

1 **Reliability of salivary cortisol and testosterone to a high-intensity cycling protocol to highlight overtraining.**

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29 **Abstract**

30 Athletes physically overload to improve performance. Unbalanced stress/recovery may induce overtraining, which  
31 is difficult to diagnosis as no diagnostic marker exists. Hormonal responses to a 55/80 cycle (30-min of alternating  
32 blocks of 1-min at 55% and 4-min at 80% maximum workrate) may highlight early stage overtraining  
33 (overreaching), as blunted cortisol and testosterone responses to 55/80 follows intensified training. However, the  
34 reliability of hormonal responses to 55/80 when not overreached is unknown. Therefore, reported blunted  
35 hormonal responses could be due to inconsistent cortisol and testosterone responses to 55/80. Participants (n = 23)  
36 completed three 55/80 bouts, >7 days apart, with no exercise 24 h pre-trials. Pre-exercise urine osmolality and  
37 stress questionnaire responses were measured. Pre, post, and 30-min post-exercise saliva samples were collected  
38 for cortisol and testosterone assessment. Salivary cortisol and testosterone responses, osmolality and wellbeing  
39 were not different between trials. Salivary cortisol and testosterone elevated from pre- to post-exercise [by 4.2  
40 nmol·L<sup>-1</sup> (cortisol) and 307 pmol·L<sup>-1</sup> (testosterone)], and 30 min post-exercise [by 160 pmol·L<sup>-1</sup> (testosterone) only].  
41 Intraclass correlation coefficients for pre to peak post-exercise cortisol (0.89; *good*) and testosterone (0.53;  
42 *moderate*) were calculated. This demonstrates the 55/80 induces reliable elevations of salivary cortisol and  
43 testosterone when in a healthy state.

44 **Keywords:** Overtraining, Overreaching, Salivary cortisol, Salivary testosterone, endurance training.  
45

## 46 Introduction

47 Individuals in high demand occupations (e.g. athletes) push the limits of their physical abilities. Athletes overload  
48 the body physically by intensifying training stress, combining an elevation of volume, duration and intensity of  
49 exercise (Wenger and Bell, 2012; Kraemer and Newton, 2000). This can lead to a physical performance decrement  
50 for a limited period but following sufficient recovery of days to weeks a “*supercompensatory*” effect may occur,  
51 with the athlete exhibiting an enhanced performance when compared to baseline levels termed “functional  
52 overreaching” (Meeusen et al., 2013). Continued intensified training can move the athlete into a state of non-  
53 functional overreaching (NFOR) or the overtraining syndrome (OTS), reducing physical performance, which may  
54 not recover for several weeks to years (Meeusen et al., 2013). Signs of overreaching have been reported to occur  
55 within a period as short as 7 days of intensified training with limited recovery (Halson et al., 2002).

56  
57 Retrospective diagnosis of NFOR/OTS is common, given that a valid and reliable protocol or a definitive  
58 diagnostic criterion are currently not available for use during the OTS progression (Meeusen et al., 2013).  
59 Therefore, an appropriate diagnostic marker and/or protocol to warn practitioners that NFOR/OTS may occur  
60 without a modification of training/competition, would be of benefit in practice. Especially, given rates of NFOR  
61 can be considered high in some circumstances, with 30-60% prevalence reported in elite athletes, elite runners,  
62 non-elite runners and adolescent swimmers (Birrer et al., 2013; Matos et al., 2011; Morgan et al., 1987).

63  
64 Hormones associated with the hypothalamus and pituitary gland are suggested as possible markers of NFOR/OTS,  
65 as hypothalamic pituitary disturbances are reported following periods of intensified training (Meeusen et al.,  
66 2004; Meeusen et al., 2010; Urhausen et al., 1998). Indeed, a short duration (30 min) cycling exercise bout,  
67 referred to as the ‘55/80’, has been developed, where a continuous 30 min cycle of alternating blocks of 1 min at  
68 55% maximum work rate and 4 min at 80% maximum work rate are completed (Hough et al., 2011). The 55/80  
69 demonstrated robust elevations in salivary cortisol (by  $\sim 7$  nmol·L<sup>-1</sup> from pre to post 55/80) and salivary testosterone  
70 (by  $\sim 400$  pmol·L<sup>-1</sup> from pre to post 55/80) concentrations in athletes not in a state of NFOR/OTS (i.e. healthy)  
71 (Hough et al., 2011; Hough et al., 2013; Hough et al., 2015). Blunted salivary cortisol (by  $\sim 70\%$ ) and salivary  
72 testosterone (by  $\sim 30\%$ ) responses to the 55/80 have been reported in physically active males and male elite  
73 triathletes following intensified training periods (i.e. possible suffering NFOR/OTS) (Hough et al., 2013; Hough  
74 et al., 2015). These blunted hormonal responses to the 55/80 were found in unison with increased fatigue and  
75 burnout scores measured by a psychological stress and recovery questionnaire (Hough et al., 2013; Hough et al.,  
76 2015). Consequently, the 55/80 was proposed as a useful tool to survey exercise-induced maladaptive cortisol and  
77 testosterone responses when in an overreached state (Hough et al., 2013; Hough et al., 2015). At rest, it is known  
78 that the intra-individual variability can be high for salivary cortisol (up to 51%) and testosterone (up to 30%)  
79 (Hough et al., 2015). Therefore, it is important to examine the simple reliability of the 55/80 hormone responses  
80 without the presence of NFOR/OTS pathologies as it is currently unknown. Knowing this reliability is important  
81 as the previously reported blunted cortisol and testosterone responses to the 55/80 following intensified training,  
82 could simple be due to an unreliable response of these hormones to the 55/80 bout. In measuring the reliability of  
83 the hormonal responses to the 55/80, it is important to understand that these responses can be influenced by  
84 physiological and psychological stress (Koolhass et al., 2011). Therefore a measure of psychological stress is  
85 important while completing this measure of hormonal reliability to the 55/80. The Recovery-Stress Questionnaire  
86 for Athletes (RESTQ) is a validated self-report of stress and recovery events that provides information on the  
87 individual’s state of well-being and predisposition to undertake physical activity (Kellmann and Kallus, 2001;  
88 Tibbert et al., 2009).

89  
90 Therefore, the current study aimed to establish the reliability of the responses of salivary cortisol and testosterone  
91 concentrations to repeated 55/80 bouts across several days. A secondary aim was to examine the physiological and  
92 perceptual strain experienced to repeated exposure to the 55/80 bout. It is hypothesised that salivary hormone  
93 concentrations and physiological and perceptual strain experienced to the 55/80 will be similar across repeated  
94 trials, within an experimental design constructed to avoid NFOR/OTS.

96 **Materials and methods**

97 **Participants**

98 Twenty-three healthy, regularly active males (means  $\pm$  SD; age:  $21 \pm 3$  years; body mass:  $80.7 \pm 8.7$  kg; height:  
99  $1.78 \pm 0.07$  m; peak oxygen uptake ( $\dot{V}O_{2peak}$ ):  $50.9 \pm 7.6$  ml $\cdot$ kg $^{-1}\cdot$ min $^{-1}$ ; maximum work rate:  $304 \pm 49$  W)  
100 volunteered for this study. This study was completed on two different laboratory sites and therefore study  
101 procedures were approved by the two local ethical advisory committees in line with the Helsinki Declaration.  
102

103 **Experimental Design**

104 A repeated measures study design was conducted. Each participant visited the laboratory on four separate  
105 occasions. The protocol is presented in Figures 1 & 2 and in briefly comprises of an incremental, continuous  $\dot{V}O_{2peak}$   
106 test in visit 1 with the remaining trials consisting of the completion of a 55/80 cycle. A written and verbal study  
107 explanation was provided and written informed consent to take part was obtained from each participant before  
108 testing began.  
109

110 **Methodology**

111 During the first laboratory visit a continuous, incremental  $\dot{V}O_{2peak}$  test was completed on a mechanically braked  
112 (Monark Ergonomic 894E, Vansbro, Sweden) or an electronically braked (Lode, Groningen, the Netherlands)  
113 cycle ergometer depending on the laboratory site visited (Figure 1). Once allocated an ergometer, the same cycle  
114 ergometer was used on each visit. Maximum work rate ( $\dot{W}_{max}$ ) was determined using the equation;  
115  $\dot{W}_{max} = \dot{W}_{final} + (t/T) \cdot \dot{W}_{inc}$  where  $\dot{W}_{final}$  = power output during the final stage completed, t = time (s) achieved  
116 in uncompleted stage, T = duration of each stage (180 s), and  $\dot{W}_{inc}$  = workrate increment (35 W). Power outputs  
117 equivalent to 55% and 80% of maximum work rate for each participant were calculated and used during the main  
118 experimental trials.

119  
120 On all remaining laboratory visits, participants consumed a standard breakfast before 09:00 and at least 500 ml of  
121 water on the trial mornings to help ensure they were in a euhydrated state (Sawka et al., 2007). When measuring  
122 the reliability of the hormonal response to exercise there is a requirement to control for hydration status and time  
123 of day of sample collection. Hypohydration is known to elevate cortisol concentrations when compared to a  
124 euhydrated state (Judelson et al., 2008). The daily pattern of cortisol and testosterone concentration release into  
125 circulation is an elevation in the morning leading to a plateau in the circulation concentrations a few hours after  
126 awakening (Crofford et al., 1997; Walton et al., 2007). Therefore, all testing sessions took place at the same time  
127 of day (11:30 – 13:00) to control for circadian rhythms in the hormones under examination. Participants remained  
128 fasted from 09:00 until the end of each trial at ~13:00. All participants abstained from exercise, caffeine and  
129 alcohol intake 24 h before each main trial and completed a food record diary. A similar diet was consumed 24 h  
130 before each main trial. Mean energy intake prior to each trial was  $10.3 \pm 3.1$  MJ with  $53 \pm 13\%$  (carbohydrate),  
131  $28 \pm 12\%$  (fat) and  $18 \pm 6\%$  (protein). The participants drank water *ad libitum* during the trials except 10 min  
132 before the collection of all saliva samples reducing risk of saliva dilution.  
133

134 Each participant completed 3 main trials (Trial 1, Trial 2 and Trial 3) separated by at least 7 days (Figure 2). The  
135 participant in each trial completed a 55/80 bout at 12:00. The 55/80 is a continuous 30 min cycle composed of  
136 alternating blocks of 1 min at 55%  $\dot{W}_{max}$  and 4 min at 70%  $\dot{W}_{max}$ . Heart rate was collected in the final 30s of  
137 each minute via short-range radio telemetry (Polar F2, Polar Electro Oy, Kempele, Finland) and ratings of  
138 perceived exertion (RPE) using a 6-20 Borg scale were recorded in the final 30s of each alternating block. A 52-  
139 item RESTQ was completed at the beginning of each main trial. The RESTQ records the frequency of stress and  
140 recovery events over a period of three days and nights and presents the participant's state of well-being and  
141 predisposition to undertake physical activity (Kellmann and Kallus, 2001). A saliva sample was collected at pre-  
142 exercise, post-exercise and 30 min post-exercise (Figure 2). Saliva samples were unstimulated and collected by  
143 passive drool into a 7 mL Bijou vial (Sterilin, Newport, UK) while seated with eyes open, head tilted slightly  
144 forward and making minimal orofacial movement. The sample was collected for 2 min to allow for collection of  
145 sufficient sample volume. The pre-exercise and 30 min post-exercise samples were collected following a 10 min  
146 seated rest. The post-exercise samples were collected immediately following the 55/80. All samples were chilled  
147 immediately after collection and were divided into aliquots within 30 min and stored at  $-80^{\circ}\text{C}$  until further analysis.  
148 For further detail on the 55/80 procedure please refer to Hough et al., (2011) or Hough et al., (2013).  
149

150 **INSERT FIGURES 1 & 2 HERE**

151

152 The salivary cortisol and testosterone concentrations were determined using commercially available Enzyme  
153 Linked Immunosorbent Assay kits (Salimetrics, PA 16803, USA). Samples from each participant were analysed  
154 on the same plate and went through 1 freeze thaw cycle only. Each sample was measured in duplicate with the  
155 mean salivary cortisol and testosterone concentrations reported. The mean inter-assay CVs were 5.1% and 6.8%  
156 for cortisol and testosterone, respectively. The mean intra-assay CVs were 4.8% and 4.4% for cortisol and  
157 testosterone, respectively.

158

## 159 **Statistical Analyses**

160

161 All data in the text, tables and figures are presented as mean values  $\pm$  standard deviation and/or range (minimum  
162 to maximum). Data were analysed using IBM® SPSS® 24.0 (IBM Corporation, Armonk NY USA). All data were  
163 checked for normality using quantile-quantile plots. Where data was not normally distributed it was log  
164 transformed and re-examined. Salivary cortisol and testosterone data were log transformed and deemed to be  
165 normally distributed after transformation. For clarity we have presented whole salivary cortisol and testosterone  
166 concentrations in the figures. All other data analyse was deemed to be normally distributed.

167

168 Linear mixed models were used to determine if there were any differences between trials (Trial 1; Trial 2 and Trial  
169 3), time (Pre; Post and 30 min Post-Exercise) and any interactions between trial and time for absolute salivary  
170 hormone data. For clarity, figures presenting salivary hormone data sets were collapsed when no significant trial  
171 effects were found. Delta hormone values from pre-exercise to peak post-exercise were analysed with linear mixed  
172 models to determine if there were differences between trials. Differences were determined between trials for urine  
173 osmolality, averaged heart rate, ratings of perceived exertion, and RESTQ responses during each main trial visit.  
174 Fixed and random effects for the linear mixed models were fit for each dependent variable (West et al., 2014).  
175 Statistical significance was set at  $p < 0.05$ . Cohen's  $d$  effect sizes are provided to supplement significant effects  
176 between trials or time. The magnitude of effect size was defined as trivial ( $d < 0.2$ ), small ( $d \geq 0.2, < 0.5$ ), medium  
177 ( $d \geq 0.5, < 0.8$ ), and large ( $d \geq 0.8$ ) (Cohen, 1988).

178

179 Reliability was analysed using intra-individual coefficient of variations (CV<sub>i</sub>) for the salivary cortisol and  
180 testosterone concentrations at each timepoint. The intra-individual mean concentrations (mean<sub>i</sub>) and standard  
181 deviations (SD<sub>i</sub>) were used to calculate the Intra-individual CV ( $CV = (SD_i/mean_i) * 100$ ). In addition, intraclass  
182 correlation coefficients (ICC) were calculated for the delta pre-exercise to peak post-exercise concentrations.  
183 These were calculated by hand using the ICC model 2,1 to measure relative reliability (Vincent and Weir, 2012).  
184 ICC values of less than 0.50 indicate poor reliability, 0.50 – 0.75 indicate moderate reliability, 0.75 – 0.90 good  
185 reliability, greater than 0.90 indicates excellent reliability (Koo and Li, 2016).

186 **Results**

187 **Hydration status, Recovery-stress questionnaires**

188 There was no difference in urine osmolality or REST-Q scores for all trials (Table 1).

189  
190 **Physiological and Perceptual Responses to Exercise**

191 No differences were found in the average heart rate and ratings of perceived exertion responses during the 55/80  
192 (Table 1). ICC values for heart rate and ratings of perceived exertion responses to the 55/80 were 0.83 and 0.75  
193 respectively.

194  
195 **\*\*\*INSERT Table 1 near here\*\*\***

196  
197 **Salivary cortisol**

198 The response of the salivary cortisol concentration to the 55/80 bouts were similar over the 3 trial days ( $F_{2, 21.271} =$   
199  $0.307, P = 0.739$ ). A time effect was found with an elevation of salivary cortisol in response to the 55/80 bouts  
200 ( $F_{2, 22.011} = 13.949, P < 0.001$ ) (Figure 3a). Acute increases in the salivary cortisol concentrations were found from  
201 Pre 55/80 to Post 55/80 ( $P = 0.01; d = 0.8$ ) with a return to baseline at 30 min Post 55/80 ( $P = 0.79$ ). There was no  
202 interaction between trial and timepoint ( $F_{4, 22.196} = 0.587, P = 0.675$ ).

203  
204 Similarly, delta salivary cortisol pre to peak post-exercise were similar over the 3 trial days ( $F_{2, 22} = 0.680, P =$   
205  $0.518$ ). A good reliability in the responses of the salivary cortisol to the exercise was found with an ICC value  
206  $0.89$  calculated for the responses to the exercise bout. The CVi calculations of the salivary cortisol concentrations  
207 at each time point fall in line with that expected and are presented in Table 2.

208  
209 **\*\*\*INSERT Table 2 near here\*\*\***

210  
211 **Salivary testosterone**

212 The responses of the salivary testosterone concentrations to the 55/80 bouts were similar across the 3 trials ( $F_{2, 22.039} =$   
213  $2.123, P = 0.144$ ) (Figure 3b). A time effect was found ( $F_{2, 21.328} = 70.914, P < 0.001$ ) with acute increases  
214 in the salivary testosterone concentrations found from Pre 55/80 to Post 55/80 ( $P < 0.001; d = 1.3$ ) and 30 min  
215 Post 55/80 ( $P < 0.001; d = 0.7$ ) (Figure 3b). There was no interaction between trial and timepoint ( $F_{4, 21.698} = 1.474,$   
216  $P = 0.245$ ).

217  
218 Delta changes pre to peak post-exercise were similar over the 3 trial days ( $F_{2, 22} = 1.324, P = 0.286$ ). A moderate  
219 ICC value for the pre to peak post-exercise delta change of  $0.53$  was calculated and the CVi calculations of the  
220 salivary testosterone concentrations at each time point are presented in Table 2.

221  
222 **\*\*\*Insert Figure 3a&b near here\*\*\***

223  
224 **Discussion**

225 This study aimed to establish the reliability of salivary cortisol, salivary testosterone, heart rate and ratings of  
226 perceived exertion responses to a short duration, high-intensity cycle bout (55/80) to determine the usefulness of  
227 the 55/80 as an exercise test to highlight alterations in exercise induced salivary cortisol and testosterone responses  
228 that may occur during NFOR/OTS. No differences in the salivary cortisol and testosterone responses to the  
229 repeated 55/80 trials were found. Therefore, the hypothesis that the salivary hormone responses to the 55/80 are  
230 similar on repeated exposure can be accepted. A secondary aim of this study was to measure the physiological  
231 (measured via heart rate responses) and perceptual (measured via ratings of perceived exertion scores) strain of  
232 the 55/80. Similar strain across the trials were found which confirms that repeated exposure to the exercise bout  
233 did not alter the strain experienced by the participants. This is important if the 55/80 is to be used as a physical  
234 stress test to examine possible dysfunction in the responses of hypothalamic pituitary adrenal and hypothalamic  
235 pituitary gonadal axes during periods of heavy training stress (i.e. NFORS/OTS).

236  
237 The findings of robust elevations of salivary cortisol and testosterone from pre- to post-55/80 in the current study  
238 corresponds with previous reports of cortisol and testosterone elevations to a 55/80 bout in a healthy state (i.e. not  
239 in a state of NFOR/OTS)(Hough et al., 2013; Hough et al., 2015). The magnitude of elevation seen from pre to  
240 peak post-exercise in the current study ( $\sim 6 \text{ nmol}\cdot\text{L}^{-1}$  for cortisol and  $\sim 315 \text{ pmol}\cdot\text{L}^{-1}$  for testosterone) is in line with  
241 that previously reported ( $\sim 7 \text{ nmol}\cdot\text{L}^{-1}$  for cortisol and  $\sim 400 \text{ pmol}\cdot\text{L}^{-1}$  for testosterone) (Hough et al., 2011; Hough  
242 et al., 2013; Hough et al., 2015). The effect sizes reported with these findings in the current study highlight a large  
243 effect ( $> 0.8$ ) of the 55/80 on both these hormones from pre- to post-exercise. Our analysis revealed no differences  
244 in the hormonal responses to repeated 55/80 bouts. It also suggests that the reliability of salivary cortisol in  
245 response to the 55/80 can be interpreted as good, however the reliability of the salivary testosterone response to

246 the 55/80 was moderate. This reliability indicates that any changes to these responses, for example when in a state  
247 of NFOR/OTS, should be viewed cautiously. The blunted responses of salivary testosterone to the 55/80 previously  
248 reported, following periods of intensified training, may have been due to the moderate reliability in the responses  
249 of this hormone in saliva to the 55/80. Examining the responses further showed a hormonal variability within the  
250 individuals of ~27% (salivary cortisol) and ~14% (salivary testosterone). This variability corresponds to that seen  
251 in resting plasma samples previously reported (Maes et al., 1997; Walton et al., 2007). Keeping in mind that the  
252 55/80 exercise bout has been reported to highlight a blunted response of cortisol (of ~70%) and testosterone (of  
253 ~30%) following an intensified training period (i.e. when the athlete is in a state of possible NFOR/OTS) (Hough  
254 et al. 2013; Hough et al. 2015). These blunted alterations are in excess of the intra-individual variability this current  
255 study reports. This suggests that the blunted hormonal responses to the 55/80 following intensified training, found  
256 previously, were not due to the intra-individual variability of the hormones measured and may be due to the  
257 elevated physical stress during a period of heavy training (i.e. possible NFOR/OTS). To conclude, the data  
258 suggests that salivary cortisol elevated in response to the 55/80 and this response has a good reliability. Therefore,  
259 this may be a useful surveillance measure to complete during training periods to help to highlight states of  
260 NFOR/OTS with the expectation that during these periods the salivary cortisol responses will be blunted as  
261 previously reported (Hough et al., 2013; Hough et al., 2015)

262  
263 The heart rate and ratings of perceived exertion analysis in the current study show that the physiological strain and  
264 the perception of exertion to the 55/80 do not differ across trials. The reliability analysis indicates a good reliability  
265 for the responses in both measures. If using this exercise stress test as a tool to highlight hormonal changes, our  
266 results indicate that hormonal alterations found are not due to changes in physiological strain or perception of  
267 exertion to the exercise bout.

## 268 **Strengths and Limitations**

269  
270 In measuring hormonal reliability, specific controls are required to help to remove external influences on the  
271 hormones being analysed. The strength of the current study is the control of these important external influences.  
272 Firstly, the RESTQ scores reported no significant disparities in stress or recovery scores within individuals during  
273 the study. This confirms that participants completed the 55/80 bouts in a similar state of well-being and  
274 predisposition to undertake physical activity (Kellmann and Kallus, 2001). It can be concluded from this that the  
275 hormonal responses have not been influenced by a change in well-being in the participants. Additionally, hydration  
276 status also influences cortisol and testosterone concentrations. Specifically, hypohydration (loss of ~5% body  
277 mass) elevates circulating cortisol and decreases testosterone when compared to a euhydrated state (Judelson et  
278 al., 2008). An indicator of euhydration is a urine osmolality value of  $< 700 \text{ mosmol}\cdot\text{kg}^{-1}$ , with each participant  
279 demonstrating an acceptable value (274 – 382  $\text{mosmol}\cdot\text{kg}^{-1}$ ) prior to completing the 55/80 (Sawka et al., 2007).  
280 Therefore, hydration status likely did not influence the hormonal responses to the 55/80 reported in this study.

281  
282 It should be noted that the measurement of salivary hormones, specifically testosterone, may be inflated if  
283 measured by immunoassay when compared with another measurement technique for salivary hormone analysis  
284 such as liquid chromatography tandem mass spectrometry (LC-MS/MS) (Welker et al., 2016). However, this  
285 inflation found in immunoassay results compared with LC-MS/MS is most evident at low concentrations ( $< 35$   
286  $\text{pmol}\cdot\text{L}^{-1}$ ). The concentrations reported in this current study were in excess of this low concentration value.  
287 However, it is important to know that different salivary hormone analysis methods may report different  
288 concentrations from the same samples. The relatively small sample size in the current research study must also be  
289 addressed. It is important to examine the power of the analysis completed within this study. A post-hoc  
290 computation of achieved power on two of the main variables in this research study was completed. These variables  
291 were the delta salivary cortisol and testosterone pre to peak post-exercise measures. The analyses achieved a power  
292 of 0.72 (cortisol) and 0.90 (testosterone). This finding details a 28% and 10% risk of committing type II errors (i.e.  
293 missing an effect if it genuinely exists). It is commonly agreed a power level of 80% is credible to determine actual  
294 effects (Field, 2009). The reader should be aware of this higher risk of missing an effect found in the cortisol data  
295 presented in this research study.

## 296 **Conclusion**

297  
298 This study confirms that the 55/80 induces reliable elevations of salivary cortisol. It also highlights a moderate  
299 reliability when measuring salivary testosterone in response to the 55/80. This supports the use of the 55/80 to  
300 survey responses of salivary cortisol (and perhaps highlight their utility within NFOR/OTS cascades). Caution  
301 must be implemented if using the 55/80 to highlight alterations in salivary testosterone concentrations. The  
302 hormonal variability within individuals found in the current study (~27% cortisol and ~14% testosterone) are lower  
303 than those reported adaptations that occur in exercise-induced salivary cortisol and testosterone responses (to the  
304  
305

306 55/80 bout) following periods of intensified training (reductions of ~72% in salivary cortisol and ~34% in salivary  
307 testosterone following periods of intensified training when compared to before the training period (Hough et al.  
308 2013; Hough et al. 2015). This finding further supports the potential use of the 55/80 as a tool for the surveillance  
309 of hormonal adaptations which may occur during periods of heavy training (e.g. NFOR/OTS).

310  
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319  
320



321 **References**

- 322 Birrer, D., Lienhard, D., Williams, C.A., Rothlin, R., & Morgan, G. (2013). Prevalence of non-functional  
323 overreaching and the overtraining syndrome in Swiss elite athletes. *Schweizerische Zeitschrift für Medizin und*  
324 *Traumatologie*, 61(4):23–29. <https://doi.org/10.1249/MSS.0b013e318207f87b>
- 325 Cohen, J. (1988). *Statistical Power Analysis for the Behavioural Sciences*. Second Edition. Hillsdale, NJ:  
326 Lawrence Erlbaum Associates.
- 327 Crofford, L.J., Kalogeras, K.T., Mastorakos, G., Magiakou, M.A., Wells, J., Kanik, K.S., Gold, P.W., Chrousos,  
328 G.P., & Wilder, R.L. (1997). Circadian relationships between interleukin (IL)-6 and hypothalamic-pituitary-  
329 adrenal axis hormones: Failure of IL-6 to cause sustained hypercortisolism in patients with early untreated  
330 rheumatoid arthritis. *Journal of Clinical Endocrinology & Metabolism*, 82(4):1279-1283.  
331 <https://doi.org/10.1210/jcem.82.4.3852>
- 332 Field, A. (2009) *Discovering Statistics Using SPSS*. Third Edition. Sage Publications Ltd., London.
- 333 Halson, S.L., Bridge, M.W., Meeusen, R., Busschaert, B., Gleeson, M., Jones, D.A., & Jeukendrup, A.E. (2002).  
334 Time course of performance changes and fatigue markers during intensified training in trained cyclists. *Journal of*  
335 *Applied Physiology*, 93(3): 947-956. <https://doi.org/10.1152/jappphysiol.01164.2001>
- 336 Hough, J.P., Papacosta, E., Wraith, E., & Gleeson, M. (2011). Plasma and salivary steroid hormone responses of  
337 men to high-intensity cycling and resistance exercise. *Journal of Strength and Conditioning Research*, 25(1):23-  
338 31. <https://doi.org/10.1519/JSC.0b013e3181fef8e7>
- 339 Hough, J., Corney, R., Kouris, A., & Gleeson, M. (2013). Salivary cortisol and testosterone responses to high-  
340 intensity cycling before and after an 11-day intensified training period. *Journal of Sports Sciences*, 31(14):1614-  
341 1623. <https://doi.org/10.1080/02640414.2013.792952>
- 342 Hough, J., Robertson, C., & Gleeson, M. (2015). A 10-day training camp blunts exercise-induced salivary  
343 testosterone in elite level triathletes. *International Journal of Sports Physiology and Performance*, Oct 10(7):935-  
344 938. <https://doi.org/10.1123/ijsp.2014-0360>
- 345 Judelson, D.A., Maresh, C.M., Yammoto, L.M., Farrell, M.J., Armstrong, L.E., Kraemer, W.J., Volek, J.S.,  
346 Spiering, B.A., Casa, D.J., & Anderson, J.M. (2008). Effect of hydration state on resistance exercise-induced  
347 endocrine markers of anabolism, catabolism and metabolism. *Journal of Applied Physiology*, 105:816-824.  
348 <https://doi.org/10.1152/jappphysiol.01010.2007>
- 349 Kellmann, M., & Kallus, K.W. (2001). *Recovery-Stress Questionnaire for Athletes: User manual*. Champaign, IL:  
350 Human Kinetics.
- 351 Koo, T.K., & Li, M.Y. (2016). A Guideline of Selecting and Reporting Intraclass Correlation Coefficients for  
352 Reliability Research. *Journal of Chiropractic Medicine*, Jun 15(2):155–163.  
353 <https://doi.org/10.1016/j.jcm.2016.02.012>
- 354 Koolhaas, J.M., Bartolomucci, A., Buwalda, B.D., De Boer, S.F., Flügge, G., Korte, S.M., Meerlo, P., Murison,  
355 R., Olivier, B., Palanza, P., Richter-Levin, G., Sgoifo, A., Steimer, T., Stiedl, O., van Dijk, G., Wöhr, M., & Fuchs,  
356 E. (2011). Stress revisited: a critical evaluation of the stress concept. *Neuroscience Biobehavioral Reviews*  
357 35:1291–301. <https://doi.org/10.1016/j.neubiorev.2011.02.003>
- 358 Kraemer, W.J. & Newton, R.U. (2000). Training for muscular power. *Physical Medicine and Rehabilitation Clinic*  
359 *of North America*, 11:341-368. [https://doi.org/10.1016/S1047-9651\(18\)30133-5](https://doi.org/10.1016/S1047-9651(18)30133-5)
- 360 Maes, M., Mommen, K., Hendrickx, D., Peeters, D., D'Hondt, P., Ranjan, R., DeMeyer, F., & Scharpe, S. (1997).  
361 Components of biological variation, including seasonality, in blood concentrations of TSH, TT3, FT4, PRL,  
362 cortisol and testosterone in healthy volunteers. *Clinical Endocrinology* 46(5):587–598.  
363 <https://doi.org/10.1046/j.1365-2265.1997.1881002.x>

- 364 Matos, N.F., Winsley, R.J., & Williams, C.A. (2011). Prevalence of nonfunctional overreaching/overtraining in  
365 young English athletes. *Medicine & Science in Sports and Exercise*, 43:287-294.  
366 <https://doi.org/10.1249/MSS.0b013e318207f87b>
- 367 Meeusen, R., Piacentini, M.F., Busschaert, B., Buyse, L., De Schutter, G., & Stray-Gundersen, J. (2004).  
368 Hormonal responses in athletes: The use of a two bout exercise protocol to detect subtle differences in  
369 (over)training status. *European Journal of Applied Physiology*, 91(2–3): 140–146. [https://doi.org/10.1007/s00421-](https://doi.org/10.1007/s00421-003-0940-1)  
370 003-0940-1
- 371 Meeusen, R., Nederhof, E., Buyse, L., Roelands, B., deSchutter, G., & Piacentini, M.F. (2010). Diagnosing  
372 overtraining in athletes using the two-bout exercise protocol. *British Journal of Sports Medicine*, 44(9):642-648.  
373 <https://doi.org/10.1136/bjism.2008.049981>  
374
- 375 Meeusen, R., Duclos, M., Forster, C., Fry, A., Gleeson, M., Nieman, D., Raglin, J., Rietjens, G., Steinacker, J., &  
376 Urhausen, A. (2013). Prevention, diagnosis, and treatment of the overtraining syndrome: Joint consensus statement  
377 of the European College of Sport Science and the American College of Sports Medicine. *Medicine & Science in*  
378 *Sports and Exercise*, Jan 45(1):186-205. <https://doi.org/10.1080/17461391.2012.730061>  
379
- 380 Morgan, W., O'Connor, P., Sparling, P., & Pate, R.R. (1987). Psychological characterization of the elite female  
381 distance runner. *International Journal of Sports Medicine*, 8:124-131. <https://doi.org/10.1055/s-2008-1025717>  
382
- 383 Sawka, M.N., Burke, L.M., Eichner, E.R., Maughan, R.J., Montain, S.J., & Stachenfeld, N.S. (2007). American  
384 College of Sports Medicine position stand. Exercise and fluid replacement. *Medicine & Science in Sports and*  
385 *Exercise*, Feb 39(2):377-390. <https://doi.org/10.1249/mss.0b013e31802ca597>  
386
- 387 Tibbert, S., Morris, T., & Andersen, M. (2009). Validity of the recovery-stress questionnaire. *Journal of Science*  
388 *and Medicine in Sport*, Jan 12: S32-S33. <https://doi.org/10.10016/j.jsams.2008.12.077>.  
389
- 390 Urhausen, A., Gabriel, H.H., & Kindermann, W. (1998). Impaired pituitary hormonal response to exhaustive  
391 exercise in overtrained endurance athletes. *Medicine & Science in Sports and Exercise*, 30(3): 407–414.  
392 <https://doi.org/10.1097/00005768-199803000-00011>  
393
- 394 Vincent, W.J. & Weir, J.P. (2012). *Statistics in kinesiology*. 4<sup>th</sup> edition. Champaign, IL: Human Kinetics  
395
- 396 Walton, M.J., Anderson, R.A., Kicman, A.T., Elton, R.A., Ossowska, K., & Baird, D.T. (2007). A diurnal variation  
397 in testicular hormone production is maintained following gonadotrophin suppression in normal men. *Clinical*  
398 *Endocrinology*, 66:123-129. <https://doi.org/10.1111/j.1365-2265.2006.02696.x>  
399
- 400 Welker, K.M., Lassetter, B., Brandes, C.M., Prasad, S., Koop, D.R., & Mehta, P.H. (2016). A comparison of  
401 salivary testosterone measurement using immunoassays and tandem mass spectrometry.  
402 *Psychoneuroendocrinology* 71: 180 – 188. <https://doi.org/10.1016/j.psyneuen.2016.05.022>  
403
- 404 Wenger, H.A. & Bell, G.J. (2012). The interactions of intensity, frequency and duration of exercise training in  
405 altering cardiorespiratory fitness. *Sports Medicine*, 3: 346 – 356. [https://doi.org/10.2165/00007256-198603050-](https://doi.org/10.2165/00007256-198603050-00004)  
406 00004  
407
- 408 West, B.T., Welch, K.B., Galecki, A.T. (2014). *Linear mixed models: A Practical Guide Using Statistical*  
409 *Software*, Second Edition. Taylor & Francis.  
410

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**Figure Captions**

**Figure 1** Schematic overview of the  $\dot{V}O_{2peak}$  test.

**Figure 2** Schematic overview of experimental trials.

**Figure 3** The collapsed (a) salivary cortisol and (b) salivary testosterone concentration responses to the 55/80 cycles in all trials.

\* Different than Pre 55/80 values

To make conversion from Syst eme International d'Unites (SI) units to gravimetric/conventional unit. To convert salivary cortisol from  $\text{nmol.L}^{-1}$  to  $\text{ng.mL}^{-1}$  multiply by 0.3625. Conversion of salivary testosterone from  $\text{pmol.L}^{-1}$  to  $\text{ng.dL}^{-1}$  multiply by .0288.