putative endonuclease	McrBC restriction endonuclease system, McrB	-
"CJ_10000538"	CJE0136	-	Cj0141c	-	-	-	0	-1	1	1	1	-1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10000539"	CJE0137	-	Cj0142c	-	CCO1737	-	0	0	1	1	1	1	1	1	1	1	ABC transporter integral membrane protein	cation ABC transporter, permease protein	ABC transporter integral membrane protein
"CJ_10000541"	CJE0138	-	Cj0143c	-	CCO1736	-	1	1	1	1	1	1	1	1	1	1	ABC transporter ATP-binding protein	cation ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein
"opCjV010000828"	CJE0139	-	-	-	CCO1735	-	1	1	1	1	1	1	1	1	1	1	periplasmic solute binding protein for ABC	cation ABC transporter, periplasmic	adhesion protein, putative
"CJ_10001578"	CJE0140	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	1	hypothetical protein	hypothetical protein	-
"CJ_10000544"	CJE0141	-	Cj1564	-	CCO1678	-	0	0	0	1	1	-1	1	-1	1	1	putative methyl-accepting chemotaxis signal	methyl-accepting chemotaxis protein	methyl-accepting chemotaxis protein (tlpA)
"CJ_10000546"	CJE0142	-	Cj0145	-	CCO1732	-	-1	0	0	1	1	-1	1	1	1	1	hypothetical protein	hypothetical protein	Tat (twin-arginine translocation) pathway signal
"CJ_10000548"	CJE0143	trx	Cj0146c	trx	CCO1731	trx	1	1	1	1	1	1	1	1	1	1	thioredoxin reductase	thioredoxin-disulfide reductase	thioredoxin reductase
"CJ_10000550"	CJE0144	-	Cj0147c	trxA	CCO1730	trx	1	1	1	1	1	1	1	1	1	1	thioredoxin	thioredoxin	thioredoxin
"CJ_10000554"	CJE0145	-	Cj0148c	-	CCO1729	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	Endonuclease
"CJ_10000564"	CJE0146	hom	Cj0149c	hom	CCO1728	-	1	1	1	1	1	1	1	1	1	1	homoserine dehydrogenase	homoserine dehydrogenase	homoserine dehydrogenase
"CJ_10000565"	CJE0147	-	Cj0150c	-	CCO1727	-	1	1	1	1	1	1	1	1	1	1	aminotransferase	aspartate aminotransferase	aspartate aminotransferase
"CJ_10000566"	CJE0148	-	Cj0151c	-	CCO1726	-	1	1	1	-1	1	1	-1	1	-1	-1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0151c
"CJ_10000567"	CJE0148	-	Cj0152c	-	CCO1725	-	1	1	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein

"CJ_10000569"	CJE0149	-	Cj0153c	-	CCO1724	-	1	1	1	1	1	1	1	1	0	1	1	putative rRNA methylase	RNA methyltransferase, TrmH family	Cj0152c RNA methyltransferase, TrmH family, group 3
"CJ_10000570"	CJE0150	-	Cj0154c	-	CCO1723	-	1	1	1	1	1	1	1	1	1	1	1	putative methylase	tetrapyrrole methylase family protein	conserved hypothetical protein TIGR00096
"CJ_10000572"	CJE0151	rpmE	Cj0155c	rpmE	CCO1722	rpmE	1	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L31	ribosomal protein L31	ribosomal protein L31
"CJ_10000574"	CJE0152	-	Cj0156c	-	CCO1721	-	0	-1	1	1	1	1	1	1	1	1	1	hypothetical protein	conserved hypothetical protein TIGR00046	conserved hypothetical protein TIGR00046
"CJ_10000576"	CJE0153	-	Cj0157c	-	CCO1720	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0157c
"CJ_10000578"	CJE0154	-	Cj0158c	-	CCO1719	-	1	1	1	1	1	1	1	1	1	1	1	putative haem-binding lipoprotein	cytochrome c family protein	conserved hypothetical protein
"CJ_10000592"	CJE0155	-	Cj0159c	-	CCO1718	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	6-pyruvoyl tetrahydrobiopterin synthase,	6-pyruvoyl tetrahydrobiopterin synthase,
"CJ_10000593"	CJE0156	-	Cj0160c	-	CCO1717	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	radical SAM domain protein	radical SAM domain protein, putative
"CJ_10000594"	CJE0157	-	Cj0161c	moaA	CCO1716	moaA	1	1	1	1	1	1	1	1	1	1	1	molybdenum cofactor biosynthesis protein A	molybdenum cofactor biosynthesis protein A	molybdopterin cofactor biosynthesis protein A
"CJ_10000595"	CJE0158	-	Cj0162c	-	CCO1715	-	0	0	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0162c
"CJ_10000596"	CJE0159	-	Cj0163c	-	CCO1714	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000597"	CJE0160	-	Cj0164c	ubiA	CCO1713	-	1	1	1	1	1	1	1	1	1	1	1	putative 4-hydroxybenzoate	4-hydroxybenzoate octaprenyltransferase	4-hydroxybenzoate octaprenyltransferase,
"CJ_10000599"	CJE0161	miaA	Cj0166	miaA	CCO1712	miaA	1	1	1	1	1	1	1	1	1	1	1	tRNA delta(2)-isopentenylpyrophosphate	tRNA delta(2)-isopentenylpyrophosphate	tRNA delta(2)-isopentenylpyrophosphate
"CJ_10000601"	CJE0162	-	Cj0167c	-	CCO1711	-	0	1	1	1	1	1	1	0	1	1	1	putative integral membrane protein	hypothetical protein	membrane protein, putative
"opCjjV010000101"	CJE0163	-	Cj0168c	-	-	-	1	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	-
"opCcV0100001164"	CJE0164	sodB	Cj0169	sodB	CCO1706	-	1	0	1	1	1	1	1	1	1	1	1	superoxide dismutase (Fe)	superoxide dismutase, Fe	superoxide dismutase (fe)
"CJ_10000628"	CJE0165	-	Cj0172c	-	CCO1705	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	saccharopine dehydrogenase	saccharopine dehydrogenase
"CJ_10000630"	CJE0166	-	Cj0173c	-	CCO1704	potA	1	1	1	1	1	0	1	1	0	1	1	putative iron-uptake ABC transport system	iron ABC transporter, ATP binding subunit	spermidine/putrescine ABC transporter,
"CJ_10000631"	CJE0167	-	Cj0174c	-	CCO1703	-	1	1	1	1	1	1	1	1	0	1	1	putative iron-uptake ABC transport system	iron ABC transporter, permease protein	iron compound ABC transporter, permease protein,
"CJ_10000632"	CJE0168	-	Cj0175c	-	CCO1702	-	1	1	1	1	1	0	1	1	1	1	1	putative iron-uptake ABC transport system	iron ABC transporter, periplasmic iron-binding	iron transport protein
"CJ_10000634"	CJE0169	-	Cj0176c	-	CCO1701	-	1	1	0	1	1	1	1	1	1	1	1	putative lipoprotein	hypothetical protein	probable lipoprotein Cj0176c -related protein
"CJ_10000636"	CJE0170	-	Cj0177	-	CCO1700	-	1	1	0	1	0	1	-1	-1	1	1	1	putative lipoprotein	TonB-dependent colicin lipoprotein, putative	lipoprotein, putative
"CJ_10000638"	CJE0171	-	Cj0178	-	CCO1699	-	1	1	-1	1	1	1	1	-1	1	1	1	putative outer membrane siderophore receptor	TonB-dependent colicin receptor protein,	TonB-dependent receptor
"CJ_10000640"	CJE0172	-	Cj0179	exbB1	CCO1698	-	1	1	-1	1	1	0	-1	-1	1	1	1	biopolymer transport protein	TonB system transport protein ExbB	biopolymer transport protein Cj0179
"CJ_10000654"	CJE0173	-	Cj0180	exbD1	CCO1697	-	0	0	1	1	1	1	1	1	-1	1	1	biopolymer transport protein	biopolymer transport protein, ExbD/ToIR family	biopolymer transport protein Cj0180
"CJ_10000656"	CJE0174	-	Cj0181	tonB1	CCO1696	-	1	1	-1	1	1	-1	-1	-1	1	1	1	possible tonB transport protein	TonB-dependent colicin receptor protein,	probable tonB transport protein Cj0181
"CJ_10000657"	CJE0175	-	Cj0182	-	CCO1695	-	1	1	1	1	1	-1	-1	1	-1	-1	-1	transmembrane transport	antibiotic transport protein,	transmembrane transport

"CJ_10000659"	CJE0176	-	Cj0183	-	CCO1693	-	1	1	1	1	1	1	1	1	1	1	protein	putative transporter, putative	protein Cj0182 transporter, putative
"CJ_10000660"	CJE0177	-	Cj0184c	-	CCO1692	-	1	-1	0	0	1	1	1	0	0	0	putative integral membrane protein with possible serine/threonine protein phosphatase	Ser/Thr protein phosphatase family protein	probable serine/threonine protein phosphatase
"CJ_10000661"	CJE0178	-	Cj0185c	-	CCO1691	phnA	1	1	1	1	1	1	1	1	1	1	phnA-like protein	PhnA domain protein	phnA protein
"CJ_10000663"	CJE0179	-	Cj0186c	-	CCO1690	-	1	1	1	1	0	1	1	1	1	1	putative integral membrane protein	integral membrane protein, TerC family	membrane protein, TerC family
"CJ_10000665"	CJE0180	purN	Cj0187c	purN	CCO1687	purN	-1	-1	-1	-1	1	-1	1	1	-1	1	phosphoribosylglycinamide formyltransferase	phosphoribosylglycinamide formyltransferase	phosphoribosylglycinamide formyltransferase
"CJ_10000667"	CJE0181	-	Cj0188c	-	CCO1686	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	YjeF-related protein N-terminus family
"CJ_10000669"	CJE0182	-	Cj0189c	-	CCO1685	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000683"	CJE0183	-	Cj0190c	-	CCO1684	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	Mg chelatase-related protein	Mg chelatase-related protein
"CJ_10000685"	CJE0184	def	Cj0191c	def	CCO1683	def	1	1	1	1	1	1	1	1	1	1	polypeptide deformylase	peptide deformylase	polypeptide deformylase
"CJ_10000687"	CJE0185	clpP	Cj0192c	clpP	CCO1682	clpP	1	1	1	1	1	1	1	1	1	1	ATP-dependent clp protease proteolytic subunit	ATP-dependent Clp protease proteolytic subunit	ATP-dependent Clp protease, proteolytic subunit
"CJ_10000689"	CJE0186	tig	Cj0193c	tig	CCO1681	tig	1	1	1	1	1	1	1	1	1	1	trigger factor (peptidyl-prolyl cis/trans	trigger factor	trigger factor
"CJ_10000690"	CJE0187	folE	Cj0194	folE	CCO1680	folE	1	1	1	1	0	1	1	1	1	1	GTP cyclohydrolase I	GTP cyclohydrolase I	GTP cyclohydrolase I
"CJ_10000691"	CJE0188	fliI	Cj0195	fliI	CCO1679	fliI	1	1	1	1	1	1	1	0	1	1	flagellum-specific ATP synthase	flagellum-specific ATP synthase	flagellum-specific ATP synthase
"CJ_10000693"	CJE0189	purF	Cj0196c	purF	CCO0286	purF	1	1	1	1	1	1	1	1	1	1	amidophosphoribosyltransferase	amidophosphoribosyltransferase	amidophosphoribosyltransferase
"CJ_10000695"	CJE0190	dapB	Cj0197c	dapB	CCO0287	dapB	1	1	1	1	1	1	1	1	1	1	dihydrodipicolinate reductase	dihydrodipicolinate reductase	dihydrodipicolinate reductase
"CJ_10000697"	CJE0191	-	Cj0198c	-	CCO0288	-	1	1	1	1	1	1	1	1	1	1	helicase-like protein	ATPase, AAA family protein	ATPase, AAA family
"CJ_10000699"	CJE0192	-	Cj0199c	-	CCO0289	-	1	1	1	1	-1	1	1	-1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0199c
"CJ_10000711"	CJE0193	-	Cj0200c	-	CCO0290	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0200c
"CJ_10000713"	CJE0194	-	Cj0201c	-	CCO0290	-	0	0	-1	1	1	-1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable periplasmic protein Cj0200c
"CJ_10000715"	CJE0195	-	Cj0202c	-	-	-	1	1	-1	1	1	-1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10000716"	CJE0196	-	Cj0203	-	-	-	1	1	1	1	1	1	1	1	1	1	putative transmembrane transport protein	pseudogene	-
"CJ_10000717"	CJE0197	-	Cj0204	-	CCO0294	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	oligopeptide transporter, OPT family	oligopeptide transporter, OPT family
"CJ_10000719"	CJE0198	-	Cj0205	bacA	CCO0295	-	1	0	1	1	1	1	1	1	1	1	putative undecaprenol kinase (bacitracin)	undecaprenol kinase, putative	undecaprenol kinase, putative
"CJ_10000721"	CJE0199	thrS	Cj0206	thrS	CCO0296	thrS	1	1	1	1	1	1	1	1	1	1	threonyl-tRNA synthetase	threonyl-tRNA synthetase	threonyl-tRNA synthetase
"CJ_10000723"	CJE0200	infC	Cj0207	infC	CCO0297	infC	1	-1	1	1	1	1	1	1	1	1	translation initiation factor IF-3	translation initiation factor IF-3	translation initiation factor IF-3
"CJ_10000725"	CJE0201	-	Cj0208	-	CCO0298	-	1	1	1	1	0	1	1	-1	1	1	DNA modification methylase (adenine-specific)	D12 class N6 adenine-specific DNA	ulcer associated adenine specific DNA
"opCjV0100001041"	CJE0202	-	-	-	-	-	1	1	1	1	1	-1	1	1	1	1	-	hypothetical protein	-
"opCjV0100000895"	CJE0203	-	-	-	-	-	-1	-1	0	0	1	-1	-1	1	1	0	-	hypothetical protein	-
"opCjV0100000671"	CJE0204	-	Cj0223	-	-	-	1	1	1	1	1	-1	1	1	1	1	pseudogene	hypothetical protein	-
"opCjV010000"	CJE0205	-	Cj0223	-	-	-	1	1	1	1	1	-1	1	1	1	1	pseudogene	hypothetical protein	-

0223"																				
"opCjV0100000062"	CJE0206	-	Cj0223	-	-	-	1	1	1	1	1	-1	1	1	1	1	pseudogene	hypothetical protein	-	
"opCjV0100000934"	CJE0207	-	Cj0223	-	-	-	1	1	0	1	1	-1	1	1	1	1	pseudogene	hypothetical protein	-	
"opCjV0100000204"	CJE0208	-	Cj0223	-	-	-	1	1	0	1	1	0	1	1	1	1	pseudogene	hypothetical protein	-	
"opCjV0100000005"	CJE0209	-	-	-	-	-	0	1	-1	1	1	-1	-1	1	1	1	-	hypothetical protein	-	
"opCjV0100000365"	CJE0210	-	Cj0223	-	-	-	0	0	-1	1	1	-1	-1	1	1	1	pseudogene	hypothetical protein	-	
"CJ_10000727"	CJE0211	-	Cj0223	-	-	-	1	1	-1	1	1	-1	0	1	1	1	pseudogene	pathogenicity domain protein	-	
"opCjV0100000640"	CJE0212	-	Cj0223	-	-	-	1	1	1	1	1	-1	1	1	1	1	pseudogene	pathogenicity protein, homolog	-	
"opCjV0100000722"	CJE0213	-	-	-	-	-	0	0	-1	-1	1	-1	1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000482"	CJE0214	-	-	-	-	-	-1	-1	0	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-	
"opCjV0100000190"	CJE0215	-	-	-	-	-	-1	-1	1	-1	1	-1	0	1	1	-1	-	phage repressor protein, putative	-	
"opCjV0100000686"	CJE0216	-	-	-	-	-	0	-1	0	-1	1	1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000915"	CJE0217	-	-	-	-	-	1	-1	1	0	1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000231"	CJE0218	-	-	-	-	-	0	-1	0	-1	1	0	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000743"	CJE0219	-	-	-	-	-	0	1	0	-1	1	-1	0	1	1	-1	-	hypothetical protein	-	
"opCjV0100000516"	CJE0220	dam	-	-	-	-	1	0	1	-1	1	-1	-1	1	1	-1	-	DNA adenine methylase	-	
"opCjV0100000891"	CJE0221	-	-	-	-	-	1	1	-1	-1	1	-1	-1	1	1	-1	-	phage virion morphogenesis protein, putative	-	
"opCjV0100000215"	CJE0222	-	-	-	-	-	1	1	-1	-1	1	-1	-1	1	1	-1	-	tail tape measure protein, TP901 family	-	
"opCjV0100000032"	CJE0223	-	-	-	-	-	1	1	-1	-1	1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000440"	CJE0224	-	-	-	-	-	1	1	-1	-1	-1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000893"	CJE0225	-	-	-	-	-	1	0	-1	-1	1	-1	0	1	1	-1	-	hypothetical protein	-	
"opCjV0100000592"	CJE0226	-	-	-	-	-	1	0	-1	-1	1	-1	1	1	1	-1	-	phage major tail tube protein, putative	-	
"opCjV0100000364"	CJE0227	-	-	-	-	-	1	0	1	-1	1	-1	-1	1	1	-1	-	major tail sheath protein	-	
"opCjV0100001020"	CJE0228	-	-	-	-	-	1	1	0	-1	1	-1	-1	1	0	-1	-	hypothetical protein	-	
"opCjV0100000959"	CJE0229	-	-	-	-	-	1	1	1	-1	-1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000217"	CJE0230	-	-	-	-	-	0	1	0	-1		-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV0100000593"	CJE0231	-	-	-	-	-	1	0	0	-1	1	-1	1	1	-1	-1	-	tail fiber protein H, putative	-	
"opCjV0100000426"	CJE0232	-	-	-	-	-	1	0	1	-1	1	-1	-1	1	1	-1	-	phage tail protein, putative	-	

"opCjV010000 0427"	CJE0233	-	-	-	-	-	0	0	0						-1	1		-	baseplate assembly protein J, putative	-
"opCjV010000 0033"	CJE0234	-	-	-	-	-	1	0	0	-1	1	-1	-1	1	1	1		-	baseplate assembly protein W, putative	-
"opCjV010000 0643"	CJE0235	-	-	-	-	-	1	1	0	-1	1	-1	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0541"	CJE0236	-	-	-	-	-	1	1	1	-1	1	-1	-1	1	1	-1		-	baseplate assembly protein V, putative	-
"opCjV010000 0173"	CJE0237	-	-	-	-	-	1	1	1	-1	0	-1	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0210"	CJE0238	-	-	-	-	-	1	1	0						1			-	hypothetical protein	-
"opCjV010000 0765"	CJE0239	-	-	-	-	-	1	1	1	-1	1	-1	-1	1	1	-1		-	lipoprotein, putative	-
"opCjV010000 0950"	CJE0240	-	-	-	-	-	1	1	1		-1	1	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0655"	CJE0241	-	-	-	-	-	1	1	1					-1	1			-	hypothetical protein	-
"opCjV010000 0798"	CJE0242	-	-	-	-	-	1	1	1	-1	1	-1	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0724"	CJE0243	-	-	-	-	-	1	1	1			1	1	-1	1			-	hypothetical protein	-
"opCjV010000 0443"	CJE0244	-	-	-	-	-	1	1	1		-1	1	-1	1	1	-1		-	Mu-like prophage I protein, putative	-
"opCjV010000 0269"	CJE0245	-	-	-	-	-	1	0	0	-1	1	-1	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0177"	CJE0246	-	-	-	-	-	1	1	1	-1	-1	1	1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0328"	CJE0247	-	-	-	-	-	1	1	0	-1		1	1	1	1	-1		-	hypothetical protein	-
"opCjV010000 1082"	CJE0248	-	-	-	-	-	1	1	0	0	1	0	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0909"	CJE0249	-	-	-	-	-	1	1	1	0	1	0	-1	1	1	-1		-	phage uncharacterized protein	-
"opCjV010000 0353"	CJE0250	-	-	-	-	-	1	1	1	-1	-1	1	-1	-1	1	1		-	hypothetical protein	-
"opCjV010000 0297"	CJE0251	-	-	-	-	-	-1	1	0	-1		1	1	-1	1	0		-	prophage MuSo1, F protein, putative	-
"opCjV010000 0585"	CJE0252	-	-	-	-	-	1	1	1		1	1	-1	1	1	-1		-	phage tail protein, putative	-
"opCjV010000 0667"	CJE0253	-	-	-	-	-	1	1	1	-1	1	-1	-1	1	1	-1		-	tail protein X, putative	-
"opCjV010000 0604"	CJE0254	-	-	-	-	-	-1	0	0	-1	1	-1	-1	1	1	-1		-	tail protein D, putative	-
"opCjV010000 0932"	CJE0255	-	-	-	-	-	0	1	0	-1	1	-1	-1	1	1	1		-	DNA-binding protein, putative	-
"opCjV010000 0957"	CJE0256	dns	-	-	-	-	1	1	0				-1	-1	1			-	extracellular deoxyribonuclease	-
"opCjV010000 0211"	CJE0257	-	-	-	-	-	1	1	0	-1	1	-1	-1	1	1	-1		-	hypothetical protein	-
"opCjV010000 0654"	CJE0258	-	-	-	-	-	0	1	0	-1	1	-1	-1	1	1	0		-	hypothetical protein	-
"opCjV010000 0071"	CJE0259	-	-	-	-	-	1	-1	0	-1	1	-1	-1	1	0	-1		-	hypothetical protein	-
"opCjV010000"	CJE0260	-	-	-	-	-	1	1	1	-1	1	-1	-1	1	1	1		-	hypothetical protein	-

Appendix

"0417"																				
"opCjV0100000431"	CJE0261	-	-	-	-	-	1	1	1	-1	1	-1	-1	-1	1	-1	-	hypothetical protein	-	
"opCjV0100000185"	CJE0262	-	-	-	-	-	-1	-1	0	-1	1	-1	-1	1	1	-1	-	conserved hypothetical protein TIGR01671	-	
"opCjV0100000791"	CJE0263	-	-	-	-	-	-1	-1	-1	-1	-1	0	-1	-1	1	-1	-	hypothetical protein	-	
"opCjV0100001084"	CJE0264	-	-	-	-	-	1	-1	-1	-1	-1	1	0	1	1	-1	-	hypothetical protein	-	
"opCjV0100000344"	CJE0265	-	-	-	-	-	1	1	1	-1	1	-1	-1	-1	1	-1	-	host-nuclease inhibitor protein Gam, putative	-	
"opCjV0100000376"	CJE0266	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	0	-1	-	hypothetical protein	-	
"opCjV0100000598"	CJE0267	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	0	-1	-	hypothetical protein	-	
"opCjV0100000580"	CJE0268	-	-	-	-	-	-1	-1	-1						-1		-	hypothetical protein	-	
"opCjV0100000034"	CJE0269	-	-	-	-	-	1	1	1					-1	1		-	bacteriophage DNA transposition protein B,	-	
"opCjV0100000708"	CJE0270	-	-	-	-	-	-1	-1	0	-1	1	-1	-1	1	0	-1	-	bacteriophage DNA transposition protein A,	-	
"opCjV0100001121"	CJE0271	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-	
"opCjV0100000076"	CJE0272	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	transcriptional regulator, putative	-	
"opCjV0100000632"	CJE0273	-	-	-	-	-	-1	-1	0	-1	1	-1	1	1	-1	-1	-	hypothetical protein	-	
"opCcV0100000484"	CJE0275	argC	Cj0224	argC	CCO0299	argC	1	1	1	1	1	1	1	0	1	1	N-acetyl-gamma-glutamyl-phosphate reductase	N-acetyl-gamma-glutamyl-phosphate reductase	N-acetyl-gamma-glutamyl-phosphate reductase	
"opCcV0100001416"	CJE0276	-	Cj0225	-	CCO0300	-	1	1	1	-1	-1	1	1	-1	1	-1	putative acetyltransferase	acetyltransferase, GNAT family	probable acetyltransferase Cj0225	
"opCcV0100000716"	CJE0277	argB	Cj0226	argB	CCO0301	argB	1	-1	0	0	1	0	-1	1	-1	1	acetylglutamate kinase	acetylglutamate kinase	acetylglutamate kinase	
"opCcV0100001452"	CJE0278	argD	Cj0227	argD	CCO0302	-	1	1	0	1	1	0	1	0	1	1	acetylnornithine aminotransferase	acetylnornithine aminotransferase	acetylnornithine transaminase Cj0227	
"CJ_10001433"	CJE0279	pem	Cj0228c	pem	CCO0303	pem	1	1	1	1	1	1	1	1	1	1	protein-L-isoaspartate O-methyltransferase	protein-L-isoaspartate O-methyltransferase	protein-L-isoaspartate O-methyltransferase	
"CJ_10001434"	CJE0280	-	Cj0229	-	CCO0304	-	1	1	1	1	1	1	1	1	1	1	putative acetyltransferase	transferase, hexapeptide repeat family	carbonic anhydrase, family 3 VC0058	
"CJ_10001435"	CJE0281	-	Cj0230c	-	CCO0305	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	nicotinate phosphoribosyltransferase	nicotinate phosphoribosyltransferase, putative	
"CJ_10001436"	CJE0282	nrdB	Cj0231c	nrdB	CCO0306	-	1	1	1	1	1	1	1	1	1	1	ribonucleoside-diphosphate reductase beta chain	ribonucleotide-diphosphate reductase beta	ribonucleoside-diphosphate reductase, beta	
"CJ_10001437"	CJE0283	-	Cj0232c	-	CCO0307	-	1	1	1	1	0	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0232c	
"CJ_10001438"	CJE0284	pyrE	Cj0233c	pyrE	CCO0308	pyrE	1	1	1	1	1	1	1	1	1	1	putative orotate phosphoribosyltransferase	orotate phosphoribosyltransferase	orotate phosphoribosyltransferase	
"CJ_10001449"	CJE0285	frr	Cj0234c	frr	CCO0309	frr	0	1	1	1	0	1	1	1	1	1	ribosome recycling factor	ribosome releasing factor	ribosome recycling factor	
"CJ_10001450"	CJE0286	secG	Cj0235c	secG	CCO0310	secG	1	1	1	1	0	1	1	1	1	1	putative protein-export membrane protein	protein-export membrane protein	protein translocation protein, low temperature	
"CJ_10001451"	CJE0287	-	Cj0236c	-	CCO0311	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0236c	
"CJ_10001452"	CJE0288	cynT	Cj0237	cynT	CCO0312	-	1	1	1	1	1	1	1	1	1	1	carbonic anhydrase	carbonic anhydrase	Carbonic anhydrase	
"CJ_10001453"	CJE0289	-	Cj0238	-	CCO0313	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane	mechanosensitive ion	probable integral membrane	

"CJ_10001454"	CJE0290	-	Cj0239c	-	CCO0314	-	1	1	1	1	1	1	1	1	1	1	protein	channel family protein	protein Cj0238
"CJ_10001455"	CJE0291	-	Cj0240c	-	CCO0315	-	1	1	1	1	1	1	1	1	1	1	nifU protein homolog	NifU family protein	nifU protein homolog Cj0239c
"CJ_10001456"	CJE0292	-	Cj0241c	-	-	-	0	1	1	1	1	1	1	1	1	1	putative aminotransferase (nifS protein)	cysteine desulfurase	cysteine desulfurase
"CJ_10001457"	CJE0293	-	Cj0243c	-	CCO0316	-	0	1	0	1	1	1	0	1	1	1	putative iron-binding protein	pseudogene	-
"CJ_10001458"	CJE0294	rpmI	Cj0244	rpmI	CCO0317	rpmI	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L35	50S ribosomal protein L35	ribosomal protein L35
"CJ_10001469"	CJE0295	rplT	Cj0245	rplT	CCO0318	rplT	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L20	50S ribosomal protein L20	ribosomal protein L20
"CJ_10001470"	CJE0296	-	Cj0246c	-	-	-	1	1	1	1	-1	1	1	0	1	1	putative MCP-domain signal transduction protein	hypothetical protein	-
"CJ_10001471"	CJE0297	-	Cj0247c	-	-	-	0	1	0	1	1	1	-1	1	1	0	hypothetical protein	pseudogene	-
"CJ_10001472"	CJE0298	-	Cj0248	-	CCO0319	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001474"	CJE0299	-	Cj0249	-	CCO0320	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001476"	CJE0300	-	Cj0250c	-	CCO0255	proP	0	0	0	1	1	1	0	0	1	1	putative transmembrane transport protein	major facilitator superfamily protein	proline/betaine transporter (proP)
"CJ_10001477"	CJE0301	-	Cj0251c	-	CCO0323	-	1	1	1	1	0	1	1	1	1	1	Highly acidic protein	hypothetical protein	hypothetical protein
"CJ_10001479"	CJE0302	moaC	Cj0252	moaC	CCO0324	moaC	1	1	1	1	1	1	0	1	1	1	molybdenum cofactor biosynthesis protein C	molybdenum cofactor biosynthesis protein C	molybdenum cofactor biosynthesis protein C
"CJ_10001480"	CJE0303	-	Cj0253	-	CCO0325	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001482"	CJE0304	-	Cj0254	-	CCO0326	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001498"	CJE0305	xth	Cj0255c	-	CCO0327	xth	1	1	0	1	1	1	1	1	1	1	exodeoxyribonuclease	exodeoxyribonuclease III	exodeoxyribonuclease III
"CJ_10001499"	CJE0306	-	Cj0256	-	CCO0328	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	sulfatase, putative	membrane protein, putative
"CJ_10001500"	CJE0307	dgkA	Cj0257	dgkA	CCO0329	-	-1	1	1	1	1	1	1	1	1	1	diacylglycerol kinase	diacylglycerol kinase	diacylglycerol kinase Cj0257
"CJ_10001501"	CJE0308	-	Cj0258	-	CCO0330	-	1	1	1	1	0	1	1	1	1	1	Putative helix turn helix motif protein	hypothetical protein	conserved hypothetical protein
"CJ_10001503"	CJE0309	pyrC	Cj0259	pyrC	CCO0331	pyrC	1	1	1	1	-1	1	1	-1	1	1	dihydroorotase	dihydroorotase	dihydroorotase, homodimeric type
"opCjV010000 0803"	CJE0310	-	-	-	-	-	-1	-1	-1	1	1	0	-1	1	1	1	-	D12 class N6 adenine- specific DNA	-
"CJ_10001506"	CJE0311	-	Cj0261c	-	CCO0333	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001510"	CJE0313	-	Cj0263	-	CCO0282	bisZ	1	1	1	1	1	1	1	1	1	-1	putative integral membrane protein	zinc transporter ZupT	biotin sulfoxide reductase VC1950
"CJ_10000543"	CJE0314	-	Cj0144	-	CCO1733	-	1	1	1	0	1	1	1	1	1	0	methyl-accepting chemotaxis signal transduction	methyl-accepting chemotaxis protein	methyl-accepting chemotaxis protein (t1pA)
"CJ_10001535"	CJE0315	-	Cj0266c	-	CCO0335	-	-1	-1	-1	-1	-1	-1	-1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0266c
"CJ_10001536"	CJE0316	-	Cj0267c	-	CCO0336	-	1	1	0	1	0	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0267c
"CJ_10001537"	CJE0317	-	Cj0268c	-	CCO0337	-	1	1	1	1	1	1	1	1	1	1	putative transmembrane protein	SPFH domain / Band 7 family protein	probable transmembrane protein Cj0268c
"CJ_10001538"	CJE0318	ilvE	Cj0269c	ilvE	CCO0338	ilvE	1	1	1	1	0	1	1	1	1	1	branched-chain amino acid aminotransferase	branched-chain amino acid aminotransferase	branched-chain amino acid aminotransferase
"CJ_10001539"	CJE0319	-	Cj0270	-	CCO0339	-	1	1	1	1	0	1	1	1	1	1	Possible isomerase	4-oxalocrotonate tautomerase family protein	4-oxalocrotonate tautomerase (dmpI)-related

"CJ_10001541"	CJE0320	-	Cj0271	-	CCO0340	-	1	1	1	1	1	1	-1	-1	-1	1	bacterioferritin comigratory protein homolog	antioxidant, AhpC/Tsa family	bacterioferritin comigratory protein homolog
"CJ_10001543"	CJE0321	-	Cj0272	-	CCO0341	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001545"	CJE0322	fabZ	Cj0273	fabZ	CCO0342	fabZ	1	1	1	1	1	1	1	1	1	1	(3R)-hydroxymyristoyl-[acyl carrier protein]	(3R)-hydroxymyristoyl ACP dehydratase	beta-hydroxyacyl-(acyl-carrier-protein)
"CJ_10001547"	CJE0323	lpxA	Cj0274	lpxA	CCO0343	lpxA	1	1	1	1	1	1	0	1	1	1	acyl-[acyl-carrier-protein]--UDP-N-	UDP-N-acetylglucosamine acyltransferase	acyl-[acyl-carrier-protein]--UDP-N-
"CJ_10001563"	CJE0324	clpX	Cj0275	clpX	CCO0344	clpX	1	1	1	1	1	1	1	1	1	1	ATP-dependent clp protease ATP-binding subunit	ATP-dependent protease ATP-binding subunit	ATP-dependent Clp protease, ATP-binding subunit
"CJ_10001564"	CJE0325	mreB	Cj0276	mreB	CCO0345	-	1	1	1	1	1	1	1	1	1	1	homolog of E. coli rod shape-determining	cell shape-determining protein MreB	rod shape-determining protein (mreB)
"CJ_10001565"	CJE0326	-	Cj0277	-	CCO0346	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	rod shape-determining protein MreC	rod shape-determining protein (mreC), putative
"CJ_10001566"	CJE0327	carB	Cj0279	carB	CCO0357	carB	1	1	1	1	1	1	1	1	1	1	carbamoyl-phosphate synthase large chain	carbamoyl-phosphate synthase large subunit	carbamoyl-phosphate synthase, large subunit
"CJ_10001567"	CJE0328	-	Cj0280	-	CCO0358	-	-1	-1	-1	-1	1	-1	1	1	1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001569"	CJE0329	tal	Cj0281c	tal	CCO0359	tal	1	1	1	1	1	1	1	1	1	1	putative transaldolase	transaldolase	transaldolase
"CJ_10001571"	CJE0330	serB	Cj0282c	serB	CCO0360	serB	1	1	1	1	1	1	1	1	1	1	putative phosphoserine phosphatase	phosphoserine phosphatase SerB	phosphoserine phosphatase SerB
"CJ_10001572"	CJE0331	cheW	Cj0283c	cheW	CCO0361	-	1	1	1	1	1	1	1	1	1	1	chemotaxis protein	purine-binding chemotaxis protein CheW	chemotaxis protein Cj0283c
"CJ_10001574"	CJE0332	cheA	Cj0284c	cheA	CCO0362	cheA	1	1	1	1	1	1	1	1	1	1	chemotaxis histidine kinase	chemotaxis protein CheA	histidine kinase (cheA)
"CJ_10001576"	CJE0333	cheV	Cj0285c	cheV	CCO0363	-	1	1	1	1	1	1	1	1	1	1	chemotaxis protein	chemotaxis protein CheV	chemotaxis protein (cheV)
"CJ_10000002"	CJE0334	-	Cj0286c	-	CCO0364	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000004"	CJE0335	greA	Cj0287c	greA	CCO0365	-	-1	-1	0	1	1	1	1	-1	-1	-1	transcription elongation factor	transcription elongation factor GreA	transcription elongation factor Cj0287c
"CJ_10000006"	CJE0336	lpxB	Cj0288c	lpxB	CCO0366	lpxB	0	1	1	1	0	1	1	1	-1	1	lipid-A-disaccharide synthase	lipid-A-disaccharide synthase	lipid-A-disaccharide synthase
"CJ_10000008"	CJE0337	-	Cj0289c	peb3	-	-	1	1	1	1	1	1	1	1	1	1	major antigenic peptide PEB3	major antigenic peptide PEB3	-
"CJ_10000009"	CJE0338	-	Cj0291c	-	CCO0371	-	1	0	0	1	1	-1	1	1	1	-1	glycerol-3-phosphate transporter (possible)	pseudogene	phosphoglycerate transporter protein pgtP
"CJ_10000010"	CJE0339	-	Cj0291c	-	CCO0372	-	1	1	1	1	1	1	1	0	1	1	glycerol-3-phosphate transporter (possible)	pseudogene	phosphoglycerate transporter protein pgtP
"CJ_10000011"	CJE0340	-	Cj0291c	-	CCO0373	-	1	1	1	1	1	-1	1	1	1	1	glycerol-3-phosphate transporter (possible)	pseudogene	phosphoglycerate transporter protein pgtP
"CJ_10000012"	CJE0341	-	Cj0291c	-	CCO0374	-	0	1	1	1	1	1	1	1	1	1	glycerol-3-phosphate transporter (possible)	pseudogene	phosphoglycerate transporter protein pgtP
"CJ_10000030"	CJE0342	panC	Cj0297c	panC	CCO0380	panC	0	0	-1	1	1	-1	-1	1	1	1	pantoate--beta-alanine ligase	pantoate--beta-alanine ligase	pantoate--beta-alanine ligase
"CJ_10000032"	CJE0343	panB	Cj0298c	panB	CCO0381	panB	1	1	-1	1	1	-1	-1	1	1	1	3-methyl-2-oxobutanoate	3-methyl-2-oxobutanoate	3-methyl-2-oxobutanoate
"CJ_10000034"	CJE0344	-	Cj0299	-	-	-	1	1	1	1	1	-1	1	1	1	1	putative periplasmic beta-lactamase	beta-lactamase	-
"CJ_10000035"	CJE0345	modC	Cj0300c	modC	CCO0385	modC	1	1	1	1	-1	1	1	-1	1	1	putative molybdenum transport ATP-binding	molybdenum ABC transporter, ATP-binding protein	molybdenum ABC transporter, ATP-binding protein
"CJ_10000036"	CJE0346	modB	Cj0301c	modB	CCO0386	-	1	1	1	1	-1	1	1	1	1	1	putative molybdenum transport system permease	molybdenum ABC transporter, permease protein	molybdenum ABC transporter, permease protein
"CJ_10000038"	CJE0347	-	Cj0302c	-	CCO0387	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	TOBE domain protein	molybdenum-pterin binding domain protein,

"CJ_1000039"	CJE0348	modA	Cj0303c	modA	CCO0388	modA	1	1	1	1	1	1	-1	-1	-1	1	putative molybdate-binding lipoprotein	molybdenum ABC transporter, periplasmic	molybdenum ABC transporter, periplasmic
"CJ_1000040"	CJE0349	-	Cj0304c	bioC	CCO0389	-	1	1	1	1	-1	1	1	-1	1	1	putative biotin synthesis protein	biotin biosynthesis protein BioC	biotin synthesis protein BioC, putative
"CJ_1000042"	CJE0350	-	Cj0305c	-	CCO0390	-	0	1	1	1	-1	1	1	0	1	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF452)
"CJ_1000049"	CJE0351	bioF	Cj0306c	bioF	CCO0391	-	0	1	1	1	-1	1	1	-1	1	-1	8-amino-7-oxononanoate synthase	8-amino-7-oxononanoate synthase	8-amino-7-oxononanoate synthase
"CJ_1000051"	CJE0352	bioA	Cj0307	bioA	CCO0393	bioA	-1	1	1	1	1	1	1	1	0	1	adenosylmethionine-8-amino-7-oxononanoate	adenosylmethionine--8-amino-7-oxononanoate	adenosylmethionine--8-amino-7-oxononanoate
"CJ_1000053"	CJE0353	bioD	Cj0308c	bioD	CCO0394	-	-1	1	0	1	1	1	1	1	1	1	putative dethiobiotin synthetase	dethiobiotin synthetase	probable dethiobiotin synthase Cj0308c
"CJ_1000055"	CJE0354	-	Cj0309c	-	CCO0395	-	1	1	0	1	1	1	1	0	1	1	putative efflux protein	multidrug resistance protein, SMR family	probable efflux protein Cj0309c
"CJ_1000056"	CJE0355	-	Cj0310c	-	CCO0396	-	-1	1	1	1	0	1	1	1	1	1	putative efflux protein	multidrug resistance protein, SMR family	probable efflux protein Cj0310c
"CJ_1000057"	CJE0356	rplY	Cj0311	-	CCO0397	-	1	0	1	1	1	1	1	1	1	1	ctc protein homolog	50S ribosomal protein L25	ribosomal 5S rRNA E-loop binding protein
"CJ_1000058"	CJE0357	pth	Cj0312	pth	CCO0398	pth	1	1	1	1	1	1	1	1	1	1	peptidyl-tRNA hydrolase	peptidyl-tRNA hydrolase	peptidyl-tRNA hydrolase
"CJ_1000059"	CJE0358	-	Cj0313	-	CCO0399	-	0	1	1	1	1	1	1	0	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0313
"CJ_1000060"	CJE0359	lysA	Cj0314	lysA	CCO0400	lysA	0	1	1	1	1	1	1	1	1	1	diaminopimelate decarboxylase	diaminopimelate decarboxylase	diaminopimelate decarboxylase
"CJ_1000061"	CJE0360	-	Cj0315	-	CCO0401	-	0	1	1	1	0	1	1	0	1	1	hypothetical protein	HAD-superfamily hydrolase, subfamily IIA	HAD-superfamily hydrolase, subfamily IIA
"CJ_1000069"	CJE0361	pheA	Cj0316	pheA	CCO0402	-	0	1	1	1	1	1	1	1	1	1	chorismate mutase/prephenate dehydratase	chorismate mutase/prephenate dehydratase	chorismate mutase / prephenate dehydratase
"CJ_1000071"	CJE0362	hisC	Cj0317	hisC	CCO0403	hisC	1	1	1	1	1	1	1	1	1	1	probable histidinol-phosphate aminotransferase	histidinol-phosphate aminotransferase	histidinol-phosphate aminotransferase
"CJ_1000073"	CJE0363	fliF	Cj0318	fliF	CCO0404	fliF	0	-1	1	1	1	1	1	1	-1	1	flagellar M-ring protein	flagellar M-ring protein	flagellar M-ring protein FliF
"CJ_1000075"	CJE0364	fliG	Cj0319	fliG	CCO0405	fliG	1	0	1	1	1	1	1	1	0	1	flagellar motor switch protein	flagellar motor protein	flagellar motor switch protein FliG
"CJ_1000076"	CJE0365	-	Cj0320	fliH	CCO0406	fliH	0	1	1	1	1	1	1	1	-1	1	putative flagellar assembly protein	flagellar assembly protein	flagellar export protein (fliH)
"CJ_1000077"	CJE0366	dxs	Cj0321	dxs	CCO0407	dxs	1	1	1	1	1	1	1	1	1	1	1-deoxyxylulose-5-phosphate synthase	1-deoxy-D-xylulose-5-phosphate synthase	deoxyxylulose-5-phosphate synthase
"CJ_1000078"	CJE0367	-	Cj0322	perR	CCO0408	furR1	1	-1	1	1	1	1	1	1	0	1	peroxide stress regulator	transcriptional regulator, Fur family	transcriptional regulator, Fur family
"CJ_1000079"	CJE0368	-	Cj0323	-	CCO0409	-	0	-1	0	1	1	1	1	1	-1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_1000080"	CJE0369	ubiE	Cj0324	ubiE	CCO0410	ubiE	1	1	1	1	1	1	1	1	1	1	ubiquinone/menaquinone biosynthesis	ubiquinone/menaquinone biosynthesis	gerC2 protein (gerC2)
"CJ_1000081"	CJE0370	xseA	Cj0325	xseA	CCO0414	xseA	1	1	1	1	1	1	1	1	1	1	exodeoxyribonuclease VII large subunit	exodeoxyribonuclease VII, large subunit	exodeoxyribonuclease VII, large subunit
"CJ_10000619"	CJE0371	serC	Cj0326	serC	CCO0415	serC	1	1	1	1	1	1	1	1	1	1	phosphoserine aminotransferase	phosphoserine aminotransferase	phosphoserine aminotransferase
"CJ_10000620"	CJE0372	-	Cj0327	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	endoribonuclease L-PSP family protein	-
"CJ_10000621"	CJE0373	fabH	Cj0328c	fabH	CCO0416	fabH	1	1	1	1	0	1	1	1	1	1	3-oxoacyl-[acyl-carrier-protein] synthase	3-oxoacyl-(acyl carrier protein) synthase	beta-ketoacyl- <i>acp</i> synthase III
"CJ_10000622"	CJE0374	plsX	Cj0329c	plsX	CCO0417	plsX	1	1	1	1	1	1	1	1	1	1	putative fatty acid/phospholipid synthesis	fatty acid/phospholipid synthesis protein	fatty acid/phospholipid synthesis protein PlsX
"opCεV0100001244"	CJE0375	rpmF	Cj0330c	rpmF	CCO0418	rpmF	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L32	50S ribosomal protein L32	ribosomal protein L32

"CJ_10000623"	CJE0376	-	Cj0331c	-	CCO0419	-	1	1		1	1		0		1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000624"	CJE0377	ndk	Cj0332c	ndk	CCO0420	-	1	1	-1	-1	1	1	1	-1	1	1	nucleoside diphosphate kinase	nucleoside diphosphate kinase	nucleoside diphosphate kinase Cj0332c
"opCcV0100000143"	CJE0378	-	Cj0333c	fdxA	CCO0421	-	1	1	1	1	1	1	1	1	1	1	ferredoxin	ferredoxin, 4Fe-4S	ferredoxin
"CJ_10000627"	CJE0379	-	Cj0334	ahpC	CCO0422	-	1	1	1	1	1	1	1	1	1	1	alkyl hydroperoxide reductase	antioxidant, AhpC/Tsa family	antioxidant, AhpC/Tsa family VC0731
"CJ_10000629"	CJE0380	flhB	Cj0335	flhB	CCO0423	flhB	1	1	1	1	1	1	1	1	1	1	flagellar biosynthetic protein	flagellar biosynthetic protein FlhB	flagellar biosynthetic protein FlhB
"CJ_10000647"	CJE0381	-	Cj0336c	motB	CCO0424	-	1	1	0	1	1	1	1	0	1	1	putative flagellar motor protein	flagellar motor protein	chemotaxis motB protein, putative
"CJ_10000648"	CJE0382	-	Cj0337c	motA	CCO0425	-	1	1	1	1	1	1	1	1	1	1	putative flagellar motor proton channel	flagellar motor protein	probable flagellar motor proton channel Cj0337c
"CJ_10000649"	CJE0383	polA	Cj0338c	polA	CCO0426	polA	1	1	1	1	1	1	1	1	1	1	DNA polymerase I	DNA polymerase I	DNA polymerase I (polA)
"CJ_10000650"	CJE0384	-	Cj0339	-	-	-	0	1	1	1	1	1	0	1	1	1	putative transmembrane transport protein	major facilitator family transporter	-
"CJ_10000651"	CJE0385	-	Cj0340	-	-	-	1	1	1	1	1	1	1	1	1	1	putative nucleoside hydrolase	inosine-uridine preferring nucleoside hydrolase	-
"CJ_10000652"	CJE0386	-	Cj0341c	-	CCO0427	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	membrane protein, putative
"opCjV0100000999"	CJE0387	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV0100000510"	CJE0388	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV0100000162"	CJE0389	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"CJ_10000653"	CJE0390	uvrA	Cj0342c	uvrA	CCO0431	uvrA	1	1	1	1	1	1	1	1	1	1	excinuclease ABC subunit A	excinuclease ABC subunit A	excinuclease ABC, A subunit
"CJ_10000655"	CJE0391	-	Cj0343c	-	CCO0432	-	1	0	0	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	conserved hypothetical integral membrane
"opCjV010000041"	CJE0392	-	Cj0344	-	-	-	0	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"opCjV0100000687"	CJE0393	-	-	-	-	-	1	1	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_10000658"	CJE0394	trpE	Cj0345	trpE	CCO0433	-	1	1	1	1	1	1	1	1	1	1	putative anthranilate synthase component I	anthranilate synthase component I	anthranilate synthase component I
"CJ_10000676"	CJE0395	trpD	Cj0346	trpD	CCO0434	-	1	1	1	1	1	1	1	1	1	1	anthranilate synthase component II	anthranilate synthase component II	anthranilate phosphoribosyltransferase,
"CJ_10000677"	CJE0396	trpF	Cj0347	trpF	CCO0435	trpF	1	1	1	1	1	1	1	1	1	1	N-(5'-phosphoribosyl)anthranilate isomerase	N-(5'phosphoribosyl)anthranilate isomerase	N-(5'phosphoribosyl)anthranilate isomerase
"CJ_10000678"	CJE0397	trpB	Cj0348	trpB	CCO0436	trpB	0	0	1	1	1	1	1	1	1	1	tryptophan synthase beta chain	tryptophan synthase subunit beta	tryptophan synthase, beta subunit
"CJ_10000679"	CJE0398	trpA	Cj0349	trpA	CCO0437	trpA	1	0	1	1	1	1	1	1	1	1	tryptophan synthase alpha chain	tryptophan synthase, alpha subunit	tryptophan synthase, alpha subunit
"CJ_10000680"	CJE0399	-	Cj0350	-	CCO0438	-	0	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000681"	CJE0400	fliN	Cj0351	fliN	CCO0439	-	1	1	1	1	1	1	1	1	1	1	flagellar motor switch protein	flagellar motor switch protein	flagellar switch protein
"CJ_10000682"	CJE0401	-	Cj0352	-	CCO0440	-	1	0	1	1	1	1	1	1	1	1	putative transmembrane protein	hypothetical protein	probable transmembrane protein Cj0352
"CJ_10000684"	CJE0402	-	Cj0353c	-	CCO0441	gppA	1	1	1	1	1	1	1	1	1	1	phosphatase	phosphatase, Ppx/GppA family	guanosine pentaphosphate phosphohydrolase
"CJ_10000686"	CJE0403	-	Cj0354c	fdxB	CCO0442	-	0	0	1	1	1	1	1	1	1	1	putative ferredoxin	ferredoxin, 4Fe-4S	ferredoxin

"CJ_10000688"	CJE0404	-	Cj0355c	-	CCO0443	-	1	1	1	1	1	1	1	1	1	1	two-component regulator	DNA-binding response regulator	response regulator
"CJ_10000705"	CJE0405	-	Cj0356c	-	CCO0444	folB	0	0	1	1	1	1	1	1	1	1	hypothetical protein	folB domain protein	dihydroneopterin aldolase
"CJ_10000706"	CJE0406	-	Cj0357c	-	CCO0445	-	0	1	0	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	conserved hypothetical protein TIGR00023
"CJ_10000707"	CJE0407	-	Cj0358	-	CCO0446	mauG	1	1	1	1	1	1	1	1	1	1	putative cytochrome C551 peroxidase	cytochrome c551 peroxidase	cytochrome c551 peroxidase
"opCcV0100001679.60"	CJE0408	-	-	-	-	-	1	1	0	-1	-1	-1	1	1	-1	1	-	hypothetical protein	-
"CJ_10000708"	CJE0409	glmM	Cj0360	-	CCO0447	-	1	1	1	1	1	1	1	1	1	1	putative phospho-sugar mutase	phosphoglucosamine mutase	phosphoglucosamine mutase
"CJ_10000709"	CJE0410	glmM	Cj0361	-	CCO0448	mrsA	0	1	1	1	1	1	1	1	1	1	putative phospho-sugar mutase	phosphoglucosamine mutase	PUTATIVE PHOSPHOSUGAR ISOMERASE PROTEIN
"CJ_10000710"	CJE0411	glmM	Cj0362	-	CCO0449	mrsA	0	1	1	1	1	1	1	0	1	1	putative phospho-sugar mutase	phosphoglucosamine mutase	phosphoglucosamine mutase
"CJ_10000712"	CJE0412	-	Cj0363c	-	CCO0452	-	1	1	0	1	1	1	1	-1	1	1	putative oxidoreductase	coproporphyrinogen III oxidase	oxygen-independent coproporphyrinogen III
"CJ_10000714"	CJE0413	-	Cj0364	-	CCO0453	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCcV0100001459"	CJE0414	cmeC	Cj0365c	-	CCO0454	-	1	1	1	1	0	0	0	0	1	1	putative outer membrane channel protein	RND efflux system, outer membrane lipoprotein	multidrug efflux pump channel protein, putative
"opCcV0100000610"	CJE0415	cmeB	Cj0366c	-	CCO0455	-			0	-1	1	1	-1	1	-1	-1	transmembrane efflux protein	RND efflux system, inner membrane transporter	transmembrane efflux protein Cj0366c
"opCcV0100001785"	CJE0416	cmeA	Cj0367c	-	CCO0456	-	1	1	0	-1	1	1	-1	1	1	-1	putative membrane fusion component of efflux	RND efflux system, membrane fusion protein CmeA	HlyD family secretion protein, putative
"CJ_10000733"	CJE0417	-	Cj0368c	-	CCO0457	-	0	1	1	1	1	1	1	1	1	1	transcriptional regulatory protein	transcriptional regulator, TetR family	transcriptional regulator, tetR family,
"CJ_10000734"	CJE0418	-	Cj0369c	-	CCO0458	-	0	0	1	1	1	1	1	1	1	1	ferredoxin domain-containing integral membrane	iron-sulfur cluster-binding domain protein	ferredoxin-like protein
"CJ_10000735"	CJE0419	rpsU	Cj0370	rpsU	CCO0459	-	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S21	30S ribosomal protein S21	ribosomal protein S21
"CJ_10000736"	CJE0420	-	Cj0371	-	CCO0460	-	1	1	1	1	1	1	1	1	1	1	putative flagellar motility protein	hypothetical protein	probable flagellar motility protein Cj0371
"CJ_10000737"	CJE0421	-	Cj0372	-	CCO0461	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	glutathionylspermidine synthase family protein	glutathionylspermidine synthase family protein
"CJ_10000738"	CJE0422	-	Cj0373	-	CCO0462	hprA	1	1	1	1	1	1	1	1	1	1	putative D-2-hydroxyacid dehydrogenase	2-hydroxyacid dehydrogenase	phosphoglycerate dehydrogenase
"CJ_10000739"	CJE0423	-	Cj0374	-	CCO0463	-	0	1	1	1	1	1	1	0	1	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF520)
"CJ_10000740"	CJE0424	-	Cj0375	-	CCO0464	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0375
"CJ_10000741"	CJE0425	-	Cj0376	-	CCO0465	-	1	1	1	1	0	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0376
"CJ_10000755"	CJE0426	-	Cj0377	-	CCO0466	-	1	0	1	1	0	1	1	1	1	1	probable AAA family ATPase	ATPase, AAA family protein	probable AAA family ATPase Cj0377
"CJ_10000757"	CJE0427	-	Cj0378c	-	CCO0467	-	1	1	1	1	0	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0378c
"CJ_10000759"	CJE0428	-	Cj0379c	-	CCO0468	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000761"	CJE0429	-	Cj0380c	-	CCO0469	-	1	1	1	1	1	-1	-1	-1	0	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000763"	CJE0430	pyrF	Cj0381c	pyrF	CCO0470	pyrF		1	1	1	-1	1	1	1	1	1	orotidine 5'-phosphate decarboxylase	orotidine 5'-phosphate decarboxylase	orotidine 5'-phosphate decarboxylase
"CJ_10000765"	CJE0431	nusB	Cj0382c	nusB	CCO0471	nusB	1	1	1	1	1	-1	-1	1	-1	-1	transcription termination	transcription	transcription antitermination

																	protein	antitermination protein NusB	factor NusB
"CJ_10000767"	CJE0432	ribH	Cj0383c	ribH	CCO0472	ribH	0	1	1	1	-1	-1	1	1		1	6,7-dimethyl-8-ribityllumazine synthase	riboflavin synthase subunit beta	6,7-dimethyl-8-ribityllumazine synthase
"CJ_10000768"	CJE0433	kdsA	Cj0384c	kdsA	CCO0473	kdsA	1	1	1	1	-1	1	1	1	1	1	2-dehydro-3-deoxyphosphoactonate aldolase	2-dehydro-3-deoxyphosphoactonate aldolase	2-dehydro-3-deoxyphosphoactonate aldolase
"CJ_10000769"	CJE0434	-	Cj0385c	-	CCO0474	-	1	1	1	0	-1	1	1	0	-1	1	putative integral membrane protein	integral membrane protein	probable integral membrane protein Cj0385c
"CJ_10000770"	CJE0435	-	Cj0386	-	CCO0475	-	-1	1	1	1	-1	1	1	1	1	-1	putative GTP-binding protein	GTP-binding protein EngA	GTPase
"CJ_10000790"	CJE0436	aroK	Cj0387	aroK	CCO0476	aroK	1	1	1	1	1	1	0	1	1	1	shikimate kinase	shikimate kinase	shikimate kinase
"CJ_10000793"	CJE0437	trpS	Cj0388	trpS	CCO0477	trpS	0	1	1	1	-1	1	1	-1	-1	1	tryptophanyl-tRNA synthetase	tryptophanyl-tRNA synthetase	tryptophanyl-tRNA synthetase
"CJ_10000796"	CJE0438	serS	Cj0389	serS	CCO0478	serS	1	1	1	1	0	1	-1	-1	1	1	seryl-tRNA synthetase	seryl-tRNA synthetase	seryl-tRNA synthetase
"CJ_10000798"	CJE0439	-	Cj0390	-	CCO0479	-	1	1	1	1	-1	-1	-1	1	1	1	putative transmembrane protein	TPR domain protein	probable transmembrane protein Cj0390
"CJ_10000800"	CJE0440	-	Cj0391c	-	CCO0480	-	0	1	1	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000802"	CJE0441	pyk	Cj0392c	pyk	CCO0481	pyk	0	0	1	1	0	1	1	1	1	1	pyruvate kinase	pyruvate kinase	pyruvate kinase
"CJ_10000804"	CJE0442	-	Cj0393c	-	CCO0482	-	1	1	1	-1	-1	1	1	1	1	-1	putative oxidoreductase	malate:quinone oxidoreductase, putative	probable oxidoreductase Cj0393c
"CJ_10000806"	CJE0443	-	Cj0394c	-	CCO0483	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	transcriptional activator, putative, Baf family	transcriptional activator, putative, Baf family
"CJ_10000807"	CJE0444	-	Cj0395c	-	CCO0484	-	1	1	1	-1	-1	1	1	1	1	1	hypothetical protein	tram-like protein	conserved hypothetical protein
"CJ_10000808"	CJE0445	-	Cj0396c	-	CCO0485	-	1	1	1	1	-1	1	1	-1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0396c
"CJ_10000819"	CJE0446	-	Cj0397c	-	CCO0486	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000822"	CJE0447	gatC	Cj0398	gatC	CCO0487	gatC	1	1	1	1	1	1	1	1	1	1	putative Glu-tRNA ^{Gln} amidotransferase subunit C	aspartyl/glutamyl-tRNA amidotransferase subunit	glutamyl-tRNA(Gln) amidotransferase, C subunit
"CJ_10000825"	CJE0448	-	Cj0399	-	CCO0488	cvpA	1	1	1	1	-1	1	1	-1	1	1	putative integral membrane protein	cvpA family protein	CvpA family protein
"CJ_10000828"	CJE0449	fur	Cj0400	fur	CCO0489	-	1	1	1	1	1	1	1	1	1	1	ferric uptake regulator	ferric uptake regulation protein	ferric uptake regulator Cj0400
"opCcV0100001338"	CJE0450	lysS	Cj0401	lysS	CCO0490	lysS	1	1	1	0	1	-1	-1	1	-1	-1	lysyl-tRNA synthetase	lysyl-tRNA synthetase	lysyl-tRNA synthetase
"opCcV0100000067"	CJE0451	glyA	Cj0402	glyA	CCO0491	glyA	-1	0	0	-1	1	1	0	1	-1	1	serine hydroxymethyltransferase	serine hydroxymethyltransferase	serine hydroxymethyltransferase
"opCcV0100001753"	CJE0452	-	Cj0403	-	CCO0492	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000833"	CJE0453	-	Cj0404	-	CCO0493	-	1	1	1	-1	-1	-1	1	1	1	-1	putative transmembrane protein	hypothetical protein	probable transmembrane protein Cj0404
"CJ_10000834"	CJE0454	aroE	Cj0405	aroE	CCO0494	aroE	0	0	1	-1	-1	1	1	1	1	1	shikimate 5-dehydrogenase	shikimate 5-dehydrogenase	shikimate 5-dehydrogenase
"CJ_10000835"	CJE0455	-	Cj0406c	-	CCO0495	-	1	1	1	1	-1	1	1	-1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0406c
"CJ_10000846"	CJE0456	lgt	Cj0407	lgt	CCO0496	lgt	1	1	1	1	1	1	1	1	1	1	prolipoprotein diacylglyceryl transferase	prolipoprotein diacylglyceryl transferase	prolipoprotein diacylglyceryl transferase
"CJ_10000849"	CJE0457	frdC	Cj0408	frdC	CCO0497	-	1	1	1	-1	-1	1	1	-1	-1	1	fumarate reductase cytochrome B subunit	fumarate reductase, cytochrome b subunit	Fumarate reductase respiratory complex,
"CJ_10000852"	CJE0458	frdA	Cj0409	frdA	CCO0498	-	1	1	1	1	-1	1	1	1	1	1	fumarate reductase flavoprotein subunit	fumarate reductase	succinate dehydrogenase flavoprotein Cj0409
"CJ_10000855"	CJE0459	frdB	Cj0410	frdB	CCO0499	sdhB	0	1	1	1	-1	-1	1	-1	1	1	fumarate reductase iron-sulfur protein	succinate dehydrogenase	fumarate reductase iron-sulfur protein

"CJ_10000857"	CJE0460	-	Cj0411	-	CCO0500	-	1	1	1	1	1	1	1	1	-1	1	0	putative ATP /GTP binding protein	GTP-binding protein	probable ATP /GTP binding protein Cj0411
"CJ_10000859"	CJE0461	-	Cj0412	-	CCO0501	-	1	1	1	1	-1	1	1	1	1	1	1	putative ATP /GTP binding protein	GTP-binding protein	probable ATP /GTP binding protein Cj0412
"CJ_10000861"	CJE0462	-	Cj0413	-	CCO0502	-	1	0	1	1	-1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	conserved hypothetical secreted protein.
"CJ_10000862"	CJE0463	-	Cj0414	-	-	-	1	1	1	1	-1	1	1	1	1	-1	-1	putative oxidoreductase subunit	hypothetical protein	-
"CJ_10000864"	CJE0464	-	Cj0415	-	-	-	1	1	1	1	-1	1	1	1	1	1	1	putative oxidoreductase subunit	oxidoreductase, putative	-
"opCjV0100000636"	CJE0465	-	-	-	-	-	0	0	0	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"opCjV0100001087"	CJE0466	-	-	-	-	-	-1	-1	1	-1	1	-1	-1	1	-1	-1	-1	-	hypothetical protein	-
"CJ_10000878"	CJE0467	-	Cj0418c	-	CCO0503	-	1	1	1	0	-1	1	1	1	-1	1	1	hypothetical protein	hypothetical protein	srpA-related protein
"CJ_10000881"	CJE0468	-	Cj0419	-	CCO0504	-	1	1	1	1	1	1	1	-1	1	1	1	hypothetical protein	HIT family protein	HIT domain protein
"CJ_10000883"	CJE0469	-	Cj0420	-	CCO0505	-	1	1	1	1	-1	1	1	-1	0	1	1	putative periplasmic protein	hypothetical protein	conserved hypothetical secreted protein
"CJ_10000884"	CJE0470	-	Cj0421c	-	CCO0506	-	-1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0421c
"CJ_10000885"	CJE0471	-	Cj0422c	-	CCO0507	-	0	1	1	1	1	1	0	-1	1	1	1	putative H-T-H containing protein	hypothetical protein	conserved hypothetical protein
"opCjV0100000549"	CJE0472	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-1	-	hypothetical protein	-
"opCjV0100000188"	CJE0473	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-1	-	hypothetical protein	-
"opCjV0100000961"	CJE0474	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-1	-	lipoprotein, putative	-
"opCcV0100001299"	CJE0475	-	Cj0426	-	CCO0512	-	1	1	1	1	1	1	1	1	1	1	1	ABC transporter ATP-binding protein	ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein (yheS)
"CJ_10001596"	CJE0476	-	Cj0427	-	CCO0513	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001597"	CJE0477	-	Cj0428	-	CCO0514	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001598"	CJE0478	-	Cj0429c	-	CCO0515	-	1	1	1	1	1	1	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCjV0100001079"	CJE0479	-	-	-	-	-	1	0	0	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_10001599"	CJE0480	-	Cj0430	-	CCO0516	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	conserved hypothetical integral membrane
"CJ_10001600"	CJE0481	-	Cj0431	-	CCO0517	-	1	1	1	1	1	1	1	1	1	1	1	putative periplasmic ATP /GTP-binding protein	hypothetical protein	conserved hypothetical protein
"CJ_10001601"	CJE0482	murD	Cj0432c	murD	CCO0524	murD	1	1	1	1	1	1	1	1	1	1	1	UDP-N-acetylmuramoylalanine--D-glutamate ligase	UDP-N-acetylmuramoyl-L-alanyl-D-glutamate	UDP-N-acetylmuramoylalanine--D-glutamate ligase
"CJ_10000001"	CJE0483	mraY	Cj0433c	mraY	CCO0525	mraY	1	1	1	1	1	1	1	1	0	1	1	phospho-N-acetylmuramoyl-pentapeptide-	phospho-N-acetylmuramoyl-pentapeptide-	phospho-N-acetylmuramoyl-pentapeptide-
"CJ_10000003"	CJE0484	pgm	Cj0434	pgm	CCO0526	gpmA	0	1	0	1	1	1	1	1	1	1	1	phosphoglycerate mutase	phosphoglyceromutase	phosphoglycerate mutase,
"CJ_10000005"	CJE0485	fabG	Cj0435	fabG	CCO0527	fabG	0	1	1	1	1	1	1	0	1	1	1	3-oxoacyl-[acyl-carrier protein] reductase	3-ketoacyl-(acyl-carrier-protein) reductase	3-oxoacyl-(acyl-carrier-protein) reductase
"CJ_10000007"	CJE0486	-	Cj0436	-	CCO0528	-	1	1	1	1	1	1	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCjV0100001038"	CJE0487	-	-	-	-	-	1	1	1	1	1	1	1	1	1	0	0	-	hypothetical protein	-

"CJ_1000022"	CJE0488	sdhA	Cj0437	sdhA	-	-	0	1	1	1	1	1	1	1	1	1	1	succinate dehydrogenase flavoprotein subunit	succinate dehydrogenase, flavoprotein subunit	-
"CJ_1000023"	CJE0489	sdhB	Cj0438	sdhB	-	-	1	0	1	1	-1	1	1	1	1	1	1	putative succinate dehydrogenase iron-sulfur	succinate dehydrogenase, iron-sulfur protein	-
"CJ_1000024"	CJE0490	sdhC	Cj0439	sdhC	-	-	0	1	1	1	1	1	0	1	-1	1	1	putative succinate dehydrogenase subunit C	succinate dehydrogenase, C subunit	-
"CJ_1000025"	CJE0491	-	Cj0440c	-	CCO0529	-	0	1	1	1	1	1	1	0	1	1	1	putative transcriptional regulator	TenA/Thi-4 family protein	transcriptional regulator TenA, putative
"opCjV010000088"	CJE0492	-	-	-	-	-	1	1	1	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_1000026"	CJE0493	acpP	Cj0441	acpP	CCO0530	acpP	1	1	1	1	1	1	1	1	0	1	1	acyl carrier protein	acyl carrier protein	acyl carrier protein
"CJ_1000027"	CJE0494	fabF	Cj0442	fabF	CCO0531	fabB	1	1	1	1	1	1	1	1	1	1	1	3-oxoacyl-[acyl-carrier-protein] synthase	3-oxoacyl-(acyl carrier protein) synthase	beta ketoacyl-acyl carrier protein synthase II
"CJ_1000028"	CJE0495	accA	Cj0443	accA	CCO0532	accA	1	-1	-1	-1	-1	-1	1	1	1	1	1	acetyl-coenzyme A carboxylase alpha subunit	acetyl-CoA carboxylase	acetyl-CoA carboxylase, carboxyl transferase,
"opCcV0100001291"	CJE0496	-	Cj0444	-	CCO0537	-	1	1	1	1	-1	1	1	0	1	1	1	pseudogene	pseudogene	iron-regulated outer membrane virulence protein,
"CJ_1000031"	CJE0497	-	Cj0447	-	CCO0538	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	NUDIX domain protein	conserved hypothetical protein TIGR00052
"CJ_1000033"	CJE0498	-	Cj0448c	-	CCO0539	-	-1	-1	1	1	0	1	1	1	1	1	1	putative MCP-type signal transduction protein	methyl-accepting chemotaxis protein	methyl-accepting chemotaxis transducer (tlpC)
"CJ_1000043"	CJE0499	-	Cj0449c	-	CCO0540	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF465) family
"opCcV0100000349"	CJE0500	rpmB	Cj0450c	rpmB	CCO0541	rpmB	1	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L28	50S ribosomal protein L28	ribosomal protein L28
"CJ_1000044"	CJE0501	rpe	Cj0451	rep	CCO0542	rpe	1	1	1	1	-1	1	1	-1	1	1	1	ribulose-phosphate 3-epimerase	ribulose-phosphate 3-epimerase	ribulose-phosphate 3-epimerase
"CJ_1000045"	CJE0502	-	Cj0452	dnaQ	CCO0543	-	1	0	1	1	1	1	1	1	1	1	1	exonuclease, possibly dna polymerase III epsilon	DNA polymerase III subunit epsilon	DNA polymerase III epsilon subunit (dnaQ)
"CJ_1000046"	CJE0503	thiC	Cj0453	thiC	CCO0544	thiC	1	1	1	1	1	1	1	1	0	1	1	thiamin biosynthesis protein ThiC	thiamine biosynthesis protein ThiC	thiamine biosynthesis protein ThiC
"CJ_1000047"	CJE0504	-	Cj0454c	-	CCO0545	-	0	1	1	1	0	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0454c
"CJ_1000048"	CJE0505	-	Cj0455c	-	CCO0546	-	0	1	1	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0455c
"CJ_1000050"	CJE0506	-	Cj0456c	-	CCO0547	-	0	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_1000052"	CJE0507	-	Cj0457c	-	CCO0548	-	0	0	0	1	1	1	1	1	0	1	1	putative lipoprotein	hypothetical protein	probable lipoprotein Cj0457c
"CJ_1000054"	CJE0508	miaB	Cj0458c	-	CCO0549	miaB	0	0	1	1	1	1	1	1	-1	1	1	hypothetical protein	tRNA-i(6)A37 thiotransferase enzyme MiaB	tRNA-i(6)A37 thiotransferase enzyme MiaB
"CJ_1000062"	CJE0509	-	Cj0459c	-	CCO0550	-	0	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_1000063"	CJE0510	nusA	Cj0460	nusA	CCO0551	nusA	1	1	1	1	1	1	1	1	1	1	1	transcription termination factor	transcription elongation factor NusA	transcription termination factor NusA
"CJ_1000064"	CJE0511	-	Cj0461c	-	CCO0552	-	1	1	1	1	0	1	1	0	1	1	1	putative integral membrane protein	transporter, putative	permease, putative
"CJ_1000065"	CJE0512	-	Cj0462	-	CCO0553	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR00423
"CJ_1000066"	CJE0513	-	Cj0463	-	CCO0554	-	0	0	1	1	0	1	1	1	1	1	1	zinc protease-like protein	peptidase, putative	processing protease (ymxG)
"CJ_1000067"	CJE0514	recG	Cj0464	recG	CCO0555	recG	1	1	1	1	1	1	1	0	1	1	1	ATP-dependent DNA helicase	ATP-dependent DNA helicase RecG	ATP-dependent DNA helicase RecG

"CJ_10000068"	CJE0515	-	Cj0465c	-	CCO0556	-	1	1	0	1	0	0	0	0	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000070"	CJE0516	-	Cj0466	-	CCO0557	-	1	1	1	1	-1	1	1	0	1	1	putative transcriptional regulator	transcriptional regulator, putative	probable transcription regulator Cj0466
"CJ_10000072"	CJE0517	-	Cj0467	-	CCO0559	glnP	1	1	1	1	1	1	1	1	0	1	amino-acid ABC transporter integral membrane	amino acid ABC transporter, permease protein,	glutamine ABC transporter, permease protein
"CJ_10000074"	CJE0518	-	Cj0468	-	CCO0560	glnP	1	1	1	1	1	1	1	1	1	1	amino-acid ABC transporter integral membrane	amino acid ABC transporter, permease protein,	glutamine ABC transporter, permease protein
"CJ_10000082"	CJE0519	-	Cj0469	-	CCO0562	glnQ	1	1	1	1	1	1	1	1	1	1	amino-acid ABC transporter ATP-binding protein	amino acid ABC transporter, ATP-binding protein	glutamine/glutamate ABC transporter, ATP-binding
"CJ_10000083"	CJE0520	tuf	Cj0470	tuf	CCO0566	tuf	1	1	1	1	1	1	1	1	1	1	elongation factor TU	elongation factor Tu	translation elongation factor Tu
"CJ_10000084"	CJE0521	rpmG	Cj0471	rpmG	CCO0567	rpmG	1	0	1	1	1	1	1	1	1	1	50S ribosomal protein L33	50S ribosomal protein L33	ribosomal protein L33
"CJ_10000085"	CJE0522	secE	Cj0472	secE	CCO0569	-	1	1	1	1	1	1	1	1	1	1	preprotein translocase SecE subunit	translocase	preprotein translocase SecE chain Cj0472
"CJ_10000086"	CJE0523	nusG	Cj0473	nusG	CCO0570	nusG	0	1	1	1	1	1	1	1	1	1	putative transcription antitermination protein	transcription antitermination protein NusG	transcription termination/antitermination factor
"CJ_10000087"	CJE0524	rplK	Cj0474	rplK	CCO0571	rplK	0	-1	1	1	1	1	1	1	1	1	50S ribosomal protein L11	50S ribosomal protein L11	ribosomal protein L11
"CJ_10000088"	CJE0525	rplA	Cj0475	rplA	CCO0572	rplA	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L1	50S ribosomal protein L1	ribosomal protein L1
"CJ_10000089"	CJE0526	rplJ	Cj0476	rplJ	CCO0573	-	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L10	50S ribosomal protein L10	50S ribosomal protein L10 Cj0476
"CJ_10000090"	CJE0527	rplL	Cj0477	rplL	CCO0574	rplL	0	1	1	1	1	1	1	1	1	1	50S ribosomal protein L7/L12	50S ribosomal protein L7/L12	ribosomal protein L7/L12
"CJ_10000091"	CJE0528	rpoB	Cj0478	rpoB	CCO0575	-	0	0	1	1	1	1	1	1	1	1	DNA-directed RNA polymerase beta chain	DNA-directed RNA polymerase, beta subunit	DNA-directed RNA polymerase, beta subunit
"CJ_10000092"	CJE0529	rpoB	Cj0479	rpoB	CCO0576	-	1	1	1	1	1	1	1	1	1	1	DNA-directed RNA polymerase beta chain	DNA-directed RNA polymerase, beta subunit	DNA-directed RNA polymerase beta chain ,
"CJ_10000093"	CJE0530	-	Cj0480c	-	CCO0578	-	0	-1	-1	1	0	-1	-1	-1	1	1	putative transcriptional regulator	transcriptional regulator, lclR family	probable transcription regulator Cj0480c
"opCv010000230"	CJE0531	-	-	-	CCO0579	-	1	1	-1	1	1	-1	1	-1	1	1	-	hypothetical protein	probable lyase Cj0481
"CJ_10000094"	CJE0532	-	Cj0481	-	CCO0580	-	1	1	-1	1	1	-1	-1	-1	1	1	putative lyase	dihydrodipicolinate synthase, putative	altronate hydrolase
"CJ_10000095"	CJE0533	uxaA	-	-	-	-	1	1	0	1	1	-1	-1	-1	1	1	-	pseudogene	-
"CJ_10000097"	CJE0534	-	Cj0484	-	CCO0582	-	0	1	-1	1	1	-1	-1	-1	1	1	transmembrane transport protein	tartrate transporter, putative	transmembrane transport protein Cj0484
"CJ_10000098"	CJE0535	-	Cj0485	-	CCO0583	-	1	0	-1	1	1	1	1	-1	1	1	putative oxidoreductase	short chain dehydrogenase	probable oxidoreductase Cj0485
"CJ_10000099"	CJE0536	fucP	Cj0486	-	CCO0584	fucP	1	1	-1	1	1	1	-1	-1	1	1	putative sugar transporter	L-fucose permease	L-fucose permease
"CJ_10000100"	CJE0537	-	Cj0487	-	CCO0585	-	0	1	-1	1	-1	-1	-1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000101"	CJE0538	-	Cj0488	-	CCO0586	-	-1	0	-1	1	1	1	-1	-1	1	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF718)
"CJ_10000109"	CJE0539	aldA	Cj0490	-	CCO0587	-	-1	1	-1	1	1	1	1	-1	1	1	pseudogene	aldehyde dehydrogenase	probable lactaldehyde dehydrogenase truncated
"CJ_10000113"	CJE0540	rpsL	Cj0491	rpsL	CCO0588	rpsL	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S12	30S ribosomal protein S12	ribosomal protein S12
"CJ_10000115"	CJE0541	rpsG	Cj0492	rpsG	CCO0589	rpsG	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S7	30S ribosomal protein S7	ribosomal protein S7
"CJ_10000116"	CJE0542	fusA	Cj0493	fusA	CCO0590	fusA	1	1	1	1	1	1	1	1	1	1	elongation factor G	elongation factor EF-2	translation elongation factor G
"opCjV0100000897"	CJE0544	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	0	-1	-	site-specific recombinase, phage integrase	-

"opCjV010000 0168"	CJE0545	-	-	-	-	-	-1	-1	-1	-1	1	1	-1	1	-1	-1	-	DNA binding domain, excisionase family	-
"opCjV010000 0553"	CJE0546	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	pentapeptide repeat family protein	-
"opCjV010000 0167"	CJE0547	-	-	-	-	-	-1	-1	-1	1	-1	1	1	-1	-1	-	-	hypothetical protein	-
"opCjV010000 0090"	CJE0548	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	phage anti-repressor protein, putative	-
"opCjV010000 0446"	CJE0549	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0163"	CJE0550	-	-	-	-	-	-1	-1	-1	0	1	-1	-1	1	-1	1	-	site-specific DNA- methyltransferase, putative	-
"opCjV010000 0554"	CJE0551	-	-	-	-	-	-1	-1	-1	-1	1	1	1	-1	-1	-	-	hypothetical protein	-
"opCjV010000 1062"	CJE0552	-	-	-	-	-	-1	-1	-1	1	0	-1	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0291"	CJE0554	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein	-
"opCjV010000 0756"	CJE0555	-	-	-	-	-							-1	-1			-	hypothetical protein	-
"opCjV010000 0790"	CJE0556	-	-	-	-	-	-1	-1	0	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV010000 0463"	CJE0557	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0608"	CJE0558	-	-	-	-	-		-1					1	-1			-	hypothetical protein	-
"opCjV010000 0149"	CJE0559	-	-	-	-	-		-1	-1	1	-1	-1	1	1	1	-	-	hypothetical protein	-
"opCjV010000 0254"	CJE0560	-	-	-	-	-		-1			-1		-1				-	hypothetical protein	-
"opCjV010000 0597"	CJE0561	-	-	-	-	-		-1	-1	-1	-1	1	-1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0602"	CJE0562	-	-	-	-	-									-1	-	-	hypothetical protein	-
"opCjV010000 0387"	CJE0563	-	-	-	-	-		-1	-1	1	-1	-1	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0421"	CJE0564	-	-	-	-	-	-1	-1	1	1	-1	-1	1	-1	1	-	-	hypothetical protein	-
"opCjV010000 0004"	CJE0565	-	-	-	-	-								-1			-	hypothetical protein	-
"opCjV010000 0955"	CJE0566	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	DNA/RNA non-specific endonuclease	-
"opCjV010000 0508"	CJE0567	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1	1	0	-	-	hypothetical protein	-
"opCjV010000 0171"	CJE0568	-	-	-	-	-		-1	-1	1	-1	-1	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0325"	CJE0569	-	-	-	-	-		-1	-1				-1	1	-1	-	-	phage repressor protein, putative	-
"opCjV010000 0973"	CJE0570	-	-	-	-	-		-1	-1	-1	-1	1	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0113"	CJE0571	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein	-
"opCjV010000 0018"	CJE0572	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	lipoprotein, putative	-
"opCjV010000"	CJE0573	-	-	-	-	-	-1	-1	-1	-1	1	-1	1	1	0	-	-	hypothetical protein	-

0063"										-1	-1	-1		1	-1	-1	1	-1	-	hypothetical protein	-	
"opCjV010000 0415"	CJE0574	-	-	-	-	-				-1	-1	-1	1	-1	-1	1	-1	-	hypothetical protein	-		
"opCjV010000 0316"	CJE0575	-	-	-	-	-				-1	-1	-1	1	-1	-1	1	-1	-	hypothetical protein	-		
"opCjV010000 0315"	CJE0576	-	-	-	-	-				-1	-1	-1	1	-1	-1	1	-1	-	hypothetical protein	-		
"opCjV010000 0129"	CJE0577	-	-	-	-	-					-1		-1	-1	-1	1	-1	-	terminase B protein, putative	-		
"opCjV010000 0083"	CJE0578	-	-	-	-	-					-1	1	1	-1	-1	1	1	1	-	hypothetical protein	-	
"opCjV010000 0745"	CJE0579	-	-	-	-	-				-1	-1	-1			1	1		-1	-	hypothetical protein	-	
"opCjV010000 0037"	CJE0580	-	-	-	-	-					-1		-1	1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV010000 0400"	CJE0581	-	-	-	-	-				-1	-1	-1	1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV010000 0694"	CJE0582	-	-	-	-	-				-1	0	-1	1	-1	-1	1	0	-1	-	hypothetical protein	-	
"opCjV010000 1059"	CJE0583	-	-	-	-	-				-1	-1	-1	1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV010000 0732"	CJE0584	-	-	-	-	-					-1	-1	1	-1	-1	1	0	1	-	hypothetical protein	-	
"opCjV010000 0491"	CJE0585	-	-	-	-	-		-1	-1	-1	-1	-1	1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV010000 0666"	CJE0586	-	-	-	-	-					-1	1	1			1		1	-	hypothetical protein	-	
"opCjV010000 0583"	CJE0587	-	-	-	-	-					-1				-1	-1			-	hypothetical protein	-	
"opCjV010000 0249"	CJE0588	-	-	-	-	-					-1	-1	-1	-1	1	-1	1	-1	-	hypothetical protein	-	
"opCjV010000 0236"	CJE0589	-	-	-	-	-					-1	-1	-1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCjV010000 0976"	CJE0590	-	-	-	-	-					-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV010000 0760"	CJE0591	-	-	-	-	-		-1	-1	-1	1	1	-1	-1	1	-1	1	1	-	hypothetical protein	-	
"opCjV010000 0011"	CJE0592	-	-	-	-	-		-1	-1	-1	0	1	-1	-1	1	-1	0	-	-	hypothetical protein	-	
"opCjV010000 0466"	CJE0593	-	-	-	-	-		-1	-1	-1	-1	1	-1	1	1	-1	-1	-	-	hypothetical protein	-	
"opCjV010000 1114"	CJE0594	-	-	-	-	-		-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein	-	
"opCjV010000 0157"	CJE0595	-	-	-	-	-					-1	-1	-1	-1	1	1	-1	-1	-	hypothetical protein	-	
"opCjV010000 0319"	CJE0596	-	-	-	-	-					-1	-1	0		1	1	1	-1	0	-	hypothetical protein	-
"opCjV010000 0852"	CJE0597	-	-	-	-	-		-1	-1	-1	0	1	-1	-1	1	-1	0	-	-	hypothetical protein	-	
"opCjV010000 0009"	CJE0598	-	-	-	-	-					-1	1	1	-1	-1	-1	1	1	-	hypothetical protein	-	
"opCjV010000 0123"	CJE0599	-	-	-	-	-		-1	-1	-1	0	-1	-1	-1	1	-1	0	-	-	hypothetical protein	-	
"opCjV010000 0091"	CJE0600	-	-	-	-	-					-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-

"opCjV0100000645"	CJE0601	-	-	-	-	-	-1	-1	0	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-	
"opCjV0100000787"	CJE0602	-	-	-	-	-	0	-1	0	0	1	1	1	1	1	1	-	hypothetical protein	-	
"CJ_10000118"	CJE0603	-	Cj0495	-	CCO0593	-	0	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000119"	CJE0604	-	Cj0496	-	CCO0594	-	0	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000120"	CJE0605	-	Cj0497	-	CCO0595	-	1	0	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	ATP-dependent nuclease (addB), putative
"CJ_10000121"	CJE0606	trpC	Cj0498	trpC	CCO0596	trpC	1	1	1	1	1	1	1	1	1	1	1	indole-3-glycerol phosphate synthase	indole-3-glycerol phosphate synthase	indole-3-glycerol phosphate synthase
"CJ_10000129"	CJE0607	-	Cj0499	-	CCO0597	hit	1	1	1	1	1	1	1	0	1	1	1	HIT-like protein	Hit family protein	hit family protein
"CJ_10000131"	CJE0608	-	Cj0500	-	CCO0598	-	1	1	1	1	1	1	1	1	1	1	1	putative ATP /GTP binding protein	GTP-binding protein	rhodanese-like domain protein, putative
"CJ_10000133"	CJE0609	-	Cj0501	-	CCO0599	amt	-	-1	0	1	1	-1	1	1	-1	1	1	pseudogene	pseudogene	ammonium transporter
"CJ_10000135"	CJE0610	hemH	Cj0503c	hemH	CCO0600	hemH	1	1	1	1	0	1	1	1	1	1	1	putative ferredoxin	ferredoxin	ferredoxin
"CJ_10000136"	CJE0611	-	Cj0504c	-	CCO0602	-	0	1	1	1	1	1	1	1	1	1	1	hypothetical protein	oxidoreductase, Gfo/Idh/MocA family	oxidoreductase, Gfo/Idh/MocA family, putative
"CJ_10000137"	CJE0612	-	Cj0505c	-	CCO0604	degT	0	1	1	1	0	1	1	1	1	1	1	putative aminotransferase (degT family)	DegT/DnrJ/EryC1/StrS aminotransferase family	A porR mutant of Pophyromonas gingivalis shows
"CJ_10000138"	CJE0613	alaS	Cj0506	alaS	CCO0605	alaS	1	1	1	1	1	1	1	1	1	1	1	alanyl-tRNA synthetase	alanyl-tRNA synthetase	alanyl-tRNA synthetase
"CJ_10000139"	CJE0614	maf	Cj0507	maf	CCO0606	-	0	1	1	1	1	1	1	1	1	1	1	MAF homolog	Maf-like protein	Maf protein, putative
"CJ_10000140"	CJE0615	pbpA	Cj0508	pbpA	CCO0607	-	1	1	1	1	1	1	1	1	1	1	1	penicillin-binding protein	penicillin-binding protein 1A	penicillin-binding protein 1A (PBP-1A)
"opCcV0100000257"	CJE0616	clpB	Cj0509c	clpB	CCO0608	-	1	-1	1	-1	1	1	1	1	1	-1	0	ATP-dependent CLP protease ATP-binding subunit	ATP-dependent chaperone protein ClpB	ATP-dependent Clp protease, ATP-binding subunit
"opCcV0100000188"	CJE0617	-	Cj0510c	-	CCO0609	-	1	1	1	0	1	0	0	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000149"	CJE0618	ctpA	Cj0511	-	CCO0611	prc	-1	1	1	1	1	1	1	1	1	1	1	putative secreted protease	carboxyl-terminal protease	protease
"CJ_10000151"	CJE0619	purC	Cj0512	purC	CCO0612	purC	1	1	1	1	0	1	1	1	1	1	1	phosphoribosylaminoimidazole-succinocarboxamide	phosphoribosylaminoimidazole-succinocarboxamide	phosphoribosylaminoimidazole-succinocarboxamide
"CJ_10000153"	CJE0620	purS	Cj0513	-	CCO0613	purS	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	phosphoribosylformylglycine amidine synthase, purS	phosphoribosylformylglycine amidine synthase, PurS
"CJ_10000154"	CJE0621	purQ	Cj0514	purQ	CCO0614	purQ	1	1	1	1	1	1	1	1	1	1	1	phosphoribosylformylglycine amidine synthase I	phosphoribosylformylglycine amidine synthase	phosphoribosylformylglycine amidine synthase I
"CJ_10000155"	CJE0622	-	Cj0515	-	CCO0615	-	1	0	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0515
"CJ_10000156"	CJE0623	-	Cj0516	plsC	CCO0616	-	0	0	1	1	0	1	1	1	1	1	1	putative 1-acyl-SN-glycerol-3-phosphate	1-acyl-sn-glycerol-3-phosphate acyltransferase,	1-acyl-sn-glycerol-3-phosphate acyltransferase,
"CJ_10000157"	CJE0624	-	Cj0517	crcB	CCO0617	crcB	1	1	1	1	1	1	1	1	1	1	1	CRCB protein homolog	CrcB	crcB protein
"CJ_10000158"	CJE0625	htpG	Cj0518	htpG	CCO0618	-	-1	1	1	1	1	1	1	0	1	1	1	hsp90 family heat shock protein	heat shock protein 90	hsp90 family heat shock protein Cj0518
"CJ_10000159"	CJE0626	-	Cj0519	-	CCO0619	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000165"	CJE0627	-	Cj0520	-	CCO0620	-	1	0	1	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0520
"CJ_10000169"	CJE0628	-	Cj0523	-	CCO0621	-	1	1	1	1	1	1	1	1	1	1	1	putative membrane protein	Na/Pi-cotransporter, putative	Na/Pi cotransporter, putative
"CJ_10000172"	CJE0629	-	Cj0525c	pbpB	CCO0622	-	0	1	1	1	0	1	1	1	1	1	1	putative penicillin-binding protein	penicillin-binding protein	cell division protein (ftsI)
"CJ_10000173"	CJE0630	fliE	Cj0526c	fliE	CCO0623	fliE	1	1	1	1	-1	1	1	-1	-1	-1	-1	putative flagellar hook-basal	flagellar basal body protein	flagellar hook-basal body

"CJ_10000174"	CJE0631	flgC	Cj0527c	flgC	CCO0624	flgC	0	0	1	1	1	1	1	1	1	1	body complex	flagellar basal-body rod protein	flagellar basal body rod protein	flagellar basal-body rod protein FlgC
"CJ_10000175"	CJE0632	flgB	Cj0528c	flgB	CCO0625	flgB	0	1	1	1	1	1	1	1	1	1	putative flagellar basal-body rod protein	flagellar basal body rod protein	flagellar basal-body rod protein FlgB	
"CJ_10000176"	CJE0633	-	Cj0529c	-	CCO0626	-	0	1	1	1	0	1	1	1	1	1	hypothetical protein	hypothetical protein	Uncharacterized BCR, YceG family COG1559	
"CJ_10000177"	CJE0634	-	Cj0530	-	CCO0627	-	1	1	1	1	1	1	0	0	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0530	
"CJ_10000781"	CJE0635	icd	Cj0531	icd	CCO0628	-	1	1	1	1	-1	1	1	-1	1	1	isocitrate dehydrogenase	isocitrate dehydrogenase, NADP-dependent	isocitrate dehydrogenase, NADP-dependent	
"CJ_10000782"	CJE0636	mdh	Cj0532	mdh	CCO0629	-	1	1	1	1	-1	1	-1	-1	1	1	malate dehydrogenase	malate dehydrogenase	malate dehydrogenase Cj0532	
"CJ_10000783"	CJE0637	sucC	Cj0533	sucC	CCO0630	sucC	1	1	1	1	-1	1	1	-1	1	1	succinyl-coA synthetase beta chain	succinyl-CoA synthase, beta subunit	succinyl-CoA synthase, beta subunit	
"CJ_10000784"	CJE0638	sucD	Cj0534	sucD	CCO0631	sucD	1	1	1	1	-1	1	-1	-1	1	-1	succinyl-coA synthetase alpha chain	succinyl-CoA synthase, alpha subunit	SucD	
"opCcV0100001932"	CJE0639	oorD	Cj0535	oorD	CCO0632	korD	1	1	1	0	-1	1	1	1	1	0	OORD subunit of 2-oxoglutarate:acceptor	2-oxoglutarate:acceptor oxidoreductase, delta	ferredoxin-like protein	
"CJ_10000786"	CJE0640	oorA	Cj0536	oorA	CCO0633	vorB	1	-1	1	1	1	1	0	1	1	1	OORA subunit of 2-oxoglutarate:acceptor	2-oxoglutarate ferredoxin oxidoreductase	ferredoxin oxidoreductase, alpha subunit	
"CJ_10000788"	CJE0641	oorB	Cj0537	oorB	CCO0634	oorB	1	1	1	1	1	0	1	1	1	1	OORB subunit of 2-oxoglutarate:acceptor	ferredoxin oxidoreductase beta subunit	ferredoxin oxidoreductase, beta subunit	
"CJ_10000791"	CJE0642	oorC	Cj0538	oorC	CCO0635	-	1	0	1	1	1	1	0	1	1	1	OORC subunit of 2-oxoglutarate:acceptor	indolepyruvate ferredoxin oxidoreductase	ferredoxin oxidoreductase, gamma subunit	
"CJ_10000794"	CJE0643	-	Cj0539	-	CCO0636	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000797"	CJE0644	-	Cj0540	-	CCO0637	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000809"	CJE0645	-	Cj0541	-	CCO0638	ispB	1	0	1		-1	1	-1	1	1	-1	polyprenyl synthetase	polyprenyl synthetase family protein	octaprenyl-diphosphate synthase (ispB)	
"CJ_10000810"	CJE0646	hemA	Cj0542	hemA	CCO0639	hemA	1	1	1	1	1	1	1	1	1	1	glutamyl-tRNA reductase	glutamyl-tRNA reductase	glutamyl-tRNA reductase	
"CJ_10000811"	CJE0647	proS	Cj0543	proS	CCO0640	proS	1	1	1	1	-1	1	1	-1	1	1	prolyl-tRNA synthetase	prolyl-tRNA synthetase	prolyl-tRNA synthetase	
"CJ_10000812"	CJE0648	-	Cj0544	-	CCO0641	-	1	1	-1	1	1	1	-1	1	-1	-1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0544	
"CJ_10000813"	CJE0649	hemC	Cj0545	hemC	CCO0642	hemC	1	1	1	0	-1	1	-1	-1	1	1	porphobilinogen deaminase	porphobilinogen deaminase	porphobilinogen deaminase	
"CJ_10000815"	CJE0650	-	Cj0546	-	CCO0643	-	1	0	1	1	-1	1	1	-1	1	1	hypothetical protein	3-octaprenyl-4-hydroxybenzoate carboxylase,	conserved hypothetical protein TIGR00148,	
"CJ_10000817"	CJE0651	flaG	Cj0547	flaG	CCO0644	-	0	1	1	1	-1	1	1	-1	-1	-1	possible flagellar protein	flagellar protein FlaG	probable flagellar protein Cj0547	
"CJ_10000820"	CJE0652	fliD	Cj0548	fliD	CCO0645	-	1	1	1	1	-1	1	1	-1	1	1	putative flagellar hook-associated protein	flagellar hook-associated protein	flagellar hook-associated protein 2 (fliD),	
"CJ_10000823"	CJE0653	fliS	Cj0549	fliS	CCO0646	fliS	1	1	1		-1	1	1	-1	1	1	flagellar protein	flagellar protein FliS	flagellar protein FliS	
"CJ_10000826"	CJE0654	-	Cj0550	-	CCO0647	-	1	1	1	1	-1	0	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000836"	CJE0655	efp	Cj0551	efp	CCO0649	efp	1	0	1	1	-1	1	1	-1	1	1	elongation factor P	elongation factor P	translation elongation factor P	
"CJ_10000837"	CJE0656	-	Cj0552	-	-	-	1	1	1	1	-1	1	1	1	1	1	hydrophobic protein	hypothetical protein	-	
"CJ_10000838"	CJE0657	-	Cj0553	-	-	-	1	1	1	1	-1	1	-1	-1	1	1	putative integral membrane protein	hypothetical protein	-	
"CJ_10000839"	CJE0658	-	Cj0554	-	-	-	1	1	1	1	-1	-1	0	1	1	-1	hypothetical protein	hypothetical protein	-	
"CJ_10000840"	CJE0659	-	Cj0555	-	-	-	-1	1	1	-1	-1	1	1	-1	1	-1	putative integral membrane protein	hypothetical protein	-	
"CJ_10000842"	CJE0660	-	Cj0556	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	-	

"CJ_10000844"	CJE0661	-	Cj0557c	-	CCO0650	-	1	1	1	-1	-1	1	-1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0557c	
"CJ_10000845"	CJE0662	-	Cj0557c	-	CCO0651	-	1	1	1	0	1	1	1	-1	0	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0557c
"CJ_10000850"	CJE0663	-	Cj0559	-	CCO0653	-	1	1	1	1	-1	1	1	-1	1	1	oxidoreductase	pyridine nucleotide-disulphide oxidoreductase	oxidoreductase Cj0559
"CJ_10000853"	CJE0665	-	Cj0560	-	CCO0655	-	1	1	1	-1	-1	1	1	-1	-1	1	putative integral membrane protein	pseudogene	MATE efflux family protein, putative
"CJ_10000865"	CJE0666	-	Cj0561c	-	CCO0656	-	1	0	1	1	-1	-1	-1	0	1	1	possible periplasmic protein	hypothetical protein	probable periplasmic protein Cj0561c
"CJ_10000866"	CJE0667	dnaB	Cj0562	dnaB	CCO0657	dnaB	1	1	1	1	1	1	1	-1	-1	1	replicative DNA helicase	replicative DNA helicase	replicative DNA helicase
"CJ_10000867"	CJE0668	-	Cj0563	-	-	-	1	1	1	-1	1	-1	1	-1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10000868"	CJE0669	-	Cj0564	-	-	-	1	1	1	1	1	1	-1	-1	1	-1	putative integral membrane protein	hypothetical protein	-
"opCjV0100000279"	CJE0670	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	GTP-binding protein	-
"opCjV0100000872"	CJE0671	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCjV0100001026"	CJE0672	-	-	-	-	-	-1	-1	-1	-1	1	-1	1	1	-1	-1	-	hypothetical protein	-
"CJ_10000882"	CJE0673	-	Cj0570	-	CCO0247	-	1	1	1	1	-1	1	1	-1	1	1	putative ATP /GTP binding protein	pseudogene	probable ATP /GTP binding protein Cj0570
"CJ_10000889"	CJE0674	-	Cj0571	-	CCO0245	-	1	1	1	1	-1	1	1	1	1	1	putative transcriptional regulator	transcriptional regulator, putative	probable transcription regulator Cj0571
"CJ_10000890"	CJE0675	ribB	Cj0572	ribA	CCO1384	ribBA	1	1	1	1	1	-1	1	1	-1	1	GTP cyclohydrolase II / 3,4-dihydroxy-2-butanone	bifunctional 3,4-dihydroxy-2-butanone	3,4-dihydroxy-2-butanone 4-phosphate
"CJ_10000891"	CJE0676	-	Cj0573	-	CCO1383	-	1	-	1	0	-1	1	1	-1	1	-1	hypothetical protein	GatB/Yqey family protein	Yqey family protein
"CJ_10000892"	CJE0677	ilvB	Cj0574	ilvI	CCO1382	ilvB	1	1	1	1	-1	0	-1	1	1	-1	acetolactate synthase large subunit	acetolactate synthase III large subunit	acetolactate synthase, large subunit,
"CJ_10000893"	CJE0678	ilvH	Cj0575	ilvH	CCO1381	ilvN	1	0	1	1	1	1	1	-1	1	1	acetolactate synthase small subunit	acetolactate synthase III small subunit	acetolactate synthase, small subunit
"CJ_10000894"	CJE0679	lpxD	Cj0576	lpxD	CCO1380	lpxD	1	1	1	1	-1	1	1	1	1	1	UDP-3-O-[3-hydroxymyristoyl] glucosamine	UDP-3-O-[3-hydroxymyristoyl] glucosamine	UDP-3-O-[3-hydroxymyristoyl] glucosamine
"CJ_10000895"	CJE0680	queA	Cj0577c	queA	CCO1379	queA	1	1	1	1	1	1	1	1	-1	1	S-adenosylmethionine:tRNA	S-adenosylmethionine:tRNA	S-adenosylmethionine:tRNA
"CJ_10000896"	CJE0681	tatC	Cj0578c	mttB	CCO1378	tatC	1	1	1	-1	-1	1	-1	-1	1	-1	sec-independent protein translocase	Sec-independent protein translocase TatC	Sec-independent protein translocase TatC
"CJ_10000897"	CJE0682	tatB	Cj0579c	-	CCO1377	-	0	1	1	1	-1	1	1	1	1	1	hypothetical protein	sec-independent translocase	Sec-independent protein translocase protein
"CJ_10000898"	CJE0683	-	Cj0580c	-	CCO1376	-	1	1	1	-1	-1	1	1	-1	-1	1	putative oxidoreductase	coproporphyrinogen III oxidase	oxygen-independent coproporphyrinogen III
"CJ_10000899"	CJE0684	nidH	Cj0581	-	CCO1375	mutT	1	1	1	1	-1	1	-1	-1	1	-1	putative NTPase	dinucleoside polyphosphate hydrolase	(di)nucleoside polyphosphate hydrolase
"CJ_10000900"	CJE0685	-	Cj0582	lysC	CCO1374	-	1	1	1	1	-1	1	1	-1	1	1	aspartokinase, alpha and beta subunits	aspartate kinase	aspartate kinase, monofunctional class
"CJ_10000901"	CJE0686	-	Cj0583	-	CCO1373	-	1	1	1	1	-1	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000902"	CJE0687	-	Cj0584	-	CCO1372	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	DNA polymerase III subunit delta	DNA polymerase III delta prime subunit (holB)
"opCcV0100000841"	CJE0688	folP	Cj0585	folP	CCO1371	folP	1	1	1	1	-1	0	-1	1	0	1	putative dihydropteroate synthase	dihydropteroate synthase	dihydropteroate synthase (folP)
"CJ_10000903"	CJE0689	ligA	Cj0586	ligA	CCO1368	-	1	1	1	1	-1	1	1	1	1	1	DNA ligase	DNA ligase	DNA ligase (NAD) Cj0586
"CJ_10000904"	CJE0690	-	Cj0587	-	CCO1366	-	1	1	1	1	-1	1	-1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0587

"CJ_10000905"	CJE0691	tlyA	Cj0588	tlyA	CCO1365	tly	1	1	1	-1	-1	1	1	1	1	1	putative haemolysin	hemolysin A	hemolysin (tly)
"CJ_10000906"	CJE0692	-	Cj0589	ribF	CCO1364	ribF	1	1	1	1	1	1	1	-1	1	1	putative riboflavin kinase /FMN	hypothetical protein	riboflavin biosynthesis protein RibF
"CJ_10000907"	CJE0693	-	Cj0590	-	CCO1363	-	1	1	1	1	-1	1	1	-1	1	0	hypothetical protein	methyltransferase, putative	methyltransferase, putative
"CJ_10000915"	CJE0694	-	Cj0591c	-	CCO1362	-	1	1	1	1	1	1	1	-1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0591c -related protein
"CJ_10000917"	CJE0695	-	Cj0592c	-	-	-	1	1	1	1	-1	1	1	1	1	-1	putative periplasmic protein	hypothetical protein	-
"CJ_10000920"	CJE0696	-	Cj0593c	-	CCO1361	-	1	1	1	1	-1	1	1	-1	-1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0593c
"CJ_10000923"	CJE0697	-	Cj0594c	-	CCO1360	-	0	1	1	1	-1	1	1	1	1	1	putative periplasmic protein	DNA/RNA non-specific endonuclease	DNA/RNA non-specific endonuclease
"CJ_10000925"	CJE0698	nth	Cj0595c	nth	CCO1359	nth	0	1	1	1	0	1	1	1	1	1	endonuclease III	endonuclease III	endonuclease III
"CJ_10000926"	CJE0699	-	Cj0596	-	CCO1358	-	1	1	1	1	-1	1	1	1	1	1	peptidyl-prolyl cis-trans isomerase	major antigenic peptide PEB4	peptidyl-prolyl cis-trans isomerase
"CJ_10000927"	CJE0700	fbaA	Cj0597	fba	CCO1357	fba	1	1	1	-1	-1	1	0	-1	1	-1	fructose-bisphosphate aldolase	fructose-bisphosphate aldolase	fructose-bisphosphate aldolase, class II
"CJ_10000929"	CJE0701	-	Cj0598	-	CCO1356	-	1	1	1	1	1	1	1	0	1	-1	putative membrane protein	hypothetical protein	probable membrane protein Cj0598
"CJ_10000931"	CJE0702	-	Cj0599	-	CCO1355	-	1	1	1	1	-1	1	1	0	1	1	putative periplasmic protein	chemotaxis protein MotB, putative	probable periplasmic protein Cj0599
"CJ_10000932"	CJE0703	-	Cj0600	-	CCO1354	-	1	1	1	1	0	1	-1	1	0	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000942"	CJE0704	-	Cj0601c	-	CCO0684	-	1	1	1	1	-1	1	1	1	1	1	putative sodium-dependent transmembrane	sodium-dependent transporter, putative	sodium- and chloride-dependent transporter
"CJ_10000944"	CJE0705	-	Cj0602c	-	CCO0685	-	1	1	1	-1	-1	1	1	0		1	hypothetical protein	MOSC domain protein	MOSC domain protein
"CJ_10000947"	CJE0706	dsbD	Cj0603c	dsbD	CCO0686	-	1	1	1	1	-1	1	1	1	1	1	putative thiol:disulfide interchange protein	thiol:disulfide interchange protein DsbD	thiol:disulfide interchange protein DsbD
"CJ_10000950"	CJE0707	-	Cj0604	-	CCO0687	-	1	1	1	1	-1	0	-1	-1	1	1	hypothetical protein	hypothetical protein	Domain of unknown function (DUF344) family
"CJ_10000952"	CJE0708	-	Cj0605	-	CCO0688	-	1	1	1	1	1	-1	-1	1	-1	-1	putative amidohydrolase	carboxypeptidase	peptidase, M20/M25/M40 family
"CJ_10000954"	CJE0709	-	Cj0606	-	CCO0689	-	1	1	1	-1	-1	0	0	-1	1	1	putative periplasmic protein	macrolide-specific efflux protein macA	probable periplasmic protein Cj0606
"CJ_10000956"	CJE0710	-	Cj0607	-	CCO0690	ybjZ	-1	0	1	1		1	-1	-1	1	1	ABC-type transmembrane transport protein	macrolide-specific efflux protein macB	ABC transporter, ATP-binding protein
"CJ_10000958"	CJE0711	-	Cj0608	-	CCO0691	-	0	1	1	1	-1	1	1	-1	1	1	putative outer membrane protein	outer membrane efflux protein	outer membrane efflux family protein, putative
"CJ_10000960"	CJE0712	-	Cj0609c	-	CCO0692	-	1	1	1	1	1	1	1	1	1	1	possible periplasmic protein	hypothetical protein	probable periplasmic protein Cj0609c
"CJ_10000961"	CJE0713	-	Cj0610c	-	CCO0693	-	1	1	1	1	-1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0610c
"CJ_10000972"	CJE0714	algI	Cj0611c	-	CCO0694	-	1	1	1	1	-1	1	-1	-1	1	-1	putative transmembrane transport protein	alginate O-acetyltransferase AlgI	probable transmembrane transport protein
"opCcV0100000246"	CJE0715	ftn	Cj0612c	cft	CCO0695	pfr	-1	-1	1	0	1	1	-1	1	-1	1	ferritin	nonheme iron-containing ferritin	ferritin
"CJ_10000976"	CJE0716	-	Cj0613	pstS	CCO0696	-	1	1	1	1	-1	1	1	1	1	-1	possible periplasmic phosphate binding protein	phosphate ABC transporter, periplasmic	probable periplasmic phosphate binding protein
"CJ_10000979"	CJE0717	-	Cj0614	pstC	CCO0697	-	1	1	1	1	0	1	1	-1	1	1	putative phosphate transport system permease	phosphate ABC transporter, permease protein	probable phosphate transport system permease
"CJ_10000981"	CJE0718	-	Cj0615	pstA	CCO0698	-	1	1	1	1	1	1	1	1	1	1	putative phosphate transport system permease	phosphate ABC transporter, permease protein	probable phosphate transport system permease
"CJ_10000983"	CJE0719	pstB	Cj0616	pstB	CCO0699	pstB	1	0	1	1	1	1	1	1	1	1	putative phosphate transport ATP-binding	phosphate ABC transporter, ATP-binding protein	phosphate ABC transporter, ATP-binding protein
"CJ_10000987"	CJE0720	-	Cj0618	-	CCO0700	-	1	1	1	1	1	-1	1	1	1	-1	hypothetical protein	pseudogene	conserved hypothetical protein

"opCjV0100001074"	CJE0721	-	-	-	-	-	-1	0	-1	1	1	-1	-1	1	1	1	-	hypothetical protein	-
"CJ_10000988"	CJE0722	-	Cj0619	-	CCO0701	-	1	1	1	-1	1	1	1	1	1	-1	putative integral membrane protein	MATE efflux family protein	probable integral membrane protein Cj0619
"CJ_10000990"	CJE0723	-	Cj0620	-	CCO0702	-	-1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001001"	CJE0724	-	Cj0621	-	CCO0703	-	0	1	1	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001003"	CJE0725	hypF	Cj0622	hypF	CCO0704	hypF	1	1	1	-1	-1	1	1	1	1	1	transcriptional regulatory protein hypF	[NiFe] hydrogenase maturation protein HypF	[NiFe] hydrogenase maturation protein HypF
"CJ_10001006"	CJE0726	hypB	Cj0623	hypB	CCO0705	hypB	1	1	1	1	1	1	1	0	1	1	hydrogenase isoenzymes formation protein	hydrogenase accessory protein HypB	hydrogenase accessory protein HypB
"CJ_10001009"	CJE0727	hypC	Cj0624	hypC	CCO0706	hypC	1	1	1	1	0	1	1	1	1	1	hydrogenase isoenzymes formation protein	hydrogenase assembly chaperone HypC/HupF	hydrogenase assembly chaperone hypC/hupF
"CJ_10001011"	CJE0728	hypD	Cj0625	hypD	CCO0707	hypD	1	1	1	1	1	1	1	1	1	1	hydrogenase isoenzymes formation protein	hydrogenase expression/formation protein HypD	hydrogenase expression/formation protein HypD
"CJ_10001013"	CJE0729	hypE	Cj0626	hypE	CCO0708	-	1	1	1	1	-1	1	1	-1	-1	-1	hydrogenase isoenzymes formation protein	hydrogenase expression/formation protein HypE	hydrogenase isoenzymes formation protein Cj0626
"CJ_10001015"	CJE0730	hypA	Cj0627	hypA	CCO0709	hypA	1	1	1	1	-1	1	1	1	1	1	hydrogenase expression /formation protein	hydrogenase nickel insertion protein HypA	hydrogenase nickel insertion protein HypA
"opCjV0100000653"	CJE0731	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1	1	1	-1	-	type III restriction/modification enzyme,	-
"opCjV0100000114"	CJE0732	-	-	-	-	-	-1	-1	-1	-1	1	-1	1	1	-1	-1	-	type III restriction-modification enzyme	-
"CJ_10001019"	CJE0733	-	Cj0630c	-	CCO0710	-	1	0	0	1	1	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000102"	CJE0734	-	Cj0631c	-	CCO0711	-	1	1	1	1	0	1	1	-1	1	1	putative ribonuclease	RNB-like protein	probable ribonuclease Cj0631c
"CJ_10000103"	CJE0735	ilvC	Cj0632	ilvC	CCO0712	ilvC	1	1	1	1	-1	1	1	1	1	1	ketol-acid reductoisomerase	ketol-acid reductoisomerase	ketol-acid reductoisomerase
"CJ_10000104"	CJE0736	-	Cj0633	-	CCO0713	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0633
"CJ_10000105"	CJE0737	dprA	Cj0634	-	CCO0714	dprA	1	1	1	1	1	1	1	1	1	1	SMF family protein	DNA processing protein A	DNA processing chain A (dprA)
"CJ_10000106"	CJE0738	-	Cj0635	-	CCO0715	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	Holliday junction resolvase-like protein	conserved hypothetical protein TIGR00250
"CJ_10000107"	CJE0739	-	Cj0636	-	CCO0716	-	1	1	1	1	1	1	1	1	1	1	NOL1/NOP2/sun family protein	NOL1/NOP2/sun family protein	Sun/nucleolar protein family protein VC1502
"CJ_10000108"	CJE0740	msrA	Cj0637c	mrsA	CCO0717	msrA	0	1	1	1	0	1	1	1	1	1	putative peptide methionine sulfoxide reductase	peptide methionine sulfoxide reductase	peptide methionine sulfoxide reductase
"CJ_10000110"	CJE0741	ppa	Cj0638c	ppa	CCO0718	ppa	-1	-1	0	1	1	1	0	1	1	1	inorganic pyrophosphatase	inorganic pyrophosphatase	inorganic pyrophosphatase
"CJ_10000112"	CJE0742	adk	Cj0639c	adk	CCO0719	-	1	1	1	1	1	1	1	1	1	1	adenylate kinase	adenylate kinase	adenylate kinase Cj0639c
"CJ_10000114"	CJE0743	aspS	Cj0640c	aspS	CCO0720	aspS	1	1	1	1	1	1	1	1	1	1	aspartyl-tRNA synthetase	aspartyl-tRNA synthetase	aspartyl-tRNA synthetase
"CJ_10000122"	CJE0744	-	Cj0641	-	CCO0721	-	1	1	1	1	1	1	1	0	1	1	hypothetical protein	ATP-NAD kinase, putative	inorganic polyphosphate/ATP-NAD kinase
"CJ_10000123"	CJE0745	recN	Cj0642	recN	CCO0722	-	1	-1	1	1	1	1	1	1	1	1	putative DNA repair protein	DNA repair protein RecN	DNA repair protein RecN, putative
"CJ_10000124"	CJE0746	-	Cj0643	-	CCO0723	-	1	1	1	1	1	1	1	1	1	1	putative two-component response regulator	response regulator/GGDEF domain protein	response regulator/GGDEF domain protein,
"CJ_10000125"	CJE0747	-	Cj0644	-	CCO0724	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hydrolase, TatD family	hydrolase, TatD family
"CJ_10000126"	CJE0748	-	Cj0645	-	CCO0725	dniR	1	1	1	1	1	1	1	1	1	1	putative secreted transglycosylase	membrane-bound lytic murein transglycosylase D,	regulatory protein DniR

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"CJ_10000127"	CJE0749	rfpA	Cj0646	-	CCO0726	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	rare lipoprotein A	lipoprotein, rare lipoprotein A family
"CJ_10000128"	CJE0750	-	Cj0647	-	CCO0727	-	1	0	1	1	1	0	1	1	1	1	hypothetical protein	phosphatase, YrbI family	phosphatase, YrbI family
"CJ_10000130"	CJE0751	-	Cj0648	-	CCO0728	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000132"	CJE0752	-	Cj0649	-	CCO0729	-	0	1	1	1	1	1	0	1	1	1	hypothetical protein	OstA family protein	ostA family protein
"CJ_10000134"	CJE0753	-	Cj0650	-	CCO0730	-	0	0	1	1	1	1	1	1	1	1	putative ATP /GTP binding protein	GTP-binding protein	GTP-binding protein EngB
"CJ_10000141"	CJE0754	-	Cj0651	-	CCO0731	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0651
"CJ_10000142"	CJE0755	pbpC	Cj0652	pbpC	CCO0732	-	1	1	1	1	1	1	1	1	1	1	penicillin-binding protein	penicillin-binding protein 2	penicillin-binding protein 2 (pbp2)
"CJ_10000143"	CJE0756	-	Cj0653c	-	CCO0733	-	1	0	1	1	1	1	1	1	1	1	putative aminopeptidase	peptidase, M24 family	peptidase, M24 family protein
"opCjV0100000750"	CJE0757	-	Cj0654c	-	-	-	1	1	1	1	1	1	1	1	1	1	pseudogene	di-/tripeptide transporter	-
"CJ_10000144"	CJE0758	-	Cj0654c	-	-	-	-1	-1	-1	1	1	1	1	1	1	1	pseudogene	pseudogene	-
"opCjV0100000166"	CJE0759	-	Cj0654c	-	-	-	0	0	-1	1	0	1	1	0	1	1	pseudogene	hypothetical protein	-
"CJ_10000145"	CJE0761	-	Cj0659c	-	CCO0734	-	1	1	0	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0659c , putative
"CJ_10000146"	CJE0762	-	Cj0660c	-	CCO0735	-	1	0	-1	1	0	1	1	1	1	1	putative transmembrane protein	hypothetical protein	probable transmembrane protein Cj0660c
"CJ_10000147"	CJE0763	era	Cj0661c	era	CCO0736	era	0	1	1	1	1	1	0	1	1	1	GTP-binding protein ERA homolog	GTP-binding protein Era	GTP-binding protein Era
"CJ_10000148"	CJE0764	hsIU	Cj0662c	hsIU	CCO0737	hsIU	0	0	1	1	1	1	1	1	1	1	putative heat shock protein	ATP-dependent protease ATP-binding subunit	heat shock protein HslIVU, ATPase subunit HslIU
"CJ_10000150"	CJE0765	hsIV	Cj0663c	hsIV	CCO0738	hsIV	1	1	1	1	1	1	1	1	1	1	putative heat shock protein	ATP-dependent protease peptidase subunit	heat shock protein
"CJ_10000152"	CJE0766	rpII	Cj0664c	rpII	CCO0739	rpII	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L9	50S ribosomal protein L9	ribosomal protein L9
"CJ_10000160"	CJE0767	argG	Cj0665c	argG	CCO0740	argG	1	1	1	1	1	1	1	1	1	1	argininosuccinate synthase	argininosuccinate synthase	argininosuccinate synthase
"CJ_10000161"	CJE0768	-	Cj0667	-	CCO0741	-	1	0	1	1	0	1	1	1	1	1	hypothetical protein	S4 domain protein	S4 domain protein
"CJ_10000162"	CJE0769	-	Cj0668	-	CCO0742	-	1	1	1	1	1	1	1	1	1	1	putative ATP /GTP-binding protein	conserved hypothetical protein TIGR00150	conserved hypothetical protein TIGR00150
"opCcV0100000267"	CJE0770	-	Cj0669	-	CCO0743	-	1	1	1	1	-1	1	1	1	1	1	ABC-transporter ATP-binding protein	ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein
"opCcV0100002014"	CJE0771	rpoN	Cj0670	rpoN	CCO0744	-	1	1	1	1	-1	1	1	1	1	-1	RNA polymerase sigma-54 factor	DNA-directed RNA polymerase subunit N	RNA polymerase sigma-54 factor, putative
"CJ_10000163"	CJE0772	dcuB	Cj0671	dcuB	CCO0745	-	1	1	1	1	1	1	1	1	1	1	putative anaerobic C4-dicarboxylate transporter	anaerobic C4-dicarboxylate transporter	anaerobic C4-dicarboxylate membrane transporter
"CJ_10000164"	CJE0773	-	Cj0672	-	-	-	1	1	1	1	1	1	1	1	1	1	Putative periplasmic protein	hypothetical protein	-
"CJ_10000166"	CJE0774	-	Cj0676	kdpA	-	-	1	1	1	1	1	1	1	1	1	1	pseudogene	pseudogene	pseudogene
"CJ_10000168"	CJE0775	-	Cj0677	kdpB	-	-	1	1	1	1	1	1	1	1	1	1	potassium-transporting ATPase B chain	pseudogene	-
"CJ_10000170"	CJE0776	-	Cj0678	kdpC	-	-	1	1	1	1	0	1	1	0	1	1	pseudogene	potassium-transporting ATPase, C subunit,	-
"CJ_10000178"	CJE0777	-	Cj0679	-	-	-	1	1	1	1	1	1	1	1	1	1	truncated KdpD protein	pseudogene	-
"CJ_10000179"	CJE0778	uvrB	Cj0680c	uvrB	CCO0746	uvrB	0	0	1	1	1	1	1	1	1	1	excinuclease ABC subunit B	excinuclease ABC subunit B	excinuclease ABC, B subunit
"CJ_10000180"	CJE0779	-	Cj0681	-	CCO0747	-	1	1	1	1	0	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000181"	CJE0780	-	Cj0682	-	CCO0748	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000182"	CJE0781	-	Cj0683	-	CCO0749	-	0	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	prepilin-like protein

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"CJ_10000183"	CJE0782	priA	Cj0684	priA	CCO0750	priA	1	1	1	1	0	1	1	1	1	1	putative primosomal protein N'	primosome assembly protein PriA	primosomal protein N'
"opCjjV010000198"	CJE0783	-	Cj0685c	-	-	-	1	1	0	1	1	1	0	0	1	1	possible sugar transferase	invasion phenotype protein	-
"CJ_10000184"	CJE0785	ispG	Cj0686	gcpE	CCO0752	ispG	1	1	1	1	1	1	1	1	1	1	gcpE protein homolog	4-hydroxy-3-methylbut-2-en-1-yl diphosphate	1-hydroxy-2-methyl-2-(E)-butenyl 4-diphosphate
"CJ_10000185"	CJE0786	flgH	Cj0687c	flgH	CCO0753	flgH	0	0	0	1	1	1	1	1	1	1	putative flagellar L-ring protein precursor	flagellar L-ring protein precursor	flagellar L-ring protein FlgH
"CJ_10000186"	CJE0787	pta	Cj0688	pta	CCO0754	carB	1	-1	0	0	1	1	0	1	1	0	putative phosphate acetyltransferase	phosphate acetyltransferase	carbamoyl-phosphate synthase, large subunit
"CJ_10000187"	CJE0788	ackA	Cj0689	ackA	CCO0755	ackA	1	1	1	1	1	1	1	1	1	1	acetate kinase	acetate kinase	acetate kinase
"CJ_10000188"	CJE0789	-	Cj0690c	-	-	-	1	1	-1	1	0	-1	1	-1	1	1	possible restriction /modification enzyme	hypothetical protein	-
"CJ_10000189"	CJE0790	-	Cj0691	-	CCO0756	-	1	1	1	1	1	1	0	0	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0691
"CJ_10000190"	CJE0791	-	Cj0692c	-	CCO0757	-	0	1	1	1	1	1	0	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0692c
"CJ_10000191"	CJE0792	mraW	Cj0693c	-	CCO0758	mraW	1	1	1	1	1	1	1	1	1	1	hypothetical protein	S-adenosyl-methyltransferase	S-adenosyl-methyltransferase MraW
"CJ_10000192"	CJE0793	-	Cj0694	-	CCO0759	-	1	1	1	1	1	1	-1	1	1	1	putative periplasmic protein	peptidyl-prolyl cis-trans isomerase D, homolog	conserved hypothetical secreted protein,
"CJ_10000193"	CJE0794	ftsA	Cj0695	ftsA	CCO0760	ftsA	0	-1	1	1	1	1	1	1	1	1	cell division protein ftsA	cell division protein FtsA	cell division protein FtsA
"CJ_10000194"	CJE0795	ftsZ	Cj0696	ftsZ	CCO0761	ftsZ	1	1	1	1	1	-1	-1	1	1	1	cell division protein ftsZ	cell division protein FtsZ	cell division protein FtsZ
"CJ_10000195"	CJE0796	-	Cj0697	flgG2	CCO0762	flgG_1	1	1	1			1				-1	putative flagellar basal-body rod protein	flagellar basal-body rod protein	flagellar basal-body rod protein (flgG)
"CJ_10000196"	CJE0797	flgG	Cj0698	flgG	CCO0763	flgG	1	0	1	1	1	1	1	1	1	1	flagellar basal-body rod protein	flagellar basal-body rod protein FlgG	flagellar basal-body rod protein (flgG)
"CJ_10000204"	CJE0798	glnA	Cj0699c	glnA	CCO0765	glnA	1	1	0	1	1	1	0	1	1	1	glutamine synthetase	glutamine synthetase, type I	glutamine synthetase, type I
"opCcV0100001719"	CJE0799	-	-	-	CCO0766	-	-1	0	1	-1	1	1	0	1	-1	-1	-	hypothetical protein	conserved hypothetical protein
"CJ_10000206"	CJE0800	-	Cj0700	-	CCO0766	-	0	1	0	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000209"	CJE0801	-	Cj0701	-	CCO0767	-	-1	-1	-1	-1	-1	-1	1	1	-1	1	putative protease	peptidase, U32 family	probable proteinase Cj0701
"CJ_10000212"	CJE0802	purE	Cj0702	purE	CCO0768	purE	1	1	-1	-1	-1	1	0	1	1	-1	phosphoribosylaminoimidazole carboxylase	phosphoribosylaminoimidazole carboxylase	phosphoribosylaminoimidazole carboxylase,
"CJ_10000214"	CJE0803	-	Cj0703	-	CCO0769	-	1	0	1	1	1	1	1	0	1	1	hypothetical protein	pseudogene	conserved hypothetical protein
"CJ_10000216"	CJE0804	glyQ	Cj0704	glyQ	CCO0770	glyQ	0	1	1	1	1	1	0	1	1	1	glycyl-tRNA synthetase alpha chain	glycyl-tRNA synthetase alpha subunit	glycyl-tRNA synthetase, alpha subunit
"CJ_10000218"	CJE0805	-	Cj0705	-	CCO0771	-	0	1	1	1	1	1	1	0	1	1	hypothetical protein	conserved hypothetical protein TIGR00486	conserved hypothetical protein TIGR00486
"CJ_10000220"	CJE0806	-	Cj0706	-	CCO0772	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000222"	CJE0807	waaA	Cj0707	kdtA	CCO0773	waaA	-1	1	1	1	1	1	1	1	1	1	3-deoxy-D-manno-octulosonic-acid transferase	3-deoxy-D-manno-octulosonic-acid transferase	3-deoxy-d-manno-octulosonic-acid transferase
"CJ_10000223"	CJE0808	-	Cj0708	-	CCO0774	-	1	1	1	1	1	1	1	1	1	1	putative ribosomal pseudouridine synthase	RNA pseudouridylation synthase family protein	ribosomal large subunit pseudouridine synthase
"CJ_10000234"	CJE0809	ffh	Cj0709	ffh	CCO0775	ffh	1	1	1	1	1	1	1	1	1	1	signal recognition particle protein	signal recognition particle protein	signal recognition particle protein
"opCcV0100000480"	CJE0810	rpsP	Cj0710	rpsP	CCO0776	rpsP		0	0	1	1	-1	-1	1	1	1	30S ribosomal protein S16	30S ribosomal protein S16	ribosomal protein S16
"CJ_10000235"	CJE0811	-	Cj0711	-	CCO0777	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000238"	CJE0812	rimM	Cj0712	rimM	CCO0778	-	0	0	1	1	1	1	1	1	1	1	putative 16S rRNA processing protein	16S rRNA processing protein RimM	16S rRNA processing protein RimM, putative

Appendix

"CJ_10000240"	CJE0813	trmD	Cj0713	trmD	CCO0779	trmD	0	1	0	1	1	1	1	1	1	1	1	tRNA (guanine-N1)-methyltransferase	tRNA (guanine-N(1))-methyltransferase	tRNA (guanine-N1)-methyltransferase
"CJ_10000242"	CJE0814	rplS	Cj0714	rplS	CCO0780	rplS	-1	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L19	50S ribosomal protein L19	ribosomal protein L19
"CJ_10000244"	CJE0815	-	Cj0715	-	CCO0781	-	1	1	1	1	1	1	1	1	1	1	1	trnthyretin-like periplasmic protein	trnthyretin-like protein	trnthyretin family protein
"CJ_10000246"	CJE0816	-	Cj0716	-	CCO0782	-	0	1	1	1	1	1	1	1	1	1	1	putative phospho-2-dehydro-3-deoxyheptonate	3-deoxy-7-phosphoheptonate synthase	phospho-2-dehydro-3-deoxyheptonate aldolase
"CJ_10000247"	CJE0817	-	Cj0717	-	CCO0783	-	1	1	1	-1	1	1	-1	-1	1	-1	-1	hypothetical protein	hypothetical protein	arsC family protein
"CJ_10000249"	CJE0818	dnaE	Cj0718	dnaE	CCO0784	dnaE	0	1	1	1	1	1	1	1	1	1	1	DNA polymerase III, alpha chain	DNA polymerase III subunit alpha	DNA polymerase III, alpha subunit
"CJ_10000258"	CJE0819	-	Cj0719c	-	CCO0785	-	1	1	1	1	1	1	1	1	0	1	1	hypothetical protein	conserved hypothetical protein TIGR00044	conserved hypothetical protein TIGR00044
"CJ_10000260"	CJE0820	flaC	Cj0720c	flaC	CCO0786	-	1	1	1	1	1	1	1	1	1	1	1	flagellin	flagellin subunit protein FlaC	flagellin
"CJ_10000262"	CJE0821	-	Cj0721c	-	CCO0787	-	1	0	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0721c
"CJ_10000265"	CJE0822	-	Cj0722c	-	CCO0788	hemG	1	1	1	1	1	1	1	1	1	1	1	putative DNA methylase	modification methylase, HemK family	protoporphyrinogen oxidase (hemK)
"CJ_10000267"	CJE0823	-	Cj0723c	-	CCO0789	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane zinc-metalloprotease	peptidase, M48 family	zinc-metallo protease (YJR117W)
"CJ_10000269"	CJE0824	-	Cj0724	-	CCO0790	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000271"	CJE0825	mogA	Cj0725c	mog	CCO0791	-	0	1	1	1	1	1	1	1	1	1	1	molybdopterin biosynthesis protein	molybdenum cofactor biosynthesis protein	molybdopterin biosynthesis protein Cj0725c
"CJ_10000272"	CJE0826	corA	Cj0726c	corA	CCO0792	corA	1	1	1	1	1	1	1	0	1	1	1	magnesium and cobalt transport protein	magnesium and cobalt transport protein CorA	magnesium and cobalt transport protein CorA
"CJ_10000274"	CJE0827	-	Cj0727	-	CCO0793	-	0	0	1	1	1	1	1	1	-1	1	1	putative periplasmic solute-binding protein	ABC transporter, periplasmic substrate-binding	ABC transporter, periplasmic binding protein
"CJ_10000276"	CJE0828	-	Cj0728	-	CCO0794	-	1	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0728
"CJ_10000287"	CJE0829	-	Cj0729	-	CCO0795	-	0	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000289"	CJE0830	-	Cj0730	-	CCO0796	-	1	1	1	1	1	1	1	1	1	1	1	putative ABC transport system permease	ABC transporter, permease protein	probable ABC transport system permease Cj0730
"CJ_10000292"	CJE0831	-	Cj0731	-	CCO0797	-	0	0	0	1	1	1	1	1	1	1	1	putative ABC transport system permease	ABC transporter, permease protein	permease protein
"CJ_10000295"	CJE0832	-	Cj0732	-	CCO0798	potA	1	1	1	1	1	1	1	1	1	1	1	ABC transport system ATP-binding protein	ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein
"CJ_10000297"	CJE0833	-	Cj0733	-	CCO0799	-	0	0	1	1	1	1	1	1	1	1	1	hypothetical protein	HAD-superfamily subfamily IB hydrolase	HAD-superfamily subfamily IB hydrolase,
"CJ_10000299"	CJE0834	cjaC	Cj0734c	hisJ	CCO0800	-	1	1	0	1	1	1	1	1	1	1	1	histidine-binding protein precursor	CjaC	histidine-binding protein precursor
"CJ_10000300"	CJE0835	-	Cj0735	-	-	-	0	0	1	1	1	1	1	1	0	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10000302"	CJE0836	-	Cj0736	-	-	-	0	1	0	-1	1	1	0	1	1	-1	-1	hypothetical protein	pseudogene	-
"CJ_10000304"	CJE0837	-	Cj0737	-	-	-	1	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	haemagglutination domain protein	-
"CJ_10000306"	CJE0838	-	Cj0738	-	-	-	1	1	1	1	1	1	1	-1	-1	-1	-1	hypothetical protein	hypothetical protein	-
"opCjV0100001024"	CJE0839	-	Cj0741	-	-	-	1	1	0	1	1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10000910"	CJE0840	-	Cj0741	-	-	-	1	1	1	1	-1	1	1	-1	1	1	1	hypothetical protein	hypothetical protein	-
"opCjV0100000193"	CJE0841	-	Cj0742	-	-	-	1	1	1	1	1	-1	1	1	1	1	1	pseudogene	hypothetical protein	-
"opCjV010000"	CJE0842	-	Cj0742	-	-	-	-1	1	0	1	1	-1	0	1	1	1	1	pseudogene	hypothetical protein	-

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"CJ_10000911"	CJE0843	-	Cj0742	-	-	-	1	1	1	-1	-1	1	1	-1	1	-1	pseudogene	hypothetical protein	-	
"CJ_10000914"	CJE0844	-	Cj0752	-	-	-	1	1	0	1	1	1	0	1	1	1	pseudogene	pseudogene	-	
"CJ_10000916"	CJE0845	tonB	Cj0753c	tonB3	CCO0809	-	1	-	1	-1	0	-1	1	-1	1	1	tonB transport protein	TonB	TonB	
"opCjV010000226"	CJE0846	-	-	-	-	-	0	1	-1	1	1			1	1	0	-	hypothetical protein	-	
"CJ_10000919"	CJE0847	cfrA	Cj0755	cfrA	CCO0810	-	1	1	1	1	-1	1	-1	1	-1	-1	putative iron uptake protein	ferric receptor CfrA	ferric receptor CfrA	
"CJ_10000922"	CJE0848	hrcA	Cj0757	hrcA	CCO0811	-	1	1	1	1	-1	-1	1	-1	-1	1	putative heat shock regulator	heat-inducible transcription repressor	probable heat shock regulator Cj0757	
"CJ_10000935"	CJE0849	grpE	Cj0758	grpE	CCO0812	grpE	1	1	1	1	-1	1	1	-1	1	1	heat shock protein grpE	co-chaperone protein GrpE	co-chaperone GrpE	
"CJ_10000936"	CJE0850	dnaK	Cj0759	dnaK	CCO0813	-	1	1	1	1	-1	1	1	0	1	1	heat shock protein dnaK	molecular chaperone DnaK	heat shock protein dnaK Cj0759	
"CJ_10000937"	CJE0851	-	Cj0760	-	CCO0816	-	1	1	1	1	0	1	1	0	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000938"	CJE0852	-	Cj0761	-	CCO0817	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000939"	CJE0853	aspC	Cj0762c	aspB	CCO0818	-	1	1	1	1	-1	1	1	-1	1	1	aspartate aminotransferase	aspartate aminotransferase	probable aspartate transaminase Cj0762c	
"CJ_10000940"	CJE0854	cysE	Cj0763c	cysE	CCO0819	-	1	1	1	1	-1	1	1	0	1	-1	serine acetyltransferase	serine acetyltransferase	serine O-acetyltransferase	
"CJ_10000941"	CJE0855	speA	Cj0764c	speA	CCO0820	speA	1	1	1	1	0	1	1	1	1	1	biosynthetic arginine decarboxylase	arginine decarboxylase	arginine decarboxylase	
"CJ_10000943"	CJE0856	hisS	Cj0765c	hisS	CCO0821	hisS	1	1	1	1	1	1	1	-1	-1	-1	histidyl-tRNA synthetase	histidyl-tRNA synthetase	histidyl-tRNA synthetase	
"CJ_10000946"	CJE0857	tmk	Cj0766c	tmk	CCO0822	tmk	1	1	1	1	-1	-1	1	0	1	1	putative thymidylate kinase	thymidylate kinase	thymidylate kinase	
"CJ_10000949"	CJE0858	coaD	Cj0767c	kdtB	CCO0823	coaD	0	1	1	1	1	1	1	1	1	1	3-deoxy-D-manno- octulosonic-acid transferase	phosphopantetheine adenylyltransferase	panetheine-phosphate adenylyltransferase	
"CJ_10000965"	CJE0859	-	Cj0768c	-	CCO0824	ubiX	1	1	1		-1		-1			-1	putative decarboxylase	3-octaprenyl-4- hydroxybenzoate carboxy- lyase	phenylacrylic acid decarboxylase,	
"CJ_10000966"	CJE0860	-	Cj0769c	-	CCO0825	-	1	1	1	1	1	1	1	-1	1	-1	putative periplasmic protein	flagellar basal body P-ring biosynthesis	probable periplasmic protein Cj0769c	
"CJ_10000967"	CJE0861	-	Cj0770c	-	CCO0826	-	1	1	1	0	-1	1	1	0	1	1	putative periplasmic protein	lipoprotein, NLPA family	D-methionine-binding lipoprotein MetQ	
"CJ_10000968"	CJE0862	-	Cj0771c	-	CCO0827	-	1	1	1	1	1	1	1	-1	1	-1	putative periplasmic protein	lipoprotein, NLPA family	D-methionine-binding lipoprotein MetQ	
"CJ_10000969"	CJE0863	-	Cj0772c	-	CCO0828	-	-1	1	1	1	0	1	0	1	0	1	putative periplasmic protein	lipoprotein, NLPA family	D-methionine-binding lipoprotein MetQ	
"CJ_10000970"	CJE0864	-	Cj0773c	-	CCO0829	-	-1	0	0	1	1	1	-1	1	1	1	binding-protein dependent transport system	ABC transporter, permease protein	ABC transporter, permease component CAC0985	
"CJ_10000971"	CJE0865	-	Cj0774c	-	CCO0830	-	1	1	1	-1	-1		1	-1	1	-1	ABC transport system ATP- binding protein	ABC transporter, ATP- binding protein	ABC transporter, ATP- binding protein	
"CJ_10000973"	CJE0866	valS	Cj0775c	valS	CCO0831	valS	1	1	1	0	-1	1	1	-1	1	1	valyl-tRNA synthetase	valyl-tRNA synthetase	valyl-tRNA synthetase	
"CJ_10000975"	CJE0867	-	Cj0776c	-	CCO0832	-	1	1	1	1	0	1	1	0	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0776c	
"CJ_10000978"	CJE0868	-	Cj0777	-	CCO0833	rep	1	1	1	1	-1	1	1	-1	1	1	putative ATP-dependent DNA helicase	ATP-dependent DNA helicase, putative	rep helicase, single-stranded DNA-dependent	
"CJ_10000994"	CJE0869	-	Cj0778	peb2	CCO0834	-	0	1	1	1	1	1	0	1	0	1	major antigenic peptide PEB2	major antigenic peptide PEB2	accessory colonization factor AcfC VC0841	
"CJ_10000995"	CJE0870	tpx	Cj0779	tpx	CCO0837	tpx	1	1	1	1	1	1	1	1	1	0	probable thiol peroxidase	thiol peroxidase	thiol peroxidase	
"CJ_10000996"	CJE0871	napA	Cj0780	napA	CCO0838	napA	1	1	1	1	1	1	1	1	1	1	periplasmic nitrate reductase	periplasmic nitrate reductase, large subunit	periplasmic nitrate reductase, large subunit	
"CJ_10000997"	CJE0872	napG	Cj0781	napG	CCO0839	-	1	1	1	1	1	1	1	1	1	1	putative ferredoxin	quinol dehydrogenase periplasmic component	probable ferredoxin Cj0781	
"CJ_10000998"	CJE0873	napH	Cj0782	napH	CCO0840	-	1	1	1	1	-1	1	1	-1	1	-1	putative ferredoxin	quinol dehydrogenase membrane component	iron-sulfur cluster-binding protein napH	

"CJ_10000999"	CJE0874	napB	Cj0783	napB	CCO0841	-	1	1	1	1	-1	1	1	-1	1	-1	periplasmic nitrate reductase small subunit	periplasmic nitrate reductase, small subunit	NapB periplasmic nitrate reductase
"CJ_10001000"	CJE0875	napL	Cj0784	-	CCO0842	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	NapL	probable periplasmic protein Cj0784
"CJ_10001002"	CJE0876	napD	Cj0785	napD	CCO0843	-	1	1	1	1	-1	1	-1	0	1	1	possible napD protein homolog	NapD	probable napD protein homolog Cj0785
"CJ_10001005"	CJE0877	-	Cj0786	-	CCO0844	-	0	1	1	1	-1	1	1	-1	1	1	small hydrophobic protein	hypothetical protein	small hydrophobic protein Cj0786 -related
"CJ_10001008"	CJE0878	-	Cj0787	-	CCO0848	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001027"	CJE0879	-	Cj0788	-	CCO0849	-	1	1	1	-1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001029"	CJE0880	-	Cj0789	-	CCO0850	cca	0	1	1	1	1	1	1	1	1	1	putative RNA nucleotidyltransferase	polyA polymerase family protein	tRNA nucleotidyltransferase
"CJ_10001031"	CJE0881	purU	Cj0790	purU	CCO0851	purU	0	1	-1	0	1	0	-1	0	1	-1	formyltetrahydrofolate deformylase	formyltetrahydrofolate deformylase	formyltetrahydrofolate deformylase
"CJ_10001033"	CJE0882	-	Cj0791c	-	CCO0965	-	1	0	1	1	1	1	1	1	1	1	putative aminotransferase	aminotransferase, putative	nifS-like protein
"CJ_10001034"	CJE0883	-	Cj0792	-	CCO0966	-	1	1	1	-1	1	-1	0	-1	-1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001035"	CJE0884	-	Cj0793	-	CCO0967	-	0	1	0	0	1	1	1	0	-1	-1	signal transduction histidine kinase	sensor histidine kinase	signal-transducing protein, histidine kinase
"CJ_10001037"	CJE0885	-	Cj0794	-	CCO0873	-	1	1	1	1	1	1	-1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001039"	CJE0886	-	Cj0795c	murF	CCO0854	murF	1	1	1	1	-1	1	1	0	-1	1	putative	Mur ligase family protein	UDP-MurNac-pentapeptide presynthetase (murF)
"CJ_10001041"	CJE0887	-	Cj0796c	-	CCO0855	-	1	1	1	1	1	1	1	-1	1	1	hypothetical protein	hypothetical protein	2-hydroxy-6-oxohepta-2,4-dienoate hydrolase
"CJ_10001043"	CJE0888	-	Cj0797c	-	CCO0856	-	1	1			-1	1	1	-1	1		hypothetical protein	prevent-host-death family protein	prevent-host-death family protein
"CJ_10001055"	CJE0889	ddl	Cj0798c	ddlA	CCO0857	-	1	1	1	1	1	1	1	1	1	1	putative D-alanine--D-alanine ligase	D-alanylalanine synthetase	D-alanine--D-alanine ligase
"CJ_10001057"	CJE0890	ruvA	Cj0799c	ruvA	CCO0858	ruvA	1	1	1	1	-1	1	1	-1	-1	1	putative Holliday junction DNA helicase	Holliday junction DNA helicase RuvA	Holliday junction DNA helicase RuvA
"CJ_10001059"	CJE0891	-	Cj0800c	-	CCO0859	-	1	1	1	1	-1	1	1	1	0	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001061"	CJE0892	mviN	Cj0801	-	CCO0860	mviN	0	1	0	1	1	1	1	1	1	1	integral membrane protein (MViN homolog)	integral membrane protein MviN	integral membrane protein MviN
"CJ_10001062"	CJE0893	cysS	Cj0802	cysS	CCO0861	cysS	1	1	1	1	1	1	1	1	1	1	cysteinyI-tRNA synthetase	cysteinyI-tRNA synthetase	cysteinyI-tRNA synthetase
"CJ_10001064"	CJE0894	-	Cj0803	msbA	CCO0862	msbA	1	1	1	0	1	1	1	-1	1	1	lipid export ABC transport protein	ABC transporter, ATP-binding protein/permease	multidrug resistance protein (msbA)
"CJ_10001066"	CJE0895	pyrD	Cj0804	pyrD	CCO0863	pyrD	1	1	1	1	1	1	1	1	1	1	dihydroorotate dehydrogenase	dihydroorotate dehydrogenase	dihydroorotate dehydrogenase
"CJ_10001068"	CJE0896	-	Cj0805	-	CCO0864	-	1	1	1	1	0	1	1	1	1	1	putative zinc protease	peptidase, M16 family	protease (pqqE)
"CJ_10001070"	CJE0897	dapA	Cj0806	dapA	-	-	1	1	1	1	-1	1	1	1	1	-1	dihydrodipicolinate synthase	dihydrodipicolinate synthase	-
"CJ_10001072"	CJE0898	-	Cj0807	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative oxidoreductase	7-alpha-hydroxysteroid dehydrogenase	-
"CJ_10001091"	CJE0899	-	Cj0808c	-	-	-	1	-1	1	1	-1	0	1	-1	1	0	small hydrophobic protein	hypothetical protein	-
"CJ_10001094"	CJE0900	-	Cj0809c	-	-	-	1	1	1	-1	-1	1	1	1	1	-1	putative hydrolase	metallo-beta-lactamase family protein	-
"CJ_10001097"	CJE0901	nadE	Cj0810	nadE	-	-	0	1	1	1	-1	1	1	-1	1	1	putative NH(3)-dependent NAD(+) synthetase	NAD+ synthetase	-
"CJ_10001099"	CJE0902	lpxK	Cj0811	lpxK	-	-	1	1	1	-1	0	1	1	-1	1	1	putative tetraacyldisaccharide 4'-kinase	tetraacyldisaccharide 4'-kinase	-

"CJ_10001101"	CJE0903	thrC	Cj0812	thrC	CCO0871	-	1	1	1	1	1	1	1	1	1	1	1	threonine synthase	threonine synthase	threonine synthase
"CJ_10001104"	CJE0904	kdsB	Cj0813	kdsB	CCO0872	kdsB	1	1	1	1	0	1	1	1	1	1	1	3-deoxy-manno- octulosonate cytidyltransferase	3-deoxy-manno- octulosonate cytidyltransferase	3-deoxy-D-manno- octulosonate
"CJ_10001107"	CJE0905	-	Cj0814	-	CCO0873	-	1	1	1	1	-1	1	1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001112"	CJE0906	-	Cj0817	glnH	CCO0876	glnH	1	1	1	1	1	1	1	1	1	1	1	glutamine-binding periplasmic protein	amino acid-binding protein	glutamine ABC transporter, periplasmic
"CJ_10001133"	CJE0907	fliP	Cj0820c	fliP	CCO0880	-	1	1	1	1	-1	-1	1	-1	1	1	1	flagellar biosynthesis protein	flagellar biosynthesis protein	flagellar biosynthesis protein Cj0820c
"CJ_10001137"	CJE0908	glmU	Cj0821	glmU	CCO0881	glmU	1	1	1	1	1	1	1	1	1	1	1	UDP-N-acetylglucosamine pyrophosphorylase	UDP-N-acetylglucosamine pyrophosphorylase	UDP-N-acetylglucosamine pyrophosphorylase
"CJ_10001140"	CJE0909	coaBC	Cj0822	dfp	CCO0882	coaBC	1	1	1	-1	0	1	1	1	0	1	1	DNA /pantothenate metabolism flavoprotein	phosphopantothenoylcysteine	phosphopantothenoylcysteine
"CJ_10001143"	CJE0910	-	Cj0823	-	CCO0883	-	1	1	1	1	-1	-1	1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001146"	CJE0911	uppS	Cj0824	uppS	CCO0884	uppS	1	1	1	1	1	1	1	1	1	1	1	putative undecaprenyl diphosphate synthase	undecaprenyl diphosphate synthase	undecaprenyl diphosphate synthase
"CJ_10001148"	CJE0912	-	Cj0825	-	CCO0885	-	1	1	1	1	1	1	1	1	1	1	1	putative processing peptidase	peptidase, A24 family	type IV prepilin peptidase, probable , putative
"CJ_10001150"	CJE0913	-	Cj0826	-	CCO0886	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	conserved hypothetical integral membrane
"CJ_10001152"	CJE0914	truA	Cj0827	truA	CCO0887	truA	1	1	1	1	-1	1	1	0	1	1	1	putative tRNA pseudouridine synthase	tRNA pseudouridine synthase A	tRNA pseudouridine synthase A
"CJ_10001169"	CJE0915	ilvA	Cj0828c	ilvA	CCO0888	ilvA	1	1	1	1	1	1	1	0	1	1	1	threonine dehydratase biosynthetic	threonine dehydratase	threonine dehydratase
"CJ_10001171"	CJE0916	-	Cj0829c	-	CCO0889	-	1	1	1	1	1	0	1	1	1	1	1	hypothetical protein	CoA-binding domain protein	CoA-binding domain protein
"CJ_10001175"	CJE0917	-	Cj0830	-	CCO0890	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0830
"CJ_10001179"	CJE0918	trmA	Cj0831c	trmA	CCO0891	-	1	1	0	1	1	1	1	1	1	1	1	tRNA (uracil-5-)- methyltransferase	tRNA (uracil-5-)- methyltransferase	tRNA (uracil-5-)- methyltransferase
"CJ_10001182"	CJE0919	-	Cj0832c	-	CCO0892	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	Na+/H+ antiporter family protein	probable integral membrane protein Cj0832c
"CJ_10001185"	CJE0920	-	Cj0833c	-	CCO0893	-	1	1	1	1	1	-1	1	-1	1	1	1	oxidoreductase	oxidoreductase, short chain	oxidoreductase, short chain
"CJ_10001187"	CJE0921	-	Cj0834c	-	CCO0894	-	1	1	1	-1	0	1	1	1	1	1	1	ankyrin repeat-containing possible periplasmic	ankyrin repeat protein	conserved hypothetical protein
"CJ_10001190"	CJE0922	acnB	Cj0835c	acnB	CCO0895	acnB	0	1	1	-1	0	1	1	1	-1	1	1	aconitate hydratase	aconitate hydratase	aconitate hydratase 2
"CJ_10001192"	CJE0923	ogt	Cj0836	ogt	CCO0896	ogt	0	1	1	1	1	1	1	1	1	1	1	methylated-DNA--protein- cysteine	methylated-DNA--protein- cysteine	methylated-DNA--protein- cysteine
"CJ_10001195"	CJE0924	-	Cj0837c	-	CCO0897	-	1	1	1	1	1	1	0	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001215"	CJE0925	metG	Cj0838c	metS	CCO0898	metS	1	0	1	1	-1	1	1	1	1	1	1	methionyl-tRNA synthetase	methionine--tRNA ligase	methionyl-tRNA synthetase
"CJ_10001218"	CJE0926	-	Cj0839c	-	CCO0899	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001222"	CJE0927	fbp	Cj0840c	fbp	CCO0900	fbp	1	1	1	1	1	1	1	1	1	1	1	putative fructose-1,6- biphosphatase	fructose-1,6-biphosphatase	fructose-1,6-biphosphatase
"CJ_10001226"	CJE0928	mobB	Cj0841c	-	CCO0901	mobB	1	1	1	1	0	1	1	0	1	1	1	putative ATP/GTP binding protein	molybdopterin-guanine dinucleotide biosynthesis	molybdopterin-guanine dinucleotide biosynthesis
"CJ_10001229"	CJE0929	-	Cj0842	-	CCO0902	-	1	1	1	1	-1	1	1	-1	1	1	1	putative lipoprotein	lipoprotein, putative	lipoprotein, putative
"CJ_10001231"	CJE0930	-	Cj0843c	-	CCO0903	-	1	1	1	1	1	1	1	1	1	1	1	putative secreted transglycosylase	soluble lytic murein transglycosylase, putative	soluble lytic murein transglycosylase (slt),
"CJ_10001234"	CJE0931	-	Cj0844c	-	CCO0904	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	YGGT family protein	probable integral membrane protein Cj0844c
"CJ_10001237"	CJE0932	gltX	Cj0845c	gltX	CCO0905	gltX	0	1	1	1	1	1	-1	1	1	1	1	glutamyl-tRNA synthetase	glutamyl-tRNA synthetase	glutamyl-tRNA synthetase

"CJ_10001239"	CJE0933	-	Cj0846	-	CCO0906	-	1	1	1	1	1	1	1	0	1	1	putative integral membrane protein	Ser/Thr protein phosphatase family protein	Predicted DNA repair exonuclease
"CJ_10001244"	CJE0934	psd	Cj0847	psd	CCO0907	psd	-1	1	0	1	1	-1	1	1	1	1	putative phosphatidylserine decarboxylase	phosphatidylserine decarboxylase	phosphatidylserine decarboxylase
"CJ_10000197"	CJE0935	-	Cj0848c	-	CCO0908	-	1	1	1	1	1	1	-1	1	1	1	hypothetical protein	FlhB domain protein	FlhB domain protein
"CJ_10000198"	CJE0936	-	Cj0849c	-	CCO0909	-	-1	-1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000199"	CJE0937	-	Cj0850c	-	CCO0910	-	1	1	1		1		0	-1	1	-1	transmembrane transport protein	major facilitator family protein	major facilitator family transporter, putative
"CJ_10000200"	CJE0938	-	Cj0851c	-	CCO0911	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0851c
"CJ_10000201"	CJE0939	-	Cj0852c	-	CCO0912	-	1	1	1	-1	1	1	-1	1		1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0852c
"CJ_10000202"	CJE0940	hemL	Cj0853c	hemL	CCO0913	hemL	1	0	0	1	1	1	1	1	-1	1	glutamate-1-semialdehyde 2,1-aminomutase	glutamate-1-semialdehyde aminotransferase	glutamate-1-semialdehyde-2,1-aminomutase
"CJ_10000203"	CJE0941	-	Cj0854c	-	CCO0914	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0854c
"CJ_10000205"	CJE0942	folD	Cj0855	folD	CCO0915	folD	1	1	0	0	1	1	1	1	1	1	flagellar biosynthesis protein	folD bifunctional protein	methylene-tetrahydrofolate dehydrogenase (folD)
"CJ_10000208"	CJE0943	lepB	Cj0856	lepP	CCO0916	-	0	0	1	1	1	1	0	1	0	1	signal peptidase I	signal peptidase I	Signal peptidase I
"CJ_10000211"	CJE0944	-	Cj0857c	moeA	CCO0987	-	1	1	0	1	1			1	1	1	putative molybdopterin biosynthesis protein	molybdopterin biosynthesis MoeA protein,	molybdopterin biosynthesis protein (moeA)
"CJ_10000227"	CJE0945	murA	Cj0858c	murA	CCO0986	murA	1	1	0	0	-1	1	1	0	-1	-1	UDP-N-acetylglucosamine	UDP-N-acetylglucosamine	UDP-N-acetylglucosamine
"CJ_10000228"	CJE0946	-	Cj0859c	-	-	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10000229"	CJE0947	-	Cj0860	-	CCO0985	-	0	0	0	1	1	1	0	1	-1	1	probable integral membrane protein	integral membrane protein	membrane protein
"CJ_10000230"	CJE0948	pabA	Cj0861c	pabA	CCO0984	pabA	1	1	1	1	1	1	1	1	1	1	para-aminobenzoate synthase glutamine	para-aminobenzoate synthase glutamine	para-aminobenzoate synthase glutamine
"CJ_10000231"	CJE0949	pabB	Cj0862c	pabB	CCO0983	-	0	1	1	1	1	1	1	1	1	1	para-aminobenzoate synthase component I	para-aminobenzoate synthase glutamine	para-aminobenzoate synthetase (pabB)
"CJ_10000232"	CJE0950	-	Cj0863c	xerD	CCO0982	-	1	1	1	1	1	1	0	1	0	0	DNA recombinase	site-specific recombinase, phage integrase	DNA recombinase Cj0863c
"CJ_10000233"	CJE0951	dsbA	Cj0864	-	CCO0980	-	1	1	1	1	1	1	0	1	1	1	putative periplasmic protein	thiol:disulfide interchange protein DsbA	thiol:disulfide interchange protein, DsbA
"opCcV010000 0804"	CJE0952	dsbB	Cj0865	dsbB	CCO0981	-	1	1	1	1	1	1	1	1	-1		putative disulfide oxidoreductase	putative disulfide oxidoreductase	disulfide bond formation protein
"opCjjV010000 091"	CJE0953	-	Cj0866	ast	-	-	1	-1	1	1	1	1	1	1	1	1	pseudogene	pseudogene	-
"CJ_10000237"	CJE0954	-	Cj0872	dsbA	CCO0980	-	0	0	1	1	1	1	1	1	1	1	putative protein disulphide isomerase	thiol:disulfide interchange protein DsbA,	thiol:disulfide interchange protein, DsbA
"CJ_10000253"	CJE0955	-	Cj0874c	-	-	-	0	1	0	0	1	1	0	1	1	1	cytochrome C	pseudogene	-
"CJ_10000255"	CJE0958	-	Cj0878	-	CCO0979	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000256"	CJE0959	-	Cj0879c	-	CCO0978	-	1	1	1	1	1	1	1	1	0	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0879c
"CJ_10000257"	CJE0960	-	Cj0880c	-	CCO0977	-	1	1	1	1	1	1	0	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000259"	CJE0961	-	Cj0881c	-	CCO0976	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCcV010000 1566"	CJE0962	flhA	Cj0882c	flhA	CCO0975	flhA	1	0	1	1	1	1	-1	1	1	1	flagellar biosynthesis protein	flagellar biosynthesis protein	flagellar biosynthesis protein FlhA
"CJ_10000264"	CJE0963	-	Cj0883c	-	CCO0974	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	Rrf2 family protein, putative	rrf2 family protein (putative transcriptional
"CJ_10000280"	CJE0964	rpsO	Cj0884	rpsO	CCO0973	rpsO	0	0	1	1	1	1	0	1	0	1	30S ribosomal protein S15	30S ribosomal protein S15	ribosomal protein S15
"CJ_10000281"	CJE0965	-	Cj0886c	ftsK	CCO0972	ftsK	1	1	1	1	1	-1	1	1	1	-1	putative cell division protein	cell division protein FtsK,	cell division protein (ftsK)

"CJ_10000282"	CJE0966	-	Cj0887c	flaD	CCO0971	-	1	0	1	1	1	1	1	1	1	1	putative flagellin	putative flagellar hook-associated protein	probable flagellin Cj0887c
"CJ_10000283"	CJE0967	-	Cj0888c	-	CCO0989	-	1	0	1	1	1	1	0	1	1	1	ABC transport system ATP-binding protein	ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein
"CJ_10000284"	CJE0968	-	Cj0889c	-	CCO0990	-	1	0	1	1	1	0	1	1	1	1	putative sensory transduction histidine kinase	sensor histidine kinase	histidine protein kinase PhoR VC0720 , putative
"CJ_10000285"	CJE0969	-	Cj0890c	-	CCO0991	drrA	0	1	1	1	1	1	1	1	1	1	putative sensory transduction transcriptional	DNA-binding response regulator	response regulator DrrA
"CJ_10000286"	CJE0970	serA	Cj0891c	serA	CCO0992	serA	1	1	1	1	1	1	1	1	1	1	D-3-phosphoglycerate dehydrogenase	D-3-phosphoglycerate dehydrogenase	D-3-phosphoglycerate dehydrogenase
"CJ_10000288"	CJE0971	-	Cj0892c	-	CCO0993	-	1	0	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0892c
"CJ_10000291"	CJE0972	rpsA	Cj0893c	rpsA	CCO0994	-	1	0	1	1	1	1	1	1	1	1	30S ribosomal protein S1	30S ribosomal protein S1	ribosomal protein S1 VC1915 , putative
"CJ_10000294"	CJE0973	ispH	Cj0894c	lytB	CCO0995	ispH	0	0	1	1	1	1	1	1	0	1	lytB homolog	4-hydroxy-3-methylbut-2-enyl diphosphate	hydroxymethylbutenyl pyrophosphate reductase
"CJ_10000310"	CJE0974	aroA	Cj0895c	aroA	CCO0996	aroA	1	1	1	1	1	1	1	0	1	1	3-phosphoshikimate 1-carboxyvinyltransferase	3-phosphoshikimate 1-carboxyvinyltransferase	3-phosphoshikimate 1-carboxyvinyltransferase
"CJ_10000311"	CJE0975	pheT	Cj0896c	pheT	CCO0997	-	1	1	1	1	1	1	1	1	1	1	phenylalanyl-tRNA synthetase beta chain	phenylalanyl-tRNA synthetase beta subunit	phenylalanyl-tRNA synthetase, beta subunit,
"CJ_10000312"	CJE0976	pheS	Cj0897c	pheS	CCO0998	pheS	1	1	1	1	1	1	1	1	1	1	phenylalanyl-tRNA synthetase alpha chain	phenylalanyl-tRNA synthetase alpha subunit	phenylalanyl-tRNA synthetase, alpha subunit
"CJ_10000313"	CJE0977	-	Cj0898	-	CCO0999	-	-1	-1	1	1	1	1	1	1	1	1	HIT-family protein	HIT family protein	HIT family protein
"CJ_10000314"	CJE0978	thiJ	Cj0899c	thiJ	CCO1000	thiJ	1	1	1	1	1	1	1	0	1	1	4-methyl-5(beta-hydroxyethyl)-thiazole	4-methyl-5(B-hydroxyethyl)-thiazole	ThiJ/PfpI family protein
"CJ_10000315"	CJE0979	-	Cj0900c	-	CCO1001	-	1	1	1	1	0	1	1	1	0	-1	small hydrophobic protein	hypothetical protein	small hydrophobic protein Cj0900c -related
"CJ_10000316"	CJE0980	-	Cj0901	-	CCO1002	-	1	1	1	1	1	1	1	1	1	1	putative amino acid ABC transporter permease	amino acid ABC transporter, permease protein,	amino acid ABC transporter, permease protein
"CJ_10000317"	CJE0981	-	Cj0902	glnQ	CCO1003	glnQ	0	1	1	1	1	1	1	-1	1	1	putative glutamine transport ATP-binding	amino acid ABC transporter, ATP-binding protein	amino acid ABC transporter, ATP-binding protein
"CJ_10000319"	CJE0982	-	Cj0903c	-	CCO1004	-	1	1	1	1	1	1	1	1	1	1	putative amino-acid transport protein	amino acid carrier protein	sodium:alanine symporter family protein
"CJ_10000321"	CJE0983	-	Cj0904c	-	CCO1006	-	-1	-1	1	1	1	1	1	1	1	1	putative RNA methylase	RNA methyltransferase, TrmH family	rRNA methylase
"CJ_10000330"	CJE0984	alr	Cj0905c	alr	CCO1007	-	1	1	1	1	1	1	1	1	1	1	alanine racemase	alanine racemase	alanine racemase, putative
"CJ_10000331"	CJE0985	-	Cj0906c	-	CCO1008	-	1	1	1	1	1	-1	1	1	1	1	putative periplasmic protein	hypothetical protein	conserved hypothetical secreted protein
"CJ_10000332"	CJE0986	-	Cj0908	-	CCO1009	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0908
"CJ_10000333"	CJE0987	-	Cj0909	-	CCO1010	-	0	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0909
"CJ_10000334"	CJE0988	-	Cj0910	-	CCO1011	-	0	0	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0910
"CJ_10000335"	CJE0989	-	Cj0911	-	CCO1012	-	1	-1	-1	-1	-1	-1	1	1	1	1	putative periplasmic protein	SCO1/SenC family protein	probable periplasmic protein Cj0911
"opCcV010000950"	CJE0990	cysK	Cj0912c	cysM	CCO1013	cysK	-1	-1	0	0	1	-1	0	1	-1	-1	cysteine synthase	cysteine synthase A	cysteine synthase A
"CJ_10000336"	CJE0991	hup	Cj0913c	hupB	CCO1014	-	1	-1	1	1	1	1	1	1	1	1	DNA-binding protein HU homolog	DNA-binding protein HU	DNA-binding protein HU
"opCcV0100001041"	CJE0992	ciaB	Cj0914c	-	CCO1015	-	1	1	1	1	1	-1	-1	1	-1	-1	CiaB protein	invasion antigen B	CiaB protein Cj0914c
"CJ_10000339"	CJE0993	-	Cj0915	-	CCO1016	-	0	-1	1	1	1	1	1	1	1	1	putative hydrolase	thioesterase family protein	probable hydrolase Cj0915

"CJ_10000348"	CJE0994	-	Cj0916c	-	CCO1017	-	1	1	0	1	1	1	1	1	0	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF466)
"CJ_10000349"	CJE0995	cstA	Cj0917c	cstA	CCO1018	-	0	0	1	1	1	1	1	1	1	1	carbon starvation protein A homolog	carbon starvation protein A homolog Cj0917c	carbon starvation protein A homolog Cj0917c
"CJ_10000350"	CJE0996	prsA	Cj0918c	prsA	CCO1019	prsA	0	0	0	1	1	1	0	-1	1	1	ribose-phosphate pyrophosphokinase	ribose-phosphate pyrophosphokinase	ribose-phosphate pyrophosphokinase
"CJ_10000352"	CJE0997	-	Cj0919c	-	CCO1020	glnP	0	1	1	1	1	1	1	1	1	1	putative ABC-type amino-acid transporter	amino acid ABC transporter, permease protein	amino acid ABC transporter, permease protein
"CJ_10000354"	CJE0998	-	Cj0920c	-	CCO1021	glnP	0	1	1	1	1	1	1	1	1	1	putative ABC-type amino-acid transporter	amino acid ABC transporter, permease protein	amino acid ABC transporter, permease protein
"CJ_10000356"	CJE0999	pebA	Cj0921c	peb1A	CCO1022	glnH	1	1	1	1	1	1	1	1	1	1	probable ABC-type amino-acid transporter	amino acid ABC transporter, periplasmic amino	amino acid ABC transporter, amino acid-binding
"CJ_10000358"	CJE1000	pebC	Cj0922c	pebC	CCO1003	glnQ	0	1	1	1	1	1	0	1	1	1	ABC-type amino-acid transporter ATP-binding	amino acid ABC transporter, ATP-binding protein	amino acid ABC transporter, ATP-binding protein
"CJ_10000359"	CJE1001	cheR	Cj0923c	cheR	CCO1024	-	1	1	1	1	1	1	1	1	1	1	putative MCP protein methyltransferase	chemotaxis protein methyltransferase CheR	chemotaxis protein methyltransferase CheR,
"CJ_10000362"	CJE1002	cheB	Cj0924c	-	CCO1025	-	1	1	0	1	1	1	1	1	1	1	putative MCP protein-glutamate methylesterase	protein-glutamate methylesterase CheB	protein-glutamate methylesterase CheB
"CJ_10000365"	CJE1003	rpiB	Cj0925	rpiB	CCO1026	-	1	1	1	1	1	1	1	1	1	1	putative ribose 5-phosphate isomerase	ribose 5-phosphate isomerase B	sugar-phosphate isomerase
"CJ_10000376"	CJE1004	-	Cj0926	-	CCO1027	-	0	0	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj0926
"CJ_10000377"	CJE1005	apt	Cj0927	apt	CCO1028	apt	1	1	1	1	-1	1	1	-1	1	1	adenine phosphoribosyltransferase	adenine phosphoribosyltransferase	adenine phosphoribosyltransferase
"CJ_10000379"	CJE1006	-	Cj0928	-	CCO1029	-	0	-1	1	1	1	1	1	1	1	1	putative integral membrane protein (dedA)	DedA family protein	conserved hypothetical integral membrane
"CJ_10000381"	CJE1007	-	Cj0929	pepA	CCO1030	pepA	0	0	1	1	1	1	1	1	1	1	aminopeptidase	leucyl aminopeptidase	cytosol aminopeptidase
"CJ_10000383"	CJE1008	-	Cj0930	-	CCO1031	ychF	1	1	1	1	1	1	1	1	1	1	putative GTP-binding protein	GTP-binding protein YchF	GTP-binding protein YchF
"opCcV010000857"	CJE1009	argH	Cj0931c	argH	CCO1032	argH	1	1	1	1	1	1	1	1	1	1	argininosuccinate lyase	argininosuccinate lyase	argininosuccinate lyase
"CJ_10000385"	CJE1010	pckA	Cj0932c	pckA	CCO1033	pckA	1	1	1	1	1	1	1	0	1	1	phosphoenolpyruvate carboxykinase (ATP)	phosphoenolpyruvate carboxykinase	phosphoenolpyruvate carboxykinase (ATP)
"CJ_10000387"	CJE1011	-	Cj0933c	pycB	CCO1034	oadA	1	1	1	1	-1	1	1	1	1	1	putative pyruvate carboxylase B subunit	oxaloacetate decarboxylase, alpha subunit,	oxaloacetate decarboxylase, alpha subunit
"CJ_10000388"	CJE1012	-	Cj0934c	-	CCO1036	-	1	1	1	1	1	1	1	1	1	1	putative transmembrane transport protein	sodium transporter, putative	sodium- and chloride-dependent transporter
"CJ_10000390"	CJE1013	-	Cj0935c	-	CCO1037	-	1	1	1	1	1	1	1	0	0	1	putative transmembrane transport protein	sodium transporter, putative	sodium- and chloride-dependent transporter
"opCcV0100001238"	CJE1014	atpE	Cj0936	atpE	CCO1038	-	1	1	1	1	0	1	1	0	1	1	ATP synthase F0 sector C subunit, putative	ATP synthase subunit C	ATP synthase F0, C subunit, putative
"CJ_10000403"	CJE1015	-	Cj0937	-	CCO0852	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0937
"CJ_10000405"	CJE1016	aas	Cj0938c	aas	CCO0853	-	1	1	1	1	1	1	1	1	1	1	putative 2-acylglycerophosphoethanolamine	2-acyl-glycerophosphoethanolamine	2-acylglycerophosphoethanolamine
"CJ_10000407"	CJE1017	-	Cj0939c	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10000409"	CJE1018	-	Cj0940c	glnP	CCO0957	glnP	1	1	1	1	1	1	1	1	1	1	putative glutamine transport system permease	amino acid ABC transporter, permease protein,	amino acid ABC transporter, permease protein
"CJ_10000410"	CJE1019	-	Cj0941c	-	CCO0954	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane	permease, putative	probable integral membrane

Appendix

"CJ_10000412"	CJE1020	secA	Cj0942c	secA	CCO0953	secA	1	1	1	1	1	1	1	1	1	1	protein	protein Cj0941c	
"CJ_10000414"	CJE1021	lolA	Cj0943	-	CCO0952	-	1	1	-1	1	1	1	-1	1	-1	-1	preprotein translocase SECA subunit	translocase	preprotein translocase, SecA subunit
"CJ_10000416"	CJE1022	-	Cj0944c	-	CCO0951	-	0	1	1	1	0	1	1	1	1	1	putative periplasmic protein	outer-membrane lipoprotein carrier protein	probable periplasmic protein Cj0943
"CJ_10000421"	CJE1023	-	Cj0945c	-	CCO0950	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0944c
"CJ_10000432"	CJE1024	-	Cj0946	-	CCO0949	-	1	1	1	-1	-1				-1		putative helicase	hypothetical protein	TPR domain protein, putative
"CJ_10000433"	CJE1025	-	Cj0947c	-	CCO0947	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj0946
"CJ_10000434"	CJE1026	-	Cj0948c	-	CCO0946	-	1	0	1	1	1	1	1	1	1	1	putative hydrolase	hydrolase, carbon-nitrogen family	probable hydrolase Cj0947c
"CJ_10000436"	CJE1027	-	Cj0949c	-	CCO0945	-	1	1	1	1	1	1	1	1	1	1	putative transmembrane transport protein	cation efflux family protein	probable transmembrane transport protein
"CJ_10000437"	CJE1028	-	Cj0950c	-	CCO0944	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	peptidyl-arginine deiminase family protein	peptidyl-arginine deiminase family protein
"opCjV0100001011"	CJE1029	-	Cj0951c	-	-	-	1	1	1	1	1	1	1	1	1	1	Putative lipoprotein	heat shock protein HsIJ, putative	secreted protein involved in flagellar motility
"CJ_10000439"	CJE1030	-	Cj0951c	-	CCO0943	-	1	1	1	1	1	1	1	1	1	1	putative MCP-domain signal transduction protein	hypothetical protein	-
"CJ_10000441"	CJE1032	-	Cj0952c	-	CCO0943	-	1	1	1	1	0	1	1	1	1	1	putative MCP-domain signal transduction protein	methyl-accepting chemotaxis protein	probable membrane protein Cj0952c
"CJ_10000443"	CJE1033	purH	Cj0953c	purH	CCO0942	purH	1	1	1	1	0	1	1	1	1	1	putative membrane protein	pseudogene	probable membrane protein Cj0952c
"CJ_10000447"	CJE1034	-	Cj0954c	-	CCO0941	-	1	1	1	1	1	1	1	1	1	1	phosphoribosylaminoimidazolecarboxamide	bifunctional	phosphoribosylaminoimidazolecarboxamide
"CJ_10000450"	CJE1035	purL	Cj0955c	purL	CCO0940	purL	1	1	1	1	0	1	1	1	1	1	putative dnaJ-like protein	DnaJ domain protein	DnaJ domain protein
"CJ_10001081"	CJE1036	trmE	Cj0956c	thdF	CCO0939	trmE	1	1	1	1	1	-1	1	1	1	1	phosphoribosylformylglycine amidine synthase	phosphoribosylformylglycine amidine synthase	phosphoribosylformylglycine amidine synthase II
"CJ_10001082"	CJE1037	-	Cj0957c	-	CCO0938	-	1	1	1	1	1	1	1	1	1	1	putative thiophene and furan oxidation protein	tRNA modification GTPase	tRNA modification GTPase TrmE
"CJ_10001083"	CJE1038	-	Cj0958c	-	CCO0937	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001085"	CJE1039	-	Cj0959c	-	CCO0936	-	1	1	1	1	-1	1	1	-1	1	-1	putative membrane protein	putative inner membrane protein translocase	60 kDa inner-membrane protein
"CJ_10001087"	CJE1040	rnpA	Cj0960c	rnpA	CCO0935	rnpA	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR00278,
"opCcV0100001871"	CJE1041	rpmH	Cj0961c	rpmH	CCO0934	rpmH	1	1	1	1	1	1	1	1	1	1	putative ribonuclease P protein component	ribonuclease P protein component	ribonuclease P protein component
"CJ_10001090"	CJE1042	-	Cj0962	-	CCO0933	-	1	1	1	1	-1	1	1	-1	1	1	putative ribonuclease P protein component	ribonuclease P protein component	ribonuclease P protein component
"CJ_10001093"	CJE1043	-	Cj0963	-	CCO0932	-	1	1	1	1	-1	1	1	-1	1	1	putative acetyltransferase	acetyltransferase, GNAT family	acetyltransferase, GNAT family
"CJ_10001096"	CJE1044	-	Cj0964	-	CCO0931	-	0	1	1	-1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	phage SPO1 DNA polymerase-related protein
"CJ_10001098"	CJE1045	-	Cj0965c	-	CCO0930	-	0	1	1	1	-1	1	1	0	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0964
"opCjV0100000416"	CJE1046	-	-	-	-	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	thioesterase family protein	thioesterase family protein, putative
"CJ_10001117"	CJE1047	-	Cj0967	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative acetyltransferase	acetyltransferase, GNAT family	acetyltransferase, GNAT family
"opCjV0100000857"	CJE1048	-	-	-	-	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10001118"	CJE1049	-	Cj0968	-	-	-	0	1	1	1	0	1	1	-1	1	1	putative periplasmic protein	hypothetical protein	-

"opCjV010000 0581"	CJE1050	-	Cj0971	-	-	-	1	0	1	0	1	0	1	1	1	0	pseudogene	hypothetical protein	-
"CJ_10001120"	CJE1051	-	Cj0970	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10001122"	CJE1052	-	Cj0971	-	-	-	1	1	1	1	0	-1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10001124"	CJE1053	-	Cj0972	-	-	-	1	1	-1	1	1	-1	-1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10001126"	CJE1054	-	Cj0973	-	-	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	-
"CJ_10001129"	CJE1055	-	Cj0974	-	-	-	-1	-1	-1	-1	1	-1	1	1	-1	1	hypothetical protein	hypothetical protein	-
"CJ_10001132"	CJE1056	-	Cj0975	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative outer-membrane protein	pseudogene	-
"CJ_10001136"	CJE1058	-	Cj0976	-	CCO1042	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	methyltransferase, putative	conserved hypothetical protein
"opCcV010000 1219"	CJE1059	-	Cj0977	-	CCO1043	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	probable lipoprotein Cj0978c -related protein
"CJ_10001161"	CJE1060	-	Cj0978c	-	CCO1044	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	thermonuclease family protein
"CJ_10001162"	CJE1061	-	Cj0979c	-	CCO1045	-	1	1	1	1	1	1	1	1	1	1	putative secreted nuclease	thermonuclease family protein	probable peptidase Cj0980
"CJ_10001164"	CJE1062	pepD	Cj0980	-	CCO1046	-	1	1	1	1	1	1	1	1	1	1	putative peptidase	aminoacyl-histidine dipeptidase	major facilitator family transporter
"CJ_10001166"	CJE1063	cjaB	Cj0981c	-	CCO0255	proP	1	1	1	1	1	1	0	0	1	1	transmembrane transport protein	transport protein CjaB	proline/betaine transporter (proP)
"opCcV010000 0748"	CJE1064	-	Cj0982c	-	CCO1048	-	1	1	0	-1	0	-1	0	1	-1	-1	putative amino-acid transporter periplasmic	surface antigen, CjaA	ABC transporter, periplasmic substrate-binding
"opCjjV010000 156"	CJE1065	jlpA	Cj0983	-	-	-	1	1	1	1	1	1	0	1	1	1	putative lipoprotein	surface-exposed lipoprotein	-
"CJ_10001170"	CJE1066	-	Cj0984	-	CCO1050	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF328)
"CJ_10001174"	CJE1067	hipO	Cj0985c	hipO	-	-	1	1	1	1	1	1	1	1	1	1	hippurate hydrolase	hippurate hydrolase	-
"CJ_10001178"	CJE1068	-	Cj0986c	-	-	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	-
"CJ_10001209"	CJE1069	-	Cj0989	-	CCO1051	-	0	0	1	0	1	-1	-1	1	-1	1	putative membrane protein	hypothetical protein	conserved hypothetical protein
"CJ_10001211"	CJE1070	-	Cj0990c	-	CCO1052	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001212"	CJE1071	-	Cj0991c	-	CCO1053	glpC	0	-1	-1	1	1	-1	1	-1	0	-1	putative oxidoreductase ferredoxin-type electron	iron-sulfur cluster-binding domain protein	anaerobic glycerol-3-phosphate dehydrogenase,
"CJ_10001213"	CJE1072	hemN	Cj0992c	hemN	CCO1054	hemN	1	1	1	1	1	1	1	1	1	1	oxygen-independent coproporphyrinogen III oxidase	coproporphyrinogen III oxidase	oxygen-independent coproporphyrinogen III
"CJ_10001214"	CJE1073	-	Cj0993c	-	CCO1055	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001217"	CJE1074	argF	Cj0994c	argF	CCO1056	argF	1	1	1	1	1	1	1	1	1	1	ornithine carbamoyltransferase	ornithine carbamoyltransferase	ornithine carbamoyltransferase
"CJ_10001221"	CJE1075	hemB	Cj0995c	hemB	CCO1057	hemB	1	1	1	1	1	1	1	1	1	1	delta-aminolevulinic acid dehydratase	delta-aminolevulinic acid dehydratase	porphobilinogen synthase
"CJ_10001225"	CJE1076	ribA	Cj0996	ribA	CCO1058	ribA	1	1	1	1	1	1	1	1	1	1	GTP cyclohydrolase II	GTP cyclohydrolase II protein	GTP cyclohydrolase II
"CJ_10001255"	CJE1077	gidB	Cj0997	-	CCO1059	gidB	-1	-1	-1	-1	-1	1	1	1	1	-1	gidB homolog	methyltransferase GidB	methyltransferase GidB
"CJ_10001256"	CJE1078	-	Cj0998c	-	CCO1060	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0998c
"CJ_10001258"	CJE1079	-	Cj0999c	-	CCO1061	-	1	1	1	0	1	0	0	1	1	-1	putative integral membrane protein	hypothetical protein	membrane protein, putative
"CJ_10001260"	CJE1080	-	Cj1000	-	CCO1062	-	1	1	1	1	1	1	1	1	1	1	putative transcriptional regulator (lysR)	transcriptional regulator, LysR family	transcription regulator LysR family VCA0542 ,
"opCcV010000	CJE1081	rpoD	Cj1001	rpoD	CCO1063	rpoD	0	1	0	1	1	1	1	1	1	1	RNA polymerase sigma	RNA polymerase sigma	RNA polymerase sigma

1743"																	factor (sigma-70)	factor	factor RpoD	
"opCcV0100001596"	CJE1082	sixA	Cj1002c	-	CCO1064	-	-1	1	1	1	1	1	-1	-1	1	-1	hypothetical protein	phosphohistidine phosphatase SixA	phosphohistidine phosphatase SixA, putative	
"CJ_10001263"	CJE1083	-	Cj1003c	-	CCO1065	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	rhomboid family protein	glp regulon protein (glpG) isolog	
"CJ_10001265"	CJE1084	-	Cj1004	-	CCO1066	-	0	1	0	0	1	1	0	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1004	
"CJ_10001268"	CJE1085	-	Cj1005c	-	CCO1067	ftsH_1	1	1	1	1	1	1	1	1	1	1	putative membrane bound ATPase	cell division protein FtsH, putative	cell division protein (ftsH)	
"CJ_10001271"	CJE1086	-	Cj1006c	-	CCO1068	-	1	1	1	1	1	1	1	1	1	1	hypothetical proteinCj1006c	MiaB-like tRNA modifying enzyme	MiaB-like tRNA modifying enzyme	
"CJ_10001284"	CJE1087	-	Cj1007c	-	CCO1069	-	1	1	1	1	1	1	1	0	1	1	putative membrane protein	mechanosensitive ion channel family protein	conserved hypothetical integral membrane	
"CJ_10001285"	CJE1088	aroB	Cj1008c	aroB	CCO1070	aroB	1	1	1	1	1	1	1	1	1	1	3-dehydroquinase synthase	3-dehydroquinase synthase	3-dehydroquinase synthase	
"CJ_10001286"	CJE1089	-	Cj1009c	-	CCO1071	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	trkA domain protein	TrkA domain protein	
"CJ_10001288"	CJE1090	tgt	Cj1010	tgt	CCO1072	tgt	-1	-1	-1	-1	-1	-1	1	1	1	-1	queuine tRNA-ribosyltransferase	queuine tRNA-ribosyltransferase	queuine tRNA-ribosyltransferase	
"CJ_10001289"	CJE1091	-	Cj1011	-	CCO1073	-	0	1	1	1	0	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1011	
"opCjV0100000026"	CJE1092	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-	
"opCcV0100001013"	CJE1093	-	-	-	CCOA0089	-	-1	-1	0	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	hypothetical protein	
"opCcV0100000010"	CJE1094	-	-	-	CCOA0088	-			0			-1	-1	-1	1		-	site-specific recombinase, phage integrase	site-specific recombinase XerC, putative	
"opCjV0100000710"	CJE1095	-	-	-	-	-		-1	0	-1	-1	1	1	1	-1	-1	-	hypothetical protein	-	
"opCcV0100001390"	CJE1096	-	-	-	CCOA0087	-	0	0	0	1	1	1	1	1	1	1	-	hypothetical protein	erythrocyte membrane-associated antigen	
"opCjV0100001060"	CJE1097	-	-	-	-	-		-1	1	-1	1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCcV0100000501"	CJE1098	-	-	-	CCOA0086	-		-1	1	-1	1	1	-1	1	-1	-1	-	hypothetical protein	hypothetical protein	
"opCcV0100000426"	CJE1099	-	-	-	CCOA0085	-			1					-1		-1	-	hypothetical protein	hypothetical protein	
"opCjV0100001086"	CJE1100	-	-	-	-	-			0				0	-1	1	-1	-	death-on-curing family protein	-	
"opCcV0100000781"	CJE1101	-	-	-	CCOA0056	-		-1	0	-1	1	-1	-1	1	1	-1	-	hypothetical protein	hypothetical protein	
"opCjV0100000105"	CJE1102	-	-	-	-	-			0		-1	-1	-1	1	1	-1	-	hypothetical protein	-	
"opCcV0100000147"	CJE1103	-	-	-	CCOA0047	-		1	1	-1	1	1	1	-1	1	0	1	-	hypothetical protein	Domain of unknown function (DUF332) superfamily
"opCjV0100000362"	CJE1104	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-	
"opCcV0100002089"	CJE1105	-	-	-	CCOA0045	-		1	1	1	1	1	1	1	1	1	-	hypothetical protein	TraG protein, putative	
"opCjV0100000980"	CJE1106	-	-	-	CCOA0045	-		-1	-1	-1	1	-1	-1	1	0	-1	-	hypothetical protein	TraG protein, putative	
"opCjV0100000731"	CJE1107	-	-	-	CCOA0045	-			1			1	1	1			-	TraG-like protein	TraG protein, putative	
"opCjV0100000054"	CJE1108	-	-	-	-	-			1	1	1	0	-1	1	1	1	-	hypothetical protein	-	
"opCjV0100000000"	CJE1109	-	-	-	-	-			-1	-1	1	-1	-1	1	0	1	-	hypothetical protein	-	

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"opCcV0100000863"	CJE1138	-	-	-	CCOA0041	-		-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	hypothetical protein
"opCcV0100001069"	CJE1139	-	-	-	CCOA0141	-	1	1	1	1	-1	-1	1	1	-1	1	-	hypothetical protein	conserved hypothetical protein
"opCjV0100000936"	CJE1140	-	-	-	-	-	0	-1	0	0	1	1	-1	1	1	0	-	pseudogene	-
"opCcV0100001183"	CJE1141	-	-	-	CCOA0129	-	1	1	1	1	-1	1	1	-1	1	1	-	hypothetical protein	vgrG protein VCA0018 , putative
"opCcV0100000261"	CJE1142	-	-	-	CCOA0099	-		-1	1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	vgrG protein VCA0123 , putative
"opCjV0100000008"	CJE1143	-	-	-	-	-		-1	-1	-1	-1	-1	-1	1	1	-1	-	hypothetical protein	-
"opCjV0100000334"	CJE1144	-	-	-	-	-		-1	-1	-1	1	0	-1	1	1	-1	-	hypothetical protein	-
"opCjV0100000499"	CJE1145	-	-	-	-	-			-1	-1	-1	1	1	-1	1	-1	-	hypothetical protein	-
"opCcV0100000277"	CJE1146	-	-	-	CCOA0095	-			-1					-1			-	hypothetical protein	hypothetical protein
"opCcV0100001147"	CJE1147	-	-	-	CCOA0094	-	-1		-1				1	-1	1		-	hypothetical protein	conserved hypothetical protein
"opCjV0100000648"	CJE1148	-	-	-	-	-		-1	-1	-1	-1	1	1	-1	1	-1	-	hypothetical protein	-
"opCjV0100000917"	CJE1149	-	-	-	-	-	-1	-1	0	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"opCcV0100000669"	CJE1150	-	-	-	CCOA0098	-			-1			-1	-1	1			-	hypothetical protein	conserved hypothetical protein
"opCcV0100001445"	CJE1151	-	-	-	CCOA0096	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	hypothetical protein
"opCjV0100000097"	CJE1152	-	-	-	-	-			-1	-1	-1	0	-1	1	1	1	-	hypothetical protein	-
"opCcV0100002028"	CJE1153	-	-	-	CCOA0091	-	1	1	1	-1	-1	1	1	1	1	-1	-	hypothetical protein	hypothetical protein
"opCcV0100000319"	CJE1154	-	-	-	CCOA0090	-	-1	-1	1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	hypothetical protein
"opCjV0100000357"	CJE1155	-	-	-	-	-	-1	-1	-1	-1	1	1	-1	1	-1	-1	-	hypothetical protein	-
"CJ_10001291"	CJE1156	-	Cj1012c	-	CCO1079	-	1	1	1	0	-1	1	-1	0	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1012c
"CJ_10001293"	CJE1157	-	Cj1013c	-	CCO1080	-	1	1	1	1	1	1	1	1	1	1	putative membrane protein	cytochrome c biogenesis protein, CcmF/CycK/CcsA	cytochrome c biogenesis protein (ycf5)
"CJ_10001295"	CJE1158	livF	Cj1014c	livF	CCO1081	livF	1	1	1	1	1	1	1	1	1	1	branched-chain amino-acid ABC transport system	high affinity branched-chain amino acid ABC	branched chain amino acid ABC transporter,
"CJ_10001298"	CJE1159	livG	Cj1015c	livG	CCO1082	livG	1	1	1	1	1	1	1	1	1	1	branched-chain amino-acid ABC transport system	high affinity branched-chain amino acid ABC	branched chain amino acid ABC transporter,
"CJ_10001300"	CJE1160	livM	Cj1016c	livM	CCO1083	livM	1	1	1	1	1	1	1	1	1	1	putative branched-chain amino-acid ABC transport	high affinity branched-chain amino acid ABC	branched chain amino acid ABC transporter,
"CJ_10001310"	CJE1161	livH	Cj1017c	livH	CCO1084	livH	1	1	1	1	1	1	1	1	1	1	branched-chain amino-acid ABC transport system	high affinity branched-chain amino acid ABC	branched chain amino acid ABC transporter,
"CJ_10001311"	CJE1162	-	Cj1018c	livK	CCO1085	livJ	1	1	1	1	1	1	1	1	1	1	branched-chain amino-acid ABC transport system	high affinity branched-chain amino acid ABC	branched chain amino acid ABC transporter,
"CJ_10001313"	CJE1163	-	Cj1019c	livJ	CCO1086	livJ	1	1	1	1	0	1	1	1	1	1	branched-chain amino-acid ABC transport system	high affinity branched-chain amino acid ABC	branched chain amino acid ABC transporter,
"CJ_10001315"	CJE1164	-	Cj1020c	-	CCO1087	-	1	1	1	1	-1	1	1	0	1	1	putative cytochrome C	cytochrome c family protein	probable cytochrome C Cj1020c
"CJ_10001318"	CJE1165	-	Cj1021c	-	CCO1088	-	1	-1	-1	-1	-1	-1	-1	1	1	1	putative periplasmic protein	porin domain protein	probable periplasmic

"CJ_10001321"	CJE1166	-	Cj1022c	-	CCO1089	-	1	1	1	1	1	1	1	1	0	1	putative integral membrane protein	hypothetical protein	protein Cj1021c -related probable integral membrane protein Cj1022c
"CJ_10001323"	CJE1167	asd	Cj1023c	asd	CCO1090	asd	1	1	1	1	1	1	1	1	1	1	aspartate-semialdehyde dehydrogenase	aspartate-semialdehyde dehydrogenase	aspartate-semialdehyde dehydrogenase
"CJ_10001326"	CJE1168	-	Cj1024c	-	CCO1091	-	1	1	1	1	0	1	1	1	1	1	signal-transduction regulatory protein	sigma-54 dependent DNA-binding response	response regulator
"CJ_10001329"	CJE1169	-	Cj1025c	-	CCO1092	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001333"	CJE1170	-	Cj1026c	-	CCO1093	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj1026c
"CJ_10001350"	CJE1171	gyrA	Cj1027c	gyrA	CCO1094	gyrA	1	1	1	1	1	1	1	1	1	1	DNA gyrase subunit A	DNA gyrase subunit A	DNA gyrase, A subunit
"CJ_10001351"	CJE1172	ctsW	Cj1028c	-	CCO1095	ctsW	1	1	1		-1	0	-1		1		possible purine/pyrimidine	transformation system protein	transformation system protein
"CJ_10001353"	CJE1173	mapA	Cj1029c	mapA	CCO1096	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein	outer membrane lipoprotein MapA	outer membrane lipoprotein MapA
"CJ_10001355"	CJE1174	lepA	Cj1030c	lepA	CCO1097	lepA	1	1	1	1	1	1	1	1	1	1	lepA GTP-binding protein homolog	GTP-binding protein LepA	GTP-binding protein LepA
"CJ_10001357"	CJE1175	-	Cj1031	-	CCO1098	-	1	1	1	1	1	1	1	1	1	1	putative outer membrane component of efflux	pseudogene	probable outer membrane component of efflux
"CJ_10001358"	CJE1176	-	Cj1032	-	CCO1099	-	1	1	1	1	1	1	1	1	1	1	putative membrane fusion component of efflux	hypothetical protein	probable membrane fusion component of efflux
"CJ_10001360"	CJE1177	-	Cj1033	-	CCO1100	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane component of efflux	AcrB/AcrD/AcrF family protein	probable integral membrane component of efflux
"CJ_10001361"	CJE1178	-	Cj1034c	-	CCO1101	-	1	1	1	1	1	1	1	1	1	1	possible dnaJ-like protein	DnaJ domain protein	DnaJ domain protein
"CJ_10001363"	CJE1179	-	Cj1035c	-	CCO1102	-	1	1	1	1	1	1	1	1	1	1	possible transferase	arginyl-tRNA-protein transferase	probable transferase Cj1035c
"CJ_10001368"	CJE1180	-	Cj1036c	-	CCO1103	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001375"	CJE1181	-	Cj1037c	pycA	CCO1104	carB	1	1	1	1	1	1	1	1	1	1	putative pyruvate carboxylase A subunit	acetyl-CoA carboxylase	carbamoyl-phosphate synthase, large subunit
"CJ_10001376"	CJE1182	-	Cj1038	-	CCO1105	-	1	1	1	1	1	1	1	1	1	1	probable cell division/peptidoglycan	cell division protein, FtsW/RodA/SpoVE family	cell cycle protein, FtsW/RodA/SpoVE family
"CJ_10001377"	CJE1183	murG	Cj1039	murG	CCO1106	murG	1	1	1	1	1	1	1	1	1	1	putative	N-acetylglucosaminyl transferase	UDP-N-acetylglucosamine-N-acetylmuramyl-transferase
"CJ_10001379"	CJE1184	-	Cj1040c	-	CCO1107	-	1	1	1	1	1	1	1	0	1	1	putative transmembrane transport protein	hypothetical protein	cyanate MFS transporter, putative
"CJ_10001380"	CJE1185	-	Cj1041c	-	CCO1108	-	1	1	1	1	1	-1	1	1	1	1	putative periplasmic ATP/GTP-binding protein	hypothetical protein	conserved hypothetical protein
"CJ_10001382"	CJE1186	-	Cj1042c	-	CCO1109	-	1	1	1	1	1	-1	1	1	1	1	putative transcriptional regulatory protein	transcriptional regulator, AraC family	probable transcription regulatory protein
"CJ_10001384"	CJE1187	-	Cj1043c	-	CCO1110	-	1	1	1	1	1	1	1	1	1	1	possible transferase	thiamine-phosphate pyrophosphorylase, putative	thiamine-phosphate pyrophosphorylase, putative
"CJ_10001386"	CJE1188	thiH	Cj1044c	thiH	CCO1111	-	1	1	1	1	1	1	1	1	1	1	thiH protein	thiamine biosynthesis protein ThiH	thiH protein Cj1044c
"CJ_10001390"	CJE1189	thiG	Cj1045c	thiG	CCO1112	-	1	1	1	1	1	1	1	1	1	1	thiG protein	thiazole synthase	thiamin biosynthesis protein thiG Cj1045c
"CJ_10001393"	CJE1190	thiF	Cj1046c	moeB	CCO1113	-	1	1	1	1	1	1	1	1	1	1	putative molybdopterin biosynthesis protein	thiamine biosynthesis protein ThiF	HesA/MoeB/ThiF family protein
"CJ_10001404"	CJE1191	thiS	Cj1047c	-	CCO1114	thiS	1	0	0	1	1	1	1	1	1	1	hypothetical protein	thiamine biosynthesis protein ThiS	thiamine biosynthesis protein ThiS
"CJ_10001405"	CJE1192	dapE	Cj1048c	dapE	CCO1116	dapE	1	0	1	1	1	1	1	1	1	1	succinyl-diaminopimelate desuccinylase	succinyl-diaminopimelate desuccinylase	succinyl-diaminopimelate desuccinylase
"CJ_10001407"	CJE1193	-	Cj1049c	-	CCO1117	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	transporter, LysE family	probable integral membrane protein Cj1049c

"CJ_10001408"	CJE1194	-	Cj1050c	-	CCO1118	-	1	1	1	1	-1	-1	1	-1	1	1	putative transferase	NAD-dependent deacetylase	probable transferase Cj1050c
"CJ_10001410"	CJE1195	-	Cj1051c	-	CCO1119	-		1	1	1	-1	-1	0	-1	1	1	restriction modification enzyme	type II restriction-modification enzyme	type I restriction modification enzyme
"CJ_10001412"	CJE1196	mutS	Cj1052c	mutS	CCO1120	-	0	1	1	1	1	1	1	1	1	1	putative mismatch repair protein	recombination and DNA strand exchange inhibitor	probable mismatch repair protein Cj1052c
"CJ_10001414"	CJE1197	-	Cj1053c	-	CCO1121	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1053c
"CJ_10001416"	CJE1198	murC	Cj1054c	murC	CCO1122	murC	1	1	1	1	1	1	1	1	1	1	UDP-N-acetylmuramate--alanine ligase	UDP-N-acetylmuramate--L-alanine ligase	UDP-N-acetylmuramate--alanine ligase
"CJ_10001422"	CJE1199	-	Cj1056c	-	CCO1127	-	1	1	1	1	-1	1	1	0	0	1	hypothetical protein	hydrolase, carbon-nitrogen family	hydrolase, carbon-nitrogen family
"CJ_10000918"	CJE1200	-	Cj1057c	-	CCO1128	-	1	1	1	1	-1	1	1	-1	1	1	putative coiled-coil protein	exodeoxyribonuclease VII, small subunit,	exodeoxyribonuclease VII, small subunit,
"CJ_10000921"	CJE1201	guaB	Cj1058c	guaB	CCO1129	guaB	1	1	1	1	1	1	1	-1	1	1	inositol-5'-monophosphate dehydrogenase	inositol-5'-monophosphate dehydrogenase	inosine-5'-monophosphate dehydrogenase
"CJ_10000924"	CJE1202	gatA	Cj1059c	gatA	CCO1130	gatA	1	1	1	1	1	1	1	1	1	1	Glu-tRNA ^{Gln} amidotransferase subunit A	glutamyl-tRNA amidotransferase subunit A	glutamyl-tRNA(Gln) amidotransferase, A subunit
"opCcV0100001814"	CJE1203	-	Cj1060c	-	CCO1131	-	1	0	0	1	1	0	1	1	1	0	-	hypothetical protein	hypothetical protein
"opCcV0100000011"	CJE1204	ileS	Cj1061c	ileS	CCO1132	ileS	1	1	1	1	-1	0	1	1	1	0	isoleucyl-tRNA synthetase	isoleucyl-tRNA synthetase	isoleucyl-tRNA synthetase
"CJ_10000928"	CJE1205	-	Cj1062	-	CCO1133	-	1	1	1	1	1	1	1	0	1	0	hypothetical protein	competence/damage-inducible domain protein	conserved hypothetical integral membrane
"CJ_10000930"	CJE1206	-	Cj1063	-	CCO1134	-	1	-1	1	-1	1	1	1	-1	1	1	possible acetyltransferase	acetyltransferase, GNAT family	probable acetyltransferase Cj1063
"opCcV0100001454"	CJE1208	-	Cj1064	-	CCO1135	-	0	-1	1	1	1	1	0	1	1	1	pseudogene	pseudogene	nitroreductase family protein
"CJ_10000933"	CJE1209	-	Cj1066	rdxA	CCO1137	-	1	-1	1	1	0	1	1	1	1	1	nitroreductase	nitroreductase family protein	nitroreductase Cj1066
"CJ_10000934"	CJE1210	pgsA	Cj1067	pgsA	CCO1138	pgsA	1	1	1	1	-1	1	1	0	1	1	CDP-diacylglycerol--glycerol-3-phosphate	CDP-diacylglycerol--glycerol-3-phosphate	CDP-diacylglycerol--glycerol-3-phosphate
"CJ_10000945"	CJE1211	-	Cj1068	-	CCO1139	-	1	-1	1	1	-1	1	1	-1	1	1	putative integral membrane protein	membrane-associated zinc metalloprotease,	membrane-associated zinc metalloprotease,
"CJ_10000948"	CJE1212	-	Cj1069	-	CCO1140	-	1	1	1	1	1	1	1	-1	1	1	hypothetical protein	hypothetical protein	LapB
"CJ_10000951"	CJE1213	rpsF	Cj1070	rpsF	CCO1144	-	1	1	1	0	-1	1	-1	1	1	1	30S ribosomal protein S6	30S ribosomal protein S6	30S ribosomal protein S6 Cj1070
"CJ_10000953"	CJE1214	ssb	Cj1071	ssb	CCO1145	-	1	1	1	1	-1	1	1	-1	1	1	single-strand DNA binding protein	single-strand DNA-binding protein	single strand DNA binding protein
"CJ_10000955"	CJE1215	rpsR	Cj1072	rpsR	CCO1146	rpsR	1	1	1	1	1	1	1	-1	1	1	30S ribosomal protein S18	30S ribosomal protein S18	ribosomal protein S18
"CJ_10000957"	CJE1216	lon	Cj1073c	lon	CCO1147	lon	1	1	1	1	-1		-1	-1	1	-1	ATP-dependent protease La	ATP-dependent protease La	ATP-dependent protease La
"CJ_10000959"	CJE1217	-	Cj1074c	-	CCO1148	-	0	1	1	-1	1	1	1	1	1	1	putative lipoprotein	probable lipoprotein, putative	probable lipoprotein Cj1074c
"CJ_10000962"	CJE1218	-	Cj1075	-	CCO1149	-	0	1	-1	-1	1	0	1	-1	-1	-1	hypothetical protein	hypothetical protein	Uncharacterized BCR, COG1699 subfamily
"CJ_10000963"	CJE1219	proC	Cj1076	proC	CCO1150	proC	1	1	1	1	1	1	-1	1	-1	-1	putative pyrroline-5-carboxylate reductase	pyrroline-5-carboxylate reductase	pyrroline-5-carboxylate reductase
"CJ_10000964"	CJE1220	ctsT	Cj1077	-	CCO1151	ctsT	1	1	1	1	-1	1	1	-1	1	-1	putative periplasmic protein	transformation system protein	transformation system protein
"CJ_10000974"	CJE1221	-	Cj1078	-	CCO1152	-	1	1	1	1	-1	1	-1	1	1	-1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1078
"CJ_10000977"	CJE1222	-	Cj1079	-	CCO1153	-	1	1	1	0	-1	1	1	-1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1079
"CJ_10000980"	CJE1223	hemD	Cj1080c	-	CCO1154	hemD	1	1	0	1	1	1	0	1	1	1	hypothetical protein	uroporphyrinogen-III synthetase	uroporphyrinogen-III synthase

"CJ_10000982"	CJE1224	thiE	Cj1081c	thiE	CCO1155	thiE	1	1	1	1	1	1	0	0	1	1	thiamin-phosphate pyrophosphorylase	thiamine-phosphate pyrophosphorylase	thiamine-phosphate pyrophosphorylase
"CJ_10000984"	CJE1225	thiD	Cj1082c	thiD	CCO1156	thiD	1	1	1	1	1	1	1	1	1	1	phosphomethylpyrimidine kinase	phosphomethylpyrimidine kinase	phosphomethylpyrimidine kinase
"CJ_10000986"	CJE1226	-	Cj1083c	-	CCO1157	nth	0	1	1	1	0	-1	-1	-1	-1	-1	possible nuclease	endonuclease III, putative	endonuclease III
"CJ_10000989"	CJE1227	-	Cj1084c	-	CCO1158	-	1	1	1		-1		1	-1	1		putative ATP/GTP-binding protein	hypothetical protein	Protein of unknown function (DUF815)
"CJ_10000991"	CJE1228	mfd	Cj1085c	mfd	CCO1159	mfd	1	1	1	1	1	1	1	1	1	1	transcription-repair coupling factor	transcription-repair coupling factor	transcription-repair coupling factor
"CJ_10000992"	CJE1229	-	Cj1086c	-	CCO1160	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	Protein of unknown function, DUF583 superfamily
"CJ_10000993"	CJE1230	-	Cj1087c	-	CCO1161	-	1	1	1	1	1	-1	1	1	1	1	putative periplasmic protein	peptidase, M23/M37 family	probable periplasmic protein Cj1087c
"CJ_10001004"	CJE1231	folC	Cj1088c	folC	CCO1162	folC	0	1	1	1	0	1	1	0	1	1	folylpolyglutamate synthase/dihydrofolate	folC bifunctional protein	folylpolyglutamate synthase (folC)
"CJ_10001007"	CJE1232	-	Cj1089c	-	CCO1163	-	0	1	1	1	-1	1	1	1	1	0	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001010"	CJE1233	-	Cj1090c	-	CCO1164	-	1	1	1	0	1	-1	-1	1	0	0	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj1090c
"CJ_10001012"	CJE1234	leuS	Cj1091c	leuS	CCO1165	leuS	1	1	1	1	-1	0	1	-1	1	-1	leucyl-tRNA synthetase	leucyl-tRNA synthetase	leucyl-tRNA synthetase
"CJ_10001014"	CJE1235	secF	Cj1092c	secF	CCO1166	-	1	1	1	1	1	1	1	1	1	1	protein-export membrane protein	protein export protein SecF	protein-export membrane protein SecF
"CJ_10001017"	CJE1236	secD	Cj1093c	secD	CCO1167	secD	0	0	1	1	1	1	1	1	0	1	protein-export membrane protein	protein export protein SecD	protein-export membrane protein SecD
"CJ_10001018"	CJE1237	yajC	Cj1094c	-	CCO1168	yajC	1	1	1	1	1	1	1	1	1	1	putative membrane protein	preprotein translocase subunit YajC	preprotein translocase, YajC subunit
"CJ_10001020"	CJE1238	cutE	Cj1095	-	CCO1169	-		1	1	1	-1	1	1	-1	1	-1	putative integral membrane protein	apolipoprotein N-acyltransferase	apolipoprotein N-acyltransferase (cute),
"CJ_10001021"	CJE1239	metK	Cj1096c	metK	CCO1174	metK	0	1	1	1	1	1	1	1	1	1	S-adenosylmethionine synthetase	S-adenosylmethionine synthetase	S-adenosylmethionine synthetase
"CJ_10001022"	CJE1240	-	Cj1097	-	CCO1175	-	1	1	1	-1	1	1	1	-1	1	1	putative transmembrane transport protein	sodium/dicarboxylate symporter	probable transmembrane transport protein Cj1097
"CJ_10001040"	CJE1241	pyrB	Cj1098	pyrB	CCO1176	pyrB	1	1	1	1	-1	1	1	-1	1	-1	aspartate carbamoyltransferase	aspartate carbamoyltransferase catalytic	aspartate carbamoyltransferase
"CJ_10001042"	CJE1242	pepF	Cj1099	-	CCO1177	pepF	0	1	1	1	1	1	1	1	1	1	peptidase (M3 family)	oligoendopeptidase F	oligoendopeptidase F
"CJ_10001044"	CJE1243	-	Cj1100	-	CCO1178	-	1	1	1	0	0	-1	1	-1	1	-1	hypothetical protein Cj1100	hypothetical protein	conserved hypothetical protein
"CJ_10001045"	CJE1244	-	Cj1101	-	CCO1179	rep	0	0	1	1	1	1	1	1	1	1	ATP-dependent DNA helicase	ATP-dependent DNA helicase, UvrD/REP family	DNA helicase II (uvrD)
"CJ_10001046"	CJE1245	truB	Cj1102	truB	CCO1180	-	1	1	1	1	1	1	1	0	1		tRNA pseudouridine synthase B	tRNA pseudouridine synthase B	tRNA pseudouridine synthase B, putative
"CJ_10001047"	CJE1246	csrA	Cj1103	csrA	CCO1181	csrA	0	1	1	1	1	1	1	1	1	1	carbon storage regulator homolog	carbon storage regulator	carbon storage regulator
"CJ_10001048"	CJE1247	ispE	Cj1104	-	CCO1182	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical proteinCj1104	4-diphosphocytidyl-2-C-methyl-D-erythritol	GHMP kinases putative ATP-binding protein domain
"CJ_10001049"	CJE1248	smpB	Cj1105	smpB	CCO1183	smpB	1	1	1	1	-1	1	1	1	-1	-1	small protein B homolog	SsrA-binding protein	SsrA-binding protein
"opCcV0100000735"	CJE1249	-	Cj1106	-	CCO1184	-	-1	0	0	0	1	1	0	1	1	1	possible periplasmic thioredoxin	thioredoxin domain protein	thioredoxin, putative
"CJ_10001050"	CJE1250	-	Cj1107	-	CCO1185	-	1	1	1	1	-1	1	1	-1	-1	1	hypothetical protein	hypothetical protein	Uncharacterized ACR, COG2127
"CJ_10001069"	CJE1251	clpA	Cj1108	clpA	CCO1186	clpA	1	1	1	1	-1	-1	1	-1	1	1	ATP-dependent CLP protease ATP-binding	ATP-dependent Clp protease, ATP-binding	ATP-dependent Clp protease, ATP-binding

"CJ_10001071"	CJE1252	aat	Cj1109	aat	CCO1187	aat	1	1	1	1	1	1	1	1	-1	-1	-1	subunit putative leucyl/phenylalanyl-tRNA-- protein	subunit leucyl/phenylalanyl-tRNA-- protein transferase	subunit leucyl/phenylalanyl-tRNA-- protein transferase
"CJ_10001073"	CJE1253	-	Cj1110c	-	-	-	1	1	1	1	0	1	1	1	1	1	1	putative MCP-type signal transduction protein	methyl-accepting chemotaxis protein	-
"CJ_10001074"	CJE1254	-	Cj1111c	-	CCO1188	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	integral membrane protein, MarC family	probable integral membrane protein Cj1111c
"CJ_10001075"	CJE1255	msrB	Cj1112c	-	CCO1189	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	methionine sulfoxide reductase B	PilB-related protein
"CJ_10001076"	CJE1256	-	Cj1113	-	CCO1190	-	1	1	1	1	-1	1	1	-1	-1	1	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF455)
"CJ_10001077"	CJE1257	pssA	Cj1114c	pssA	CCO1191	-	1	1	1	1	-1	1	1	1	1	-1	-1	CDP-diacylglycerol--serine	CDP-diacylglycerol--serine	CDP-diacylglycerol--serine
"CJ_10001078"	CJE1258	-	Cj1115c	-	CCO1192	-	1	1	1	-1	-1	1	1	0	1	1	1	putative membrane protein	phosphatidylserine decarboxylase-related	phosphatidylserine decarboxylase-related
"CJ_10001079"	CJE1259	ftsH	Cj1116c	ftsH	CCO1193	ftsH	0	1	1	1	0	1	1	0	-1	1	1	membrane bound zinc metallopeptidase	cell division protein FtsH	cell division protein FtsH
"CJ_10001080"	CJE1260	-	Cj1117c	prmA	CCO1194	prmA	1	1	1	1	-1	1	1	1	1	-1	-1	possible ribosomal protein methyltransferase	ribosomal protein L11 methyltransferase	ribosomal protein L11 methyltransferase VC0293
"opCcV010000 1838"	CJE1261	cheY	Cj1118c	cheY	CCO1195	-	1	1	1	1	1	1	1	1	1	1	1	chemotaxis regulatory protein	chemotaxis protein CheY	chemotaxis regulatory protein Cj1118c
"opCjjV010000 092"	CJE1262	pglG	Cj1119c	wlaM	-	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	general glycosylation pathway protein	-
"opCcV010000 1169"	CJE1263	pglF	Cj1120c	wlaL	CCO1196	pglF	1	1	1	1	1	1	1	1	1	1	1	putative sugar epimerase/dehydratase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 1599"	CJE1264	pglE	Cj1121c	wlaK	CCO1197	pglE	1	1	1	1	1	1	1	1	1	1	1	putative aminotransferase (degT family)	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 0909"	CJE1265	pglD	Cj1123c	wlaI	CCO1198	pglD	1	1	0	1	1	-1	0	0	1	1	1	putative transferase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 0601"	CJE1266	pglC	Cj1124c	wlaH	CCO1199	pglC	1	-1	0	1	1	-1	1	-1	1	1	1	putative galactosyltransferase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 1943"	CJE1267	pglA	Cj1125c	wlaG	CCO1200	pglA	1	1	1	1	-1	1	1	1	1	1	1	putative galactosyltransferase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 1680"	CJE1268	pglB	Cj1126c	wlaF	CCO1201	pglB	1	1	1	1	1	1	1	1	0	1	1	putative integral membrane protein (possible)	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 0963"	CJE1269	pglJ	Cj1127c	wlaE	CCO1202	pglJ	1	-1	-1	0	1	0	1	1	1	1	1	putative glycosyltransferase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 1010"	CJE1270	pglI	Cj1128c	wlaD	CCO1203	pglI	1	1	1	1	1	1	1	1	1	1	1	putative glycosyltransferase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 0192"	CJE1271	pglH	Cj1129c	wlaC	CCO1204	pglH	1	1	1	1	1	1	1	1	1	1	1	putative glycosyltransferase	general glycosylation pathway protein	general glycosylation pathway protein
"opCcV010000 0153"	CJE1272	wlaB	Cj1130c	wlaB	CCO1205	wlaB	1	1	1	1	1	1	1	1	1	1	1	ABC-type transport protein	ABC transporter, ATP- binding/permease protein	ABC transporter, ATP- binding/permease protein
"opCcV010000 1263"	CJE1273	galE	Cj1131c	galE	CCO1206	galE	1	0	1	1	1	1	1	1	1	1	1	UDP-glucose 4-epimerase	UDP-glucose 4-epimerase	UDP-glucose 4-epimerase
"CJ_10001156"	CJE1274	wlaX	Cj1132c	-	CCO1207	wlaX	1	1	1	1	0	0	1	-1	1	1	1	hypothetical protein	polysaccharide biosynthesis protein	polysaccharide biosynthesis protein
"opCcV010000 1916"	CJE1275	waaC	Cj1133	waaC	CCO1208	-	1	1	1	1	-1	1	1	-1	1	1	1	putative lipopolysaccharide heptosyltransferase	lipopolysaccharide heptosyltransferase I	lipopolysaccharide heptosyltransferase-1 (rfaC)
"CJ_10001157"	CJE1276	waaM	Cj1134	htrB	CCO1209	-	1	1	1	1	1	1	1	1	1	1	1	putative lipid A biosynthesis lauroyl acyltransferase	lipid A biosynthesis lauroyl acyltransferase	heat shock protein B (ibpB), putative
"CJ_10001158"	CJE1277	-	Cj1135	-	CCO1210	-	1	1	-1	1	0	-1	-1	-1	1	1	1	putative two-domain glycosyltransferase	lipooligosaccharide biosynthesis	glycosyl transferase, group 2 family protein
"opCjV010000 0275"	CJE1278	-	-	-	-	-	-1	-1	-1	-1	-1	1	1	-1	-1	-1	-1	-	lipooligosaccharide biosynthesis	-

"opCjV0100000610"	CJE1279	-	-	-	-	-	-	-	-1	-	-1	-1	1	-1	-	-	lipooligosaccharide biosynthesis	-	
"CJ_10001197"	CJE1280	-	Cj1139c	-	CCO1213	-	1	1	1	0	-1	-1		1	1	-1	putative galactosyltransferase	lipooligosaccharide biosynthesis	glycosyl transferase, group 2 family protein.
"opCjV0100000624"	CJE1281	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	-
"CJ_10001206"	CJE1282	waaV	Cj1146c	waaV	CCO1219	-	1	1	1	1	1	1	1	1	1	1	putative glucosyltransferase	lipooligosaccharide biosynthesis	glycosyl transferase, putative
"CJ_10001207"	CJE1283	waaF	Cj1148	waaF	CCO1220	rfaF	0	1	0	1	0	1	1	0	1	1	ADP-heptose--LPS heptosyltransferase	ADP-heptose--LPS heptosyltransferase II	ADP-heptose-lps heptosyltransferase II (rfaF)
"opCjV0100000220"	CJE1284	-	-	-	-	-	-1	-1	-1	-1	1	1	-1	1	1	-1	-	lipooligosaccharide biosynthesis	-
"CJ_10001241"	CJE1285	-	Cj1149c	gmhA	CCO1222	gmhA	1	1	1	1	1	1	1	0	0	1	Phosphoheptose isomerase	phosphoheptose isomerase	phosphoheptose isomerase
"CJ_10001243"	CJE1286	hdE	Cj1150c	waaE	CCO1223	-	1	1	1	1	1	1	1	-1	1	1	putative ADP-heptose synthase	D,D-heptose 1-phosphate	rfaE protein
"CJ_10001246"	CJE1287	waaD	Cj1151c	waaD	CCO1224	-	1	1	1	1	-1	1	1	1	1	1	ADP-L-glycero-D-mannoheptose-6-epimerase	ADP-L-glycero-D-mannoheptose-6-epimerase	ADP-L-glycero-D-mannoheptose-6-epimerase (rfaD)
"CJ_10001248"	CJE1288	-	Cj1152c	-	CCO1225	-		1	1	0	-1	1	1	1	1	1	putative phosphatase	hydrolase, putative	histidinol phosphatase domain protein
"CJ_10001249"	CJE1289	cyf	Cj1153	-	CCO1226	-	1	1	0	1	1	1	1	1	1	1	putative periplasmic cytochrome C	cytochrome c553	probable periplasmic cytochrome C Cj1153
"CJ_10001250"	CJE1290	-	Cj1154c	-	CCO1227	-	1	1	1	1	1	1	1	1	1	1	small hydrophobic protein	cytochrome oxidase maturation protein.	small hydrophobic protein Cj1154c -related
"CJ_10001251"	CJE1291	-	Cj1155c	-	CCO1228	fixI	0	1	0	1	1	1	1	1	1	1	putative cation-transporting ATPase	heavy metal translocating P-type ATPase	cation-transporting ATPase, P-type (copA)
"CJ_10001252"	CJE1292	rho	Cj1156	rho	CCO1229	-	1	1	0	1	1	1	1	0	1	1	transcription termination factor	transcription termination factor Rho	transcription termination factor Cj1156
"CJ_10001253"	CJE1293	dnaX	Cj1157	dnaX	CCO1231	dnaZX	-1	1	1	0	-1	1	1	-1	1	1	putative DNA polymerase III subunit gamma	DNA polymerase III subunits gamma and tau	DNA polymerase III gamma and tau subunits
"CJ_10000207"	CJE1294	-	Cj1159c	-	-	-	1	1	1	1	0	1	1	1	1	1	small hydrophobic protein	pseudogene	-
"CJ_10000213"	CJE1295	-	Cj1161c	-	CCO1232	-	1	1	1	1	1	1	1	1	1	1	putative cation-transporting ATPase	copper-translocating P-type ATPase	copper-translocating P-type ATPase
"CJ_10000215"	CJE1296	-	Cj1162c	-	CCO1233	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	heavy-metal-associated domain, putative
"CJ_10000217"	CJE1297	-	Cj1163c	-	CCO1234	-	0	1	1	1	1	1	1	1	1	1	putative cation transport protein	cation efflux family protein	cation efflux family protein
"CJ_10000219"	CJE1298	-	Cj1164c	-	CCO1235	-	1	1	1	1	1	1	1	0	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000221"	CJE1299	-	Cj1165c	-	CCO1236	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1165c
"CJ_10000224"	CJE1300	-	Cj1166c	-	CCO1237	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1166c
"CJ_10000225"	CJE1301	ldh	Cj1167	ldh	CCO1238	-	0	1	1	1	1	1	1	1	1	1	putative L-lactate dehydrogenase	L-lactate dehydrogenase	L-lactate dehydrogenase
"CJ_10000226"	CJE1302	-	Cj1168c	-	CCO1239	-	1	1	0	1	1	1	1	1	-1	1	putative integral membrane protein (dedA)	DedA family protein	probable integral membrane protein (dedA)
"opCjV010000049"	CJE1303	-	Cj1169c	-	-	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10000236"	CJE1304	-	Cj1170c	-	-	-	1	1	1	1	1	-1	1	1	0	1	outer membrane protein	hypothetical protein	-
"CJ_10000239"	CJE1305	ppiB	Cj1171c	ppi	CCO1240	-	0	0	1	1	1	1	1	0	1	1	peptidyl-prolyl cis-trans isomerase	peptidyl-prolyl cis-trans isomerase B	peptidyl-prolyl cis-trans isomerase.
"CJ_10000241"	CJE1306	-	Cj1172c	-	CCO1241	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR01033
"CJ_10000243"	CJE1307	-	Cj1173	-	CCO1242	-	0	1	1	1	1	1	1	1	1	1	putative efflux protein	multidrug resistance	probable efflux protein

"CJ_10000245"	CJE1308	-	Cj1174	-	CCO1243	-	0	1	1	1	1	1	1	1	1	1	putative efflux protein	protein, SMR family multidrug resistance protein, SMR family	Cj1173 probable efflux protein Cj1174
"CJ_10000248"	CJE1309	argS	Cj1175c	argS	CCO1244	argS	1	1	1	1	1	1	1	1	1	1	arginyl-tRNA synthetase	arginyl-tRNA synthetase	arginyl-tRNA synthetase
"CJ_10000250"	CJE1310	-	Cj1176c	-	CCO1245	-	1	-1	1	1	1	1	1	-1	-1	-1	hypothetical protein	twin-arginine translocation protein, TatA/E	Sec-independent protein translocase protein
"CJ_10000251"	CJE1311	-	Cj1177c	gmk	CCO1246	-	1	1	1	1	1	1	1	1	1	1	guanylate kinase	guanylate kinase	guanylate kinase Cj1177c
"CJ_10000252"	CJE1312	-	Cj1178c	-	CCO1247	-	0	1	1	1	1	1	1	1	1	1	highly acidic protein	hypothetical protein	highly acidic protein Cj1178c
"CJ_10000261"	CJE1313	fliR	Cj1179c	fliR	CCO1248	fliR	1	1	1	1	1	1	1	1	1	1	flagellar biosynthetic protein	flagellar biosynthesis protein	flagellar biosynthetic protein FliR
"CJ_10000263"	CJE1314	-	Cj1180c	-	CCO1249	-	1	1	1	1	1	1	1	1	1	1	putative ABC transporter ATP binding protein	ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein
"CJ_10000266"	CJE1315	tsf	Cj1181c	tsf	CCO1250	tsf	1	1	1	1	1	-1	1	-1	-1	-1	elongation factor TS	elongation factor Ts	translation elongation factor Ts
"CJ_10000268"	CJE1316	rpsB	Cj1182c	rpsB	CCO1251	rpsB	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S2	30S ribosomal protein S2	ribosomal protein S2
"CJ_10000270"	CJE1317	-	Cj1183c	cfa	CCO1252	-	1	1	1	1	1	1	1	1	1	1	putative cyclopropane-fatty-acyl-phospholipid	cyclopropane-fatty-acyl-phospholipid synthase,	cyclopropane fatty acid synthase (cfa)
"CJ_10000273"	CJE1318	petC	Cj1184c	petC	CCO1253	petC	1	1	1	1	1	1	1	1	1	1	putative ubiquinol-cytochrome C reductase	ubiquinol--cytochrome c reductase, cytochrome c1	ubiquinol cytochrome c oxidoreductase,
"CJ_10000275"	CJE1319	petB	Cj1185c	petB	CCO1254	petB	0	0	1	1	1	-1	1	1	1	1	putative ubiquinol-cytochrome C reductase	ubiquinol--cytochrome c reductase, cytochrome b	ubiquinol cytochrome c oxidoreductase,
"CJ_10000277"	CJE1320	petA	Cj1186c	petA	CCO1256	-	1	0	-1	0	1	1	0	0	1	1	putative ubiquinol-cytochrome C reductase	ubiquinol--cytochrome c reductase, iron-sulfur	ubiquinol cytochrome c oxidoreductase, Rieske
"CJ_10000278"	CJE1321	arsB	Cj1187c	arsB	CCO1257	-	1	1	1	1	1	1	1	1	1	1	putative arsenical pump membrane protein	arsenical pump membrane protein	arsenic efflux pump arsB
"CJ_10000279"	CJE1322	gidA	Cj1188c	gidA	CCO1258	gidA	1	1	1	1	1	1	1	1	1	1	glucose inhibited division protein A homolog	glucose-inhibited division protein A	glucose inhibited division protein A
"CJ_10000290"	CJE1323	-	Cj1189c	-	CCO1259	-	1	1	1	1	1	1	1	0	1	1	putative signal-transduction sensor protein	methyl-accepting chemotaxis protein	methyl-accepting chemotaxis protein
"CJ_10000293"	CJE1324	-	Cj1190c	-	CCO1260	-	1	1	1	1	-1	1	1	0	1	1	putative MCP-domain signal transduction protein	methyl-accepting chemotaxis protein	methyl-accepting chemotaxis protein (tlpA)
"CJ_10000296"	CJE1325	-	Cj1191c	-	CCO1261	-	-1	-1	1	1	1	1	1	1	1	1	putative signal-transduction sensor protein	methyl-accepting chemotaxis protein	methyl-accepting chemotaxis protein
"CJ_10000298"	CJE1326	dctA	Cj1192	dctA	CCO1262	dctA	0	1	1	1	0	1	1	1	1	1	putative C4-dicarboxylate transport protein	C4-dicarboxylate transport protein	C4-dicarboxylate transport protein
"CJ_10000301"	CJE1327	-	Cj1193c	-	CCO1263	-	-1	-1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1193c
"CJ_10000303"	CJE1328	-	Cj1194	-	CCO1264	-	1	0	1	1	1	1	1	1	1	1	possible phosphate permease	phosphate transporter family protein	probable phosphate permease Cj1194
"CJ_10000305"	CJE1329	-	Cj1195c	pyrC2	CCO1265	-	1	1	1	-1	1	1	-1	1	-1	-1	putative dihydroorotase	dihydroorotase	dihydroorotase, putative
"CJ_10000307"	CJE1330	gpsA	Cj1196c	gpsA	CCO1266	-	1	1	1	1	1	1	1	1	0	1	glycerol-3-phosphate dehydrogenase [NAD(P)+]	NAD(P)H-dependent glycerol-3-phosphate	glycerol-3-phosphate dehydrogenase (NAD
"CJ_10000308"	CJE1331	gatB	Cj1197c	gatB	CCO1267	gatB	0	0	1	1	1	1	-1	1	1	1	Glu-tRNA ^{Gln} amidotransferase subunit B	aspartyl/glutamyl-tRNA amidotransferase subunit	glutamyl-tRNA(Gln) amidotransferase, B subunit
"CJ_10000309"	CJE1332	luxS	Cj1198	-	CCO1268	luxS	1	1	1	1	0	1	1	0	1	1	hypothetical protein	S-ribosylhomocysteinase	autoinducer-2 production protein LuxS
"CJ_10000318"	CJE1333	-	Cj1199	-	CCO1269	-	1	1	1	1	-1	1	1	-1	-1	-1	putative iron/ascorbate-dependent	oxidoreductase, 2OG-Fe(II) oxygenase family	oxidoreductase, 2OG-Fe(II) oxygenase family
"CJ_10000320"	CJE1334	-	Cj1200	-	CCO1271	-	0	1	-1	1	1	-1	1	1	1	1	putative periplasmic protein	lipoprotein, NLPa family	D-methionine-binding lipoprotein MetQ
"CJ_10000322"	CJE1335	metE	Cj1201	metE	CCO1272	metE	1	1	1	1	1	1	1	0	1	1	5-methyltetrahydropteroyltryptophan	5-methyltetrahydropteroyltryptophan	5-methyltetrahydropteroyltryptophan
"CJ_10000323"	CJE1336	metF	Cj1202	metF	CCO1273	-	1	1	1	1	1	1	-1	1	-1	-1	5,10-	5,10-	5,10-

																	methylenetetrahydrofolate reductase	methylenetetrahydrofolate reductase	methylenetetrahydrofolate reductase
"CJ_10000324"	CJE1337	-	Cj1203c	-	CCO1274	-	-1	1	1	-1	-1	1	1	0	1	-1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1203c
"CJ_10000325"	CJE1338	atpB	Cj1204c	atpB	CCO1275	atpB	1	1	1	1	1	1	1	1	1	1	ATP synthase F0 sector A subunit	ATP synthase subunit A	ATP synthase F0, A subunit
"CJ_10000326"	CJE1339	radA	Cj1205c	radA	CCO1276	radA	0	1	1	1	1	1	1	1	1	1	putative DNA repair protein RadA (ATP-dependant)	DNA repair protein RadA	DNA repair protein RadA
"CJ_10000327"	CJE1340	ftsY	Cj1206c	ftsY	CCO1282	ftsY	0	0	0	1	1	1	1	1	1	1	putative signal recognition particle protein	signal recognition particle-docking protein	signal recognition particle-docking protein
"CJ_10000328"	CJE1341	-	Cj1207c	-	CCO1283	-	1	1	1	1	1	1	1	1	1	1	putative lipoprotein thiredoxin	lipoprotein, putative	thiol:disulfide interchange protein DsbE,
"CJ_10000329"	CJE1342	-	Cj1208	-	CCO1284	-	1	1	1	1	1	1	1	1	0	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000337"	CJE1343	-	Cj1209	-	CCO1285	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	HDIg/HD/KH domain protein
"CJ_10000338"	CJE1344	-	Cj1210	-	CCO1286	dedA	-1	0	1	1	1	1	1	0	1	1	putative integral membrane protein	DedA family protein	conserved hypothetical integral membrane
"CJ_10000340"	CJE1345	-	Cj1211	-	CCO1287	-	-1	1	1	1	0	1	1	1	1	1	putative integral membrane protein	ComEC/Rec2 family protein	competence locus E (comE3), putative
"CJ_10000341"	CJE1346	rbn	Cj1212c	rbn	CCO1288	-	1	1	1	1	1	1	1	1	1	1	putative ribonuclease BN	ribonuclease BN	ribonuclease BN, putative
"CJ_10000342"	CJE1347	glcD	Cj1213c	glcD	CCO1289	-	0	0	1	1	1	1	1	1	1	1	putative glycolate oxidase subunit D	glycolate oxidase, subunit GlcD	probable glycolate oxidase chain D Cj1213c
"CJ_10000343"	CJE1348	-	Cj1214c	-	CCO1290	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCjV010000 0308"	CJE1349	-	-	-	-	-	0	1	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_10000344"	CJE1350	-	Cj1215	-	CCO1291	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	peptidase, M23/M37 family	probable periplasmic protein Cj1215
"CJ_10000345"	CJE1351	-	Cj1216c	-	CCO1292	-	1	0	1	1	1	1	1	1	-1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000346"	CJE1352	-	Cj1217c	-	CCO1293	-	1	1	1	1	-1	1	-1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000347"	CJE1353	ribE	Cj1218c	ribA	CCO1294	ribE	1	1	1	1	1	1	1	1	1	1	putative riboflavin synthase alpha chain	riboflavin synthase subunit alpha	riboflavin synthase, alpha subunit
"CJ_10000360"	CJE1354	-	Cj1219c	-	CCO1295	-	-1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1219c
"CJ_10000364"	CJE1355	groES	Cj1220	groES	CCO1296	groES	1	1	1	1	0	1	1	1	1	1	10 kD chaperonin (cpn10)	co-chaperonin GroES	chaperonin, 10 kDa
"opCecV010000 0358"	CJE1356	groEL	Cj1221	groEL	CCO1297	groEL	0	0	1	1	1	1	1	1	1	1	60 kD chaperonin (cpn60)	chaperonin GroEL	chaperonin, 60 kDa
"CJ_10000368"	CJE1357	-	Cj1222c	-	CCO1300	-	1	1	1	1	1	1	1	1	1	1	putative two-component sensor	sensor histidine kinase	signal-transducing protein, histidine kinase,
"CJ_10000370"	CJE1358	-	Cj1223c	-	CCO1301	-	1	1	1	1	1	1	1	1	1	1	putative two-component regulator	DNA-binding response regulator	response regulator
"CJ_10000371"	CJE1359	-	Cj1224	-	CCO1302	-	0	1	1	1	1	1	1	1	1	1	putative iron-binding protein	non-heme iron protein, hemerythrin family	methyl-accepting chemotaxis protein, putative
"CJ_10000372"	CJE1360	-	Cj1225	-	-	-	1	0	1	1	1	1	1	1	0	1	hypothetical protein	hypothetical protein	-
"CJ_10000373"	CJE1361	-	Cj1226c	-	CCO1314	-	1	1	1	1	1	0	1	1	1	1	putative two-component sensor	sensor histidine kinase	signal-transducing protein, histidine kinase
"CJ_10000374"	CJE1362	-	Cj1227c	-	CCO1315	-	1	1	1	1	0	1	1	1	1	1	putative two-component regulator	DNA-binding response regulator	outer membrane protein R (ompR)
"CJ_10000375"	CJE1363	htrA	Cj1228c	htrA	CCO1316	-	1	0	1	1	1	1	1	1	0	1	serine protease (protease DO)	protease DO	serine protease (htrA)
"CJ_10000389"	CJE1364	-	Cj1229	cbpA	CCO1317	-	1	0	1	1	1	1	1	1	1	1	putative curved-DNA binding protein	co-chaperone protein DnaJ	co-chaperone-curved DNA binding protein A

"CJ_10000391"	CJE1365	-	Cj1230	hspR	CCO1318	-	1	1	1	1	1	1	1	1	1	1	putative heat shock transcriptional regulator	transcriptional regulator, MerR family	probable heat shock transcription regulator
"CJ_10000393"	CJE1366	-	Cj1231	kefB	CCO1319	kefB	1	1	1	1	1	1	1	1	1	1	putative glutathione-regulated potassium-efflux	sodium/hydrogen exchanger family protein	glutathione-regulated potassium-efflux system
"CJ_10000395"	CJE1367	-	Cj1232	-	CCO1320	-	0	1	1	1	0	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000397"	CJE1368	-	Cj1233	-	CCO1321	-	1	1	1	1	1	1	1	1	1	1	putative hydrolase	HAD-superfamily hydrolase, subfamily IA, variant	probable hydrolase Cj1233
"CJ_10000398"	CJE1369	glyS	Cj1234	glyS	CCO1322	glyS	0	1	1	1	1	1	1	1	1	1	glycyl-tRNA synthetase beta chain	glycyl-tRNA synthetase beta subunit	glycyl-tRNA synthetase, beta subunit
"CJ_10000399"	CJE1370	-	Cj1235	-	CCO1323	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	peptidase, M23/M37 family	peptidase, M23/M37 family
"CJ_10000400"	CJE1371	-	Cj1236	-	CCO1324	-	0	0	-1	1	1	1	-1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000401"	CJE1372	-	Cj1237c	-	CCO1327	-	1	1	1	1	1	1	1	1	1	1	possible phosphatase	phosphatase, Ppx/GppA family	phosphatase, Ppx/GppA family, putative
"CJ_10000402"	CJE1373	pdxJ	Cj1238	pdxJ	CCO1328	pdxJ	1	1	1	1	1	1	1	1	1	1	putative pyridoxal phosphate biosynthetic	pyridoxal phosphate biosynthetic protein	pyridoxal phosphate biosynthetic protein PdxJ
"CJ_10000417"	CJE1374	pdxA	Cj1239	pdxA	CCO1329	pdxA	0	0	1	1	1	1	1	1	1	1	putative pyridoxal phosphate biosynthetic	4-hydroxythreonine-4-phosphate dehydrogenase	4-hydroxythreonine-4-phosphate dehydrogenase
"opCcV0100000600"	CJE1375	-	Cj1240c	-	CCO1330	-	1	1	0	1	-1	1	-1	1	1	-1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1240c
"CJ_10000419"	CJE1376	-	Cj1240c	-	-	-	0	0	-1	1	0	0	0	-1	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10000422"	CJE1377	-	Cj1241	-	-	-	1	1	1	1	0	1	1	1	1	1	putative transmembrane transport protein	transporter, putative	-
"CJ_10000424"	CJE1378	-	Cj1242	-	CCO1332	-	1	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000426"	CJE1379	hemE	Cj1243	hemE	CCO1333	hemE	1	1	1	1	1	1	1	1	1	1	uroporphyrinogen decarboxylase	uroporphyrinogen decarboxylase	uroporphyrinogen decarboxylase
"CJ_10000427"	CJE1380	-	Cj1244	-	CCO1334	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	radical SAM domain protein	MoaA/NifB/PqqE family protein, putative
"CJ_10000428"	CJE1381	-	Cj1245c	-	CCO1335	-	1	-1	1	1	1	1	1	1	-1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1245c
"CJ_10000429"	CJE1382	uvrC	Cj1246c	uvrC	CCO1336	uvrC	1	0	1	1	1	1	1	1	1	1	excinuclease ABC subunit C	excinuclease ABC subunit C	excinuclease ABC, C subunit
"CJ_10000430"	CJE1383	-	Cj1247c	-	CCO1337	-	-1	1	-1	-1	-1	-1	1	1	1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCjV0100000727"	CJE1384	-	-	-	-	-	1	1	1	1	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_10000431"	CJE1385	guaA	Cj1248	guaA	CCO1338	guaA	0	0	1	1	1	1	1	0	1	1	GMP synthase (glutamine-hydrolyzing)	bifunctional GMP synthase/glutamine	GMP synthase
"CJ_10000445"	CJE1386	-	Cj1249	-	CCO1343	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000448"	CJE1387	purD	Cj1250	purD	CCO1344	purD	1	1	1	1	1	1	1	1	1	1	phosphoribosylamine--glycine ligase	phosphoribosylamine--glycine ligase	phosphoribosylamine--glycine ligase
"CJ_10000451"	CJE1388	-	Cj1251	-	CCO1345	-	1	1	1	1	1	1	1	0	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000453"	CJE1389	-	Cj1252	-	CCO1346	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	organic solvent tolerance protein, putative	conserved hypothetical secreted protein,
"CJ_10000455"	CJE1390	pnp	Cj1253	pnp	CCO1347	-	1	0	1	1	1	1	1	1	1	1	polyribonucleotide nucleotidyltransferase	polyribonucleotide nucleotidyltransferase	polyribonucleotide nucleotidyltransferase
"CJ_10000456"	CJE1391	-	Cj1254	-	CCO1348	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000457"	CJE1392	-	Cj1256c	-	CCO1351	-	1	1	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1256c

"CJ_10000458"	CJE1393	-	Cj1257c	-	CCO1352	-	1	1	1	1	1	1	1	1	1	1	putative efflux pump	pseudogene	multi-drug resistance protein
"CJ_10000459"	CJE1394	-	Cj1258	-	CCO1353	-	1	1	1	1	1	1	1	1	1	1	possible phosphotyrosine protein phosphatase	low molecular weight phosphotyrosine protein	low molecular weight protein
"opCcV0100001939"	CJE1395	porA	Cj1259	porA	CCO0683	-	1	-1	1	-1	1	-1	-1	1	1	-1	major outer membrane protein	major outer membrane protein	major outer membrane protein
"opCcV0100001977"	CJE1396	-	Cj1260c	dnaJ	CCO0682	dnaJ	-1	-1	1	0	1	1	1	1	-1	1	chaperone DnaJ	co-chaperone protein DnaJ	heat shock protein
"opCcV0100000324"	CJE1397	racR	Cj1261	racR	CCO0681	drpA	1	1	1	1	1	1	1	1	1	1	two-component regulator	DNA-binding response regulator	response regulator (ompR)
"CJ_10001102"	CJE1398	racS	Cj1262	racS	CCO0680	-	1	1	1	1	1	1	-1	1	1	1	two-component sensor (histidine kinase)	sensor histidine kinase	signal-transducing protein, histidine kinase
"CJ_10001105"	CJE1399	recR	Cj1263	recR	CCO0679	recR	1	1	1	1	-1	-1	1	0	1	-1	recombination protein	recombination protein RecR	recombination protein RecR
"CJ_10001108"	CJE1400	hydD	Cj1264c	hydD	CCO0678	-	1	1	1	1	-1	1	1	-1	-1	1	putative hydrogenase maturation protease	hydrogenase maturation protease HydD	hydrogenase expression/formation protein
"CJ_10001111"	CJE1401	hydC	Cj1265c	hydC	CCO0677	hyaC	1	1	1	-1	-1	1	1	1	1	-1	Ni/Fe-hydrogenase B-type cytochrome subunit	quinone-reactive Ni/Fe hydrogenase, cytochrome b	quinone-reactive Ni/Fe hydrogenase, cytochrome b
"CJ_10001113"	CJE1402	hydB	Cj1266c	hydB	CCO0676	-	1	-1	1	1	1	1	-1	1	1	-1	Ni/Fe-hydrogenase large subunit	quinone-reactive Ni/Fe-hydrogenase, large	hydrogenase (NiFe) large chain Cj1266c
"CJ_10001114"	CJE1403	hydA	Cj1267c	hydA	CCO0675	-	1	1	1	1	-1	1	1	0	1	1	Ni/Fe-hydrogenase small chain	quinone-reactive Ni/Fe-hydrogenase, small	quinone-reactive Ni/Fe hydrogenase, small
"CJ_10001115"	CJE1404	-	Cj1268c	-	CCO0674	-	1	0	1	1	-1	1	1	-1	1	1	hypothetical protein	oxidoreductase, FAD-dependent	oxidoreductase, FAD-binding, putative
"CJ_10001131"	CJE1405	-	Cj1269c	amiA	CCO0673	amiA	1	1	1	-1	-1	1	1	1	1	1	putative N-acetylmuramoyl-L-alanine amidase	N-acetylmuramoyl-L-alanine amidase	N-acetylmuramoyl-L-alanine amidase (amiA)
"CJ_10001135"	CJE1406	-	Cj1270c	-	CCO0672	-	1	1	1	1	-1	1	1	1	1	-1	hypothetical protein	oxidoreductase, 2-nitropropane dioxygenase	oxidoreductase, 2-nitropropane dioxygenase
"CJ_10001139"	CJE1407	tyrS	Cj1271c	tyrS	CCO0671	tyrS	1	1	1	1	-1	1	1	1	1	-1	tyrosyl-tRNA synthetase	tyrosyl-tRNA synthetase	tyrosyl-tRNA synthetase
"CJ_10001142"	CJE1408	-	Cj1272c	spoT	CCO0670	spoT	1	1	1	1	1	1	1	1	1	1	putative guanosine-3',5'-bis(diphosphate)	RelA/SpoT family protein	penta-phosphate
"CJ_10001145"	CJE1409	rpoZ	Cj1273c	rpoZ	CCO0669	rpoZ	1	1	1	1	1	1	1	1	1	1	putative DNA-directed RNA polymerase omega	DNA-directed RNA polymerase omega subunit	DNA-directed RNA polymerase, omega subunit
"CJ_10001149"	CJE1410	pyrH	Cj1274c	pyrH	CCO0668	pyrH	1	1	1	1	1	1	1	1	1	1	uridylate kinase	uridylate kinase	uridylate kinase
"CJ_10001151"	CJE1411	-	Cj1275c	-	CCO0667	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	peptidase, M23/M37 family	probable periplasmic protein Cj1275c
"CJ_10001153"	CJE1412	-	Cj1276c	-	CCO0665	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	cell division protein FtsX, putative	cell division membrane protein (ftsX)
"CJ_10001154"	CJE1413	lolD	Cj1277c	-	CCO0664	-	1	1	1	1	1	1	1	1	1	1	putative ABC transporter ATP-binding protein	lipoprotein releasing system ATP-binding protein	cell division protein (ftsE)
"CJ_10001155"	CJE1414	trmB	Cj1278c	-	CCO0663	-	1	1	1	1	-1	1	1	0	1	1	hypothetical protein	tRNA (guanine-N(7))-methyltransferase	methyltransferase, putative, putative
"CJ_10001173"	CJE1415	-	Cj1279c	-	CCO0662	-	1	1	1	1	1	1	1	1	1	1	putative fibronectin domain-containing	fibronectin type III domain protein	Fibronectin type III domain protein
"CJ_10001177"	CJE1416	-	Cj1280c	-	CCO0661	rhuD	1	1	1	1	1	1	1	1	1	1	putative ribosomal pseudouridine synthase	ribosomal large subunit pseudouridine synthase,	ribosomal large subunit pseudouridine synthase,
"CJ_10001181"	CJE1417	-	Cj1282	mrdB	CCO0660	-	1	1	1	1	1	1	1	1	1	1	RodA protein homolog	rod shape-determining protein RodA, putative	RodA protein homolog Cj1282
"opCjV0100000721"	CJE1418	-	-	-	-	-	1	-1	1	1	-1	0	1	-1	1	-	-	site-specific recombinase, phage integrase	-
"opCjV0100000575"	CJE1419	-	-	-	-	-	1	-1	1	1	-1	-1	1	1	1	-	-	DNA binding protein, putative	-
"opCjV0100000641"	CJE1420	-	-	-	-	-	1	-1	1	1	-1	-1	1	-1	1	-	-	hypothetical protein	-
"opCjV010000"	CJE1421	-	-	-	-	-	-1	-1	-1	0	1	-1	-1	1	-1	0	-	site-specific DNA-	-

0968"																			methyltransferase	
"opCjV0100000601"	CJE1422	-	-	-	-	-	-1	1	1	1	1	1	1	1	-1	1	-		emm-like protein	-
"opCjV0100000882"	CJE1423	-	-	-	-	-		1	-1	1	1	-1	-1	-1	-1	1	-		hypothetical protein	-
"opCjV0100000896"	CJE1424	-	-	-	-	-	-1	1	-1	1	1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000764"	CJE1425	-	-	-	-	-		1	-1	1				-1		1	-		hypothetical protein	-
"opCjV0100000038"	CJE1426	-	-	-	-	-		1	-1	1	1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000024"	CJE1427	-	-	-	-	-		1	-1	1	1	1	-1	1	1	1	-		hypothetical protein	-
"opCjV0100000303"	CJE1428	-	-	-	-	-		1	-1	1	-1	1	-1	-1	1	1	-		hypothetical protein	-
"opCjV0100000019"	CJE1429	-	-	-	-	-		1	-1	1	1	-1	-1	1	1	1	-		hypothetical protein	-
"opCjV0100000825"	CJE1430	-	-	-	-	-		1	-1	1	1	-1	-1	1	-1	1	-		RloG protein, putative	-
"opCjV0100000964"	CJE1431	-	-	-	-	-		1	-1	1				1		1	-		hypothetical protein	-
"opCjV0100000453"	CJE1432	-	-	-	-	-		1	-1	1	1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000230"	CJE1433	-	-	-	-	-		1		1				-1		1	-		hypothetical protein	-
"opCjV0100000258"	CJE1434	-	-	-	-	-		1	-1	1	1	-1	-1	-1	-1	1	-		hypothetical protein	-
"opCjV0100000086"	CJE1435	-	-	-	-	-		1	-1	1	-1	-1	-1	-1	1	1	-		DNA-binding protein Roi	-
"opCjV0100000850"	CJE1436	-	-	-	-	-		1	-1	1	-1	-1	1	1	1	1	-		hypothetical protein	-
"opCjV0100000261"	CJE1437	-	-	-	-	-		0	-1	1	-1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000511"	CJE1438	-	-	-	-	-		1	-1	1	1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000359"	CJE1439	-	-	-	-	-		-1	-1	0	1	-1	-1	1	0	-1	-		hypothetical protein	-
"opCjV0100000561"	CJE1440	-	-	-	-	-	-1	-1	-1	1	1	-1	-1	1	-1	-1	-		signal peptidase I, putative	-
"opCjV0100000910"	CJE1441	-	-	-	-	-	-1	-1	0	0	1	0	-1	1	0	-1	-		DNA/RNA non-specific endonuclease	-
"opCjV0100001085"	CJE1442	-	-	-	-	-	-1	-1	-1	1	1	-1	-1	-1	-1	-1	-		hypothetical protein	-
"opCjV0100000221"	CJE1444	-	-	-	-	-		-1	-1	0	1	-1	-1	1	1	0	-		hypothetical protein	-
"opCjV0100000498"	CJE1445	-	-	-	-	-		-1	-1	0	1	-1	1	-1	1	-1	-		hypothetical protein	-
"opCjV0100000808"	CJE1447	-	-	-	-	-			-1	1	1	-1	-1	1	0	1	-		hypothetical protein	-
"opCjV0100000711"	CJE1448	-	-	-	-	-	-1	-1	-1	1	1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000548"	CJE1452	-	-	-	-	-			-1	1	1	-1	-1	1	-1	1	-		hypothetical protein	-
"opCjV0100000031"	CJE1453	-	-	-	-	-				1				-1		1	-		hypothetical protein	-

"opCjV0100000719"	CJE1454	-	-	-	-	-	-1	-1	-1	1	1	-1	-1	1	-1	1	-	phage head-tail adaptor, putative	-
"opCjV0100000405"	CJE1455	-	-	-	-	-	-1	-1	1	1	-1	-1	0	-1	1	-	hypothetical protein	-	
"opCjV0100000757"	CJE1456	-	-	-	-	-	-1	-1	-1	1	1	-1	-1	1	-1	1	-	hypothetical protein	-
"opCjV0100000469"	CJE1457	-	-	-	-	-	-1	-1	1	1	-1	-1	1	0	1	-	hypothetical protein	-	
"opCjV0100000506"	CJE1458	-	-	-	-	-	-1	-1	1	1	-1	-1	1	-1	1	-	major capsid protein, HK97 family	-	
"opCjV0100000621"	CJE1459	-	-	-	-	-	-1	-1	1	1	-1	-1	1	-1	1	-	hypothetical protein	-	
"opCjV0100000657"	CJE1460	-	-	-	-	-	-1	-1	1	1	-1	-1	1	-1	1	-	hypothetical protein	-	
"opCjV0100000148"	CJE1461	-	-	-	-	-	-1	-1	1	0	-1	-1	1	1	-	hypothetical protein	-		
"opCjV0100000902"	CJE1462	-	-	-	-	-	-1	1	-1	1	1	-1	-1	1	0	1	-	hypothetical protein	-
"opCjV0100000773"	CJE1463	-	-	-	-	-	1	-1	1	-1	-1	-1	1	1	1	-	hypothetical protein	-	
"opCjV0100000301"	CJE1464	-	-	-	-	-	1	-1	1	-1	-1	1	1	1	1	-	hypothetical protein	-	
"opCjV0100000369"	CJE1465	-	-	-	-	-	1	1	1	1	1	1	-1	1	1	-	hypothetical protein	-	
"opCjV0100000730"	CJE1466	-	-	-	-	-	1	-1	1	1	-1	-1	1	1	1	-	hypothetical protein	-	
"opCjV0100000128"	CJE1467	-	-	-	-	-	1	-1	1	1	1	-1	1	1	-	hypothetical protein	-		
"opCjV0100000675"	CJE1468	-	-	-	-	-	1	-1	1	1	-1	-1	1	-1	1	-	phage protein, HK97 gp10 family	-	
"opCjV0100000861"	CJE1469	-	-	-	-	-	-1	1	-1	1	1	-1	-1	0	-1	1	-	portal protein, HK97 family	-
"opCjV0100001119"	CJE1470	-	-	-	-	-	1	-1	1	1	-1	-1	1	1	1	-	toxin-antitoxin protein, putative	-	
"opCjV0100000368"	CJE1471	-	-	-	-	-	1	1	1	1	1	1	-1	1	1	-	phage terminase, large subunit, putative	-	
"opCcV0100000560.80"	CJE1472	-	-	-	-	-	1	1	-1	0	1	-1	-1	1	1	-1	-	"phage terminase, small subunit, putative"	-
"opCjV0100000064"	CJE1473	-	-	-	-	-	-1	0	-1	1	1	-1	-1	1	-1	1	-	HNH endonuclease domain protein	-
"opCjV0100000059"	CJE1474	-	-	-	-	-	-1	1	-1	1	1	-1	-1	1	-1	1	-	hypothetical protein	-
"CJ_10001184"	CJE1475	-	Cj1283	ktrB	CCO1388	-	1	1	1	0	1	1	1	1	1	-1	putative K+ uptake protein	potassium uptake protein, TrkH family	probable K+ uptake protein Cj1283
"CJ_10001189"	CJE1476	-	Cj1284	ktrA	CCO1389	-	1	1	1	1	1	1	1	1	1	1	putative K+ uptake protein	potassium uptake protein TrkA, putative	probable K+ uptake protein Cj1284
"CJ_10001191"	CJE1477	-	Cj1285c	-	CCO1390	-	1	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	Uncharacterized ACR, COG1427
"CJ_10001193"	CJE1478	upp	Cj1286c	upp	CCO1391	upp	0	1	1	1	1	1	1	1	1	1	uracil phosphoribosyltransferase	uracil phosphoribosyltransferase	uracil phosphoribosyltransferase
"CJ_10001196"	CJE1479	-	Cj1287c	-	CCO1392	maeB	1	-1	1	1	1	1	-1	1	-1	-1	malate oxidoreductase	NADP-dependent malic enzyme, truncation	malic enzyme
"CJ_10001199"	CJE1480	gltX	Cj1288c	gltX2	CCO1393	gltX	1	1	1	1	1	1	1	1	1	1	glutamyl-tRNA synthetase	glutamyl-tRNA synthetase	glutamyl-tRNA synthetase
"CJ_10001201"	CJE1481	-	Cj1289	-	CCO1394	-	0	1	1	1	0	1	1	1	1	1	possible periplasmic protein	hypothetical protein	probable periplasmic protein Cj1289

"CJ_10001220"	CJE1482	-	Cj1290c	accC	CCO1395	accC	0	1	1	1	1	1	1	1	1	1	1	biotin carboxylase	acetyl-CoA carboxylase	acetyl-CoA carboxylase, biotin carboxylase
"CJ_10001224"	CJE1483	accB	Cj1291c	accB	CCO1396	accB	1	1	1	1	1	1	1	1	1	1	1	putative biotin carboxyl carrier protein of	acetyl-CoA carboxylase, biotin carboxyl carrier	acetyl-CoA carboxylase, biotin carboxyl carrier
"CJ_10001228"	CJE1484	-	Cj1292	dcd	CCO1397	-	1	1	1	1	1	1	1	1	1	1	1	possible deoxycytidine triphosphate deaminase	deoxycytidine triphosphate deaminase	probable dCTP deaminase Cj1292
"CJ_10001233"	CJE1485	flmA	Cj1293	-	CCO1398	-	1	1	1	1	0	1	1	0	1	1	1	possible sugar nucleotide epimerase/dehydratase	polysaccharide biosynthesis protein	polysaccharide biosynthesis protein
"CJ_10001236"	CJE1486	-	Cj1294	-	CCO1399	-	1	1	0	1	0	1	1	1	1	1	1	putative aminotransferase (degT family)	aminotransferase, DegT/DnrJ/EryC1/StrS family	probable aminotransferase (degT family) Cj1294
"CJ_10001238"	CJE1487	-	Cj1295	-	CCO1400	-	1	1	1	1	-1	1	1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001245"	CJE1488	-	Cj1298	-	CCO1402	-	1	1	-1	1	1	1	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001247"	CJE1489	-	Cj1299	acpP2	CCO1403	-	1	1	-1	1	-1	-1	1	0	1	1	1	putative acyl carrier protein	acyl carrier protein, putative	probable acyl carrier protein Cj1299 -related
"CJ_10001267"	CJE1490	-	Cj1300	-	CCO1404	-	1	1	0	1	-1	1	1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001274"	CJE1491	-	Cj1302	-	CCO1405	-	1	1	-1	1	1	-1	1	1	1	1	1	hypothetical protein	HAD-superfamily phosphatase, subfamily IIIC	HAD-superfamily phosphatase, subfamily IIIC
"CJ_10001276"	CJE1492	-	Cj1303	fabH2	CCO1406	-	1	1	1	1	1	1	1	1	1	1	1	putative 3-oxoacyl-[acyl-carrier-protein]	3-oxoacyl-(acyl carrier protein) synthase	3-oxoacyl-(acyl-carrier-protein) synthase III,
"CJ_10001278"	CJE1493	-	Cj1304	acpP3	CCO1407	-	1	1	1	-1	0	1	-1	1	1	-1	-1	putative acyl carrier protein	acyl carrier protein, putative	probable acyl carrier protein Cj1304 -related
"opCjV010000 0925"	CJE1494	-	Cj1306c	-	CCO1408	-	1	1	0	1	1	-1	-1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001280"	CJE1495	-	Cj1306c	-	CCO1410	-	1	1	-1	1	1	-1	-1	-1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001281"	CJE1496	-	Cj1307	-	CCO1411	-	1	1	0	1	1	1	-1	1	1	1	1	putative amino acid activating enzyme	amino acid adenylation domain protein	amino acid adenylation domain protein
"opCjV010000 1065"	CJE1497	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	0	-1	-	-	acetyltransferase, GNAT family	-
"opCjV010000 0777"	CJE1498	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	formyl transferase domain protein	-
"CJ_10001282"	CJE1499	-	Cj1308	acpP4	CCO1414	-	1	1	0	1	-1	1	-1	0	1	1	1	putative acyl carrier protein	acyl carrier protein, putative	probable acyl carrier protein Cj1308 -related
"opCjV010000 0515"	CJE1500	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	polysaccharide deacetylase family protein	-
"opCjV010000 0278"	CJE1501	-	-	-	-	-	-1	-1	-	-1	-1	-	-	1	1	-1	-	-	hypothetical protein	-
"opCjV010000 0441"	CJE1502	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	1	-1	-	-	aminoglycoside N3-acetyltransferase domain	-
"opCjV010000 0035"	CJE1503	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein	-
"CJ_10001297"	CJE1504	-	Cj1310c	-	CCO1419	-	1	1	1	1	-1	1	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001302"	CJE1506	neuA	Cj1311	neuA2	CCO1421	-	1	1	1	1	0	1	1	0	1	1	1	acylneuramate cytidyltransferase	CMP-N-acetylneuraminic acid synthetase	CMP-N-acetylneuraminic acid synthetase (neuA)
"CJ_10001304"	CJE1507	-	Cj1312	-	CCO1422	-	0	1	0	1	0	1	-1	1	-1	1	1	possible flagellar protein	flagellar protein, putative	probable flagellar protein Cj1312
"CJ_10001306"	CJE1508	-	Cj1313	-	CCO1423	-	-1	0	0	1	1	1	1	1	1	1	1	possible flagellar protein	acetyltransferase, GNAT family	probable flagellar protein Cj1313
"opCcV010000 0429"	CJE1509	-	Cj1314c	-	CCO1424	-	1	1	1	1	1	1	1	1	1	1	1	putative cyclase	imidazoleglycerol phosphate synthase, cyclase	probable cyclase Cj1314c

"opCcV0100000900"	CJE1510	-	Cj1315c	-	CCO1425	hisH	1	1	1	1	1	1	1	1	1	1	amidotransferase	imidazole glycerol phosphate synthase, glutamine	imidazole glycerol phosphate synthase, glutamine
"opCcV0100000878"	CJE1511	-	Cj1316c	-	CCO1426	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	flagellin modification protein, PseA	PseA protein
"CJ_10001308"	CJE1512	neuB	Cj1317	neuB3	CCO1427	-	1	1	1	1	1	1	1	1	1	1	N-acetylneuraminic acid synthetase	N-acetylneuraminic acid synthetase	spore coat polysaccharide biosynthesis protein
"CJ_10001309"	CJE1513	-	Cj1319	-	CCO1430	-	1	1	-1	1	1	1	-1	1	1	1	putative nucleotide sugar epimerase/dehydratase family	NAD-dependent epimerase/dehydratase family	probable nucleotide sugar dehydratase Cj1319
"CJ_10001332"	CJE1514	-	Cj1320	-	CCO1431	-	1	1	-1	1	1	1	-1	1	1	1	putative aminotransferase (degT family)	aminotransferase, DegT family	probable aminotransferase (degT family) Cj1320
"opCjV0100000414"	CJE1515	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	formyltransferase, putative	-
"CJ_10001347"	CJE1516	ptmC	Cj1327	neuB2	CCO1433	-	1	1	-1	1	1	1	-1	1	1	1	N-acetylneuraminic acid synthetase	N-acetylneuraminic acid synthetase	capsular polysaccharide biosynthesis protein
"CJ_10001348"	CJE1517	neuC	Cj1328	neuC2	CCO1434	-	0	1	1	1	1	0	1	1	1	1	putative N-acetylglucosamine-6-phosphate	UDP-N-acetylglucosamine 2-epimerase	UDP-N-acetylglucosamine 2-epimerase
"CJ_10001349"	CJE1518	-	Cj1329	-	CCO1435	mpg	1	1	1	1	-1	1	1	1	1	1	putative sugar-phosphate nucleotide transferase	nucleotidyltransferase family protein	Mannose-1-phosphate guanylyltransferase
"CJ_10001364"	CJE1519	-	Cj1330	-	CCO1436	-	-1	0	-1	1	1	-1	-1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001366"	CJE1520	ptmB	Cj1331	ptmB	CCO1437	-	1	1	-1	1	1	1	1	1	1	1	acylneuraminate cytidyltransferase (flagellin)	posttranslational flagellin modification protein	acylneuraminate cytidyltransferase, putative
"CJ_10001369"	CJE1521	ptmA	Cj1332	ptmA	CCO1438	-	1	1	1	1	1	1	-1	1	1	1	putative oxidoreductase (flagellin)	flagellin modification protein A	probable oxidoreductase (flagellin modification)
"opCcV0100001710"	CJE1522	-	Cj1318	-	CCO1428	-	1	1	-1	1	0	1	1	-1	1	1	hypothetical protein	motility accessory factor	conserved hypothetical protein
"CJ_10001373"	CJE1523	-	Cj1334	-	CCO1440	-	0	1	0	-1	-1	-1	-1	1	1	1	hypothetical protein Cj1334 (1318 family)	motility accessory factor	hypothetical protein Cj1334 (1318 family)
"opCcV0100000701"	CJE1524	-	Cj1318	-	CCO1429	-	1	1	1	1	-1	1	-1	1	1	1	hypothetical protein	motility accessory factor	conserved hypothetical protein
"CJ_10001374"	CJE1525	-	Cj1337	-	CCO1442	-	1	1	-1	1	1	1	1	-1	1	1	hypothetical protein	motility accessory factor	conserved hypothetical protein
"opCjjV010000192"	CJE1526	flaB	Cj1338c	flaB	-	-	1	1	1	1	1	1	0	1	1	1	flagellin	flagellin	-
"opCjV0100000504"	CJE1527	-	-	-	-	-	0	0	1	1	1	0	0	1	1	0	-	hypothetical protein	-
"opCjjV010000104"	CJE1528	flaA	Cj1339c	flaA	-	-	1	1	1	1	1	1	1	0	1	1	flagellin	flagellin	-
"CJ_10001388"	CJE1529	-	Cj1340c	-	CCO1445	-	1	1	1	1	-1	1	1	1	1	-1	hypothetical protein	motility accessory factor	conserved hypothetical protein
"CJ_10001394"	CJE1531	-	Cj1342c	-	CCO1448	-	1	-1	-1	1	1	1	-1	0	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001396"	CJE1532	ctsG	Cj1343c	-	CCO1449	ctsG	1	-1	1	1	1	1	-1	1	-1	-1	putative periplasmic protein	transformation system protein	transformation system protein
"CJ_10001398"	CJE1533	gcp	Cj1344c	-	CCO1450	gcp	1	1	1	1	1	1	1	0	1	1	putative glycoprotease	O-sialoglycoprotein endopeptidase	O-sialoglycoprotein endopeptidase
"CJ_10001399"	CJE1534	-	Cj1345c	-	CCO1451	-	1	1	1	1	1	-1	1	1	0	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1345c
"CJ_10001400"	CJE1535	dxr	Cj1346c	dxr	CCO1452	dxr	1	1	1	1	1	1	1	-1	1	1	putative 1-deoxy-D-xylulose 5-phosphate	1-deoxy-D-xylulose 5-phosphate reductoisomerase	1-deoxy-D-xylulose 5-phosphate reductoisomerase
"CJ_10001401"	CJE1536	cdsA	Cj1347c	cdsA	CCO1453	cdsA	1	1	1	1	1	1	1	1	1	1	phosphatidate	phosphatidate	phosphatidate

"CJ_10001402"	CJE1537	-	Cj1348c	-	CCO1454	-	1	1	1	1	1	1	1	1	1	1	1	cytidylyltransferase putative coiled-coil protein	cytidylyltransferase hypothetical protein	cytidylyltransferase conserved hypothetical protein
"CJ_10001403"	CJE1538	-	Cj1349c	-	CCO1455	-	1	1	1	1	1	1	1	1	1	1	1	possible fibronectin/fibrinogen- binding protein	fibronectin/fibrinogen binding protein,	fibronectin/fibrinogen- binding protein,
"CJ_10001417"	CJE1539	-	Cj1350	mobA	CCO1456	-	1	1	1	1	1	1	1	1	1	1	1	putative molybdopterin- guanine dinucleotide	molybdopterin-guanine dinucleotide biosynthesis	molybdopterin-guanine dinucleotide biosynthesis
"CJ_10001420"	CJE1540	pIdA	Cj1351	pIdA	CCO1457	-	1	1	1	1	1	1	1	1	1	1	1	phospholipase A	phospholipase A	phospholipase A
"CJ_10001423"	CJE1541	ceuB	Cj1352	ceuB	CCO1458	ceuB	1	1	1	1	1	-1	-1	1	-1	-1	1	enterochelin uptake permease	enterochelin ABC transporter, permease protein	enterochelin ABC transporter, permease protein
"CJ_10001425"	CJE1542	ceuC	Cj1353	ceuC	CCO1459	ceuC	1	1	1	1	1	1	1	0	1	1	1	enterochelin uptake permease	enterochelin ABC transporter, permease protein	enterochelin ABC transporter, permease protein
"CJ_10001427"	CJE1543	ceuD	Cj1354	ceuD	CCO1460	ceuD	1	1	1	1	1	1	1	1	1	1	1	enterochelin uptake ATP- binding protein	enterochelin ABC transporter, ATP-binding	enterochelin ABC transporter, ATP-binding
"CJ_10001428"	CJE1544	-	Cj1355	ceuE	CCO1461	ceuE	0	0	1	1	1	1	1	1	1	1	1	enterochelin uptake periplasmic binding protein	pseudogene	enterochelin ABC transporter, periplasmic
"CJ_10001429"	CJE1545	-	Cj1356c	-	CCO1462	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	conserved hypothetical protein
"CJ_10001430"	CJE1546	nrfA	Cj1357c	-	CCO1464	-	1	1	1	1	1	1	1	1	1	1	1	putative periplasmic cytochrome C	cytochrome c552	probable periplasmic cytochrome C Cj1357c
"CJ_10001431"	CJE1547	nrfH	Cj1358c	-	CCO1465	-	1	1	1	1	1	1	0	1	1	1	1	putative periplasmic cytochrome C	cytochrome c-type protein nrfH	probable periplasmic cytochrome C Cj1358c
"CJ_10001432"	CJE1548	ppk	Cj1359	ppk	CCO1466	ppk	1	1	1	1	1	1	1	1	1	1	1	polyphosphate kinase	polyphosphate kinase	polyphosphate kinase
"opCjV010000 0483"	CJE1549	-	-	-	-	-	1	-1	1	-1	1	1	1	1	-1	-1	-	-	pseudogene	-
"opCjV010000 0871"	CJE1550	-	-	-	-	-	1	-1	1	-1	1	1	1	1	-1	-1	-	-	hypothetical protein	-
"opCjV010000 0227"	CJE1551	-	-	-	-	-	1	-1	1	-1	1	1	-1	1	-1	-1	-	-	hypothetical protein	-
"opCjV010000 0140"	CJE1552	-	-	-	-	-	-1	-1	1	-1	1	1	-1	1	-1	-1	-	-	hypothetical protein	-
"CJ_10000351"	CJE1553	-	Cj1361c	-	CCO1471	-	0	-1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000353"	CJE1554	ruvB	Cj1362	ruvB	CCO1472	ruvB	1	1	1	1	1	1	1	1	1	1	1	holliday junction DNA helicase	Holliday junction DNA helicase RuvB	Holliday junction DNA helicase RuvB
"CJ_10000355"	CJE1555	-	Cj1363	amaA	CCO1473	-	1	0	1	1	1	1	1	1	1	1	1	acid membrane antigen A	hypothetical protein	acid membrane antigen A Cj1363
"CJ_10000357"	CJE1556	fumC	Cj1364c	fumC	CCO1474	fumC	1	0	1	0	1	1	1	1	1	1	1	fumarate hydratase	fumarate hydratase, class II	fumarate hydratase, class II
"CJ_10000361"	CJE1557	-	Cj1365c	-	-	-	1	1	1	1	1	1	1	1	1	1	1	putative secreted serine protease	serine protease, subtilase family	-
"CJ_10000363"	CJE1558	glmS	Cj1366c	glmS	CCO1475	glmS	0	1	1	1	1	1	1	1	1	1	1	glucosamine--fructose-6- phosphate	D-fructose-6-phosphate amidotransferase	glucosamine--fructose-6- phosphate
"CJ_10000366"	CJE1559	-	Cj1367c	-	CCO1476	-	0	-1	1	1	1	1	1	1	1	1	1	possible nucleotidyltransferase	GlnD family protein	probable nucleotidyltransferase Cj1367c
"CJ_10000367"	CJE1560	-	Cj1368	-	CCO1477	-	1	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR00423
"CJ_10000369"	CJE1561	-	Cj1369	-	CCO1478	-	0	-1	-1	1	1	-1	1	1	0	1	1	putative transmembrane transport protein	xanthine/uracil permease family protein	probable transmembrane transport protein Cj1369
"CJ_10000378"	CJE1562	-	Cj1370	-	CCO1479	-	1	1	1	1	1	1	1	1	1	0	1	putative nucleotide phosphoribosyltransferase	nucleotide phosphoribosyltransferase, putative	probable nucleotide phosphoribosyltransferase

"CJ_10000380"	CJE1563	-	Cj1371	-	CCO1480	-	1	1	1	1	1	1	1	1	1	putative periplasmic protein (vacJ homolog)	lipoprotein, VacJ family	probable periplasmic protein (vacJ homolog)
"CJ_10000382"	CJE1564	-	Cj1372	-	CCO1481	-	0	1	1	1	1	1	1	1	1	putative periplasmic protein	toluene tolerance protein, putative	probable periplasmic protein Cj1372
"CJ_10000384"	CJE1565	-	Cj1373	-	CCO1482	-	0	1	1	1	0	1	1	0	1	putative integral membrane protein	hypothetical protein	conserved hypothetical integral membrane
"CJ_10000386"	CJE1566	-	Cj1374c	-	CCO1483	-	0	0	1	1	1	1	1	1	-1	hypothetical protein	non-canonical purine NTP pyrophosphatase,	Ham1 family
"opCjjV010000122"	CJE1567	-	Cj1375	-	-	-	1	1	1	1	1	1	1	1	1	putative efflux protein	multidrug resistance efflux transporter,	-
"CJ_10000392"	CJE1568	-	Cj1377c	-	CCO1490	-	1	1	1	1	0	1	1	1	0	putative ferredoxin	iron-sulfur cluster-binding domain protein	iron-sulfur cluster-binding protein, putative
"CJ_10000394"	CJE1569	selA	Cj1378	selA	CCO1489	selA	0	1	1	1	1	1	1	1	1	L-seryl-tRNA(SeC) selenium transferase	selenocysteine synthase	L-seryl-tRNA selenium transferase
"CJ_10000396"	CJE1570	selB	Cj1379	selB	CCO1488	selB	0	1	1	1	0	1	1	1	1	putative selenocysteine-specific elongation	selenocysteine-specific elongation factor	selenocysteine-specific translation elongation
"CJ_10000404"	CJE1571	-	Cj1380	-	CCO1487	dsbC	1	0	0	1	0	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1380
"CJ_10000406"	CJE1572	-	Cj1381	-	CCO1486	-	0	1	0	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj1381
"CJ_10000408"	CJE1573	fldA	Cj1382c	fldA	CCO1492	-	0	1	1	1	1	1	1	1	1	flavodoxin	flavodoxin	flavodoxin
"CJ_10000411"	CJE1574	-	Cj1383c	-	CCO1493	-	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000413"	CJE1575	-	Cj1384c	-	CCO1494	-	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000415"	CJE1576	katA	Cj1385	katA	CCO1495	-	0	1	1	1	1	1	1	1	1	catalase	catalase	catalase
"CJ_10000418"	CJE1577	-	Cj1386	-	CCO1496	-	0	0	0	1	1	1	1	1	1	ankyrin-repeat containing protein	ankyrin repeat protein	ankyrin domain protein
"CJ_10000420"	CJE1578	-	Cj1387c	-	CCO1497	-	1	1	1	1	1	1	1	1	1	helix-turn-helix containing protein	hypothetical protein	conserved hypothetical protein
"CJ_10000423"	CJE1579	-	Cj1388	-	CCO1499	-	1	1	1	1	1	1	1	1	1	hypothetical protein	endoribonuclease L-PSP, putative	endoribonuclease L-PSP, putative
"CJ_10000425"	CJE1580	-	Cj1389	-	CCO1500	-	0	1	1	1	0	1	1	1	1	pseudogene	pseudogene	C4-dicarboxylate anaerobic carrier, putative
"CJ_10000435"	CJE1581	metC	Cj1393	-	CCO1502	metC	1	1	1	1	1	1	1	1	1	pseudogene	cystathionine beta-lyase	rhodanese domain protein/cystathionine
"CJ_10000438"	CJE1582	-	Cj1394	-	CCO1503	purB	0	1	0	1	1	1	1	0	-1	putative fumarate lyase	adenylosuccinate lyase	adenylosuccinate lyase
"CJ_10000440"	CJE1583	-	Cj1395	-	CCO1504	-	1	1	0	1	0	1	1	1	1	pseudogene	MmgE/PrpD family protein	MmgE/PrpD family protein, putative
"CJ_10000442"	CJE1584	-	Cj1397	-	CCO1505	-	1	1	1	1	1	1	1	1	1	hypothetical protein	feoA family protein	ferrous iron transport protein A, putative
"CJ_10000444"	CJE1585	-	Cj1398	feoB	CCO1506	feoB	1	1	1	1	0	1	1	0	1	ferrous iron transport protein	pseudogene	ferrous iron transport protein B
"CJ_10000446"	CJE1586	-	Cj1399c	hydA2	CCO1507	-	-1	-1	1	1	1	1	1	1	1	putative Ni/Fe-hydrogenase small subunit	hydrogenase, (NiFe)/(NiFeSe) small subunit	quinone-reactive Ni/Fe hydrogenase, small
"CJ_10000449"	CJE1587	fabI	Cj1400c	fabI	CCO1508	fabI	1	1	1	1	1	1	1	1	1	putative enoyl-[acyl-carrier-protein] reductase	enoyl-(acyl carrier protein) reductase	enoyl-(acyl-carrier-protein) reductase
"CJ_10000452"	CJE1588	tpiA	Cj1401c	tpiA	CCO1509	tpiA	1	1	1	1	1	1	1	1	-1	putative triosephosphate isomerase	triosephosphate isomerase	triosephosphate isomerase
"CJ_10000454"	CJE1589	pgk	Cj1402c	pgk	CCO1510	pgk	1	1	1	1	1	1	1	1	0	phosphoglycerate kinase	phosphoglycerate kinase	phosphoglycerate kinase
"CJ_10000477"	CJE1590	gapA	Cj1403c	gapA	CCO1511	gap	1	1	1	1	1	1	1	1	1	glyceraldehyde 3-phosphate dehydrogenase	glyceraldehyde 3-phosphate dehydrogenase A	glyceraldehyde-3-phosphate dehydrogenase, type
"CJ_10000478"	CJE1591	nadD	Cj1404	-	CCO1512	nadD	1	0	1	1	1	1	1	1	1	hypothetical protein	nicotinate (nicotinamide) nucleotide	nicotinate (nicotinamide) nucleotide
"CJ_10000479"	CJE1592	-	Cj1405	-	CCO1513	-	1	1	0	1	1	1	1	1	1	hypothetical protein	hypothetical protein	iojap-related protein

"CJ_10000480"	CJE1593	-	Cj1406c	-	-	-	0	0	0	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10000481"	CJE1594	algC	Cj1407c	-	CCO1515	-	1	0	1	1	1	1	1	1	1	1	putative phospho-sugar mutase	phosphomannomutase/phosphoglucomutase	phosphomannomutase (algC){Pseudomonas
"CJ_10000482"	CJE1595	fliL	Cj1408	fliL	CCO1516	-	1	1	1	1	1	1	1	1	1	1	possible flagellar protein	flagellar basal body-associated protein	probable flagellar protein Cj1408
"CJ_10000483"	CJE1596	acpS	Cj1409	acpS	CCO1517	acpS	0	0	0	1	1	1	1	1	1	1	putative holo-[acyl-carrier protein] synthase	4'-phosphopantetheinyl transferase	holo-(acyl-carrier-protein) synthase
"CJ_10000484"	CJE1597	-	Cj1410c	-	CCO1518	-	1	0	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1410c
"CJ_10000485"	CJE1598	-	Cj1411c	-	CCO1519	-	1	1	1	1	1	1	1	1	1	1	putative cytochrome P450	cytochrome P450 family protein	probable cytochrome P450 Cj1411c
"CJ_10000486"	CJE1599	-	Cj1412c	-	CCO1520	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1412c
"CJ_10000497"	CJE1600	kpsS	Cj1413c	-	CCO1521	-	1	1	1	1	1	1	1	1	1	1	possible polysaccharide modification protein	capsule polysaccharide export protein KpsS	capsule polysaccharide biosynthesis protein
"CJ_10000498"	CJE1601	kpsC	Cj1414c	-	CCO1522	-	1	1	1	1	1	1	1	1	1	1	possible polysaccharide modification protein	capsule polysaccharide export protein KpsC	capsule polysaccharide biosynthesis protein
"opCjV010000467"	CJE1602	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	capsular polysaccharide biosynthesis protein,	-
"opCjV010000584"	CJE1603	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1				-	capsular polysaccharide biosynthesis protein,	-
"opCjV010000374"	CJE1604	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1	1	-1	-	-	capsular polysaccharide biosynthesis protein,	-
"opCjV010000251"	CJE1605	-	-	-	-	-	-1	-1	-1	-1	1						-	capsular polysaccharide biosynthesis protein,	-
"opCjV010000333"	CJE1606	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1	-1	-1	-	-	haloacid dehalogenase-like hydrolase	-
"opCjV010000661"	CJE1607	-	-	-	-	-	-1	-1	-1	-1	-1	-1	1	0	-1	-	-	hypothetical protein	-
"CJ_10000523"	CJE1608	-	Cj1423c	-	-	-	1	1	1	1	-1	1	-1	-1	1	1	putative sugar-phosphate nucleotidyltransferase	capsular biosynthesis nucleotidyltransferase,	-
"CJ_10000525"	CJE1609	-	Cj1424c	gmhA2	-	-	1	1	0	1	-1	1	-1	1	1	1	putative phosphoheptose isomerase	phosphoheptose isomerase	-
"CJ_10000527"	CJE1610	-	Cj1425c	-	-	-	0	0	1	1		1	1	-1	1	1	putative sugar kinase	capsular biosynthesis sugar kinase, putative	-
"opCjV0100001098"	CJE1611	webK	-	-	-	-	0	0	1	-1	1	1	-1	1	-1	-1	-	GDP-mannose 4,6-dehydratase	-
"opCjV010000619"	CJE1612	fel	Cj1428c	fel	-	-	-1	-1	-1	-1	-1	-1	-1	1	1	-1	-	GDP-L-fucose synthetase	-
"opCjV010000987"	CJE1613	-	-	-	-	-	-1	-1	-1	1	-1		1	1	-1	-	-	capsular polysaccharide biosynthesis protein,	-
"opCjV010000978"	CJE1614	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	0	-1	-	-	hypothetical protein	-
"opCjV0100001043"	CJE1615	-	-	-	-	-	-1	-1	-1	1	-1	-1	1	0	-1	-	-	capsular polysaccharide biosynthesis	-
"opCjV010000633"	CJE1616	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	capsular polysaccharide biosynthesis protein,	-
"CJ_10000581"	CJE1617	kpsF	Cj1443c	kpsF	CCO1550	-	1	1	1	1	1	1	1	1	1	1	KpsF protein	arabinose-5-phosphate isomerase	KpsF protein Cj1443c
"CJ_10000583"	CJE1618	kpsD	Cj1444c	kpsD	CCO1551	-	0	0	1	1	1	1	1	1	0	1	putative capsule polysaccharide export system	capsular polysaccharide ABC transporter,	polysaccharide biosynthesis protein, putative
"CJ_10000585"	CJE1619	kpsE	Cj1445c	kpsE	CCO1552	-	1	0	1	1	1	1	1	1	0	1	putative capsule polysaccharide export system	capsular polysaccharide ABC transporter	probable capsule polysaccharide export system

"opCcV0100000731"	CJE1620	kpsT	Cj1447c	kpsT	CCO1553	-	1	0	1	1	1	1	1	1	0	1	putative capsule polysaccharide export	capsular polysaccharide ABC transporter,	lipopolysaccharide ABC export system,
"opCcV0100001067"	CJE1621	kpsM	Cj1448c	kpsM	CCO1554	abcT3	-1	-1	0	1	1	-1	1	1	-1	1	-	capsular polysaccharide ABC transporter,	KpsM
"CJ_10000587"	CJE1622	-	Cj1449c	-	CCO1555	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000588"	CJE1623	-	Cj1450	-	CCO1556	-	1	0	1	1	1	1	1	1	1	-1	putative ATP/GTP-binding protein	hypothetical protein	conserved hypothetical protein
"CJ_10000589"	CJE1624	-	Cj1451	dut	CCO1557	-	1	1	0	1	1	1	1	1	1	1	putative dUTPase	deoxyuridine triphosphatase domain protein	probable dUTP diphosphatase Cj1451
"CJ_10000590"	CJE1625	-	Cj1452	-	CCO1558	-	0	1	0	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1452
"CJ_10000591"	CJE1626	-	Cj1453c	-	CCO1559	-	1	0	1	1	0	1	1	0	0	1	hypothetical protein	PP-loop family protein	conserved hypothetical protein
"CJ_10000607"	CJE1627	yliG	Cj1454c	-	CCO1560	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	MiaB-like tRNA modifying enzyme YliG, TIGR01125	MiaB-like tRNA modifying enzyme YliG, TIGR01125
"CJ_10000609"	CJE1628	prfB	Cj1455	prfB	CCO1561	prfB	1	1	1	1	1	1	1	1	1	1	peptide chain release factor 2	peptide chain release factor 2	peptide chain release factor 2
"opCcV0100000633"	CJE1629	-	Cj1456c	-	CCO1562	-	1	1	0	0	-1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	hypothetical protein
"opCcV0100000250"	CJE1630	-	-	-	CCO1563	-	1	0	0	1	1	0	1	0	1	1	-	hypothetical protein	lipoprotein, putative
"CJ_10000612"	CJE1631	-	Cj1457c	-	CCO1564	-	1	0	1	1	1	1	1	1	0	1	hypothetical protein	tRNA pseudouridine synthase D	tRNA pseudouridine synthase D, putative
"CJ_10000613"	CJE1632	-	Cj1458c	thiL	CCO1565	-	1	0	1	1	1	1	1	1	0	1	putative thiamin-monophosphate kinase	thiamine monophosphate kinase	thiamin-monophosphate kinase, putative
"CJ_10000614"	CJE1633	-	Cj1459	-	CCO1566	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000615"	CJE1634	-	Cj1460	-	CCO1567	-	1	0	1	1	0	1	0	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000616"	CJE1635	-	Cj1461	-	CCO1568	-	1	1	1	1	0	1	1	1	1	1	possible DNA methylase	site-specific DNA methyltransferase, putative	probable DNA methylase Cj1461
"CJ_10000617"	CJE1636	flgI	Cj1462	flgI	CCO1569	flgI	1	1	1	1	1	1	1	1	1	1	flagellar P-ring protein	flagellar P-ring protein precursor	flagellar P-ring protein FlgI
"CJ_10000618"	CJE1637	-	Cj1463	-	CCO1570	-	1	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001312"	CJE1638	-	Cj1464	-	CCO1571	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001314"	CJE1639	-	Cj1465	-	CCO1572	-	1	1	1	1	0	1	1	-1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001317"	CJE1640	-	Cj1466	flgK	CCO1573	flgK	1	1	1	1	1	1	1	1	1	1	putative flagellar hook-associated protein	flagellar hook-associated protein	flagellar hook-associated protein 1 (HAP1)
"CJ_10001320"	CJE1641	-	Cj1467	-	CCO1574	-	0	1	0	1	0	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001325"	CJE1642	-	Cj1468	-	CCO1575	-	1	1	-1	1	0	1	1	1	1	1	putative integral membrane protein	hypothetical protein	membrane protein, putative
"CJ_10001328"	CJE1643	ctsF	Cj1470c	-	CCO1576	ctsF	1	1	1	1	1	1	1	1	1	1	pseudogene	general secretory pathway protein F	general secretory pathway protein F
"CJ_10001331"	CJE1644	ctsE	Cj1471c	-	CCO1577	ctsE	0	1	1	1	-1	1	1	1	1	1	putative type II protein secretion system E	general secretory pathway protein E	general secretory pathway protein E
"CJ_10001335"	CJE1645	ctsX	Cj1472c	-	CCO1578	ctsX	1	1	1	1	1	1	1	1	1	1	putative membrane protein	transformation system protein	transformation system protein
"CJ_10001338"	CJE1646	ctsP	Cj1473c	-	CCO1579	ctsP	1	1	1	1	1	1	1	1	1	1	putative ATP/GTP-binding protein	transformation system protein	transformation system protein
"CJ_10001341"	CJE1647	ctsD	Cj1474c	-	CCO1580	ctsD	1	1	1	1	1	1	1	1	1	1	putative type II protein	general secretory pathway	general secretory pathway

"CJ_10001352"	CJE1648	ctsR	Cj1475c	-	CCO1581	ctsR	1	1	1	1	1	1	1	1	1	secretion system D hypothetical protein	protein D transformation system protein	protein D transformation system protein	
"CJ_10001354"	CJE1649	-	Cj1476c	-	CCO1582	-	1	-1	1	1	1	1	-1	1	-1	pyruvate-flavodoxin oxidoreductase	pyruvate ferredoxin/flavodoxin oxidoreductase	pyruvate ferredoxin/flavodoxin oxidoreductase	
"CJ_10001356"	CJE1650	-	Cj1477c	-	CCO1583	-	1	1	1	1	1	1	1	1	1	putative hydrolase	HAD-superfamily hydrolase, subfamily IA, variant	phosphoglycolate phosphatase	
"CJ_10001359"	CJE1651	cadF	Cj1478c	cadF	CCO1584	cadF	1	1	1	1	1	1	1	1	1	outer membrane fibronectin-binding protein	fibronectin-binding protein	fibronectin-binding protein	
"opCcV010000 0256"	CJE1652	rpsI	Cj1479c	rpsI	CCO1585	rpsI	1	1	1	1	1	1	1	1	1	30S ribosomal protein S9	30S ribosomal protein S9	ribosomal protein S9	
"CJ_10001362"	CJE1653	rplM	Cj1480c	rplM	CCO1586	rplM	1	1	1	1	1	1	1	1	1	50S ribosomal protein L13	50S ribosomal protein L13	ribosomal protein L13	
"CJ_10001365"	CJE1654	-	Cj1481c	-	CCO1589	-	1	1	1	1	1	1	1	1	1	putative helicase	ATP-dependent DNA helicase, UvrD/REP family	helicase, putative	
"CJ_10001367"	CJE1655	-	Cj1482c	-	CCO1590	-	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10001370"	CJE1656	-	Cj1483c	-	CCO1591	-	1	0	1	1	1	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj1483c	
"CJ_10001372"	CJE1657	-	Cj1484c	-	CCO1592	-	1	1	1	1	1	1	1	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1484c	
"opCcV010000 0330"	CJE1658	-	Cj1485c	-	CCO1593	-	0	1	1	0	1	0	1	1	0	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1485c -related	
"CJ_10001378"	CJE1659	-	Cj1486c	-	CCO1594	-	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1486c -related	
"CJ_10001381"	CJE1660	ccoP	Cj1487c	ccoP	CCO1595	ccoP	1	1	1	1	1	1	1	1	1	cb-type cytochrome C oxidase subunit III	cytochrome c oxidase, cbb3-type, subunit III	cytochrome c oxidase, cbb3-type, subunit III	
"CJ_10001383"	CJE1661	ccoQ	Cj1488c	ccoQ	CCO1596	-	1	1	1	1	1	1	1	1	1	cb-type cytochrome C oxidase subunit IV	cytochrome c oxidase, cbb3-type, subunit IV	conserved hypothetical protein	
"CJ_10001385"	CJE1662	ccoO	Cj1489c	ccoO	CCO1597	ccoO	1	1	1	1	1	1	1	1	1	cb-type cytochrome C oxidase subunit II	cytochrome c oxidase, cbb3-type, subunit II	cytochrome c oxidase, cbb3-type, subunit II	
"CJ_10001387"	CJE1663	ccoN	Cj1490c	ccoN	CCO1598	ccoN	1	1	0	0	-1	0	1	0	1	cb-type cytochrome C oxidase subunit I	cytochrome c oxidase, cbb3-type, subunit I	cytochrome c oxidase, cbb3-type, subunit I	
"CJ_10001389"	CJE1664	-	Cj1491c	-	CCO1599	-	1	1	1	1	1	1	0	1	1	putative two-component regulator	DNA-binding response regulator	transcriptional regulatory protein KdpE,	
"CJ_10001392"	CJE1665	-	Cj1492c	-	CCO1600	-	1	1	1	1	-1	1	1	1	1	putative two-component sensor	sensory box sensor histidine kinase, putative	signal-transducing histidine kinase, putative	
"CJ_10001395"	CJE1666	-	Cj1493c	-	CCO1601	-	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1493c	
"CJ_10001397"	CJE1667	carA	Cj1494c	carA	CCO1602	carA	0	1	1	1		1	-1		1	carbamoyl-phosphate synthase small chain	carbamoyl-phosphate synthase small subunit	carbamoyl-phosphate synthase, small subunit	
"CJ_10001406"	CJE1668	-	Cj1495c	-	CCO1603	-	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10001409"	CJE1669	-	Cj1496c	-	CCO1604	-	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1496c	
"CJ_10001411"	CJE1670	-	Cj1497c	-	CCO1605	-	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10001413"	CJE1671	purA	Cj1498c	purA	CCO1606	purA	1	1	1	1	1	1	1	1	1	adenylosuccinate synthetase	adenylosuccinate synthetase	adenylosuccinate synthetase	
"opCcV010000 0651"	CJE1672	-	-	-	CCO1608	-	1	1	1	1	1	1	1	-1	1	0	-	hypothetical protein	probable integral membrane protein Cj1500
"CJ_10001415"	CJE1673	-	Cj1500	-	CCO1609	-	1	1	1	1	1	1	1	1	1	putative integral membrane protein	hypothetical protein	conserved hypothetical protein	
"opCjjV010000 193"	CJE1674	-	Cj1501	-	-	-	1	1	0	1	1	1	0	1	1	1	hypothetical protein	hypothetical protein	-

"CJ_10001418"	CJE1675	putP	Cj1502c	putP	CCO1611	putP	1	1	1	1	1	1	1	1	1	1	sodium/proline symporter	sodium/proline permease	proline permease (putP)
"CJ_10001421"	CJE1676	-	Cj1503c	putA	CCO1612	-	1	1	1	1	1	1	1	1	1	1	putative proline	bifunctional putA protein, putative	proline dehydrogenase
"CJ_10001424"	CJE1677	selD	Cj1504c	selD	CCO1613	selD	1	1	1	1	1	-1	-1	1	-1	-1	putative selenide, water dikinase	selenide, water dikinase	selenide, water dikinase
"CJ_10001426"	CJE1678	-	Cj1505c	-	CCO1614	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	predicted Transcriptional regulator
"CJ_10001439"	CJE1679	-	Cj1506c	-	-	-	1	1	1	1	1	1	1	1	1	1	putative MCP-type signal transduction protein	methyl-accepting chemotaxis protein	-
"CJ_10001440"	CJE1680	-	Cj1507c	-	CCO1615	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	ModE repressor domain protein	N-terminal HTH domain of molybdenum-binding
"CJ_10001441"	CJE1681	fdhD	Cj1508c	fdhD	CCO1616	fdhD	1	1	1	1	1	1	1	1	1	1	FdhD protein	formate dehydrogenase accessory protein	formate dehydrogenase family accessory protein
"CJ_10001442"	CJE1682	fdhC	Cj1509c	fdhC	CCO1617	-	1	1	1	1	1	1	1	1	1	1	putative formate dehydrogenase, cytochrome B	formate dehydrogenase, cytochrome b subunit	formate dehydrogenase cytochrome B chain
"CJ_10001443"	CJE1683	fdhB	Cj1510c	fdhB	CCO1618	-	1	0	1	1	1	1	1	1	1	1	putative formate dehydrogenase iron-sulfur	formate dehydrogenase, iron-sulfur subunit	formate dehydrogenase, iron-sulfur subunit
"CJ_10001444"	CJE1684	fdhA	Cj1511c	fdhA	-	-	1	1	1	1	1	1	1	1	1	1	putative formate dehydrogenase large subunit	formate dehydrogenase, alpha subunit,	-
"CJ_10001445"	CJE1685	-	Cj1513c	-	CCO1620	-	1	1	1	1	1	1	1	1	1	1	possible periplasmic protein	tat domain protein	probable periplasmic protein Cj1513c -related
"CJ_10001446"	CJE1686	-	Cj1514c	-	CCO1621	-	1	1	1	1	1	1	1	1	0	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"opCjV0100000299"	CJE1687	-	-	-	-	-	0	0	0	1	1	0	0	1	0	1	-	hypothetical protein	-
"CJ_10001447"	CJE1688	nspC	Cj1515c	-	CCO1622	nspC	1	1	1	1	1	1	1	1	1	1	putative decarboxylase	carboxynorspermidine decarboxylase	carboxynorspermidine decarboxylase
"CJ_10001448"	CJE1689	-	Cj1516	-	CCO1623	-	1	1	1	1	1	1	1	0	1	1	putative periplasmic oxidoreductase	oxidoreductase, putative	probable periplasmic oxidoreductase Cj1516
"CJ_10001459"	CJE1690	-	Cj1517	moaD	CCO1624	-	0	1	1	1	1	1	0	1	1	1	possible molybdopterin converting factor,	thiS family protein	probable molybdopterin converting factor, chain
"CJ_10001460"	CJE1691	-	Cj1518	moaE	CCO1625	-	1	1	1	1	1	1	1	1	1	1	possible molybdopterin converting factor,	molybdopterin converting factor, subunit 2	molybdopterin biosynthesis protein E chain
"opCcV0100001980"	CJE1692	-	-	-	CCO1626	-	1	1	1	1	-1	1	1	1	1	1	-	hypothetical protein	probable molybdopterin biosynthesis protein
"CJ_10001461"	CJE1693	-	Cj1519	moeA2	-	-	1	1	1	1	1	1	1	1	1	1	putative molybdopterin biosynthesis protein	molybdopterin biosynthesis MoeA protein,	-
"CJ_10001463"	CJE1694	cas2	Cj1521c	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	CRISPR-associated protein Cas2	-
"CJ_10001464"	CJE1695	cas1	Cj1522c	-	-	-	1	0	0	1	1	1	1	1	0	1	hypothetical protein	CRISPR-associated protein Cas1	-
"CJ_10001465"	CJE1697	-	Cj1523c	-	-	-	0	1	0	1	1	1	0	1	1	1	hypothetical protein Cj1523c	pseudogene	-
"opCjV0100000404"	CJE1698	-	-	-	-	-	1	1	0	1	1	1	-1	-1	1	1	-	hypothetical protein	-
"CJ_10001466"	CJE1699	-	Cj1528	-	CCO1630	-	1	1	1	1	1	1	1	1	1	1	pseudogene	pseudogene	C4-dicarboxylate anaerobic carrier, putative
"CJ_10001467"	CJE1700	purM	Cj1529c	purM	CCO1632	purM	0	0	1	1	1	1	1	1	1	1	phosphoribosylformylglycin amidine cyclo-ligase	phosphoribosylaminoimidazole synthetase	phosphoribosylformylglycin amidine cyclo-ligase
"CJ_10001468"	CJE1701	-	Cj1530	-	CCO1633	-	0	-1	1	1	-1	1	1	-1	1	1	putative ATP/GTP-binding protein	dephospho-CoA kinase	dephospho-CoA kinase
"CJ_10001484"	CJE1702	dapF	Cj1531	dapF	CCO1634	dapF	1	1	1	1	1	1	1	1	1	1	putative diaminopimelate epimerase	diaminopimelate epimerase	diaminopimelate epimerase
"CJ_10001486"	CJE1703	-	Cj1532	-	CCO1635	-	0	1	0	1	1	1	1	1	1	1	possible periplasmic protein	mannosyl-glycoprotein	bax protein, putative

"CJ_10001488"	CJE1704	-	Cj1533c	-	CCO1636	-	1	0	1	1	1	1	1	1	0	1	putative helix-turn-helix containing protein	hypothetical protein	conserved hypothetical ATP-binding protein,
"CJ_10001490"	CJE1705	-	Cj1534c	-	CCO1641	-	1	-1	0	1	1	1	0	1	1	1	possible bacterioferritin	bacterioferritin, putative	bacterioferritin, putative
"CJ_10001492"	CJE1706	pgi	Cj1535c	pgi	CCO1642	pgi	1	1	1	1	1	1	1	1	1	1	putative glucose-6-phosphate isomerase	glucose-6-phosphate isomerase	glucose-6-phosphate isomerase
"CJ_10001493"	CJE1707	galU	Cj1536c	galU	CCO1643	galU	1	1	1	1	1	1	1	1	1	1	UTP--glucose-1-phosphate uridylyltransferase	UTP-glucose-1-phosphate uridylyltransferase	UTP-glucose-1-phosphate uridylyltransferase
"CJ_10001494"	CJE1708	acs	Cj1537c	acs	CCO1644	-	1	1	1	1	1	1	1	1	1	1	acetyl-coenzyme A synthetase	acetyl-coenzyme A synthetase	acetyl-CoA synthetase
"CJ_10001495"	CJE1709	-	Cj1538c	-	CCO1645	-	1	1	1	1	1	1	1	1	1	1	putative anion-uptake ABC-transport system	tungsten ABC transporter, ATP-binding protein,	glutamine ABC transporter, ATP-binding protein
"CJ_10001496"	CJE1710	-	Cj1539c	-	CCO1646	-	1	1	1	1	1	1	1	1	1	1	putative anion-uptake ABC-transport system	tungstate ABC transporter, permease protein,	ABC transporter, permease protein
"CJ_10001497"	CJE1711	-	Cj1540	-	CCO1647	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	tungstate ABC transporter, periplasmic	probable periplasmic protein Cj1540
"CJ_10001513"	CJE1712	-	Cj1541	-	CCO1648	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	tungstate ABC transporter, periplasmic	hypothetical protein
"CJ_10001515"	CJE1713	-	Cj1542	-	CCO1652	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR00370
"CJ_10001517"	CJE1714	-	Cj1543	-	CCO1653	-	1	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	urea amidolyase-related protein
"CJ_10001518"	CJE1715	-	Cj1544c	-	CCO1655	-	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	integral membrane protein	probable integral membrane protein Cj1544c
"CJ_10001520"	CJE1716	-	Cj1545c	-	-	-	1	1	1	1	1	-1	1	1	1	1	MdaB protein homolog	flavodoxin-like fold domain protein	-
"CJ_10001521"	CJE1717	-	Cj1546	-	-	-	-1	1	1	1	1	1	1	1	1	1	hypothetical protein	transcriptional regulator, putative	-
"CJ_10001522"	CJE1718	-	Cj1547	-	CCO1659	-	0	1	1	1	1	1	1	-1	1	1	homolog of BLC protein	pseudogene	outer membrane lipoprotein BLC, putative
"opCcV0100001745"	CJE1719	-	Cj1548c	-	CCO1660	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	putative NADP-dependent alcohol dehydrogenase	oxidoreductase, zinc-binding dehydrogenase	probable alcohol dehydrogenase (NADP) Cj1548c
"opCjV0100000360"	CJE1720	-	-	-	-	-	-1	-1	1	-1	1	-1	-1	1	-1	-1	-	pseudogene	-
"opCjV0100000526"	CJE1721	-	-	-	-	-	-1	-1	1	1	-1	-1	1	1	-1	-	-	RloC protein, putative	-
"opCjV0100000237"	CJE1722	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	pseudogene	-
"opCjV0100000763"	CJE1723	-	-	-	-	-	1	-	1	-1	1	-1	-1	1	1	-1	-	MloA protein, putative	-
"opCjV0100000327"	CJE1724	hsdM	-	-	-	-	-1	-1	-1	-1	-1	-1	1	1	-1	-	-	type I restriction-modification system, M	-
"opCjV0100000827"	CJE1725	-	-	-	-	-	1	-1	0	1	1	1	0	1	1	1	-	4-carboxymuconolactone decarboxylase, putative	-
"opCjV0100000389"	CJE1726	-	-	-	-	-	0	-1	0	-1	1	1	0	1	-1	-1	-	hypothetical protein	-
"opCjV0100000776"	CJE1727	-	-	-	-	-	1	-1	1	-1	1	1	1	1	-1	-1	-	hypothetical protein	-
"opCjV0100000501"	CJE1728	-	-	-	-	-	1	-1	1	-1	1	1	1	1	0	-1	-	transporter, putative	-
"CJ_10001555"	CJE1729	-	Cj1555c	-	-	-	-1	1	1	1	1	-1	-1	-1	1	1	hypothetical protein	pseudogene	-
"CJ_10001558"	CJE1730	-	Cj1558	-	-	-	-1	1	0	1	1	1	0	0	1	1	putative membrane protein	permease, putative	-
"CJ_10001560"	CJE1731	arsR	Cj1561	-	CCO1673	-	-1	1	1	1	0	1	1	0	1	1	putative transcriptional regulator	arsenical resistance operon repressor	transcriptional regulator, ArsR family
"opCjV010000"	CJE1732	arsC	-	-	-	-	1	-1	-1	1	1	1	1	1	-1	-1	-	arsenate reductase	-

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"opCjV0100000235"	CJE1733	-	-	-	-	-	1	-1	-1	-1	1	1	1	1	-1	-1	-	arsenical-resistance protein, putative	-	
"CJ_10001562"	CJE1734	-	Cj1563c	-	CCO1677	-	1	1	1	1	0	1	1	1	1	1	putative transcriptional regulator	pseudogene	probable transcription regulator Cj1563c	
"CJ_10001580"	CJE1736	pflA	Cj1565c	pflA	CCO0279	-	1	1	1	1	0	1	1	1	1	1	paralysed flagellum protein	paralyzed flagella protein PflA	paralysed flagella protein (pflA), putative	
"CJ_10001582"	CJE1737	nuoN	Cj1566c	nuoN	CCO0278	nuoN	1	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain N	NADH-quinone oxidoreductase, N subunit	NADH-ubiquinone oxidoreductase, NQO14 subunit	
"CJ_10001584"	CJE1738	nuoM	Cj1567c	nuoM	CCO0277	nuoM	0	1	1	1	1	1	0	1	1	0	NADH dehydrogenase I chain M	NADH-quinone oxidoreductase, M subunit	NADH-ubiquinone oxidoreductase, NQO13 subunit	
"CJ_10001586"	CJE1739	nuoL	Cj1568c	nuoL	CCO0276	-	1	1	0	1	1	1	1	1	1	0	NADH dehydrogenase I chain L	NADH dehydrogenase subunit L	NADH2 dehydrogenase (ubiquinone) I chain L	
"CJ_10001587"	CJE1740	nuoK	Cj1569c	nuoK	CCO0275	-	1	1	1	1	1	1	0	-1	1	1	NADH dehydrogenase I chain K	NADH-quinone oxidoreductase, K subunit	NADH2 dehydrogenase (ubiquinone) I chain K	
"CJ_10001588"	CJE1741	nuoJ	Cj1570c	nuoJ	CCO0274	nuoJ	1	1	1	1	1	1	0	1	1	1	NADH dehydrogenase I chain J	NADH dehydrogenase subunit J	NADH-ubiquinone oxidoreductase, NQO10 subunit	
"CJ_10001590"	CJE1742	nuoI	Cj1571c	nuoI	CCO0273	-	1	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain I	NADH dehydrogenase subunit I	NADH2 dehydrogenase (ubiquinone) I chain I	
"CJ_10001591"	CJE1743	nuoH	Cj1572c	nuoH	CCO0272	-	1	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain H	NADH dehydrogenase subunit H	NADH2 dehydrogenase (ubiquinone) chain I	
"CJ_10001593"	CJE1744	nuoG	Cj1573c	nuoG	CCO0271	-	1	1	0	1	1	1	1	0	1	1	probable NADH dehydrogenase I chain G	NADH dehydrogenase gamma subunit	probable NADH2 dehydrogenase (ubiquinone) I	
"CJ_10000511"	CJE1745	-	Cj1574c	-	CCO0270	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000513"	CJE1746	-	Cj1575c	-	CCO0269	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	
"CJ_10000515"	CJE1747	nuoD	Cj1576c	nuoD	CCO0268	-	-1	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain D	NADH dehydrogenase delta subunit	NADH2 dehydrogenase (ubiquinone) I chain D	
"CJ_10000518"	CJE1748	nuoC	Cj1577c	nuoC	CCO0267	-	1	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain C	NADH dehydrogenase subunit C	NADH2 dehydrogenase (ubiquinone) I chain C	
"opCcV0100000039"	CJE1749	nuoB	Cj1578c	nuoB	CCO0266	-	0	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain B	NADH dehydrogenase beta subunit	NADH2 dehydrogenase (ubiquinone) I chain B	
"CJ_10000521"	CJE1750	nuoA	Cj1579c	nuoA	CCO0265	-	1	1	1	1	1	1	1	1	1	1	NADH dehydrogenase I chain A	NADH dehydrogenase alpha subunit	NADH2 dehydrogenase (ubiquinone) I chain A	
"CJ_10000522"	CJE1751	-	Cj1580c	-	CCO0264	abcT11	1	1	1	1	1	1	1	0	1	1	putative peptide ABC-transport system	peptide ABC transporter, ATP-binding protein	oligopeptide ABC-transporter, ATP-binding	
"CJ_10000524"	CJE1752	-	Cj1581c	-	CCO0263	dppD		0	1	1	1	1	1	0	1	1	putative peptide ABC-transport system	peptide ABC transporter, ATP-binding protein	peptide ABC transporter, ATP-binding protein	
"CJ_10000526"	CJE1753	-	Cj1582c	-	CCO0262	-	1	1	1	1	1	1	1	1	1	1	putative peptide ABC-transport system permease	peptide ABC transporter, permease protein	peptide ABC transporter, permease protein	
"CJ_10000528"	CJE1754	-	Cj1583c	-	CCO0261	-	1	1	1	1	1	1	1	1	1	1	putative peptide ABC-transport system permease	peptide ABC transporter, permease protein	peptide ABC transporter, permease protein,	
"CJ_10000540"	CJE1755	-	Cj1584c	-	CCO0260	-	1	-1	1	1	1	1	1	1	1	1	putative peptide ABC-transport system	peptide ABC transporter, periplasmic	peptide ABC transporter, peptide-binding	
"CJ_10000542"	CJE1756	-	Cj1585c	-	CCO0259	-	1	1	-1	1	1	-1	-1	-1	1	1	putative oxidoreductase	oxidoreductase, FAD-binding, iron-sulfur	probable oxidoreductase Cj1585c	
"CJ_10000545"	CJE1757	-	Cj1586	-	CCO0258	hmpA	1	1	1	1	1	1	1	1	1	1	putative bacterial haemoglobin	flavoheomprotein, truncation	flavoheomprotein	
"CJ_10000547"	CJE1758	-	Cj1587c	-	CCO0256	-	0	0	0	1	1	1	1	1	0	1	putative ABC transporter	cyclic peptide ABC transporter, ATP-binding	pyoverdine ABC export system,	

"CJ_10000549"	CJE1759	-	Cj1588c	-	-	-	0	1	1	1	0	1	0	0	1	1	putative transmembrane transport protein	pseudogene	-
"opCjV0100001112"	CJE1760	-	-	-	CCO0255	proP	0	1	0	1	1	1	1	1	1	1	-	hypothetical protein	proline/betaine transporter (proP)
"CJ_10000551"	CJE1761	-	Cj1589	-	-	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	metallo-beta-lactamase domain protein	-
"CJ_10000552"	CJE1762	infA	Cj1590	infA	CCO0244	infA	1	1	1	1	1	1	-1	1	1	1	translation initiation factor IF-1	translation initiation factor IF-1	translation initiation factor IF-1
"opCjV010000151"	CJE1763	rpmJ	Cj1591	rpmJ	-	-	1	1	1	1	1	1	1	1	1	1	50S ribosomal protein L36	ribosomal protein L36	-
"CJ_10000555"	CJE1764	rpsM	Cj1592	rpsM	CCO0243	rpsM	1	1	1	1	0	1	1	1	1	1	30S ribosomal protein S13	30S ribosomal protein S13	ribosomal protein S13p/S18e
"opCjV0100001449"	CJE1765	rpsK	Cj1593	rpsK	CCO0242	rpsK	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S11	30S ribosomal protein S11	ribosomal protein S11
"CJ_10000568"	CJE1766	rpsD	Cj1594	rpsD	CCO0241	rpsD	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S4	30S ribosomal protein S4	ribosomal protein S4
"CJ_10000571"	CJE1767	rpoA	Cj1595	rpoA	CCO0240	rpoA	1	1	1	1	1	1	1	1	1	1	DNA-directed RNA polymerase alpha chain	DNA-directed RNA polymerase alpha subunit	DNA-directed RNA polymerase, alpha subunit
"CJ_10000573"	CJE1768	rplQ	Cj1596	rplQ	CCO0239	rplQ	1	1	1	1	1	1	0	1	1	1	50S ribosomal protein L17	50S ribosomal protein L17	ribosomal protein L17
"CJ_10000575"	CJE1769	hisG	Cj1597	hisG	CCO0238	hisG	1	1	1	1	1	1	1	1	1	1	ATP phosphoribosyltransferase	ATP phosphoribosyltransferase	ATP phosphoribosyltransferase
"CJ_10000577"	CJE1770	hisD	Cj1598	hisD	CCO0237	hisD	-1	0	1	1	1	1	1	1	1	1	histidinol dehydrogenase	histidinol dehydrogenase	histidinol dehydrogenase
"CJ_10000579"	CJE1771	hisB	Cj1599	hisB	CCO0236	hisB	1	1	1	1	1	1	1	1	1	1	imidazoleglycerol-phosphate	imidazole glycerol-phosphate	imidazoleglycerol-phosphate
"CJ_10000580"	CJE1772	-	Cj1600	hisH	CCO0235	hisH	1	1	1	1	1	1	1	1	1	1	amidotransferase HisH	imidazole glycerol phosphate synthase, glutamine	imidazole glycerol phosphate synthase, glutamine
"CJ_10000582"	CJE1773	hisA	Cj1601	hisA	CCO0234	-	1	1	1	1	1	1	1	1	-1	1	phosphoribosylformimino-5-aminoimidazole	phosphoribosylformimino-5-aminoimidazole	phosphoribosylformimino-5-aminoimidazole
"CJ_10000584"	CJE1774	-	Cj1602	-	-	-	1	1	0	1	1	0	0	0	1	1	hypothetical protein	HrgA protein	-
"CJ_10000586"	CJE1775	-	Cj1603	hisF	CCO0233	hisF	1	1	1	1	1	1	1	1	0	1	cyclase	imidazole glycerol phosphate synthase subunit	imidazoleglycerol phosphate synthase, cyclase
"CJ_10000598"	CJE1776	hisI	Cj1604	hisI	CCO0232	-	0	1	0	1	1	1	0	0	1	1	phosphoribosyl-AMP cyclohydrolase/possible	phosphoribosyl-ATP	phosphoribosyl-ATP
"CJ_10000600"	CJE1777	-	Cj1605c	dapD	CCO0230	dapD	1	1	1	1	1	1	1	0	1	1	possible	2,3,4,5-tetrahydropyridine-2-carboxylate	tetrahydrodipicolinate N-succinyltransferase
"CJ_10000602"	CJE1778	mrp	Cj1606c	mrp	CCO0229	-	1	1	1	1	1	1	1	1	1	1	putative ATP/GTP-binding protein (mrp protein)	ATP/GTP-binding protein	ATP-binding protein (mpr)
"CJ_10000603"	CJE1779	ispDF	Cj1607	-	CCO0228	-	1	1	0	1	1	1	1	1	1	1	hypothetical protein	bifunctional 2-C-methyl-D-erythritol 4-phosphate	ispD/ispF bifunctional enzyme
"CJ_10000604"	CJE1780	-	Cj1608	-	CCO0227	-	1	1	1	1	1	1	1	1	1	1	possible two-component regulator	DNA-binding response regulator, putative	response regulator, putative
"CJ_10000605"	CJE1781	-	Cj1609	-	CCO0226	-	1	1	1	1	1	1	1	1	1	1	possible sulfate adenylyltransferase	ATP-sulfurylase family protein	sulfate adenylyltransferase, putative
"CJ_10000606"	CJE1782	pgpA	Cj1610	pgpA	CCO0225	pgpA	1	0	1	1	1	1	-1	1	0	1	putative phosphatidylglycerophosphatase	phosphatidylglycerophosphatase A	phosphatidylglycerophosphatase A
"CJ_10000608"	CJE1783	rpsT	Cj1611	rpsT	CCO0224	rpsT	1	1	1	1	1	1	1	1	1	1	30S ribosomal protein S20	30S ribosomal protein S20	ribosomal protein S20
"CJ_10000610"	CJE1784	prfA	Cj1612	prfA	CCO0223	prfA	1	1	1	1	1	1	1	1	1	1	peptide chain release factor 1	peptide chain release factor 1	peptide chain release factor 1
"CJ_10000611"	CJE1785	-	Cj1613c	-	CCO0222	-	1	1	1	1	1	1	1	1	0	1	hypothetical protein	hypothetical protein	Protein of unknown function (DUF319) family
"CJ_10000633"	CJE1786	chuA	Cj1614	chuA	CCO0221	-	0	1	-1	1	1	1	1	0	1	0	haemin uptake system outer membrane receptor	TonB-dependent heme receptor	iron-regulated outer membrane virulence protein,
"CJ_10000635"	CJE1787	-	Cj1615	chuB	CCO0220	-	1	1	1	1	1	1	1	1	1	1	putative haemin uptake	hemin ABC transporter,	probable heme uptake

"CJ_10000637"	CJE1788	-	Cj1616	chuC	CCO0219	-	1	0	1	1	1	1	1	1	1	1	system permease protein	permease protein,	system permease protein
"CJ_10000639"	CJE1789	-	Cj1617	chuD	CCO0218	-	1	0	0	0	1	0	1	0	1	1	putative haemin uptake system ATP-binding	hemin ABC transporter, ATP-binding protein,	iron (III) ABC transporter, ATP-binding protein
"CJ_10000641"	CJE1790	-	Cj1618c	-	CCO0217	-	1	1	1	1	1	1	1	1	1	1	putative haemin uptake system periplasmic	hemin ABC transporter, periplasmic hemin-binding	iron ABC transporter, periplasmic
"CJ_10000642"	CJE1791	kgtP	Cj1619	kgtP	CCO0215	kgtP	1	1	1	1	1	1	1	1	1	1	hypothetical protein	pseudogene	conserved hypothetical protein
"CJ_10000642"	CJE1791	kgtP	Cj1619	kgtP	CCO0215	kgtP	1	1	1	1	1	1	1	1	1	1	alpha-ketoglutarate permease	alpha-ketoglutarate permease	dicarboxylic acid transport protein
"CJ_10000643"	CJE1792	mutY	Cj1620c	mutY	CCO0214	mutY	1	1	1	1	1	1	1	0	1	1	A/G-specific adenine glycosylase	A/G-specific adenine glycosylase	A/G-specific adenine glycosylase
"CJ_10000644"	CJE1793	-	Cj1621	-	CCO0209	-	1	1	0	1	1	1	0	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1621
"CJ_10000645"	CJE1794	-	Cj1622	ribD	CCO0202	ribD	1	1	1	1	0	1	1	1	1	1	putative riboflavin-specific deaminase	riboflavin biosynthesis protein RibD, putative	riboflavin biosynthesis protein (ribG)
"CJ_10000646"	CJE1795	-	Cj1623	-	CCO0201	-	1	1	1	1	1	1	1	0	1	1	putative membrane protein	hypothetical protein	probable membrane protein Cj1623
"CJ_10000662"	CJE1796	sdaA	Cj1624c	sdaA	CCO0200	sdaA	1	1	1	1	1	1	1	0	1	1	L-serine dehydratase	L-serine ammonia-lyase	L-serine dehydratase 1
"CJ_10000664"	CJE1797	sdaC	Cj1625c	sdaC	CCO0199	sdaC	1	1	1	1	1	1	1	1	1	1	serine transporter	serine transporter	serine transporter (sdaC)
"CJ_10000666"	CJE1798	-	Cj1626c	-	CCO0198	-	1	1	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1626c
"CJ_10000668"	CJE1799	-	Cj1627c	-	CCO0197	-	0	0	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000670"	CJE1800	-	Cj1628	exbB2	CCO0195	-	1	1	1	1	1	1	1	1	1	1	putative exbB/tolQ family transport protein	TonB system transport protein ExbB	probable exbB/tolQ family transport protein
"CJ_10000671"	CJE1801	exbD	Cj1629	exbD2	CCO0194	-	-1	-1	0	1	1	1	1	1	1	1	putative exbD/tolR family transport protein	biopolymer transport exbD protein	tolR protein, putative
"CJ_10000672"	CJE1802	-	Cj1630	tonB2	-	-	0	1	0	1	1	1	1	1	1	1	putative tonB transport protein	pseudogene	-
"CJ_10000673"	CJE1803	-	Cj1631c	-	CCO0193	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000674"	CJE1804	-	Cj1632c	-	CCO0192	-	1	1	1	1	-1	1	1	-1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1632c -related
"CJ_10000675"	CJE1805	-	Cj1633	-	CCO0191	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000692"	CJE1806	aroC	Cj1634c	aroC	CCO0180	aroC	0	0	1	1	1	1	1	1	1	1	chorismate synthase	chorismate synthase	chorismate synthase
"CJ_10000694"	CJE1807	rncS	Cj1635c	rnc	CCO0179	-	1	1	1	1	1	1	1	1	1	1	ribonuclease III	ribonuclease III	ribonuclease III Cj1635c
"CJ_10000696"	CJE1808	rnhA	Cj1636c	rnhA	CCO0178	rnhA	1	1	1	1	1	1	1	1	1	1	ribonuclease HI	ribonuclease H	RNase H
"CJ_10000698"	CJE1809	-	Cj1637c	-	CCO0177	-	1	0	1	1	1	1	1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1637c
"CJ_10000700"	CJE1810	dnaG	Cj1638	dnaG	CCO0176	-	1	1	1	1	1	1	1	1	1	1	DNA primase	DNA primase	DNA primase, probable CP0919, putative
"opCcV0100001481"	CJE1811	-	Cj1639	-	CCO0175	-	1	1	0	1	1	0	0	1	-1	1	nifU protein homolog	NifU family protein	nifU protein homolog Cj1639
"CJ_10000701"	CJE1812	-	Cj1640	-	CCO0174	-	0	0	0	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000702"	CJE1813	murE	Cj1641	murE	CCO0173	murE	1	1	1	1	1	1	1	1	1	1	UDP-N-acetylmuramoylalanyl-D-glutamate--2,	UDP-N-acetylmuramoylalanyl-D-glutamate--2,	UDP-N-acetylmuramoylalanyl-D-glutamate--2,
"CJ_10000703"	CJE1814	-	Cj1642	-	CCO0172	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein TIGR00103
"CJ_10000704"	CJE1815	-	Cj1643	-	CCO0171	-	0	0	1	1	1	1	1	1	1	1	putative periplasmic protein	PDZ domain protein	probable periplasmic protein Cj1643
"CJ_10000718"	CJE1816	ispA	Cj1644	ispA	CCO0170	ispA	1	-1	-1	-1	-1	-1	-1	-1	-1	-1	geranyltranstransferase	geranyltranstransferase	geranyltranstransferase (ispA)

"CJ_10000720"	CJE1817	tkf	Cj1645	tkf	CCO0169	tkf	1	1	1	1	1	1	1	1	1	1	1	transketolase	transketolase	transketolase
"CJ_10000722"	CJE1818	-	Cj1646	iamB	CCO0168	-	1	1	1	1	1	1	1	1	1	1	1	putative ABC transport system permease protein	ABC transporter, permease protein, putative	conserved hypothetical integral membrane
"CJ_10000724"	CJE1819	-	Cj1647	iamA	CCO0167	-	1	1	1	1	1	1	1	1	1	1	1	putative ABC transport system ATP-binding	ABC transporter, ATP-binding protein	ABC transporter
"CJ_10000726"	CJE1820	-	Cj1648	-	CCO0166	-	1	1	1	1	1	1	1	1	1	1	1	possible ABC transport system periplasmic	ABC transporter, periplasmic substrate-binding	conserved hypothetical protein
"CJ_10000728"	CJE1821	-	Cj1649	-	CCO0165	-	-1	0	0	1	1	1	0	1	1	1	1	putative lipoprotein	lipoprotein, putative	probable lipoprotein Cj1649
"CJ_10000729"	CJE1822	-	Cj1650	-	CCO0164	-	0	0	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000730"	CJE1823	map	Cj1651c	map	CCO0163	map	0	1	1	1	1	1	1	1	1	1	1	methionine aminopeptidase	methionine aminopeptidase	methionine aminopeptidase, type I
"CJ_10000731"	CJE1824	murI	Cj1652c	murI	CCO0162	murI	1	1	1	1	0	1	1	1	1	1	1	glutamate racemase	glutamate racemase	glutamate racemase
"CJ_10000732"	CJE1825	nlpC	Cj1653c	-	CCO0161	-	1	1	1	1	1	1	-1	-1	1	1	1	probable lipoprotein	lipoprotein NlpC	lipoprotein, NLP/P60 family
"CJ_10000742"	CJE1826	-	Cj1654c	nhaA2	CCO0158	nhaA	0	1	1	1	1	0	-1	-1	1	1	1	Na(+)/H(+) antiporter	Na+/H+ antiporter NhaA	Na+/H+ antiporter NhaA
"CJ_10000743"	CJE1827	-	Cj1654c	nhaA3	CCO0159	-	1	1	1	1	1	1	0	1	1	1	1	Na(+)/H(+) antiporter	Na+/H+ antiporter NhaA	Na+/H+ antiporter Cj1654c
"CJ_10000744"	CJE1828	-	Cj1654c	nhaA4	CCO0160	-	1	1	1	1	1	1	1	1	1	1	1	Na(+)/H(+) antiporter	Na+/H+ antiporter NhaA	Na+/H+ antiporter Cj1654c
"opCjV010000794"	CJE1829	-	-	-	-	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	hypothetical protein	hypothetical protein	-
"CJ_10000745"	CJE1830	-	Cj1658	-	CCO0156	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	iron permease, FTR1 family	probable integral membrane protein Cj1658
"CJ_10000746"	CJE1831	-	Cj1659	p19	CCO0155	-	1	1	1	1	1	1	1	1	1	1	1	periplasmic protein p19	hypothetical protein	periplasmic protein p19 Cj1659
"CJ_10000747"	CJE1832	-	Cj1660	-	CCO0154	-	1	1	1	1	1	1	1	0	1	1	1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj1660
"CJ_10000748"	CJE1833	-	Cj1661	-	CCO0153	-	1	1	1	1	1	1	1	1	1	1	1	possible ABC transport system permease protein	ABC transporter, permease protein	ABC transporter, ATP-binding protein, putative
"CJ_10000749"	CJE1834	-	Cj1662	-	CCO0152	-	1	1	1	1	1	1	1	1	1	1	1	putative integral membrane protein	ABC transporter, permease protein	permease protein, putative
"CJ_10000750"	CJE1835	-	Cj1663	-	CCO0151	-	0	1	1	1	1	1	1	1	1	1	1	putative ABC transport system ATP-binding	ABC transporter, ATP-binding protein	ABC transporter, ATP-binding protein
"CJ_10000751"	CJE1836	-	Cj1664	-	CCO0150	-	1	1	1	1	1	1	1	1	1	1	1	possible periplasmic thioredoxin	thioredoxin, homolog	probable periplasmic thioredoxin Cj1664
"CJ_10000771"	CJE1837	-	Cj1665	-	CCO0149	-	1	1	1	1	-1	1	1	0	0	1	1	possible lipoprotein thioredoxin	thioredoxin family protein	thioredoxin, putative
"CJ_10000772"	CJE1838	-	Cj1666c	-	-	-	1	1	1	1	-1	-1	1	-1	1	-1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10000773"	CJE1839	-	Cj1667c	-	-	-	1	1	1	1	-1	1	1	0	1	1	1	repA protein homolog	hypothetical protein	-
"CJ_10000774"	CJE1840	-	Cj1668c	-	-	-	1	1	1	1	0	0	-1	0	1	1	1	putative periplasmic protein	hypothetical protein	-
"CJ_10000775"	CJE1841	-	Cj1669c	-	CCO1793	-	1	1	1	1	-1	1	1	-1	1	1	1	putative ATP-dependent DNA ligase	DNA ligase	DNA ligase
"CJ_10000776"	CJE1842	-	Cj1670c	-	CCO1794	-	1	1	1	1	-1	1	1	-1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1670c
"CJ_10000777"	CJE1843	-	Cj1671c	-	CCO1795	-	0	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000778"	CJE1844	eno	Cj1672c	eno	CCO1796	eno	1	-	1	-1	1	-1	1	-1	1	0	0	enolase	phosphopyruvate hydratase	enolase
"CJ_10000779"	CJE1845	recA	Cj1673c	recA	CCO1797	recA	1	1	1	1	-1	1	1	-1	1	1	1	recA protein	recombinase A	recA protein
"CJ_10000780"	CJE1846	-	Cj1674	-	CCO1798	-	0	1	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10001473"	CJE1847	fliQ	Cj1675	fliQ	CCO1799	fliQ	1	1	1	1	1	1	1	1	1	1	1	flagellar biosynthetic protein	flagellar biosynthesis protein	flagellar biosynthetic protein FliQ
"CJ_10001475"	CJE1848	murB	Cj1676	murB	CCO1800	murB2	1	1	1	1	-1	1	1	0	1	1	1	putative UDP-N-acetylenolpyruvoylglucosa	UDP-N-acetylenolpyruvoylglucosa	UDP-N-acetylenolpyruvoylglucosam

Appendix

"CJ_10001483"	CJE1849	-	Cj1680c	-	CCO1802	-	1	1	1	1	0	1	1	1	1	mine	mine reductase	ine reductase
"CJ_10001485"	CJE1850	cysQ	Cj1681c	cysQ	CCO1803	-	1	1	1	1	1	1	1	0	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1680c
"CJ_10001487"	CJE1851	gltA	Cj1682c	gltA	CCO1804	gltA	1	1	1	1	1	1	0	1	1	citrate synthase	citrate synthase	citrate synthase I
"CJ_10001489"	CJE1852	-	Cj1684c	-	CCO1805	-	1	1	1	1	1	1	1	1	1	putative transmembrane transport protein	sodium/hydrogen exchanger family protein	probable transmembrane transport protein
"CJ_10001491"	CJE1853	bioB	Cj1685c	bioB	CCO1806	bioB	1	1	1	1	1	1	1	1	1	putative biotin synthase	biotin synthase	biotin synthase
"CJ_10001502"	CJE1854	topA	Cj1686c	topA	CCO1807	topA	1	1	1	1	1	0	1	0	1	DNA topoisomerase I	DNA topoisomerase I	DNA topoisomerase I
"CJ_10001505"	CJE1855	-	Cj1687	-	CCO1808	-	0	1	0	1	1	1	1	1	1	putative efflux protein	major facilitator superfamily protein	probable efflux protein Cj1687
"CJ_10001507"	CJE1856	secY	Cj1688c	secY	CCO1809	-	1	1	0	1	1	1	1	1	1	preprotein translocase subunit	preprotein translocase SecY	preprotein translocase, SecY subunit
"CJ_10001509"	CJE1857	rplO	Cj1689c	rplO	CCO1810	rplO	0	1	1	1	1	1	1	1	1	50S ribosomal protein L15	50S ribosomal protein L15	ribosomal protein L15
"opCcV0100000232"	CJE1858	rpsE	Cj1690c	rpsE	CCO1811	rpsE	1	1	1	1	1	1	1	1	1	30S ribosomal protein S5	30S ribosomal protein S5	ribosomal protein S5
"CJ_10001512"	CJE1859	rplR	Cj1691c	rplR	CCO1812	rplR	1	1	1	1	1	1	1	1	1	50S ribosomal protein L18	50S ribosomal protein L18	ribosomal protein L18
"CJ_10001514"	CJE1860	rplF	Cj1692c	rplF	CCO1813	-	1	1	1	1	1	1	1	1	1	50S ribosomal protein L6	50S ribosomal protein L6	50S ribosomal protein L6 Cj1692c
"CJ_10001516"	CJE1861	rpsH	Cj1693c	rpsH	CCO1814	rpsH	1	1	1	1	1	1	1	1	1	30S ribosomal protein S8	30S ribosomal protein S8	ribosomal protein S8
"opCcV0100000311"	CJE1862	rpsN	Cj1694c	rpsN	CCO1815	rpsN	1	1	1	1	1	1	1	1	1	30S ribosomal protein S14	ribosomal protein S14	ribosomal protein S14p/S29e
"CJ_10001519"	CJE1863	rplE	Cj1695c	rplE	CCO1816	-	1	1	1	1	1	1	1	1	0	50S ribosomal protein L5	50S ribosomal protein L5	50S ribosomal protein L5 Cj1695c
"opCcV0100001194"	CJE1864	rplN	Cj1696c	rplN	CCO1818	rplN	1	1	1	1	1	1	1	1	1	50S ribosomal protein L24	50S ribosomal protein L24	ribosomal protein L14
"CJ_10001540"	CJE1865	rplX	Cj1697c	rplX	CCO1817	rplX	1	1	1	1	1	1	1	1	1	50S ribosomal protein L14	50S ribosomal protein L14	ribosomal protein L24
"CJ_10001542"	CJE1866	rpsQ	Cj1698c	rpsQ	CCO1819	rpsQ	1	1	1	1	1	1	1	1	1	30S ribosomal protein S17	30S ribosomal protein S17	ribosomal protein S17
"CJ_10001544"	CJE1867	rpmC	Cj1699c	rpmC	CCO1820	rpmC	1	1	1	1	1	1	1	1	1	50S ribosomal protein L29	50S ribosomal protein L29	ribosomal protein L29
"CJ_10001546"	CJE1868	rplP	Cj1700c	rplP	CCO1821	rplP	1	1	1	1	1	1	1	1	1	50S ribosomal protein L16	50S ribosomal protein L16	ribosomal protein L16
"CJ_10001548"	CJE1869	rpsC	Cj1701c	rpsC	CCO1822	rpsC	1	1	1	1	1	1	0	1	1	30S ribosomal protein S3	30S ribosomal protein S3	ribosomal protein S3
"CJ_10001550"	CJE1870	rplV	Cj1702c	rplV	CCO1823	-	1	1	1	1	1	1	1	1	1	50S ribosomal protein L22	50S ribosomal protein L22	50S ribosomal protein L22 Cj1702c
"CJ_10001552"	CJE1871	rpsS	Cj1703c	rpsS	CCO1824	rpsS	1	1	1	1	1	1	1	1	1	30S ribosomal protein S19	30S ribosomal protein S19	ribosomal protein S19
"CJ_10001553"	CJE1872	rplB	Cj1704c	rplB	CCO1825	rplB	1	1	1	1	1	1	1	1	1	50S ribosomal protein L2	50S ribosomal protein L2	ribosomal protein L2
"CJ_10001556"	CJE1873	rplW	Cj1705c	rplW	CCO1826	rplW	1	1	1	1	1	1	0	1	1	50S ribosomal protein L23	50S ribosomal protein L23	ribosomal protein L23
"CJ_10001568"	CJE1874	rplD	Cj1706c	rplD	-	-	1	1	1	1	1	1	1	1	1	50S ribosomal protein L4	50S ribosomal protein L4	-
"CJ_10001570"	CJE1875	rplC	Cj1707c	rplC	-	-	1	1	1	1	1	1	1	1	1	50S ribosomal protein L3	50S ribosomal protein L3	-
"opCcV0100000037"	CJE1876	rpsJ	Cj1708c	rpsJ	CCO0002	rpsJ	1	1	1	1	1	1	1	1	1	30S ribosomal protein S10	30S ribosomal protein S10	ribosomal protein S10
"opCjV0100000962"	CJE1877	-	-	-	-	-	0	1	0	1	1	1	1	1	1	-	hypothetical protein	-
"CJ_10001573"	CJE1878	-	Cj1709c	-	CCO0003	-	1	1	1	1	1	1	1	1	1	putative ribosomal pseudouridine synthase	RNA pseudouridylation synthase family protein	RNA pseudouridine synthase, RsuA family
"CJ_10001575"	CJE1879	-	Cj1710c	-	CCO0004	-	1	1	1	1	1	1	1	1	1	hypothetical protein	metallo-beta-lactamase family protein	conserved hypothetical protein subfamily
"CJ_10001577"	CJE1880	ksgA	Cj1711c	ksgA	CCO0005	ksgA	1	1	1	1	1	1	0	1	1	putative dimethyladenosine transferase (16S rRNA)	dimethyladenosine transferase	dimethyladenosine transferase
"CJ_10001579"	CJE1881	-	Cj1712	-	CCO0006	-	1	1	1	1	1	1	1	1	1	hypothetical protein	hypothetical protein	purine nucleoside phosphorylase (punB)
"CJ_10001581"	CJE1882	-	Cj1713	-	CCO0007	-	1	1	1	1	1	1	1	1	1	hypothetical protein	radical SAM enzyme, Cfr family	radical SAM enzyme, Cfr family
"CJ_10001583"	CJE1883	-	Cj1714	-	CCO0008	-	1	1	1	1	0	1	-1	1	1	small hydrophobic protein	hypothetical protein	hypothetical protein

"opCjV0100000081"	CJE1884	-	-	-	-	-	-1	-1	1	-1	1	1	1	1	0	1	-	lipopolysaccharide core biosynthesis protein,	-	
"CJ_10001585"	CJE1885	-	Cj1715	-	CCO0014	-	1	1	1	1	1	1	1	1	1	1	1	putative acetyltransferase	acetyltransferase, GNAT family	acetyltransferase, GNAT family, putative
"opCcV0100001061"	CJE1886	leuD	Cj1716c	leuD	CCO0015	leuD	0	0	1	1	1	1	1	1	1	1	1	putative 3-isopropylmalate dehydratase small	3-isopropylmalate dehydratase, small subunit	3-isopropylmalate dehydratase, small subunit
"opCcV0100000744"	CJE1887	leuC	Cj1717c	leuC	CCO0016	leuC	1	-1	0	-1	1	-1	-1	1	1	1	1	3-isopropylmalate dehydratase large subunit	isopropylmalate isomerase large subunit	3-isopropylmalate dehydratase, large subunit
"CJ_10000014"	CJE1888	leuB	Cj1718c	leuB	CCO0017	leuB	1	1	1	1	1	1	1	0	1	1	1	3-isopropylmalate dehydrogenase	3-isopropylmalate dehydrogenase	3-isopropylmalate dehydrogenase
"CJ_10000016"	CJE1889	leuA	Cj1719c	leuA	CCO0018	leuA	1	1	1	1	1	1	1	1	1	1	1	2-isopropylmalate synthase	2-isopropylmalate synthase	2-isopropylmalate synthase
"CJ_10000018"	CJE1890	-	Cj1720	-	CCO0019	-	-1	1	-1	-1	-1	-1	1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein
"CJ_10000019"	CJE1891	-	Cj1721c	-	CCO0020	-	1	1	1	1	-1	1	1	-1	1	1	1	possible outer membrane protein	hypothetical protein	probable outer membrane protein Cj1721c
"CJ_10000021"	CJE1892	-	Cj1724c	-	CCO0022	-	1	0	1	1	0	1	1	1	1	1	1	hypothetical protein	GTP cyclohydrolase 1 family protein	GTP cyclohydrolase 1 subfamily, putative
"opCcV0100000585"	CJE1893	-	Cj1725	-	CCO0023	-	1	1	-1	1	1	-1	-1	-1	1	1	1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj1725
"CJ_10000037"	CJE1894	metA	Cj1726c	metA	CCO0024	metA	0	0	-1	1	1	1	1	-1	1	1	1	putative homoserine O-succinyltransferase	homoserine O-succinyltransferase	homoserine O-succinyltransferase
"opCcV0100001193"	CJE1895	metX	Cj1727c	metY	CCO0027	metC	0	1	-1	1	1	-1	1	-1	1	1	1	putative O-acetylhomoserine (thiol)-lyase	homoserine O-acetyltransferase	O-acetylhomoserine sulfhydrylase
"CJ_10000041"	CJE1896	-	Cj1729c	flgE2	CCO0029	-	1	1	1	1	-1	1	1	-1	1	1	1	flagellar hook subunit protein	flagellar hook protein	flagellar hook protein FlgE
"opCcV0100001311"	CJE1897	ruvC	Cj1731c	ruvC	CCO0031	ruvC	1	1	0	1	1	1	1	1	1	1	1	crossover junction endodeoxyribonuclease	Holliday junction resolvase	crossover junction endodeoxyribonuclease RuvC
"CJ_10000460"	-	-	-	virB8	-	-	1	-1	1	0	1	-1	1	1	0	0	0	VirB8	-	-
"CJ_10000461"	-	-	-	virB9	-	-	1	1	0	1	1	0	1	1	1	1	1	VirB9	-	-
"CJ_10000462"	-	-	-	virB10	-	-	1	1	1	1	0	1	1	-1	1	1	1	VirB10	-	-
"CJ_10000464"	-	-	-	virB11	-	-	1	1	1	1	-1	1	1	-1	1	1	1	VirB11	-	-
"CJ_10000465"	-	-	-	virD4	-	-	1	1	1	1	-1	1	1	-1	1	0	0	VirD4	-	-
"CJ_10000753"	-	-	-	topA	-	-	1	1	1	1	1	1	1	-1	1	0	0	TopA	-	-
"CJ_10000756"	-	-	-	ssb	-	-	1	1	1	1	-1	1	1	1	-1	1	1	Ssb	-	-
"CJ_10000803"	-	-	-	repA	-	-	-1	1	1	1	-1	1	1	0	0	-1	-1	pseudogene	-	-
"CJ_10000877"	-	-	-	virB4	-	-	0	1	1	1	-1	1	1	1	1	0	0	VirB4	-	-
"CJ_10001589"	-	-	-	repA	-	-	1	1	1	1	-1	1	1	1	0	0	0	putative replication protein A	-	-
"CJ_10001592"	-	-	-	repB	-	-	1	1	0	0	0	1	0	1	1	1	1	replication protein B	-	-
"CJ_10001594"	-	-	-	orf4	-	-	1	1	0	-1	-1	-1	1	1	1	1	1	unknown	-	-
"CJ_10001595"	-	-	-	orf3	-	-	1	1	1	1	1	-1	-1	1	1	0	0	unknown	-	-
"CJ_10001532"	-	-	-	Cj0008	-	-	1	1	1	1	-1	1	1	-1	1	1	1	hypothetical protein	-	-
"CJ_10000625"	-	-	-	Cj0170	-	-	1	1	1	1	-1	1	1	-1	1	1	1	hypothetical protein	-	-
"CJ_10000626"	-	-	-	Cj0171	-	-	1	1	1	1	0	1	1	0	1	1	1	hypothetical protein	-	-
"CJ_10001504"	-	-	-	Cj0260c	-	-	1	1	1	1	-1	1	1	-1	1	1	1	small hydrophobic protein	-	-
"CJ_10001534"	-	-	-	Cj0265c	-	-	1	1	1	1	-1	1	1	0	1	-1	-1	putative cytochrome C-type haem-binding	-	-
"opCjV010000018"	-	-	-	Cj0416	-	-	1	1	-1	1	-1	1	1	-1	1	1	1	hypothetical protein	-	-
"CJ_10000875"	-	-	-	Cj0417	-	-	1	1	1	1	-1	1	1	-1	1	1	1	hypothetical protein	-	-
"CJ_10000886"	-	-	-	Cj0423	-	-	1	1	1	1	-1	-1	1	-1	1	1	1	putative integral membrane protein	-	-

"CJ_10000887"	-	-	Cj0424	-	-	-	1	1	1	1	1	1	-1	-1	1	1	putative acidic periplasmic protein	-	-
"CJ_10000888"	-	-	Cj0425	-	-	-	1	0	1	1	1	1	1	1	1	1	putative periplasmic protein	-	-
"CJ_10000869"	-	-	Cj0565	-	-	-	1	1	1	-1	-1	0	-1	-1	-1	-1	pseudogene	-	-
"CJ_10000871"	-	-	Cj0566	-	-	-	0	1	1	1	-1	1	1	0	-1	1	hypothetical protein	-	-
"CJ_10000873"	-	-	Cj0567	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000876"	-	-	Cj0568	-	-	-	1	1	1	-1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000880"	-	-	Cj0569	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10001016"	-	-	Cj0628	-	-	-	1	1	1	1	0	-1	1	-1	-1	-1	putative lipoprotein	-	-
"opCjjV010000135"	-	-	Cj0629	-	-	-	1	1	0	1	-1	-1	1	-1	1	1	putative lipoprotein	-	-
"CJ_10000912"	-	-	Cj0747	-	-	-	0	-1	1	1	-1	1	1	0	1	1	hypothetical protein	-	-
"CJ_10000913"	-	-	Cj0748	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000102"	-	-	Cj0873c	-	-	-	1	1	1	1	0	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000254"	-	-	Cj0876c	-	-	-	1	0	1	1	1	1	1	1	1	1	putative periplasmic protein	-	-
"opCjjV010000034"	-	-	Cj0877c	-	-	-	1	1	0	1	1	-1	0	0	1	1	hypothetical protein	-	-
"opCjjV010000204"	-	-	Cj0969	-	-	-		1	1	1	0	1	1	0	1	1	pseudogene	-	-
"CJ_10001208"	-	-	Cj0987c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative integral membrane protein	-	-
"opCjjV010000207"	-	-	Cj0988c	-	-	-	0	1	1	1	1	1	1	1	1	0	-	-	-
"CJ_10001419"	-	-	Cj1055c	-	-	-	1	1	1	1	-1	-1	1	1	1	1	putative integral membrane protein	-	-
"CJ_10001116"	-	-	Cj1122c	wlaJ	-	-	0	1	1	1	-1	1	-1	-1	1	1	putative integral membrane protein	-	-
"CJ_10001159"	-	-	Cj1136	-	-	-	-1	1	1	-1	-1	-1	1	1	1	1	putative galactosyltransferase	-	-
"CJ_10001160"	-	-	Cj1137c	-	-	-	1	1	1	1	-1	1	1	-1	-1	1	hypothetical protein Cj1137c	-	-
"CJ_10001194"	-	-	Cj1138	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative galactosyltransferase	-	-
"CJ_10001198"	-	-	Cj1140	-	-	-	0	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10001200"	-	-	Cj1141	neuB1	-	-	1	1	1	1	-1	-1	-1	-1	1	1	N-acetylneuraminic acid synthetase	-	-
"CJ_10001202"	-	-	Cj1142	neuC1	-	-	1	1	1	1	1	1	0	-1	1	1	putative N-acetylglucosamine-6-phosphate	-	-
"CJ_10001203"	-	-	Cj1143	neuA1	-	-	1	1	-1	1	-1	-1	0	-1	1	-1	acylneuramate cytidyltransferase	-	-
"CJ_10001204"	-	-	Cj1144c	-	-	-	1	1	1	-1	-1	1	1	1	1	-1	hypothetical protein	-	-
"CJ_10001205"	-	-	Cj1145c	-	-	-	1	1	1	1	-1	1	1	0	1	-1	hypothetical protein	-	-
"CJ_10001270"	-	-	Cj1301	-	-	-	1	-1	1	1	-1	1	-1	1	1	1	hypothetical protein	-	-
"CJ_10001336"	-	-	Cj1321	-	-	-	1	1	1	1	-1	0	1	-1	1	1	putative transferase	-	-
"CJ_10001339"	-	-	Cj1322	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10001342"	-	-	Cj1323	-	-	-	1	1	1	1	-1	1	1	1	1	-1	hypothetical protein	-	-
"CJ_10001344"	-	-	Cj1324	-	-	-	1	1	-1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10001345"	-	-	Cj1325	-	-	-		1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10001346"	-	-	Cj1326	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000499"	-	-	Cj1415c	cysC	-	-	1	1	1	1	-1	1	0	-1	1	1	possible adenylylsulfate kinase	-	-
"CJ_10000500"	-	-	Cj1416c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative sugar nucleotidyltransferase	-	-

"CJ_10000501"	-	-	Cj1417c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000502"	-	-	Cj1418c	-	-	-	1	1	1	1	-1	1	1	-1	1	-1	hypothetical protein Cj1418c	-	-
"CJ_10000503"	-	-	Cj1419c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	possible methyltransferase	-	-
"CJ_10000504"	-	-	Cj1420c	-	-	-	1	1	1	1	1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000505"	-	-	Cj1421c	-	-	-	1	1	1	1	-1	1	-1	-1	-1	-1	possible sugar transferase	-	-
"CJ_10000506"	-	-	Cj1422c	-	-	-	1	1	1	1	-	1	1	-1	1	1	possible sugar transferase	-	-
"CJ_10000529"	-	-	Cj1426c	-	-	-	-	1	0	1	1	1	-1	-1	1	1	hypothetical protein	-	-
"CJ_10000530"	-	-	Cj1427c	-	-	-	-	1	1	1	-1	-1	1	-1	1	1	putative sugar-nucleotide	-	-
"CJ_10000531"	-	-	Cj1428c	fcI	-	-	1	1	1	1	1	0	1	-1	1	1	putative fucose synthetase	-	-
"CJ_10000532"	-	-	Cj1429c	-	-	-	-1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000533"	-	-	Cj1430c	-	-	-	1	1	1	1	1	1	-1	-1	1	1	putative nucleotide-sugar epimerase/dehydratase	-	-
"CJ_10000534"	-	-	Cj1431c	-	-	-	1	1	1	1	1	-1	-1	-1	1	1	hypothetical protein	-	-
"CJ_10000535"	-	-	Cj1432c	-	-	-	1	1	1	1	-1	-1	-1	-1	1	1	putative sugar transferase	-	-
"CJ_10000553"	-	-	Cj1433c	-	-	-	1	1	1	1	-1	1	1	1	1	-1	hypothetical protein	-	-
"CJ_10000554"	-	-	Cj1434c	-	-	-	-	1	0	1	1	1	1	-1	1	1	putative sugar transferase	-	-
"CJ_10000556"	-	-	Cj1435c	-	-	-	1	1	0	-1	-1	1	1	-1	1	-1	hypothetical protein	-	-
"CJ_10000557"	-	-	Cj1436c	-	-	-	0	1	0	1	-1	1	1	-1	1	1	putative aminotransferase	-	-
"CJ_10000558"	-	-	Cj1437c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative aminotransferase	-	-
"CJ_10000559"	-	-	Cj1438c	-	-	-	1	1	1	-	-1	-1	1	1	-	-	putative sugar transferase	-	-
"CJ_10000560"	-	-	Cj1439c	glf	-	-	1	1	1	-1	-1	1	1	-1	-1	-	UDP-galactopyranose mutase	-	-
"CJ_10000561"	-	-	Cj1440c	-	-	-	1	1	1	1	-1	1	1	-1	1	0	putative sugar transferase	-	-
"CJ_10000562"	-	-	Cj1441c	kfiD	-	-	1	1	1	-	-1	1	1	1	-1	1	putative UDP-glucose 6-dehydrogenase	-	-
"CJ_10000563"	-	-	Cj1442c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10001462"	-	-	Cj1520	-	-	-	0	1	-1	0	1	1	0	1	-1	1	hypothetical protein Cj1520	-	-
"CJ_10001523"	-	-	Cj1549c	-	-	-	1	1	0	1	-1	1	1	-1	1	1	putative type I restriction enzyme R protein	-	-
"CJ_10001525"	-	-	Cj1550c	-	-	-	1	1	0	1	-1	1	-1	-1	1	1	putative ATP/GTP-binding protein	-	-
"CJ_10001549"	-	-	Cj1551c	-	-	-	1	1	0	1	-1	0	1	-1	1	1	putative type I restriction enzyme S protein	-	-
"CJ_10001551"	-	-	Cj1552c	-	-	-	1	-	0	-	-	1	1	-	-	-1	hypothetical protein	-	-
"CJ_10001554"	-	-	Cj1553c	-	-	-	1	1	1	1	-1	1	1	-1	-1	-1	putative type I restriction enzyme M protein	-	-
"CJ_10001557"	-	-	Cj1556	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10001478"	-	-	Cj1677	-	-	-	1	1	1	1	-1	1	1	-1	1	-1	putative lipoprotein	-	-
"opCjjV010000086"	-	-	Cj1678	-	-	-	1	1	0	-1	-1	0	-1	-1	1	-1	-	-	-
"CJ_10001481"	-	-	Cj1679	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000072"	-	-	Cj1723c	-	-	-	1	1	1	1	-1	1	1	-1	1	1	putative periplasmic protein	-	-
"opCjjV010000036"	-	-	Cjp03	-	-	-	1	1	1	1	0	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000463"	-	-	Cjp04	-	-	-	-1	-1	0	1	1	1	1	0	1	1	hypothetical protein	-	-
"opCjjV010000083"	-	-	Cjp04	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000099"	-	-	Cjp04	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000008"	-	-	Cjp05	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000046"	-	-	Cjp06	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-

"opCjjV010000040"	-	-	Cjp07	-	-	-	1	1	1	-1	-1	1	-1	1	0	1	hypothetical protein	-	-
"opCjjV010000067"	-	-	Cjp07	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000467"	-	-	Cjp08	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000024"	-	-	Cjp08	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000469"	-	-	Cjp09	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000004"	-	-	Cjp09	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000752"	-	-	Cjp10	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000073"	-	-	Cjp10	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000103"	-	-	Cjp11	rnxB	-	-	1	1	1	1	1	1	1	1	1	1	ribonuclease P, RNA component	-	-
"CJ_10000754"	-	-	Cjp12	-	-	-	1	1	1	1	-1	1	1	-1	1	-1	hypothetical protein	-	-
"opCjjV010000022"	-	-	Cjp13	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000758"	-	-	Cjp14	-	-	-	0	0	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000760"	-	-	Cjp15	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000762"	-	-	Cjp16	-	-	-	1	1	1	1	-1	1	1	1	-1	1	hypothetical protein	-	-
"CJ_10000764"	-	-	Cjp17	-	-	-	1	1	1	1	-1	1	-1	-1	1	-1	hypothetical protein	-	-
"CJ_10000766"	-	-	Cjp18	-	-	-	1	1	1	1	-1	1	1	0	1	1	hypothetical protein	-	-
"opCjjV010000110"	-	-	Cjp19	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000113"	-	-	Cjp19	-	-	-	1	1	1	0	1		-1	1	1	-1	hypothetical protein	-	-
"CJ_10000785"	-	-	Cjp20	-	-	-	1	1	1	1	-1			-1	1	1	hypothetical protein	-	-
"opCjjV010000141"	-	-	Cjp20	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000787"	-	-	Cjp21	-	-	-	1	1	1	1	1	-1	-1	-1	-1	-1	hypothetical protein	-	-
"opCjjV010000005"	-	-	Cjp21	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000789"	-	-	Cjp22	-	-	-	1	0	0	1	-1	1	-1	-1	1	1	hypothetical protein	-	-
"opCjjV010000206"	-	-	Cjp22	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000792"	-	-	Cjp23	-	-	-	1	1	1	1	-1	1	1	1	1	-1	hypothetical protein	-	-
"opCjjV010000205"	-	-	Cjp23	-	-	-	0	1	0	1	1	1	0	1	1	1	hypothetical protein	-	-
"CJ_10000795"	-	-	Cjp24	-	-	-	0	1	1	1	-1	0	1	1	1	1	hypothetical protein	-	-
"opCjjV010000014"	-	-	Cjp24	-	-	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000006"	-	-	Cjp25	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000079"	-	-	Cjp25	-	-	-	1	1	0	-1	-1	1	0	1	1	-1	hypothetical protein	-	-
"CJ_10000799"	-	-	Cjp26	-	-	-	1	1	1	-1	-1	-1	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000179"	-	-	Cjp26	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000801"	-	-	Cjp27	-	-	-	1	1	1	1	0	1	1	1	-1	1	hypothetical protein	-	-
"opCjjV010000070"	-	-	Cjp27	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000080"	-	-	Cjp28	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-

"CJ_10000805"	-	-	Cjp29	-	-	-	1	1	1	1	-1	0	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000023"	-	-	Cjp29	-	-	-	1	1	1	1	1	1	1	1	0	1	hypothetical protein	-	-
"CJ_10000814"	-	-	Cjp30	-	-	-	1	1	1	1	-1	1	-1	-1	1	-1	hypothetical protein	-	-
"opCjjV010000064"	-	-	Cjp30	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000816"	-	-	Cjp31	-	-	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000818"	-	-	Cjp32	-	-	-	0	1	1	1	-1	-1	-1	-1	1	1	hypothetical protein	-	-
"opCjjV010000052"	-	-	Cjp32	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000821"	-	-	Cjp33	-	-	-	1	1	1	1	1	1	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000051"	-	-	Cjp33	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000824"	-	-	Cjp34	-	-	-	1	1	1	1	0	1	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000050"	-	-	Cjp34	-	-	-	0	1	0	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000827"	-	-	Cjp35	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000197"	-	-	Cjp35	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000829"	-	-	Cjp36	-	-	-	1	1	1	1	0	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000175"	-	-	Cjp36	-	-	-	0	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000830"	-	-	Cjp37	-	-	-	1	1	1	1	1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000831"	-	-	Cjp38	-	-	-	1	1	1	0	-1	1	-1	-1	-1	1	hypothetical protein	-	-
"CJ_10000832"	-	-	Cjp39	-	-	-	1	1	1	1	-1	1	-1	-1	-1	1	hypothetical protein	-	-
"CJ_10000841"	-	-	Cjp40	-	-	-	1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000843"	-	-	Cjp41	-	-	-	0	1	1	1	1	0	1	0	0	0	hypothetical protein	-	-
"CJ_10000848"	-	-	Cjp43	-	-	-	1	1	1	1	-1	-1	1	1	-1	-1	hypothetical protein	-	-
"CJ_10000851"	-	-	Cjp44	-	-	-	1	1	1	1	1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000854"	-	-	Cjp45	-	-	-	1	1	1	1	1	1	1	1	1	0	hypothetical protein	-	-
"CJ_10000856"	-	-	Cjp46	-	-	-	1	0	1	-1	-1	1	1	0	-1	-1	hypothetical protein	-	-
"CJ_10000858"	-	-	Cjp47	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"CJ_10000860"	-	-	Cjp48	-	-	-	0	1	1	1	0	1	0	-1	1	1	hypothetical protein	-	-
"CJ_10000863"	-	-	Cjp49	-	-	-	1	1	1	1	1	1	1	1	1	0	hypothetical protein	-	-
"CJ_10000870"	-	-	Cjp50	-	-	-	1	-1	1	1	1	-1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000872"	-	-	Cjp51	-	-	-	-1	1	1	1	-1	1	1	-1	1	1	hypothetical protein	-	-
"CJ_10000874"	-	-	Cjp52	-	-	-	1	1	1	1	-1	-1	1	1	1	1	hypothetical protein	-	-
"CJ_10000879"	-	-	Cjp54	-	-	-	1	1	1	1	-1	1	1	-1	1	-1	VirB7	-	-
"opCjjV010000109"	-	-	Cjr03	-	-	-	1	1	1	1	0	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000127"	-	-	Cjr06	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000130"	-	-	Cjr09	-	-	-	1	1	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000105"	-	-	Cjt02	-	-	-	1	1	1	0	0	1	1	1	1	1	hypothetical protein	-	-
"opCcV0100000317"	-	-	-	-	CCO0001	-	1	1	1	1	1	1	1	1	1	1	-	-	similar to 50S ribosomal protein L3
"opCcV0100000640"	-	-	-	-	CCO0011	-	-1	-1	1	-1	1	1	1	1	-1	-1	-	-	pseudogene
"opCcV0100001650"	-	-	-	-	CCO0025	-	1	1	1	-1	1	1	1	1	-1	1	-	-	conserved hypothetical protein
"opCcV0100001876"	-	-	-	-	CCO0026	-	1	1	1	1	1	1	1	-1	1	-1	-	-	YeeE/YedE family protein family

"opCcV010000 2071"	-	-	-	-	CCO0035	-	1	1	1	1	-1	1	1	-1	1	1	-	-	Tat (twin-arginine translocation) pathway signal
"opCcV010000 2054"	-	-	-	-	CCO0036	-	0	1	1	-1	1	1	1	1	1	1	-	-	Chain B, Stru
"opCcV010000 1228"	-	-	-	-	CCO0037	-	1	1	1	0	-1		1	-1	1	0	-	-	Chain B, Stru
"opCcV010000 1744"	-	-	-	-	CCO0038	-	1	1	1	1	-1	1	1	-1	1	-1	-	-	hypothetical protein
"opCcV010000 1285"	-	-	-	-	CCO0039	-	-1	1	1	0	1	1	-1	1	0	0	-	-	sodium:solute symporter family protein
"opCcV010000 0410"	-	-	-	-	CCO0040	-	1	1	1	1	1	-1	-1	1	1	-1	-	-	Protein of unknown function, DUF485 superfamily
"opCcV010000 0937"	-	-	-	-	CCO0048	-	1	1	1	1	1	1	1	1	1	1	-	-	hypothetical protein
"opCcV010000 0276"	-	-	-	-	CCO0067	-	1	1	0	1	-1	-1	-1	1	1	-1	-	-	conserved hypothetical protein
"opCcV010000 0178"	-	-	-	-	CCO0068	-	1	1	0	1	-1	-1	-1	1	1	-1	-	-	conserved hypothetical protein
"opCcV010000 1696"	-	-	-	-	CCO0069	-	1	1	0	1	-1	1	1	-1	1	1	-	-	conserved hypothetical protein
"opCcV010000 0483"	-	-	-	-	CCO0072	-	1	1	1	1	1	1	1	1	1	1	-	-	probable integral membrane protein Cj0033
"opCcV010000 1621"	-	-	-	-	CCO0074	-	1	1	1	1	-1	1	1	1	1	0	-	-	conserved hypothetical protein
"opCcV010000 1498"	-	-	-	-	CCO0092	-	1	1	1	1	-1	1	1	0	1	1	-	-	hypothetical protein
"opCcV010000 0901"	-	-	-	-	CCO0093	-	1	1	1	1	1	1	1	1	1	1	-	-	probable periplasmic protein Cj0057
"opCcV010000 1086"	-	-	-	-	CCO0094	-	1	1	-1	1	1	-1	-1	1	1	1	-	-	hypothetical protein
"opCcV010000 0235"	-	-	-	-	CCO0095	-	1	1	0	-1	1	1	-1	1	-1	1	-	-	probable periplasmic protein Cj0057
"opCcV010000 0045"	-	-	-	-	CCO0096	-	1	1	0	-1	1	1	-1	1	1	1	-	-	conserved hypothetical protein
"opCcV010000 0580"	-	-	-	-	CCO0097	-	1	1	1	1	-1	1	1	1	1	1	-	-	hypothetical protein
"opCcV010000 0649"	-	-	-	-	CCO0098	-	1	1	-1	1	-1	1	0	-1	1	1	-	-	hypothetical protein
"opCcV010000 0634"	-	-	-	-	CCO0099	-	1	1	1	1	1	0	1	0	1	-1	-	-	probable periplasmic protein Cj0057
"opCcV010000 0623"	-	-	-	-	CCO0100	-	-1	-1	1	-1	1	-1	-1	1	-1	-1	-	-	conserved hypothetical protein
"opCcV010000 0499"	-	-	-	-	CCO0101	-	1	1	1	-1	-1	1	-1	1	0	1	-	-	conserved hypothetical protein
"opCcV010000 1681"	-	-	-	-	CCO0102	-	1	1	0	1	-1	0	1	-1	1	1	-	-	hypothetical protein
"opCcV010000 0103"	-	-	-	-	CCO0104	-	1	1	0	-1	1	0	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV010000 1477"	-	-	-	-	CCO0105	-	1	1	0	-1	-1	1	1	-1	1	-1	-	-	hypothetical protein
"opCcV010000 1918"	-	-	-	-	CCO0106	-	0	1	1	1	-1	1	1	-1	1	1	-	-	hypothetical protein
"opCcV010000"	-	-	-	-	CCO0107	-	1	1	1	-1	1	-1	1	1	1	0	-	-	conserved hypothetical

1220"																				protein
"opCcV0100000418"	-	-	-	-	CCO0108	-	1	1	1	1	-1	1	1	1	1	-1	-	-	-	conserved hypothetical protein
"opCcV0100000721"	-	-	-	-	CCO0109	-	1	1	0	1	-1	1	1	-1	1	1	-	-	-	hypothetical protein
"opCcV0100001335"	-	-	-	-	CCO0110	-	1	1	0		-1	1	1	1	1	1	-	-	-	probable periplasmic protein Cj0057
"opCcV0100000581"	-	-	-	-	CCO0111	-	1	1	1	1	-1	1	1	0	1	-1	-	-	-	conserved hypothetical protein
"opCcV0100001091"	-	-	-	-	CCO0112	-	1	1	1	1	1	1	0	1	1	1	-	-	-	hypothetical protein
"opCcV0100000775"	-	-	-	-	CCO0127	-	1	1	0	1	1	1	0	1	1	-1	-	-	-	methyl-accepting chemotaxis protein, putative
"opCcV0100000715"	-	-	-	-	CCO0128	-	1	1	0	1	-1	1	-1	0	1	1	-	-	-	conserved hypothetical protein
"opCcV0100001622"	-	-	-	-	CCO0136	-	-1	-1	-1	0	1	1	-1	1	-1	0	-	-	-	hypothetical protein
"opCcV0100001293"	-	-	-	-	CCO0137	-	1	-1	0	1	1	1	-1	1	-1	1	-	-	-	transcriptional regulator, Crp family, putative
"opCcV0100000793"	-	-	-	-	CCO0138	-	1	1	1	1	1	1	1	1	-1	-1	-	-	-	conserved hypothetical protein
"opCcV0100000316"	-	-	-	-	CCO0181	-	1	1	0	-1	-1	1	0	1	1	1	-	-	-	filamentous hemagglutinin, intein-containing,
"opCcV0100001392"	-	-	-	-	CCO0182	-		-1	0			1		-1			-	-	-	hemagglutinin/hemolysin-related protein
"opCcV0100001501"	-	-	-	-	CCO0183	-	0	0	0	1	1	-1	0	1	1	1	-	-	-	Hemolysin, putative
"opCcV0100001076"	-	-	-	-	CCO0184	-	1	1	1	1	-1	1	1	1	1	-1	-	-	-	Hemolysin, putative
"opCcV0100001872"	-	-	-	-	CCO0185	-		1	1	1	-1	1	1	1	0	1	-	-	-	hypothetical protein
"opCcV0100001181"	-	-	-	-	CCO0186	-	1	1	1	1	-1	1	-1	1	1	1	-	-	-	hypothetical protein
"opCcV0100000939"	-	-	-	-	CCO0187	-	1	1	1	1	-1	1	1	0	1	1	-	-	-	filamentous hemagglutinin 1, putative
"opCcV0100002052"	-	-	-	-	CCO0188	-	0	1	1	-1	-1	1	1	0	1	1	-	-	-	hypothetical protein
"opCcV0100000029"	-	-	-	-	CCO0189	-			-1			-1	-1	1	1	-1	-	-	-	hypothetical protein
"opCcV0100001039"	-	-	-	-	CCO0190	-	1	1	1	1	0	0	0	-1	1	1	-	-	-	hemolysin activation protein HecB, putative
"opCcV0100001254"	-	-	-	-	CCO0203	-	1	1	1		1	0	1	1	1	-1	-	-	-	hypothetical protein
"opCcV0100000013"	-	-	-	-	CCO0204	-	1	1	1	0	-1	1	-1	1	0	-1	-	-	-	hypothetical protein
"opCcV0100000114"	-	-	-	-	CCO0210	-	1	-1	1	0	1	1	1	1	1	1	-	-	-	conserved hypothetical protein
"opCcV0100001920"	-	-	-	-	CCO0212	-	1	1	1	1	-1	-1	1	-1	-1	-1	-	-	-	conserved hypothetical protein
"opCcV0100001217"	-	-	-	-	CCO0213	-	1	0	1	1	1	1	1	1	1	1	-	-	-	methyl-accepting chemotaxis protein, putative
"opCcV0100000117"	-	-	-	-	CCO0231	-	1	1	1	1	-1	1	-1	-1	1	-1	-	-	-	hypothetical protein
"opCcV0100000991"	-	-	-	-	CCO0246	-	1	1	1	1	-1	-1	1	1	1	-1	-	-	-	hypothetical protein

"opCcV0100000193"	-	-	-	-	CCO0248	-	1	1	1	0	-1	0	0	-1	1	-1	-	-	conserved hypothetical protein
"opCcV0100000024"	-	-	-	-	CCO0249	-	1	1	1	0	1	-1	1	-1	0	-1	-	-	conserved hypothetical protein
"opCcV0100000266"	-	-	-	-	CCO0250	-	1	1	1	1	-1	-1	1	1	0	1	-	-	hypothetical protein
"opCcV0100000545"	-	-	-	-	CCO0251	-	1	1	1	1	0	1	1	-1	1	1	-	-	lipoprotein, putative
"opCcV0100000645"	-	-	-	-	CCO0252	-	-1	0	1	1	1	1	1	1	1	1	-	-	probable integral membrane protein Cj0564
"opCcV0100001358"	-	-	-	-	CCO0253	-	1	1	1	1	1	0	1	1	1	1	-	-	hypothetical protein
"opCcV0100000876"	-	-	-	-	CCO0254	-	1	1	1	1	-1	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100000630"	-	-	-	-	CCO0257	-	1	1	1	1	1	1	1	1	1	1	-	-	glcG protein
"opCcV0100000875"	-	-	-	-	CCO0280	-	0	0	1	1	1	1	0	1	1	1	-	-	methyl-accepting chemotaxis protein (tlpA)
"opCcV0100000323"	-	-	-	-	CCO0281	-	1	1	1	-1	-1	0	-1	1	-1	1	-	-	hypothetical protein
"opCcV0100000310"	-	-	-	-	CCO0284	-	1	1	1	1	-1	-1	1	0	1	1	-	-	hypothetical protein
"opCcV0100000982"	-	-	-	-	CCO0285	-	1	1	1	1		1	1			1	-	-	CAAX amino terminal protease family protein
"opCcV0100001967"	-	-	-	-	CCO0291	-	1	1	1	-1	0	1	1	1	-1	-1	-	-	Hypothetical cytosolic protein, putative
"opCcV0100000695"	-	-	-	-	CCO0292	-	1	-1	1	1	-1	1	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100000694"	-	-	-	-	CCO0293	-	1	1	1	-1	-1	-1	0	1	-1	-1	-	-	penicillin-binding protein, putative
"opCcV0100001345"	-	-	-	-	CCO0332	-	1	1	1	1	0	1	0	1	1	1	-	-	TM2 domain protein, putative
"opCcV0100000810"	-	-	-	-	CCO0347	-	1	1	0	-1	-1	1	-1	-1	1	1	-	-	hypothetical protein
"opCcV0100001609"	-	-	-	-	CCO0349	-	1	1	-1	-1	1	1	1	0	1	1	-	-	carboxyphosphoenolpyruvate phosphonmutase
"opCcV0100000265"	-	-	-	-	CCO0350	citZ	1	1	1	-1	-1	1	0	-1	1	0	-	-	2-methylcitrate synthase
"opCcV0100000345"	-	-	-	-	CCO0351	b0334	1	1	0	-1	1	1	0	1	-1	-1	-	-	pseudogene
"opCcV0100001597"	-	-	-	-	CCO0352	-	1	1	1	-1	0	0	1	0	1	1	-	-	ID867
"opCcV0100001722"	-	-	-	-	CCO0353	-	1	1	1	1	-1	1	1	-1	-1	1	-	-	membrane protein , putative
"opCcV0100001475"	-	-	-	-	CCO0354	-	1	1	1	1	1	1	1	1	-1	-1	-	-	hypothetical protein
"opCcV0100001552"	-	-	-	-	CCO0355	-	1	-1	0	-1	1	1	1	-1	1	1	-	-	probable periplasmic protein Cj0413 , putative
"opCcV0100001845"	-	-	-	-	CCO0356	-	1	1	-1	-1	1		1	1	1	-1	-	-	hypothetical protein
"opCcV0100001068"	-	-	-	-	CCO0368	cdtC	1	-1	0	-1	1	1	-1	1	1	1	-	-	cytolethal distending toxin C
"opCcV0100001565"	-	-	-	-	CCO0382	-	1	1	0	-1	-1	1	0	-1	1	-1	-	-	lipoprotein, putative
"opCcV0100000000"	-	-	-	-	CCO0383	-	1	1	1	0	-1	-1	-1	1	1	-1	-	-	conserved hypothetical

"opCcV0100001532"	-	-	-	-	CCO0815	-	1	1	0	-1	0	-1	0	1	1	1	-	-	hypothetical protein
"opCcV0100000380"	-	-	-	-	CCO0845	-	1	1	1	1	0	1	1	-1	1	1	-	-	azlC protein, putative
"opCcV0100000914"	-	-	-	-	CCO0846	-	1	0	0	1	-1	1	1	1	-1	1	-	-	Branched-chain amino acid transport protein,
"opCcV0100001559"	-	-	-	-	CCO0847	-	0	-1	1	1	1	1	1	1	1	1	-	-	transporter, LysE family
"opCcV0100001442"	-	-	-	-	CCO0865	dapA	1	-1	1	1	1	1	0	1	-1	1	-	-	dihydrodipicolinate synthase
"opCcV0100002024"	-	-	-	-	CCO0866	-	1	1	1	-1	-1	1	1	0	1	1	-	-	probable oxidoreductase Cj0807
"opCcV0100000315"	-	-	-	-	CCO0867	-	0	-1	1	1	1	1	1	1	-1	1	-	-	small hydrophobic protein Cj0808c
"opCcV0100000086"	-	-	-	-	CCO0868	-	1	1	1	-1	-1	-1	1	1	-1	1	-	-	probable hydrolase Cj0809c
"opCcV0100001659"	-	-	-	-	CCO0869	nadE	1	1	1	1	1	1	1	1	1	1	-	-	NAD ⁺ synthetase
"opCcV0100001955"	-	-	-	-	CCO0870	lpxK	1	1	1	1	-1	1	1	-1	1	1	-	-	tetraacyldisaccharide-1-P 4'-kinase
"opCcV0100001395"	-	-	-	-	CCO0917	-	1	1	-1	1	1	1	1	-1	1	1	-	-	ferric uptake regulation protein, putative
"opCcV0100001593"	-	-	-	-	CCO0918	-	1	1	-1	1	1	-1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100001708"	-	-	-	-	CCO0919	fimA	1	1	1	0	-1	-1	0	-1	0	1	-	-	periplasmic solute binding protein for ABC
"opCcV0100000916"	-	-	-	-	CCO0921	-	1	1	1	1	-1	1	1	1	1	1	-	-	membrane protein, putative
"opCcV0100001855"	-	-	-	-	CCO0922	-	1	1		1	1	1	1	1	0	1	-	-	hypothetical protein
"opCcV0100001245"	-	-	-	-	CCO0923	-	0	-1	0	1	1	0	1	1	0	1	-	-	hypothetical protein
"opCcV0100000808"	-	-	-	-	CCO0924	-	1	1	1	-1	1	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100000122"	-	-	-	-	CCO0925	-			-1	-1	-1		-1	-1	-1	1	-1	-	hypothetical protein
"opCcV0100001262"	-	-	-	-	CCO0926	-	1	1	1	0	1	-1	-1	1	1	0	-	-	hypothetical protein
"opCcV0100000529"	-	-	-	-	CCO0927	-			1	-1	-1	1	-1	-1	1	1	1	-	hypothetical protein
"opCcV0100001870"	-	-	-	-	CCO0928	-	-1	-1	1	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein
"opCcV0100001938"	-	-	-	-	CCO0929	-	1	1	1	-1	-1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000722"	-	-	-	-	CCO0955	-	1	1	1	1	-1	1	1	1	1	-1	-	-	inner membrane protein, putative
"opCcV0100001617"	-	-	-	-	CCO0956	-			1	1	1	0	0	0	1	-1	0	-	conserved hypothetical protein
"opCcV0100001038"	-	-	-	-	CCO0968	-	1	1	1	1	1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100001944"	-	-	-	-	CCO0970	-	1	1	1	1	-1	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100000061"	-	-	-	-	CCO1005	-	1	1	1	1	-1	0	1	1	1	-1	-	-	sodium/alanine symporter VC2356
"opCcV010000"	-	-	-	-	CCO1040	-	1	1	0	-1	-1	1	0	-1	1	0	-	-	conserved hypothetical

Appendix

1105"																				protein
"opCcV0100000131"	-	-	-	-	CCO1041	-	1	1	0	-1	1	-1	1	1	1	1	-	-	-	conserved hypothetical protein
"opCcV0100001819"	-	-	-	-	CCO1049	-	1	-1	0	-1	1	0	-1	1	0	1	-	-	-	surface-exposed lipoprotein
"opCcV0100001438"	-	-	-	-	CCO1075	-	1	1	1	-1	-1	1	0	1		-1	-	-	-	conserved hypothetical protein
"opCcV0100001590"	-	-	-	-	CCO1076	-	1	1	1	0	-1	1	1	-1	1	1	-	-	-	hypothetical protein
"opCcV0100001714"	-	-	-	-	CCO1077	-	1	1	-1	1	0	1	-1	1	1	1	-	-	-	hypothetical protein
"opCcV0100000376"	-	-	-	-	CCO1078	-	1	1	1	1	-1	1	1	1	1	0	-	-	-	hypothetical protein
"opCcV0100000365"	-	-	-	-	CCO1115	-	1	1	0	0	-1	1	-1	-1	1	1	-	-	-	adenine specific DNA methyltransferase
"opCcV0100001628"	-	-	-	-	CCO1123	-	1	0	1	1	1	1	-1	1	1	1	-	-	-	hypothetical protein
"opCcV0100000022"	-	-	-	-	CCO1124	-	1	1	1	1	-1	1	0	1	1	1	-	-	-	hypothetical protein
"opCcV0100001229"	-	-	-	-	CCO1125	-	1	-1	1	1	0	1	1	1	0	1	-	-	-	VgrG protein, putative
"opCcV0100000141"	-	-	-	-	CCO1126	-	1	-1	1	1	1	1	1	1	1	1	-	-	-	pseudogene
"opCcV0100000525"	-	-	-	-	CCO1142	-	1	1	-1	1	1	1	1	1	0	1	-	-	-	conserved hypothetical protein
"opCcV0100001697"	-	-	-	-	CCO1143	-	1	1	0		1	-1	1	1	1	1	-	-	-	transporter, MFS superfamily
"opCcV0100001165"	-	-	-	-	CCO1170	-	1	1	0	0	1	1	1	1	1	1	-	-	-	oxidoreductase, short-chain
"opCcV0100002057"	-	-	-	-	CCO1171	-	1	1	1	1	-1	1	1	-1	1	1	-	-	-	hypothetical protein
"opCcV0100000357"	-	-	-	-	CCO1172	-	1	1	1	1	-1	-1	1	1	1	1	-	-	-	conserved hypothetical protein
"opCcV0100000331"	-	-	-	-	CCO1173	-	1	1	1	1	-1	0	1	-1	1	-1	-	-	-	hypothetical protein
"opCcV0100000771"	-	-	-	-	CCO1211	-	1	1	1	1	-1	-1	0	-1	1	-1	-	-	-	glycosyl transferase, group 1 family protein
"opCcV0100001761"	-	-	-	-	CCO1212	-	1	0	1	-1	-1	1	-1	1	1	-1	-	-	-	general stress protein A, putative
"opCcV0100001473"	-	-	-	-	CCO1214	-	1	1	-1	0	-1	1	-1	1	1	-1	-	-	-	conserved hypothetical protein
"opCcV0100000308"	-	-	-	-	CCO1215	-	1	1	1	-1	1	-1	1	1	-1	-1	-	-	-	bifunctional alpha-2,3/-2,8-sialyltransferase
"opCcV0100000837"	-	-	-	-	CCO1216	-	-1	1	1	0	1	1	-1	1	0	0	-	-	-	hypothetical protein
"opCcV0100000084"	-	-	-	-	CCO1217	-	1	1	0	0	-1	1	-1	1	1	0	-	-	-	sialyl transferase
"opCcV0100001797"	-	-	-	-	CCO1218	-	1	1	1	0	0	1	1	0	1	1	-	-	-	conserved hypothetical protein
"opCcV0100001035"	-	-	-	-	CCO1221	-	-1				1	1	1				-	-	-	lipooligosaccharide 5G8 epitope
"opCcV0100000093"	-	-	-	-	CCO1277	-	1	1	1	1	1	1	1	1	1	-1	-	-	-	hypothetical protein
"opCcV0100000726"	-	-	-	-	CCO1278	-	1	1	1	1	-1	-1	1	-1	-1	1	-	-	-	hydrolase, carbon-nitrogen family

"opCcV0100000994"	-	-	-	-	CCO1279	-	1	1	1	1	-1	1	0	-1	1	1	-	-	polysaccharide deacetylase family protein
"opCcV0100001233"	-	-	-	-	CCO1280	-	0	0	1	1	1	1	0	1	1	-	-	-	conserved hypothetical protein
"opCcV0100001155"	-	-	-	-	CCO1281	-	1	1	0	1	0	1	1	-1	1	1	-	-	cobalamin synthesis protein/P47K family protein
"opCcV0100000977"	-	-	-	-	CCO1298	-	1	1	0	-1	1	0	0	1	-1	1	-	-	sodium/pantothenate symporter , putative
"opCcV0100000688"	-	-	-	-	CCO1299	-	1	1	0	1	1	-1	-1	1	-1	1	-	-	hypothetical protein
"opCcV0100000196"	-	-	-	-	CCO1303	-	1	1	-1	-1		1	1	1	1	-1	-	-	Ribbon-helix-helix protein, copG family domain
"opCcV0100000897"	-	-	-	-	CCO1304	-	1	1	1	1	1	-1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100002006"	-	-	-	-	CCO1305	-	1	1	1	-1	0	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100000805"	-	-	-	-	CCO1306	-	1	1	1	1	1	1	1	1	-1	1	-	-	cytochrome c family protein
"opCcV0100001726"	-	-	-	-	CCO1307	-	1	1	1	1	0	1	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001573"	-	-	-	-	CCO1308	-	1	1	1	-1	1	1	-1	1	1	1	-	-	putative periplasmic protein
"opCcV0100001188"	-	-	-	-	CCO1309	-	1	1	1	1	-1	1	-1	1	1	-1	-	-	putative periplasmic protein
"opCcV0100000386"	-	-	-	-	CCO1310	-	1	1	1	0	-1	1	1	-1	1	1	-	-	putative periplasmic protein
"opCcV0100000226"	-	-	-	-	CCO1311	-	1	-1	1	1	1	1	1	1	-1	1	-	-	hypothetical protein
"opCcV0100001368"	-	-	-	-	CCO1312	-	1	-	0		1	1	-1	1		-1	-	-	filamentous haemagglutinin domain protein
"opCcV0100000823"	-	-	-	-	CCO1325	-	1	1	1	-1	-1	-1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000145"	-	-	-	-	CCO1326	-	1	1	1	-1	1	1	-1	1	-1	-1	-	-	hypothetical protein
"opCcV0100000048"	-	-	-	-	CCO1331	-	1	1	1	0	-1	1	1	-1	1	-1	-	-	hypothetical protein
"opCcV0100001666"	-	-	-	-	CCO1339	-	1	1	1	0	-1	1	-1	1	1	1	-	-	ISCCo1, transposase
"opCcV0100000302"	-	-	-	-	CCO1340	-	1	1	1	1	-1	1	1	1	1	-1	-	-	methyltransferase Atu0936 , putative
"opCcV0100001923"	-	-	-	-	CCO1341	-	1	-1	1	0	-1	1	1	1	1	1	-	-	phospholipid N-methyltransferase, putative
"opCcV0100000839"	-	-	-	-	CCO1342	-	1	1	1	1	-1	-1	0	-1	1	-1	-	-	hypothetical protein
"opCcV0100000125"	-	-	-	-	CCO1349	-	1	1	-1	-1	1	1	1	1	1	-1	-	-	membrane protein, putative
"opCcV0100000978"	-	-	-	-	CCO1369	-	1	1	1	1	-1	1	1	-1	1	1	-	-	conserved domain protein
"opCcV0100000088"	-	-	-	-	CCO1370	-	1	1	1	1		1	0	1	1	-1	-	-	ABC transporter ATP-binding protein
"opCcV0100002007"	-	-	-	-	CCO1409	-	1	1	1	1	-1	1	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001457"	-	-	-	-	CCO1412	-		-1	-1	-1	0	-1	-1	1	0	-1	-	-	acetyltransferase, GNAT family family
"opCcV010000"	-	-	-	-	CCO1413	-	0	-1	0	1	1	0	1	1	-1	0	-	-	formyltransferase, putative

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"opCcV0100000382"	-	-	-	-	CCO1535	-	1	1	1	1	1	0	0	1	-1	1	-	-	glycosyl transferase, group 2 family protein
"opCcV0100001735"	-	-	-	-	CCO1536	-	1	1	1	1	-1	1	0	1	1	-1	-	-	hypothetical protein
"opCcV0100000112"	-	-	-	-	CCO1537	-	1	1	0	-1	1	1	-1	1	0	-1	-	-	DcbE, putative
"opCcV0100000971"	-	-	-	-	CCO1538	-	1	1	1	1	-1	1	1	1	1	-1	-	-	alpha-2,3-sialyltransferase
"opCcV0100000350"	-	-	-	-	CCO1539	cysC	1	-1	0	1	1	1	0	1	-1	0	-	-	adenylsulfate kinase
"opCcV0100001905"	-	-	-	-	CCO1540	-	1	1	1	1	-1	0	1	-1	1	1	-	-	transporter, sodium/sulfate symporter family,
"opCcV0100000496"	-	-	-	-	CCO1541	-	1	1	-1	-1	1	0	0	1	1	0	-	-	sulfate adenyltransferase, subunit
"opCcV0100001898"	-	-	-	-	CCO1542	-	1	0	1	1	1	1	1	1	-1	0	-	-	sulfate adenyltransferase, subunit 2
"opCcV0100000677"	-	-	-	-	CCO1543	cysQ	1	1	1	1	-1	1	-1	-1	1	-1	-	-	3'(2'),5'-bisphosphate nucleotidase
"opCcV0100001820"	-	-	-	-	CCO1544	-	1	1	0	1	-1	0	1	-1	1	-1	-	-	alpha-2,3-sialyltransferase
"opCcV0100000849"	-	-	-	-	CCO1546	-	1	1	1	-1	-1	1	-1	0	1	-1	-	-	Glycosyl transferase family 8 family
"opCcV0100000616"	-	-	-	-	CCO1547	ggaB	1	1	1	0	-1	0	-1	1	1	-1	-	-	capsular polysaccharide synthesis-C
"opCcV0100000636"	-	-	-	-	CCO1548	ggaB	1	1	1	1	-1	1	1	-1	1	-1	-	-	minor teichoic acids biosynthesis protein ggaB
"opCcV0100002058"	-	-	-	-	CCO1549	-	-1	1	1	1	-1	-1	1	0	1	1	-	-	capsule biosynthesis protein, putative
"opCcV0100001140"	-	-	-	-	CCO1587	-	1	1	1	1	0	1	1	1	1	1	-	-	DnaJ-related protein
"opCcV0100000059"	-	-	-	-	CCO1588	-	1	-1	0	-1	1	-1	1	1	-1	0	-	-	conserved hypothetical protein
"opCcV0100002045"	-	-	-	-	CCO1610	-	0	1	1	1	-1	0	-1	1	1	1	-	-	hypothetical protein
"opCcV0100000233"	-	-	-	-	CCO1627	-	1	1	1	1	-1	1	1	-1	1	-1	-	-	AgrC, putative
"opCcV0100000893"	-	-	-	-	CCO1628	-	1	1	0	-1	-1	1	1	-1	1	0	-	-	hypothetical protein
"opCcV0100001511"	-	-	-	-	CCO1629	-	1	-1	1		0	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100000974"	-	-	-	-	CCO1631	-	1	1	1	1	1	1	-1	1	1	-1	-	-	hypothetical protein
"opCcV0100000632"	-	-	-	-	CCO1637	-	1	1	0	1	-1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000523"	-	-	-	-	CCO1638	-	1	0	1	1	-1	-1	0	1	1	0	-	-	ISccol, transposase orfB
"opCcV0100001018"	-	-	-	-	CCO1639	-	1	1	0	-1	1	1	0	1	1	-1	-	-	ISccol, transposase orfA
"opCcV0100000802"	-	-	-	-	CCO1651	-	1	1	1	-1	0	-1	1	1	1	1	-	-	pseudogene
"opCcV0100000373"	-	-	-	-	CCO1654	-	1	1	1	-1	-1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100001856"	-	-	-	-	CCO1656	mdaB	1	-1	1	0	1	1	1	1	-1	0	-	-	modulator of drug activity (mda66)
"opCcV0100000000"	-	-	-	-	CCO1657	mdaB	0	1	-1	0	1	1	1	1	1	-1	-	-	modulator of drug activity

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1037"																			(mda66)
"opCcV0100001139"	-	-	-	-	CCO1658	-	1	1	1	-1	1	-1	-1	1	1	0	-	-	major facilitator family transporter, putative
"opCcV0100001384"	-	-	-	-	CCO1661	-	1	0	0	1	1	1	1	1	1	1	-	-	HsdR
"opCcV0100000533"	-	-	-	-	CCO1662	-	1	1	0	1	-1	0	1	-1	1	1	-	-	conserved hypothetical protein
"opCcV0100001548"	-	-	-	-	CCO1663	-	1	1	1	1		1		1	1	0	-	-	Helicase conserved C-terminal domain protein
"opCcV0100000565"	-	-	-	-	CCO1664	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	HsdS
"opCcV0100001895"	-	-	-	-	CCO1665	-	1	-1	1	-1	1	-1	-1	1	-1	-1	-	-	MloA
"opCcV0100000986"	-	-	-	-	CCO1666	-	1	-	0	-1	1	0	1	1	1	-1	-	-	HsdM
"opCcV0100001132"	-	-	-	-	CCO1667	-	1	1	1	1	-1	1	1	-1	1	1	-	-	conserved hypothetical protein
"opCcV0100000572"	-	-	-	-	CCO1668	-	1	1	1	1	-1	1	1	1	1	1	-	-	sarcosine oxidase, putative
"opCcV0100001564"	-	-	-	-	CCO1669	-	1	-1	1	1	1	1	1	1	1	1	-	-	NADP(H) oxidoreductase CC0205
"opCcV0100001197"	-	-	-	-	CCO1670	-	1	1	1	-1	1	1	1	1	0	1	-	-	hypothetical protein
"opCcV0100001574"	-	-	-	-	CCO1671	-	1	1	1	-1	1	1	1	1	1	1	-	-	catalase-like protein
"opCcV0100000460"	-	-	-	-	CCO1674	arsC	1	-1	-1	-1	0	1	1	1	-1	-1	-	-	arsC
"opCcV0100002074"	-	-	-	-	CCO1675	arsC	0	1	1	1	-1	1	1	-1	1	1	-	-	arsenite efflux transporter
"opCcV0100001754"	-	-	-	-	CCO1676	arsC	1	-1	-1	-1	1	1	1	1	-1	-1	-	-	arsenite efflux transporter
"opCcV0100000794"	-	-	-	-	CCO1688	-	1	1	1	1	-1	1	-1	1	-1	-1	-	-	hypothetical protein
"opCcV0100001259"	-	-	-	-	CCO1689	-	1	1	1	-1	-1	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100000936"	-	-	-	-	CCO1694	-	1	1	1	-1	-1	0	-1	1	1	-1	-	-	major facilitator family transporter, putative
"opCcV0100001518"	-	-	-	-	CCO1707	-	1	1	1	1	-1	1	1	-1	1	1	-	-	integral membrane protein, putative
"opCcV0100002061"	-	-	-	-	CCO1708	-	1	1	1	1	-1	1	1	-1	1	1	-	-	integral membrane protein, putative
"opCcV0100002049"	-	-	-	-	CCO1710	-	1	1	1	1	-1	-1	1	1	1	1	-	-	hypothetical protein
"opCcV0100002027"	-	-	-	-	CCO1734	-	0	1	1	1	-1	1	-1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001016"	-	-	-	-	CCO1738	-	1	1	1	1	-1	-1	-1	1	1	1	-	-	Protein of unknown function DUF262 family
"opCcV0100001826"	-	-	-	-	CCO1739	-	1	1	1	0	-1	1	-1	-1	1	1	-	-	hypothetical protein
"opCcV0100001409"	-	-	-	-	CCO1740	-	1	1	0	1	1	1	1	1	-1	-1	-	-	hypothetical protein
"opCcV0100000462"	-	-	-	-	CCO1758	-	1	-1	0	0	1	1	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001365"	-	-	-	-	CCO1784	-	1	1	1	1	-1	1	1		1	1	-	-	hypothetical protein

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"opCcV0100001864"	-	-	-	-	CCO1785	-	1	1	1	1	0	1	1	-1	1	-1	-	-	hypothetical protein
"opCcV0100002070"	-	-	-	-	CCO1786	-	1	1	1	1	1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000420"	-	-	-	-	CCO1801	-	1	1	1	0	-1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100000124"	-	-	-	-	CCOA_CcrnpB2	-		-1	0	-1	1	-1	-1	1	1	1	-	-	sRNA
"opCcV0100001087"	-	-	-	-	CCOA_CcrnpB3	-	-1	-1	0	-1	1	-1	-1	1	-1	0	-	-	sRNA
"opCcV0100001665"	-	-	-	-	CCOA_CctmRNA2	-	1	1	0	1	1	1	1	0	1	1	-	-	sRNA
"opCcV0100001861"	-	-	-	-	CCOA0001	-	1	1	0	-1	1	1	0	1	-1	1	-	-	replication protein
"opCcV0100000083"	-	-	-	-	CCOA0002	-	1	1	0	-1	-1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100000854"	-	-	-	-	CCOA0003	-	1	1	1	0	-1	1	-1	-1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001773"	-	-	-	-	CCOA0004	-	1	1	1	-1	-1	-1	-1	1	0	1	-	-	hypothetical protein
"opCcV0100001736"	-	-	-	-	CCOA0005	-	-1	1	1	1	1	1	1	1	0	1	-	-	hypothetical protein
"opCcV0100000493"	-	-	-	-	CCOA0006	-	1	1	1	1	-1	-1	-1	1	1		-	-	conserved hypothetical protein
"opCcV0100000042"	-	-	-	-	CCOA0007	-			0	-1	-1	-1	-1	1	1	-1	-	-	hypothetical protein
"opCcV0100000980"	-	-	-	-	CCOA0010	-	1	1	1	1	-1	1	-1	1	1	-1	-	-	helicase, Snf2 family
"opCcV0100000134"	-	-	-	-	CCOA0011	-	1	1	1	-1	1	1	-1	1	1	-1	-	-	hypothetical protein
"opCcV0100000200"	-	-	-	-	CCOA0012	-	1	1	1	0	-1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000902"	-	-	-	-	CCOA0013	-	1	1	1	-1	1	1	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001913"	-	-	-	-	CCOA0016	-	1	1	1	1	-1	1	-1	-1	-1	1	-	-	TraH protein
"opCcV0100001144"	-	-	-	-	CCOA0017	-	1	1	1	1	-1	1	-1	1	-1	-1	-	-	hypothetical protein
"opCcV0100000842"	-	-	-	-	CCOA0018	-	1	1	1	0	-1	1	0	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100002059"	-	-	-	-	CCOA0019	-	1	1	1	1	-1	0	1	-1	1	1	-	-	conserved hypothetical protein
"opCcV0100001136"	-	-	-	-	CCOA0020	-	1	1	1	1	1	-1	1	1	1	-1	-	-	882aa long hypothetical purine NTPase, putative
"opCcV0100001603"	-	-	-	-	CCOA0021	-		-1	1	-1	1	-1	-1	1	-1	-1	-	-	DNA primase TraC
"opCcV0100000729"	-	-	-	-	CCOA0022	-		-1	0	-1	1	-1	-1	1	-1	-1	-	-	DNA primase, putative
"opCcV0100001108"	-	-	-	-	CCOA0023	-	1	1	1	1	1	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100001673"	-	-	-	-	CCOA0024	-		-1	0		-1	1	1	-1	1	-1	-	-	hypothetical protein
"opCcV0100000619"	-	-	-	-	CCOA0025	-			1	-1	-1	-1	1	-1	-1	-1	-	-	hypothetical protein
"opCcV010000"	-	-	-	-	CCOA0026	-	1	1	1	1	-1	1	1	-1	-1	-1	-	-	conserved hypothetical

1721"																				protein
"opCcV010000 1306"	-	-	-	-	CCOA0027	-	1	1	1	1	-1	1	1	1	0	1	-	-	-	hypothetical protein
"opCcV010000 1844"	-	-	-	-	CCOA0028	-	-1	-1	1	-1	-1	-1	1	1	1	-1	-	-	-	hypothetical protein
"opCcV010000 0173"	-	-	-	-	CCOA0029	-			0	1	1	1	1	1	1	1	-	-	-	hypothetical protein
"opCcV010000 1116"	-	-	-	-	CCOA0030	-			-1	-1	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein
"opCcV010000 2023"	-	-	-	-	CCOA0031	-	1	1	1	1	-1	1	1	1	1	1	-	-	-	hypothetical protein
"opCcV010000 1560"	-	-	-	-	CCOA0032	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	-	hypothetical protein
"opCcV010000 2082"	-	-	-	-	CCOA0033	-	1	1	1	1	1	1	-1	-1	1	-1	-	-	-	hypothetical protein
"opCcV010000 0028"	-	-	-	-	CCOA0034	-			0					-1	1	-1	-	-	-	hypothetical protein
"opCcV010000 1968"	-	-	-	-	CCOA0035	-	0	1	1	-1	-1	1	1	1	1	1	-	-	-	hypothetical protein
"opCcV010000 1381"	-	-	-	-	CCOA0036	-	1	1	1	1	1	1	1	1	1	1	-	-	-	hypothetical protein
"opCcV010000 0052"	-	-	-	-	CCOA0039	-		1	1	1	1	-1		1	1	-1	-	-	-	hypothetical protein
"opCcV010000 0466"	-	-	-	-	CCOA0046	-	-1	-1	-1	-1	1	-1	-1	1	-1	-1	-	-	-	hypothetical protein
"opCcV010000 0891"	-	-	-	-	CCOA0048	-	1	1	0	1	1	-1	1	1	-1	1	-	-	-	hypothetical protein
"opCcV010000 0012"	-	-	-	-	CCOA0049	-	1	1	1	-1	0	1	1	1	-1	-1	-	-	-	hypothetical protein
"opCcV010000 0224"	-	-	-	-	CCOA0050	-	1	-1	1		-1	0	1	1	1	-1	-	-	-	hypothetical protein
"opCcV010000 0332"	-	-	-	-	CCOA0051	-	1	1	1	1	-1	1	1	1	0	0	-	-	-	conserved hypothetical protein
"opCcV010000 1725"	-	-	-	-	CCOA0052	-	1	1	1	-1	1	0	-1	1	-1	1	-	-	-	hypothetical protein
"opCcV010000 1608"	-	-	-	-	CCOA0053	-	1	1	1	1	-1	1	1	1	1	-1	-	-	-	conserved hypothetical protein
"opCcV010000 0456"	-	-	-	-	CCOA0054	-	1	1	1	1	-1	1	1	1	1	1	-	-	-	hypothetical protein
"opCcV010000 0146"	-	-	-	-	CCOA0055	-	1	1	0	0		1	1	-1	1	-1	-	-	-	hypothetical protein
"opCcV010000 2055"	-	-	-	-	CCOA0057	-	0	1	1	1	-1	1	1	-1	1	1	-	-	-	DNA repair protein RAD50, putative
"opCcV010000 1777"	-	-	-	-	CCOA0058	-	1	1	1	1	-1	1	-1	1	1	-1	-	-	-	hypothetical protein
"opCcV010000 0712"	-	-	-	-	CCOA0059	-	1	1	1	-1	1	1	1	1	1	1	-	-	-	hypothetical protein
"opCcV010000 2018"	-	-	-	-	CCOA0060	-	0	1	1	-1	-1	1	1	1	0	-1	-	-	-	hypothetical protein
"opCcV010000 0880"	-	-	-	-	CCOA0061	-	1	1	1	-1	-1	-1	1	0	1	-1	-	-	-	hypothetical protein
"opCcV010000 1073"	-	-	-	-	CCOA0062	-	1	1	1	1	1	1	1	0	1	1	-	-	-	conserved hypothetical protein
"opCcV010000 0740"	-	-	-	-	CCOA0063	-	1	-1	0	-1	1	1	-1	1	-1	-1	-	-	-	TnpY

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"opCcV0100001142"	-	-	-	-	CCOA0064	-	1	1	1	1	-1	-1	0	-1	0	-1	-	-	hypothetical protein
"opCcV0100001700"	-	-	-	-	CCOA0065	-	1	-1	1	1	1	1	1	1	0	0	-	-	mobilization/transfer protein
"opCcV0100001662"	-	-	-	-	CCOA0066	-	1	0	1	-1	1	1	1	1	-1	1	-	-	conserved hypothetical protein
"opCcV0100000366"	-	-	-	-	CCOA0067	-	1	1	1	1	0	-1	1	1	1	1	-	-	aminoglycoside 3'-phosphotransferase
"opCcV0100001497"	-	-	-	-	CCOA0068	-	1	1	0	-1	1	-1	0	1	0	1	-	-	aminoglycoside 3'-phosphotransferase,
"opCcV0100000812"	-	-	-	-	CCOA0069	-	1	1	1	-1	-1	1	1	1	1	-1	-	-	pyrrolidone-carboxylate peptidase
"opCcV0100001534"	-	-	-	-	CCOA0070	-	1	1	-1	-1	-1	1	0	1	1	-1	-	-	hygromycin-B-phosphotransferase
"opCcV0100000960"	-	-	-	-	CCOA0071	-	1	1	1	-1	-1	-1	-1	1	-1	-	-	-	conserved hypothetical protein
"opCcV0100001317"	-	-	-	-	CCOA0072	-	1	1	1	-1	0	1	1	1	-1	1	-	-	TnpV
"opCcV0100001506"	-	-	-	-	CCOA0073	-	1	-1	1	1	1	1	1	1	1	0	-	-	site-specific recombinase, resolvase family,
"opCcV0100001341"	-	-	-	-	CCOA0074	-	1	0	1	1	1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100000522"	-	-	-	-	CCOA0075	-	1	1	1	1	0	1	-1	1	1	1	-	-	transcriptional regulator, Cro/C1 family
"opCcV0100000944"	-	-	-	-	CCOA0076	-	1	1	1	-1	0	1	1	-1	1	1	-	-	hypothetical protein
"opCcV0100000407"	-	-	-	-	CCOA0077	-	1	1	1	1	-1	0	-1	-1	1	1	-	-	hypothetical protein
"opCcV0100000685"	-	-	-	-	CCOA0078	-	0	-1	1	1	1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000016"	-	-	-	-	CCOA0079	-	1	1	1	-1	-1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100000543"	-	-	-	-	CCOA0080	-	1	1	1	1	-1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100001312"	-	-	-	-	CCOA0081	-	1	0	1	1	1	-1	-1	1	-1	1	-	-	hypothetical protein
"opCcV0100001690"	-	-	-	-	CCOA0083	-	1	1	1	1	0	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100001250"	-	-	-	-	CCOA0084	-	1	1	1	-1	-1	0	1	1	-1	1	-	-	Domain of unknown function (DUF332) superfamily
"opCcV0100000139"	-	-	-	-	CCOA0101	-	1	1	1	1	1	1	1	0	1	-1	-	-	hypothetical protein
"opCcV0100002042"	-	-	-	-	CCOA0102	-	0	-1	1	1	0	1	1	1	1	0	-	-	conserved hypothetical protein
"opCcV0100001766"	-	-	-	-	CCOA0103	-	1	1	-1	1	-1	1	1	-1	1	0	-	-	conserved hypothetical protein
"opCcV0100000705"	-	-	-	-	CCOA0104	-	1	1	1	-1	-1	0	-1	-1	1	1	-	-	hypothetical protein
"opCcV0100001289"	-	-	-	-	CCOA0105	-	1	1	1	1	-1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100002072"	-	-	-	-	CCOA0106	-	0	1	1	-1	-1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100000094"	-	-	-	-	CCOA0107	-	1	1	1	1	-1	1	1	1	1	-1	-	-	hypothetical protein

"opCcV0100001675"	-	-	-	-	CCOA0108	-	1	1	0	1	-1	0	1	1	-1	1	-	-	hypothetical protein
"opCcV0100001626"	-	-	-	-	CCOA0109	-	0	0	1	1	1	1	0	1	1	1	-	-	conserved hypothetical protein
"opCcV0100000453"	-	-	-	-	CCOA0110	-	1	1	1	-1	1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100001929"	-	-	-	-	CCOA0111	-	1	1	1	-1	0	-1	1	-1	0	1	-	-	hypothetical protein
"opCcV0100002037"	-	-	-	-	CCOA0112	-	1	-1	1	1	-1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000811"	-	-	-	-	CCOA0113	-	1	1	1	-1	1	1	-1	-1	-1	1	-	-	hypothetical protein
"opCcV0100000556"	-	-	-	-	CCOA0114	-	1	1	1	-1	1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100000251"	-	-	-	-	CCOA0115	-	1	1	1		-1	1	0	1	1	1	-	-	hypothetical protein
"opCcV0100001717"	-	-	-	-	CCOA0116	-	1	1	1		-1	1	-1	1	0	-1	-	-	hypothetical protein
"opCcV0100001258"	-	-	-	-	CCOA0117	-	1	1	1	-1	-1	1	-1	1	0	0	-	-	hypothetical protein
"opCcV0100001028"	-	-	-	-	CCOA0118	-	1	1	1		1	1	1	1	1	1	-	-	hypothetical protein
"opCcV0100000289"	-	-	-	-	CCOA0119	-	1	1	0	1	-1	1	0	-1	1	-1	-	-	hypothetical protein
"opCcV0100001894"	-	-	-	-	CCOA0120	-	1	-1	0	1	1	1	1	1	0	0	-	-	hypothetical protein
"opCcV0100000073"	-	-	-	-	CCOA0121	-	1	1	1	-1	-1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100000906"	-	-	-	-	CCOA0122	-	1	1	1	1	-1	1	1	1	1	-1	-	-	hypothetical protein
"opCcV0100000799"	-	-	-	-	CCOA0123	-	1	1	1	1	0	1	-1	1		1	-	-	hypothetical protein
"opCcV0100000283"	-	-	-	-	CCOA0124	-	1	1	1	-1	-1	1	-1	1	1	-1	-	-	hypothetical protein
"opCcV0100001464"	-	-	-	-	CCOA0125	-	1	1	1	1	-1	1	1	-1	1	-1	-	-	hypothetical protein
"opCcV0100000189"	-	-	-	-	CCOA0126	-	1	1	0	1	-1	1	-1	1	1	1	-	-	hypothetical protein
"opCcV0100000285"	-	-	-	-	CCOA0127	-	1	1	-1	-1	0	-1	0	-1	-1	1	-	-	conserved hypothetical protein
"opCcV0100000270"	-	-	-	-	CCOA0128	-	1	1	1	1	-1	1	-1	1	-1	1	-	-	hypothetical protein
"opCcV0100001759"	-	-	-	-	CCOA0130	-	1	1	1	1	1	1	1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100000641"	-	-	-	-	CCOA0131	-	1	0	1	-1	-1	1	-1	1	1	-1	-	-	conserved hypothetical protein
"opCcV0100001107"	-	-	-	-	CCOA0132	-	1	1	1	1	1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100001807"	-	-	-	-	CCOA0133	-		1	1	1	1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100001253"	-	-	-	-	CCOA0134	-	1	1	1	1	-1	1	1	1	1	1	-	-	conserved hypothetical protein
"opCcV0100000724"	-	-	-	-	CCOA0135	-	1	0	1	1	1	1	1	1	1	1	-	-	hypothetical protein
"opCcV010000"	-	-	-	-	CCOA0136	-		1	1	0	0	1	1	1	1	1	-	-	lipoprotein, putative

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0477"	-	-	-	-	CCOA0137	-	1	1	1	1	-1	-1	1	-1	1	1	-	-	conserved hypothetical protein	
"opCcV010000 0468"	-	-	-	-	CCOA0138	-	1	1	1	1	-1	-1	0	1	1	-1	-	-	conserved hypothetical protein	
"opCcV010000 0670"	-	-	-	-	CCOA0139	-	1	1	1	1	1	1	1	1	1	1	-	-	hcp protein	
"opCcV010000 0044"	-	-	-	-	CCOA0140	-	1	1	1	1	-1	1	1	1	1	1	-	-	conserved hypothetical protein	
"opCcV010000 1716"	-	-	-	-	CCOA0142	-	-1	1	1	1	-1	0	1	1	1	1	-	-	hypothetical protein	
"opCcV010000 1906"	-	-	-	-	CCOA0145	-	1	1	1	1		0	-1	1	1	1	-	-	pseudogene	
"opCcV010000 0920"	-	-	-	-	CCOA0146	-	1	1	1	1	0		-1	1	1	1	-	-	ISCCo1, transposase orfB	
"opCcV010000 0347"	-	-	-	-	CCOA0147	-	1	1	1	-1	-1	-1	1	1	1	-1	-	-	Fic family protein, putative	
"opCcV010000 0938"	-	-	-	-	CCOA0148	-	1	1	1	-1	1	-1	-1	1	-1	0	-	-	pseudogene	
"opCcV010000 1282"	-	-	-	-	CCOA0149	-	1	1	0	1	1		-1	1	1	1	-	-	ISCCo1, transposase orfB	
"opCcV010000 0502"	-	-	-	-	CCOA0151	-			0	1	-1	-1	-1	1	1	-1	-	-	lipase family protein	
"opCcV010000 0442"	-	-	-	-	CCOA0152	-	1	1	1	1	-1	1	1	-1	1	1	-	-	hypothetical protein	
"opCcV010000 0253"	-	-	-	-	CCOA0153	-	1	1	1	0	-1	-1	1	1	1	0	-	-	hypothetical protein	
"opCcV010000 0500"	-	-	-	-	CCOA0154	-	1	1	1	0	-1	1	1	-1	1	1	-	-	hypothetical protein	
"opCcV010000 1883"	-	-	-	-	CCOA0155	-	1	1	1	1	-1	0	-1	1	1	1	-	-	conserved hypothetical protein	
"opCcV010000 0306"	-	-	-	-	CCOA0156	-	1	1	1	-1	-1	1	-1	0	1	-1	-	-	hypothetical protein	
"opCcV010000 0892"	-	-	-	-	CCOA0157	-	1	-1	1	1	1	0	1	1	-1	0	-	-	conserved hypothetical protein	
"opCcV010000 1354"	-	-	-	-	CCOA0158	-	1	1	1	-1	-1	1	1	-1	1	-1	-	-	hypothetical protein	
"opCcV010000 1811"	-	-	-	-	CCOA0159	-		-1	0	-1	1	1	-1	1	1	-1	-	-	DNA primase, putative	
"opCcV010000 0322"	-	-	-	-	CCOA0160	-	-1	-1	0	-1	1	-1	-1	1	-1	-1	-	-	hypothetical protein	
"opCcV010000 1892"	-	-	-	-	CCOA0161	-			0	-1	1	-1	-1	1	1	-1	-	-	DNA primase, putative	
"opCcV010000 0646"	-	-	-	-	CCOA0162	-		-1	1	-1	1	-1	-1	1	-1	-1	-	-	DNA primase trac	
"opCcV010000 0935"	-	-	-	-	CCOA0163	-			-1	1	-1	0	-1	-1	1	-1	1	-	-	conserved hypothetical protein
"opCcV010000 0552"	-	-	-	-	CCOA0164	-		-1	-1	-1	1	-1	-1	1	-1	-1	-	-	conserved hypothetical protein	
"opCcV010000 1496"	-	-	-	-	CCOA0165	-			1		-1	1	-1	1	1	-1	-	-	hypothetical protein	
"opCcV010000 0772"	-	-	-	-	CCOA0167	-			1			1	-1	-1	1	-1	-	-	hypothetical protein	
"opCcV010000 1485"	-	-	-	-	CCOA0168	-	1	0	1	1	1	1	1	-1	1	1	-	-	conserved hypothetical protein	
"opCcV010000 2017"	-	-	-	-																

"opCcV0100002056"	-	-	-	-	CCOA0169	-	1	1	1	0	-1	1	-1	-1	1	1	-	-	conserved hypothetical protein
"opCcV0100001829"	-	-	-	-	CCOA0170	-	1	1	1	1	-1	1	1	1	1	0	-	-	hypothetical protein
"opCcV0100001579"	-	-	-	-	CCOA0171	-	1	0	1	-1	1	1	1	-1	1	1	-	-	conserved hypothetical protein
"opCcV0100000325"	-	-	-	-	CCOA0172	-	1	1	1	1	-1	1	-1	1	-1	1	-	-	hypothetical protein
"opCcV0100000449"	-	-	-	-	CCOA0173	-	1	1	1	1	-1	1	1	-1	1	1	-	-	conserved hypothetical protein
"opCcV0100001185"	-	-	-	-	CCOA0174	-	1	-1	1	-1	1	1	1	1	-1	1	-	-	conserved hypothetical protein
"opCcV0100000097"	-	-	-	-	CCOA0175	-	1	0	0	-1	-1	-1	0	-1	1	0	-	-	VapD-related protein
"opCcV0100002075"	-	-	-	-	CCOA0176	-	-1	-1	1	1	-1	1	1	0	1	-1	-	-	plasmid replication protein, putative
"opCcV0100000102"	-	-	-	-	CCOA0177	-	1	1	1	1	1	1	1	-1	1	-1	-	-	site-specific recombinase, resolvase family
"opCcV0100001361"	-	-	-	-	CCOA0178	-	1	0	1	1	1	1	1	1	0	1	-	-	virulence-associated protein 2
"opCcV0100000271"	-	-	-	-	CCOA0179	-	1	1	-1	1	-1	1	1	1		-1	-	-	hypothetical protein
"opCcV0100001182"	-	-	-	-	CCOA0180	-	1	1	1	1	1	1	1	1	1	-1	-	-	TraC protein
"opCcV0100000528"	-	-	-	-	CCOA0181	-	1	-1	1	1	0	0	0	1	-1	0	-	-	type IV secretion system protein VirB4
"opCcV0100001249"	-	-	-	-	CCOA0182	-	1	1	0	1	1	1	1	-1	1	-1	-	-	antirepressor, putative
"opCcV0100000515"	-	-	-	-	CCOA0184	-		1	0	1	1	1	-1	1	1	1	-	-	single-strand binding protein, putative
"opCcV0100000773"	-	-	-	-	CCOA0185	-	1	1	0	-1	1	-1	-1	1	0	-1	-	-	conserved hypothetical protein
"opCcV0100002012"	-	-	-	-	CCOA0186	-	0	1	1	1	-1	1	1	-1	1	1	-	-	type IV secretion system protein, putative
"opCcV0100001201"	-	-	-	-	CCOA0187	-	1	1	0	0	-1	-1	-1	1	1	-1	-	-	TrbL/VirB6 plasmid conjugal transfer protein
"opCcV0100001056"	-	-	-	-	CCOA0188	-	1	1	0	-1	-1	1	0	-1	1	-1	-	-	lipoprotein, putative
"opCcV0100000925"	-	-	-	-	CCOA0189	-	1	1	1	-1	1	1	-1	1	1	1	-	-	type IV secretion system protein VirB8,
"opCcV0100002034"	-	-	-	-	CCOA0190	-	1	1	1	-1	-1	1	1	1	1	0	-	-	type IV secretion system protein VirB9
"opCcV0100001206"	-	-	-	-	CCOA0191	-	1	1	1	-1	1	-1	0	1	-1	1	-	-	type IV secretion system protein VirB10,
"opCcV0100001020"	-	-	-	-	CCOA0192	-	1	1	1	-1	-1	1	-1	1	1	1	-	-	type IV secretion system protein VirB11
"opCcV0100001962"	-	-	-	-	CCOA0193	-	1	1	1	1	0	1	1	-1	1	1	-	-	cag pathogenicity island protein (cag5),
"opCcV0100000388"	-	-	-	-	CCOA0194	-	1	1	-1	-1	1	0	1	1	1	1	-	-	cag island protein, putative
"opCcV0100000537"	-	-	-	-	CCOA0195	-	1	1	1	0	-1	1	1	-1	1	-1	-	-	YggA-like protein
"opCcV0100001638"	-	-	-	-	CCOA0196	-	1	1	1	1	0	-1	0	1	-1	1	-	-	conserved hypothetical protein
"opCcV010000"	-	-	-	-	CCOA0197	-	1	1	1	1	1	1	1	-1	1	1	-	-	hypothetical protein

"opCcV010000 1321"	-	-	-	-	pCC31p13	-	-1	-1	1	-1	1	1	-1	1	1	-1	-	-	cpp13
"opCcV010000 2080"	-	-	-	-	pCC31p14	-	1	1	1	1	-1	1	1	-1	1	1	-	-	cpp14
"opCcV010000 1336"	-	-	-	-	pCC31p15	-	1	0	0	-1	-1	0	-1	1	0	1	-	-	cpp15
"opCcV010000 0825"	-	-	-	-	pCC31p16	-	1	1	1	1	0	1	1	-1	1	-1	-	-	cpp16
"opCcV010000 1960"	-	-	-	-	pCC31p17	-	1	1	1	1	-1	1	1	1	1	1	-	-	cpp17
"opCcV010000 1732"	-	-	-	-	pCC31p18	-	1	1	1	1	-1			0	1	-1	-	-	cpp18
"opCcV010000 1909"	-	-	-	-	pCC31p19	-	1	1	1	1	0	1	1	1	1	1	-	-	cpp19
"opCcV010000 2068"	-	-	-	-	pCC31p21	-	1	1	1	1	-1	1	1	0	1	1	-	-	cpp22
"opCcV010000 0675"	-	-	-	-	pCC31p22	-			0			1	1	1		-1	-	-	cpp23
"opCcV010000 1373"	-	-	-	-	pCC31p23	-	1	1	1	1	0	1	1	1	1	1	-	-	cpp24
"opCcV010000 1757"	-	-	-	-	pCC31p24	-			0	1		-1		-1	1	-1	-1	-	cpp25
"opCcV010000 0985"	-	-	-	-	pCC31p25	-	1	1	-1	1	-1	1	1	-1	1	0	-	-	cpp26
"opCcV010000 2035"	-	-	-	-	pCC31p26	-	1	0	1	1	-1	1	-1	0	1	1	-	-	cpp27
"opCcV010000 1362"	-	-	-	-	pCC31p27	-	1	1	1	0	-1	1	1	-1	1	1	-	-	vapd
"opCcV010000 1318"	-	-	-	-	pCC31p28	-	1	0	0	1	1	1	1	1	-1	0	-	-	cpp29
"opCcV010000 1969"	-	-	-	-	pCC31p29	-	1	0	1	1	0	1	1	1	1	1	-	-	cmgb2
"opCcV010000 1153"	-	-	-	-	pCC31p30	-	1	1	-1	-1	-1	1	1	-1	1	-1	-	-	cmgb3/4
"opCcV010000 1760"	-	-	-	-	pCC31p31	-	1	1	0	-1	0	1	-1	1	1	1	-	-	cpp32
"opCcV010000 0972"	-	-	-	-	pCC31p32	-	1	1	1	-1	0	0	1	1	1	-1	-	-	cpp33
"opCcV010000 0194"	-	-	-	-	pCC31p33	-	1	1	1		0	-1	1	1	1	1	-	-	ssb1
"opCcV010000 1287"	-	-	-	-	pCC31p34	-	1	1	0	-1	-1	1	1		1	-1	-	-	cpp35
"opCcV010000 1463"	-	-	-	-	pCC31p35	-	1	1	1	0	1	1	1	1	-1	1	-	-	cmgb5
"opCcV010000 1487"	-	-	-	-	pCC31p36	-	1	1	0	1		0	-1	1	1	1	-	-	cmgb6
"opCcV010000 0790"	-	-	-	-	pCC31p38	-	1	1	1	-1	-1	1	-1	1	-1	1	-	-	cmgb8
"opCcV010000 2021"	-	-	-	-	pCC31p39	-	1	1	1	0	-1	1	1	0	1	1	-	-	cmgb9
"opCcV010000 2005"	-	-	-	-	pCC31p40	-	1	1	1	1	-1	1	1	1	1	1	-	-	cmgb10
"opCcV010000 0177"	-	-	-	-	pCC31p41	-		1	1	1	1	1	1	1	1	1	-	-	cmgb11
"opCcV010000"	-	-	-	-	pCC31p42	-	1	0	1	1	-1	1	1	-1	1	1	-	-	cmgd4

"opCjjV010000053"	-	-	pTet_21	-	-	-			1	-1			-1	1			hypothetical protein	-	-
"opCjjV010000043"	-	-	pTet_22	-	-	-	1	1	1	-1	0	1	-1	-1	0	-1	hypothetical protein	-	-
"opCjjV010000069"	-	-	pTet_23	-	-	-	1	1	1	1	1	1	-1	-1	1	1	hypothetical protein	-	-
"opCjjV010000029"	-	-	pTet_24	-	-	-	1	1	0	-1	1	1	-1	1	1	-1	hypothetical protein	-	-
"opCjjV010000202"	-	-	pTet_25	-	-	-	1	-	0	1	1	-1	0	1	1	0	hypothetical protein	-	-
"opCjjV010000063"	-	-	pTet_26	-	-	-	1	1	1	1	-1	1	1	1		0	hypothetical protein	-	-
"opCjjV010000178"	-	-	pTet_27	-	-	-	1	1	1	1	-1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000129"	-	-	pTet_28	-	-	-	1	1	0		-1	0	-1	1	1	1	hypothetical protein	-	-
"opCjjV010000153"	-	-	pTet_29	-	-	-	1	1		1	-1	0	1	-1	1	1	hypothetical protein	-	-
"opCjjV010000111"	-	-	pTet_30	-	-	-	1	-	1	1	1	1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000056"	-	-	pTet_31	-	-	-	1	1	1	1	1	1	0	1	1	1	hypothetical protein	-	-
"opCjjV010000106"	-	-	pTet_32	-	-	-	1	-1	1	0	1	-1	-1	1	1	1	hypothetical protein	-	-
"opCjjV010000190"	-	-	pTet_33	-	-	-	1	1	1	1	-1	1	-1	1	-1	-1	hypothetical protein	-	-
"opCjjV010000140"	-	-	pTet_34	-	-	-	0	-1	1	-1	1	1	0	1	-1	1	hypothetical protein	-	-
"opCjjV010000042"	-	-	pTet_35	-	-	-	1	1	1	0	-1	1	0	1	1	-1	hypothetical protein	-	-
"opCjjV010000145"	-	-	pTet_36	-	-	-	1	1	-1	-1	-1	-1	1	1	1	1	hypothetical protein	-	-
"opCjjV010000185"	-	-	pTet_37	-	-	-	1	1	1	1	-1	1	1	1	-1	1	hypothetical protein	-	-
"opCjjV010000015"	-	-	pTet_38	-	-	-	1	1	1	1	1	0	-1	1	-1	1	hypothetical protein	-	-
"opCjjV010000149"	-	-	pTet_39	-	-	-	1	1	1	-1	1	1	0	1	1	1	hypothetical protein	-	-
"opCjjV010000077"	-	-	pTet_40	-	-	-	1	1	1	-1	1	1	1	-1	1	-1	hypothetical protein	-	-
"opCjjV010000118"	-	-	pTet_41	-	-	-	1	0	1	-1	1	-1	-1	1		1	hypothetical protein	-	-
"opCjjV010000057"	-	-	pTet_42	-	-	-	1	1	1	1	0	1	1	-1	1	-1	hypothetical protein	-	-
"opCjjV010000012"	-	-	pTet_43	-	-	-			1	-1		1	1	1	1		hypothetical protein	-	-
"opCjjV010000097"	-	-	pTet_44	-	-	-			1		-1			1	1		hypothetical protein	-	-
"opCcV0100000864"	CJE_Cj16S A	rrsA	Cjr01	-	-	CCO_Cc16 SB	rrsB	1	1	1	1	1	1	1	1	1	16S ribosomal RNA	16S ribosomal RNA	16S ribosomal RNA
"opCcV0100000237"	CJE_Cj23S A	rrlA	Cjr05	-	-	CCO_Cc23 SB	rrlB	1	1	0	1	1	1	1	1	0	23S ribosomal RNA	23S ribosomal RNA	23S ribosomal RNA
"opCjV0100001047"	CJE_Cjtmp B1	-	-	-	-	-	-	1	1	1	1	1	1	1	1	1	-	sRNA	-
"opCjjV0100000"	CJE_Cjtm	-	Cjs01	-	-	-	-	1	1	1	1	1	1	1	1	1	tmRNA	sRNA	-

"opCcV010000 1380"	CJE_tRNA -Ser-2	-	-	-	CCO_tRNA -Ser-2	-		0	0	1	1	-1	0	0	1	1	-	tRNA-Ser	tRNA-Ser
"opCcV010000 1987"	CJE_tRNA -Ser-3	-	Cjt06	-	CCO_tRNA -Ser-3	-		-1	-1		-1	-1	-1	1	1	-1	tRNA-Ser	tRNA-Ser	tRNA-Ser
"opCcV010000 1276"	CJE_tRNA -Thr-1	-	-	-	CCO_tRNA -Thr-1	-	1	1	1	1	1	1	1	1	1	1	-	tRNA-Thr	tRNA-Thr
"opCjjV010000 157"	CJE_tRNA -Thr-2	-	Cjt1	-	-	-	1	1	1	1	1	1	1	1	1	1	tRNA-Thr	tRNA-Thr	-
"opCcV010000 1423"	CJE_tRNA -Trp-1	-	-	-	CCO_tRNA -Trp-1	-	1	1	1	1	1	1	1	1	1	1	-	tRNA-Trp	tRNA-Trp
"opCcV010000 1928"	CJE_tRNA -Tyr-1	-	-	-	CCO_tRNA -Tyr-1	-	1	0	1	1	-1	1	1	1	1	-1	-	tRNA-Tyr	tRNA-Tyr
"opCcV010000 1203"	CJE_tRNA -Val-1	-	-	-	CCO_tRNA -Val-1	-	-1	1	1	1	0	1	1	1	1	1	-	tRNA-Val	tRNA-Val
"opCcV010000 0443"	CJE_tRNA -Val-3	-	-	-	CCO_tRNA -Val-3	-	1	1	1	1	1	1	1	1	1	1	-	tRNA-Val	tRNA-Val

1=present; -1=absent/highly divergent; 0=slightly divergent

	Lipooligosaccharide region
	Flagellar modification region
	Capsule region in <i>C. jejuni</i> RM1221
	<i>C. jejuni</i> NCTC11168 specific capsule genes

Supplementary table 4: Genes present and highly divergent/absent in the hyperinvasive and low invasive *C. jejuni*.

Gene name/number			Hyperinvasive <i>C. jejuni</i>						Low-invasive <i>C. jejuni</i>				Role/function			COG functional grouping
GROUP 3: Genes present in all low-invasive <i>C. jejuni</i> and absent/highly divergent form five of six hyperinvasive <i>C. jejuni</i>			01_10	01_35	01_04	01_41	01_51	EX114	01_30	01_32	01_46	01_39	<i>C. jejuni</i> NCTC11168	<i>C. jejuni</i> RM1221	<i>C. coli</i> RM2228	Fouts <i>et al.</i> , 2005
<i>C. jejuni</i> NCTC 11168	<i>C. jejuni</i> RM1221	<i>C. coli</i> RM2228														
<i>accA</i> (Cj0443)	<i>accA</i> (CJE0495)	<i>accA</i> (CCO0532)	1	-1	-1	-1	-1	-1	1	1	1	1	acetyl-coenzyme A carboxylase carboxyl	acetyl-CoA carboxylase alpha subunit	acetyl-CoA carboxylase, carboxyl transferase,	Lipid transport and metabolism
Cj0911	CJE0989	CCO1012	1	-1	-1	-1	-1	-1	1	1	1	1	putative periplasmic protein	SCO1/SenC family protein	probable periplasmic protein Cj0911	General function prediction only
.....	CCOA0043	-1	-1	-1	-1	1	-1	1	1	1	1	hypothetical protein	hypothetical protein	Hypothetical
Cj1720	CJE1890	CCO0019	-1	1	-1	-1	-1	-1	1	1	1	1	hypothetical protein	hypothetical protein	conserved hypothetical protein	Hypothetical
GROUP 4: Genes absent/highly divergent in all low-invasive <i>C. jejuni</i> and present in five of six hyperinvasive <i>C. jejuni</i>																
<i>infB</i> (Cj0136)	<i>infB</i> (CJE0131)	<i>infB</i> (CCO1743)	1	1	1	-1	1	1	-1	-1	-1	-1	translation initiation factor IF-2	translation initiation factor IF-2	translation initiation factor IF-2	Translation
Cj0380c	1	1	1	1	1	-1	-1	-1	-1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein	Hypothetical
Cjp21	1	1	1	1	1	-1	-1	-1	-1	-1	hypothetical protein	Hypothetical
GROUP 5: Genes present in five of six hyperinvasive <i>C. jejuni</i> and absent/highly divergent in three of four low invasive <i>C. jejuni</i>																
Cj1002c	<i>sixA</i> (CJE1082)	<i>sixA</i> (CCO1064)	-1	1	1	1	1	1	-1	-1	1	-1	conserved hypothetical protein	phosphohistidine phosphatase SixA	phosphohistidine phosphatase SixA, putative	Signal transduction mechanism
Cj1176c	CJE1310	CCO1245	1	-1	1	1	1	1	1	-1	-1	-1	hypothetical protein	twin-arginine translocation protein, TatA/E	Sec-independent protein translocase protein	Intracellular trafficking and secretion

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Cj1343c	<i>ctsG</i> (CJE1532)	<i>ctsG</i> (CCO1449)	1	-1	1	1	1	1	-1	1	-1	-1	putative periplasmic protein	transformation system protein	transformation system protein	Cell wall/membrane biogenesis
Cj1287c	CJE1479	<i>maeB</i> (CCO1392)	1	-1	1	1	1	1	-1	1	-1	-1	malate oxidoreductase	NADP-dependent malic enzyme, truncation	malic enzyme	Energy production and conversion
Cj1476c	CJE1649	CCO1582	1	-1	1	1	1	1	-1	1	-1	-1	pyruvate-flavodoxin oxidoreductase	pyruvate ferredoxin/flavodoxin oxidoreductase	pyruvate ferredoxin/flavodoxin oxidoreductase	Energy production and conversion
Cj0544	CJE0648	CCO0641	1	1	-1	1	1	1	-1	1	-1	-1	putative integral membrane protein	hypothetical protein	probable integral membrane protein Cj0544	Cell wall/membrane biogenesis
Cj0943	<i>lolA</i> (CJE1021)	CCO0952	1	1	-1	1	1	1	-1	1	-1	-1	putative periplasmic protein	outer-membrane lipoprotein carrier protein	probable periplasmic protein Cj0943	Cell wall/membrane biogenesis
.....	CCO1326	1	1	1	-1	1	1	-1	1	-1	-1	hypothetical protein	Hypothetical
Cj0717	CJE0817	CCO0783	1	1	1	-1	1	1	-1	-1	1	-1	hypothetical protein	hypothetical protein	<i>arsC</i> family protein	Arsenic resistance
.....	CCOA0113	1	1	1	-1	1	1	-1	-1	-1	1	hypothetical protein	Hypothetical
Cj0151c	CJE0147	CCO1726	1	1	1	-1	1	1	-1	1	-1	-1	putative periplasmic protein	hypothetical protein	probable periplasmic protein Cj0151c	Hypothetical
<i>pyrC2</i> (Cj1195c)	CJE1329	CCO1265	1	1	1	-1	1	1	-1	1	-1	-1	putative dihydroorotase	dihydroorotase	dihydroorotase, putative	Nucleotide transport and metabolism
Cjp17	1	1	1	1	-1	1	-1	-1	1	-1	hypothetical protein	Hypothetical
<i>sucD</i> (Cj0534)	<i>sucD</i> (CJE0638)	<i>sucD</i> (CCO0631)	1	1	1	1	-1	1	-1	-1	1	-1	succinyl-coA synthetase alpha chain	succinyl-CoA synthase, alpha subunit	SucD	Energy production and conversion
Cjp30	1	1	1	1	-1	1	-1	-1	1	-1	hypothetical protein	Hypothetical
Cjp39	1	1	1	1	-1	1	-1	-1	-1	1	hypothetical protein	Hypothetical

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Cj0581	<i>nidH</i> (CJE0684)	<i>mutT</i> (CCO1375)	1	1	1	1	-1	1	-1	-1	1	-1	putative NTPase	dinucleoside polyphosphate hydrolase	(di)nucleoside polyphosphate hydrolase	General function prediction only
<i>cfrA</i> (Cj0755)	<i>cfrA</i> (CJE0847)	CCO0810	1	1	1	1	-1	1	-1	1	-1	-1	putative iron uptake protein	ferric receptor CfrA	ferric receptor CfrA	Inorganic iron transport and metabolism
Cj1553c" <i>hsdM</i> "	1	1	1	1	-1	1	1	-1	-1	-1	putative type I restriction enzyme M protein	Defense mechanisms
.....	CCO1531	1	1	1	1	-1	1	-1	1	-1	-1	UDP-glucose 4-epimerase, putative	Amino acid transport and metabolism
.....	CCO0231	1	1	1	1	-1	1	-1	-1	1	-1	hypothetical protein	Hypothetical
.....	<i>cysQ</i> in RM2228 only	1	1	1	1	-1	1	-1	-1	1	-1	3'(2),5'-bisphosphate nucleotidase
.....	CCO1688	1	1	1	1	-1	1	-1	1	-1	-1	hypothetical protein	Hypothetical
.....	CCOA0017	1	1	1	1	-1	1	-1	1	-1	-1	hypothetical protein	Hypothetical
.....	CCOA0026	1	1	1	1	-1	1	1	-1	-1	-1	conserved hypothetical protein	Hypothetical
.....	CCO1269	1	1	1	1	-1	1	-1	-1	-1	1	TraH protein, Conjugative relaxosome accessory transposon protein
<i>fliE</i> (Cj0526c)	<i>fliE</i> (CJE0630)	<i>fliE</i> (CCO0623)	1	1	1	1	-1	1	1	-1	-1	-1	putative flagellar hook-basal body complex	flagellar basal body protein	flagellar hook-basal body complex protein	Cell motility/Intracellular trafficking and secretion
	CJE1333	CCO1269	1	1	1	1	-1	1	1	-1	-1	-1	putative iron/ascorbate-dependent	oxidoreductase, 2OG-Fe(II) oxygenase family	oxidoreductase, 2OG-Fe(II) oxygenase family	General function prediction only
pTet_33			1	1	1	1	-1	1	-1	1	-1	-1	hypothetical protein	Hypothetical

<i>tsf</i> (Cj1181c)	<i>tsf</i> (CJE1315)	<i>tsf</i> (CCO1250)	1	1	1	1	1	-1	1	-1	-1	-1	elongation factor TS	elongation factor Ts	translation elongation factor Ts	Translation
Cj0182	CJE0175	CCO1695	1	1	1	1	1	-1	-1	1	-1	-1	transmembrane transport protein	antibiotic transport protein, putative	transmembrane transport protein Cj0182	Lipid transport and metabolism
Cj0605	CJE0708	CCO0688	1	1	1	1	1	-1	-1	1	-1	-1	putative amidohydrolase	carboxypeptidase	peptidase, M20/M25/M40 family	General function prediction only
Cj0111	CJE0106	CCO1770	1	1	1	1	1	-1	1	-1	-1	-1	periplasmic protein	TonB domain protein	periplasmic protein Cj0111	Transport and binding protein
<i>nusB</i> (Cj0382c)	<i>nusB</i> (CJE0431)	<i>nusB</i> (CCO0471)	1	1	1	1	1	-1	-1	1	-1	-1	transcription termination protein	transcription antitermination protein NusB	transcription antitermination factor NusB	Transcription
<i>selD</i> (Cj1504c)	<i>selD</i> (CJE1677)	<i>selD</i> (CCO1613)	1	1	1	1	1	-1	-1	1	-1	-1	putative selenide, water dikinase	selenide, water dikinase	selenide, water dikinase	Translation
<i>ceuB</i> (Cj1352)	<i>ceuB</i> (CJE1541)	<i>ceuB</i> (CCO1458)	1	1	1	1	1	-1	-1	1	-1	-1	enterochelin uptake permease	enterochelin ABC transporter, permease protein	enterochelin ABC transporter, permease protein	Inorganic iron transport and metabolism
GROUP 6: Genes absent/highly divergent in five of six hyperinvasive <i>C. jejuni</i> and present in three of four low invasive <i>C. jejuni</i>																
Cj1021c	CJE1165	CCO1088	1	-1	-1	-1	-1	-1	-1	1	1	1	putative periplasmic protein	porin domain protein	probable periplasmic protein Cj1021c-related	Cell wall/membrane biogenesis
Cj0102	CJE0097	CCO1779	1	-1	-1	-1	-1	-1	1	1	-1	1	ATP synthase F0 sector B' subunit	ATP synthase subunit B	ATP synthase F0, subunit b' (atpF'), putative	Energy production and conversion
<i>ispA</i> (Cj1644)	<i>ispA</i> (CJE1816)	<i>ispA</i> (CCO0170)	1	-1	-1	-1	-1	-1	-1	1	1	1	geranyltranstransferase	geranyltranstransferase	geranyltranstransferase (<i>ispA</i>)	Coenzyme transport and metabolism

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Cj1247c	CJE1383	CCO1337	-1	1	-1	-1	-1	-1	1	1	1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein	Hypothetical
.....	CCOA0028	-1	-1	1	-1	-1	-1	1	1	1	-1	hypothetical protein	Hypothetical
Cj0208	CJE0328	CCO0358	-1	-1	-1	-1	1	-1	1	1	1	-1	hypothetical protein	hypothetical protein	conserved hypothetical protein	Hypothetical
<i>purN</i> (Cj0187c)	<i>purN</i> (CJE0180)	<i>purN</i> (CCO1687)	-1	-1	-1	-1	1	-1	1	1	-1	1	phosphoribosyl glycynamide formyltransferase	phosphoribosylglycinamide formyltransferase	phosphoribosyl glycynamide formyltransferase	Nucleotide transport and metabolism
Cj0974	CJE1055	-1	-1	-1	-1	1	-1	1	1	-1	1	hypothetical protein	hypothetical protein	Hypothetical
Cj0997	<i>gidB</i> (CJE1077)	<i>gidB</i> (CCO1059)	-1	-1	-1	-1	-1	1	1	1	1	-1	<i>gidB</i> homolog	methyltransferase GidB	methyltransferase GidB	Cell wall/membrane biogenesis

1=present;-1=absent/highly divergent

Supplementary table 5: *C. jejuni* 01/10 prophage 1 and 2 genes, ORFs with best match in campylobacters other than *C. jejuni* RM1221 and LOS biosynthesis loci identified by BLASTx annotation tool.

Gene homolog/ortholog	Annotated function
Prophage	
CJJ81176_1532	putative homolog of BLC protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
	replication-associated A domain protein [<i>Vibrio cholerae</i> HE39]
MOY_16467	hypothetical protein [<i>Halomonas</i> strain GFAJ-1]
MOY_16472	hypothetical protein [<i>Halomonas</i> strain GFAJ-1]
MOY_16477	hypothetical protein [<i>Halomonas</i> strain GFAJ-1]
MOY_16482	hypothetical protein, partial [<i>Halomonas</i> strain GFAJ-1]
CJJ81176_1534	type I restriction-modification enzyme, R subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
Other genes	
conserved hypothetical protein	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CF93-6
conserved hypothetical protein	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CF93-6
CJSA_0241	molybdopterin containing oxidoreductase in [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]
Cj0264c	molybdopterin containing oxidoreductase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cj1679	hypothetical protein in <i>C. jejuni</i> NCTC 11168
ICDCCJ07001_615	K ⁺ -transporting ATPase, B subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]. Two component system

Cj0736	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
CJ1055c	sulfatase family protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
CJSA_1363	putative sugar transferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]
CJSA_1352	putative sugar transferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]
ICDCCJ07001_1325	cryptic C4-dicarboxylate transporter DcuD [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
Cj0628	lipoprotein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cj1137c	glycosyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cj1136	Cj1136 in <i>C. jejuni</i> NCTC1168 except 2 nucleotide bases mismatch in the beginning atgaaaaccgt in 01/51 and atgaaaaaagt in NCTC11168
Cj1324	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cj1323	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cj1321	transferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
	putative glycosyltransferase <i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 87330
CJJ81176_1534	type I restriction-modification enzyme, R subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
CJJ81176_1534	type I restriction-modification enzyme, R subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
CJJ81176_1536	hypothetical protein in [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
Cj1552c	<i>mloB</i> hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
CJJ81176_1539	type I restriction-modification system, M subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]

CJE1600_KpsS	<i>kpsS</i> capsule polysaccharide export protein KpsS [<i>Campylobacter jejuni</i> RM1221]	
CJE1601_KpsC	<i>kpsC</i> capsule polysaccharide export protein KpsC [<i>Campylobacter jejuni</i> RM1221]	
CJSA_1346	<i>cysC</i> adenylylsulfate kinase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]	
JJD26997_1749	putative sugar-1-phosphate nucleotidyltransferase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]	
CJJ81176_1416	class I glutamine amidotransferase, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]	
Cj1418c	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]	
Cj1419c	methyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]	
Cj1420c	methyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]	
CJSA_1247	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]	
CJ0628	lipoprotein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]	
CJ1305c	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]	
CJS3_0200	putative integral membrane protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> S3]	
CJSA_1357	putative sugar nucleotidyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> IA3902]	
Cj0055c	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]	
CJJ81176_0291	biotin sulfoxide reductase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]	
LOS region <i>C. jejuni</i> 01/51		
Gene name	Gene homolog	Annotated function

<i>wlaB</i>	CJE1272	ABC transporter, ATP-binding/permease protein [<i>Campylobacter jejuni</i> RM1221]
<i>galE</i>	CJE1273	UDP-glucose 4-epimerase [<i>Campylobacter jejuni</i> RM1221]
<i>wlaX</i>	CJE1274	polysaccharide biosynthesis protein [<i>Campylobacter jejuni</i> RM1221]
<i>waaC</i>	CJE1275	lipopolysaccharide heptosyltransferase I [<i>Campylobacter jejuni</i> RM1221]
<i>waaM</i>	CJE1276	lipid A biosynthesis lauroyl acyltransferase [<i>Campylobacter jejuni</i> RM1221]
	CJE1277	lipooligosaccharide biosynthesis glycosyltransferase, putative [<i>Campylobacter jejuni</i> RM1221]
	CJE1278	lipooligosaccharide biosynthesis galactosyltransferase, putative [<i>Campylobacter jejuni</i> RM1221]
	CJ1138	galactosyltransferase in NCTC11168 and putative galactosyltransferase in 84-25
<i>wlaN</i>		beta-1,3 galactosyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
<i>cgtB</i>		glycosyltransferase [<i>Campylobacter jejuni</i> DFVF1099]
<i>cstIII</i>		alpha-2,3-/2,8-sialyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 84-25]
<i>neuBI</i>		N-acetylneuraminic acid synthetase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 84-25]
<i>neuCI</i>		UDP-N-acetylglucosamine 2-epimerase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 84-25]
<i>neuAI</i>		two-domain bifunctional protein [beta-1,4-N-acetylgalactosaminyltransferase/CMP-Neu5Acsynthase [<i>Campylobacter</i>

		<i>jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
	CJE1281	conserved hypothetical protein [<i>Campylobacter jejuni</i> RM1221]
<i>waaV</i>	CJE1282	lipooligosaccharide biosynthesis glycosyltransferase [<i>Campylobacter jejuni</i> RM1221]
<i>waaF</i>	CJE1283	ADP-heptose--LPS heptosyltransferase II [<i>Campylobacter jejuni</i> RM1221]
<i>gmhA-1</i>	CJE1285	phosphoheptose isomerase [<i>Campylobacter jejuni</i> RM1221]
<i>hldE</i>	CJE1286	D,D-heptose 1-phosphate adenosyltransferase/7-phosphate kinase [<i>Campylobacter jejuni</i> RM1221]
<i>waaD</i>	CJE1287	ADP-L-glycero-D-mannoheptose-6-epimerase [<i>Campylobacter jejuni</i> RM1221]
	CJE1288	hydrolase, putative [<i>Campylobacter jejuni</i> RM1221]

Supplementary table 6: *C. jejuni* 01/10 prophage 1 and 2 genes, ORFs with best match in campylobacters other than *C. jejuni* RM1221 and LOS biosynthesis loci identified by BLASTx annotation tool.

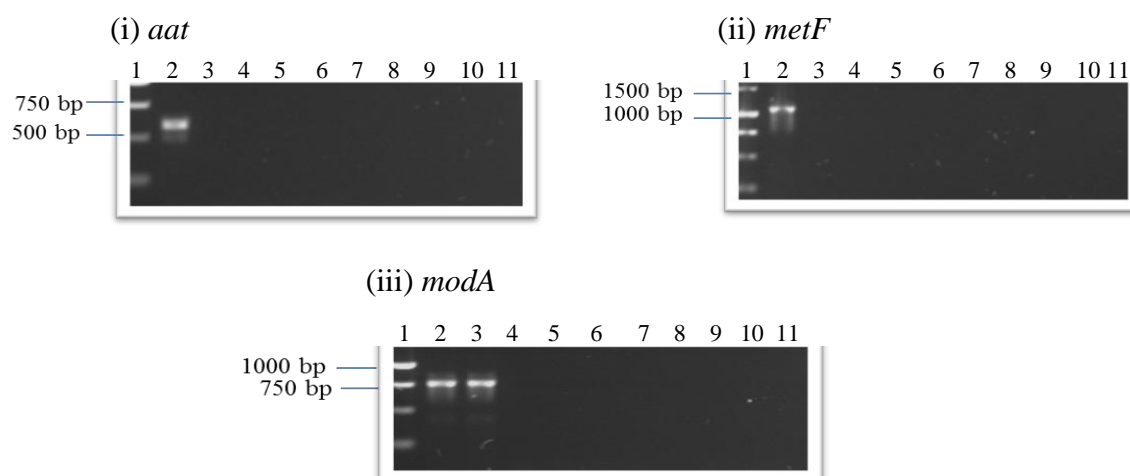
Gene homolog/ortholog		Annotated function
Prophage 1		
JJD26997_0292		phage integrase family site specific recombinase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]
Cje102_04044		hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
Cje102_04054		hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
Type I RM system		type I restriction-modification system, M subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
Anticodon nuclease		anticodon nuclease [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
RM system		restriction modification system DNA specificity subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
HsdR		type I site-specific deoxyribonuclease, HsdR family protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
Prophage 2		
CJS3_0736		phage repressor protein, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> S3]
<i>dam</i>		DNA adenine methylase [<i>Campylobacter jejuni</i> RM1221]
CJE0221		phage virion morphogenesis protein, putative [<i>Campylobacter jejuni</i> RM1221]
CJE0221		phage virion morphogenesis protein, putative [<i>Campylobacter jejuni</i> RM1221]
CJE0225		hypothetical protein [<i>Campylobacter jejuni</i> RM1221]
CJE0226		phage major tail tube protein [<i>Campylobacter jejuni</i> RM1221]
CJE0227		major tail sheath protein [<i>Campylobacter jejuni</i> RM1221]
Cje140_03187		hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 9217]
CJJ26094_0481		hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 260.94]
ICDCCJ07001_687		tail fiber protein H [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]

ICDCCJ07001_686	phage tail protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
CJS3_0717	baseplate assembly protein J, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> S3]
ICDCCJ07001_684	baseplate assembly protein V [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
Cje120_04436	hypothetical protein cje120_04436 [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 9879]
CJE0244	Mu-like prophage I protein [<i>Campylobacter jejuni</i> RM1221]
Cje104_04971	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23223]
Cje109_07168	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23263]
ICDCCJ07001_676	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
CJE0248	hypothetical protein [<i>Campylobacter jejuni</i> RM1221]
Cje160_04023	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-979]
Cje34_01110	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 87459]
ICDCCJ07001_672	prophage MuSo1, F protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
CJE0252	phage tail protein [<i>Campylobacter jejuni</i> RM1221]
ICDCCJ07001_670	tail protein D [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
Cje145_09019	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-1025]
<i>dns</i>	extracellular deoxyribonuclease [<i>Campylobacter jejuni</i> RM1221]
Cco16_02733	hypothetical protein [<i>Campylobacter coli</i> 86119]
CSS_0559	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 305]
Cje160_04285	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-979]
ICDCCJ07001_665	host-nuclease inhibitor protein Gam [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
ICDCCJ07001_663	bacteriophage DNA transposition protein B [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
CJE0269	bacteriophage DNA transposition protein B [<i>Campylobacter jejuni</i> RM1221]
CJE0270	bacteriophage DNA transposition protein A [<i>Campylobacter jejuni</i> RM1221]
Other genes	
C8J_0013	putative integral membrane protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81116]

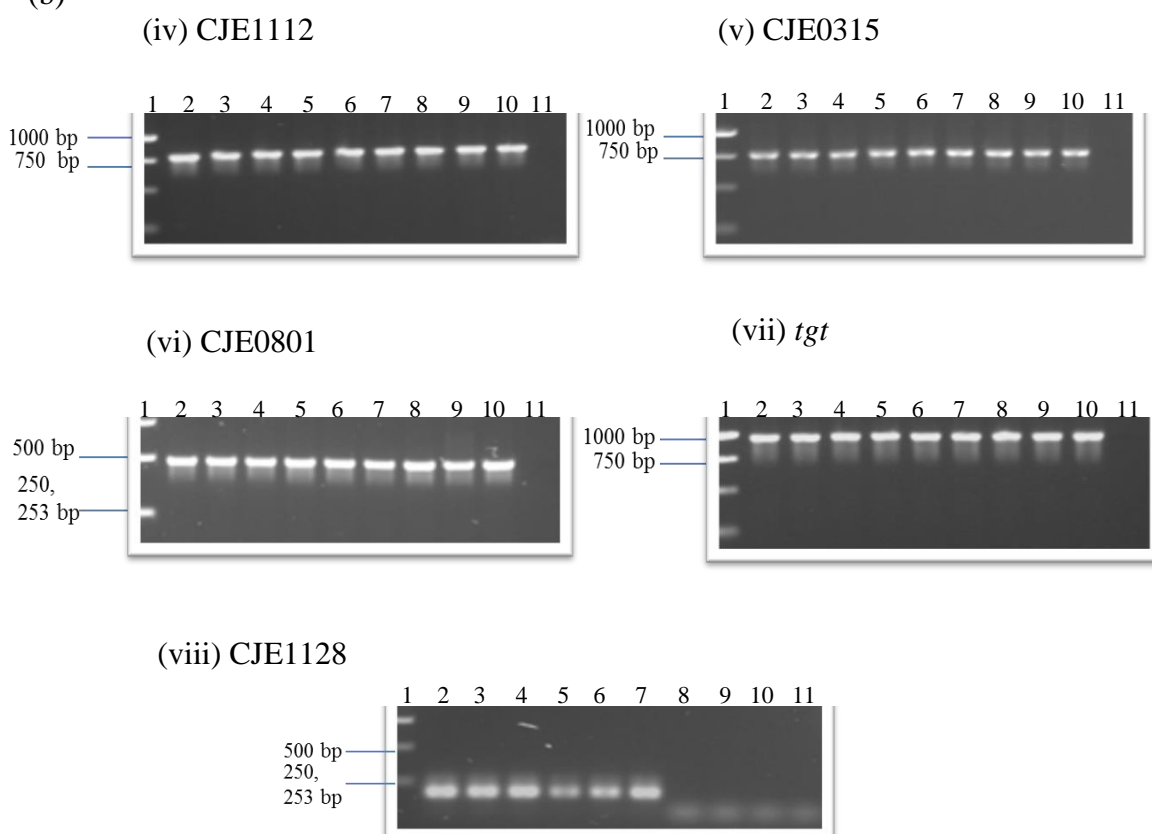
<i>cipA</i>	invasion protein CipA [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cje102_08266	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
Cje52_03044	conserved domain protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CF93-6]
CJ81176_0764	putative outer membrane protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
Cje11_06363	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 60004]
Cje102_01575	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
Cj1055c	sulfatase family protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cje135_06326	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ATCC 33560]
ICDCCJ07001_1244	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
Cje14_08577	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 53161]
Cj1324	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
Cj1355	<i>ceuE</i> enterochelin uptake periplasmic binding protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
C8J_1278	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81116]
Cje160_07455	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-979]
Cje102_05088	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
ICDCCJ07001_660	signal peptidase I [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
ICDCCJ07001_659	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001]
Cco19_02101	hypothetical protein [<i>Campylobacter coli</i> 1091]
Cje135_02178	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ATCC 33560]
Cco76_02321	hypothetical protein [<i>Campylobacter coli</i> LMG 23336]
type I RM system	type I restriction-modification system, S subunit, putative [<i>Campylobacter coli</i> 317/04]
<i>mloA</i>	MloA protein, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1336]
<i>hsdM</i>	type I restriction-modification system, M subunit [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1997-4]
Cje102_06884	hypothetical protein [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23218]
C8J_1455	transporter, putative [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81116]
<i>fkbM</i>	JJD26997_1251_FkbM family methyltransferase [<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97]

<i>tet</i>		tetracycline gene [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
LOS region 01/10		
Gene name	Gene homolog	Annotated function
<i>wlaB</i>	CJE1272	ABC transporter, ATP-binding/permease protein [<i>Campylobacter jejuni</i> RM1221]
<i>galE</i>	CJE1273	UDP-glucose 4-epimerase [<i>Campylobacter jejuni</i> RM1221]
<i>wlaX</i>	CJE1274	polysaccharide biosynthesis protein [<i>Campylobacter jejuni</i> RM1221]
<i>waaC</i>	CJE1275	lipopolysaccharide heptosyltransferase I [<i>Campylobacter jejuni</i> RM1221]
<i>waaM</i>	CJE1276	lipid A biosynthesis lauroyl acyltransferase [<i>Campylobacter jejuni</i> RM1221]
	CJE1277	lipooligosaccharide biosynthesis glycosyltransferase, putative [<i>Campylobacter jejuni</i> RM1221]
	CJ1138	galactosyltransferase in <i>C. jejuni</i> NCTC11168 and putative galactosyltransferase in <i>C. jejuni</i> 84-25
<i>cgta</i>		beta-1,4-N-acetylgalactosaminyltransferase (CgtA) [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> M1]
<i>wlaN</i>		beta-1,3 galactosyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168]
CJJ81176_1157		alpha-2,3-sialyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
ICDCCJ07001_1099		sialic acid synthase [<i>Campylobacter jejuni</i> ICDCCJ07001]
CJ81176_1159		UDP-N-acetylglucosamine 2-epimerase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
CJ81176_1160		beta-1,4-N-acetylgalactosaminyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
CJ81176_1161		CMP-Neu5Ac synthetase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
CJ81176_1162		acetyltransferase [<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81-176]
<i>waaV</i>	CJE1282	lipooligosaccharide biosynthesis glycosyltransferase [<i>Campylobacter jejuni</i> RM1221]
<i>waaF</i>	CJE1283	ADP-heptose--LPS heptosyltransferase II [<i>Campylobacter jejuni</i> RM1221]
<i>gmhA-1</i>	CJE1285	phosphoheptose isomerase [<i>Campylobacter jejuni</i> RM1221]
<i>hldE</i>	CJE1286	D,D-heptose 1-phosphate adenosyltransferase/7-phosphate kinase [<i>Campylobacter jejuni</i> RM1221]
<i>waaD</i>	CJE1287	ADP-L-glycero-D-mannoheptose-6-epimerase [<i>Campylobacter jejuni</i> RM1221]
	CJE1288	hydrolase, putative [<i>Campylobacter jejuni</i> RM1221]

(a)



(b)



Supplementary figure 1: Detection of 15 genes by PCR analysis identified by CGH.

(a) Genes present in all hyperinvasive *C. jejuni* (Group 1; table 4.1a); (i) *aat* (ii) *metF* (iii) *modA*. Other six genes (*i.e.* *hisS*, *proC*, CJE0838, CJE0669, CCOA0033 and CJE0320) were not detected in any of nine low invasive *C. jejuni* strains therefore no gel images are shown. (b) Genes highly divergent/absent from all the hyperinvasive *C. jejuni* strains (Group 2; table 4.1b); (iv) CJE1112 (v) CJE0315 (vi) CJE0801 (vii) *tgt* (viii) CJE1128. One gene, CJE0731 yielded no PCR product for

any of the nine low invasive *C. jejuni* hence the gel image is now presented. The PCR primers and expected product sizes are listed in table 2.2.1.

Each gel was loaded: Lane 1; 1kbp DNA marker (Promega, UK); lanes 2-10; *C. jejuni* 01/05, 01/08, 01/11, 01/36, C2/3, C12/11, C27/14, C69/2, C110/4; Lane 11; negative control.

Supplementary file 1: The analysis run to identify loci specific to the hyperinvasive *C. jejuni* strains.

```

program data_preproc;
uses windows;

{$R+}
{$X+}

label mm1,mm2;

const
    n_seq=10;
    tab=#09;

var
    inf,inf1,inf2:text;
    tt1:string[1];
    name:string;
    name1:string;
    tt2:string;
    i,j,k,current,al,kk,tab_name:integer;
    name_tab:array[1..45000] of string;
    tab_tab:array[1..45000] of string;
    name_name:array[1..10] of string;
    high:array[1..45000,1..10] of integer;
    status:integer;
    high1,high0,low1,low0:integer;

begin
    Writeln('Start');
    readln;

    name_name[1]:='01_10';
    name_name[2]:='01_35';
    name_name[3]:='01_04';
    name_name[4]:='01_41';

```

```

name_name[5]:='01_51';
name_name[6]:='EX114';
name_name[7]:='01_30';
name_name[8]:='01_32';
name_name[9]:='01_46';
name_name[10]:='01_39';

tab_name:=0;
current:=0;
assignfile(inf1,'c:\Alan\all_data');rewrite(inf1);{ append(inf1); }
for al:=1 to n_seq do
begin
name:='c:\ALAN\data\' + copy(name_name[al],1,length(name_name[al])) + '.dat';
writeln(name);

assignfile(inf,name);reset(inf);
readln(inf,tt2);
repeat
name:='';
repeat
read(inf,tt1);
name:=name+tt1;
until tt1=tab;
{ writeln(name,'#'); }

if tab_name=0 then begin tab_name:=1;name_tab[tab_name]:=name;current:=1;goto mm1;end else
begin
for k:=1 to tab_name do
begin

```

```

        if (copy(name,1,length(name))=copy(name_tab[k],1,length(name))) then begin
current:=k;goto mm1;end;

        end;

        tab_name:=tab_name+1;

        name_tab[tab_name]:=name;current:=tab_name;

        end;

mm1:

name1:="";

repeat
read(inf,tt1);
name1:=name1+tt1;
until tt1=tab;
tab_tab[current]:=name1;

tt2:="";

repeat
read(inf,tt1);
tt2:=tt2+tt1;
until ((tt1=tab) or (EOLN=TRUE));
status:=2;
if copy(tt2,1,1)='1' then status:=1;
if copy(tt2,1,1)='-' then status:=0;
{ val(tt2,status,kk);
writeln(tt2); }
readln(inf,tt2);

high[current,al]:=status;
until EOF(inf)=TRUE;
closefile(inf);

```

```
end;

for i:=1 to current do
begin
name:=name_tab[i]+tab+tab_tab[i]+tab;
for j:=1 to 10 do
begin
str(high[i,j]:1,name1);
if high[i,j]<>2 then name:=name+name1+tab else goto mm2;
end;
writeln(inf1,name);
mm2:
end;
closefile(inf1);
assignfile(inf2,'c:\Alan\res.txt');{rewrite(inf2);}append(inf2);
for i:=1 to current do
begin
name:=name_tab[i]+tab+tab_tab[i]+tab;
high1:=0;high0:=0;
low1:=0;low0:=0;

for j:=1 to 6 do
begin
if high[i,j]=1 then high1:=high1+1;
if high[i,j]=0 then high0:=high0+1;
end;

for j:=7 to 10 do
begin
if high[i,j]=1 then low1:=low1+1;
```



```
if high[i,j]=0 then low0:=low0+1;
end;
if (((high1=5) and (low0=3)) or ((high0=5) and (low1=3))) then begin
    write(inf2,name);
    for k:=1 to 10 do
        begin
            str(high[i,k]:1,name1);
            write(inf2,name1,tab);
        end;
    writeln(inf2);
end;
end;

closefile(inf2);

writeln('OK');
readln;
end.
```