

1 **Title:** Hyporheic invertebrates as bioindicators of ecological health in temporary rivers:  
2 a meta-analysis

3

4 **Short title:** Hyporheic bioindicators in temporary rivers

5

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18 **Abstract**

19 Worldwide, many rivers cease flow and dry either naturally or owing to human  
20 activities such as water extraction. However, even when surface water is absent, diverse  
21 assemblages of aquatic invertebrates inhabit the saturated sediments below the river bed  
22 (hyporheic zone). In the absence of surface water or flow, biota of this zone may be  
23 sampled as an alternative to surface water-based ecological assessments. The potential  
24 of hyporheic invertebrates as ecological indicators of river health, however, is largely  
25 unexplored. We analysed hyporheic taxa lists from the international literature on  
26 temporary rivers to assess compositional similarity among broad-scale regions and  
27 sampling conditions, including the presence or absence of surface waters and flow, and  
28 the regional effect of hydrological phase (dry channel, non-flowing waters, surface  
29 flow) on richness. We hypothesized that if consistent patterns were found, then effects  
30 of human disturbances in temporary rivers may be assessable using hyporheic  
31 bioindicators. Assemblages differed geographically and by climate, but hydrological  
32 phase did not have a strong effect at the global scale. However, hyporheic assemblage  
33 composition within regions varied along a gradient of higher richness during wetter  
34 phases. This indicates that within geographic regions, hyporheic responses to surface  
35 drying are predictable and, by extension, hyporheic invertebrates are potentially useful  
36 ecological indicators of temporary river health. With many rivers now experiencing, or  
37 predicted to experience, lower flows and longer dry phases owing to climate change, the  
38 development of ecological assessment methods specific to flow intermittency is a  
39 priority. We advocate expanded monitoring of hyporheic zones in temporary rivers and  
40 recommend hyporheic invertebrates as potential bioindicators to complement surface  
41 water assessments.

42

43 **Keywords**

44 Ecological assessment; low river flows; aquatic invertebrates; river health; climate  
45 change; flow intermittency

46

47 **Abbreviations**

48 ANOSIM, analysis of similarities; CAP, canonical analysis of principal coordinates;  
49 EPT, Ephemeroptera, Plecoptera and Trichoptera; DC, dry channel; NMDS, non-metric  
50 multi-dimensional scaling; NSF, no surface flow; SF, surface flow

51

## 52 **1. Introduction**

53 Temporary rivers experience varying periods of flow cessation and surface  
54 drying (Larned et al., 2010) and are the major inland water component of many regions,  
55 including Australia (Kennard et al., 2010), southern Africa (Davies et al., 1995), North  
56 America (Poff and Ward, 1989), South America and the Mediterranean basin (Bonada  
57 et al., 2008). This widespread occurrence, along with the increase in flow intermittency  
58 occurring through climate change across much of the world (Kundzewicz et al., 2008)  
59 and the escalating human demand for water (Vörösmarty et al., 2010), makes  
60 understanding ecological consequences of intermittency in river systems increasingly  
61 important (Datry et al., 2011).

62 However, flow intermittency challenges our ability to monitor and assess the  
63 ecological integrity of temporary rivers. First, variation in the presence and timing of  
64 flow creates considerable spatial and temporal variation in these rivers' physical,  
65 chemical and biological attributes, such that many conventional indicators of river  
66 health may not detect anthropogenic changes (Datry et al., 2011). For example,  
67 taxonomic richness is expected to decline in temporary rivers as their waters decline and  
68 channels dry (Larned et al., 2010), but this response is not consistent among rivers or  
69 through time (Rolls et al., 2012). Therefore, this variation must be incorporated into the  
70 assessment process so that variation owing to natural wetting and drying can be  
71 distinguished from that caused by human activities (Sheldon, 2005), such as a reduction  
72 in taxonomic richness associated with land use change (Boulton et al., 1997). Second,  
73 the unpredictable spatio-temporal presence of surface waters means that monitoring  
74 programs based on sampling surface waters at specific locations or times of year

75 produce incomplete datasets (Steward et al., 2012), complicating analyses and creating  
76 gaps in reporting.

77 To avoid these problems, monitoring environments other than surface waters in  
78 temporary rivers have been suggested, including dry riverbeds (Steward et al., 2011)  
79 and the hyporheic zone, defined as the saturated sediments beneath the surface channel  
80 and adjacent banks. A major advantage of the hyporheic zone as a monitoring  
81 environment in temporary rivers is its persistence. Streams with dry surface channels  
82 can have substantial hyporheic zones (Valett et al., 1990; Claret and Boulton, 2003),  
83 and hyporheic invertebrates of temporary rivers have been collected from beneath both  
84 dry and wet channels, and across multiple seasons (e.g. Boulton et al., 1992a; del  
85 Rosario and Resh, 2000; Young et al., 2011). Although water can be lost from the  
86 subsurface sediments of some rivers within days of flow cessation (Datry, 2012),  
87 aquatic invertebrates often persist beneath surface channels in moist or dry sediments,  
88 even during long dry phases (Stubbington et al., 2009). These features suggest that  
89 hyporheic fauna are a viable alternative for temporary river bioassessment.

90 The potential for hyporheic invertebrates to act as indicators of health in  
91 temporary rivers has long been recognised (Boulton et al., 1992a), comparable to the  
92 use of macroinvertebrate richness and composition in permanent waters as indicators of  
93 overall river health (e.g. Barbour et al., 1999 (USA); Davies, 2000 (Australia); Clarke et  
94 al., 2003 (UK)). However, only a few attempts have been made to include hyporheic  
95 invertebrates in river health assessments (e.g. Nelson and Roline, 2003; Moldovan et al.,  
96 2013). This may reflect the cryptic nature of hyporheic fauna ('out of sight, out of  
97 mind'), a reluctance to accept new sampling methods, and a lack of appreciation of the  
98 ecological interactions between surface and hyporheic ecosystems in most rivers.

99 Further, in the context of temporary rivers, there is a need to determine the extent of  
100 hyporheic physical, chemical and biological variation attributable to surface flow  
101 conditions (Stubbington et al., 2011a). Factors known to affect hyporheic invertebrate  
102 distribution and composition, such as sediment characteristics and interstitial flow  
103 patterns, and the selectivity of sampling techniques (Fraser and Williams, 1997), also  
104 require consideration.

105 We aimed to assess the potential of hyporheic invertebrates of temporary rivers  
106 as ecological indicators of river health. We analysed hyporheic invertebrate data from  
107 temporary rivers across the world to determine whether assemblage composition and  
108 richness showed consistent patterns of variation that could be attributed to: (a) factors  
109 that could be controlled in a survey program, such as geographical location, climate  
110 zone and sampling techniques, and (b) factors that vary such as hydrological conditions  
111 at the time of sampling (hydrological phase). Our rationale was that if patterns of  
112 variation were consistent, and therefore predictable and quantifiable, then hyporheic  
113 invertebrates of temporary rivers could be used as bioindicators of variation owing to  
114 anthropogenic disturbance. We hypothesized that the broad-scale factors of climate and  
115 geographical region would have strong effects on hyporheic assemblage composition  
116 and, within these factors, surface water and surface flow conditions would also be  
117 important drivers (Fig. 1). In addition, we hypothesized that hyporheic invertebrate  
118 richness would be lower when the surface channel was dry or there was no surface  
119 water flow, and lower still when the system was also affected by anthropogenic  
120 disturbance (Fig. 1).

121

122 **2. Methods**

123 *2.1. Literature search*

124 We searched for relevant studies using the electronic databases Science Citation  
125 Index Expanded and Conference Proceedings Citation Index-Science within ISI Web of  
126 Science (Thomson Reuters), and the Boolean search statement: Topic = (invertebrate\*  
127 OR macroinvertebrate\*) AND (dry\* OR temporar\* or ephemeral\* or intermitten\* or  
128 episodic\*) AND (stream\* OR river\*) AND (hyporhe\* OR intersti\* OR vertical\*),  
129 where \* indicates all possible word endings. This yielded 75 studies, which we  
130 examined individually to confirm suitability. Studies were excluded if they were not  
131 field-based (i.e. experimental microcosm studies or review papers), were from  
132 perennially flowing rivers, did not collect hyporheic invertebrates, only examined  
133 certain taxa, and/or taxonomic resolution was coarser than family level for the Insecta.  
134 Where taxa lists or detail on collection methods or hydrological conditions were not  
135 given, we contacted the authors to access the data. This refined the 75 studies to 14,  
136 which we expanded to 21 by including data from two independent, unpublished studies  
137 (Leigh, Stubbington) and from five other published studies cited within those from the  
138 original search. Four of the 21 studies included rivers within primarily agricultural  
139 landscapes (Table 1). All other studies were conducted in areas with minimal  
140 anthropogenic impact, confirmed by the studies' authors (pers. comm.) or as inferred  
141 from the study-region descriptions (e.g. nature reserves, national parks).

142 We standardised the invertebrate records to presence-absence data using the  
143 lowest levels of within-group taxonomic resolution consistent across studies. Separate  
144 taxa lists were created for samples collected during different hydrological phases,  
145 classed as: dry channel (DC), flowing (surface flow, SF) and non-flowing waters (no

146 surface flow but surface water present, NSF). When a study's taxa list was drawn from  
147 samples taken during multiple hydrological phases including SF, it was allocated to the  
148 category 'mix'. Broad-scale geographical region (Antarctica, Australia, Europe, New  
149 Zealand and North America), climate zone (arid, mediterranean, polar, subarctic,  
150 temperate and tropical), collection method and depth were also used to categorise the  
151 data. Collection methods were classed as wells (invertebrates pumped from pipes sunk  
152 into the subsurface sediments), cages (invertebrates collected from buried colonisation  
153 pots), pits (invertebrates collected from pits in the hyporheic zone) and dug  
154 (invertebrates picked from sediments dug from the beneath the channel). Depth was  
155 categorised as either  $\leq 30$  cm or  $> 30$  cm. This yielded 24 taxa lists (termed 'cases') for  
156 our meta-analysis (Table 1). We also compiled accompanying information on direction  
157 of surface-subsurface flow during sampling (upwelling, downwelling or neutral), mesh  
158 size used to screen the invertebrate samples, and substrate composition.

159

## 160 *2.2. Meta-analysis*

161 To examine patterns in assemblage composition, we calculated Bray-Curtis  
162 similarities between all pairs of cases from the presence-absence data. The resultant  
163 similarity matrix formed the basis of all subsequent analyses involving assemblage  
164 composition (performed in PRIMER v6 with the PERMANOVA+ add-on; Clarke and  
165 Gorley, 2006; Anderson et al., 2008).

166 We tested the hypotheses that assemblage composition would be significantly  
167 associated with climate zone, geographical region, collection method, collection depth  
168 and hydrological phase (e.g. Fig. 1A), using separate one-way ANOSIM (analyses of  
169 similarities). Data collected from agricultural landscapes were not included in these



170 analyses as these particular hypotheses did not concern the potential effects of  
171 anthropogenic impacts. Differences were evaluated based on the ANOSIM  $R$  statistic  
172 (with  $R > 0.25$  and, when there were  $> 1000$  possible permutations of cases,  $P$ -values  $<$   
173  $0.05$  indicative of substantial differences between groups (Clarke and Warwick, 2001)).  
174 Although multi-factor models and interactions were not analysed owing to limited  
175 degrees of freedom, we created a joint climate and geographical region factor to test for  
176 differences in composition that were associated with their combination. Patterns of  
177 variation in assemblage composition among the cases, as indicated by the ANOSIM  
178 analyses, were visualised using non-metric multi-dimensional scaling (NMDS)  
179 ordination, based on 100 random starts. The two-dimensional solution was displayed if  
180 stress (goodness of fit) was  $< 0.2$  (Clarke and Warwick, 2001).

181 Canonical analysis of principal coordinates (CAP) was used to explore the  
182 relationship between assemblage richness and variation in composition (based on the  
183 Bray-Cutis similarity matrix) among cases, excluding those from agricultural  
184 landscapes. CAP is a constrained ordination technique designed to visualise multivariate  
185 patterns pertaining to specific hypotheses, and can be used as tool for prediction to place  
186 new data in ordination space (Anderson and Willis, 2003; Anderson et al., 2008). We  
187 used CAP to analyse how well the assemblage composition data could predict the  
188 positions of cases along a gradient of richness (as a proxy for river health) and the  
189 model's predictive capacity was tested using new cases (the cases from agricultural  
190 landscapes).

191 Under our hypothesis that hyporheic invertebrate richness, if acting as a good  
192 indicator of river health, would be lower under dry compared with wet conditions, and  
193 lower still under conditions of anthropogenic impact (Fig. 1B), the position of cases

194 from low-impact study regions along the gradient should indicate where ‘healthy’ rivers  
195 lie given the hydrological phase at the time of sampling. A decline in these rivers’  
196 health should lower their position, and ‘unhealthy’ rivers disturbed by human activities  
197 should be lower on the gradient than ‘healthy’ (relatively undisturbed) rivers with  
198 comparable features (e.g. similar flow regimes and matched hydrological phases) (Fig.  
199 1C). CAP model performance was evaluated based on the percentage of variation in the  
200 similarity matrix explained by the model, the trace statistic to test the null hypothesis of  
201 no difference in composition along the richness gradient, and a ‘leave-one-out’  
202 procedure to check for overparameterisation by choosing the number ( $m$ ) of principal  
203 coordinate axes for the analysis that minimises the ‘leave-one-out’ residual sums of  
204 squares (Anderson and Robinson, 2003; Anderson et al., 2008).

205         Patterns in richness data were also examined graphically to evaluate  
206 consistencies in the relationship between hydrological phase and richness metrics within  
207 climate and geographical regions (Fig. 1B), and to assess overall differences among  
208 those regions and among collection methods. Metrics comprised overall (raw absolute)  
209 richness and the mean richness and relative richness (proportion of total richness) of the  
210 cases’ most taxonomically rich groups (Mollusca, Crustacea, Insecta), including the  
211 EPT group (Ephemeroptera, Plecoptera and Trichoptera) within the Insecta. We  
212 included EPT metrics because EPT taxa are routinely used as bioindicators in river  
213 health assessment (e.g. Barbour et al., 1999) owing to their sensitivity to pollutants and  
214 changes in water quality.

215         All comparisons and analyses involving richness were based on taxa lists as  
216 reported by each study. Although sampling effort and taxonomic abundance may affect  
217 richness measures (Gotelli and Colwell, 2001), it was not possible to use standardisation

218 techniques (e.g. taxon sampling curves) prior to our analyses because many of the lists  
219 on which the richness (presence-absence) data were based were aggregations of taxa  
220 identified across samples (i.e. one list of taxa per case rather than separate lists for each  
221 sample collected per case) and abundance data were not consistently available.  
222 However, when there were enough cases within regions to compare sampling effort and  
223 richness, no clear trend was observed (Fig. 2). Therefore, although we acknowledge this  
224 limitation of the data, we consider raw taxon richness the best measure available for the  
225 purposes of our study.

226

### 227 **3. Results**

228         Assemblage composition was significantly associated with climate, both  
229 individually (ANOSIM  $R = 0.464$ ,  $P = 0.0003$ ) and in combination with broad  
230 geographical region ( $R = 0.641$ ,  $P = 0.0001$ ). Pairwise comparisons between cases  
231 grouped by the joint factor of climate and geographical region indicated that differences  
232 were present between all groups (pairwise  $R$  range: 0.333-1), except temperate New  
233 Zealand and Australian groups, Australian arid and temperate zone groups, arid  
234 Australian and North American groups, and tropical Australian and arid North  
235 American groups (pairwise  $R$  all  $< 0.2$ ;  $P$ -values not informative owing to low numbers  
236 of possible permutations). In NMDS ordination space, cases from temperate climates  
237 tended to align positively along the first axis (Fig. 3A). Cases from the high and low  
238 latitudes (tropical, subarctic and polar regions) tended to have lower representation of  
239 taxonomic groups than those from elsewhere (Fig. 4A,B). Cases from temperate  
240 climates generally had greater richness and/or relative richness of EPT and Insecta than  
241 those from other climates (Fig. 4A,B).

242 Depth of collection and the hydrological phase during sampling were not  
243 significantly associated with assemblage composition (ANOSIM  $P = 0.4680$  and  
244  $0.3940$ , respectively) at the global scale (i.e. among rather than within climate and  
245 geographical regions). However, there was a significant relationship between the  
246 method used to collect hyporheic invertebrates (wells, pits, cages, dug) and assemblage  
247 composition ( $R = 0.351$ ,  $P = 0.0030$ ). Pairwise comparisons indicated differences  
248 between all methods except for pits and cages (for which  $R < 0.05$ ), which could be  
249 visualised on the NMDS ordination (Fig. 3B). Pit- and cage-collected cases had lower  
250 richness and relative richness of crustacean taxa compared with those collected from  
251 wells (Fig. 4C,D). The one ‘dug’ case was from Antarctica and was taxonomically  
252 distinct from all other cases, containing only Rotifera, Nematoda and Tardigrada.  
253 Therefore, we repeated the above analyses without this case; results did not change  
254 (climate:  $R = 0.390$ ,  $P = 0.0009$ ; climate-geographical region:  $R = 0.599$ ,  $P = 0.0003$ ;  
255 method:  $R = 0.255$ ,  $P = 0.0015$ ), and both depth and hydrological phase were non-  
256 significant. Further, pairwise  $R$  statistics indicated that assemblage compositions were  
257 similar ( $R < 0.25$ ) between the same pairs of regions and collection methods listed  
258 above.

259 There was a strong and statistically significant relationship between assemblage  
260 richness and variation in composition (CAP, Fig. 5), with the canonical correlation  
261 explaining 98.3% of the variation in the similarity matrix of cases from systems classed  
262 as undisturbed by agricultural land use ( $m = 10$ , CAP trace statistic =  $0.97$ ,  $P = 0.0001$ ).  
263 Assemblages from Europe and from temperate climates tended to have higher richness  
264 than those from higher latitudes or from mediterranean or arid climates (Fig. 5A).  
265 Within climate and geographical regions, richness was usually higher when flow or

266 surface water was present during sample collection (Figs. 5A, 6). The greatest deviation  
267 from this trend involved the 'mix' case from arid North America, collected under  
268 conditions that included some surface flow. Richness of this case was low compared  
269 with the other 'mix' and 'surface-flowing' cases from the same climate and  
270 geographical region (Fig. 5A, 6). However, the invertebrates in this case had been  
271 collected from among the deepest hyporheic zones (mean collection depth = 93 cm;  
272 Boulton et al., 1992a) of all cases included in the analysis.

273         Within climate and geographical regions, a similar trend of lower richness in  
274 'dry-channel' or 'non-flowing' cases (DC or NSF) compared with 'mix' or 'surface-  
275 flowing' cases (mix or SF) was observed for EPT taxa (Fig. 7A). However, when these  
276 comparisons were based on relative rather than absolute EPT richness, the differences  
277 between DC/NSF and mix/SF cases within regions were generally smaller (Fig. 7B).  
278 This suggested that relative EPT richness in the hyporheic zone may, in some instances,  
279 vary less in response to changes in surface hydrology than absolute EPT richness.  
280 However, comparison of EPT absolute and relative richness between the two  
281 anthropogenically disturbed and the two undisturbed cases from temperate Europe (Fig.  
282 7) showed that while absolute richness of the disturbed cases was always lower than the  
283 undisturbed cases, relative richness was only lower for one of the disturbed cases.

284         Total richness for all four of the anthropogenically disturbed cases was predicted  
285 successfully by the CAP model. Based on their composition data, the richness of these  
286 'new' cases from agricultural landscapes was predicted within  $\pm 5$  taxa of the observed  
287 values (Figs. 5B, 6; Table 1). The positions of these cases along the gradient were also  
288 consistent with patterns among the other cases; European temperate zone cases had  
289 higher richness than other cases, and SF cases had higher richness than NSF and DC

290 cases. Further, in support of our hypotheses and consistent with observed values, the  
291 new cases were successfully predicted to have lower richness than those from  
292 undisturbed locations within the same climate zone (mediterranean) or climate-  
293 geographical region (temperate Europe) (Figs. 5B, 6; Table 1).

294

#### 295 **4. Discussion**

296 *4.1. The potential of hyporheic invertebrates as bioindicators of ecological health in*  
297 *temporary rivers*

298 Our meta-analysis of trends in the composition and richness of hyporheic  
299 assemblages from across the world suggests that there may be sufficient predictability in  
300 the responses of hyporheic invertebrates to surface drying and anthropogenic  
301 disturbance to support their use as ecological indicators in temporary rivers. Although  
302 assemblages differed between broad-scale climate and geographical regions, there was  
303 consistency in the trends observed between richness, hydrological phase and level of  
304 anthropogenic disturbance (as indicated by agricultural land use). Within regions, higher  
305 richness of hyporheic invertebrates was associated with surface flow presence than  
306 absence of surface flow or water, and the richness of cases from agricultural landscapes  
307 relative to this pattern was always lower.

308 Human activities have long been known to affect ecological processes and biotic  
309 communities in the hyporheic zone (e.g. Boulton et al., 1997; Trayler and Davis, 1998),  
310 and the mechanisms by which these effects occur are manifold. Agriculture, land  
311 clearing, urban development and river regulation can all modify sediment transport,  
312 promote colmation (clogging of interstices) and interfere with hydrological exchange  
313 between the surface and subsurface (Boulton et al., 1998). These processes in turn affect

314 hyporheic metabolism, water quality and invertebrate assemblages (Brunke and Gonsler,  
315 1997; Hancock, 2002). However, natural alternation between wet and dry phases in  
316 surface waters can also affect the composition of hyporheic assemblages (e.g. Boulton  
317 et al., 1992b; Mori et al., 2012). Our meta-analysis has shown that ecological effects of  
318 agriculture on temporary rivers, as indicated by changes in hyporheic invertebrate  
319 assemblages, can be distinguished from natural wetting and drying cycles, suggesting  
320 that this biota is a potential ecological indicator of river health for these systems.

321

#### 322 *4.2. Hyporheic invertebrate richness and EPT metrics as potential bioindicators*

323 The success of any monitoring or assessment program lies in its ability to detect  
324 changes in river health, diagnose the causes of poor health and instigate action to  
325 improve health. The choice of indicator(s) plays a major role in determining this success  
326 (Bunn et al., 2010). Indicators should be easy to measure, pertinent to the  
327 spatiotemporal scale of the assessment, and respond to anthropogenic impacts in a  
328 predictable and interpretable way (Boulton, 1999; Boulton et al., 2010).

329 While our study showed that total invertebrate richness and the richness and  
330 relative richness of EPT responded consistently to hydrological phase within broad-  
331 scale climate and geographical regions, there was less difference between wet and dry  
332 phases in relative than absolute EPT richness. Therefore, the proportion of EPT taxa in a  
333 hyporheic assemblage may be more stable as surface hydrology varies than the absolute  
334 number of EPT taxa. If this property of proportional richness is found to exist in any  
335 one site, system or group of systems targeted for bioassessment, the metric may provide  
336 a relatively reliable indication of health in temporary rivers.

337           However, while absolute EPT richness of anthropogenically-disturbed cases was  
338 lower than that of undisturbed cases from the same broad-scale region (temperate  
339 Europe), relative EPT richness of one of the disturbed cases was comparable with that  
340 of the undisturbed cases. This may reflect a relationship between the ability of  
341 hyporheic bioindicators, such as EPT richness, to detect anthropogenic disturbances and  
342 the type, severity or combination of the disturbances involved. In a Colorado stream  
343 affected by multiple human impacts, hyporheic EPT richness was a poor indicator and  
344 could not distinguish between impact types (Nelson and Roline, 2003). Taxonomic  
345 composition, however, was indicative of flow regulation effects, and high abundances  
346 of one particular taxon (a stonefly) were specifically indicative of mining effects  
347 (Nelson and Roline, 2003). Our findings and studies such as Nelson and Roline (2003)  
348 highlight the need for further investigation into the potential use of EPT metrics in  
349 hyporheic bioassessments, and into the development of hyporheic bioindicators more  
350 generally.

351

#### 352 *4.3 Caveats to and recommendations on the use of hyporheic invertebrates as* 353 *bioindicators*

354           Hyporheic sampling methods can be selective (Fraser and Williams, 1997;  
355 Boulton et al., 1998) and the general influence of sampling methods on ecological  
356 assessment outcomes is a well-known caveat of bioassessment (Cao and Hawkins,  
357 2011). Our study indicated that sampling method and assemblage composition were  
358 associated. Crustacea, for example, were better represented in cases for which samples  
359 had been collected from wells rather than pits or cages. Differences in sampling  
360 methods among the cases may even have played a role in structuring the differences



361 observed between regions. First, we included all reported taxa in our analysis, although  
362 some studies were primarily interested in macroinvertebrates and the collection and  
363 identification of meiofauna was therefore unlikely to be consistent across regions.  
364 Second, the mesh size used to screen invertebrates probably influenced sample  
365 composition and richness. The absence of Crustacea from temperate North American  
366 cases (Fig. 4), for example, may have partially resulted from the relatively large mesh  
367 size used (250  $\mu\text{m}$ ; Table 1), potentially precluding collection of small invertebrates  
368 such as microcrustaceans.

369 Therefore, while the technical capacity and funding level of any assessment  
370 program will dictate the collection methods, sampling effort, taxonomic resolution and  
371 other identification protocols implemented (Lindenmayer et al., 2012), the potential  
372 effects of these factors on assessment outcomes must be acknowledged. Based on the  
373 techniques commonly used by most studies (Table 1) and from our own experiences of  
374 sampling hyporheic fauna, we recommend standardized protocols such as sampling  
375 from wells inserted 30-60 cm in the streambed and using self-priming hand-pumps to  
376 collect 5-6 L, filtered through a maximum mesh size of 125  $\mu\text{m}$ . Consideration of  
377 factors beyond the control of the operator that influence the composition and  
378 distribution of hyporheic fauna, such as sediment characteristics and direction of  
379 vertical hydrological exchange (Brunke and Gonser, 1997; Boulton et al., 1998), will  
380 also help to discriminate anthropogenically induced changes in hyporheic bioindicators.  
381 Pilot studies and the strategic development of sampling and analytical methods (e.g.  
382 Buss et al., 2009; Downes 2010) will be essential to ensure success.

383 Finally, we suggest that temporary river assessment programs incorporating  
384 hyporheic bioindicators will benefit during developmental stages from a conceptual

385 understanding of how surface flow variation mediates changes in those indicators (e.g.  
386 Fig. 8), both in disturbed and undisturbed locations. We suggest that in many rivers,  
387 particularly ‘losing’ systems where downwelling water predominates, the loss of surface  
388 water may be followed by a gradual reduction in the volume of the saturated hyporheic  
389 zone (Fig. 8A, B). As surface-subsurface flow exchange uncouples and the size of the  
390 saturated subsurface continues to decrease, changes in hyporheic water quality occur  
391 (e.g. reduction in dissolved oxygen; Fig. 8C), followed by potentially substantial change  
392 in invertebrate assemblage composition, distribution and diversity (Boulton and Stanley,  
393 1995; Stanley and Boulton, 1995). Our study suggests that this process may manifest as  
394 a marked but gradual decline in richness along the drying gradient, with anthropogenic  
395 disturbance compounding the ecological response (Fig. 8D). Therefore, initial  
396 assessment data must be collected over adequate spatial and temporal scales that span  
397 wet, dry and transitional phases in flow intermittency so that the full range of  
398 invertebrate responses to surface flow variation can be described, tested against the  
399 conceptual understanding and, if possible, modelled for use in future assessments.

400

## 401 **5. Conclusion**

402 Our global analysis provides evidence that invertebrate assemblage  
403 characteristics within hyporheic zones have the potential to act as ecological health  
404 indicators of temporary rivers. While this supports the broader suggestion that patterns  
405 and processes within hyporheic zones are important indicators of the health of  
406 connected surface- and groundwater ecosystems (Boulton and Stanley, 1996; Boulton,  
407 2000), a lack of baseline data and uptake of protocols to develop, test and use hyporheic  
408 indicators will continue to hinder their routine use (Boulton et al., 2010). Increased

409 efforts to compile knowledge and gather data on hyporheic fauna will help to resolve  
410 this issue and improve our understanding of hyporheic responses to surface system  
411 disturbances (Marmonier et al., 2012; Wood et al., 2012). We advocate expanded  
412 monitoring of hyporheic zones in temporary rivers and recommend hyporheic  
413 invertebrates as potential bioindicators to complement surface water assessments.

414

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424

#### 425 **References**

- 426 Anderson, M.J., Robinson, J., 2003. Generalised discriminant analysis based on  
427 distances. *Aust. N. Z. J. Stat.* 45, 301-318.
- 428 Anderson, M.J., Willis, T.J., 2003. Canonical analysis of principal coordinates: a useful  
429 method of constrained ordination for ecology. *Ecology* 84, 511-525.
- 430 Anderson, M.J., Gorley, R.N., Clarke, R.K., 2008. PERMANOVA+ for PRIMER:  
431 Guide to Software and Statistical Methods. PRIMER-E, Plymouth, UK.

432 Barbour, M.T., Gerritsen, J., Snyder, B.D., Stribling, J.B., 1999. Rapid Bioassessment  
433 Protocols for Use in Streams and Wadeable Rivers: Periphyton, Benthic  
434 Macroinvertebrates, and Fish. Second ed. EPA 841-B-99-002. US EPA, Office  
435 of Water, Washington, DC.

436 Bonada, N., Rieradevall, M., Dallas, H., Davis, J., Day, J., Figueroa, R., Resh, V.H.,  
437 Prat, N., 2008. Multi-scale assessment of macroinvertebrate richness and  
438 composition in Mediterranean-climate rivers. *Freshw. Biol.* 53, 772–788.

439 Boulton, A.J., 1999. An overview of river health assessment: Philosophies, practice,  
440 problems and prognosis. *Freshw. Biol.* 41, 469-479.

441 Boulton, A.J., 2000. River ecosystem health down under: assessing ecological condition  
442 in riverine groundwater zones in Australia. *Ecosystem Health* 6, 108-118.

443 Boulton, A.J., Stanley, E.H., 1995. Hyporheic processes during flooding and drying in a  
444 Sonoran Desert stream. II. Faunal dynamics. *Arch. Hydrobiol.* 134, 27-52.

445 Boulton, A.J., Stanley, E.H., 1996. But the story gets better: subsurface invertebrates in  
446 stream ecosystems. *Trends Ecol. Evol.* 11, 430.

447 Boulton, A.J., Stibbe, S.E., Grimm, N.B., Fisher, S.G., 1991. Invertebrate recolonization  
448 of small patches of defaunated hyporheic sediments in a Sonoran Desert stream.  
449 *Freshw. Biol.* 26, 267-278.

450 Boulton, A.J., Valett, H.M., Fisher, S.G., 1992a. Spatial distribution and taxonomic  
451 composition of the hyporheos of several Sonoran Desert streams. *Arch.*  
452 *Hydrobiol.* 125, 37-61.

453 Boulton, A.J., Stanley, E.H., Fisher, S.G., Lake, P.S., 1992b. Over-summering strategies  
454 of macroinvertebrates in intermittent streams in Australia and Arizona, in:  
455 Robarts, R.D., Bothwell, M.L. (Eds.), *Aquatic Ecosystems in Semi-arid*

456 Regions: Implications for Resource Management. Environment Canada,  
457 Saskatoon, Saskatchewan, pp. 227-237.

458 Boulton, A.J., Scarsbrook, M.R., Quinn, J.M., Burrell, G.P., 1997. Land-use effects on  
459 the hyporheic ecology of five small streams near Hamilton, New Zealand. *N. Z.*  
460 *J. Mar. Freshw. Res.* 31, 609-622.

461 Boulton, A.J., Findlay, S., Marmonier, P., Stanley, E.H., Valett, H.M., 1998. The  
462 functional significance of the hyporheic zone in streams and rivers. *Annu. Rev.*  
463 *Ecol. Syst.* 29, 59-81.

464 Boulton, A.J., Datry, T., Kasahara, T., Mutz, M., Stanford, J.A., 2010. Ecology and  
465 management of the hyporheic zone: Stream-groundwater interactions of running  
466 waters and their floodplains. *J. N. Am. Benthol. Soc.* 29, 26-40.

467 Brunke, M., Gonser, T., 1997. The ecological significance of exchange processes  
468 between rivers and groundwater. *Freshw. Biol.* 37, 1-33.

469 Bunn, S.E., Abal, E.G., Smith, M.J., Choy, S.C., Fellows, C.S., Harch, B.D., Kennard,  
470 M.J., Sheldon, F., 2010. Integration of science and monitoring of river  
471 ecosystem health to guide investments in catchment protection and  
472 rehabilitation. *Freshw. Biol.* 55, 223-240.

473 Buss, S., Cai, Z., Cardenas, B., Fleckenstein, J., Hannah, D., Heppell, K., Hulme, P.,  
474 Ibrahim, T., Kaeser, D., Krause, S., Lawler, D., Lerner, D., Mant, J., Malcolm,  
475 I., Old, G., Parkin, G., Pickup, R., Pinay, G., Porter, J., Rhodes, G., Ritchie, A.,  
476 Riley, J., Robertson, A., Sear, D., Shields, B., Smith, J., Tellam, J., Wood, P.,  
477 2009. *The Hyporheic Handbook: A handbook on the groundwater–surface water*  
478 *interface and hyporheic zone for environmental managers. Science Report,*  
479 *SC0500. Environment Agency, Bristol, UK.*

480 Cao, Y., Hawkins, C.P., 2011. The comparability of bioassessments: a review of  
481 conceptual and methodological issues. *J. N. Am. Benthol. Soc.* 30, 680-701.

482 Claret, C., Boulton, A.J., 2003. Diel variation in surface and subsurface microbial  
483 activity along a gradient of drying in an Australian sand-bed stream. *Freshw.*  
484 *Biol.* 48, 1739-1755.

485 Clarke, K.R., Gorley, R.N., 2006. PRIMER v6: User Manual/Tutorial. PRIMER-E,  
486 Plymouth, UK.

487 Clarke, K.R., Warwick, R.M., 2001. Change in marine communities: An approach to  
488 statistical analysis and interpretation, fourth ed. PRIMER-E, Plymouth, UK.

489 Clarke, R.T., Wright, J.F., Furse, M.T., 2003. RIVPACS models for predicting the  
490 expected macroinvertebrate fauna and assessing the ecological quality of rivers.  
491 *Ecol. Model.* 160, 219-233.

492 Clinton, S.M., Grimm, N.B., Fisher, S.G., 1996. Response of a hyporheic invertebrate  
493 assemblage to drying disturbance in a desert stream. *J. N. American Benthol.*  
494 *Soc.* 15, 700-712.

495 Collins, B.M., Sobczak, W.V., Colburn, E.A., 2007. Subsurface flowpaths in a forested  
496 headwater stream harbor a diverse macroinvertebrate community. *Wetlands* 27,  
497 319-325.

498 Cooling, M.P., Boulton, A.J., 1993. Aspects of the hyporheic zone below the terminus  
499 of a South Australian arid-zone stream. *Aust. J. Mar. Freshw. Res.* 44, 411-426.

500 Crossman, J., Bradley, C., Milner, A., Pinay, G., 2012. Influence of environmental  
501 instability of groundwater-fed streams on hyporheic fauna, on a glacial  
502 floodplain, Denali National Park, Alaska. *River Res. Appl.* doi:  
503 10.1002/rra.1619

504 Datry, T., 2012. Benthic and hyporheic invertebrate assemblages along a flow  
505 intermittence gradient: effects of duration of dry events. *Freshw. Biol.* 57, 563-  
506 574.

507 Datry, T., Larned, S.T., Scarsbrook, M.R., 2007. Responses of hyporheic invertebrate  
508 assemblages to large-scale variation in flow permanence and surface–subsurface  
509 exchange. *Freshw. Biol.* 52, 1452-1462.

510 Datry, T., Arscott, D., Sabater, S., 2011. Recent perspectives on temporary river  
511 ecology. *Aquat. Sci.* 73, 453-457.

512 Davies, B.R., Thoms, M.C., Walker, K.F., O'Keefe, J.H., Gore, J.A., 1995. Dryland  
513 rivers: Their ecology, conservation and management, in: Calow, P., Petts, G.E.  
514 (Eds.), *The Rivers Handbook: Hydrological and Ecological Principles*.  
515 Blackwell Scientific Publishing, Oxford, pp. 484-511.

516 Davies, P.E., 2000. Development of a national river bioassessment system  
517 (AUSRIVAS), in: Wright, J.F., Sutcliffe, D.W., Furse, M.T. (Eds.), *Assessing*  
518 *the Biological Quality of Freshwaters: RIVPACS and Other Techniques*.  
519 Freshwater Biological Association, Ambleside, U.K., pp. 113-124.

520 del Rosario, R.B., Resh, V.H., 2000. Invertebrates in intermittent and perennial streams:  
521 is the hyporheic zone a refuge from drying? *J. N. Am. Benthol. Soc.* 19, 680-  
522 696.

523 del Rosario, R.B., Resh, V.H., 2001. Interstitial invertebrate assemblages associated  
524 with small-scale subsurface flowpaths in perennial and intermittent California  
525 streams. *Arch. Hydrobiol.* 150, 629-640.

526 Downes, B.J., 2010. Back to the future: little-used tools and principles of scientific  
527 inference can help disentangle effects of multiple stressors on freshwater  
528 ecosystems. *Freshw. Biol.* 55, 60-79.

529 Fraser, B.G., Williams, D.D., 1997. Accuracy and precision in sampling hyporheic  
530 fauna. *Canadian Journal of Fisheries and Aquatic Sciences* 54, 1135-1141.

531 Gotelli, N.J., Colwell, R.K., 2001. Quantifying biodiversity: Procedures and pitfalls in  
532 the measurement and comparison of species richness. *Ecol. Lett.* 4, 379-391.

533 Griffith, M.B., Perry, S.A., 1993. The distribution of macroinvertebrates in the  
534 hyporheic zone of two small Appalachian headwater streams. *Arch. Hydrobiol.*  
535 126, 373-384.

536 Hancock, P., 2002. Human impacts on the stream-groundwater exchange zone. *Environ.*  
537 *Manag.* 29, 761-781.

538 Kennard, M.J., Pusey, B.J., Olden, J.D., Mackay, S.J., Stein, J.L., Marsh, N., 2010.  
539 Classification of natural flow regimes in Australia to support environmental flow  
540 management. *Freshw. Biol.* 55, 171–193.

541 Kundzewicz, Z.W., Mata, L.J., Arnell, N.W., Döll, P., Jimenez, B., Miller, K., Oki, T.,  
542 Şen, Z., Shiklomanov, I., 2008. The implications of projected climate change for  
543 freshwater resources and their management. *Hydrol. Sci. J.* 53, 3-10.

544 Larned, S.T., Datry, T., Arscott, D.B., Tockner, K., 2010. Emerging concepts in  
545 temporary-river ecology. *Freshw. Biol.* 55, 717-738.

546 Lindenmayer, D.B., Gibbons, P., Bourke, M., Burgman, M., Dickman, C.R., Ferrier, S.,  
547 Fitzsimons, J., Freudenberger, D., Garnett, S.T., Groves, C., Hobbs, R.J.,  
548 Kingsford, R.T., Krebs, C., Legge, S., Lowe, A.J., McLean, R., Montambault, J.,  
549 Possingham, H., Radford, J., Robinson, D., Smallbone, L., Thomas, D., Varcoe,



550 T., Vardon, M., Wardle, G., Woinarski, J., Zerger, A., 2012. Improving  
551 biodiversity monitoring. *Austral Ecol.* 37, 285-294.

552 Marmonier, P., Archambaud, G., Belaidi, N., Bougon, N., Breil, P., Chauvet, E., Claret,  
553 C., Cornut, J., Datry, T., Dole-Olivier, M.-J., Dumont, B., Flipo, N., Foulquier,  
554 A., Gérino, M., Guilpart, A., Julien, F., Maazouzi, C., Martin, D., Mermillod-  
555 Blondin, F., Montuelle, B., Namour, P., Navel, S., Ombredane, D., Pelte, T.,  
556 Piscart, C., Pusch, M., Stroffek, S., Robertson, A., Sanchez-Pérez, J.-M.,  
557 Sauvage, S., Taleb, A., Wantzen, M., Vervier, P., 2012. The role of organisms in  
558 hyporheic processes: Gaps in current knowledge, needs for future research and  
559 applications. *Ann. Limnol. Int. J. Limnol.* 48, 253-266.

560 Moldovan, O.T., Meleg, I.N., Levei, E., Terente, M., 2013. A simple method for  
561 assessing biotic indicators and predicting biodiversity in the hyporheic zone of a  
562 river polluted with metals. *Ecol. Indic.* 24, 412-420.

563 Mori, N., Simčič, T., Žibrat, U., Brancelj, A., 2012. The role of river flow dynamics and  
564 food availability in structuring hyporheic microcrustacean assemblages: A reach  
565 scale study. *Fundam. Appl. Limnol.* 180, 335-349.

566 Nelson, S.M., Roline, R.A., 2003. Effects of multiple stressors on hyporheic  
567 invertebrates in a lotic system. *Ecol. Indic.* 3, 65-7.

568 Paltridge, R.M., Dostine, P.L., Humphrey, C.L., Boulton, A.J., 1997. Macroinvertebrate  
569 recolonization after re-wetting of a tropical seasonally-flowing stream (Magela  
570 Creek, Northern Territory, Australia). *Mar. Freshw. Res.* 48, 633-645.

571 Poff, N.L., Ward, J.V., 1989. Implications of streamflow variability and predictability  
572 for lotic community structure: A regional analysis of streamflow patterns. *Can.*  
573 *J. Fish. Aquat. Sci.* 46, 1805-1818.

574 Rolls, R.J., Leigh, C., Sheldon, F., 2012. Mechanistic effects of low-flow hydrology on  
575 riverine ecosystems: Ecological principles and consequences of alteration.  
576 *Freshw. Sci.* 31, 1163-1186.

577 Sheldon, F., 2005. Incorporating natural variability into the assessment of ecological  
578 health in Australian dryland rivers. *Hydrobiologia* 552, 45-56.

579 Stanley, E.H., Boulton, A.J., 1995. Hyporheic processes during flooding and drying in a  
580 Sonoran Desert stream. I. Hydrologic and chemical dynamics. *Arch. Hydrobiol.*  
581 134, 1-26.

582 Steward, A., Marshall, J., Sheldon, F., Harch, B., Choy, S., Bunn, S., Tockner, K., 2011.  
583 Terrestrial invertebrates of dry river beds are not simply subsets of riparian  
584 assemblages. *Aquat. Sci.* 73, 551-566.

585 Steward, A.L., von Schiller, D., Tockner, K., Marshall, J.C., Bunn, S.E., 2012. When  
586 the river runs dry: human and ecological values of dry riverbeds. *Frontiers in*  
587 *Ecology and the Environment* 10, 202-209.

588 Stubbington, R., 2011. The hyporheic zone as a refugium for benthic invertebrates in  
589 groundwater-dominated streams. PhD thesis, Loughborough University,  
590 Loughborough, UK.

591 Stubbington, R., Wood, P.J., Boulton, A.J., 2009. Low flow controls on benthic and  
592 hyporheic macroinvertebrate assemblages during supra-seasonal drought.  
593 *Hydrol. Process.* 23, 2252-2263.

594 Stubbington, R., Wood, P., Reid, I., 2011a. Spatial variability in the hyporheic zone  
595 refugium of temporary streams. *Aquat. Sci.* 73, 499-511.

596 Stubbington, R., Wood, P.J., Reid, I., Gunn, J., 2011b. Benthic and hyporheic  
597 invertebrate community responses to seasonal flow recession in a groundwater-  
598 dominated stream. *Ecohydrology* 4, 500-511.

599 Trayler, K.M., Davis, J.A., 1998. Forestry impacts and the vertical distribution of  
600 stream invertebrates in south-western Australia. *Freshw. Biol.* 40, 331-342.

601 Treonis, A.M., H.Wall, D., Virginia, R.A., 1999. Invertebrate biodiversity in Antarctic  
602 dry valley soils and sediments. *Ecosystems* 2, 482–492.

603 Valett, H.M., Fisher, S.G., Stanley, E.H., 1990. Physical and chemical characteristics of  
604 the hyporheic zone of a Sonoran Desert stream. *J. N. Am. Benthol. Soc.* 9, 201-  
605 215.

606 Vörösmarty, C.J., McIntyre, P.B., Gessner, M.O., Dudgeon, D., Prusevich, A., Green,  
607 P., Glidden, S., Bunn, S.E., Sullivan, C.A., Liermann, C.R., Davies, P.M., 2010.  
608 Global threats to human water security and river biodiversity. *Nature* 467, 555-  
609 561.

610 Wood, P.J., Boulton, A.J., Little, S., Stubbington, R., 2010. Is the hyporheic zone a  
611 refugium for aquatic macroinvertebrates during severe low flow conditions?  
612 *Fundam. Appl. Limnol.* 176, 377-390.

613 Wood, P.J., Gilvear, D.J., Willby, N., Robertson, A.L., Gledhill, T., Boon, P.J., 2012.  
614 Improvements in understanding the contribution of hyporheic zones to  
615 biodiversity and ecological functioning of UK rivers, in: Boon, P.J., Raven, P.J.  
616 (Eds.), *River Conservation and Management*. John Wiley & Sons, Ltd,  
617 Chichester, UK, pp. 159-173.

618 Young, B.A., Norris, R.H., Sheldon, F., 2011. Is the hyporheic zone a refuge for  
619 macroinvertebrates in drying perennial streams? *Mar. Freshw. Res.* 62, 1373-  
620 1382.

621 Table 1: Characteristics of systems used in the meta-analysis of hyporheic invertebrate assemblage composition and richness, separated  
 622 into twenty-four cases based on climate, geographical location, anthropogenic disturbance<sup>a</sup>, hydrological phase during sampling and  
 623 collection particulars.

Climate	Broad geographical location	River	Anthropogenic disturbance <sup>a</sup>	Maximum flow cessation period (mo y <sup>-1</sup> ) <sup>b</sup>	Hydrological phase	Collection method	Collection depth (cm)	Mesh size (µm)	Vertical hydrological exchange direction	Stream bed composition <sup>c</sup>	Number of samples	Total richness <sup>d</sup>	Source <sup>e</sup>
Temperate	Europe (France)	Albarine River	low	6	SF	wells	≤30	90	?	coarse alluvium	100	45	Datry, 2012
Temperate	Europe (UK)	River Lathkill	low	5	SF	wells	≤30	90	D,N	cobble, gravel, sand	167	36	Stubbington et al., 2011a, b
Temperate	Europe (UK)	River Glen	other	5	SF	wells	≤30	90	D	cobble, gravel, sand	120	35 (32)	Stubbington, 2011; Stubbington et al., 2011a
Temperate	Europe (UK)	Little Stour River	other	only dries during supra-seasonal droughts	SF	wells	≤30	90	?	coarse alluvium	99	27 (32)	Stubbington et al., 2009; Wood et al., 2010
Temperate	New Zealand	Selwyn River	low	11	SF	wells	≤30	90	?	coarse alluvium	82	33	Datry et al., 2007
Temperate	Australia (Australian Capital Territory, ACT)	Burke and Condor Creeks	low	1	SF	cages	≤30	n/a	?	?	6	25	Young et al., 2011
Temperate	Australia (ACT)	Burke and Condor Creeks	low	1	DC	cages	≤30	n/a	?	cobble, boulder, gravel, sand	6	11	Young et al., 2011
Temperate	Australia (Victoria)	Lerderderg and Werribee Rivers	low	2	DC	pits	≤30	50	D	gravel, pebble, cobble, boulder	5	8	Boulton et al., 1992b
Temperate	North America (West Virginia)	Two unnamed tributaries of Elklick Run	low	3	SF	cages	≤30	250	?	cobble, boulder, sand	15	22	Griffith and Perry, 1993

Temperate	North America (Massachusetts)	Bigelow Brook tributary	low	12	DC	pits	≤30	n/a	?	cobble, gravel, sand, silt	6	15	Collins et al., 2007
Mediterranean	North America (California)	Cronin Creek	low	5	NSF+DC	wells	>30	63	D	cobble, gravel	82	28	del Rosario and Resh, 2000
Mediterranean	North America (California)	Cronin Creek	low	5	DC	wells	>30	63	N	cobble, gravel	10	18	del Rosario and Resh, 2001
Mediterranean	Australia (South Australia)	Finniss, Light, Marne, Onkaparinga and Wakefield Rivers	other	9	SF	wells	>30	75	mix	sand, silt, gravel, cobble	9	16 (17)	C. Leigh, unpubl. data
Mediterranean	Australia (South Australia)	Angas, Marne and Wakefield Rivers	other	9	NSF	wells	>30	75	mix	sand, silt, gravel, cobble	7	10 (13)	C. Leigh, unpubl. data
Arid	Australia (South Australia)	Brachina Creek	low	9	NSF+DC	wells	>30	50	D,N	cobble, gravel	88	18	Cooling and Boulton, 1993
Arid	North America (Arizona)	Sycamore and Bridle Creeks	low	9	SF	cages	≤30	63	D,U	gravel	80	12	Boulton et al., 1991
Arid	North America (Arizona)	Sycamore and Bridle Creeks	low	9	mix	wells	≤30	50	mix	gravel, sand	17	20	Boulton et al., 1992a
Arid	North America (Arizona)	Sycamore and Bridle Creeks	low	9	mix	wells	>30	50	mix	gravel, sand	17	7	Boulton et al., 1992a
Arid	North America (Arizona)	Sycamore and Bridle Creeks	low	9	DC	wells	>30	50	mix	gravel, sand	17	7	Boulton et al. 1992a
Arid	North America (Arizona)	Sycamore Creek	low	9	DC	pits	?	50	D,U	gravel, pebble, cobble, boulder	10	9	Boulton et al., 1992b

Arid	North America (Arizona)	Rock Creek	low	8	NSF+DC	wells	>30	63	?	sand	209	16	Clinton et al., 1996
Tropical	Australia (Northern Territory)	Magela Creek	low	6	DC	pits	>30	63	D	sand	3	7	Paltridge et al., 1997
Subarctic	North America (Alaska)	Toklat River	low	?	SF	cages	≤30	65	U	cobble, gravel	4	8	Crossman et al., 2012
Polar	Antarctica	Von Guerard Stream and Harnish Creek	low	11	SF	dug	≤30	n/a	?	coarse alluvium	18	3	Treonis et al., 1999

624 <sup>a</sup> ‘low’ indicates study areas in nature reserves, national parks, native woodlands, protected national recreation areas, or in areas that have  
625 been defined by the studies’ authors as under low influence of anthropogenic impact (pers. comm. T. Datry). ‘Other’ indicates study  
626 regions in primarily agricultural landscapes. However, flow losses can be exacerbated in some reaches of the River Lathkill owing to  
627 disused mine-drainage soughs, and on the Glen by extractions for human use

628 <sup>b</sup> approximate, based on information provided in the studies, and applicable only to the study sites used in this study

629 <sup>c</sup> as defined in each publication or by the studies’ authors

630 <sup>d</sup> based on the taxonomic resolution used in this study. Values in parentheses are the predicted values from canonical analysis of principal  
631 coordinates (*see* Results)

632 <sup>e</sup> unpublished data by Stubbington (2011; PhD thesis) were consolidated with data from Stubbington et al. (2011a) collected from the same  
633 river, sites and sampling period (River Glen); as were data from the River Lathkill (Stubbington et al., 2011a, b) and data from the Little  
634 Stour River (Stubbington et al., 2009; Wood et al., 2010)

635 NSF, no surface flow but surface water present

636 SF, surface flow

637 DC, dry surface channel

638 mix, mixture of hydrological phases that includes surface flow, or an unspecified mix of vertical hydrological exchange directions

639 D, downwelling

640 N, neutral

641 U, upwelling

642 ?, data not available

643 n/a, not applicable

644 **Figure captions**

645 Figure 1: Conceptual diagrams of hypotheses on hyporheic invertebrate assemblages of  
646 temporary rivers. **A**: relationships, illustrated as if in two-dimensional ordination space,  
647 between assemblages of taxa from different climate and geographical regions (encircled  
648 diamonds, triangles and squares) collected under different hydrological phases (open  
649 symbols indicate assemblages beneath dry surface channels). **B**: relationships between  
650 taxonomic richness and these same factors. **C**: hypothetical gradient of taxonomic richness  
651 of assemblages from different climates and regions, showing how dry phases and  
652 disturbance by human activities deflect samples down the gradient. Climate 'A' is drier  
653 than 'B'. 'Undisturbed' and 'Disturbed' reflect river systems subject to different levels of  
654 anthropogenic impact. 'Wet' vs 'Dry' refers to surface water flow vs no surface water flow,  
655 surface water presence vs absence, or surface water flow vs surface water absence.

656

657 Figure 2: Relationship between taxonomic richness versus sampling effort within climate  
658 and geographical regions examined in this study that had > 3 taxa lists ('cases'): temperate  
659 Europe and arid North America.

660

661 Figure 3: Two-dimensional non-metric multi-dimensional scaling (NMDS) ordination  
662 (stress = 0.158) of hyporheic invertebrate assemblages ('cases') collected using different  
663 methods and from different climate and geographical regions, not including those from  
664 agricultural landscapes. **A**: encircled symbols show climate and geographical regions with  
665 at least two cases, including at least one dry channel (DC) case. **B**: dashed line encircles  
666 cases for which samples were collected from wells, solid line from pits and cages.



667

668 Figure 4: Richness and relative richness (mean  $\pm$  1 standard deviation) of taxonomic groups  
669 identified to higher levels of taxonomic resolution (Mollusca: family, Crustacea: order and  
670 family, EPT: family and Insecta: family) by climate and geographical region (**A**, **B**) and by  
671 collection method (**C**, **D**). EPT refers to Ephemeroptera, Plecoptera and Trichoptera;  
672 relative richness is a unitless measure showing Mollusca, Crustacea and EPT richness  
673 proportional to the richness of all invertebrate taxa.

674

675 Figure 5: Canonical analysis of principal coordinates (CAP) ordination relating hyporheic  
676 assemblages ('cases') to a taxonomic richness gradient. **A**: CAP model based on sampling  
677 locations with low anthropogenic disturbance. Ellipses show trend of higher richness for  
678 cases sampled during wet phases (solid line) and lower richness during dry phases (dashed  
679 line), exceptions include the two high-latitude, low richness cases (subarctic and polar  
680 cases) and the deep-zone case from arid North America. **B**: predicted placement of cases  
681 from agricultural landscapes ('disturbed' cases) onto the gradient, in comparison with  
682 'undisturbed' cases from similar regions or climates. Hydrological phase during sampling  
683 (SF, mix, NSF, DC): *see* Table 1.

684

685 Figure 6: Total richness of invertebrates in hyporheic zones sampled in different climate  
686 and geographical regions, and in 'undisturbed' and 'disturbed' (primarily agricultural)  
687 landscapes. Hydrological phase during sampling: dry channels (DC), non-flowing surface  
688 waters (NSF), surface flow (SF and mix): *see* Table 1. Closed, black bars show SF data,  
689 unless indicated as mix.

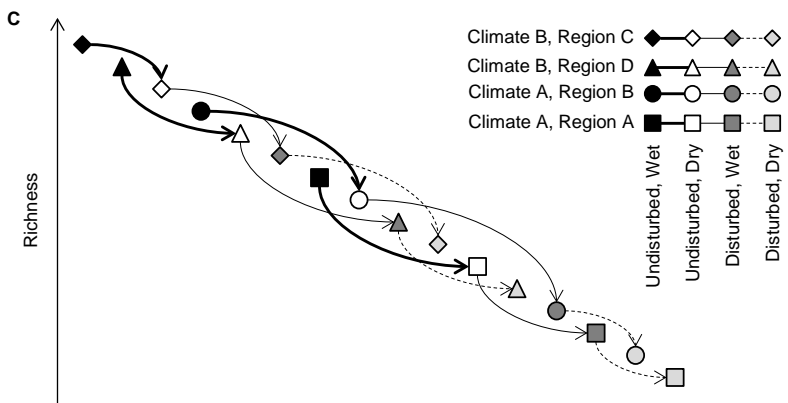
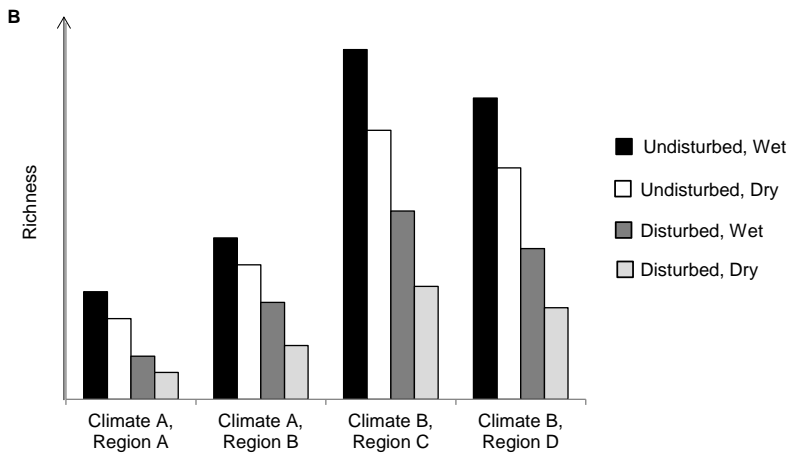
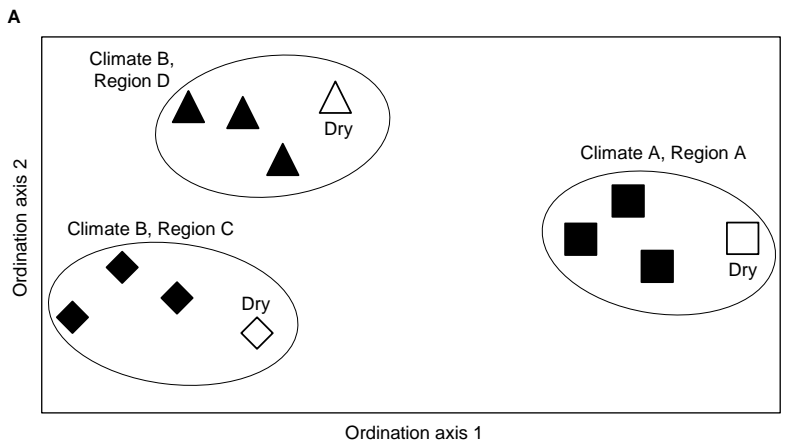
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691 Figure 7: Richness of Ephemeroptera, Plecoptera and Trichoptera (EPT) in hyporheic zones  
692 sampled in different climate and geographical regions, and in ‘undisturbed’ and ‘disturbed’  
693 (primarily agricultural) landscapes. **A**: raw EPT richness; **B**: relative richness, a unitless  
694 measure of EPT richness proportional to the richness of all invertebrate taxa. Hydrological  
695 phase during sampling: dry channels (DC), non-flowing surface waters (NSF), surface flow  
696 (SF and mix): *see* Table 1. Closed, black bars show SF data, unless indicated as mix.

697

698 Figure 8: Conceptual model of different conditions (A, B, C, D) in the hyporheic zone of a  
699 temporary river, unimpacted or impacted by human activities, during a complete surface-  
700 flow cycle through time. Consistent subsurface flow is assumed, and variations on this  
701 general model will occur in association with differences in climate, geographical location,  
702 and both small- and large-scale river characteristics (units of measure are therefore not  
703 provided). **A**: surface flow magnitude; **B**: hyporheic saturation (depth to water table); **C**:  
704 hyporheic water quality (e.g. dissolved oxygen concentration); **D**: invertebrate richness in  
705 the hyporheic zone.

706

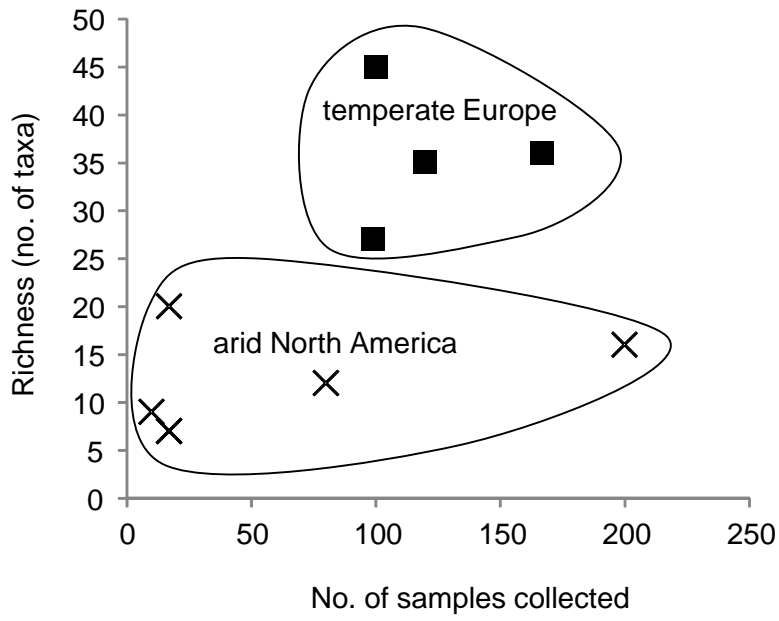


Hyporheic assemblages grouped by broad-scale climate and geographical region, human disturbance and hydrological phase during sampling

707

708 Figure 1

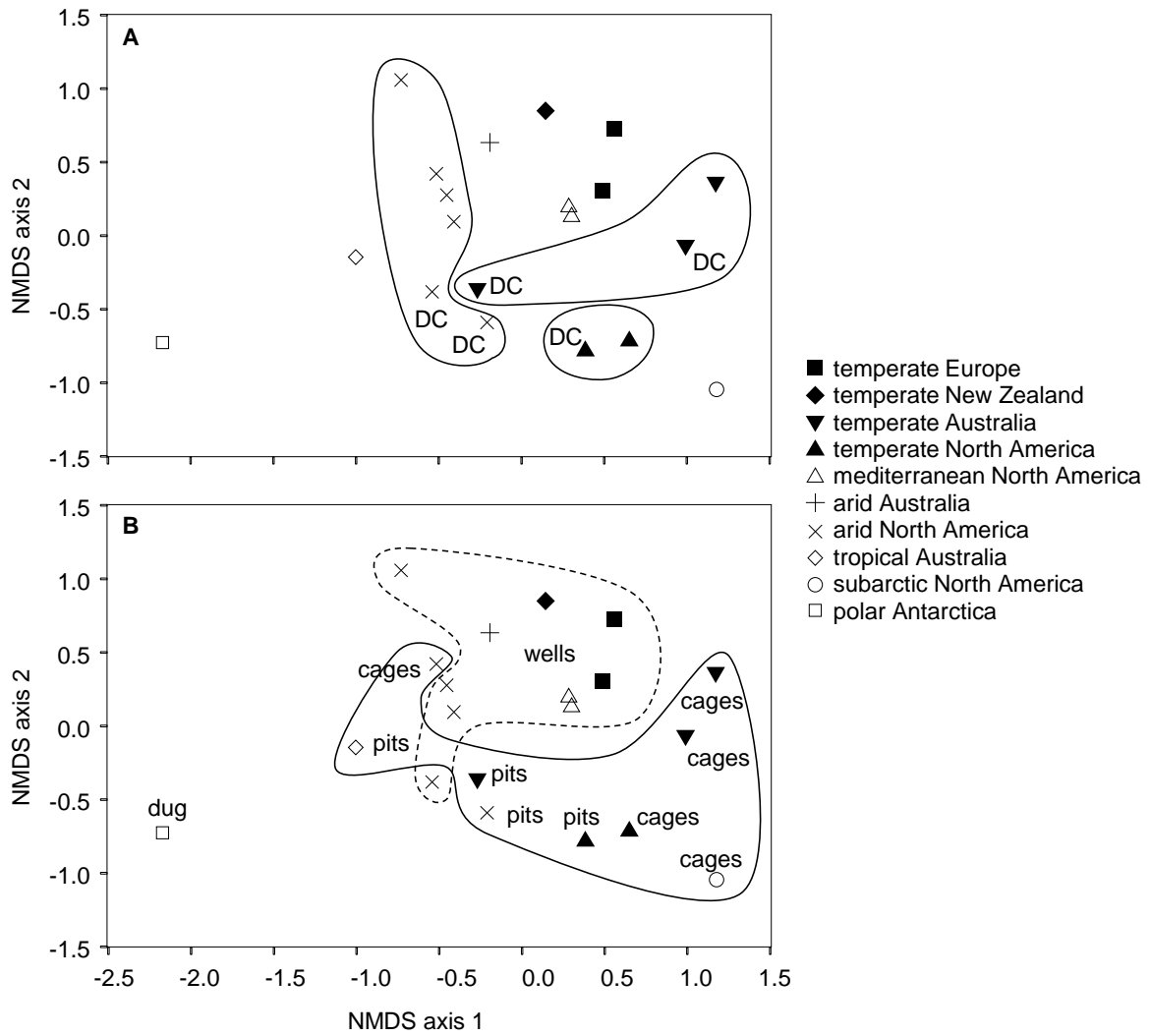
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711 Figure 2

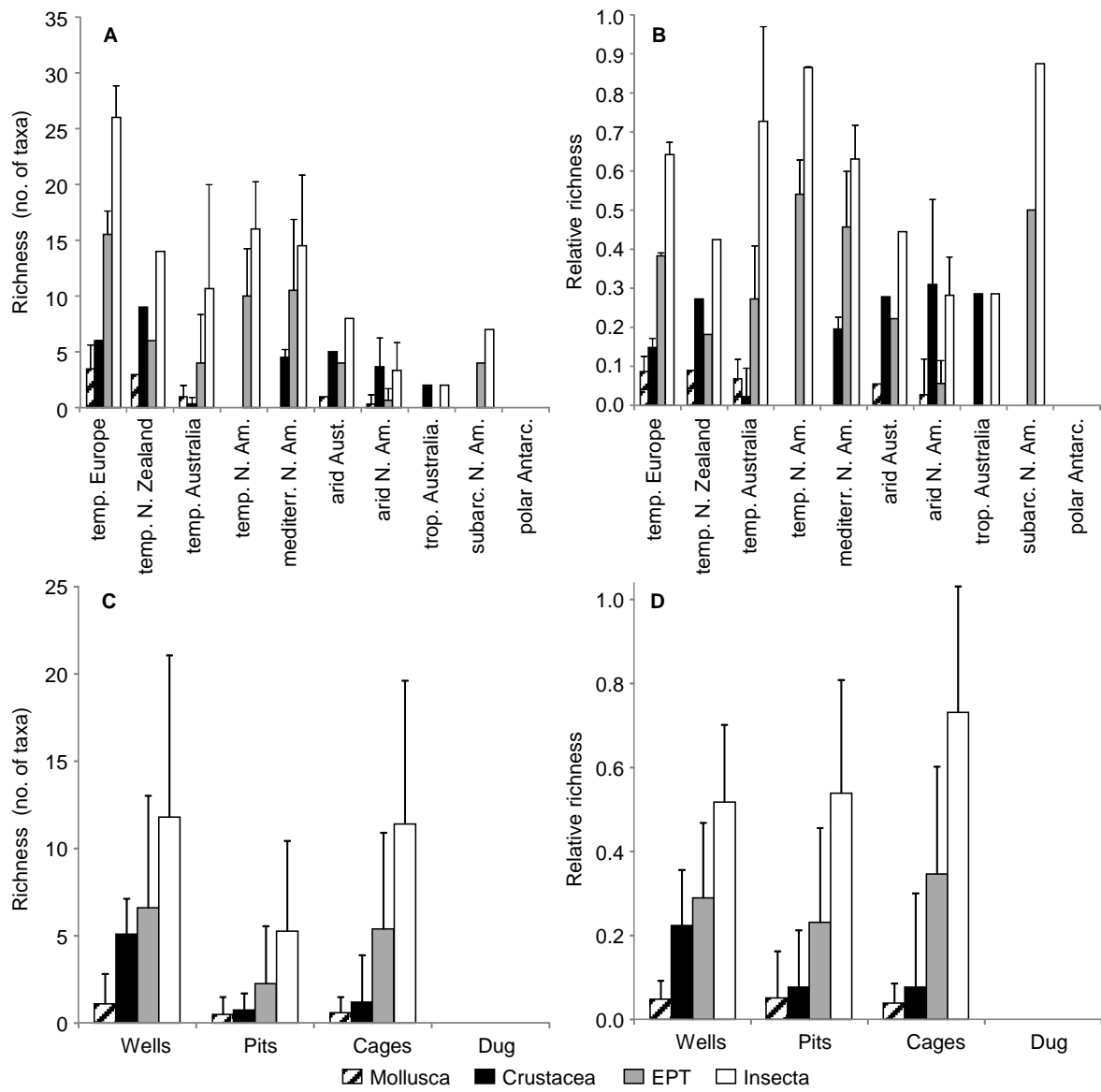
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714 Figure 3

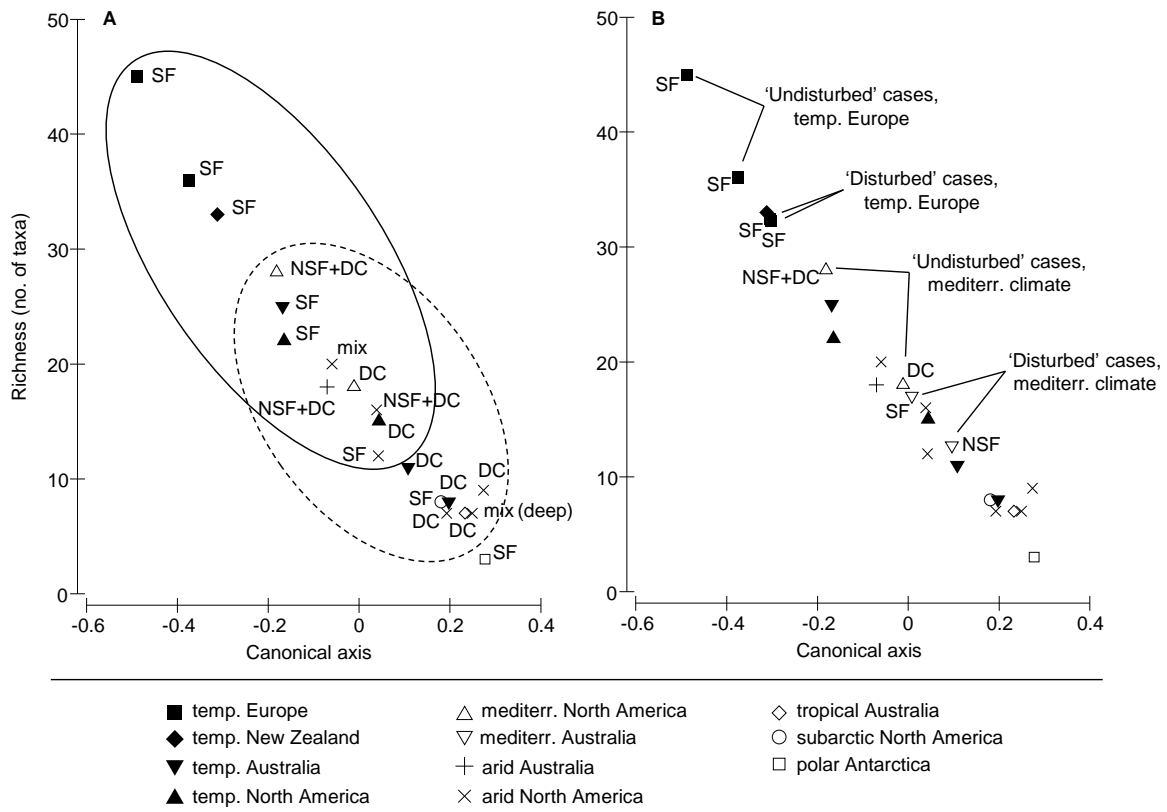
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717 Figure 4

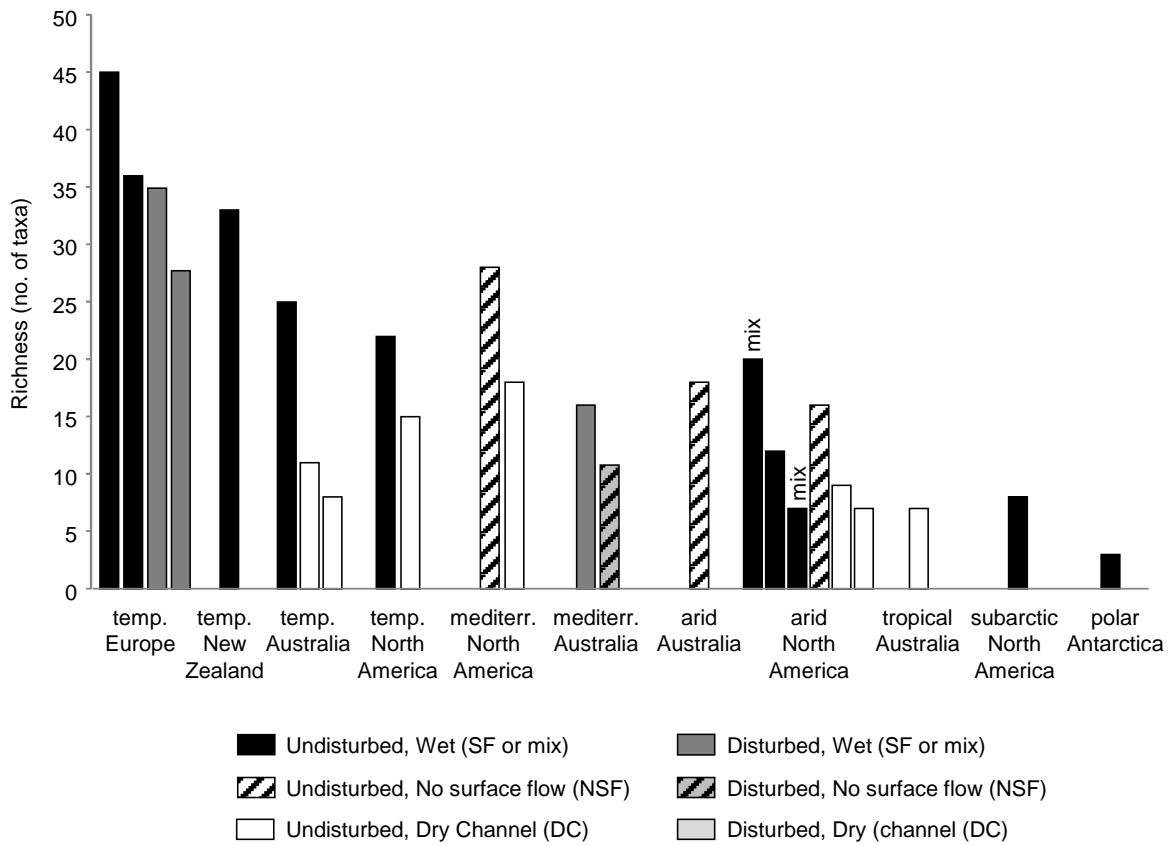
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719

720 Figure 5

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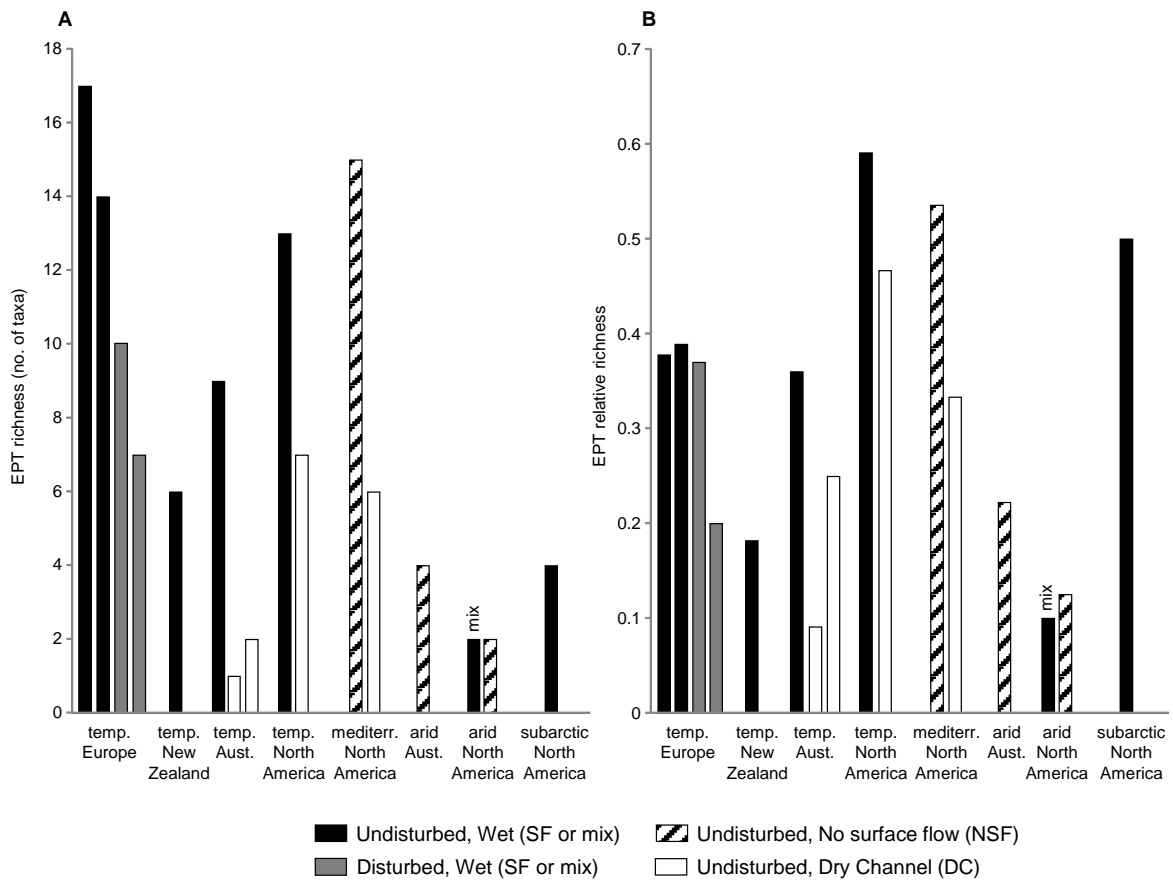


722

723 Figure 6

724

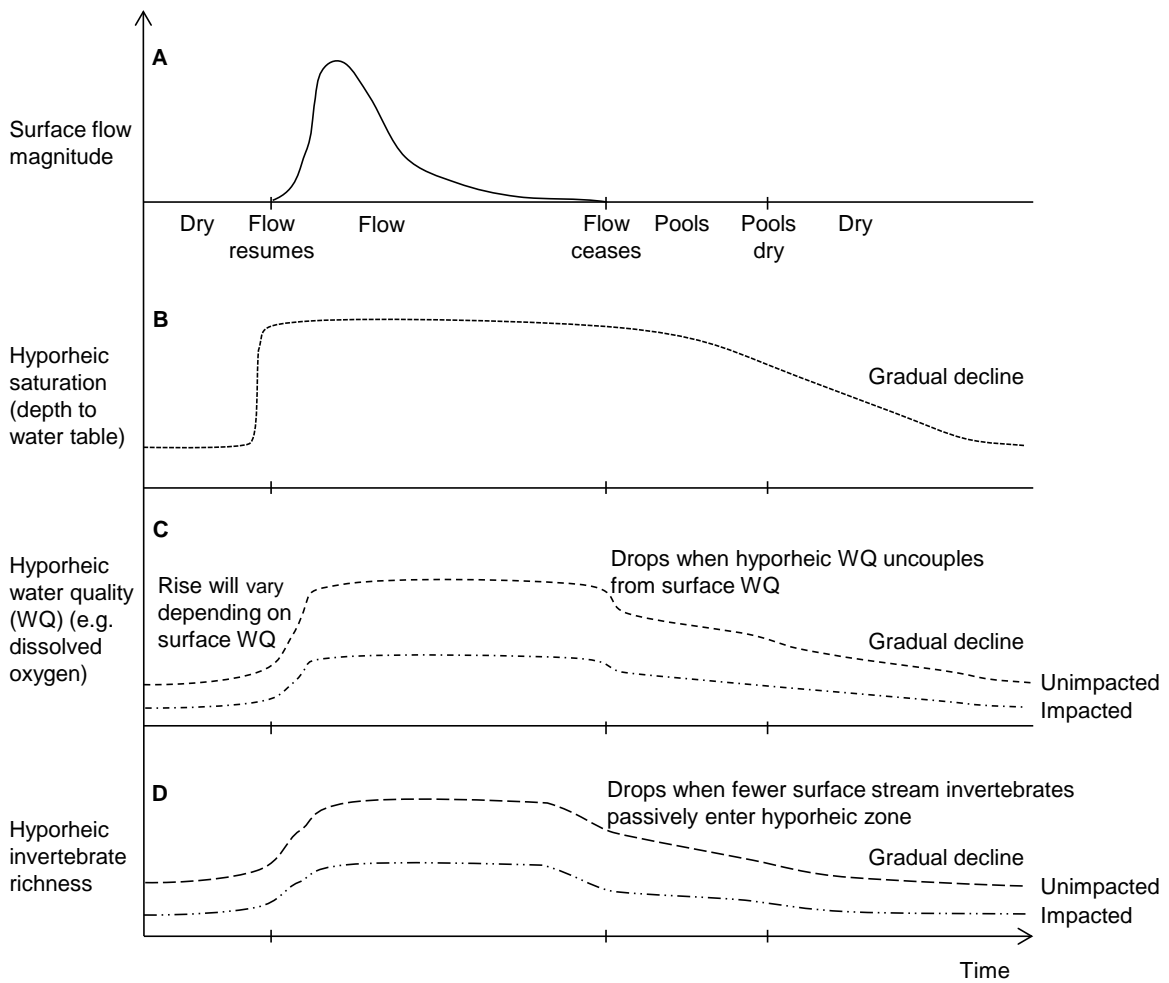




725

726 Figure 7

727



728

729 Figure 8