

## 1 **Abstract**

2 Evidence suggests that periods of heavy intense training can result in impaired immune cell  
3 function, whether this leaves elite athletes at greater risk of infections and upper respiratory  
4 symptoms is still debated. There is some evidence that episodes of upper-respiratory symptoms  
5 do cluster around important periods of competition and intense periods of training. Since  
6 reducing upper respiratory symptoms, primarily from an infectious origin, may have  
7 implications for performance, a large amount of research has focused on nutritional strategies  
8 to improve immune function at rest and in response to exercise. Although there is some  
9 convincing evidence that meeting requirements of high intakes in carbohydrate and protein and  
10 avoiding deficiencies in nutrients such as vitamin D and antioxidants is integral for optimal  
11 immune health, well-powered randomised controlled trials reporting improvements in upper-  
12 respiratory symptoms beyond such intakes are lacking. Consequently, there is a need to first  
13 understand whether the nutritional practices adopted by elite athletes increases their risk of  
14 upper respiratory symptoms. Second, promising evidence in support of efficacy and  
15 mechanisms of immune-enhancing nutritional supplements (probiotics, bovine colostrum) on  
16 upper respiratory symptoms needs to be followed up with more randomised controlled trials in  
17 elite athletes with sufficient participant numbers and rigorous procedures with clinically  
18 relevant outcome measures of immunity.

19

## 20 **Highlights**

- 21 - Evidence suggests that upper-respiratory symptoms in athletes typically cluster around  
22 intense periods of training, with greater risk during the winter months.

- 23 - Emerging evidence supports the use of probiotics and bovine colostrum to enhance  
24 immune health and reduce URS, to further elucidate mechanisms and efficacy well-  
25 powered randomised control trials in athletes are warranted
- 26 - Exercise in a state of mild hypohydration (1-3%) may not detrimentally affect mucosal  
27 immunity, with little evidence for an association between hypohydration and self-  
28 reported URS
- 29 - Randomised control trials to establish how periodised carbohydrate intake can impact  
30 self-reported URS and *in-vivo* measures of immune function in athletes are warranted
- 31 - Limited evidence to support to the use of protein and amino acid supplementation to  
32 reduced URS
- 33 - Antioxidant and vitamin D supplementation may be warranted in those who are  
34 deficient and exposed to extreme unaccustomed acute physical stress, however  
35 randomised control trials tracking changes in URS and immunological markers in  
36 athletes are needed.

37

### 38 **Key words**

39 Immunology, Nutrition, Exercise

40

### 41 **Introduction**

42 The relationship between infection risk and exercise training load has long been described as a  
43 J-shaped curved (Figure 1), with high training loads believed to increase the risk of  
44 opportunistic infections, particularly of the upper respiratory tract (Nieman, 1994). However,  
45 there is limited empirical evidence that elite athletes experience more infections than the  
46 general population (Svendsen, Taylor, Tonnessen, Bahr & Gleeson, 2016). Based on re-

47 evaluation of published data, Malm (2006) proposed that elite training is associated with a  
48 lower susceptibility to infection compared to high exercise workloads, whereby the relationship  
49 between infection risk and training load instead resembles an S-shaped curve (Figure 1)(Malm,  
50 2006). Whilst this remains to be verified in prospective studies, it is hypothesised that to  
51 maintain elite athlete status there is a pre-requisite to have a robust immune system capable of  
52 withstanding infections even during heightened physical and psychological stress. However, it  
53 may be that the reduced infection risk in elite athletes observed in previous studies may not  
54 reflect a lower impact of physical stress (exercise workload) per se but rather a reflection of  
55 better preventive and treatment strategies in place within the studied elite settings.  
56 Nevertheless, although elite athletes may not experience a greater annual incidence rate of  
57 infections, there is increasing recognition that episodes of upper-respiratory symptoms (URS)  
58 typically cluster around intense periods of training (Hellard, Avalos, Guimaraes, Toussaint &  
59 Pyne, 2015; Moreira, Delgado, Moreira & Haahtela, 2009; Svendsen, Gleeson, Haugen &  
60 Tønnessen, 2015; Svendsen et al., 2016) with a greater risk during winter months (Hellard et  
61 al., 2015; Spence et al., 2007).

62 URS is the most common medical complaint affecting athletes, and with medals often being  
63 decided by the smallest of margins, even minor illnesses can have a meaningful, negative  
64 impact on competition outcomes. Indeed, fewer days of illness appears to be one factor that  
65 differentiates World and Olympic medallists from other international-level athletes (Svendsen  
66 et al., 2015). URS in athletes is likely to involve both infectious and non-infectious causes,  
67 previous reports suggest between 31% (Spence et al., 2007) and 82% (Hanstock et al., 2016)  
68 of URS episodes during winter months occur with an infectious pathogen. Non-infectious URS  
69 in athletes may be related to allergic rhinitis, asthma and/or exercise-induced  
70 bronchoconstriction.

71 Since reducing URS, particularly of infectious origin, may have implications for athletic  
72 performance, it is not surprising that a large amount of research has focused on nutritional  
73 strategies to improve immune function at rest, and/or favourably modify the immune response  
74 to exercise. Although the impact of a chronically high training load on immune function is still  
75 debated, it is documented that a single bout of prolonged, intense exercise transiently modifies  
76 a large number of immune variables. Following intense exercise an individual's capacity to  
77 defend against pathogens is altered, resulting in what is referred to as an "open window" for  
78 infectious causes of URS, lasting up to 72 hours post-exercise depending on the intensity and  
79 duration of the exercise, and the immune marker measured (Moreira et al., 2009).

80 Quantifying immunocompetence in athletes in the field and identifying changes that predict  
81 infection risk as a result of interventions is challenging. The gold standard (or most relevant  
82 outcome) may be clinical symptoms such as whether or not an individual actually contracts an  
83 infection, as confirmed by pathological tests, assuming that pathogen exposure is similar across  
84 intervention groups. However, in order to elucidate the underlying mechanisms that mediate  
85 any potential changes in infection risk following a nutrition intervention, it would also be  
86 pertinent to include immunological markers in laboratory research trials.

87 This short review will provide an updated summary on selected immune nutrition topics  
88 including dietary carbohydrate and protein intake, hydration status, antioxidants, vitamin D,  
89 bovine colostrum, and probiotics, including when evidence is available practical  
90 recommendations for the sport and exercise nutrition practitioner and/or athlete and coach  
91 (Table 1). For a more in-depth review, readers are referred to the latest consensus statement of  
92 the ISEI (Bermon et al., 2017).

93

94 ***Carbohydrate***

95 It is well documented that carbohydrate intake is a fundamental part of an athlete's diet, both  
96 in its ability to enhance physical performance and its role in recovery following exercise.  
97 Athletes adopting a train-low compete-high CHO intake approach may increase their risk of  
98 immune impairment during periods of restricted CHO intake (Burke, 2010). Previous work has  
99 shown that participants on low carbohydrate diets (less than 10% of energy from CHO) for 48  
100 – 72 hrs have larger circulating stress hormone (cortisol, adrenaline) and cytokine (IL1ra, IL6,  
101 IL10) responses when compared to normal or high CHO diets (Bishop, Walsh, Haines,  
102 Richards & Gleeson, 2001b). Whilst it should be noted that it is unusual for elite athletes to  
103 regularly have intakes as low as that outlined in the Bishop paper (<10% of daily energy intake  
104 from CHO equated to ~1.1 g/kg body mass per day); it is not unheard of for this to be the case  
105 for short periods. Athletes may adopt a low CHO intake lasting 1-3 days in specific scenarios  
106 e.g. making weight or during periods of tapering where training volume may decrease for  
107 competition preparations (Reale, Slater & Burke, 2017). In contrast, high CHO diets have been  
108 associated with blunted plasma cortisol responses to exercise due to preservation of plasma  
109 glucose, better maintenance of post-exercise plasma glutamine concentrations and attenuated  
110 exercise-induced disturbances in the number of circulating leukocytes, neutrophils and  
111 lymphocytes (Bishop, Walsh, Haines, Richards & Gleeson, 2001a; Gleeson, Blannin, Walsh,  
112 Bishop & Clark, 1998).

113 The majority of evidence from the literature suggests that increasing CHO availability will act  
114 indirectly to reduce the stress hormone response to exercise and therefore limit exercise-  
115 induced immune impairment. There is also some evidence to suggest the beneficial effects of  
116 consuming CHO during exercise can occur without any effect of plasma cortisol levels,  
117 although this is likely to be dependent upon intensity and duration (Green, Croaker &  
118 Rowbottom, 2003). Sixty grams per hour of CHO attenuates the rise in plasma cytokines during  
119 exercise, and reduces the trafficking of leukocyte subsets during prolonged (2.5h) endurance

120 running (Henson et al., 1998). In contrast, however, CHO feeding during marathon running  
121 appears ineffective in altering salivary secretory immunoglobulin-A secretion and reducing  
122 self-reported symptoms of URS (Nieman et al., 2002). Recent evidence also found that acute  
123 CHO ingestion before, during and after prolonged exercise had no benefit on in vivo immune  
124 responses with a novel antigen (Davison, Kehaya, Diment & Walsh, 2016).

125 Whilst a substantial body of evidence supports the influence of carbohydrate availability (in  
126 terms of dietary intake or acute supplementation) on stress hormone responses and in vitro  
127 markers of immune function (Bishop, Walsh, Haines, Richards & Gleeson, 2001a; Gleeson et  
128 al., 1998), evidence is lacking with use of clinically relevant outcomes (integrated in vivo  
129 measures, incidence of URS). Despite the lack of studies showing a benefit of CHO on URS it  
130 is important to acknowledge that CHO intake and supplementation have consistently been  
131 shown to enhance aspects of performance and recovery. Subsequently if athletes achieve  
132 recommended intakes for CHO (Table 1) during training and competition, it may help control  
133 for any proposed impact of CHO availability on immune function. Periodised CHO intake to  
134 match training intensity and competition with periods of restriction to enhance training  
135 adaptations (Bartlett, Hawley & Morton, 2015; Burke, 2010) could offer a suitable compromise  
136 between fuelling and enhancing training adaptations and limiting the negative effects of low  
137 CHO availability upon the stress hormone response.

138

### 139 ***Protein***

140 Immune function is reliant on rapid cell replication and generation of proteins such as cytokines  
141 and immunoglobulins. It is therefore not surprising that inadequate protein intake has been  
142 associated with compromised host defence and susceptibility to opportunistic infection.  
143 Although athletes typically appear to consume adequate amounts of protein, some individuals

144 may experience sub-optimal protein intake, for example during periods of heavy training and/or  
145 weight loss. Even in endurance athletes meeting the recommended daily protein intake of 1.6  
146 g/kg body mass (Tarnopolsky, 2004), there may be immunological benefits of further  
147 increasing dietary protein during intense training periods. Witard et al. (2014) found that during  
148 intensified training increasing daily dietary protein intake from 1.5 to 3 g/kg body mass (whilst  
149 maintaining a carbohydrate rich diet of 6 g/kg body mass per day) blunted the exercise-induced  
150 impairment in CD8+ T lymphocyte, total leukocyte and granulocyte redistribution observed in  
151 participants following a lower protein diet (Witard et al., 2014). Importantly the high protein  
152 diet resulted in significantly fewer self-reported URS, with the authors proposing that  
153 consumption of a high protein diet may help maintain immune surveillance during high-  
154 intensity training. The exact dose for optimal immunological benefits during periods of  
155 intensified training may be somewhere between 1.5 and 3g/kg body mass, depending on the  
156 type of exercise conducted, but further research is warranted to establish clearer  
157 recommendations.

158 There has also been interest in single amino acid supplementation to influence immune  
159 competence in athletes. Specifically the non-essential amino acid glutamine is an important  
160 fuel for immune cells in particular lymphocytes and macrophages (Castell & Newsholme,  
161 1997). Endurance exercise can reduce plasma glutamine (Castell & Newsholme, 1998) and it  
162 has been hypothesized that oral glutamine supplementation may enhance post-exercise  
163 immunity (Castell & Newsholme, 1997). However, the majority of studies have found that  
164 reduced plasma glutamine does not meaningfully contribute to exercise-induced immune  
165 impairment (Gleeson, 2008). Despite an attractive hypothesis, there is little evidence that  
166 glutamine supplementation influences immune responses to exercise.

167 There remains a lack of randomised controlled trials assessing increased protein intake upon  
168 self-reported URS, and despite some mechanistic evidence for glutamine supplementation

169 showing immune benefits, to date there is limited evidence that glutamine supplementation is  
170 effective in abolishing the post-exercise immune cell impairment and URS.

171

## 172 *Hydration*

173 It is not unusual for athletes to commence training in a pre-existing fluid deficit. A combination  
174 of factors can lead to this predisposition, including failure to rehydrate between sessions or  
175 specific weight making strategies. The potential negative effects of hypohydration during  
176 exercise in laboratory settings are well documented; increased cardiovascular strain, elevated  
177 core temperature and increased perception of effort (Sawka & Coyle, 1999). Furthermore, a  
178 pre-exercise fluid deficit of as little as a 1.5-2.0% body mass loss (BML) has been suggested  
179 to negatively affect laboratory based exercise trials (Maughan & Shirreffs, 2004). In contrast,  
180 recent evidence suggests that in appropriate and representative environmental conditions of  
181 outdoor exercise where effective evaporative cooling can be maintained body mass losses of  
182 up to 3% can be well tolerated and have little negative impact upon exercise performance (Wall  
183 et al., 2015).

184 With regard to the immune response to moderate hypohydration there appears to be no  
185 negative effect on total or differential leukocyte numbers, lymphocyte function (Mitchell,  
186 Dugas, McFarlin & Nelson, 2002), neutrophil function, or antigen-stimulated cytokine  
187 production (Svendsen, Killer & Gleeson, 2014). As such, undertaking exercise in a  
188 hypohydrated state, at least within the range generally applicable to athletes, does not appear  
189 to have meaningful implications for cellular immunity.

190 In contrast, significant reductions in salivary flow rates have been observed in exercising  
191 participants at a BML of 1.3-3% (Killer, Svendsen & Gleeson, 2015). Saliva contains numerous  
192 antimicrobial proteins (AMPs) that play an important role in mucosal immunity, and reductions

193 in salivary flow rate may therefore have implications for host defence. To date, secretory IgA  
194 (SIgA) has been the most studied marker of mucosal immunity within athletic populations. The  
195 importance of salivary lysozyme (SLys) and salivary lactoferrin (SLac) have also gained  
196 recognition as both are present in mucosal secretions of the upper respiratory tract and  
197 understood to play an integral role in the innate immune system.

198 Fortes and colleagues investigated SIgA and SLys following exercise-induced dehydration and  
199 subsequent overnight fluid restriction (3% BML) (Fortes, Diment, Di Felice & Walsh, 2012).  
200 Dehydration resulted in a significant decrease in SIgA concentration, with no change in  
201 secretion rate and conversely, no change in SLys concentrations but a significant reduction in  
202 secretion rate. Research has also identified transient changes in salivary AMPs during and  
203 immediately post-90 min exercise following 24 h fluid restriction, which had mostly returned  
204 to baseline values by 3 h post-exercise (Killer et al., 2015). Exercise in a state of mild  
205 hypohydration caused a reduction in saliva flow rate, yet induced greater secretion rates of  
206 SLac and higher concentrations of SIgA and SLys.

207 These data suggest that prolonged exercise in a state of mild hypohydration (1-3%) may not  
208 detrimentally affect mucosal immunity. Whilst there remains a lack of evidence into incidence  
209 of URS and hydration status, it is unlikely that the reported small transient fluctuations in  
210 salivary AMPs would translate into clinical relevance. Furthermore, inconsistencies in the  
211 measurement of AMP concentrations vs secretion rates, variation in dehydration protocols  
212 (fluid restriction vs exercise-induced) and a wide range of levels of dehydration (percentage  
213 BML) are likely to contribute to the lack of clarity around the impact of hydration on mucosal  
214 immunity, in particular when deciphering any clinical significance. Future research should look  
215 to address some of these issues and establish if exercise-induced hypohydration in a range of  
216 environments (laboratory and field) can have a detrimental impact upon exercise-induced  
217 immune impairment.

218

219 *Antioxidants*

220 Strenuous exercise is associated with an acute increase in the production of free radicals  
221 (reactive oxygen species (ROS), and reactive nitrogen species (Powers, Nelson & Hudson,  
222 2011). An endogenous network of enzymatic (e.g. superoxide dismutase, glutathione  
223 peroxidase, catalase) and non-enzymatic antioxidants (e.g. vitamins A, C and E) exist to  
224 provide intracellular and extracellular protection against oxidant damage (Powers, Deruisseau,  
225 Quindry & Hamilton, 2004). Whilst it is acknowledged these antioxidant defences adapt with  
226 training, it has long been debated whether they are sufficient to counter oxidant production  
227 during strenuous exercise. Early investigations highlighting the damaging effects of oxidants  
228 on muscle and cells led to a proposed role of antioxidant status in exercise-induced immune  
229 dysfunction following prolonged exercise (Powers et al., 2011).

230 Of all the potential exogenous antioxidant supplements, the essential nutrient vitamin C has  
231 received the greatest attention as a strategy to support immune health in athletes (Nieman et  
232 al., 2002). Initially, interest was also partly due to preliminary evidence of the prophylactic  
233 benefit of vitamin C on the common cold. The current evidence, however, provided by the  
234 latest Cochrane review, reports that routine vitamin C supplementation (> 0.2 g per day) does  
235 not reduce the risk of developing a cold in the general population but such regular  
236 supplementation (as opposed to upon onset of symptoms) appears to reduce the duration and  
237 severity of colds (Hemilä & Chalker, 2013). In contrast, pre-specified sub-group analysis of  
238 trials in this review concluded that there is firm evidence that vitamin C supplementation  
239 between 0.25 and 1.0 g/day results in reduced number of participants reporting URS under  
240 periods of physical stress with or without cold stress (marathon runners, skiers and soldiers on  
241 subarctic operations). The underlying mechanism(s) of such effects remains unclear,

242 particularly as any role of exercise-induced oxidant production in alterations of immune  
243 dysfunction has not been shown consistently (Nieman et al., 2002). Additional evidence has  
244 purported benefits of vitamin C in non-infectious causes of URS (e.g. exercise-induced  
245 bronchoconstriction) following exercise (Hemilä, 2013).

246 Investigation of other essential nutrients with antioxidant potential (e.g. vitamin E) or multiple  
247 vitamins have largely been unsuccessful with concerns over pro-oxidant/pro-inflammatory  
248 effects in large doses or interference with the role of ROS in key signalling processes of training  
249 adaptation (Nieman & Mittlemeier, 2017). Focus in this area has shifted towards other  
250 nutritional compounds in the human diet such as polyphenols, albeit the emerging evidence of  
251 the effects of these interventions on URS risk in athletes also appear to be independent of any  
252 antioxidant properties (e.g. direct anti-pathogenic pathways) (Somerville, Braakhuis &  
253 Hopkins, 2016). There is lack of conclusive evidence that exercise-induced oxidant production  
254 is detrimental to athlete health, including host defence. Nevertheless, the additional evidence  
255 of the effect of vitamin C on duration and severity of URS means evaluation on an individual  
256 athlete basis may be clinically worthwhile. The evidence of higher regular intake of vitamin C  
257 and reduced incidence of URS, however, should not be ignored. It is important to stress that  
258 these benefits were evident within a range of doses (0.25 – 1.0 g per day) that were not  
259 particularly high, and thus excess consumption may be easily achieved through use of over-the-  
260 counter vitamin C supplements. It appears that these benefits are only apparent in those exposed  
261 to short-term unaccustomed physical stress. Such findings may have limited application to the  
262 trained athlete who has regular (long-term) exposure to such stress (i.e. training and  
263 competition).

264

265 ***Vitamin D***

266 Over the past decade, there has been emerging evidence highlighting the role that vitamin D  
267 may have in athlete health (Owens, Fraser & Close, 2015). Commonly known for its role in  
268 bone health (Ebeling, 2014) and muscle function (Owens et al., 2015) it is also increasingly  
269 recognised for its role in inflammation and aspects of innate and acquired immunity (He et al.,  
270 2016).

271 Unlike other vitamins that are primarily obtained through diet, physiological sufficiency for  
272 vitamin D can be met through endogenous synthesis via UV irradiation of the skin's dermis.  
273 The cutaneous production of vitamin D is highly variable and dependent upon both  
274 environmental and individual factors. These include season, time of day, amount of cloud  
275 cover, skin pigmentation, age, clothing, and use of high-factor sunscreen (Chen et al., 2007).  
276 Furthermore, vitamin D synthesis drops in winter months at latitudes greater than 35-37° due  
277 to insufficient UVB photons reaching the Earth's surface (Webb, Kline & Holick, 1988).

278 Adequate concentration has been previously defined as serum 25 hydroxy vitamin D  
279 (25(OH)D) >50 nmol/L by the US Institute of Medicine. However, within the literature, there  
280 is a lack of consensus as to what constitutes vitamin D deficiency and what might be classified  
281 as an insufficiency for elite athlete health and performance. It is beyond the scope of this review  
282 to discuss what constitutes sufficient or optimal circulating concentrations of 25(OH)D, so  
283 readers are referred to He et al (2016) for more information (He, Aw Yong, Walsh & Gleeson,  
284 2016). The prevalence of deficiency and sufficiency in athletes varies by, training location,  
285 sport (Larson-Meyer & Willis, 2010) and skin colour (Pollock, Dijkstra, Chakraverty &  
286 Hamilton, 2012), with deficiency being greater in the winter months (Farrokhyar et al., 2015).  
287 There is growing evidence that vitamin D likely plays a key role in both innate and acquired  
288 immunity through its modulation of gene expression (Kamen & Tangpricha, 2010). Vitamin  
289 D upregulates gene expression of antimicrobial peptides, which are important regulators in  
290 innate immunity, and downregulate expression of inflammatory cytokines (He et al., 2016).

291 Furthermore, vitamin D is also found to have an immunomodulatory effect on T and B-  
292 lymphocytes in acquired immunity (Von Essen et al., 2010).

293 A small number of studies have reported negative associations between vitamin D  
294 concentration and self-reported URS in athletes (He et al., 2013) and military personnel (Laaksi  
295 et al., 2007). In a study of endurance athletes, those in a Vitamin D deficient status group  
296  $25(\text{OH})\text{D} < 30\text{nmol/L}$ , reported greater number of URS days and higher symptom-severity  
297 scores compared to counterparts with greater circulating vitamin D concentrations (He et al.,  
298 2013). Elite athletes reporting with URS who had a positive virology/bacteriology result  
299 (infectious group) or athletes with a mild to moderate leucocytosis (suggestive group) had  
300 significantly lower levels of circulating  $25(\text{OH})\text{D}$  levels than athletes with a negative  
301 virology/bacteriology count and normal differential leukocyte count (Cox et al., 2008). In a  
302 military setting young Finnish conscripts who had low circulating  $25(\text{OH})\text{D}$  concentrations  
303 (defined by the authors as  $<40\text{ nmol/L}$ ) had significantly more duty days lost to respiratory  
304 infection during 6 months of training and were 1.6 times more likely to miss duty due to  
305 respiratory infection than those with a circulating  $25(\text{OH})\text{D} >40\text{ nmol/L}$  (Laaksi et al., 2007).

306 Although causality cannot be established from these cross-sectional comparison studies of  
307 physically active individuals, they are in agreement with RCTs of general populations that  
308 show reduced respiratory infections with daily or weekly vitamin D supplementation,  
309 particularly in those with deficiency ( $< 25\text{-}30\text{ nmol/L}$  circulating  $25(\text{OH})\text{D}$ ) (Berry, Hesketh,  
310 Power & Hyppönen, 2011).

311

### 312 ***Bovine colostrum***

313 Bovine colostrum (COL) is the initial milk produced by a cow in the first few days following  
314 parturition. In addition to a different composition of macronutrients (higher percentage of

315 protein, lower percentage of lactose and fat) compared to mature milk (Ontsouka, Bruckmaier  
316 & Blum, 2003), COL is richer in antimicrobial, growth and immune factors (Uruakpa, Ismond  
317 & Akobundu, 2002). In fact, the bioactivity of COL is at its greatest in the first milking with  
318 the concentrations of such components decreasing over the subsequent days (Korhonen,  
319 Marnila & Gill, 2000). Although sharing a homologous composition to human colostrum, the  
320 concentrations of immune factors in COL are in vastly greater concentrations (Shing, Peake,  
321 Suzuki, Jenkins & Coombes, 2013). Such bioactivity has led to suggestions that COL could  
322 enhance human immune function and hence aid prophylaxis of infections.

323 A recent meta-analysis (Jones, March, Curtis & Bridle, 2016) of five randomised controlled  
324 trials concluded COL supplementation reduces the incidence rate of episodes and total number  
325 of days of URS during exercise training (cyclists, distance runners, recreational athletes,  
326 swimmers). The magnitude of these reductions (URS days: 44%; URS episodes 38%) are  
327 greater than the smallest clinically important difference, but the low precision of the individual  
328 study estimates (as a result of small sample sizes and hence low number of events) means that  
329 further trials will likely change the best estimate of the average effect of COL. The minimum  
330 and/or the optimum dose of COL for benefit on incidence of URS is yet to be confirmed, but  
331 there is preliminary tentative (observational) evidence suggesting 20 g per day may result in  
332 superior protection than 10 g (Jones et al., 2016) (Table 1). There remains a lack of evidence  
333 to determine whether COL supplementation can reduce duration or severity of URS episodes  
334 in athletes.

335 Given the somewhat uncertainty surrounding the causes of self-reported URS with exercise,  
336 there may be a number of potential mechanisms responsible for the effects of COL on URS  
337 during exercise training. In-depth reviews of the underlying mechanisms in the effects of COL  
338 have been discussed extensively elsewhere (Bermon et al., 2017; Davison, 2012). Briefly, one  
339 proposed mechanism is that the small bioactive constituents of COL, or their metabolites,

340 appear in the circulation after consumption and have immune-enhancing effects on host  
341 immunity (Jones et al., 2016). Recently, COL supplementation induced greater sensitivity of  
342 in vivo immune responses to a novel antigen (experimental CHS) following prolonged exercise  
343 (Jones et al., 2018). Whilst recognising that the specific mechanisms of action of COL may  
344 differ between populations, reports of reduced incidence of respiratory infections in other at  
345 risk groups (with immune deficiency/recurrent infections) are further examples of available  
346 evidence supporting an hypothesis that use of COL can lead to changes in host defence against  
347 pathogenic causes of URS (Cesarone et al., 2007; Patiroğlu & Kondolot, 2013).

348

#### 349 *Probiotics and Prebiotics*

350 The human intestine represents the largest mass of lymphoid tissue in the body and is resident  
351 to thousands of bacterial taxa (Wylie et al., 2012). The adult gastrointestinal immune system  
352 comprises of a stable alliance among the commensal microbiota, immune mediators, and the  
353 epithelial barrier. All three components are essential for function and maintenance of a stable  
354 and mature intestinal immune system. Nutritional supplementation to support the gut  
355 microbiota is a proposed means to maintain immune competence and reduce URS risk (Hao,  
356 Dong & Wu, 2015).

357 It is now recognised that the species composition of the microbiota can be modified by  
358 alterations in dietary intake. Regular consumption of probiotic bacteria can positively modify  
359 the composition of the gut microbiota and influence immune health (Round & Mazmanian,  
360 2009). Alternatively, a prebiotic, a selectively fermented ingredient that allows specific  
361 changes, both in the composition and/or activity in the gastrointestinal microbiota can confer  
362 benefits upon host well-being and health (Gibson et al., 2017). However, to date there are  
363 currently no published studies on the efficacy of prebiotics to reduce URS in athletes. Non-

364 infectious causes of URS such as exercise-induced bronchoconstriction has shown a favourable  
365 response to prebiotics (Williams et al., 2016).

366 A large number of studies have been conducted investigating the effects of probiotics on URS  
367 in the non-athlete general population, and readers are referred to the latest Cochrane systematic  
368 review for more detail (Hao et al., 2015). The review concluded that probiotics were better than  
369 placebo with fewer participants experiencing at least one episode of acute URS, but there was  
370 no difference when measuring rate of episodes of URS or the duration of episodes (Hao et al.,  
371 2015).

372 With regard to probiotic use and athletes, there are few well-conducted large scale randomised  
373 controlled trials, but readers are referred to a 2015 review of the probiotic literature in athletes  
374 for more detail (Pyne et al., 2015). Briefly, they identified 15 relevant experimental studies  
375 from 2006 to 2014 that investigated the clinical and immunological effects of probiotic  
376 supplementation in trained individuals; five randomised placebo controlled studies reported  
377 reductions in self-reported URS frequency, with three reporting trivial to no effects (Pyne et  
378 al., 2015).

379 A randomised crossover trial showed benefit from a daily dose of  $1.3 \times 10^{10}$  colony forming  
380 units (CFU) *Lactobacillus fermentum* for 28 days in distance runners during a winter training  
381 period (Cox, Pyne, Saunders & Fricker, 2010). The number of days, and severity of self-  
382 reported URS was less (~50%) in those receiving the probiotic compared to placebo. This was  
383 coupled with a two-fold greater change in whole blood culture interferon- $\gamma$  with the probiotic;  
384 however, there were no changes in salivary IgA, or IL-4 and IL-12 (Cox et al., 2010). Further  
385 evidence in support of probiotic feeding showed that 16 weeks of *Lactobacillus casei Shirota*  
386 ( $1.3 \times 10^{11}$  cells per day) reduced the proportion of active individuals reporting URS by 36%,  
387 reduced the number of URS episodes (1.2 vs 2.1), and increased salivary IgA over the course

388 of the study (Gleeson, Bishop, Oliveira & Tauler, 2011). However, there was no difference in  
389 duration of symptoms (Gleeson et al., 2011). In contrast, a follow up study using a probiotic  
390 bacteria strain of *Lactobacillus salivarius*,  $2 \times 10^{10}$  CFU for 16 weeks failed to reduce the  
391 frequency of URS in an athletic cohort or modify markers of immune function (Gleeson et al.,  
392 2012), highlighting issues with strain specificity. Further issues arise with potential sex  
393 differences in responsiveness to probiotic treatment as 11 weeks of *Lactobacillus fermentum*  
394 ( $1.0 \times 10^9$  cells per day) was able to reduce illness load (severity x duration) of URS by 31% in  
395 males but not females (West et al., 2011).

396 It appears there is a growing body of evidence that probiotic supplementation may be beneficial  
397 in reducing the frequency of URS during periods of high training load. A greater number of  
398 well-controlled studies with probiotics are required to clarify dose response, strain choice and  
399 elucidate mechanisms of action within athlete populations. Furthermore, prebiotics, which act  
400 by increasing the growth and activity of non-pathogenic commensal bacteria at a genus level  
401 maybe a viable alternative or have an additive effect as a synbiotic (combined probiotic and  
402 prebiotic intervention) and research into their use is warranted.

403

#### 404 ***Conclusions***

405 The risk of URS in athletes typically cluster around important periods of travel, competition  
406 and intense periods of training. In order to limit the detrimental effects of URS on training  
407 completion or competitive performance, elite athletes seek strategies to prevent or manage such  
408 events. There is a need to understand whether the nutritional practices adopted by elite athletes'  
409 increase their risk of URS. The nutritional interventions discussed in this review show some  
410 promising mechanistic evidence for an immunomodulatory effect within athletes, yet well-  
411 powered randomised controlled trials reporting reduced incidence in URS are not widely

412 available. There is need for more randomised controlled trials to establish the efficacy of  
413 nutrient interventions in elite athletes with sufficient participant numbers, rigorous procedures  
414 and use of validated assessment of clinical symptoms confirmed with pathological tests where  
415 appropriate. Studies investigating interventions with purported immune modulatory  
416 mechanisms of action are recommended to couple measurement of URS with clinically  
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629 Table 1 summary of the effects of nutritional interventions on upper respiratory symptoms in athletes and practical recommendations

Nutrient	Markers of immune function			Clinical symptoms (URS)	Research recommendations	Practical recommendations
	In vitro	Ex vivo	In vivo			
CARBOHYDRATE	[✓]	[✓]	[✱]	<p>●●●●●</p> <p>Lack of RCTs investigating effect on URS</p>	Further RCTs to establish if training in a low CHO state or CHO feeding during and after prolonged exercise impacts on URS and integrated in vivo measures of immune function	<p>Athletes consuming 30–60 g CHO per hour during sustained intensive exercise will aid the demands of physical and metabolic recovery. Immediately post exercise (0-2 hours) athletes are recommended to consume 1.0-1.2 g/kg body mass, however the absolute amount should be adjusted depending on the nature of the training session they have completed and the duration of the recovery period before the next training session.</p> <p>These intakes may attenuate the rise in stress hormones and indirectly limit the degree of exercise-induced immune impairment.</p> <p>Athletes undergoing train-low strategies should carefully periodise these sessions within their season to limit any potential impact this may have on immunity and thus on their ability to perform in competition.</p>
PROTEIN/AMINO ACIDS	[✓]	[✓]	[✱]	<p>◎●●●●</p> <p>Limited number of RCTs showing benefit of additional total protein or glutamine supplementation.</p>	Further RCTs to establish if additional total protein or glutamine supplementation impacts on URS and integrated in vivo measures of immune function	<p>Athletes are recommended to consume adequate daily amounts of protein (1.2 - 1.7 g/kg body mass), depending on the nature of their training, to help maintain sufficient whole body protein metabolism. Subsequently this may support correct immune function.</p> <p>It should also be noted that the pattern of ingested protein can affect whole-body protein metabolism, ~20-30 g at regular (~3 h) intervals throughout the</p>

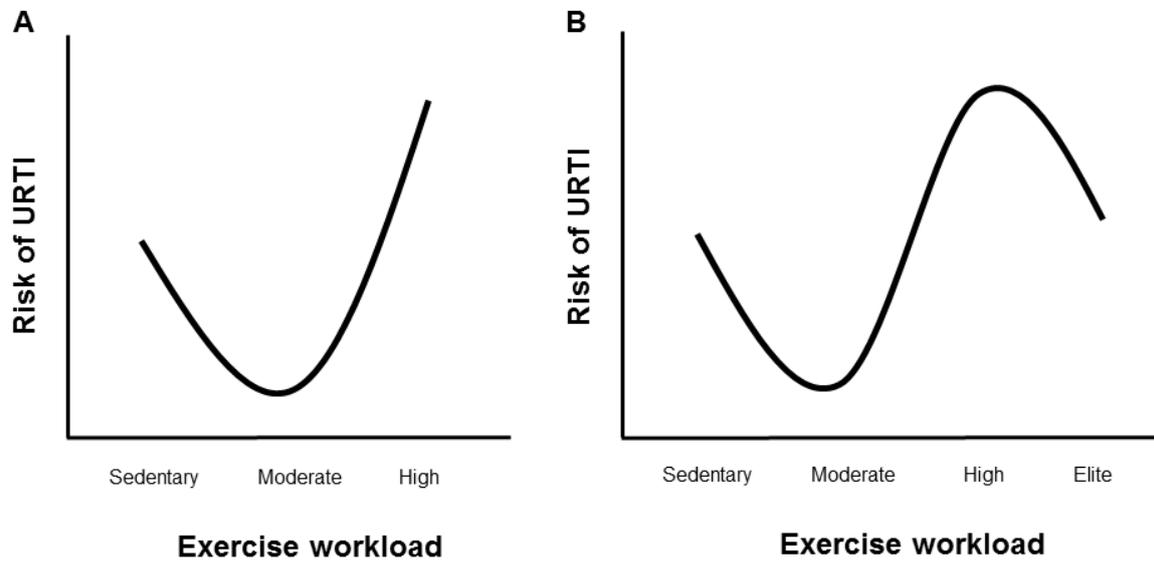
						day is recommended for maximising net protein balance. Further supplementation of protein intake or single/multiple amino acids is not recommended to improve immunity and reduce URS incidence.
HYDRATION	[✓]	[?]	[*]	○○○○○ Lack of RCTs investigating effect on URS	Lack of evidence for an association between hypohydration and self-reported URS, future research should look to establish if higher levels of hypohydration can impair immune function and if this is environment dependent.	Athletes should be advised to maintain fluid balance throughout day-to-day training to ensure optimal performance and health, especially when away at training camps either in the heat or at altitude where fluid losses may be elevated, and infection risk increased.  Daily monitoring of body mass during training camps is a simple and inexpensive method commonly used to monitor changes in fluid balance. In addition, regular monitoring of either urine osmolality or specific gravity can indicate normal ranges for individual athletes and therefore highlight fluid imbalances quickly and effectively.
ANTIOXIDANT SUPPLEMENTATION	[✓]	[✓]	[*]	◎◎◎○○ Meta-analysis of RCTs of heavy acute exercise (and/cold) stress	Given the lack of evidence of vitamin C supplementation to reduce reporting URS in general population, further RCTs are needed during periods of short-term and long-term physical stress with changes in URS supported by clinically relevant immunological markers.	Athletes are recommended to consume a nutrient-dense diet with a variety of fresh fruits and vegetables. In the absence of any rare nutritional (e.g. vitamin C) deficiencies, most athletes are recommended to avoid excessive supplementation with antioxidant vitamins. Supplementation of 0.25 – 1.0 g/day of vitamin C to reduce URS may be useful in some athletes when exposed to extreme unaccustomed acute physical stress.
VITAMIN D SUPPLEMENTATION	[✓]	[✓]	[*]	◎◎◎○○	RCTs of vitamin D supplementation (to correct deficiency) to establish	Practical recommendations seem to be effective in those who are deficient (< 30 nmol/L) and these can be made for both summer and winter months,

				Meta-analyses of RCTs in general population	whether effect on URS in other populations can be shown in athletes. Assessment of URS should be supported by integrated in vivo measures of immune function	<p>although considerations must be made for latitude and skin type.</p> <p>Seasonal screening for vitamin D deficiency is recommended throughout the year in athletes. Bespoke strategies can then be put in to place which either involve maintenance (1000 - 2000 IU/day) or increasing intake to reverse a deficiency. Studies show that consuming a 1,000 to 2,000 IU/day vitamin D3 supplement during winter can achieve sufficiency in most individuals. However, up to 4,000 IU/day may be needed if starting from deficiency. Furthermore, those training indoors or individuals required to wear protective or religious clothing in the summer may also benefit from the 1,000 IU/day vitamin D3 recommendation.</p> <p>In the absence of deficiencies, most athletes are recommended to avoid excessive intake of vitamin D.</p>
BOVINE COLOSTRUM	[✓]	[✓]	[✓]	⊙⊙⊙⊙⊙ Meta-analyses of RCTs in athlete populations	Low precision of estimates of effect on URS need to be followed up with appropriately designed and adequately powered RCTs. Key mechanisms of action need to be elucidated.	Consider daily supplementation (10-20 g) of bovine colostrum particularly during periods of greatest URS risk (e.g. winter period, training camps, long haul travel and competition).
PROBIOTICS	[✓]	[✓]	[?]	⊙⊙⊙⊙⊙ Numerous RCTs in athletes and meta-analyses of	Well-controlled research studies are required to establish dose and strain specific responses of probiotic interventions. Furthermore,	To ensure colonisation of bacterial species in the gut, implementation of probiotic supplementation is recommended to commence at least 14 days prior to overseas travel or competition. With a strain specific consensus lacking, a multi-strain probiotic

				RCTs in general population	mechanisms in elite athletes need to be elucidated. A viable alternative treatment may be a synbiotic (combined probiotic and prebiotic intervention) and research into their use is warranted.	combining species from the genus's lactobacillus and bifidobacterium with the viable number of cells per species greater than $1 \times 10^9$ CFU per day should be considered to ensure the greatest survival to the gut, and subsequent immune modulation.
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633 Figure 1 The proposed J-shaped (A) (Nieman, 1994) and S-shaped (B) (Malm, 2006)

634 relationship between exercise and risk of (respiratory) infection