



RESEARCH ARTICLE

Changes in labial capillary density on ascent to and descent from high altitude [version 1; referees: 2 approved]

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v1 First published: 30 Aug 2016, 5:2107 (doi: [10.12688/f1000research.7649.1](https://doi.org/10.12688/f1000research.7649.1))
 Latest published: 30 Aug 2016, 5:2107 (doi: [10.12688/f1000research.7649.1](https://doi.org/10.12688/f1000research.7649.1))

Abstract

Present knowledge of how the microcirculation is altered by prolonged exposure to hypoxia at high altitude is incomplete and modification of existing analytical techniques may improve our knowledge considerably. We set out to use a novel simplified method of measuring *in vivo* capillary density during an expedition to high altitude using a CytoCam incident dark field imaging video-microscope.

The simplified method of data capture involved recording one-second images of the mucosal surface of the inner lip to reveal data about microvasculature density in ten individuals. This was done on ascent to, and descent from, high altitude. Analysis was conducted offline by two independent investigators blinded to the participant identity, testing conditions and the imaging site. Additionally we monitored haemoglobin concentration and haematocrit data to see if we could support or refute mechanisms of altered density relating to vessel recruitment. Repeated sets of paired values were compared using Kruskal Wallis Analysis of Variance tests, whilst comparisons of values between sites was by related samples Wilcoxon Signed Rank Test. Correlation between different variables was performed using Spearman's rank correlation coefficient, and concordance between analysing investigators using intra-class correlation coefficient.

There was a significant increase in capillary density from London on ascent to high altitude; median capillaries per field of view area increased from 22.8 to 25.3 (p=0.021). There was a further increase in vessel density during the six weeks spent at altitude (25.3 to 32.5, p=0.017). Moreover, vessel density remained high on descent to Kathmandu (31.0 capillaries per field of view area), despite a significant decrease in haemoglobin concentration and haematocrit.

Using a simplified technique, we have demonstrated an increase in capillary density on early and sustained exposure to hypobaric hypoxia at thigh altitude, and that this remains elevated on descent to normoxia. The technique is simple, reliable and reproducible.

Open Peer Review

Referee Status:

| | Invited Referees | |
|--|------------------|------------|
| | 1 | 2 |
| version 1 published 30 Aug 2016 | report | report |
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How to cite this article: Gilbert-Kawai E, Coppel J, Phillip H *et al.* **Changes in labial capillary density on ascent to and descent from high altitude [version 1; referees: 2 approved]** *F1000Research* 2016, 5:2107 (doi: [10.12688/f1000research.7649.1](https://doi.org/10.12688/f1000research.7649.1))

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Grant information: Xtreme Everest 2 was supported by the Royal Free Hospital NHS Trust Charity, the Special Trustees of University College London Hospital NHS Foundation Trust, the Southampton University Hospital Charity, the UCL Institute of Sports Exercise and Health, The London Clinic, University College London, University of Southampton, Duke University Medical School, the United Kingdom Intensive Care Society, the National Institute of Academic Anaesthesia, the Rhinology and Laryngology Research Fund, The Physiological Society, Smiths Medical, Deltex Medical, Atlantic Customer Solutions and the Xtreme Everest 2 volunteer participants who trekked to Everest Base Camp. Some of this work was undertaken at University College London Hospital- University College London Biomedical Research Centre, which received a proportion of funding from the United Kingdom Department of Health's National Institute for Health Research Biomedical Research Centers funding scheme. Some of this work was undertaken at University Hospital Southampton-University of Southampton Respiratory Biomedical Research Unit, which received a proportion of funding from the United Kingdom Department of Health's National Institute for Health Research Biomedical Research Units funding scheme.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: Braedius Medical, a company owned by a relative of Can Ince, has developed and designed a hand held microscope called CytoCam-IDF imaging. Can Ince has no financial relation with Braedius Medical of any sort; he never owned shares, or received consultancy or speaker fees from Braedius Medical.

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List of Abbreviations

| | |
|------------------|------------------------------|
| EBC | Everest Base Camp |
| FOV | Field of view area |
| [Hb] | Haemoglobin concentration |
| Hct | Haematocrit |
| IDF | Incident Dark Field |
| KTM | Kathmandu |
| LON | London |
| SpO ₂ | Peripheral oxygen saturation |

Introduction

The physiological processes involved in acclimatisation to high altitude attempt to maintain adequate oxygen delivery as the partial pressure of oxygen decreases. Traditionally, research has concentrated on global haemodynamics and the macrocirculation, variables such as cardiac output¹, oxygen saturations² and haemoglobin concentration [Hb]³. Far fewer studies have focused on the microcirculation, which regulates blood flow to match micro-regional oxygen demand. Disruption of microvascular blood flow could explain a failure of acclimatisation in some individuals as well as the well-documented exercise limitation that occurs at altitude despite normalisation of systemic oxygen delivery⁴. The precise role of the microcirculation in acclimatisation to hypoxia, however, remains unclear.

Teleological reasoning would suggest that increasing capillary density could provide a means to augment oxygen flux and tissue oxygenation through a reduction in the inter-capillary distance⁵. Whilst plausible, data on this theory remains contradictory, though this may in part relate to the dissimilar tissues observed. In human skeletal muscle biopsy samples previously exposed to hypobaric hypoxia, no evidence of neovascularisation has been demonstrated⁶⁻⁹. Interestingly, in each instance whereby the capillary density was initially thought to increase, no change in the capillary-to-fibre ratio was observed. The perceived rise in capillary density were therefore interpreted as being secondary occurrences in response to a reduction in skeletal muscle mass. Conversely, an increase in the density of sublingual microcirculatory vessels to >25 μm was demonstrated on ascent to high altitude^{10,11}, a response that was further amplified after prolonged exposure to hypoxia¹⁰. In this instance, what remains to be determined is whether the observed changes in vessel density are due to microvascular recruitment secondary to increased blood viscosity (and thus quickly reversible), or neovascularization (which is likely to be sustained). Moreover, the question of what happens to vessel density following re-exposure to normoxia remains to be elucidated.

We therefore piloted a novel modification of a previously described technique for calculating changes in capillary density^{12,13} on ten individuals, to see if we could firstly support or refute previous findings on ascent to high altitude, and secondly see if the changes

observed persist on descent. Additionally we monitored haemoglobin concentration and haematocrit data to see if we could support or refute mechanisms of altered density relating to vessel recruitment.

Methods

The study was undertaken as part of the Xtreme Everest 2 research expedition (XE2)¹⁴. The study design, risk management plan and protocol were approved (in accordance with the declaration of Helsinki) both by the University College London Committee and the Ethics of Non-National Health Service Human Research, and the Nepal Health Research Council (Reg no. 139/2012). Written consent was obtained from all participants. Baseline images of the labial capillaries were initially obtained from ten individuals in London (LON) (35m) in December 2012 and January 2013. Sequential images were taken after an 11 day ascent to Everest Base Camp (EBC-early) (5300m), then after 6 weeks residence at Everest Base Camp (EBC-late), and finally on descent, over 5 days, to Kathmandu (KTM) (1300m) in May 2013.

Images were obtained using a CytoCam-IDF video microscope (Braedius, Medical BV, Netherlands). This new device is based on the principle of Incident Dark Field (IDF) imaging, which uses polarized green light (wavelength 548nm) produced from LEDs to visualize, in real time, the sublingual microvasculature. Its high resolution imaging sensor (14 Mpixel) allows for a 50% increase in optical resolution (300 lines/mm) compared to previous Sidestream Dark Field imaging devices, and it generates a far larger field of view. With the participant lying in the supine position having rested for a minimum of 10 minutes, the CytoCam-IDF device's probe was introduced into their mouth and placed on the mucosal surface of the inner lip. Once a suitable image was visualised on the screen of the CytoCam-IDF monitor, (Figure 1), 1 second of digital video footage was recorded.

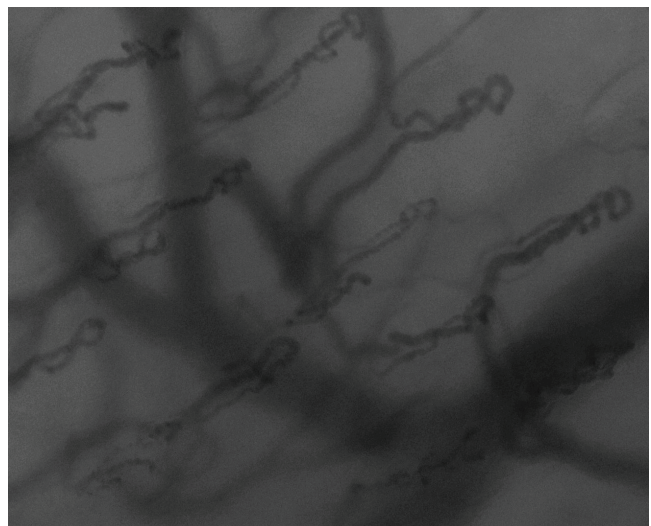


Figure 1. An example of a labial capillary density image.

This process was conducted on all four lip quadrants (right upper lip, left upper lip, right lower lip, left lower lip), and at each quadrant four separate videos were acquired. Two trained investigators (EGK, PH) obtained all the data.

To determine capillary density, analysis was conducted offline by two independent investigators blinded to the participant identity, the testing conditions and the imaging site (EGK, JC). Using the company's own video software (CytoCamTools V1, Braedius, Netherlands), a single still frame was projected on the computer screen and the number of capillary loops per image frame was counted manually. Partly visualised capillaries were included if the observer was assured that the vessel was a capillary due to its morphology. Subsequently, the mean capillary density was calculated from the four images obtained in each lip quadrant, and from these four results, the mean total lip density obtained. Capillary density was defined as the number of capillaries counted per field of view area (FOV), which equates to 1.79 mm². The haemoglobin concentration ([Hb]) (Hemocue AB, Hemocue, Sweden) and haematocrit (Hct) (Sigma 1–14 microcentrifuge, Sigma, Germany) were obtained from whole blood samples, and peripheral arterial oxygen saturation (SpO₂) measured (Nonin Onyx 9500, Nonin Medical Inc, Minnesota, USA) on the same days as microcirculatory imaging was performed.

Statistical analysis

As data were not normally distributed, they were described by median and interquartile range. Repeated sets of paired values were compared using Kruskal Wallis ANOVA, whilst comparisons of values between LON baseline and other sites was by Wilcoxon Signed Rank Test. Correlation between different variables was performed using Spearman's rank correlation coefficient, and concordance between analysing investigators using intra-class correlation coefficient. All statistical analysis was undertaken on SPSS version 21 (SPSS Inc., Chicago, IL, USA), and a P value of <0.05 was taken to indicate statistical significance.

Results

Dataset 1. IDF values

<http://dx.doi.org/10.5256/f1000research.7649.d134021>

The number of loop capillaries seen per field of view

Dataset 2. Physiological values

<http://dx.doi.org/10.5256/f1000research.7649.d134022>

The haemoglobin, haematocrit and peripheral oxygen saturations at each laboratory.

CytoCam-IDF imaging was conducted on all ten individuals on the first two occasions, however only eight individuals had data captured on descent. No problems were encountered with the device or image acquisition. Mean laboratory barometric pressure and mean temperature for each location is shown in [Table 1](#).

Table 1. Mean (standard deviation) atmospheric pressure and temperature at each laboratory.

| Site | Atmospheric Pressure (kPa) | Temperature (°C) | Humidity (%) |
|-------------------|----------------------------|------------------|--------------|
| London | 100.6 (0.2) | 16.9 (1.8) | 35.4 (6.5) |
| Everest Base Camp | 53.0 (0.2) | 12.9 (8.2) | 37.8 (17.5) |
| Kathmandu | 86.8 (0.4) | 23.8 (3.4) | 47.4 (15.7) |

Changes in labial capillary density are shown in [Figure 2](#). Compared with LON (median 22.8 capillaries per field of view area (20.7–26.8)), capillary density was significantly increased at EBC-early 25.3 (24.5–30.6; p=0.021), EBC-late 32.5, (28.4–36.63; p=0.012), and on descent in KTM 31.0 (24.0–35.13; P=0.017). Between EBC-early and EBC-late, capillary density increased significantly (p=0.017), however there was no significant decline in density between EBC-late and KTM (p = 0.069).

Changes in [Hb], Hct and SpO₂ at each site are shown in [Table 2](#). There was a significant increase in [Hb] between LON and EBC-early (p=0.007), and EBC-early and EBC-late (p=0.011), and a decrease between EBC-late and KTM (p=0.008). There was also a significant increase in Hct between LON and EBC-early (p=0.007), and EBC-late and KTM (p=0.012), but no significant change between EBC-early and EBC-late (p=0.191). Between the sites on ascent, the increase in vessel density demonstrated an inverse relationship with the SpO₂, however at each altitude there was no correlation between vessel density and [Hb], Hct or SpO₂.

To assess whether an image capture time of 1 second was indicative of that captured over longer periods of time, we obtained 30 seconds of footage from four individuals at two different locations. From this we randomly selected one frame per five seconds of footage, and counted the number of capillaries per field of view area. The values of these may be seen in [Table 3](#), as too can the mean and standard deviations for each set of frames, the latter of which demonstrates a highest value of only 0.52 capillaries per field of view area.

Discussion

This study demonstrates for the first time, persistence of *in vivo* sublingual microvascular density increase on re-exposure to normoxia after a prolonged period of hypobaric hypoxia at high altitude. We utilised an infrequently used imaging and analysis technique that we had purposefully adapted to suit our needs, and found our data aligned with previously published work on blood vessel density at altitude¹⁰.

Using the data obtained from corresponding blood samples, it is possible to speculate on the adaptive processes occurring at each measurement point. As previously described, we observed a significant rise in [Hb] (14.5 g/dl to 16.4g/dl; p=0.007) and Hct (44%

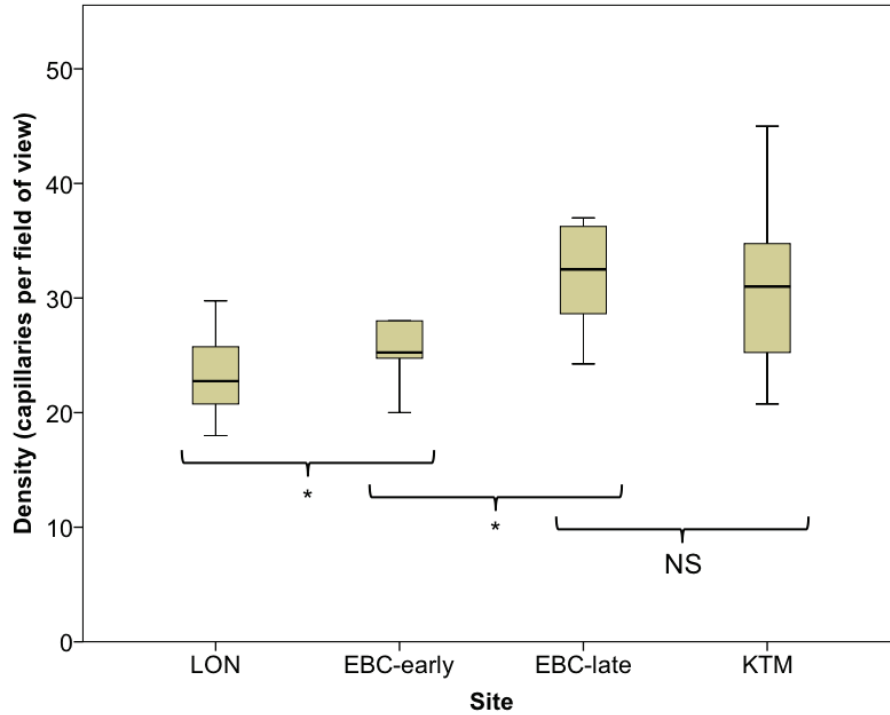


Figure 2. Box-whisker plots of labial capillary density on ascent from London to Everest Base Camp, after six weeks at Everest Base Camp, and on descent to Kathmandu.

Table 2. Median (interquartile range) haemoglobin concentration, haematocrit and peripheral oxygen saturations at each measurement point.

| Site | Hemoglobin concentration (g/dl) | Hematocrit (%) | Oxygen saturation (%) |
|---------------------------|---------------------------------|------------------|-----------------------|
| London | 14.5 (13.5–15.3) | 44.0 (41.5–47.5) | 99 (98–100) |
| Everest Base Camp - early | 16.4 (15.7–17.2) | 52.0 (51.5–56.3) | 81 (79–86) |
| Everest Base Camp - late | 18.3 (17.1–19.7) | 56.0 (51.0–61.5) | 88 (85–91) |
| Kathmandu | 16.0 (14.0–17.0) | 50.0 (46.5–55.0) | 99 (99–100) |

Table 3. Number of capillaries counted per field of view area from one randomly selected frame per five seconds of footage, with mean and standard deviation demonstrated.

| Code | Site | Frame 1 (0–5 s) | Frame 2 (5–10 s) | Frame 3 (10–15 s) | Frame 4 (15–20 s) | Frame 5 (20–25 s) | Frame 6 (25–30 s) | Mean | SD |
|------|------|-----------------|------------------|-------------------|-------------------|-------------------|-------------------|------|------|
| 1 | LON | 30 | 30 | 30 | 30 | 30 | 30 | 30.0 | 0.00 |
| 2 | LON | 24 | 25 | 24 | 24 | 24 | 25 | 24.3 | 0.52 |
| 3 | KTM | 27 | 27 | 27 | 27 | 26 | 27 | 26.8 | 0.41 |
| 4 | KTM | 22 | 23 | 23 | 23 | 23 | 22 | 22.7 | 0.52 |

s = seconds; SD = standard deviation

to 52%; $p=007$) on ascent to altitude. Whilst this polycythaemia increases arterial oxygen content, blood viscosity also rises, altering its rheology. Under normal physiological conditions, a considerable proportion of the microcirculation is thought to be 'unrecruited', acting as a reservoir for times of increased metabolic needs¹⁵. As Hct rises, these reserve vessels are recruited, and microvascular density increases, along with functional capillary density¹⁵⁻¹⁸. Thus a secondary benefit to increased Hct is achieved; a reduction in the diffusion distance from capillaries to mitochondria. Importantly however, it should be noted that in normal capillary Hct is generally 50% less than systemic Hct owing to the streamlined blood flow in narrow capillaries¹⁹. The effect of hypoxia on this association is unknown. Whether or not neovascularisation had occurred on arrival at high altitude is difficult to say, although due to the short time between measurement points the chances of this being the case are low²⁰.

After 6 weeks spent at altitude, a further, and far greater, increase in vessel density was apparent; EBC-early 25.3 capillaries per field of view area, EBC-late 32.5 ($p=0.017$). Over the same time period, [Hb] had significantly risen, whilst Hct had not. As Hct is a more reliable indicator of viscosity between the two variables²¹, it seems unlikely that further recruitment of the microvasculature had occurred, yet it is plausible that neovascularisation had. Increased levels of vascular endothelial growth factor (VEGF) have been detected in subjects ascending to high altitude²²; its role in angiogenesis perhaps explaining the observed rise in microvascular density¹⁰. Such adaptations lead to improved tissue oxygenation by a reduction of the inter-capillary distance, whilst maintaining a sufficiently low, and thus fluid Hct to permit flow of red blood cells in the microvasculature.

On descent to a lower altitude (KTM) there was no significant fall in microvascular density when compared to EBC-late ($p = 0.069$), however, a much greater number of vessels (36% increase) was evident when compared with baseline testing in LON. Whilst vessel density was thus unaltered on descent, over the same time point [Hb] and Hct values significantly declined ($p=0.012$). When compared to the original LON values, Hct on descent to KTM was significantly higher (44.0% and 50.0% respectively ($p=0.011$)), however, [Hb] was not (14.5 and 16.0g/dl ($p=0.052$)). Teasing apart the relative contributions of vessel recruitment and neovascularisation to the observed changes in sublingual microcirculatory density is challenging. Whilst the failure of vessel density to return to baseline after descent suggests some neovascularisation, neither [Hb] nor Hct had normalised at the time of the final readings so a raised blood viscosity could perhaps be maintaining a heightened level of capillary recruitment. A combination of the two processes would make sense as continually increasing [Hb] to improve oxygen delivery would eventually be counter productive. Indeed in Tibetans, who have been exposed to environmental hypoxia for many generations, there is a clear reduction in [Hb] compared to populations who have been exposed to these conditions for less time²³⁻²⁶. This suggests Tibetans utilize alternative long-term strategies for chronic adaptation to hypobaric hypoxia, ones that do not rely on maintaining a high [Hb]. It is plausible that one such means would be to increase their capillary density.

Technique and analysis

The use of the described methodology was also novel. A similar technique has been used twice previously; once in the assessment of coronary artery disease in diabetes¹³ and the other in a study investigating hypertension and rarefaction during treatment with Telatinib¹². In these instances, data capture involved recording sublingual images for 1 minute¹³ or 30 seconds¹² per quadrant, however we altered this time period by using an extremely short capture phase for data acquisition (< 1 second). Crucially, this allowed us to readily obtain snap shot images to reveal data about microvasculature density, whilst avoiding concerns surrounding probe and patient movement, in addition to issues relating to pressure artefact. Analysis was rapid, simple and reproducible; in this study it had an observer mean intra-class correlation coefficient of 0.91 (95% CI 0.84 – 0.96). Previously no difference in capillary density was observed in ten individuals between lip quadrants, and the reproducibility of the technique to determine capillary density was moderate to high with a coefficient of variation of 4.6%¹². Of note, the technique does not allow assessment of microvascular flow, nor does it yield information on heterogeneity of microvascular blood flow, however, we propose it to be a robust method for the assessment of labial vessel density that could be conducted after only a short user training period.

Study limitations

The small number of participants used in this study could be considered a study limitation. As we were both employing a newly-adapted data acquisition technique, and using a novel device at altitude, no power calculation was performed. This therefore increases the risk of a type 2 error. Other limiting factors include the environmental considerations associated with high altitude research in a remote field environment. These include fluctuations in laboratory temperature, humidity (Table 1) and participant hydration status, all factors that may alter microvascular blood flow and density. Attempts were made to limit these potential confounding factors by performing CytoCam-IDF imaging at the same time of day in heated purpose-built laboratories, and encouraging participants to maintain a good state of hydration. Previous studies at altitude have also raised concerns over the development of tissue oedema^{10,11} that can occur on ascent to altitude²⁷. Whilst this could potentially reduce image quality and lead to false measurements of flow and density, our IDF camera provided us with a depth of focus reading, thus allowing us to confirm that we were recording at the same depth under the tongue on each time point. Finally, we have discussed alterations in capillary or vessel density. It is important however to clarify this nomenclature. IDF imaging cannot image blood vessels directly but rather uses the fact that polarized green light is optimally absorbed by red blood cells within the microvasculature regardless of oxygenation status. Absorption of light by haemoglobin, but not by surrounding tissues, therefore creates a distinct contrast of dark and light colour respectively, and red blood cells moving through the mucosal microcirculation thus appear as dark globules moving along the axis of flow. All vessels visualized are therefore only seen if they contain erythrocytes. The variables measured by IDF imaging (and its precursor SDF imaging) include a measure of total vessel density (TVD) and perfused vessel density (PVD)²⁸.

A distinction is made between the two depending on the speed of red blood cell flow within the observed vessels. TVD includes vessels which contain erythrocytes flowing at any velocity (or even at standstill), whilst PVD only includes vessels with continuously moving erythrocytes. As we cannot measure erythrocyte velocity with this simplified method, our observations therefore describe the TVD.

Conclusions

This study demonstrated an increase in sublingual microvascular vessel density on early and sustained exposure to hypobaric hypoxia; and, for the first time, that no significant change in vessel density occurred on immediate descent. The technique used to capture the images provided a rapid and reliable means for assessing changes in vessel density, and could be applied in future studies of microcirculatory vessel density. Further research in this area may allow a more complete comprehension of the multidimensional response to sustained hypoxia that occurs during pathophysiological situations.

Data availability

F1000Research: Dataset 1. IDF values, [10.5256/f1000research.7649.d134021](https://doi.org/10.5256/f1000research.7649.d134021)²⁹

F1000Research: Dataset 2. Physiological values, [10.5256/f1000research.7649.d134022](https://doi.org/10.5256/f1000research.7649.d134022)³⁰

Author contributions

E G-K: design of study, collection of data, analysis of data, writing manuscript

JC: collection of data, analysis of data, writing manuscript

PH: analysis of data, writing manuscript

MG: design of study, writing manuscript

CI: design of study, writing manuscript

DM: design of study, analysis of data, writing manuscript

All authors have seen and agreed to the final content of the manuscript

Competing interests

Braedius Medical, a company owned by a relative of Can Ince, has developed and designed a hand held microscope called CytoCam-IDF imaging. Can Ince has no financial relation with Braedius Medical of any sort; he never owned shares, or received consultancy or speaker fees from Braedius Medical.

Grant information

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The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgements

Xtreme Everest 2 is a research project coordinated by the Caudwell Xtreme Everest Hypoxia Research Consortium, collaboration between the UCL Centre for Altitude, Space, and Extreme Environment Medicine, the Centre for Human Integrative Physiology at the University of Southampton and the Duke University Medical Centre. Membership, roles and responsibilities of the Xtreme Everest 2 Research Group can be found at www.xtreme-everest.co.uk/team.

References

- Wagner PD: **Reduced maximal cardiac output at altitude—mechanisms and significance.** *Respir Physiol.* 2000; **120**(1): 1–11.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Stoneham MD, Pethybridge RJ: **Acclimatization to altitude: effects on arterial oxygen saturation and pulse rate during prolonged exercise at altitude.** *J R Nav Med Serv.* 1993; **79**(1): 3–9.
[PubMed Abstract](#)
- Pugh LG: **Blood volume and haemoglobin concentration at altitudes above 18,000 ft. (5500 m).** *J Physiol.* 1964; **170**(2): 344–354.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Levett DZ, Fernandez BO, Riley HL, *et al.*: **The role of nitrogen oxides in human adaptation to hypoxia.** *Sci Rep.* 2011; **1**: 109.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Hochachka PW, Stanley C, Merkt J, *et al.*: **Metabolic meaning of elevated levels of oxidative enzymes in high altitude adapted animals: an interpretive hypothesis.** *Respir Physiol.* 1983; **52**(3): 303–313.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Hoppeler H, Kleinert E, Schlegel C, *et al.*: **Morphological adaptations of human skeletal muscle to chronic hypoxia.** *Int J Sports Med.* 1990; **11**(Suppl 1): S3–S9.
[PubMed Abstract](#) | [Publisher Full Text](#)

7. MacDougall JD, Green HJ, Sutton JR, *et al.*: **Operation Everest II: structural adaptations in skeletal muscle in response to extreme simulated altitude.** *Acta Physiol Scand.* 1991; **142**(3): 421–427.
[PubMed Abstract](#) | [Publisher Full Text](#)
8. Lundby C, Pilegaard H, Andersen JL, *et al.*: **Acclimatization to 4100 m does not change capillary density or mRNA expression of potential angiogenesis regulatory factors in human skeletal muscle.** *J Exp Biol.* 2004; **207**(pt 22): 3865–3871.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. Mizuno M, Savard GK, Areskog NH, *et al.*: **Skeletal muscle adaptations to prolonged exposure to extreme altitude: a role of physical activity?** *High Alt Med Biol.* 2008; **9**(4): 311–317.
[PubMed Abstract](#) | [Publisher Full Text](#)
10. Martin DS, Goedhart P, Vercueil A, *et al.*: **Changes in sublingual microcirculatory flow index and vessel density on ascent to altitude.** *Exp Physiol.* 2010; **95**(8): 880–891.
[PubMed Abstract](#) | [Publisher Full Text](#)
11. Martin DS, Ince C, Goedhart P, *et al.*: **Abnormal blood flow in the sublingual microcirculation at high altitude.** *Eur J Appl Physiol.* 2009; **106**(3): 473–478.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. Steeghs N, Gelderblom H, Roodt JO, *et al.*: **Hypertension and rarefaction during treatment with telatinib, a small molecule angiogenesis inhibitor.** *Clin Cancer Res.* 2008; **14**(11): 3470–3476.
[PubMed Abstract](#) | [Publisher Full Text](#)
13. Djaberri R, Schuijff JD, de Koning EJ, *et al.*: **Non-invasive assessment of microcirculation by sidestream dark field imaging as a marker of coronary artery disease in diabetes.** *Diab Vasc Dis Res.* 2013; **10**(2): 123–134.
[PubMed Abstract](#) | [Publisher Full Text](#)
14. Martin DS, Gilbert-Kawai E, Levett DZh, *et al.*: **Xtreme Everest 2: unlocking the secrets of the Sherpa phenotype?** *Extrem Physiol Med.* 2013; **2**(1): 30.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Parthasarathi K, Lipowsky HH: **Capillary recruitment in response to tissue hypoxia and its dependence on red blood cell deformability.** *Am J Physiol.* 1999; **277**(6 Pt 2): H2145–H2157.
[PubMed Abstract](#)
16. Genzel-Boroviczeny O, Christ F, Glas V: **Blood transfusion increases functional capillary density in the skin of anemic preterm infants.** *Pediatr Res.* 2004; **56**(5): 751–755.
[PubMed Abstract](#) | [Publisher Full Text](#)
17. Sakr Y, Chierigo M, Piagnerelli M, *et al.*: **Microvascular response to red blood cell transfusion in patients with severe sepsis.** *Crit Care Med.* 2007; **35**(7): 1639–1644.
[PubMed Abstract](#) | [Publisher Full Text](#)
18. Yuruk K, Almac E, Bezemer R, *et al.*: **Blood transfusions recruit the microcirculation during cardiac surgery.** *Transfusion.* 2011; **51**(5): 961–967.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Duling BR, Klitzman B: **Local control of microvascular function: role in tissue oxygen supply.** *Annu Rev Physiol.* 1980; **42**: 373–382.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. Lindeboom JA, Mathura KR, Aartman IH, *et al.*: **Influence of the application of platelet-enriched plasma in oral mucosal wound healing.** *Clin Oral Implants Res.* 2007; **18**(1): 133–139.
[PubMed Abstract](#) | [Publisher Full Text](#)
21. Pries AR, Secomb TW, Gaehtgens P: **Biophysical aspects of blood flow in the microvasculature.** *Cardiovasc Res.* 1996; **32**(4): 654–667.
[PubMed Abstract](#) | [Publisher Full Text](#)
22. Walter R, Maggiorini M, Scherrer U, *et al.*: **Effects of high-altitude exposure on vascular endothelial growth factor levels in man.** *Eur J Appl Physiol.* 2001; **85**(1–2): 113–117.
[PubMed Abstract](#) | [Publisher Full Text](#)
23. Adams WH, Strang LJ: **Hemoglobin levels in persons of tibetan ancestry living at high altitude.** *Proc Soc Exp Biol Med.* 1975; **149**(4): 1036–1039.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. Beall CM, Reichsman AB: **Hemoglobin levels in a Himalayan high altitude population.** *Am J Phys Anthropol.* 1984; **63**(3): 301–306.
[PubMed Abstract](#) | [Publisher Full Text](#)
25. Samaja M, Veicsteinas A, Cerretelli P: **Oxygen affinity of blood in altitude Sherpas.** *J Appl Physiol Respir Environ Exerc Physiol.* 1979; **47**(2): 337–341.
[PubMed Abstract](#)
26. Wu T, Wang X, Wei C, *et al.*: **Hemoglobin levels in Qinghai-Tibet: different effects of gender for Tibetans vs. Han.** *J Appl Physiol (1985).* 2005; **98**(2): 598–604.
[PubMed Abstract](#) | [Publisher Full Text](#)
27. Maggiorini M, Böhler B, Walter M, *et al.*: **Prevalence of acute mountain sickness in the Swiss Alps.** *BMJ.* 1990; **301**(6756): 853–855.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
28. De Backer D, Hollenberg S, Boerma C, *et al.*: **How to evaluate the microcirculation: report of a round table conference.** *Crit Care.* 2007; **11**(5): R101.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
29. Gilbert-Kawai E, Coppel J, Phillip H, *et al.*: **Dataset 1 in: Changes in labial capillary density on ascent to and descent from high altitude.** *F1000Research.* 2016.
[Data Source](#)
30. Gilbert-Kawai E, Coppel J, Phillip H, *et al.*: **Dataset 2 in: Changes in labial capillary density on ascent to and descent from high altitude.** *F1000Research.* 2016.
[Data Source](#)

Open Peer Review

Current Referee Status:



Version 1

Referee Report 28 September 2016

doi:10.5256/f1000research.8238.r15955



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The authors report on the use of a relatively novel method to determine capillary density during acute and chronic exposure to hypobaric hypoxia. Previously reported results of an increase in capillary density during acclimatization are confirmed, but it is for the first time demonstrated that upon immediate descent some changes persist.

Overall, this is a well-conducted study that adds to the current knowledge base of microcirculatory changes during adaptation to hypobaric hypoxia, and provides an innovative approach to vessel density measurement.

Specific comments:

Introduction

In the introduction the absence of neovascularization in muscle samples during adaptation to high altitude is discussed. It is however also known that mitochondrial density increases during this process (PMID 26339730). The authors may want to include this in the discussion about capillary-to-fibre ratio, the mitochondria arguably representing the circulation's endpoint.

Methods

The KTM time point seems difficult to me for use as a “post-hypoxia” measurement, since it is not comparable to the LON baseline. This is not a severe limitation since “immediate descent” was one of the study endpoints rather than a comparison to the initial (LON) baseline. I still suggest mentioning this in the limitations.

Microcirculation capture time is very short (1 sec). I am not sure if the analysis in four subjects given in table 3 is sufficient support for the use of much shorter capture intervals than recommended by current consensus guidelines. This could be discussed in more detail in the limitations.

Hb and Hematocrit values, as well as SpO₂ were determined “on the same day as microcirculatory imaging was performed”. Given the influence of hydration status changes throughout a day, especially in a mountain setting, either specify the time frame more clearly or mention this time relation in the limitations.

Results

Figure 2: It is unclear to me what the results of the Kruskal-Wallis ANOVA described in the methods section were for vessel density measurements, consider mentioning these in the results section. Further, are the P values given in the text references to Wilcoxon Signed Rank tests as mentioned in the methods section? Consider describing these tests as post-hoc analysis for the non-parametric ANOVA. And if so, was a correction for multiple testing applied (Bonferroni, or one of the newer algorithms)? Please describe this more clearly.

It is mentioned that at each altitude there was no correlation between vessel density and Hb, Hct or SpO₂. In the methods section it is stated that Spearman's rank correlation coefficient was calculated. Consider adding correlation coefficients and P values to the results section to provide the readers some insight into the data.

In addition to Hb, Hct and SpO₂ given in Table 2, it would be of interest to know more about the subjects' physiologic parameters, such as blood pressure, cardiac output and even RV/RA – including a discussion about the changes in macrocirculation in relation to the changes in microcirculation.

Discussion

Given borderline significance ($p=0.069$) for a decrease in vessel density from EBC-late to KTM and the lack of a power calculation, as well as the lack of real comparability to the initial (LON) baseline it is somewhat difficult to argue that there really was no change between EBC-late and KTM. This is discussed in the discussion to some extent, but it should be stated more clearly that a verification of these results is needed in the future.

The method for vessel density estimation used in this study differs from the effective calculation of total vessel density (TVD), being the total length of small vessels divided by the surface area covered in the recording, that is recommended by current consensus guidelines. Please discuss the relationship between vessel density as measured in this study with vessel density as assessed by TVD, and potential confounders.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.

Referee Report 14 September 2016

doi:[10.5256/f1000research.8238.r16269](https://doi.org/10.5256/f1000research.8238.r16269)



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Edward Gilbert-Kawai and colleagues involved in the Xtreme Everest 2 study report further work on the effects of altitude and hypoxaemia on the microcirculation. They describe a technique for rapidly measuring microcirculatory capillary density, using a method of counting the number of capillary loops in the buccal mucosa of healthy volunteers at sea level, progressive altitude acclimatisation and descent.

They also report systemic haemoglobin and haematocrit values in an attempt to elucidate whether changes in microcirculatory parameters are paralleled by changes in blood viscosity. Their results show, in line with previous work, that microcirculatory vessel density increases after prolonged exposure to hypoxic conditions. However, they also show that there is some persistence of this effect on partial descent. The authors pose a question as to whether this increased vessel density is due to neo-vascularisation or recruitment of existing capillaries secondary to an increase in haematocrit.

This is generally a scientifically sound paper and a well written manuscript. I have several comments for the authors:

Major Comments

- It strikes me that the authors are trying to achieve two things in this paper. Firstly, to confirm that there is microcirculatory adaptation to altitude and hypoxaemia and to provide more information regarding the mechanisms surrounding this and secondly to describe a method of rapidly assessing vessel density. Both subjects are not quite covered in enough detail to do either justice.
- With regard to the assessment method, this has been described previously in both buccal mucosa but also in other areas such as the nail bed. It has the advantage of being quick to perform but only provides information on one variable – number of capillary loops. The authors state and assume that this measure equates to Total Vessel Density (TVD) but it would be very useful to confirm this by actually measuring capillary TVD using sublingual IDF video capture and analysis. More importantly, the described method provides no measure of microcirculatory flow. This is crucial when discussing the impact of capillary haematocrit and viscosity on microcirculatory performance. Personally I think the paper would have been improved had the investigators used sublingual images and analyzed these to produce data on a fuller range of flow and density parameters.
- Although the authors give two possible mechanisms for the observed increase in capillary density, I would be interested to know if these changes could possibly be accounted for by an increase in systemic haemodynamic parameters, such as cardiac output. Unless there is a loss of haemodynamic coherence one would expect that micro would follow macro. Measurement of these values using a non invasive technique such as trans thoracic echocardiography would have been interesting and added value to the paper.
- The authors discuss that systemic haematocrit and haemoglobin do not necessarily equate with capillary haemoglobin. This is a crucial point and one often observes that “shocked” capillaries have a visibly lower number of erythrocytes even in the presence of a reasonably preserved systemic Hb level, especially post resuscitation. Being able to quantify this difference would allow us to answer crucial questions but is not possible with current analysis methods using this technology.
- I would be very interested to know how long this microcirculatory adaptation lasts. Did the authors consider repeating the measures after return to sea level and after a more prolonged period – e.g. one month?

Minor Comments

- Southampton is spelt incorrectly in several places within the list of authors
- Citing a validation paper of the IDF camera (e.g. Hutchings S, Watts S, Kirkman E. The Cytocam video microscope. A new method for visualising the microcirculation using Incident Dark Field

technology. Clin Hemorheol Microcirc. IOS Press; 2015 Oct 16;62(3):261–71) would be useful as the authors have stated that it is a new device.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Competing Interests: No competing interests were disclosed.
