

Biopolymeric Sustainable Materials and their Emerging Applications

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Abstract

Advancements in polymer science and engineering have helped the scientific community to shift its attention towards the use of environmentally benign materials for reducing the environmental impact of conventional synthetic plastics. Biopolymers are environmentally benign, chemically versatile, sustainable, biocompatible, biodegradable, inherently functional, and ecofriendly materials that exhibit tremendous potential for a wide range of applications including food, electronics, agriculture, textile, biomedical, and cosmetics. This review also inspires the researchers toward more consumption of biopolymer-based composite materials as an alternative to synthetic composite materials. Herein, an overview of the latest knowledge of different natural- and synthetic-based biodegradable polymers and their fiber-reinforced composites is presented. The review discusses different degradation mechanisms of biopolymer-based composites as well as their sustainability aspects. This review also elucidates current challenges, future opportunities, and emerging applications of biopolymeric sustainable composites in numerous engineering fields. Finally, this review proposes biopolymeric sustainable materials as a propitious solution to the contemporary environmental crisis.

Keywords: Biopolymers, Biodegradable polymers, Biodegradability, Biocomposites, Biomedical applications

List of abbreviations

ACC	All-cellulose composite
AEMO	Acrylate epoxidized mustard oil
APHAS	American Public Health Association standard
ASTM	American society for testing of materials
BF	Bamboo fiber
BG	Bioglass
BOD	Biochemical oxygen demand
CA	Cellulose acetate
CB	Composite bioplastic
CCG	Chemically converted graphene
CG	Cashew gum
CMC	Carboxymethyl cellulose
CNC	Cellulose nanocrystals
CNT	Carbon nanotube
COVID-19	Coronavirus disease 2019
DMT	Dimethyl terephthalate
DS	Degree of substitution
ES	Extruded sheets
FCC	Fiber cement composites
FFF	Fused filament fabrication
GCF	Green Coconut Fiber
GelMA	Methacrylated gelatin

HA	Hyaluronic acid
HAP	Hydroxyapatite
KF	Kenaf Fiber
LCA	Life cycle assessment
MB	Mater-Bi [®]
MC	Microcrystalline
MOSF	Moringa Oleifera (Sahajana) seed filler
MSCs	Mesenchymal stem cells
MNP	Melanin nanoparticles
MWCNT	Multi-walled carbon nanotube
NFs	Natural fibers
nHAP	Nanohydroxyapatite
NPs	Nanoparticles
NR	Natural rubber
NVP	N-vinyl-2-pyrrolidone
OPEFB	Oil palm empty fruit bunch
PA	Polyamide
PBAT	Polybutylene adipate terephthalate
PBH	Polyhydroxy butyrate
PBSA	Poly(butylene succinate-co-butylene adipate)
PCL	Polycaprolactone
PDO	Propanediol
PE	Polyethylene
PEO	Poly (ethylene oxide)
PET	Polyethylene terephthalate
PGA	Poly(glycolic acid)
PHA	Poly(hydroxyalkanoate)
PHB	Poly(hydroxybutyrate)
PHBV	Polyhydroxybutyrate-valerate
PBS	Poly(butylene succinate)
PLA	Poly(lactic acid)
PLLA	Poly-L-lactic acid
PLGA	Poly(lactide-co-glycolide)
PP	Polypropylene
PTA	Pure terephthalic acid
PTT	Poly(trimethylene terephthalate)
PVA	Poly(vinyl alcohol)
PVAc	Poly(vinyl acetate)
PWBF	Plain woven banana fabric
rPP	Recycled polypropylene
RS-g-PMMA	Rice straw-g-poly methyl methacrylate
SB	Simple bioplastic
SEM	Scanning electron microscope
SPNCC	Sugar palm nanocrystalline cellulose
SPS	Sugar palm starch
TPS	Thermoplastic starch
UV	Ultraviolet
WPC	Wood Plastic composite

1. Introduction

Nowadays, the extensive use of conventional polymers, including polyethylene (PE), thermoset epoxy resins, and polyethylene terephthalate (PET) are polluting our environment [1]–[3]. More importantly, their role in the life of both flora and fauna have some serious negative impacts due to their stability against microorganisms, humidity, and temperature. Thus, piling up a huge amount of plastic waste [4]–[6]. Additionally, there are rising global

health concerns among the scientific community about toxic chemicals, including bisphenol A, phthalates, or polychlorinated biphenyls, which are employed for the production of plastics [7]. Thus, plastic waste management is a major environmental issue and requires suitable methods and materials for plastic decomposition [8]. Other synthetic polymers are extensively consumed globally, and it is estimated that the non-biodegradable polymer waste will reach a volume of 25 billion metric tons by 2050. Additionally, the production of plastics has exceeded 8 billion tons of plastic in the last seventy years [9]–[11]. From 2020 to 2023, the growth of natural and synthetic plastics will be 13 % per annum as per estimation. Thus, synthetic polymers pose a risk to the environment due to their exceptionally stable nature, which causes the accumulation of waste [12]. To address the issue of plastic waste, the recycling of synthetic plastics is a sustainable environmental solution. However, the recyclable polymers possess lower mechanical characteristics and cannot be employed for high-end applications [13]–[15]. Landfilling of these materials should be given the least priority as prescribed by European Union (EU) waste management directives. Therefore, recently, more emphasis was given to the use of naturally available polymers that overcome plastic waste issues due to their biodegradable nature [16]–[18].

The utilization of biodegradable polymers or biopolymers is a prominent alternative to overcome the consumption of synthetic plastic [19]. These bio-based polymers, inevitably key biopolymers derived from renewable sources including biomass, corn, sugarcane, and molasses, have gained significant attraction. Upon degradation process, these polymers reduce their pH and molecular weight as well as increase their crystallinity [20]. These polymers are renewable materials which undergo rapid degradation through natural micro-organisms without releasing any toxic or distinguishable residue under appropriate conditions, including moisture, oxygen, and temperature [21]. The reinforcement of different natural fillers/additives significantly enhances the physical as well as mechanical properties of these biomaterials [22]. These polymers are becoming indispensable and applications of biopolymers have been greatly enhanced in the last decade [23]–[25]. These biopolymers are vastly applied in broad spectrum of engineering applications such as pharmaceutical, food packaging, electronics, textile, agriculture, construction, biomedical, aerospace, and automotive industries [26]–[28]. Bio-based polymeric materials are classified into three diverse groups on the basis of their sources and production method, as presented in **Table 1**. The first group incorporates natural polymers, including polysaccharides, proteins, and lipids extracted from animals or plants. In the second group, synthetic polymers are developed through different ring-opening polymerization or condensation methods by using bio-based monomers [29]–[31]. These polymers include polycaprolactone (PCL), polybutylene adipate terephthalate (PBAT), polylactic acid (PLA), poly(butylene succinate-co-butylene adipate) (PBSA), and poly(butylene succinate) (PBS) [32]–[34]. Whereas the third group contains polymers including poly(hydroxybutyrate) (PHB), poly(hydroxyalkanoate) (PHA), and polyhydroxybutyrate-valerate (PHBV), which are developed through genetically modified bacteria or micro-organisms [35]–[37].

Table 1 Classification of biopolymeric materials

Classification	Source	Biopolymers	Ref.
Polypeptides/Proteins	Animals	Fish protein, collagen, silk, gelatin, casein, whey	[38]
	Plants	Soy protein, zein, wheat gluten	[39]
Polysaccharides	Animals	Hyaluronic acid, chitosan/chitin	[40]

	Plants	Starch, agar, cellulose, carrageenan, pectin, alginate	[41]
	Lipids	Fatty acid, carnauba wax, beeswax, oil	[42]
Biopolymers produced through microorganisms	Bio-derived monomers	PLA, PGA	[43]
	Microbial polyesters	PHA, PCL, PHB, and PHBV	[44]
	Bacterial	Cellulose, gelatin, dextran, Xanthan, polygalactosamine	[45]
Synthetic	-	PVA, PVAc	[46]
	Aliphatic polyesters	PLA, PGA, PCL	[47]

Nowadays, different biopolymers, including poly(lactic acid) (PLA), and poly(hydroxyalkanoate)s (PHAs) are producing a capacity growth of approximately 22 %-300 %. These biopolymers offer an eco-friendly pathway to plastic production due to their lower associated carbon emissions from petroleum refinement and extraction [48]. The extensive use of biopolymers has been in composite materials due to their unique attributes, as illustrated in **Figure 1**. These biopolymers act as matrices in composite materials and can be used to replace different thermosets like epoxy resin [49]–[51]. Different methods including in-situ, infiltration, and electrospinning techniques have been used to develop biopolymer composites and have gained considerable attention [52]–[54]. The complete degradation of different biopolymers and their composites is completely achieved in the natural environment due to different micro-organisms, moisture, and ultraviolet (UV) sunlight [55]–[57]. The degradation process performs changes in crystallinity of the biopolymer-based matrix phase and different natural fibers (NFs) or natural residue as reinforcement in composite materials [58].

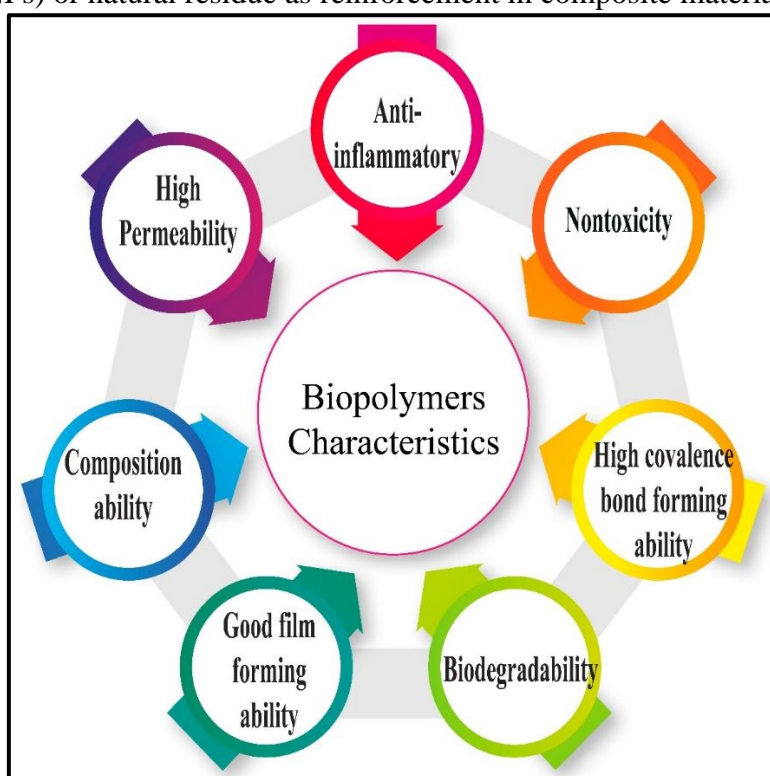


Figure 1. Main characteristics of biopolymeric materials.

The world is moving towards a sustainable, cleaner, and greener environment by substituting non-biodegradable polymers with bio-based polymers. These polymers possess distinct

features such as renewability, non-toxicity, sustainability, and bio-degradability [59]–[61]. Therefore, myriad biopolymers are replacing synthetic polymers in different commercial sectors, including packaging, biomedical, and agricultural industries. **Figure 2** presents the major consumption of bio-based polymers in different engineering sectors. The global market indicates that the value of biopolymers increased from \$15.6 billion in 2016 to \$22.3 billion by 2022.

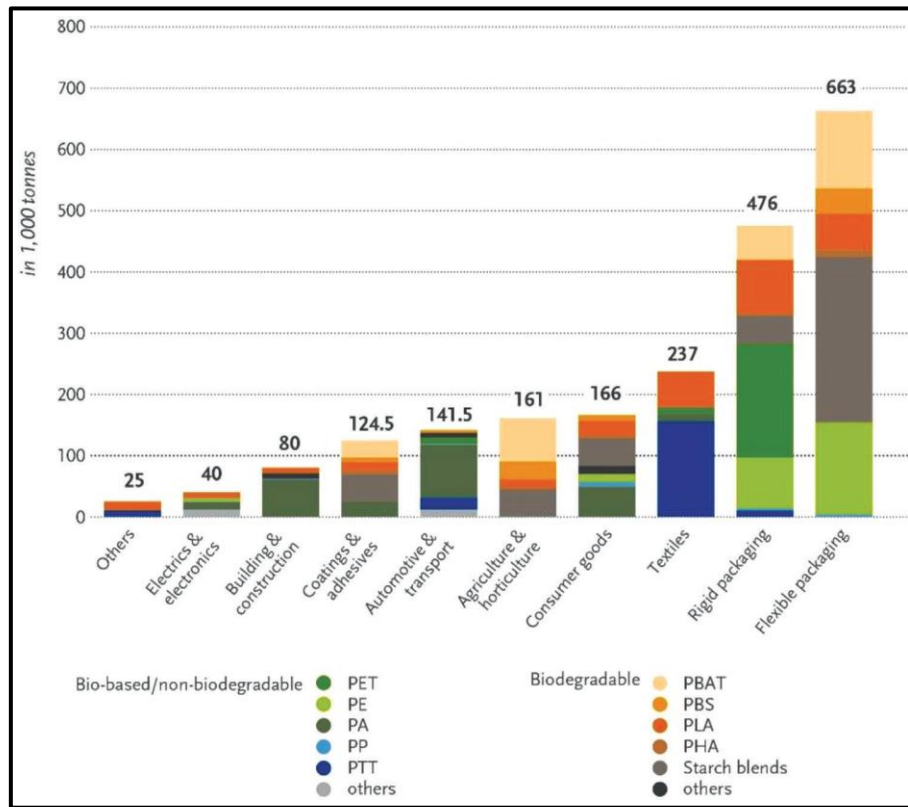


Figure 2. Global biopolymer production capacity in different markets (Adapted from [62])

Different natural- and synthetic-based biodegradable polymers have been recently utilized by the research community to evaluate their properties for promoting sustainable solutions. Considering all the aforementioned information about the negative impact of non-biodegradable polymers and positive feedback of biopolymers, the proposed review is written with the aim of highlighting recent developments regarding their biodegradability and the emerging applications of biopolymers in different engineering disciplines.

1. Biodegradability

Even the reusing or recycling of non-biodegradable polymers or polymer composites generates harmful and poisonous substances. To overcome this issue, composites are manufactured by using biodegradable-based polymers and fibers, which provide sustainable solutions [63]. Crystallinity and molecular weight are two prime parameters that influence the degradation as well as mechanical properties of biopolymer-based films/coatings. [64]. It is difficult for microorganisms to attack biopolymers exhibiting molecular weights higher than anti-microbial agents [65]. Thus, the mechanical strength of biopolymers remains unaffected, as anti-microbial molecules do not influence polymeric compatibility. However, any physical or chemical interaction between polymer matrix and additives considerably affects the tensile properties. To overcome this problem, hydrophobic agents, including essential oils, are incorporated as anti-microbial agents to reduce the permeability of the biopolymers. According

to the American Society for Testing of Materials (ASTM), the chemical structure of the biodegradable polymer is significantly transformed by environmental factors. Different enzymes/microorganisms including fungi, bacteria, and algae, deteriorate the chemical and physical properties of biopolymers and reduce their molecular mass by yielding H₂O, CO₂, and other inorganic products due to anaerobic and aerobic conditions. Additionally, biodegradation produced not toxic or distinguishable residue [66]–[68]. Different degradation strategies, including mechanical degradation, thermo-oxidative degradation, hydrolytic degradation, and photodegradation, also help in promoting biodegradation [69]–[74]. Biodegradation of biopolymers occurs after two steps (depolymerization and mineralization), as elaborated in **Table 2**.

Table 2. Chemistry of biodegradation process

Step	Major stages	Schematic diagram
Depolymerization	In this stage, macromolecule-based polymeric chains are shortened and weakened under the action of moisture, sunlight, heat, or enzymes [75]. Mostly, exo- or endo-enzymes are abiotic reactions that produce cleavage among the polymer networks. The contact area between biopolymer and micro-organism increases during this stage [76].	
Mineralization	In this step, transportation of small-sized oligomeric fragments into the cells occurs, where these fragments are bio-assimilation through the microbial population [78]. Depending upon the presence of oxygen, biodegradation is further divided into two types: anaerobic degradation (in the absence of air/oxygen) and aerobic degradation (in the presence of air/oxygen). No residue exists upon the completion of the mineralization process and biopolymer is transformed into biomass, CO ₂ , water, and methane [79].	

Adapted with permission [77]

1.1. Biodegradable Polymers

Biodegradable polymers are obtained from renewable resources and petroleum resources and have recently gained tremendous attraction as these polymers promote a sustainable environment. These polymers are decomposed into minerals, biomass, CO₂, H₂O, and CH₄ through microbial degradation [80]. It is essential to utilize biodegradable polymers to lessen the environmental impact of the climate. On the other hand, biodegradable biopolymers are macromolecular compounds extracted from animals, plants, or micro-organisms and commonly include proteins, lipids, and polysaccharides [81]. Proteins incorporate gelatin, gluten, keratin, and casein, while, lipids include fatty acids or esters, vegetable- and animal-based oils like soya, and sunflower oil [82]. Polysaccharides exist in chitosan, starch, chitin/chitosan, and alginate forms [83].

These biopolymers have drawn significant attention due to their eco-friendliness, non-toxicity, biocompatibility, chemical versatility, and inherent functionality [84]–[86]. On the basis of biodegradability, these polymers are divided into biodegradable and non-biodegradable polymers [87]. Synthetic biodegradable polymer chemical compositions include amides, esters,

diacids, and anhydrides [88]. Repeating units of these polymers are connected through covalent bonds and are fabricated via micro-organisms [89]. Synthetic biopolymers including PLA, PHA, PCL, and PBS are fabricated through chemical processing of different renewable biological sources. Synthetic biopolymers are made up of hydrolyzable links and disintegrate into monomers upon biodegradation [90]. Traditional biodegradable synthetic polymers are synthesized from renewable resources through a hybrid approach that involves chemical and biological reactions [91]. Additionally, valiant efforts have been made for the development of sustainable and green platforms by microbial one-step fermentation of synthetic biopolymers, including PGA, PLA, and multiple types of PHAs [92].

Non-biodegradable biopolymers include polycarbonates, polyethylene, polyether esters, and polyamides (PAs), which are conventionally synthesized from bio-based monomers. Non-biodegradable polymers have been shown to be more usable than biodegradable biopolymers [93]–[95]. The incorporation of polysaccharide’s functional groups into different fillers imparts distinct applications. There are also some non-biodegradable plastics, including bio-PET (bio-ethylene terephthalate), bio-PE (bio-polyethylene), and bio-PTT (bio-polytrimethylene terephthalate) [96]–[98]. **Table 3** summarizes the distinct properties of different natural- and synthetic-based biopolymers, degradation time, and their applications.

Table 3. General overview of different biopolymers along with their source, different characteristics, and degradation time

Biopolymer	Source / composition	Characteristics	Degradation Time	Applications	Ref.
Collagen	Animal	(i) Naturally available protein (ii) Biocompatibility (iii) Excellent physiochemical properties	12h	(i) Tissue engineering (ii) Cosmetic industry (iii) Food industry	[99]
Alginate	Plant	(i) Naturally available polysaccharides (ii) Excellent antimicrobial properties	80 days	(i) Textile industry (ii) Drug delivery system (iii) Tissue engineering (iv) Wound dressing	[100]
Chitosan	Animal	(i) Abundantly available polysaccharides (ii) Biocompatibility (iii) Excellent solubility (iv) Excellent viscosity (v) Extraordinary polyelectrolytic behavior (vi) Structurally versatile	> 20 weeks	(i) Food industry (ii) Pharmaceutical industry (iii) Agricultural industry	[101]
Silk	Animal	(i) Naturally available protein (ii) Excellent mechanical properties (iii) High luster	6 weeks	(i) Tissue engineering (ii) Food applications (iii) Textile industry	[102]
Starch	Plant	(i) Naturally available polysaccharides (ii) Excellent water vapor properties (iii) High viscosity (iv) Excellent storage modulus	Several weeks	(i) Drug delivery systems (ii) Textile industry (iii) Paper industry (iv) Food packaging	[103]
HA	Animal	(i) Abundantly available polysaccharides (ii) Biocompatibility (iii) High viscosity	4 months	(i) Tissue engineering (ii) Cosmetic industry (iii) Wound dressing (iv) Food packaging	[104]
Cellulose	Plant fibers	(i) Abundantly available in nature (ii) Highly crystalline in nature	Weeks to months	(i) Biomedical (ii) Drug delivery (iii) Cosmetics	[105], [106]

		(iii) Good insulation and anti-static properties (iv) Hydrophilic in nature		(iv) Clothes (v) Food packaging	
PTT	Petroleum-based monomers, Produced from 1,3- PDO and PTA/DMT	(i) Type of bio-polyester (ii) Possess extraordinary mechanical properties (iii) Excellent processability (iv) Good thermal steadiness	-	(i) Textile industry (ii) Automotive parts (iii) Mobile phone housings	[107]
PLA	Corn starch, wheat, sugarcane	(i) Very low in price (ii) Good resistance to UV radiation (iii) Low thermal stability (iv) Excellent use as matrix in composite material (v) Non-toxic to the environment	20 months	(i) Food packaging (ii) Drug delivery (iii) Highly used in composite materials	[108] – [110]
PHA	Obtained through bacterial fermentation of sugars or lipids	(i) Water insoluble (ii) Good ultraviolet resistance (iii) Soluble in chloroform and other chlorinated hydrocarbons (iv) Suitable for many medical applications due to its biocompatibility	12 months	(i) Food Packaging applications (ii) Industrial fermentation (iii) Animal nutrients (iv) Implant material	[111] – [114]
PHB	Produced and stored by bacteria	(i) Partially crystalline material (ii) Stiff and brittle (iii) PHB is compatible with the blood and tissues of mammals	6 to 10 months	(i) Wound dressing (ii) Act as absorbable nerve guides, (iii) Tissue scaffolding bone and nerve regeneration	[115] – [118]
PGA	Aliphatic polyester	(i) High degree of crystallinity (45–55%), (ii) High thermal stability (220–230 °C) (iii) High gas barrier (three times higher than ethylene vinyl alcohol)	5 months	(i) Drug delivery, (ii) Dental, and orthopedic applications (iii) Tissue engineering	[119]
PBS	Sugarcane, sugar beet, corn, potato, wheat	(i) High heat resistance among the general biodegradability resin (ii) PBS has excellent compatibility with a fiber.	2 to 3 months	(i) Bags, food and cosmetic packaging. (ii) Disposable products (iii) Agriculture industries	[55]
PVAc	Polyvinyl acetate	(i) Nontoxic and thermoplastic polymer (ii) Partially crystalline (iii) High tensile strength and flexibility (iv) Soluble in water	Several months to years	(i) Textile and paper industry (ii) Food packaging (iii) Biomedical applications	[120]
PCL	Petroleum-based products	(i) Excellent chemical and solvent resistance (ii) High employed in composite material (iii) Highly elastic in nature	6 to 28 months	(i) Surgical implants (ii) Drug delivery devices (iii) Regenerative medicine	[121] – [123]

1.1.1 Biodegradation mechanisms

Biodegradation of biopolymers occurs through the action of microbial microorganisms including algae, fungi, or bacteria. For instance, Kalita et al. [124] evaluated the biodegradation mechanism of algae/PLA-based composites and compared their performance with PLA-based composites. **Figure 3(a₁)** depicts steps involved in the biodegradation of biopolymer composite. ASTM standard D5338–15 was employed to evaluate the degradation behavior

under thermophilic composting and abiotic conditions. Scanning electron microscope (SEM) results provided in **Figure 3(a₂)** showed that rough patches containing holes and cracks were developed in biopolymer due to the excellent biodegradation. Additionally, PLA/ 5% algae extruded sheets (ES) specimens exhibited higher microbial attachment after two weeks of biodegradation, as illustrated in **Figure 3(a₃)**. The excellent degradation of biopolymers occurred due to the presence of high nitrogen content in biomass (algae).

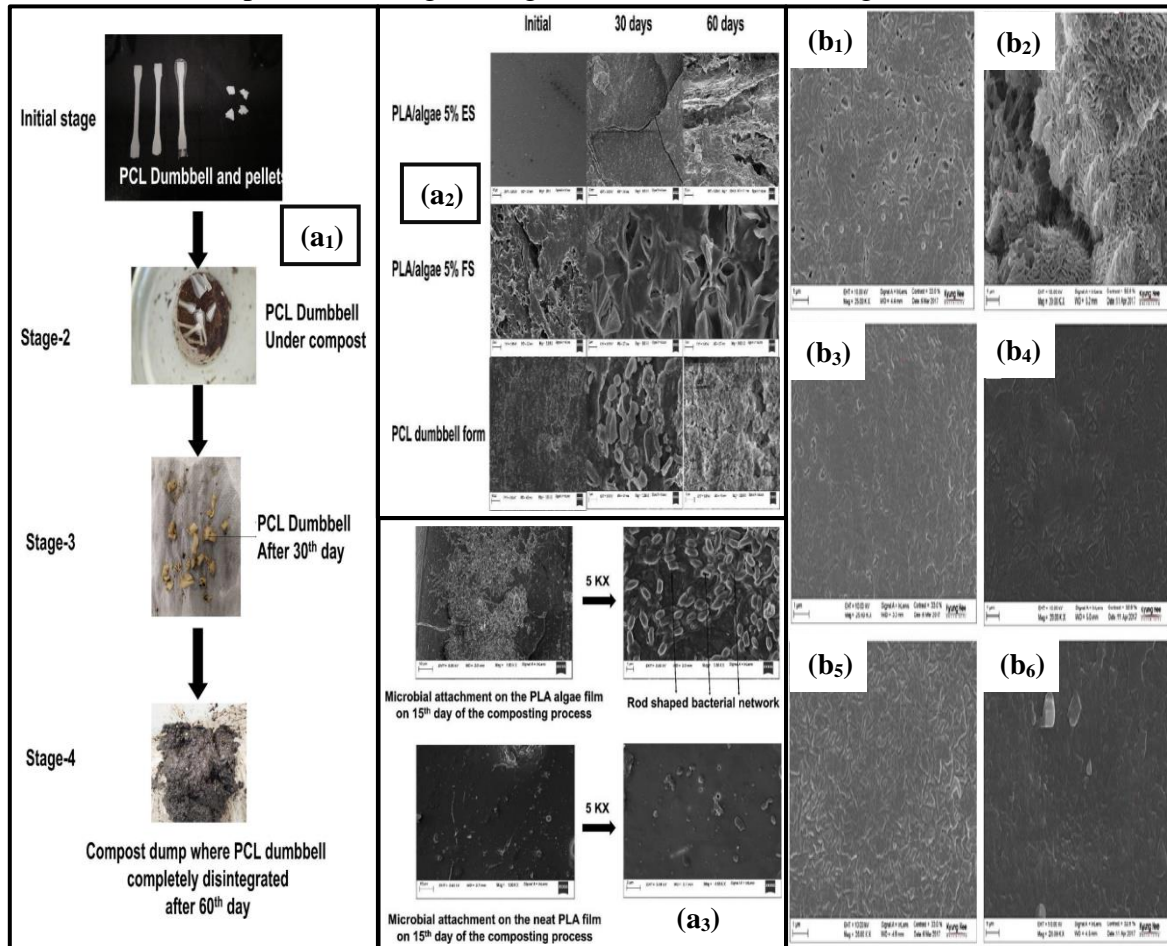


Figure 3. (a₁) Flowchart showing different stages of the biodegradation mechanism of PCL-based biopolymer; (a₂) SEM images of the biodegraded PLA-based polymers; (a₃) Microbial attachment comparison on the PLA/5 % algae ES and neat PLA ES films on the fifteen day of the composting process. (Adapted from [124]); SEM images depicting ungraded and degraded samples; (b₁, b₂) PEO40/PLA60/CNT1-based biopolymer before and after four weeks degradation; (b₃, b₄) PEO25/PLA75/CNT1-based biopolymer before and after four weeks degradation; (b₅, b₆) PEO10/PLA90/CNT1-based biopolymer before and after four weeks degradation [125] (**adapted with permission**)

The combination of inorganic nanomaterials, including nanowires, carbon nanotubes (CNTs), carbon nanofibers, or graphene, has opened a new research field, which helps in developing high-performance nano-devices [126]–[128]. For example, Zare et al. [125] developed poly(ethylene oxide)(PEO)/PLA-based nano biosensor by incorporating CNTs. SEM results observed for this study are shown in **Figure 3(b)** which demonstrated that the addition of CNTs accelerated degradation in PLA/PEO-based biopolymers by dwindling the interphase thickness during degradation.

Similarly, Urbanek et al. [2] proposed a low temperature biodegradable study of PCL, PBS, and PBSA using Antarctic microorganisms, as illustrated in **Figure 4(a)**. The reported results confirmed that highest biodegradations were achieved in the order of 49.68% for PBSA and 33.7% for PCL, 45.99% for PBSA, and 49.65% for PCL, respectively at 20°C under antarctic microorganisms, as depicted in **Figure 4(b)**. Finally, these antarctic microorganisms are best suited for effective plastic degradation at relatively low temperatures.

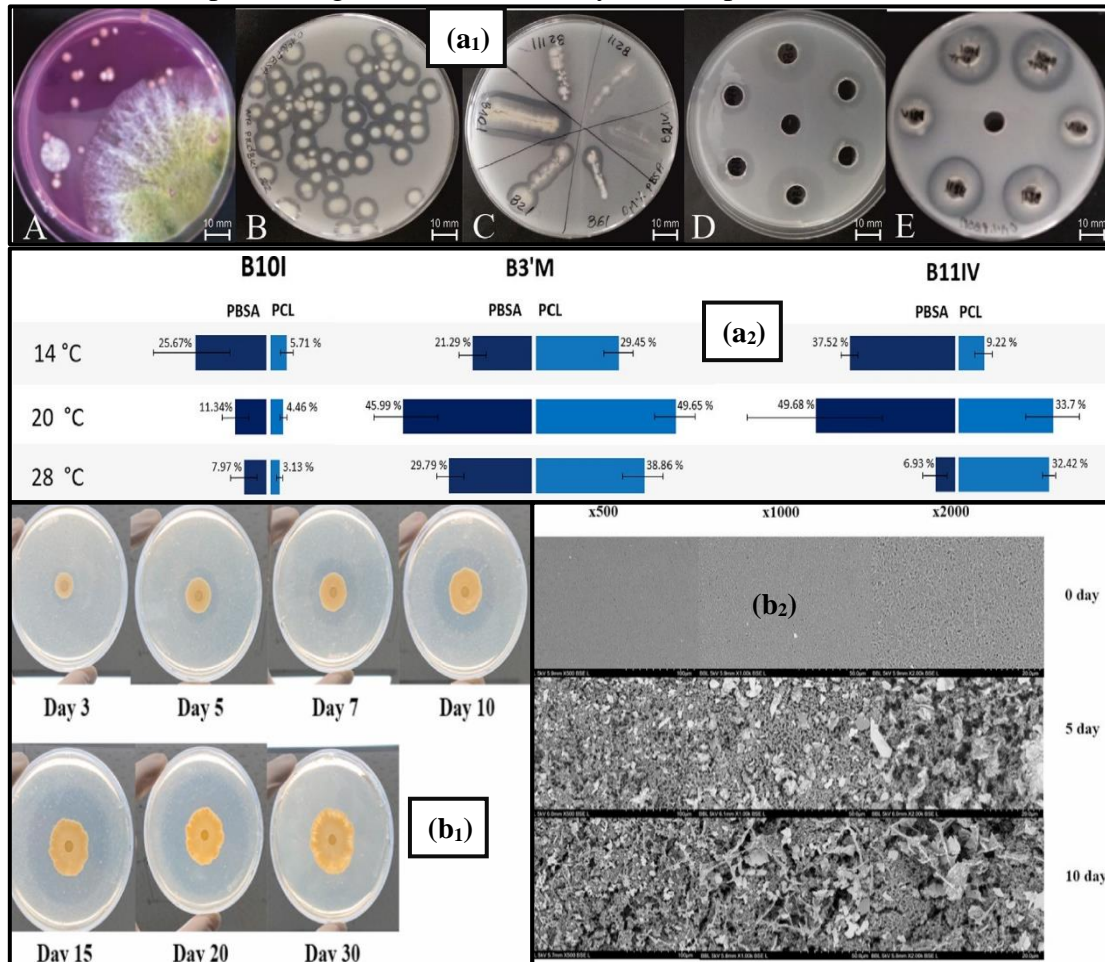


Figure 4. (a₁) Isolated antarctic microorganisms and halo zones that appeared around the colonies on FMM medium supplemented with 0.1 % PBSA (B, C, D, E) showing biodegradation; (a₂) Biodegradation rates for different studied biopolymers at different temperatures [2] (Adapted with permission); (b₁) Clear zones images representation generated by *Bacillus* sp. JY14; (b₂) SEM images depicting the surface changes as a result of degradation [129] (adapted with permission)

Cho et al. [129] evaluated different properties of PHB-based biopolymer through six different microorganisms commonly found in marine soil at different temperature and salinity levels. The PHB-degradation was monitored by noting the changes in the physical and chemical properties of PHB films incubated with *Bacillus* sp. JY14, as depicted in **Figure 4(b₁)**. **SEM results provided in Figure 4(b₂)** indicated transformation in the surface structure, molecular network, and weight of PHB-based biopolymer upon degradation. Additionally, a maximum of 98 % PHB degradation was achieved in just 14 days.

Similarly, Ruggero et al. [130] performed lab-scale degradation testing on Mater-Bi® (MB) film starch-based biopolymer and PBAT under different temperature and moisture conditions. **Figure 5(a)** depicts SEM images of degradation behavior in which circular spots disappeared and formed small holes. The results demonstrated that the biodegradation of PBAT is highly

affected by temperature and moisture conditions. Furthermore, among all three replicates, both starch and additives are completely decomposed within the first few days of the degradation process.

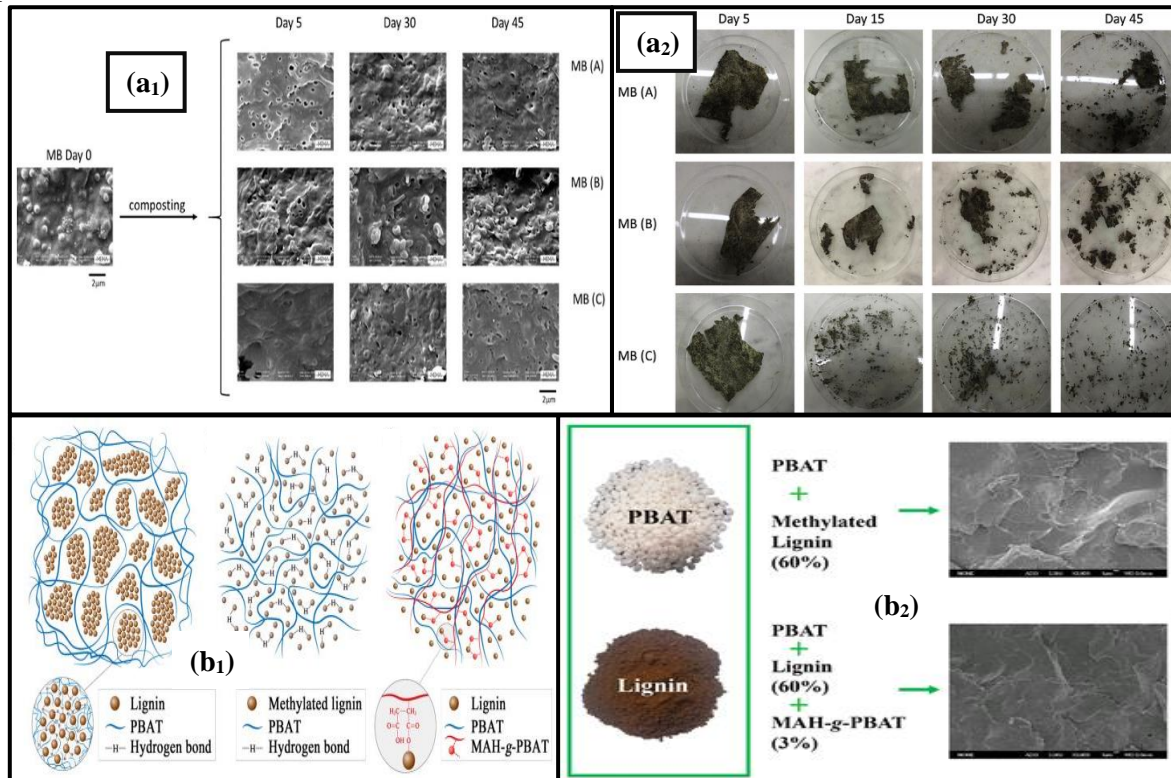


Figure 5. (a₁) SEM images of different A, B and C degraded samples during 5, 30, and 45 days of the composting process; (a₂) SEM images of the degraded samples during 5, 15, 30, and 45 days of the composting process [130] (Adapted with permission); (b₁) Lignin/PBAT-based biodegradable composite films/materials (Adapted with permission from [131]).

Wang et al. [132] developed lignin-based biodegradable composites by incorporating lignin into the PBAT matrix. For improving the compatibility, a green esterification reaction under microwave-assisted solvent-free conditions was adopted. These cost-efficient biodegradable materials with controlled mechanical properties are best suited for UV-shielding properties for packaging applications. In another study, Xiong et al. [131] evaluated the effects of lignin methylation and compatibilizer on biodegradable PBAT/lignin composites produced through melt extrusion, as depicted in **Figure 5(b)**. It was reported that the degree of lignin aggregation played a crucial role in the mechanical properties of these biodegradable composites. These biopolymer-based composites have a strong potential to be manufactured into rubbish bags, packaging bags, and soil remediation films. To summarize this, biopolymers exhibit excellent biodegradation behavior and comparable mechanical properties. These polymers can be composted leaving environment-friendly by-products including CO₂ and water.

1.2. Biodegradable composites

Biocomposites are environmentally benign, sustainable, renewable, carbon-neutral materials and eco-composites, which use NFs and biopolymers. In the last decade, these materials have spellbound the world due to their lightweight nature, recyclability, and cost-effectiveness [133]. Additionally, these composites contain fillers/additives and bio-based polymers that originate from renewable sources [134]. The decomposition of biocomposites occurs by soil burial or outdoor climate. Degradation occurs due to the complete breakdown of NFs that

damages the bonding between the fibers and biopolymer matrix. The biodegradability of biocomposites is influenced by UV light, temperature, air dampness, and microorganisms [135].

Nowadays, in biopolymeric composites, different NFs including plant-based (jute, cotton, bamboo, and flax), animal-based (silk and wood), mineral-based (asbestos), and microbial-based (polycyclic aromatic hydrocarbons, and bacterial cellulose), are vastly applied as reinforcement materials [136]. These fibers promote a sustainable green environment due to their appropriate mechanical characteristics and availability [137]. Because both fibres (reinforcement) and biopolymers (matrix) are derived from renewable sources and are compostable, biocomposites are excellent substitutes for carbon- and glass-reinforced polymer composites [138]. For instance, Duigou et al. [139] evaluated the mechanical performance of PLA/flax-based composites which exhibited mechanical properties comparable to polypropylene (PP)/hemp, PP/glass, and PP/sisal composites. Additionally, the shortening of fibers and separation of fiber bundles due to the recycling of composites did not influence the mechanical properties. Hence, these biocomposites are promising candidates for promoting a green and eco-friendly environment.

Biocomposites are mostly manufactured through the electrospinning technique which exhibits the versatility to use incorporate/reinforce continuous nano-fibers (2 – 500 nm) into different biopolymers, compared to other nano-fiber processing routes [140]. Additionally, this process possesses other advantages including tailorable physical properties, fiber functionalization, deposition upon substrate, material combination, and mass production capability [141]–[143]. Electrospun manufactured composites show high micro-sized pores and high surface area to volume ratio making them suitable for biomedical and air-filtration applications [144].

Similarly, the reinforcement of NFs in biocomposites as additives or fillers can be done through fused filament fabrication (FFF). The properties of the 3D-printed biocomposites depend upon the type of fibers, their strength, stiffness, and interfacial bonding with the biopolymer matrix [145]. **Figure 6** depicts different sustainable reinforcements and fillers which are employed for the fabrication of biocomposite materials [146]. The primary purpose of these reinforcements is to reuse different recyclable materials [147]. It lowers manufacturing costs while increasing biodegradability in biocomposites [148]–[150].

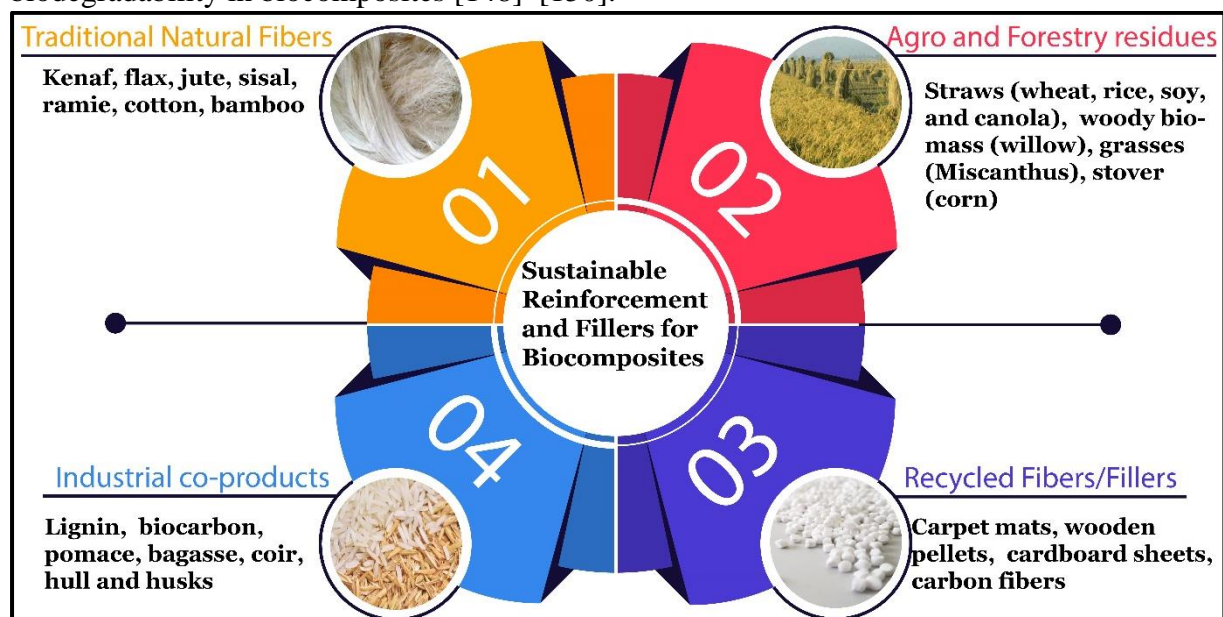


Figure 6. Different types of natural fibers employed for the manufacturing of biocomposites

The infusion of bio-derived nanofibers and nanoparticles (NPs) significantly improves mechanical and physical properties, including heat deformation temperature, modulus, gas permeability, tensile behavior, and decomposition of the developed composites. Most biocomposites contain starch, cellulose, PLA, PCL, PBS, PVA, and polyhydroxy butyrate (PBH). For instance, the reinforcement of sugar palm fiber in the biopolymeric composite significantly improved the mechanical characteristics [151]. The matrix constituent of the biocomposites is responsible for imparting better mechanical features with regard to their morphology, chemical constituents, molecular weight, and processing approach.

Different biodegradable fiber/polymer-based composites are evaluated by using various ASTM standards. For instance, soil burial testing calculates biodegradation by assessing the mass loss of polymers/fibers after testing [152]–[154]. Additionally, soil degradation occurs upon exposure to soil microbiomes. Soil burial testing is performed as per ISO 17556 and ASTM D5988-18 standards. Different influencing parameters, including moisture-holding limit, damping content, pH level, ash content, and carbon-to-nitrogen ratio have also been encountered during testing. Burial period and damping content have considerably influenced biodegradability, out of all these parameters [155]–[157]. **Table 4** incorporates the most recent studies of biodegradable NFs- and biopolymer-based composites, their testing techniques, degradation percentage, and time.

Table 4. Compilation of the recent work on the biodegradability of different NFs- and biopolymer-based composites

Biodegradable fiber- and polymer-based composites	Source	Biodegradation	Degradation time	Testing method	Ref.
CA	Wood Pulp	DS* of 1.8 achieved	Varies from several days to week	Reaction of CA with biocatalysts	[158]
PBS/jute composites	Cellulose	62.5 %	180 days	Soil burial	[159]
PLA	Corn, starch, sugar	100 %	9-12 days	Incubator with <i>Burkholderia cepacia</i> composite	[160]
Sisal fiber and TPS	Agave sisalana, starch	62 %	28 days	Soil burial	[161]
OPEFB fiber/ACC films	Palm oil plant	56.56 %	28 days	Soil burial	[162]
Polyester–Banana Fiber	Banana/euphorbia coagulum	41 %	90 days	ASTM D 5988	[163]
PLA-grafted-MA/GCF	Corn, starch, sugar	92 %	21 days	Incubator with <i>Burkholderia cepacia</i> compost	[160]
PE/NR composite	NR	38.3 %	90 days	Full-scale soil test (ASTM D 5988)	[164]
TPS composite	Starch	88.57 %	30 days	-	[165]
Feathers nonwoven reinforced polyester composite	Chicken feather fiber	62 %	60 days	Soil burial period	[166]
BF-reinforced tapioca starch PVA composite	Starch, cellulose	27 %	15 days	ASTM G160	[167]
Garden waste cellulose in starch/PVA	Starch, cellulose	80-91 %	14 days	Soil burial	[168]
Rich husk filler PLA composite	Rice husk, PLA	3.3 %	1 day	ASTM D-5488-94d	[169]
MOSF based PVA composites	Crude protein, crude oil	5-35 %	90 days	Soil burial	[170]

Corn based SB and CB, Cassava based SB and CB	Starch	12-57 %	14 days	Soil burial	[30]
Starch/MC composite	Starch, banana stem	82-96 %	28 days	Soil burial	[171]
SPNCC-based nanocomposites and SPS films	Cellulose, Starch	100 %	9-12 days	Soil burial	[172]
Starch/Chitosan/PVA composite	Chitosan, Starch	90 %	90 days	ISO 846:1997	[173]
PBS/levan composite	Polyester	Holes due to biodegradation were visible	4 days	Soil burial	[174]
RS-g-PMMA bio composite	Lignocellulosic	11 %	15 days	APHAS method	[175]
MNP/PVA Composite	MNP	0.141, 0.046, and 0.027 mg	Per hour	<i>Zophobas morios</i> Larvae (Superworms)	[176]
Ramie-based PLA composite	PLA	100 %	60 days	Soil burial	[154]
CNC/chitosan composite	Cellulose	Moderate biodegradability	15 days	Soil burial	[177]
WPC	Cellulose	3.55 weight loss per gain	-	ASTM D5338-11	[178]
BF/PLA composite	Lignocellulosic, PLA	12-28 %	10-70 days	ASTM D5338-11	[179]
AEMO-NVP composite	Leather	1.25-5 %	7-28 days	Soil burial	[180]
Oxo-PP composite	Transition metal ions	28.87 %	45 days	ASTM D 5338	[181]
PWBF-PVA composite	Cellulose, PVA	3.5 g weight loss was observed	60 days	Soil burial	[182]
Jute composite	Cellulose, rice bran	0.2-2.91%	7-28 days	Soil burial	[183]
Polyester composite	Shorea robusta	6.2 %	15-120 days	Soil burial	[184]
FCC	Cellulose	48 NI kg ⁻¹ net biogas produced as a result of biodegradation.	21 days	German standard DIN 38414	[185]
rPP and KF composites	Cellulose	11.82 %	4 months	Soil burial	[186]
Coffee wastewater	Carbohydrates	80-90 % BOD	10-60 days	Simulation technique	[187]
CG and gelatin films	Gelatin	45 %	22 days	ASTM D5988-12	[53]

*DS: Another way of measuring biodegradability. A level of 1.8 DS shows good biodegradability.

In summary, biopolymeric composites are sustainable and eco-friendly materials, which exhibit excellent biodegradability and mechanical properties. These composites promote a cleaner and greener environment. The development of composites by using recycled fibers gives comparable mechanical properties and can be applied for high-performance applications.

3. Life cycle of biopolymers

Life cycle assessment (LCA) is a system employed for analyzing the environmental and ecological footprints of products and materials. This is usually done through the evaluation of different life stages of products, including extraction of raw materials, fabrication, use, and end-of-life stage [188]. The ultimate life cycle of biodegradable materials is depicted in **Figure 7**. In the first step, biopolymers are often derived from agricultural products and involve the cultivation of different raw materials, such as potatoes for starch, wood for cellulose, or any crop used. These polymers are manufactured in substages: fertilizers, pesticides, and herbicides

also help in the manufacturing of biopolymers. Production and milling are the other two key processes, which help in developing biopolymers [189]. In the second step, these polymers are employed to develop different consumer products which exhibit improved physical and mechanical properties. Additionally, the mechanical properties of these composites can be enhanced by infusing organic nano-materials [190]. In the third step, the EOL of biopolymer-based consumer products seeks to evaluate the impacts associated with the production and disposal of biopolymers compared to fossil-based plastics. Different routes, including landfill, compost, and recycling, are used for different biopolymers, whereas some compostable biopolymers including PLA and TPS are modeled using compost and landfill scenarios [191]. In the last step, biopolymer degradation occurs in the biological environment through different microorganisms including algae, fungi, and bacteria [192]. Consequently, more than 90 % of biodegradable polymers are transformed into water, CO₂, and minerals within six months as per EU standard.

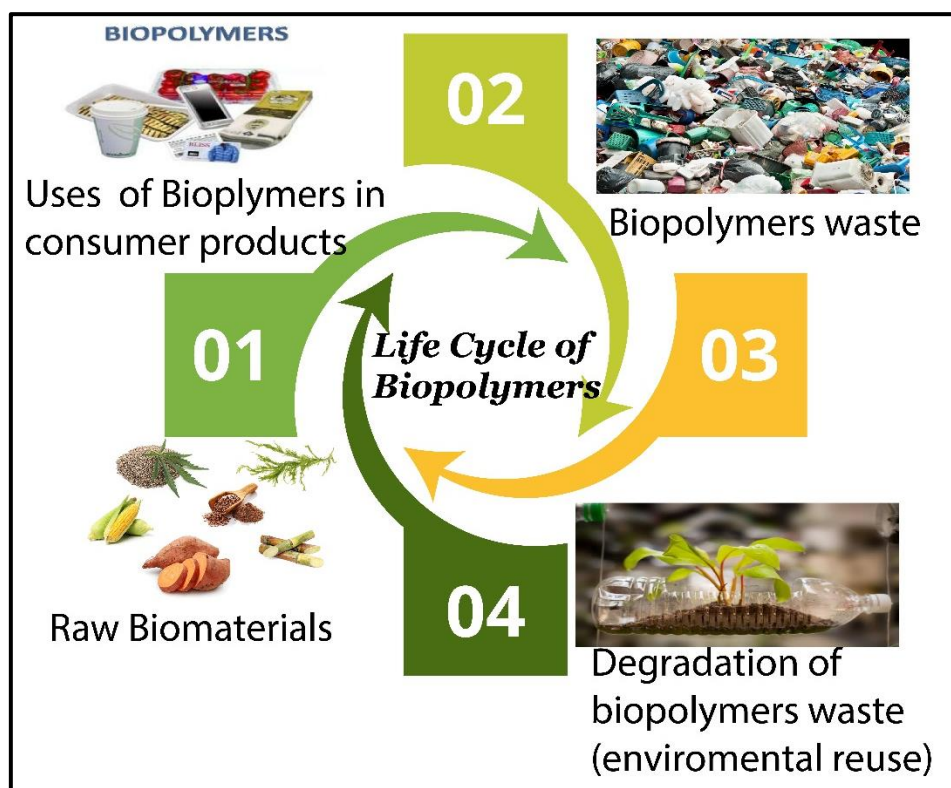


Figure 7. Life cycle of biopolymers.

4. Applications of Biopolymers

Biopolymers have exhibited huge potential due to their flexibility in chemistry that gives rise to materials with a great diversity of physical and mechanical properties [193]–[195]. Since their emergence, biopolymers have been effectively utilized in many biomedical and other engineering applications including controlled drug delivery, food packaging, construction industry, regenerative medicine, wearable electronics, orthopedic, and long-term implants [196]–[198]. Rapid advancement in materials science and technologies have opened up new windows of opportunities to discover the novel applications of biopolymers in engineering and biomedical fields [199]–[201]. The focused researches on these materials are reflected by a significant upsurge of the biodegradable polymer-based marketed products and ongoing clinical trials of these materials [202]–[204]. Particularly, in the last decade, biopolymers

experienced a renaissance due to the limitation of fossil resources in combination with public demands for environmentally-friendly and sustainable processes, which has led to the formation of a market for bio-based plastics [205].

These eco-friendly biopolymers and their composites can be classified based on their various sources, numerous techniques of manufacturing, and possible forms of usage [206]. Because of their excellent biocompatibility and biodegradability, these polymers are ideal for use in environmental applications [207]–[209]. Biopolymer-based materials are also responsible for reduced carbon footprint during their manufacturing and their microbial degradation improved overall resource efficiency [210].

4.1 Medical applications

Numerous biodegradable polymers and composites including PLA, PVA, PCL, and PHB are being used in tissue engineering [211], orthopedic implants [212], and drug delivery [213]. These biodegradable materials have no harmful impact on the human body, thus, making them highly appropriate for medical applications [214]. PCL-based materials have a wide scope in wound healing applications, drug delivery systems, and tissue engineering applications, especially in the formation of hard tissues, soft tissues, and load-bearing tissues [215]–[217]. **Table 5** contains some of the most recent studies containing the applications of biodegradable biopolymers and their composites.

Table 5. Recently adopted different biopolymers based single/hybrid materials and their engineering/biomedical applications.

Researcher	Year	Biopolymers	Added materials / fillers	Engineering applications
Zare et al. [125]	2022	PLA/PEO	CNT	Degradation-based nanobiosensor
Yang et al. [218]	2020	PLLA	Xeolitic imidazolate framework-8 (ZIF-8),	Bone scaffold
Yuan et al. [219]	2019	Bacterial cellulose and chitosan	Collagen	Hemostatic material
Mishra et al.[220]	2021	Chitosan-casein	Nanofibrous polyelectrolyte complex	Hemorrhage treatment, provide rapid blood clotting
Yalcin et al.[221]	2021	PHB	siRNAs	Gene delivery system
Malik et al. [222]	2018	PCL	Organo modified montmorillonite clay	Fixation devices
Mucke et al.[223]	2021	TPS	PBAT	Packaging
Kundu et al. [224]	2021	PCL	nHAP fibers	Tissue engineering
Diaz et al. [225]	2020	PCL	nHAP/MWCNT	Bone regeneration
Villamagna et al. [226]	2020	PEA	Peroxisome proliferator-activated receptor δ	Drug delivery
Das et al. [227]	2019	PLA	MWCNTs	Chemo, photothermal therapy and tissue engineering
Mondal et al. [228]	2020	PLA	HAP	Drug delivery and tissue engineering
Singh et al. [229]	2019	Chitosan	Nano-sized BG	Bone scaffold and bone tissue regeneration
Lucik et al. [230]	2020	PLA/PCL	Thymol and carvacrol mixture	Food packaging,
Mouro et al. [231]	2021	PCL/PVA	Chelidoniummajus L.	Wound dressing
Chugh et al. [232]	2020	PEA	Dimer acid	Hot-melt adhesive

Tseng et al. [233]	2020	PLA	-	cutlery products
Zhu et al. [234]	2020	PVA/CMC	Na ₂ CO ₃	Paper processing, adhesives and coatings, water-soluble films, emulsifiers, textiles
Ge et al. [235]	2021	Gelatin	Agarose gel	Gelling agent
Mirzaeei et al. [236]	2021	PCL and PLGA	Metronidazole and amoxicillin	Drug-delivery systems
Dietrich et al. [237]	2019	PHB	Hardwood holocellulose hydrolysate	Detoxification system
Zhao et al. [238]	2020	Chitosan	EDC and NHS	Artificial liver
Zhao et al. [239]	2019	PLGA	MSCs	Ligament/Tendon Repair
Buyuksungur et al. [240]	2021	PCL	GelMA	Dentistry
Parvizifard and Karbasi [241]	2019	PHB	MWCNT	Tissue engineering
Ferlic et al. [242]	2020	PHB	ZrO ₂	Surgical implant
Deng et al. [243]	2021	PCL	PEG/chitosan-keratin	Sutures
Arjona et al. [244]	2021	PHB	-	Microcapsules
Subramanian et al. [245]	2020	PCL	SMA hydrogel	Contraceptive devices
Caballero et al. [246]	2021	Alginate	-	Theragnostics
Robles et al. [247]	2019	PLA	Lignin	Wound healing

Besides these, biopolymers are also promising materials for many filter applications due to their desirable chemical and physical properties which are best suited for absorption and the removal of certain chemical contaminants, stipulate bactericidal, or viricidal functionality. Different functional groups present in proteins and polysaccharides permit highly selective filtration for pollutants and other contaminants [248], as illustrated in **Figure 8(a)**. Hemorrhage or excessive blood loss during childbirth, accidents, and complex surgeries is the leading cause of death. Bleeding management is an effective strategy to control excessive blood loss, which can be achieved using topical hemostatic agents through different biopolymers [249].

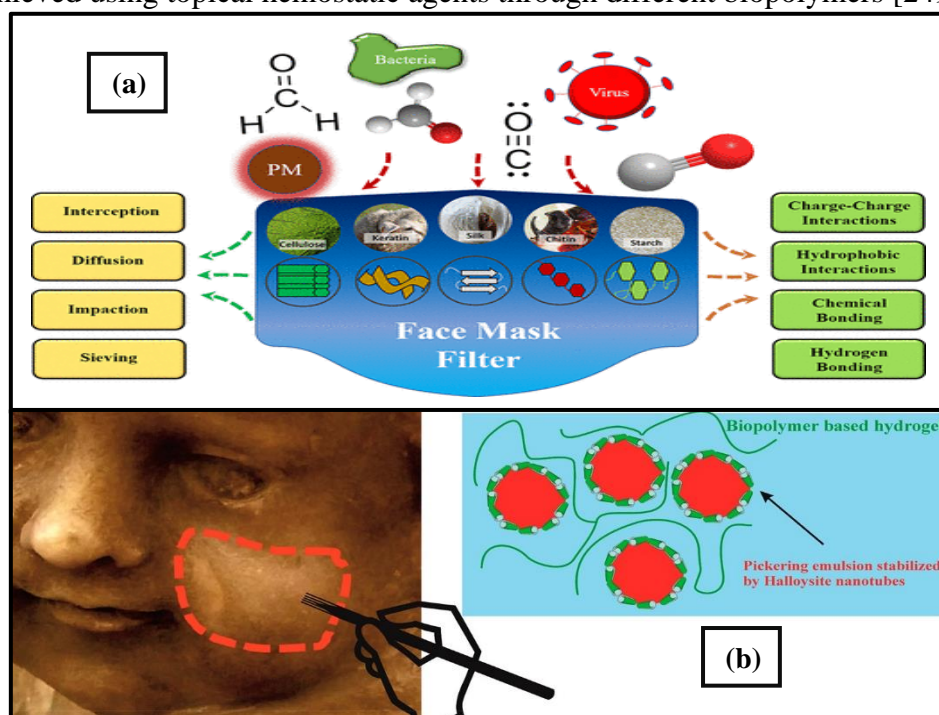


Figure 8. (a) Filtration materials produced from different biopolymer including variety of protein cellulose, silk, keratin, chitin, and starch [248] (adapted with permission); (b) Pickering emulsions based on halloysite nanotubes and ionic biopolymers (chitosan and pectin) [250] (adapted with permission).

Cavallaro et al. [250] fabricated sustainable Pickering emulsions using halloysite nanotubes and ionic biopolymers including chitosan and pectin. The reported results showed that Pickering emulsion in a pectin-based gel phase has the potential for wax layer removal from a marble surface, as depicted in **Figure 8(b)**. Furthermore, pectin improves the Pickering emulsion stability, while a nonhomogeneous gel and phase separation were detected in the presence of chitosan.

4.1.1 Tissue Engineering

Tissue engineering is a reliable method for supplying perfectly matched organs for the necessary transplantation of the patient due to injury, trauma, or any chronic disease [251]. Thus, tissue engineering is a lifesaving technique due to the shortage of donors at vital times when a patient's life can be in severe danger [252]–[255]. To date, different organs/parts including kidney, liver, orthopedic, nerve, pancreas, ear, and nose have been implanted. This is entirely possible due to scaffolds fabrication with the use of 3D-printed biodegradable polymers [256]–[259]. These biopolymer-based printed scaffolds have excellent biodegradability, compatibility, porosity, osteointegration, size, and bioactivity [193], [260]–[262].

For instance, Jinga et al. [263] developed composite scaffolds by using PCL-based biodegradable polymer along with other inorganic mineral powders including zinc oxide (ZnO), titanium dioxide (TiO₂), and hydroxyapatite (HAP) via electrospinning method. Upon successful experimentations, it was noted that these fiber-based composites are accepted by cell structures. Hence, these scaffolds could be used for regeneration of damaged parts and wound healing applications. The PCL/collagen-based electrospun fibrous membrane was evaluated for wound healing applications through an in-vitro study [264], as depicted in **Figure 9(a)**. The results indicated that biopolymer-based exhibited polarized and ordered fibroblasts, and these fibroblasts possessed excellent cell proliferation and focal adhesion. A randomized controlled clinical trial targeted to estimate the feasibility and effectiveness of PCL-based scaffold in fresh extraction sockets for ridge preservation was employed [265], as depicted in **Figure 9(c)**.

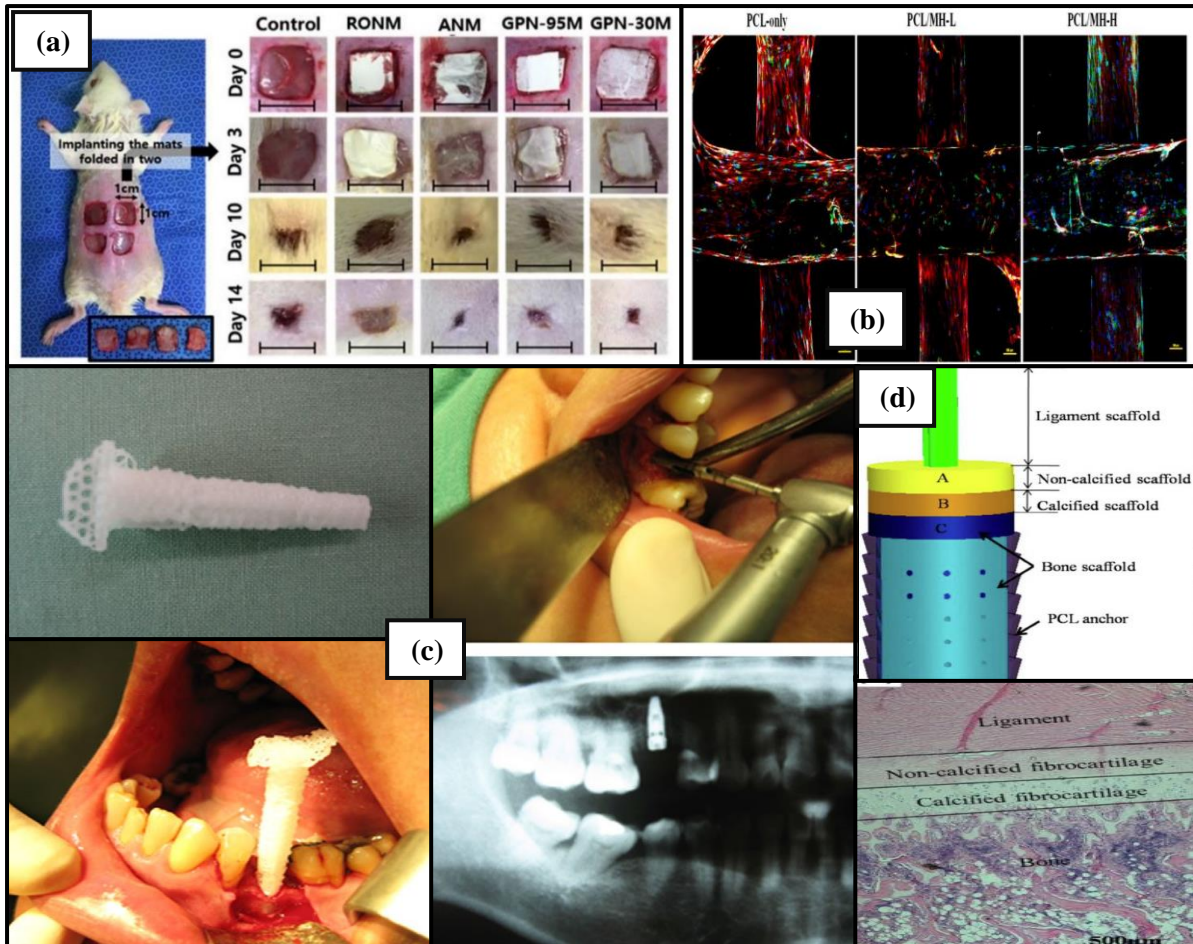


Figure 9. (a) Time dependent healing process of rat wound using PCL/collagen mats [264] (Reproduced with permission); (b) Immunostaining for the osteogenic marker images of osteoblasts on 3D-printed scaffolds (Adapted with permission from [266]); (c) PCL scaffold during surgical implant and tooth extraction site [265] (Reproduced with permission); (d) Design of a scaffold for ligament, cartilage, and bone generation [267] (Reproduced with permission)

Similarly, PCL/HAP/simvastatin electrospun nanofiber-based composite coating was deposited on a magnesium substrate by Rezk et al. [212] and observed to have excellent biodegradability and enhanced mechanical properties, which could be helpful for their utilization in orthopedic implants. In another study, Abdal-hay et al. [266] 3D-printed a PCL-based biodegradable composite by incorporating bioresorbable magnesium hydroxide (MH) NPs. The results revealed that the inclusion of MH significantly improved the mechanical properties and accelerated the degradation rate of PCL in phosphate-buffered saline solution. Furthermore, the cell bridging was greater on the PCL/MH of 20 wt.% compared to PCL- and PCL/MH (5 wt.%) scaffolds. Additionally, full bridging of scaffold pores was noted in the PCL/MH of 20 wt.% scaffolds. The cell structure and bioactivity were also examined using immunofluorescence staining, as shown in **Figure 9(b)**. Thus, these materials have broad implications for the manufacturing of bioactive 3D-printed scaffolds with tailorable degradation properties and are ideal candidates for the regenerative treatment of various injuries or bone diseases.

Soni et al. [268] proposed a promising technique for the healing of bone tissues using biocompatible and biodegradable bioactive glass/PCL-based thin membranes and observed excellent mechanical properties and degradation. **Figure 9(d)** illustrates the multiphase

scaffolds designed by He et al. [267] for the bone regeneration. In another study, Makkar et al. [269] developed biocompatible and biodegradable PCL-based coatings for their potential use in orthopedic applications. Magnesium alloy (Mg) was immersed in hydrofluoric acid solution (F_2) which yielded MgF_2 . **Figure 10(a)** depicts in-vitro analysis performed on the rabbit model using duplex MgF_2 /PCL coating. The results displayed excellent cell adhesion, good cell viability, and cell proliferation, compared to uncoated Mg samples.

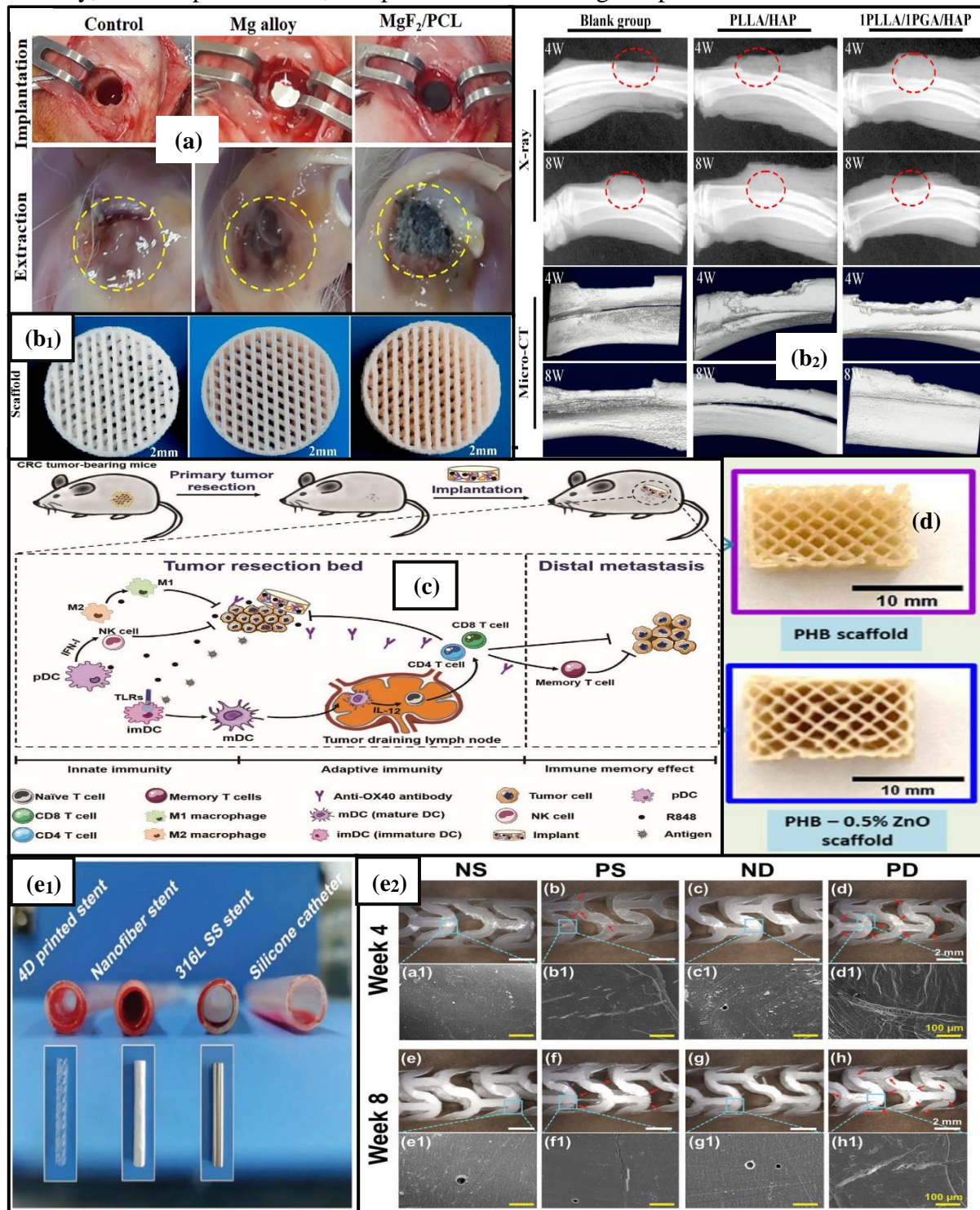


Figure 10. (a) Figure shows the photos of rabbit's damaged part at the time of implantation and extraction (Reproduced with permission from [269]); (b1) Optical images of the PLLA (90 wt.)/HAP (10 wt.), PLLA (45 wt.)/PGA (45 wt.)/HAP (10 wt.) and PGA (90 wt.)/HAP (10 wt.)

scaffolds (adapted with permission [270]); (b₂) X-ray radiographs and micro-CT images of the different PLLA (45 wt.)/PGA (45 wt.)/HAP (10 wt.%) scaffold, PLLA (90 wt.)/HAP (10 wt.%) and blank group after four- and eight-weeks implantation. (Adapted from [270]); (c) Schematic diagram explaining the biopolymer immune implant co-loaded with R848 and anti-OX40 antibody [271] (Adapted with permission); (d). PHB- and PHB/ZnO-based composite scaffolds manufactured through 3D printing technique [272] (Adapted with permission) (e₁) Cross-section images of different stents models including PLA nanofiber stents after circulation for 2 hours (e₂) Comparison of degradation behavior of PLA stent in different environment through optical and SEM images for 4 and 8 weeks (red arrows point to the cracks) [273](adapted with permission)

Shuai et al. [270] investigated HAP/poly-l-lactic acid (PLLA)-based bone scaffolds by blending poly(glycolic acid) (PGA) through 3D printing, as depicted in **Figure 10(b₁)** which increased its bioactivity and osteoconductivity similar to the living bone. **Figure 10(b₂)** depicts implantation results demonstrating that the inclusion of PGA improved the degradation rate of scaffolds and weight loss increased from 3.3 % to 25 % after immersion for 28 days. Additionally, numerous pores produced by the degradation of the scaffolds helped the exposure of HAP from the matrix, which not only activated the deposition of bone-like apatite on the scaffold but also accelerated apatite growth.

Hanas et al. [274] developed electrospun PCL/HAP-coated AZ31-based composites using a friction stir process. The developed composite exhibited controlled degradation at a normal pH of 7.2 and physiological temperature. The in-vitro study showed better proliferation and adhesion as compared to uncoated scaffolds. Thus, these composites have promising nature of being used in implant applications for tissue regeneration. Likewise, Ji et al. [271] studied biopolymer-based immune implants for colorectal cancer (CRC) post-surgical therapy, as depicted in **Figure 10(c)**. Immunological results demonstrated that the biopolymer immune implant treatment was performed in a two-stage action, with enhanced natural killer cell infiltration and activation of dendritic cells during the first few days. Consequently, an increased population of infiltrating T cells established immune memory effects which prevented tumor recurrence. In summary, biopolymeric sustainable composites are extensively applied for tissue regeneration of different organs due to their excellent biocompatibility, biodegradability, non-toxicity, and sustainability. Wang et al. [273] studied degradation behavior of PLA stents and their failure behavior in a dynamic condition after self-expandable deployment as depicted in **Figure 10(e₁)**. Results showed that PLA stents exhibited excellent compression force and recovery ratio. **Figure 10(e₂)** illustrates PLA stents complete degradation behavior in different environment and degradation conditions (NS, PS, ND and PD in **Figure 10(e₂)** refers to no/pre deformation + static/dynamic degradation conditions respectively). PLA stents demonstrated minimal micro structural damage at 60 °C followed by 8-week degradation tests after their implantation in blood vessels. Finally, this microstructure damage triggered by deployment were responsible for acceleration in the degradation of PLA stents than fluid shear stress.

4.1.2. Drug Delivery Systems

Drug delivery is a core pharmaceutical research area in which biopolymers can be employed in microsphere, microcapsule, hydrogel, or implant forms. These drug delivery systems do not require any invasive procedures for removing devices after drug release [275]. Biopolymers with various drug formulations influence the drug delivery process in various ways because of several side effects and a high level of toxicity of synthetic polymers [276]. Synthetic polymers are associated with many harmful effects, including delays in treatment, and dose reduction

[277]. The controlled release of a therapeutic agent in a tunable dose is considered a prerequisite for any drug delivery system. This can be achieved through conjugation between a therapeutic agent and a drug delivery vehicle [278].

Biopolymeric materials possess extraordinary features including biocompatibility, non-toxicity, water-solubility, and biodegradability, which make them highly suitable for controlled drug delivery [279]. These drug delivery systems also prohibit enzymatic decomposition before delivering therapeutic media at the targeted locations. A variety of stimuli including chemical, physical, thermal, or combination of any of these stimuli can help the release of therapeutic medicine [280]–[282]. Biopolymeric materials in combination with HAP are often employed as potential nano-carriers. It is due to its high biocompatibility, high surface activity, high surface-to-volume ratio, and adequate biodegradability of HAP. For instance, Macha et al. [283] developed PLA/coralline HAP-based biodegradable composites for their potential use in drug delivery systems. This approach holds a good strategy to repair the defected bones using medicines and minerals through biodegradable polymer-based microcapsules. These drug delivery devices exhibited excellent ability to load and release protein, nucleic, antibiotics, and cancer drugs at a slow rate in the human body. In summary, biopolymer-based microsphere and microcapsule can encapsulate hydrophilic as well as hydrophobic drugs and regulates their targeted and controlled release at the desired location.

4.2 Electronic applications

Flexible sensing devices have been attracted worldwide due to their extensive uses in wearable sensors, smart textiles, actuators, electronic skins, flexible displays, and medical devices [284]–[286]. These devices can monitor human motions and physiological signals (referring to **Figure 11(a)**) in long term for providing clinical information for the diagnosis and prevention of diseases, as well as rehabilitation therapy. These devices are usually fabricated with synthetic polymers, metal oxides, and semiconductors [287]–[289].

Different nanofiller-based biodegradable polymer composites have found their applications, including solar-assisted cells, electromagnetic devices, and electronics [290]. The rapid growth in the daily usage of electronic devices will cause serious environmental problems due to the chemical toxicity of unused electronic devices [291]. However, the use of biodegradable polymeric composite-based devices can tackle this problem.

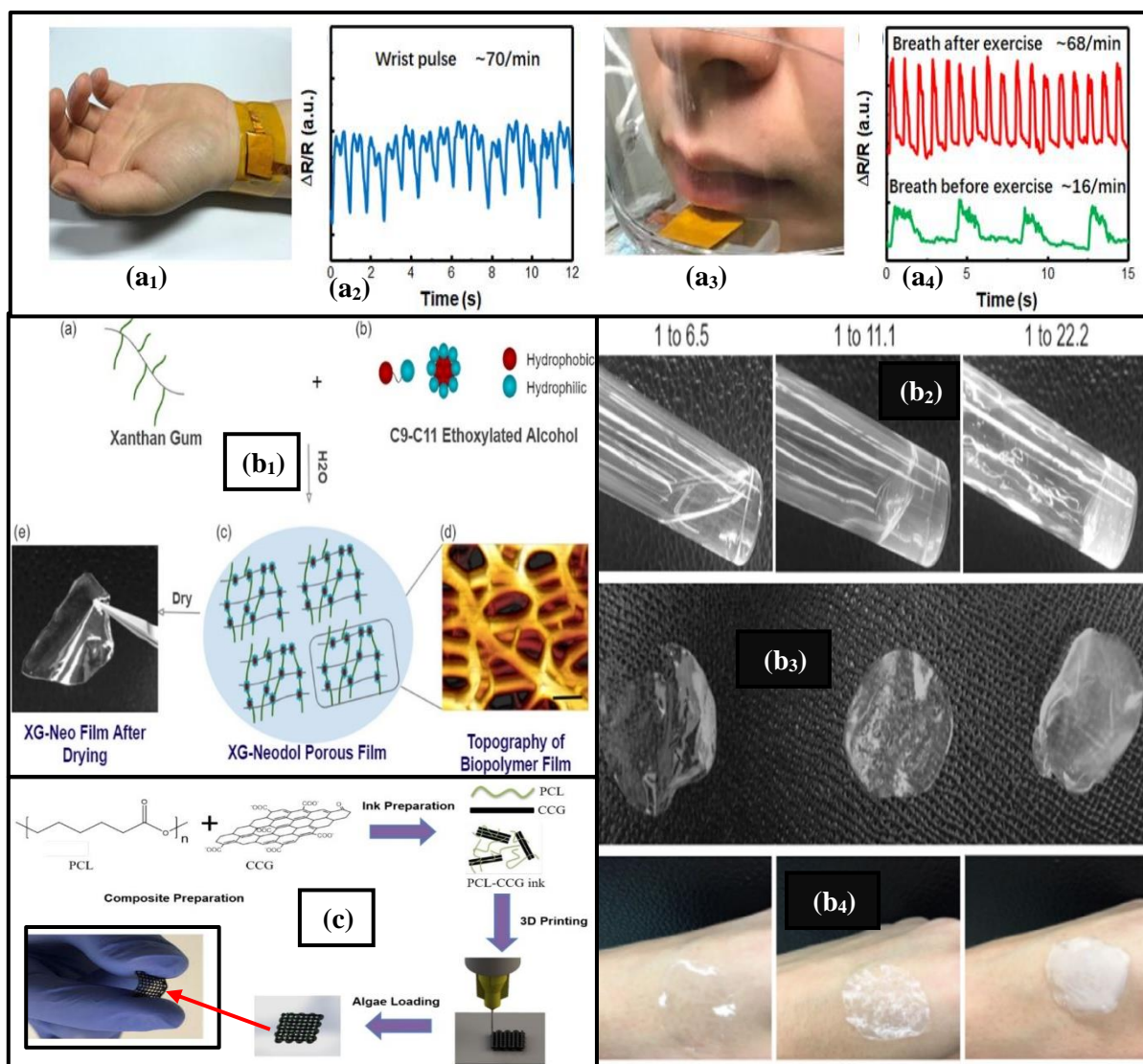


Figure 11. Different applications of naturally available materials in electronics; (a₁) Wrist pulse detection application; (a₂) Pulse waveform of the tester; (a₃) Application in respiration detection; (a₄) Results of breathing before and after exercise recorded by respiration detector [292] (Adapted with permission); (b₁) Fabrication of porous biopolymer films from the polysaccharide, xanthan gum and the ethoxylated alcohol, Neodol; (b₂) Macroscopic images of the viscoelastic solutions and gels before and after drying at room temperature; (b₃) Films fabrication after taking 500 µL of each solution and allowing to dry yields films of different thicknesses, elasticities and transparencies; (b₄) Dry films placed on skin. [293] (adapted with permission); (c) Schematic diagram of procedure to form multilayer 3D-printed PCL/graphene composite electrodes, and the subsequent examination of electrochemical response via immobilization of algae (adapted with permission from [294]).

Fabijanac et al. [293] produced UV-protected film through different novel biopolymers including Neodol, xanthan gum, anionic polysaccharide, and a non-ionic surfactant, as depicted in **Figure 11(b)**. Results from electron plasmon resonance showed the free-radical reducing ability (3.5 times), while liquid chromatography-mass spectroscopy quantifies the UV-decomposition of sinapyl alcohol. Thus, this material exhibits excellent potential for its utilization as biosensors due to its inherent tenability and flexibility. In another study, PCL/chemically converted graphene (CCG)-based composite electrodes containing 10 wt.% of graphene were developed through 3D printing by Lee et al. [294]. These novel printable eco-

friendly composite electrodes are shown in **Figure 11(d)** exhibited excellent printability, processability, and robustness. Furthermore, the biocompatibility of electrodes was illustrated through electrochemical response by growing the unicellular microalgae onto PCL/CCG substrate. The printed PCL/CCG product can be potentially applied in bioelectronic applications.

4.3 Miscellaneous applications

The utilization of protective facemasks has increased rapidly due to the coronavirus disease 2019 (COVID-19) pandemic, and these facemasks are usually fabricated using non-biodegradable synthetic polymers, which are causing environmental problems [295]. Biopolymers are effectively applied to manufacture efficient air filters for facemasks [296]. These biopolymer-based facemasks are manufactured by incorporating nanofibers through the electrospinning process, which helps in controlling nanofiber dimensions, fiber length, and pore size [297]. Additionally, silver NPs can be used for doping fiber membranes of facemasks which permits antimicrobial characteristics as well as increases surface roughness. **Figure 12** depicts the proposed processes, biopolymers, and requirements for developing biodegradable facemasks [135].

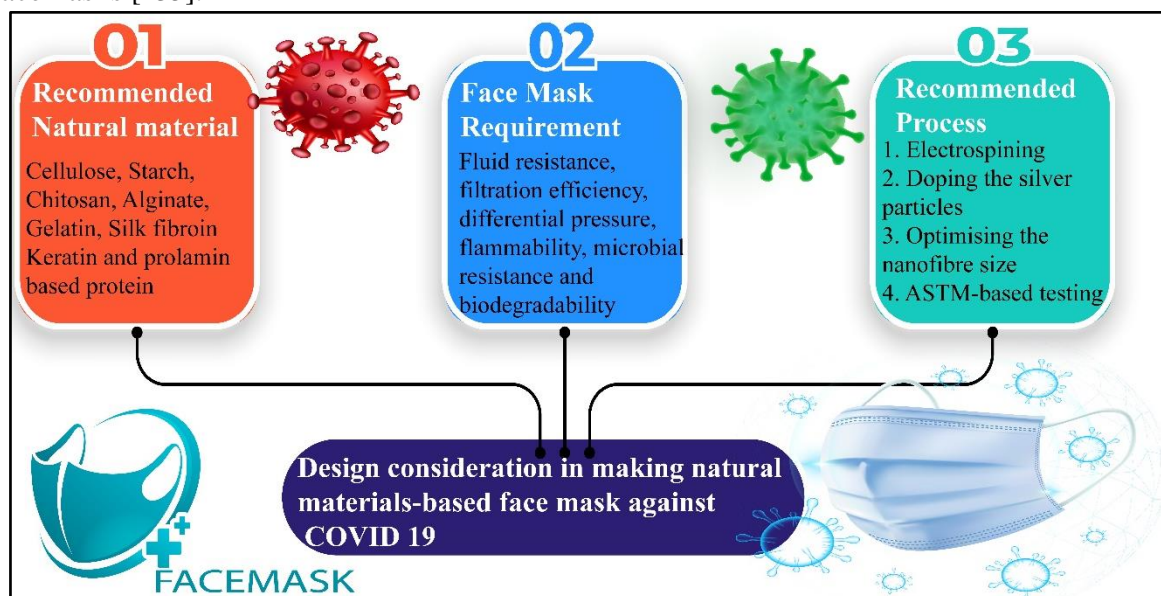


Figure 12. Proposed natural biopolymers, requirements, and processes for the fabrication of environmentally benign facemasks.

In the contemporary world, biopolymers are vastly applied in modern packaging and cosmetics industries due to their non-skin irritant and non-toxic nature. For instance, soy protein exhibits improved color profiles [298]. Additionally, biopolymers can be added for manicure preparation, sunscreen, scrubbing agents, and cleaning products [299]. Similarly, bivalent chelating-mineral ability and anti-microbial behavior make chitosan an excellent choice to be used as a packaging material for the preservation of edible foods [300]. Biopolymers including PLA, PHA, PBAT, and PBS exhibit comparable mechanical properties, compared to non-biodegradable polymers, and can be excellent substitutes for fabricating hygienic products and packaging materials [301].

In the textile industry, biopolymers have also gained significant attraction in the form of bed linens, towels, bandages, wipes, as well as geotextiles [302]. Superior antimicrobial properties of collagen, and alginates, permit their utilization for fabricating textile products [303].

Similarly, chitosan is commonly applied in healthcare or medical textiles for developing wound care dressings and bandages due to its biocompatibility, cationic biodegradability, and antimicrobial features. Bio-based PTT and PLA have also been employed for the manufacturing of textiles due to their extraordinary dyeability and better strength [304]. Whereas, in the agricultural industry, biopolymers mostly exist in the form of biodegradable plant pots and mulch films [305]. Similarly, the controlled release of fertilizer in the soil can be achieved using biodegradable composites [306]. **Figure 13** depicts the most recent applications of biopolymers in different engineering sectors.

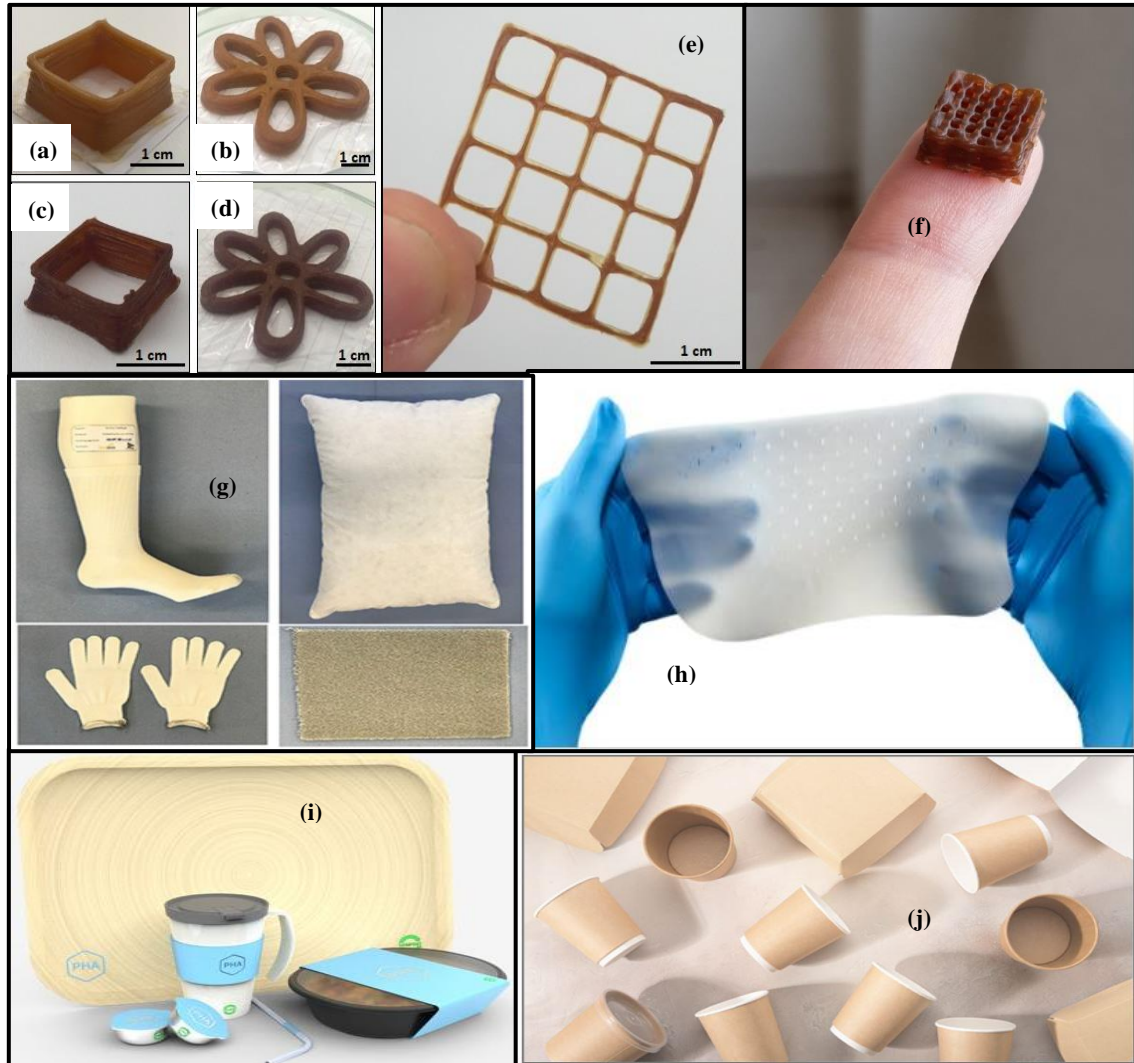


Figure 13. Commercially available products from biopolymers; (a) Appearance of a hollow cube right after the printing process; (b) At drying conditions at ambient for 24 h; (c) A flower model before drying; (d) After drying at ambient conditions for 24 h; (e) Three-layered grid structure printed from hemicelluloses; (f) A scaffold prototype printed from hemicellulosic pastes (a-f adapted from [307]); (g) Textile products from biopolymers (@AMIBM Maastricht University 2018); (h) Collagen matrices or sponges can be used to treat wounds for tissue regrowth and reinforcement [308]; (i) PHA-based biopolymer in food packaging (according to Cambridge Consultants)

In the last few decades, there has been an increasing concern over food contamination and most fatal contaminations occur due to the presence of microorganisms [309]. Biopolymers are blended with NPs, including nanocellulose, CNTs, zinc, silver, magnesium, copper, and gold NPs, to develop biopolymeric nanocomposites [310]. Biopolymeric sustainable

nanocomposites are extensively applied in food packaging applications to extend the shelf life of food due to improved physical, thermal, mechanical, and biodegradable properties [311]. **Figure 14** depicts the various biopolymeric materials used for food packaging applications.



Figure 14. Commercially available products from biopolymers; (a) The Midwestern Pet Foods-branded PlantBag containing at least 30% bio-based PE extruded with fossil fuel-based PE (adopted from [312]); (b) Image of the PlantBottle™ made up to 30% from biomass and 100% recyclable. (Picture Courtesy of the Coca-Cola Company (Atlanta, USA)); (c) 100% Recyclable Bio-PET Bottle (adopted from [313]); (d) Biodegradable food tray made of PHB obtained by injection molding (adopted from [314]); (e) Biodegradable packaging articles based on starch [314]; (f) CNC-based biopolymers for packaging applications (adopted from [315]); (g) PLA based products for packaging (adopted from [316])

5. Future challenges and opportunities

Despite the tremendous progress made in the field of biodegradable biopolymers, some areas of research still require further exploration for addressing challenges. Biopolymers are intriguing raw materials for promoting eco-friendliness, non-toxicity, and sustainability. There is a need to conduct focused research for improving the mechanical characteristics and biodegradation mechanisms of complex working environments. Additionally, biopolymers are beneficial for the ecological environment only if their degradation products are non-toxic. To

date, no study has been found in the literature that evaluates the harmful and toxic impact upon the biodegradation of biopolymers.

Furthermore, several weaknesses in some biopolymers including poor processability and solubility in water or common organic solvents have been observed. These areas need to be explored to encourage the utilization of biopolymer materials. Biodegradable packaging for agricultural products also needs more attention in terms of gas barrier properties and water resistance. In biomedical areas, tissue biomimicry still remains a key challenge. Thus, more development and focus on 3D printing of biopolymers-based tissues or organs are required. This is because of some challenges related to material printability, the exact geometry of organs, functionality, and safety (biosafety and environmental safety). **Figure 15** summarizes the challenges in terms of application requirements that need attention in the future.

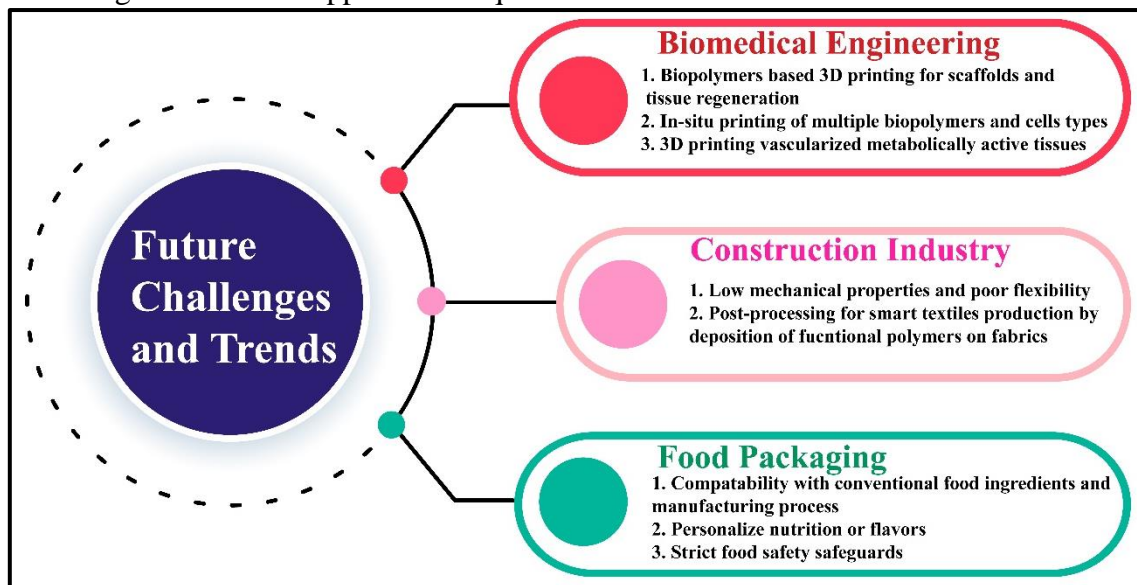


Figure 15. Future challenges and trends for biopolymers.

6. Conclusions

Recent advancements toward promoting a sustainable environment require the utilization of green materials, especially biopolymers and their composites, which possess the ability to replace synthetic polymers due to their extraordinary biodegradability, processability, renewability, biocompatibility, and non-toxicity. Besides these advantages, these biopolymers also have excellent technological and economic perspectives. The developed biopolymeric composites were characterized through spectroscopic, microscopic, diffraction, and degradation testing. These biopolymeric composites have exhibited their efficiency and integrity to be applied in textiles, implanting devices, tissue engineering, drug delivery systems, and agricultural industries. Non-biodegradable polymer composites are usually preferred over biodegradable biopolymer composites due to their superior mechanical characteristics. However, these plastics cannot be degraded and are severely influencing the ecological environment. Different biopolymers, including PLA, PHA, cellulose, and other naturally occurring biopolymers and their composites, demonstrated excellent degradation behavior, implying that these advantageous biopolymers will not only help to reduce the accumulation of plastic waste, but will also help to reduce the global warming effect.

The world needs to shift its attention toward the environmental pollution problem caused by plastic waste and its possible remedies. Reasonable and comparable mechanical characteristics of biodegradable biopolymer composites help their possible utilization in all engineering fields

to produce a cleaner and greener environment by cutting down on the accumulation of plastic waste.

Conflict of interest statement

The authors declare no conflict of interest.

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References

- [1] C. T. Ferreira, C. A. B. Perez, D. Hirayama, and C. Saron, "Recycling of polyamide (PA) from scrap tires as composites and blends," *J. Environ. Chem. Eng.*, vol. 1, no. 4, pp. 762–767, 2013, doi: <https://doi.org/10.1016/j.jece.2013.07.016>.
- [2] A. K. Urbanek, M. C. Strzelecki, and A. M. Mirończuk, "The potential of cold-adapted microorganisms for biodegradation of bioplastics," *Waste Manag.*, vol. 119, pp. 72–81, 2021, doi: <https://doi.org/10.1016/j.wasman.2020.09.031>.
- [3] D. N. Moholkar, P. S. Sadalage, D. Peixoto, A. C. Paiva-Santos, and K. D. Pawar, "Recent advances in biopolymer-based formulations for wound healing applications," *Eur. Polym. J.*, vol. 160, p. 110784, 2021, doi: <https://doi.org/10.1016/j.eurpolymj.2021.110784>.
- [4] M. Y. Khalid, A. Al Rashid, Z. U. Arif, W. Ahmed, H. Arshad, and A. A. Zaidi, "Natural fiber reinforced composites: Sustainable materials for emerging applications," *Results Eng.*, vol. 11, p. 100263, Sep. 2021, doi: [10.1016/J.RINENG.2021.100263](https://doi.org/10.1016/J.RINENG.2021.100263).
- [5] J. Jiang *et al.*, "From plastic waste to wealth using chemical recycling: A review," *J. Environ. Chem. Eng.*, vol. 10, no. 1, p. 106867, 2022, doi: <https://doi.org/10.1016/j.jece.2021.106867>.
- [6] D. Rahangdale and A. Kumar, "Acrylamide grafted chitosan based ion imprinted polymer for the recovery of cadmium from nickel-cadmium battery waste," *J. Environ. Chem. Eng.*, vol. 6, no. 2, pp. 1828–1839, 2018, doi: <https://doi.org/10.1016/j.jece.2018.02.027>.
- [7] V. V. Devadas *et al.*, "Algae biopolymer towards sustainable circular economy," *Bioresour. Technol.*, vol. 325, p. 124702, 2021, doi: <https://doi.org/10.1016/j.biortech.2021.124702>.
- [8] J. Payne, P. McKeown, and M. D. Jones, "A circular economy approach to plastic waste," *Polym. Degrad. Stab.*, vol. 165, pp. 170–181, 2019, doi: <https://doi.org/10.1016/j.polymdegradstab.2019.05.014>.
- [9] Y. G. Thyavihalli Girijappa, S. Mavinkere Rangappa, J. Parameswaranpillai, and S. Siengchin, "Natural Fibers as Sustainable and Renewable Resource for Development of Eco-Friendly Composites: A Comprehensive Review," *Front. Mater.*, vol. 6, p. 226, 2019, doi: [10.3389/fmats.2019.00226](https://doi.org/10.3389/fmats.2019.00226).
- [10] C. M. Chan *et al.*, "Composites of Wood and Biodegradable Thermoplastics: A Review," *Polym. Rev.*, vol. 58, no. 3, pp. 444–494, Jul. 2018, doi: [10.1080/15583724.2017.1380039](https://doi.org/10.1080/15583724.2017.1380039).
- [11] C. Jönsson *et al.*, "Biocatalysis in the Recycling Landscape for Synthetic Polymers and Plastics towards Circular Textiles," *ChemSusChem*, vol. 14, no. 19, pp. 4028–4040, Oct. 2021, doi: <https://doi.org/10.1002/cssc.202002666>.
- [12] S. S. Siwal *et al.*, "Key ingredients and recycling strategy of personal protective equipment (PPE): Towards sustainable solution for the COVID-19 like pandemics," *J. Environ. Chem. Eng.*, vol. 9, no. 5, p. 106284, 2021, doi: <https://doi.org/10.1016/j.jece.2021.106284>.
- [13] Z. U. Arif, M. Y. Khalid, W. Ahmed, H. Arshad, and S. Ullah, "Recycling of the glass/carbon fibre reinforced polymer composites: A step towards the circular economy," *Polym. Technol. Mater.*, pp. 1–28, Jan. 2022, doi: [10.1080/25740881.2021.2015781](https://doi.org/10.1080/25740881.2021.2015781).
- [14] M. Y. Khalid, Z. U. Arif, W. Ahmed, and H. Arshad, "Recent trends in recycling and reusing techniques of different plastic polymers and their composite materials," *Sustain. Mater. Technol.*, vol. 31, p. e00382, Apr. 2022, doi: [10.1016/J.SUSMAT.2021.E00382](https://doi.org/10.1016/J.SUSMAT.2021.E00382).
- [15] M. Y. Khalid, Z. U. Arif, and A. Al Rashid, "Investigation of Tensile and Flexural Behavior of Green Composites along with their Impact Response at Different Energies," *Int. J. of Precis. Eng. and Manuf.-Green Tech.*, 2021, doi: [10.1007/s40684-021-00385-w](https://doi.org/10.1007/s40684-021-00385-w).
- [16] A. Závodská, L. Benešová, B. Smyth, and A. J. Morrissey, "A comparison of biodegradable municipal waste (BMW) management strategies in Ireland and the Czech Republic and the lessons learned," *Resour. Conserv. Recycl.*, vol. 92, pp. 136–144, Nov. 2014, doi: [10.1016/J.RESCONREC.2014.09.007](https://doi.org/10.1016/J.RESCONREC.2014.09.007).
- [17] V. Sommer and G. Walther, "Recycling and recovery infrastructures for glass and carbon fiber reinforced plastic waste from wind energy industry: A European case study," *Waste Manag.*, vol. 121, pp. 265–275, Feb. 2021, doi: [10.1016/J.WASMAN.2020.12.021](https://doi.org/10.1016/J.WASMAN.2020.12.021).
- [18] M. Solis and S. Silveira, "Technologies for chemical recycling of household plastics – A technical review

- and TRL assessment,” *Waste Manag.*, vol. 105, pp. 128–138, 2020, doi: <https://doi.org/10.1016/j.wasman.2020.01.038>.
- [19] B. K. H. Lim and E. S. Thian, “Biodegradation of polymers in managing plastic waste — A review,” *Sci. Total Environ.*, vol. 813, p. 151880, 2022, doi: <https://doi.org/10.1016/j.scitotenv.2021.151880>.
- [20] I. Chiulan, E. B. Heggset, Ş. I. Voicu, and G. Chinga-Carrasco, “Photopolymerization of Bio-Based Polymers in a Biomedical Engineering Perspective,” *Biomacromolecules*, vol. 22, no. 5, pp. 1795–1814, May 2021, doi: 10.1021/acs.biomac.0c01745.
- [21] J. Li *et al.*, “Emerging Biopolymer-Based Bioadhesives,” *Macromol. Biosci.*, vol. 22, no. 2, p. 2100340, Feb. 2022, doi: <https://doi.org/10.1002/mabi.202100340>.
- [22] M. Y. Khalid *et al.*, “Developments in Chemical Treatments, Manufacturing Techniques and Potential Applications of Natural-Fibers-Based Biodegradable Composites,” *Coatings*, vol. 11, no. 3. 2021, doi: 10.3390/coatings11030293.
- [23] C. Li *et al.*, “Fiber-Based Biopolymer Processing as a Route toward Sustainability,” *Adv. Mater.*, vol. 34, no. 1, p. 2105196, Jan. 2022, doi: <https://doi.org/10.1002/adma.202105196>.
- [24] M. Y. Khalid, A. Al Rashid, Z. U. Arif, W. Ahmed, and H. Arshad, “Recent advances in nanocellulose-based different biomaterials: types, properties, and emerging applications,” *J. Mater. Res. Technol.*, vol. 14, pp. 2601–2623, Sep. 2021, doi: 10.1016/J.JMRT.2021.07.128.
- [25] B. Aaliya, K. V. Sunooj, and M. Lackner, “Biopolymer composites: a review,” *Int. J. Biobased Plast.*, vol. 3, no. 1, pp. 40–84, Jan. 2021, doi: 10.1080/24759651.2021.1881214.
- [26] Z. U. Arif, M. Y. Khalid, W. Ahmed, and H. Arshad, “A review on four-dimensional bioprinting in pursuit of advanced tissue engineering applications,” *Bioprinting*, p. e00203, Mar. 2022, doi: 10.1016/J.BPRINT.2022.E00203.
- [27] M. Y. Khalid, Z. U. Arif, and W. Ahmed, “Four-dimensional (4D) printing: technological and manufacturing renaissance,” *Macromol. Mater. Eng.*, vol. n/a, no. n/a, 2022, doi: <https://doi.org/10.1002/mame.202200003>.
- [28] G. E. Luckachan and C. K. S. Pillai, “Biodegradable Polymers- A Review on Recent Trends and Emerging Perspectives,” *J. Polym. Environ.*, vol. 19, no. 3, pp. 637–676, 2011, doi: 10.1007/s10924-011-0317-1.
- [29] K. W. Meereboer, M. Misra, and A. K. Mohanty, “Review of recent advances in the biodegradability of polyhydroxyalkanoate (PHA) bioplastics and their composites,” *Green Chem.*, vol. 22, no. 17, pp. 5519–5558, 2020.
- [30] Y. Zoungran, E. Lynda, K. K. Dobi-Brice, E. Tchirioua, C. Bakary, and D. D. Yannick, “Influence of natural factors on the biodegradation of simple and composite bioplastics based on cassava starch and corn starch,” *J. Environ. Chem. Eng.*, vol. 8, no. 5, p. 104396, Oct. 2020, doi: 10.1016/J.JECE.2020.104396.
- [31] V. Pagno *et al.*, “Heat treatment of polymeric PBAT/PCL membranes containing activated carbon from Brazil nutshell biomass obtained by electrospinning and applied in drug removal,” *J. Environ. Chem. Eng.*, vol. 8, no. 5, p. 104159, 2020, doi: <https://doi.org/10.1016/j.jece.2020.104159>.
- [32] A. P. Karmanov *et al.*, “Chemical structure and polymer properties of wheat and cabbage lignins – Valuable biopolymers for biomedical applications,” *Polymer (Guildf.)*, vol. 220, p. 123571, Apr. 2021, doi: 10.1016/J.POLYMER.2021.123571.
- [33] M. S. Meiabadi *et al.*, “Modeling the Producibility of 3D Printing in Polylactic Acid Using Artificial Neural Networks and Fused Filament Fabrication,” *Polymers*, vol. 13, no. 19. 2021, doi: 10.3390/polym13193219.
- [34] M. Askari, M. Afzali Naniz, M. Kouhi, A. Saberi, A. Zolfagharian, and M. Bodaghi, “Recent progress in extrusion 3D bioprinting of hydrogel biomaterials for tissue regeneration: a comprehensive review with focus on advanced fabrication techniques,” *Biomater. Sci.*, vol. 9, no. 3, pp. 535–573, 2021, doi: 10.1039/D0BM00973C.
- [35] A. Morales, J. Labidi, P. Gullón, and G. Astray, “Synthesis of advanced biobased green materials from renewable biopolymers,” *Curr. Opin. Green Sustain. Chem.*, vol. 29, p. 100436, Jun. 2021, doi: 10.1016/J.COGSC.2020.100436.
- [36] Y. P. R., M. Das, and S. K. Maiti, “Recent progress and challenges in cyanobacterial autotrophic production of polyhydroxybutyrate (PHB), a bioplastic,” *J. Environ. Chem. Eng.*, vol. 9, no. 4, p. 105379, 2021, doi: <https://doi.org/10.1016/j.jece.2021.105379>.
- [37] S. Ibrahim, G. A. M. Nawwar, and M. Sultan, “Development of bio-based polymeric hydrogel: Green, sustainable and low cost plant fertilizer packaging material,” *J. Environ. Chem. Eng.*, vol. 4, no. 1, pp. 203–210, 2016, doi: <https://doi.org/10.1016/j.jece.2015.10.028>.
- [38] P. R. Yaashikaa, P. Senthil Kumar, and S. Karishma, “Review on biopolymers and composites – Evolving material as adsorbents in removal of environmental pollutants,” *Environ. Res.*, vol. 212, p. 113114, 2022, doi: <https://doi.org/10.1016/j.envres.2022.113114>.
- [39] K. Wen, L. Wei, Z. Ren, B. Wang, and J. Lu, “Enhanced photocatalytic degradation of cationic and

- anionic dyes by Ag-modified g-C₃N₄ composite: Insights on different mechanisms under visible light,” *J. Mater. Res.*, vol. 36, no. 7, pp. 1549–1560, Apr. 2021, doi: 10.1557/S43578-021-00203-8.
- [40] J. Liu, L. Sun, W. Xu, Q. Wang, S. Yu, and J. Sun, “Current advances and future perspectives of 3D printing natural-derived biopolymers,” *Carbohydr. Polym.*, vol. 207, pp. 297–316, Mar. 2019, doi: 10.1016/J.CARBPOL.2018.11.077.
- [41] A. M. Díez-Pascual, “Biopolymer Composites: Synthesis, Properties, and Applications,” *International Journal of Molecular Sciences*, vol. 23, no. 4, MDPI, p. 2257, 2022.
- [42] M. Bilal and H. M. N. Iqbal, “Naturally-derived biopolymers: Potential platforms for enzyme immobilization,” *Int. J. Biol. Macromol.*, vol. 130, pp. 462–482, 2019, doi: <https://doi.org/10.1016/j.ijbiomac.2019.02.152>.
- [43] S. Mítura, A. Sionkowska, and A. Jaiswal, “Biopolymers for hydrogels in cosmetics: review,” *J. Mater. Sci. Mater. Med.*, vol. 31, no. 6, p. 50, 2020, doi: 10.1007/s10856-020-06390-w.
- [44] A. Kashirina, Y. Yao, Y. Liu, and J. Leng, “Biopolymers as bone substitutes: a review,” *Biomater. Sci.*, vol. 7, no. 10, pp. 3961–3983, 2019, doi: 10.1039/C9BM00664H.
- [45] G. Crini, “Historical review on chitin and chitosan biopolymers,” *Environ. Chem. Lett.*, vol. 17, no. 4, pp. 1623–1643, 2019, doi: 10.1007/s10311-019-00901-0.
- [46] T. Biswal, “Biopolymers for tissue engineering applications: A review,” *Mater. Today Proc.*, vol. 41, pp. 397–402, 2021, doi: <https://doi.org/10.1016/j.matpr.2020.09.628>.
- [47] S. Bose, C. Koski, and A. A. Vu, “Additive manufacturing of natural biopolymers and composites for bone tissue engineering,” *Mater. Horiz.*, vol. 7, no. 8, pp. 2011–2027, 2020, doi: 10.1039/D0MH00277A.
- [48] J. Liu *et al.*, “Biodegradation and up-cycling of polyurethanes: Progress, challenges, and prospects,” *Biotechnol. Adv.*, vol. 48, p. 107730, 2021, doi: <https://doi.org/10.1016/j.biotechadv.2021.107730>.
- [49] K. K. Sadasivuni, P. Saha, J. Adhikari, K. Deshmukh, M. B. Ahamed, and J. J. Cabibihan, “Recent advances in mechanical properties of biopolymer composites: a review,” *Polym. Compos.*, vol. 41, no. 1, pp. 32–59, 2020, doi: 10.1002/pc.25356.
- [50] V. Bátor, D. Åkesson, A. Zamani, M. J. Taherzadeh, and I. Sárvári Horváth, “Anaerobic degradation of bioplastics: A review,” *Waste Manag.*, vol. 80, pp. 406–413, Oct. 2018, doi: 10.1016/J.WASMAN.2018.09.040.
- [51] P. Kanmani, J. Aravind, M. Kamaraj, P. Sureshbabu, and S. Karthikeyan, “Environmental applications of chitosan and cellulosic biopolymers: A comprehensive outlook,” *Bioresour. Technol.*, vol. 242, pp. 295–303, Oct. 2017, doi: 10.1016/J.BIORTECH.2017.03.119.
- [52] A. George, M. R. Sanjay, R. Srisuk, J. Parameswaranpillai, and S. Siengchin, “A comprehensive review on chemical properties and applications of biopolymers and their composites,” *Int. J. Biol. Macromol.*, vol. 154, pp. 329–338, Jul. 2020, doi: 10.1016/J.IJBIOMAC.2020.03.120.
- [53] M. A. Oliveira *et al.*, “Performance evaluation of cashew gum and gelatin blend for food packaging,” *Food Packag. Shelf Life*, vol. 17, pp. 57–64, Sep. 2018, doi: 10.1016/J.FPSL.2018.05.003.
- [54] A. Kartik *et al.*, “A critical review on production of biopolymers from algae biomass and their applications,” *Bioresour. Technol.*, vol. 329, p. 124868, Jun. 2021, doi: 10.1016/J.BIORTECH.2021.124868.
- [55] E. Delamarche, A. Mattlet, S. Livi, J.-F. Gérard, R. Bayard, and V. Massardier, “Tailoring Biodegradability of Poly(Butylene Succinate)/Poly(Lactic Acid) Blends With a Deep Eutectic Solvent,” *Front. Mater.*, vol. 7, p. 7, 2020, doi: 10.3389/fmats.2020.00007.
- [56] S. Kazemi Najafi, “Use of recycled plastics in wood plastic composites – A review,” *Waste Manag.*, vol. 33, no. 9, pp. 1898–1905, Sep. 2013, doi: 10.1016/J.WASMAN.2013.05.017.
- [57] T. G. Kebede, S. Dube, and M. M. Nindi, “Biopolymer electrospun nanofibres for the adsorption of pharmaceuticals from water systems,” *J. Environ. Chem. Eng.*, vol. 7, no. 5, p. 103330, 2019, doi: <https://doi.org/10.1016/j.jece.2019.103330>.
- [58] L. S. Anthony, M. Vasudevan, V. Perumal, M. Ovinis, P. B. Raja, and T. N. J. I. Edison, “Bioresource-derived polymer composites for energy storage applications: Brief review,” *J. Environ. Chem. Eng.*, vol. 9, no. 5, p. 105832, 2021, doi: <https://doi.org/10.1016/j.jece.2021.105832>.
- [59] T. R. Abdou, A. B. Botelho Junior, D. C. R. Espinosa, and J. A. S. Tenório, “Recycling of polymeric composites from industrial waste by pyrolysis: Deep evaluation for carbon fibers reuse,” *Waste Manag.*, vol. 120, pp. 1–9, Feb. 2021, doi: 10.1016/J.WASMAN.2020.11.010.
- [60] K. Yu, Q. Shi, M. L. Dunn, T. Wang, and H. J. Qi, “Carbon Fiber Reinforced Thermoset Composite with Near 100% Recyclability,” *Adv. Funct. Mater.*, vol. 26, no. 33, pp. 6098–6106, Sep. 2016, doi: <https://doi.org/10.1002/adfm.201602056>.
- [61] M. Y. Khalid *et al.*, “Interlaminar Shear Strength (ILSS) Characterization of Fiber Metal Laminates (FMLs) manufactured through VARTM Process,” *Forces Mech.*, p. 100038, Jul. 2021, doi: 10.1016/J.FINMEC.2021.100038.
- [62] A. Mtibe, M. P. Motloug, J. Bandyopadhyay, and S. S. Ray, “Synthetic Biopolymers and Their

- Composites: Advantages and Limitations—An Overview,” *Macromol. Rapid Commun.*, vol. 42, no. 15, p. 2100130, Aug. 2021, doi: <https://doi.org/10.1002/marc.202100130>.
- [63] L. Liu, M. Xu, Y. Ye, and B. Zhang, “On the degradation of (micro)plastics: Degradation methods, influencing factors, environmental impacts,” *Sci. Total Environ.*, vol. 806, p. 151312, 2022, doi: <https://doi.org/10.1016/j.scitotenv.2021.151312>.
- [64] G. Ali, M. Sharma, E.-S. Salama, Z. Ling, and X. Li, “Applications of chitin and chitosan as natural biopolymer: potential sources, pretreatments, and degradation pathways,” *Biomass Convers. Biorefinery*, 2022, doi: 10.1007/s13399-022-02684-x.
- [65] K. Bano *et al.*, “Microbial enzymatic degradation of biodegradable plastics,” *Curr. Pharm. Biotechnol.*, vol. 18, no. 5, pp. 429–440, 2017.
- [66] S. Kakadellis, J. Woods, and Z. M. Harris, “Friend or foe: Stakeholder attitudes towards biodegradable plastic packaging in food waste anaerobic digestion,” *Resour. Conserv. Recycl.*, vol. 169, p. 105529, Jun. 2021, doi: 10.1016/J.RESCONREC.2021.105529.
- [67] Q. Xie *et al.*, “A new biodegradable sisal fiber–starch packing composite with nest structure,” *Carbohydr. Polym.*, vol. 189, pp. 56–64, Jun. 2018, doi: 10.1016/J.CARBPOL.2018.01.063.
- [68] S. S. Athukoralalage, R. Balu, N. K. Dutta, and N. Roy Choudhury, “3D Bioprinted Nanocellulose-Based Hydrogels for Tissue Engineering Applications: A Brief Review,” *Polymers*, vol. 11, no. 5, 2019, doi: 10.3390/polym11050898.
- [69] Y. Xu *et al.*, “Development and properties of bacterial cellulose, curcumin, and chitosan composite biodegradable films for active packaging materials,” *Carbohydr. Polym.*, vol. 260, p. 117778, May 2021, doi: 10.1016/J.CARBPOL.2021.117778.
- [70] J. Chen *et al.*, “Chitosan-based recyclable composite aerogels for the photocatalytic degradation of rhodamine B,” *Carbohydr. Polym.*, vol. 273, p. 118559, Dec. 2021, doi: 10.1016/J.CARBPOL.2021.118559.
- [71] D. Jubinville, E. Esmizadeh, C. Tzoganakis, and T. Mekonnen, “Thermo-mechanical recycling of polypropylene for the facile and scalable fabrication of highly loaded wood plastic composites,” *Compos. Part B Eng.*, vol. 219, p. 108873, Aug. 2021, doi: 10.1016/J.COMPOSITESB.2021.108873.
- [72] R. N. Oosterbeek, X. C. Zhang, S. M. Best, and R. E. Cameron, “A technique for improving dispersion within polymer–glass composites using polymer precipitation,” *J. Mech. Behav. Biomed. Mater.*, vol. 123, p. 104767, Nov. 2021, doi: 10.1016/J.JMBBM.2021.104767.
- [73] J. Xie, L. Huang, R. Wang, S. Ye, and X. Song, “Novel visible light-responsive graphene oxide/Bi₂WO₆/starch composite membrane for efficient degradation of ethylene,” *Carbohydr. Polym.*, vol. 246, p. 116640, Oct. 2020, doi: 10.1016/J.CARBPOL.2020.116640.
- [74] E. P. C. G. Luz *et al.*, “In vitro degradability and bioactivity of oxidized bacterial cellulose-hydroxyapatite composites,” *Carbohydr. Polym.*, vol. 237, p. 116174, Jun. 2020, doi: 10.1016/J.CARBPOL.2020.116174.
- [75] S. Miri, R. Saini, S. M. Davoodi, R. Pulicharla, S. K. Brar, and S. Magdouli, “Biodegradation of microplastics: Better late than never,” *Chemosphere*, vol. 286, p. 131670, 2022, doi: <https://doi.org/10.1016/j.chemosphere.2021.131670>.
- [76] N. K. Kalita, A. Sarmah, S. M. Bhasney, A. Kalamdhad, and V. Katiyar, “Demonstrating an ideal compostable plastic using biodegradability kinetics of poly(lactic acid) (PLA) based green biocomposite films under aerobic composting conditions,” *Environ. Challenges*, vol. 3, p. 100030, 2021, doi: <https://doi.org/10.1016/j.envc.2021.100030>.
- [77] M. R. Havstad, “Chapter 5 - Biodegradable plastics,” T. M. B. T.-P. W. and R. Letcher, Ed. Academic Press, 2020, pp. 97–129.
- [78] B. P. Frank *et al.*, “Biodegradation of Functionalized Nanocellulose,” *Environ. Sci. Technol.*, vol. 55, no. 15, pp. 10744–10757, Aug. 2021, doi: 10.1021/acs.est.0c07253.
- [79] O. García-Depraect *et al.*, “Biodegradation of bioplastics under aerobic and anaerobic aqueous conditions: Kinetics, carbon fate and particle size effect,” *Bioresour. Technol.*, vol. 344, p. 126265, 2022, doi: <https://doi.org/10.1016/j.biortech.2021.126265>.
- [80] D. P. Stephen and S. B. Palanisamy, “Advances in biopolymer composites and biomaterials for the removal of emerging contaminants,” *Phys. Sci. Rev.*, 2021, doi: doi:10.1515/psr-2021-0056.
- [81] R. Gao, H. Pan, L. Kai, K. Han, and J. Lian, “Microbial degradation and valorization of poly(ethylene terephthalate) (PET) monomers,” *World J. Microbiol. Biotechnol.*, vol. 38, no. 5, p. 89, 2022, doi: 10.1007/s11274-022-03270-z.
- [82] N. Morin-Crini, E. Lichtfouse, G. Torri, and G. Crini, “Applications of chitosan in food, pharmaceuticals, medicine, cosmetics, agriculture, textiles, pulp and paper, biotechnology, and environmental chemistry,” *Environ. Chem. Lett.*, vol. 17, no. 4, pp. 1667–1692, 2019, doi: 10.1007/s10311-019-00904-x.
- [83] M. J. Mochane, J. S. Sefadi, T. S. Motsoeneng, T. E. Mokoena, T. G. Mofokeng, and T. C. Mokhena, “The effect of filler localization on the properties of biopolymer blends, recent advances: A review,”

- Polym. Compos.*, vol. 41, no. 7, pp. 2958–2979, Jul. 2020, doi: <https://doi.org/10.1002/pc.25590>.
- [84] M. Horue, I. Rivero Berti, M. L. Cacicedo, and G. R. Castro, “Microbial production and recovery of hybrid biopolymers from wastes for industrial applications- a review,” *Bioresour. Technol.*, vol. 340, p. 125671, Nov. 2021, doi: 10.1016/J.BIORTECH.2021.125671.
- [85] M. Daria, L. Krzysztof, and M. Jakub, “Characteristics of biodegradable textiles used in environmental engineering: A comprehensive review,” *J. Clean. Prod.*, vol. 268, p. 122129, Sep. 2020, doi: 10.1016/J.JCLEPRO.2020.122129.
- [86] A. Vinod, M. R. Sanjay, S. Suchart, and P. Jyotishkumar, “Renewable and sustainable biobased materials: An assessment on biofibers, biofilms, biopolymers and biocomposites,” *J. Clean. Prod.*, vol. 258, p. 120978, Jun. 2020, doi: 10.1016/J.JCLEPRO.2020.120978.
- [87] P. Heidarian *et al.*, “Dynamic plant-derived polysaccharide-based hydrogels,” *Carbohydr. Polym.*, vol. 231, p. 115743, 2020, doi: <https://doi.org/10.1016/j.carbpol.2019.115743>.
- [88] N. B. Jildeh and M. Matouq, “Nanotechnology in packing materials for food and drug stuff opportunities,” *J. Environ. Chem. Eng.*, vol. 8, no. 5, p. 104338, 2020, doi: <https://doi.org/10.1016/j.jece.2020.104338>.
- [89] N. S. Rejinold, G. Choi, and J.-H. Choy, “Chitosan hybrids for cosmeceutical applications in skin, hair and dental care: an update,” *Emergent Mater.*, vol. 4, no. 5, pp. 1125–1142, 2021, doi: 10.1007/s42247-021-00280-9.
- [90] S. S. Gupta, V. Mishra, M. Das Mukherjee, P. Saini, and K. R. Ranjan, “Amino acid derived biopolymers: Recent advances and biomedical applications,” *Int. J. Biol. Macromol.*, vol. 188, pp. 542–567, 2021, doi: <https://doi.org/10.1016/j.ijbiomac.2021.08.036>.
- [91] R. D. Silva, L. T. de Carvalho, R. M. de Moraes, S. de F. Medeiros, and T. M. Lacerda, “Biomimetic Biomaterials Based on Polysaccharides: Recent Progress and Future Perspectives,” *Macromol. Chem. Phys.*, vol. n/a, no. n/a, p. 2100501, Mar. 2022, doi: <https://doi.org/10.1002/macp.202100501>.
- [92] S. Thomas, S. Gopi, and A. Amalraj, *Biopolymers and Their Industrial Applications: From Plant, Animal, and Marine Sources, to Functional Products*. Elsevier, 2020.
- [93] S. Takeshita, A. Sadeghpour, D. Sivaraman, S. Zhao, and W. J. Malfait, “Solvents, CO₂ and biopolymers: Structure formation in chitosan aerogel,” *Carbohydr. Polym.*, vol. 247, p. 116680, Nov. 2020, doi: 10.1016/J.CARBPOL.2020.116680.
- [94] Z. Barati, S. Latif, and J. Müller, “Biocatalysis and Agricultural Biotechnology Enzymatic hydrolysis of cassava peels as potential pre-treatment for peeling of cassava tubers,” *Biocatal. Agric. Biotechnol.*, vol. 20, no. July, p. 101247, 2019, doi: 10.1016/j.bcab.2019.101247.
- [95] M. Niaounakis, “Recycling of biopolymers – The patent perspective,” *Eur. Polym. J.*, vol. 114, pp. 464–475, 2019, doi: <https://doi.org/10.1016/j.eurpolymj.2019.02.027>.
- [96] I. Alves Lopes, L. Coelho Paixão, L. J. Souza da Silva, A. Almeida Rocha, A. K. Allan, and A. Amorim Santana, “Elaboration and characterization of biopolymer films with alginate and babassu coconut mesocarp,” *Carbohydr. Polym.*, vol. 234, p. 115747, Apr. 2020, doi: 10.1016/J.CARBPOL.2019.115747.
- [97] A. Sarkar *et al.*, “Preparation of novel biodegradable starch/poly(vinyl alcohol)/bentonite grafted polymeric films for fertilizer encapsulation,” *Carbohydr. Polym.*, vol. 259, p. 117679, May 2021, doi: 10.1016/J.CARBPOL.2021.117679.
- [98] H. Zhou *et al.*, “Preparation of bio-based cellulose acetate/chitosan composite film with oxygen and water resistant properties,” *Carbohydr. Polym.*, vol. 270, p. 118381, Oct. 2021, doi: 10.1016/J.CARBPOL.2021.118381.
- [99] E. M. N. Polman, G.-J. M. Gruter, J. R. Parsons, and A. Tietema, “Comparison of the aerobic biodegradation of biopolymers and the corresponding bioplastics: A review,” *Sci. Total Environ.*, vol. 753, p. 141953, 2021, doi: <https://doi.org/10.1016/j.scitotenv.2020.141953>.
- [100] M. Karimi-Avargani, F. Bazooyar, D. Biria, A. Zamani, and M. Skrifvars, “The special effect of the *Aspergillus flavus* and its enzymes on biological degradation of the intact polylactic acid (PLA) and PLA-Jute composite,” *Polym. Degrad. Stab.*, vol. 179, p. 109295, Sep. 2020, doi: 10.1016/J.POLYMDEGRADSTAB.2020.109295.
- [101] E. Chiellini, A. Corti, S. D’Antone, and R. Solaro, “Biodegradation of poly (vinyl alcohol) based materials,” *Prog. Polym. Sci.*, vol. 28, no. 6, pp. 963–1014, Jun. 2003, doi: 10.1016/S0079-6700(02)00149-1.
- [102] P. Rastogi and B. Kandasubramanian, “Breakthrough in the printing tactics for stimuli-responsive materials: 4D printing,” *Chem. Eng. J.*, vol. 366, pp. 264–304, 2019, doi: <https://doi.org/10.1016/j.cej.2019.02.085>.
- [103] J. Tarique, S. M. Sapuan, A. Khalina, S. F. K. Sherwani, J. Yusuf, and R. A. Ilyas, “Recent developments in sustainable arrowroot (*Maranta arundinacea* Linn) starch biopolymers, fibres, biopolymer composites and their potential industrial applications: A review,” *J. Mater. Res. Technol.*, vol. 13, pp. 1191–1219, Jul. 2021, doi: 10.1016/J.JMRT.2021.05.047.
- [104] I. Taniguchi, S. Yoshida, K. Hiraga, K. Miyamoto, Y. Kimura, and K. Oda, “Biodegradation of PET:

- Current Status and Application Aspects,” *ACS Catal.*, vol. 9, no. 5, pp. 4089–4105, May 2019, doi: 10.1021/acscatal.8b05171.
- [105] D. Chmolowska, N. Hamda, and R. Laskowski, “Cellulose decomposed faster in fallow soil than in meadow soil due to a shorter lag time,” *J. Soils Sediments*, vol. 17, no. 2, pp. 299–305, 2017, doi: 10.1007/s11368-016-1536-9.
- [106] P. Heidarian, A. Kaynak, M. Paulino, A. Zolfagharian, R. J. Varley, and A. Z. Kouzani, “Dynamic nanocellulose hydrogels: Recent advancements and future outlook,” *Carbohydr. Polym.*, vol. 270, p. 118357, 2021, doi: <https://doi.org/10.1016/j.carbpol.2021.118357>.
- [107] R. Grujić, D. Vujadinović, and D. Savanović, “Biopolymers as food packaging materials,” *Adv. Appl. Ind. Biomater.*, pp. 139–160, 2017.
- [108] M. Murariu, A.-L. Dechief, R. Ramy-Ratiarison, Y. Paint, J.-M. Raquez, and P. Dubois, “Recent advances in production of poly(lactic acid) (PLA) nanocomposites: a versatile method to tune crystallization properties of PLA,” 2015, doi: 10.1179/2055033214Y.0000000008.
- [109] L. Aliotta, P. Cinelli, M. B. Coltelli, M. C. Righetti, M. Gazzano, and A. Lazzeri, “Effect of nucleating agents on crystallinity and properties of poly (lactic acid) (PLA),” *Eur. Polym. J.*, vol. 93, pp. 822–832, Aug. 2017, doi: 10.1016/J.EURPOLYJM.2017.04.041.
- [110] A. K. Urbanek, W. Rymowicz, M. C. Strzelecki, W. Kociuba, Ł. Franczak, and A. M. Mirończuk, “Isolation and characterization of Arctic microorganisms decomposing bioplastics,” *AMB Express*, vol. 7, p. 148, 2017, doi: 10.1186/s13568-017-0448-4.
- [111] P. Cinelli, M. Seggiani, N. Mallegni, V. Gigante, and A. Lazzeri, “Processability and Degradability of PHA-Based Composites in Terrestrial Environments,” *International Journal of Molecular Sciences*, vol. 20, no. 2, 2019, doi: 10.3390/ijms20020284.
- [112] Y. X. Weng, L. Wang, M. Zhang, X. L. Wang, and Y. Z. Wang, “Biodegradation behavior of P(3HB,4HB)/PLA blends in real soil environments,” *Polym. Test.*, vol. 32, no. 1, pp. 60–70, Feb. 2013, doi: 10.1016/J.POLYMERTESTING.2012.09.014.
- [113] A. N. Boyandin *et al.*, “Microbial degradation of polyhydroxyalkanoates in tropical soils,” *Int. Biodeterior. Biodegradation*, vol. 83, pp. 77–84, Sep. 2013, doi: 10.1016/J.IBIOD.2013.04.014.
- [114] C.-S. Wu, “Preparation and Characterization of Polyhydroxyalkanoate Bioplastic-Based Green Renewable Composites from Rice Husk,” *J. Polym. Environ.*, vol. 22, no. 3, pp. 384–392, 2014, doi: 10.1007/s10924-014-0662-y.
- [115] P. Mousavioun, G. A. George, and W. O. S. Doherty, “Environmental degradation of lignin/poly(hydroxybutyrate) blends,” *Polym. Degrad. Stab.*, vol. 97, no. 7, pp. 1114–1122, Jul. 2012, doi: 10.1016/J.POLYMDEGRADSTAB.2012.04.004.
- [116] M. C. S. Corrêa, M. L. Rezende, D. S. Rosa, J. A. M. Agnelli, and P. A. P. Nascente, “Surface composition and morphology of poly(3-hydroxybutyrate) exposed to biodegradation,” *Polym. Test.*, vol. 27, no. 4, pp. 447–452, Jun. 2008, doi: 10.1016/J.POLYMERTESTING.2008.01.007.
- [117] S. M. Emadian, T. T. Onay, and B. Demirel, “Biodegradation of bioplastics in natural environments,” *Waste Manag.*, vol. 59, pp. 526–536, Jan. 2017, doi: 10.1016/J.WASMAN.2016.10.006.
- [118] N. Altaee, G. A. El-Hiti, A. Fahdil, K. Sudesh, and E. Yousif, “Biodegradation of different formulations of polyhydroxybutyrate films in soil,” *Springerplus*, vol. 5, no. 1, p. 762, 2016, doi: 10.1186/s40064-016-2480-2.
- [119] P. K. Samantaray *et al.*, “Poly (glycolic acid)(PGA): a versatile building block expanding high performance and sustainable bioplastic applications,” *Green Chem.*, vol. 22, no. 13, pp. 4055–4081, 2020.
- [120] N. Jain, V. K. Singh, and S. Chauhan, “A review on mechanical and water absorption properties of polyvinyl alcohol based composites/films,” *J. Mech. Behav. Mater.*, vol. 26, no. 5–6, pp. 213–222, 2017, doi: doi:10.1515/jmbm-2017-0027.
- [121] M. R. Pfau, K. G. McKinzey, A. A. Roth, and M. A. Grunlan, “PCL-Based Shape Memory Polymer Semi-IPNs: The Role of Miscibility in Tuning the Degradation Rate,” *Biomacromolecules*, vol. 21, no. 6, pp. 2493–2501, Jun. 2020, doi: 10.1021/acs.biomac.0c00454.
- [122] A. C. Vieira, J. C. Vieira, J. M. Ferra, F. D. Magalhães, R. M. Guedes, and A. T. Marques, “Mechanical study of PLA–PCL fibers during in vitro degradation,” *J. Mech. Behav. Biomed. Mater.*, vol. 4, no. 3, pp. 451–460, Apr. 2011, doi: 10.1016/J.JMBBM.2010.12.006.
- [123] M. Prambauer, C. Wendeler, J. Weitzenböck, and C. Burgstaller, “Biodegradable geotextiles – An overview of existing and potential materials,” *Geotext. Geomembranes*, vol. 47, no. 1, pp. 48–59, Feb. 2019, doi: 10.1016/J.GEOTEXMEM.2018.09.006.
- [124] N. K. Kalita, N. A. Damare, D. Hazarika, P. Bhagabati, A. Kalamdhad, and V. Katiyar, “Biodegradation and characterization study of compostable PLA bioplastic containing algae biomass as potential degradation accelerator,” *Environ. Challenges*, vol. 3, p. 100067, 2021, doi: <https://doi.org/10.1016/j.envc.2021.100067>.
- [125] Y. Zare, S. S. Rhim, and K. Y. Rhee, “The interphase degradation in a nanobiosensor including

- biopolymers and carbon nanotubes,” *Sensors Actuators A Phys.*, vol. 331, p. 112967, 2021, doi: <https://doi.org/10.1016/j.sna.2021.112967>.
- [126] A. Al Rashid, S. A. Khan, S. G. Al-Ghamdi, and M. Koç, “Additive manufacturing of polymer nanocomposites: Needs and challenges in materials, processes, and applications,” *Journal of Materials Research and Technology*, vol. 14. Elsevier Editora Ltda, pp. 910–941, Sep. 01, 2021, doi: [10.1016/j.jmrt.2021.07.016](https://doi.org/10.1016/j.jmrt.2021.07.016).
- [127] A. Al Rashid, W. Ahmed, M. Y. Khalid, and M. Koç, “Vat photopolymerization of polymers and polymer composites: Processes and applications,” *Addit. Manuf.*, vol. 47, p. 102279, Nov. 2021, doi: [10.1016/J.ADDMA.2021.102279](https://doi.org/10.1016/J.ADDMA.2021.102279).
- [128] B. Nasri-Nasrabadi *et al.*, “An electroactive polymer composite with reinforced bending strength, based on tubular micro carbonized-cellulose,” *Chem. Eng. J.*, vol. 334, pp. 1775–1780, 2018, doi: <https://doi.org/10.1016/j.cej.2017.11.140>.
- [129] J. Y. Cho *et al.*, “Polyhydroxyalkanoates (PHAs) degradation by the newly isolated marine *Bacillus* sp. JY14,” *Chemosphere*, vol. 283, p. 131172, 2021, doi: <https://doi.org/10.1016/j.chemosphere.2021.131172>.
- [130] F. Ruggero, E. Carretti, R. Gori, T. Lotti, and C. Lubello, “Monitoring of degradation of starch-based biopolymer film under different composting conditions, using TGA, FTIR and SEM analysis,” *Chemosphere*, vol. 246, p. 125770, 2020, doi: <https://doi.org/10.1016/j.chemosphere.2019.125770>.
- [131] S.-J. Xiong *et al.*, “Economically Competitive Biodegradable PBAT/Lignin Composites: Effect of Lignin Methylation and Compatibilizer,” *ACS Sustain. Chem. Eng.*, vol. 8, no. 13, pp. 5338–5346, Apr. 2020, doi: [10.1021/acssuschemeng.0c00789](https://doi.org/10.1021/acssuschemeng.0c00789).
- [132] H.-M. Wang *et al.*, “Tunable, UV-shielding and biodegradable composites based on well-characterized lignins and poly(butylene adipate-co-terephthalate),” *Green Chem.*, vol. 22, no. 24, pp. 8623–8632, 2020, doi: [10.1039/D0GC03284K](https://doi.org/10.1039/D0GC03284K).
- [133] V. Shanmugam *et al.*, “Circular economy in biocomposite development: State-of-the-art, challenges and emerging trends,” *Compos. Part C Open Access*, vol. 5, p. 100138, 2021, doi: <https://doi.org/10.1016/j.jcomc.2021.100138>.
- [134] S. Kamarian, M. Bodaghi, A. Pourasghar, and S. Talebi, “Vibrational behavior of non-uniform piezoelectric sandwich beams made of CNT-reinforced polymer nanocomposite by considering the agglomeration effect of CNTs,” *Polym. Compos.*, vol. 38, no. S1, pp. E553–E562, Sep. 2017, doi: <https://doi.org/10.1002/pc.23933>.
- [135] V. Shanmugam *et al.*, “Potential natural polymer-based nanofibres for the development of facemasks in countering viral outbreaks,” *J. Appl. Polym. Sci.*, vol. 138, no. 27, p. 50658, Jul. 2021, doi: <https://doi.org/10.1002/app.50658>.
- [136] S. M. Rangappa and S. Siengchin, “Moving towards biofiber-based composites: Knowledge gaps and insights,” *Express Polym. Lett.*, vol. 16, no. 5, pp. 451–452, 2022.
- [137] A. Grozdanov, M. Avella, A. Buzarovska, G. Gentile, and M. E. Errico, “Reuse of natural fiber reinforced eco-composites in polymer mortars,” *Polym. Eng. Sci.*, vol. 50, no. 4, pp. 762–766, Apr. 2010, doi: <https://doi.org/10.1002/pen.21582>.
- [138] M. Y. Khalid, Z. U. Arif, W. Ahmed, and H. Arshad, “Evaluation of tensile properties of fiber metal laminates under different strain rates,” *Proc. Inst. Mech. Eng. Part E J. Process Mech. Eng.*, p. 09544089211053063, Oct. 2021, doi: [10.1177/09544089211053063](https://doi.org/10.1177/09544089211053063).
- [139] A. Le Duigou, I. Pillin, A. Bourmaud, P. Davies, and C. Baley, “Effect of recycling on mechanical behaviour of biocompostable flax/poly(l-lactide) composites,” *Compos. Part A Appl. Sci. Manuf.*, vol. 39, no. 9, pp. 1471–1478, 2008, doi: <https://doi.org/10.1016/j.compositesa.2008.05.008>.
- [140] S. Mitra, T. Mateti, S. Ramakrishna, and A. Laha, “A Review on Curcumin-Loaded Electrospun Nanofibers and their Application in Modern Medicine,” *JOM*, 2022, doi: [10.1007/s11837-022-05180-9](https://doi.org/10.1007/s11837-022-05180-9).
- [141] C. Gandhimathi *et al.*, “Fabrication and characterization of high flux poly(vinylidene fluoride) electrospun nanofibrous membrane using amphiphilic polyethylene-block-poly(ethylene glycol) copolymer,” *J. Appl. Polym. Sci.*, vol. 138, no. 17, p. 50296, May 2021, doi: <https://doi.org/10.1002/app.50296>.
- [142] S. Malik, S. Sundarrajan, T. Hussain, A. Nazir, F. Berto, and S. Ramakrishna, “Electrospun biomimetic polymer nanofibers as vascular grafts,” *Mater. Des. Process. Commun.*, vol. 3, no. 6, p. e203, Dec. 2021, doi: <https://doi.org/10.1002/mdp2.203>.
- [143] M. Rahmati *et al.*, “Electrospinning for tissue engineering applications,” *Prog. Mater. Sci.*, vol. 117, p. 100721, 2021, doi: <https://doi.org/10.1016/j.pmatsci.2020.100721>.
- [144] S.-M. Mousavi *et al.*, “Bioactive Agent-Loaded Electrospun Nanofiber Membranes for Accelerating Healing Process: A Review,” *Membranes*, vol. 11, no. 9, 2021, doi: [10.3390/membranes11090702](https://doi.org/10.3390/membranes11090702).
- [145] S. Valvez, P. N. B. Reis, L. Susmel, and F. Berto, “Fused Filament Fabrication-4D-Printed Shape Memory Polymers: A Review,” *Polymers*, vol. 13, no. 5, 2021, doi: [10.3390/polym13050701](https://doi.org/10.3390/polym13050701).
- [146] M. A. K., V. Singaravelu, P. Jean-Mathieu, and M. Manjusri, “Composites from renewable and

- sustainable resources: Challenges and innovations,” *Science (80-.)*, vol. 362, no. 6414, pp. 536–542, Nov. 2018, doi: 10.1126/science.aat9072.
- [147] J.-P. Zhang, C.-C. Zhang, and F.-S. Zhang, “A novel process for waste polyvinyl chloride recycling: Plant growth substrate development,” *J. Environ. Chem. Eng.*, vol. 9, no. 4, p. 105475, 2021, doi: <https://doi.org/10.1016/j.jece.2021.105475>.
- [148] M. Y. Khalid, A. Al Rashid, Z. U. Arif, M. F. Sheikh, H. Arshad, and M. A. Nasir, “Tensile strength evaluation of glass/jute fibers reinforced composites: An experimental and numerical approach,” *Results Eng.*, vol. 10, p. 100232, 2021, doi: <https://doi.org/10.1016/j.rineng.2021.100232>.
- [149] M. Y. Khalid, Z. U. Arif, M. F. Sheikh, and M. A. Nasir, “Mechanical characterization of glass and jute fiber-based hybrid composites fabricated through compression molding technique,” *Int. J. Mater. Form.*, 2021, doi: 10.1007/s12289-021-01624-w.
- [150] M. Y. Khalid, A. Al Rashid, Z. Abbas, N. Akram, Z. U. Arif, and F. P. G. Márquez, “Evaluation of tensile properties of glass/sisal and glass/jute fibers reinforced hybrid composites at different stacking sequences,” *Polym.*, vol. 45, no. 3, pp. 390–397, 2021.
- [151] R. A. Ilyas *et al.*, “Chapter 7 - Mechanical Testing of Sugar Palm Fiber Reinforced Sugar Palm Biopolymer Composites,” F. M. Al-Oqla and S. M. B. T.-A. P. Sapuan Properties, and Applications of Starch and Other Bio-Based Polymers, Eds. Elsevier, 2020, pp. 89–110.
- [152] A. Hazarika, M. Mandal, and T. K. Maji, “Dynamic mechanical analysis, biodegradability and thermal stability of wood polymer nanocomposites,” *Compos. Part B Eng.*, vol. 60, pp. 568–576, Apr. 2014, doi: 10.1016/J.COMPOSITESB.2013.12.046.
- [153] N. A. Rosli, W. H. Wan Ishak, and I. Ahmad, “Eco-friendly high-density polyethylene/amorphous cellulose composites: Environmental and functional value,” *J. Clean. Prod.*, vol. 290, Mar. 2021, doi: 10.1016/J.JCLEPRO.2021.125886.
- [154] S. Sharma, A. Majumdar, and B. S. Butola, “Tailoring the biodegradability of polylactic acid (PLA) based films and ramie- PLA green composites by using selective additives,” *Int. J. Biol. Macromol.*, vol. 181, pp. 1092–1103, Jun. 2021, doi: 10.1016/J.IJBIOMAC.2021.04.108.
- [155] M. F. Shockley and A. H. Muliana, “Modeling temporal and spatial changes during hydrolytic degradation and erosion in biodegradable polymers,” *Polym. Degrad. Stab.*, vol. 180, p. 109298, Oct. 2020, doi: 10.1016/J.POLYMDEGRADSTAB.2020.109298.
- [156] Y. Tachibana, T. Tsutsuba, K. Kageyama, and K. ichi Kasuya, “Biodegradability of poly(butylene n-alkylenedionate)s composed of long-methylene chains as alternative polymers to polyethylene,” *Polym. Degrad. Stab.*, vol. 190, p. 109650, Aug. 2021, doi: 10.1016/J.POLYMDEGRADSTAB.2021.109650.
- [157] M. Y. Khalid, A. Al Rashid, Z. U. Arif, N. Akram, H. Arshad, and F. P. García Márquez, “Characterization of Failure Strain In Fiber Reinforced Composites: Under On-Axis and Off-Axis Loading,” *Crystals*, vol. 11, no. 2, 2021, doi: 10.3390/cryst11020216.
- [158] O. Haske-Cornelius *et al.*, “Enzymatic Systems for Cellulose Acetate Degradation,” doi: 10.3390/catal7100287.
- [159] L. Liu, J. Yu, L. Cheng, and X. Yang, “Biodegradability of poly(butylene succinate) (PBS) composite reinforced with jute fibre,” *Polym. Degrad. Stab.*, vol. 94, no. 1, pp. 90–94, Jan. 2009, doi: 10.1016/J.POLYMDEGRADSTAB.2008.10.013.
- [160] C.-S. Wu, “Renewable resource-based composites of recycled natural fibers and maleated polylactide bioplastic: Characterization and biodegradability,” *Polym. Degrad. Stab.*, vol. 94, no. 7, pp. 1076–1084, 2009, doi: <https://doi.org/10.1016/j.polymdegradstab.2009.04.002>.
- [161] Q. Xie *et al.*, “A new biodegradable sisal fiber–starch packing composite with nest structure,” *Carbohydr. Polym.*, vol. 189, pp. 56–64, 2018, doi: <https://doi.org/10.1016/j.carbpol.2018.01.063>.
- [162] N. L. I. Zailuddin, A. F. Osman, and R. Rahman, “Morphology, mechanical properties, and biodegradability of all-cellulose composite films from oil palm empty fruit bunch,” *SPE Polym.*, vol. 1, no. 1, pp. 4–14, Nov. 2020, doi: <https://doi.org/10.1002/pls2.10008>.
- [163] S. Kumari, R. Kumar, B. Rai, and G. Kumar, “Morphology and Biodegradability Study of Natural Latex-Modified Polyester–Banana Fiber Composites,” *J. Nat. Fibers*, vol. 18, no. 5, pp. 763–771, May 2021, doi: 10.1080/15440478.2019.1652131.
- [164] E. Mastalygina, I. Varyan, N. Kolesnikova, M. I. Gonzalez, and A. Popov, “Effect of Natural Rubber in Polyethylene Composites on Morphology, Mechanical Properties and Biodegradability,” *Polymers*, vol. 12, no. 2, 2020, doi: 10.3390/polym12020437.
- [165] D. R, S. N, Zulnazri, and R. M, “Mechanical and Biodegradability Properties of Bio Composite from Sago Starch and Straw Filler,” *Eurasian J. Anal. Chem.*, vol. 13, no. 6, pp. 244–248, 2018, [Online]. Available: <http://www.eurasianjournals.com/Mechanical-and-Biodegradability-Properties-of-Bio-Composite-from-Sago-Starch-and,104481,0,2.html>.
- [166] O. Mrajji, M. E. L. Wazna, Z. Samouh, A. E. L. Bouari, O. Cherkaoui, and R. El Moznine, “The effect of nonwoven structure on thermomechanical properties of feather waste reinforced polyester composite,” *J.*

- Ind. Text.*, p. 1528083720947734, Aug. 2020, doi: 10.1177/1528083720947734.
- [167] F. M. Yusof, N. 'Aini Wahab, N. L. Abdul Rahman, A. Kalam, A. Jumahat, and C. F. Mat Taib, "Properties of treated bamboo fiber reinforced tapioca starch biodegradable composite," *Mater. Today Proc.*, vol. 16, pp. 2367–2373, Jan. 2019, doi: 10.1016/J.MATPR.2019.06.140.
- [168] W. Bin Chan, S. W. Phang, and L. W. Yoon, "Thermal and biodegradability study for garden waste cellulose fibre in thermoplastic starch composite," *AIP Conf. Proc.*, vol. 2233, no. 1, p. 40001, May 2020, doi: 10.1063/5.0001496.
- [169] M. H. C. Yeong and R. A. Kristanti, "Potential Use of Rise Husk as Filler in Poly Lactic Acid Bio-composite: Mechanical Properties, Morphology and Biodegradability," *Front. Water Environ.*, vol. 1, no. 1, pp. 28–34, 2021.
- [170] K. Mishra and S. Sinha, "Biodegradable green composite film developed from Moringa Oleifera (Sahajana) seed filler and PVA: Surface functionalization, characterization and barrier properties," *J. Thermoplast. Compos. Mater.*, p. 08927057211007550, Apr. 2021, doi: 10.1177/08927057211007550.
- [171] S. Fatrozi, L. Purwanti, S. K. Sari, M. N. Ariesta, T. Kusumaningsih, and S. D. Marliyana, "Starch-based biofoams reinforced with microcrystalline cellulose from banana stem: Hydrophobicity and Biodegradability," *IOP Conf. Ser. Mater. Sci. Eng.*, vol. 858, p. 12037, 2020, doi: 10.1088/1757-899x/858/1/012037.
- [172] R. A. Ilyas, S. M. Sapuan, M. R. Ishak, and E. S. Zainudin, "Sugar palm nanocrystalline cellulose reinforced sugar palm starch composite: Degradation and water-barrier properties," *IOP Conf. Ser. Mater. Sci. Eng.*, vol. 368, p. 12006, 2018, doi: 10.1088/1757-899x/368/1/012006.
- [173] N. D. L. N.E. Kochkina, "Structure and properties of biodegradable maize starch/chitosan composite films as affected by PVA additions," *Int. J. Biol. Macromol.*, vol. 157, pp. 377–384, Aug. 2020, doi: 10.1016/J.IJBIOMAC.2020.04.154.
- [174] S. Manatsittipan, K. Kuttiyawong, and S. Tiptipakorn, "Effects of Levan on the Biodegradability and Thermal Properties of Polybutylene Succinate," *Key Eng. Mater.*, vol. 737, pp. 294–298, 2017, doi: 10.4028/www.scientific.net/KEM.737.294.
- [175] A. Mukherjee, D. Datta, and G. Halder, "Synthesis and characterisation of rice-straw-based grafted polymer composite by free radical copolymerisation," *Indian Chem. Eng.*, vol. 61, no. 2, pp. 105–119, Apr. 2019, doi: 10.1080/00194506.2018.1490930.
- [176] T. Eom *et al.*, "Naturally Derived Melanin Nanoparticle Composites with High Electrical Conductivity and Biodegradability," *Part. Part. Syst. Charact.*, vol. 36, no. 10, p. 1900166, Oct. 2019, doi: <https://doi.org/10.1002/ppsc.201900166>.
- [177] P. G. Gan, S. T. Sam, M. F. Abdullah, M. F. Omar, and W. K. Tan, "Water resistance and biodegradation properties of conventionally-heated and microwave-cured cross-linked cellulose nanocrystal/chitosan composite films," *Polym. Degrad. Stab.*, vol. 188, p. 109563, Jun. 2021, doi: 10.1016/J.POLYMDEGRADSTAB.2021.109563.
- [178] A. Vedrtnam, S. Kumar, and S. Chaturvedi, "Experimental study on mechanical behavior, biodegradability, and resistance to natural weathering and ultraviolet radiation of wood-plastic composites," *Compos. Part B Eng.*, vol. 176, p. 107282, Nov. 2019, doi: 10.1016/J.COMPOSITESB.2019.107282.
- [179] M. R. N. Fazita, K. Jayaraman, D. Bhattacharyya, M. S. Hossain, M. K. M. Haafiz, and A. K. H.P.S., "Disposal Options of Bamboo Fabric-Reinforced Poly(Lactic) Acid Composites for Sustainable Packaging: Biodegradability and Recyclability," *Polymers*, vol. 7, no. 8, 2015, doi: 10.3390/polym7081465.
- [180] N. C. Jadhav and R. D. Kale, "Scrap leather valorization through composite fabrication using mustard oil resin and N-vinyl-2-pyrrolidone," *Iran. Polym. J.*, vol. 29, no. 9, pp. 771–784, 2020, doi: 10.1007/s13726-020-00838-0.
- [181] S. Sable, S. Ahuja, and H. Bhunia, "Preparation and Characterization of Oxo-degradable Polypropylene Composites Containing a Modified Pro-oxidant," *J. Polym. Environ.*, vol. 29, no. 3, pp. 721–733, 2021, doi: 10.1007/s10924-020-01910-9.
- [182] S. B. Kivade, A. Gunge, M. Nagamadhu, and S. Rajole, "Mechanical and dynamic mechanical behavior of acetylation-treated plain woven banana reinforced biodegradable composites," *Adv. Compos. Hybrid Mater.*, 2021, doi: 10.1007/s42114-021-00247-8.
- [183] R. D. Kale, N. C. Jadhav, and S. Pal, "Fabrication of green composites based on rice bran oil and anhydride cross-linkers," *Iran. Polym. J.*, vol. 28, no. 6, pp. 471–482, 2019, doi: 10.1007/s13726-019-00715-5.
- [184] P. Vimalanathan, N. Venkateshwaran, and V. Santhanam, "Mechanical, dynamic mechanical, and thermal analysis of Shorea robusta-dispersed polyester composite," *Int. J. Polym. Anal. Charact.*, vol. 21, no. 4, pp. 314–326, May 2016, doi: 10.1080/1023666X.2016.1155818.
- [185] M. Everaert, K. Broos, P. Nielsen, W. Van Dun, M. Boone, and M. Quaghebeur, "Carbonation is affecting biodegradability testing of fiber cement composites," *Sci. Total Environ.*, vol. 686, pp. 888–892, Oct.

- 2019, doi: 10.1016/J.SCITOTENV.2019.06.018.
- [186] N. S. Suharty, I. P. Almanar, Sudirman, K. Dihadjo, and N. Astasari, “Flammability, Biodegradability and Mechanical Properties of Bio-Composites Waste Polypropylene/Kenaf Fiber Containing Nano CaCO₃ with Diammonium Phosphate,” *Procedia Chem.*, vol. 4, pp. 282–287, Jan. 2012, doi: 10.1016/J.PROCHE.2012.06.039.
- [187] E. Novita, “Biodegradability Simulation of Coffee Wastewater Using Instant Coffee,” *Agric. Agric. Sci. Procedia*, vol. 9, pp. 217–229, Jan. 2016, doi: 10.1016/J.AASPRO.2016.02.138.
- [188] F. Bandini *et al.*, “Anaerobic digestion and aerobic composting of rigid biopolymers in bio-waste treatment: fate and effects on the final compost,” *Bioresour. Technol.*, vol. 351, p. 126934, 2022, doi: <https://doi.org/10.1016/j.biortech.2022.126934>.
- [189] R. Santhosh, D. Nath, and P. Sarkar, “Novel food packaging materials including plant-based byproducts: A review,” *Trends Food Sci. Technol.*, vol. 118, pp. 471–489, 2021, doi: <https://doi.org/10.1016/j.tifs.2021.10.013>.
- [190] A. L. Patrício Silva, “Future-proofing plastic waste management for a circular bioeconomy,” *Curr. Opin. Environ. Sci. Heal.*, vol. 22, p. 100263, 2021, doi: <https://doi.org/10.1016/j.coesh.2021.100263>.
- [191] Y. J. Sohn *et al.*, “Recent Advances in Sustainable Plastic Upcycling and Biopolymers,” *Biotechnol. J.*, vol. 15, no. 6, p. 1900489, Jun. 2020, doi: <https://doi.org/10.1002/biot.201900489>.
- [192] S. Ali, A. Paul Peter, K. W. Chew, H. S. H. Munawaroh, and P. L. Show, “Resource recovery from industrial effluents through the cultivation of microalgae: A review,” *Bioresour. Technol.*, vol. 337, p. 125461, 2021, doi: <https://doi.org/10.1016/j.biortech.2021.125461>.
- [193] A. A. Rostam-Alilou, H. Jafari, A. Zolfagharian, A. Serjouei, and M. Bodaghi, “Experimentally validated vibro-acoustic modeling of 3D bio-printed grafts for potential use in human tympanic membrane regeneration,” *Bioprinting*, vol. 25, p. e00186, 2022, doi: <https://doi.org/10.1016/j.bprint.2021.e00186>.
- [194] H. P. S. Abdul Khalil *et al.*, “Biodegradable Films for Fruits and Vegetables Packaging Application: Preparation and Properties,” *Food Eng. Rev.*, vol. 10, no. 3, pp. 139–153, 2018, doi: 10.1007/s12393-018-9180-3.
- [195] A. Kaynak and A. Zolfagharian, “Functional Polymers in Sensors and Actuators: Fabrication and Analysis,” *Polymers*, vol. 12, no. 7. 2020, doi: 10.3390/polym12071569.
- [196] G. P. Udayakumar *et al.*, “Biopolymers and composites: Properties, characterization and their applications in food, medical and pharmaceutical industries,” *J. Environ. Chem. Eng.*, vol. 9, no. 4, p. 105322, Aug. 2021, doi: 10.1016/J.JECE.2021.105322.
- [197] M. Arastouei, M. Khodaei, S. M. Atyabi, and M. Jafari Nodoushan, “The in-vitro biological properties of 3D printed poly lactic acid/akermanite composite porous scaffold for bone tissue engineering,” *Mater. Today Commun.*, vol. 27, p. 102176, Jun. 2021, doi: 10.1016/J.MTCOMM.2021.102176.
- [198] M. Shirzad, A. Zolfagharian, A. Matbouei, and M. Bodaghi, “Design, evaluation, and optimization of 3D printed truss scaffolds for bone tissue engineering,” *J. Mech. Behav. Biomed. Mater.*, vol. 120, p. 104594, 2021, doi: <https://doi.org/10.1016/j.jmbbm.2021.104594>.
- [199] S. Sivakanthan, S. Rajendran, A. Gamage, T. Madhujith, and S. Mani, “Antioxidant and antimicrobial applications of biopolymers: A review,” *Food Res. Int.*, vol. 136, p. 109327, Oct. 2020, doi: 10.1016/J.FOODRES.2020.109327.
- [200] S. Zhang *et al.*, “Biopolymers and nanostructured materials to develop pectinases-based immobilized nano-biocatalytic systems for biotechnological applications,” *Food Res. Int.*, vol. 140, p. 109979, Feb. 2021, doi: 10.1016/J.FOODRES.2020.109979.
- [201] W. Xu, X. Wang, N. Sandler, S. Willför, and C. Xu, “Three-Dimensional Printing of Wood-Derived Biopolymers: A Review Focused on Biomedical Applications,” *ACS Sustain. Chem. Eng.*, vol. 6, no. 5, pp. 5663–5680, May 2018, doi: 10.1021/acssuschemeng.7b03924.
- [202] N. Teramoto, “Biomacromolecules, Biobased and Biodegradable Polymers (2017–2019),” *Polymers*, vol. 12, no. 10. 2020, doi: 10.3390/polym12102386.
- [203] C. Feng, T. Lotti, R. Canziani, Y. Lin, C. Tagliabue, and F. Malpei, “Extracellular biopolymers recovered as raw biomaterials from waste granular sludge and potential applications: A critical review,” *Sci. Total Environ.*, vol. 753, p. 142051, Jan. 2021, doi: 10.1016/J.SCITOTENV.2020.142051.
- [204] M. Azizi-Lalabadi and S. M. Jafari, “Bio-nanocomposites of graphene with biopolymers; fabrication, properties, and applications,” *Adv. Colloid Interface Sci.*, vol. 292, p. 102416, Jun. 2021, doi: 10.1016/J.CIS.2021.102416.
- [205] P. Rekhi, M. Goswami, S. Ramakrishna, and M. Debnath, “Polyhydroxyalkanoates biopolymers toward decarbonizing economy and sustainable future,” *Crit. Rev. Biotechnol.*, pp. 1–25, Oct. 2021, doi: 10.1080/07388551.2021.1960265.
- [206] V. Destefano, S. Khan, and A. Tabada, “Applications of PLA in modern medicine,” *Eng. Regen.*, vol. 1, no. April, pp. 76–87, 2020, doi: 10.1016/j.engreg.2020.08.002.
- [207] G. P. Udayakumar *et al.*, “Ecofriendly biopolymers and composites: Preparation and their applications in

- water-treatment,” *Biotechnol. Adv.*, vol. 52, p. 107815, Nov. 2021, doi: 10.1016/J.BIOTECHADV.2021.107815.
- [208] M. E. S. Hassan, J. Bai, and D.-Q. Dou, “Biopolymers; Definition, Classification and Applications,” *Egypt. J. Chem.*, vol. 62, no. 9, pp. 1725–1737, 2019, doi: 10.21608/ejchem.2019.6967.1580.
- [209] S. RameshKumar, P. Shaiju, K. E. O’Connor, and R. B. P., “Bio-based and biodegradable polymers - State-of-the-art, challenges and emerging trends,” *Curr. Opin. Green Sustain. Chem.*, vol. 21, pp. 75–81, Feb. 2020, doi: 10.1016/J.COGSC.2019.12.005.
- [210] C. Andreeßen and A. Steinbüchel, “Recent developments in non-biodegradable biopolymers: Precursors, production processes, and future perspectives,” *Appl. Microbiol. Biotechnol.*, vol. 103, no. 1, pp. 143–157, 2019, doi: 10.1007/s00253-018-9483-6.
- [211] D. C. Aduba *et al.*, “Vat photopolymerization 3D printing of acid-cleavable PEG-methacrylate networks for biomaterial applications,” *Mater. Today Commun.*, vol. 19, no. October 2018, pp. 204–211, 2019, doi: 10.1016/j.mtcomm.2019.01.003.
- [212] A. I. Rezk, H. M. Mousa, J. Lee, C. H. Park, and C. S. Kim, “Composite PCL / HA / simvastatin electrospun nanofiber coating on biodegradable Mg alloy for orthopedic implant application,” *J. Coatings Technol. Res.*, vol. 16, no. 2, pp. 477–489, 2019, doi: 10.1007/s11998-018-0126-8.
- [213] B. D. Ulery, L. S. Nair, and C. T. Laurencin, “Biomedical Applications of Biodegradable Polymers,” pp. 832–864, 2011, doi: 10.1002/polb.22259.
- [214] J. S. Id, J. Shen, S. Chen, M. A. Cooper, D. Wu, and Z. Yang, “Nanofiller Reinforced Biodegradable PLA / PHA Composites : Current Status and Future Trends,” pp. 1–22, doi: 10.3390/polym10050505.
- [215] E. Malikmammadov, T. E. Tanir, A. Kiziltay, and V. Hasirci, “PCL and PCL-based materials in biomedical applications,” *J. Biomater. Sci. Polym. Ed.*, vol. 5063, pp. 1–31, 2018, doi: 10.1080/09205063.2017.1394711.
- [216] Y. S. Alshebly, M. Nafea, M. S. Mohamed Ali, and H. A. F. Almurib, “Review on recent advances in 4D printing of shape memory polymers,” *Eur. Polym. J.*, vol. 159, p. 110708, Oct. 2021, doi: 10.1016/J.EURPOLYMJ.2021.110708.
- [217] D. Puppi and F. Chiellini, “Biodegradable Polymers for Biomedical Additive Manufacturing,” *Appl. Mater. Today*, vol. 20, p. 100700, Sep. 2020, doi: 10.1016/J.APMT.2020.100700.
- [218] Y. Yang *et al.*, “Metal organic frameworks as a compatible reinforcement in a biopolymer bone scaffold,” *Mater. Chem. Front.*, vol. 4, no. 3, pp. 973–984, 2020, doi: 10.1039/C9QM00772E.
- [219] H. Yuan, L. Chen, and F. F. Hong, “A Biodegradable Antibacterial Nanocomposite Based on Oxidized Bacterial Nanocellulose for Rapid Hemostasis and Wound Healing,” *ACS Appl. Mater. Interfaces*, vol. 12, no. 3, pp. 3382–3392, Jan. 2020, doi: 10.1021/acsami.9b17732.
- [220] B. Mishra, S. Hossain, S. Mohanty, M. K. Gupta, and D. Verma, “Fast acting hemostatic agent based on self-assembled hybrid nanofibers from chitosan and casein,” *Int. J. Biol. Macromol.*, vol. 185, pp. 525–534, Aug. 2021, doi: 10.1016/J.IJBIOMAC.2021.06.116.
- [221] S. Yalcin and U. Gündüz, “Synthesis and biological activity of siRNA and Etoposide with magnetic nanoparticles on drug resistance model MCF-7 Cells: Molecular docking study with MRP1 enzyme,” *Nanomedicine J.*, vol. 8, no. 2, pp. 98–105, 2021.
- [222] N. Malik, S. Shrivastava, and S. B. Ghosh, “Moisture Absorption Behaviour of Biopolymer Polycaprolactone (PCL) / Organo Modified Montmorillonite Clay (OMMT) biocomposite films,” *IOP Conf. Ser. Mater. Sci. Eng.*, vol. 346, p. 12027, 2018, doi: 10.1088/1757-899x/346/1/012027.
- [223] N. Mücke *et al.*, “Use of Water-Soluble Curcumin in TPS/PBAT Packaging Material: Interference on Reactive Extrusion and Oxidative Stability of Chia Oil,” *Food Bioprocess Technol.*, vol. 14, no. 3, pp. 471–482, 2021, doi: 10.1007/s11947-021-02584-4.
- [224] K. Kundu, A. Afshar, D. R. Katti, M. Edirisinghe, and K. S. Katti, “Composite nanoclay-hydroxyapatite-polymer fiber scaffolds for bone tissue engineering manufactured using pressurized gyration,” *Compos. Sci. Technol.*, vol. 202, p. 108598, Jan. 2021, doi: 10.1016/J.COMPSCITECH.2020.108598.
- [225] E. Díaz, J. Aresti, and J. León, “Evaluation of physicochemical and mechanical properties with the in vitro degradation of PCL/nHA/MWCNT composite scaffolds,” *J. Reinf. Plast. Compos.*, vol. 40, no. 3–4, pp. 134–142, Jul. 2020, doi: 10.1177/0731684420943304.
- [226] I. J. Villamagna *et al.*, “GSK3787-Loaded Poly(Ester Amide) Particles for Intra-Articular Drug Delivery,” *Polymers*, vol. 12, no. 4. 2020, doi: 10.3390/polym12040736.
- [227] A. Das, J. Adhikari, and P. Saha, “Electrospun polymeric nanocarbon nanomats for tissue engineering,” *Nanocarbon its Compos. Prep. Prop. Appl.*, pp. 91–122, Jan. 2019, doi: 10.1016/B978-0-08-102509-3.00004-3.
- [228] S. Mondal *et al.*, “Hydroxyapatite nano bioceramics optimized 3D printed poly lactic acid scaffold for bone tissue engineering application,” *Ceram. Int.*, vol. 46, no. 3, pp. 3443–3455, Feb. 2020, doi: 10.1016/J.CERAMINT.2019.10.057.
- [229] B. N. Singh *et al.*, “Design and evaluation of chitosan/chondroitin sulfate/nano-bioglass based composite

- scaffold for bone tissue engineering,” *Int. J. Biol. Macromol.*, vol. 133, pp. 817–830, Jul. 2019, doi: 10.1016/J.IJBIOMAC.2019.04.107.
- [230] I. Lukic, J. Vulic, and J. Ivanovic, “Antioxidant activity of PLA/PCL films loaded with thymol and/or carvacrol using scCO₂ for active food packaging,” *Food Packag. Shelf Life*, vol. 26, p. 100578, Dec. 2020, doi: 10.1016/J.FPSL.2020.100578.
- [231] C. Mouro, A. P. Gomes, M. Ahonen, R. Fangueiro, and I. C. Gouveia, “Chelidoniummajus L. Incorporated Emulsion Electrospun PCL/PVA_PEC Nanofibrous Meshes for Antibacterial Wound Dressing Applications,” *Nanomaterials*, vol. 11, no. 7. 2021, doi: 10.3390/nano11071785.
- [232] K. Chugh, G. Phalak, and S. Mhaske, “Fatty acid based novel precursors for polyesteramide hot melt adhesive,” *J. Adhes. Sci. Technol.*, vol. 34, no. 17, pp. 1871–1884, Sep. 2020, doi: 10.1080/01694243.2020.1732763.
- [233] H. C. Tseng, N. Fujimoto, and A. Ohnishi, “Biodegradability and methane fermentability of polylactic acid by thermophilic methane fermentation,” *Bioresour. Technol. Reports*, vol. 8, p. 100327, Dec. 2019, doi: 10.1016/J.BITEB.2019.100327.
- [234] J. Zhu *et al.*, “Effect of Na₂CO₃ on the Microstructure and Macroscopic Properties and Mechanism Analysis of PVA/CMC Composite Film,” *Polymers*, vol. 12, no. 2. 2020, doi: 10.3390/polym12020453.
- [235] H. Ge, Y. Wu, L. L. Woshnak, and S. H. Mitmesser, “Effects of hydrocolloids, acids and nutrients on gelatin network in gummies,” *Food Hydrocoll.*, vol. 113, p. 106549, Apr. 2021, doi: 10.1016/J.FOODHYD.2020.106549.
- [236] S. Mirzaeei, M. Mansurian, K. Asare-Addo, and A. Nokhodchi, “Metronidazole- and Amoxicillin-Loaded PLGA and PCL Nanofibers as Potential Drug Delivery Systems for the Treatment of Periodontitis: In Vitro and In Vivo Evaluations,” *Biomedicines*, vol. 9, no. 8. 2021, doi: 10.3390/biomedicines9080975.
- [237] K. Dietrich, M.-J. Dumont, V. Orsat, and L. F. Del Rio, “Consumption of sugars and inhibitors of softwood hemicellulose hydrolysates as carbon sources for polyhydroxybutyrate (PHB) production with *Paraburkholderia sacchari* IPT 101,” *Cellulose*, vol. 26, no. 13, pp. 7939–7952, 2019, doi: 10.1007/s10570-019-02664-x.
- [238] C. Zhao, Y. Li, G. Peng, X. Lei, G. Zhang, and Y. Gao, “Decellularized liver matrix-modified chitosan fibrous scaffold as a substrate for C3A hepatocyte culture,” *J. Biomater. Sci. Polym. Ed.*, vol. 31, no. 8, pp. 1041–1056, May 2020, doi: 10.1080/09205063.2020.1738690.
- [239] T. Zhao *et al.*, “Integration of mesenchymal stem cell sheet and bFGF-loaded fibrin gel in knitted PLGA scaffolds favorable for tendon repair,” *J. Mater. Chem. B*, vol. 7, no. 13, pp. 2201–2211, 2019, doi: 10.1039/C8TB02759E.
- [240] S. Buyuksungur, V. Hasirci, and N. Hasirci, “3D printed hybrid bone constructs of PCL and dental pulp stem cells loaded GelMA,” *J. Biomed. Mater. Res. Part A*, vol. 109, no. 12, pp. 2425–2437, Dec. 2021, doi: <https://doi.org/10.1002/jbm.a.37235>.
- [241] M. Parvizifard and S. Karbasi, “Physical, mechanical and biological performance of PHB-Chitosan/MWCNTs nanocomposite coating deposited on bioglass based scaffold: Potential application in bone tissue engineering,” *Int. J. Biol. Macromol.*, vol. 152, pp. 645–662, Jun. 2020, doi: 10.1016/J.IJBIOMAC.2020.02.266.
- [242] P. W. Ferlic, M. Nogler, A. M. Weinberg, K. D. Kühn, M. Liebensteiner, and D. C. Coraça-Huber, “Material modifications enhancing the antibacterial properties of two biodegradable poly(3-hydroxybutyrate) implants,” *Biomed. Mater.*, vol. 16, no. 1, p. 15030, 2020, doi: 10.1088/1748-605x/abbec6.
- [243] X. Deng, M. Gould, and M. A. Ali, “Fabrication and characterisation of melt-extruded chitosan/keratin/PCL/PEG drug-eluting sutures designed for wound healing,” *Mater. Sci. Eng. C*, vol. 120, p. 111696, Jan. 2021, doi: 10.1016/J.MSEC.2020.111696.
- [244] J. de Carvalho Arjona, M. das Graças Silva-Valenzuela, S.-H. Wang, and F. R. Valenzuela-Diaz, “Biodegradable Nanocomposite Microcapsules for Controlled Release of Urea,” *Polymers*, vol. 13, no. 5. 2021, doi: 10.3390/polym13050722.
- [245] B. Subramanian *et al.*, “Synthesis and characterization of PCL-DA:PEG-DA based polymeric blends grafted with SMA hydrogel as bio-degradable intrauterine contraceptive implant,” *Mater. Sci. Eng. C*, vol. 116, p. 111159, Nov. 2020, doi: 10.1016/J.MSEC.2020.111159.
- [246] D. Caballero, C. M. Abreu, A. C. Lima, N. N. Neves, R. L. Reis, and S. C. Kundu, “Precision biomaterials in cancer theranostics and modelling,” *Biomaterials*, vol. 280, p. 121299, Jan. 2022, doi: 10.1016/J.BIOMATERIALS.2021.121299.
- [247] J. Domínguez-Robles *et al.*, “Antioxidant PLA Composites Containing Lignin for 3D Printing Applications: A Potential Material for Healthcare Applications,” *Pharmaceutics*, vol. 11, no. 4. 2019, doi: 10.3390/pharmaceutics11040165.
- [248] C. R. Gough *et al.*, “Biopolymer-Based Filtration Materials,” *ACS Omega*, vol. 6, no. 18, pp. 11804–11812, May 2021, doi: 10.1021/acsomega.1c00791.

- [249] E. Mohamed, A. Fitzgerald, and T. Tsuzuki, "The role of nanoscale structures in the development of topical hemostatic agents," *Mater. Today Nano*, vol. 16, p. 100137, Dec. 2021, doi: 10.1016/J.MTANO.2021.100137.
- [250] G. Cavallaro, S. Milioto, L. Nigamatzyanova, F. Akhatova, R. Fakhrullin, and G. Lazzara, "Pickering Emulsion Gels Based on Halloysite Nanotubes and Ionic Biopolymers: Properties and Cleaning Action on Marble Surface," *ACS Appl. Nano Mater.*, vol. 2, no. 5, pp. 3169–3176, May 2019, doi: 10.1021/acsnm.9b00487.
- [251] T. C. Mokhena and J. S. Sefadi, "applied sciences Morphology and Properties of Electrospun PCL and Its Composites for Medical Applications : A Mini Review," pp. 1–17.
- [252] V. Bertana, F. Catania, M. Cocuzza, S. Ferrero, L. Scaltrito, and C. F. Pirri, "Medical and biomedical applications of 3D and 4D printed polymer nanocomposites," *3D 4D Print. Polym. Nanocomposite Mater. Process. Appl. Challenges*, pp. 325–366, Jan. 2019.
- [253] S. Bose, D. Ke, H. Sahasrabudhe, and A. Bandyopadhyay, "Additive manufacturing of biomaterials," *Prog. Mater. Sci.*, vol. 93, pp. 45–111, 2018, doi: <https://doi.org/10.1016/j.pmatsci.2017.08.003>.
- [254] S. K. Hedayati, A. H. Behraves, S. Hasannia, A. Bagheri Saed, and B. Akhondi, "3D printed PCL scaffold reinforced with continuous biodegradable fiber yarn: A study on mechanical and cell viability properties," *Polym. Test.*, vol. 83, p. 106347, Mar. 2020, doi: 10.1016/J.POLYMERTESTING.2020.106347.
- [255] A. Cano-Vicent *et al.*, "Fused deposition modelling: Current status, methodology, applications and future prospects," *Addit. Manuf.*, vol. 47, p. 102378, Nov. 2021, doi: 10.1016/J.ADDMA.2021.102378.
- [256] C. Lin, L. Liu, Y. Liu, and J. Leng, "4D printing of shape memory polybutylene succinate/polylactic acid (PBS/PLA) and its potential applications," *Compos. Struct.*, vol. 279, p. 114729, Jan. 2022.
- [257] F. Mokhtari, B. Azimi, M. Salehi, S. Hashemikia, and S. Danti, "Recent advances of polymer-based piezoelectric composites for biomedical applications," *J. Mech. Behav. Biomed. Mater.*, vol. 122, p. 104669, Oct. 2021, doi: 10.1016/J.JMBBM.2021.104669.
- [258] R. Velu, D. K. Jayashankar, and K. Subburaj, "Additive processing of biopolymers for medical applications," *Addit. Manuf.*, pp. 635–659, Jan. 2021, doi: 10.1016/B978-0-12-818411-0.00019-7.
- [259] S. Mohandesnezhad *et al.*, "In vitro evaluation of novel Zeolite-hydroxyapatite blended scaffold for dental tissue engineering." Research Square, 2020, doi: 10.21203/rs.2.20452/v1.
- [260] D. Veeman *et al.*, "Additive Manufacturing of Biopolymers for Tissue Engineering and Regenerative Medicine: An Overview, Potential Applications, Advancements, and Trends," *Int. J. Polym. Sci.*, vol. 2021, p. 4907027, 2021, doi: 10.1155/2021/4907027.
- [261] S. Malekmohammadi *et al.*, "Smart and Biomimetic 3D and 4D Printed Composite Hydrogels: Opportunities for Different Biomedical Applications," *Biomedicines*, vol. 9, no. 11. 2021, doi: 10.3390/biomedicines9111537.
- [262] A. A. Rostam-Alilou, H. Jafari, A. Zolfagharian, A. Serjouei, and M. Bodaghi, "Using fibrin/collagen composite hydrogel and silk for bio-inspired design of tympanic membrane grafts: A vibro-acoustic analysis," *Compos. Part C Open Access*, vol. 6, p. 100205, 2021, doi: <https://doi.org/10.1016/j.jcomc.2021.100205>.
- [263] S. Jinga, A. Zamfirescu, G. Voicu, M. Enculescu, A. Evangelidis, and C. Busuioc, "PCL-ZnO/TiO₂/HAp Electrospun Composite Fibers with Applications in Tissue Engineering," pp. 1–12, 2019.
- [264] J. In Kim and C. S. Kim, "Harnessing nanotopography of PCL/collagen nanocomposite membrane and changes in cell morphology coordinated with wound healing activity," *Mater. Sci. Eng. C*, vol. 91, pp. 824–837, 2018, doi: <https://doi.org/10.1016/j.msec.2018.06.021>.
- [265] B. T. Goh, L. Y. Teh, D. B. P. Tan, Z. Zhang, and S. H. Teoh, "Novel 3D polycaprolactone scaffold for ridge preservation – a pilot randomised controlled clinical trial," *Clin. Oral Implants Res.*, vol. 26, no. 3, pp. 271–277, Mar. 2015, doi: <https://doi.org/10.1111/clr.12486>.
- [266] A. Abdal-hay, N. T. Raveendran, B. Fournier, and S. Ivanovski, "Fabrication of biocompatible and bioabsorbable polycaprolactone/ magnesium hydroxide 3D printed scaffolds: Degradation and in vitro osteoblasts interactions," *Compos. Part B Eng.*, vol. 197, p. 108158, Sep. 2020, doi: 10.1016/J.COMPOSITESB.2020.108158.
- [267] J. He, W. Zhang, Y. Liu, X. Li, D. Li, and Z. Jin, "Design and fabrication of biomimetic multiphased scaffolds for ligament-to-bone fixation," *Mater. Sci. Eng. C*, vol. 50, pp. 12–18, 2015, doi: <https://doi.org/10.1016/j.msec.2015.01.088>.
- [268] R. Soni, N. V. Kumar, S. Chameettachal, and F. Pati, "ScienceDirect Synthesis and Optimization of PCL-Bioactive Glass Composite Scaffold for Bone Tissue Engineering," *Mater. Today Proc.*, vol. 15, pp. 294–299, 2019, doi: 10.1016/j.matpr.2019.05.008.
- [269] P. Makkar, H. J. Kang, A. R. Padalhin, I. Park, B. Moon, and B. T. Lee, "Development and properties of duplex MgF₂ / PCL coatings on biodegradable magnesium alloy for biomedical applications," pp. 1–21, 2018.

- [270] C. Shuai, W. Yang, P. Feng, S. Peng, and H. Pan, "Accelerated degradation of HAP/PLLA bone scaffold by PGA blending facilitates bioactivity and osteoconductivity," *Bioact. Mater.*, vol. 6, no. 2, pp. 490–502, 2021, doi: <https://doi.org/10.1016/j.bioactmat.2020.09.001>.
- [271] G. Ji *et al.*, "Biopolymer Immune Implants' Sequential Activation of Innate and Adaptive Immunity for Colorectal Cancer Postoperative Immunotherapy," *Adv. Mater.*, vol. 33, no. 3, p. 2004559, Jan. 2021, doi: <https://doi.org/10.1002/adma.202004559>.
- [272] N. G. S. Silva *et al.*, "A promising sustainable PHB-ZnO composite for development of biodegradable filaments," *Polym. Compos.*, vol. 43, no. 1, pp. 144–159, Jan. 2022, doi: <https://doi.org/10.1002/pc.26363>.
- [273] X. Wang *et al.*, "Preparation of 4D printed peripheral vascular stent and its degradation behavior under fluid shear stress after deployment," *Biomater. Sci.*, 2022, doi: 10.1039/D2BM00088A.
- [274] T. Hanas, T. S. S. Kumar, G. Perumal, M. Doble, and S. Ramakrishna, "Electrospun PCL/HA coated friction stir processed AZ31/HA composites for degradable implant applications," vol. 252, no. October 2017, pp. 398–406, 2018, doi: 10.1016/j.jmatprotec.2017.10.009.
- [275] A. Melocchi *et al.*, "Shape memory materials and 4D printing in pharmaceuticals," *Adv. Drug Deliv. Rev.*, vol. 173, pp. 216–237, Jun. 2021, doi: 10.1016/J.ADDR.2021.03.013.
- [276] A. Koyyada and P. Orsu, "Natural gum polysaccharides as efficient tissue engineering and drug delivery biopolymers," *J. Drug Deliv. Sci. Technol.*, vol. 63, p. 102431, Jun. 2021, doi: 10.1016/J.JDDST.2021.102431.
- [277] R. Anand and A. Kumar, "Significant biopolymers and their applications in buccal mediated drug delivery," *J. Biomater. Sci. Polym. Ed.*, vol. 32, no. 9, pp. 1203–1218, Jun. 2021, doi: 10.1080/09205063.2021.1902175.
- [278] M. C. Biswas *et al.*, "Recent Advancement of Biopolymers and Their Potential Biomedical Applications," *J. Polym. Environ.*, 2021, doi: 10.1007/s10924-021-02199-y.
- [279] K. Aoki and N. Saito, "Biodegradable Polymers as Drug Delivery Systems for Bone Regeneration," *Pharmaceutics*, vol. 12, no. 2. 2020, doi: 10.3390/pharmaceutics12020095.
- [280] S. Zu *et al.*, "4D printing of core-shell hydrogel capsules for smart controlled drug release," *Bio-Design Manuf.*, 2022, doi: 10.1007/s42242-021-00175-y.
- [281] E. Pei and G. H. Loh, "Technological considerations for 4D printing: an overview," *Prog. Addit. Manuf.*, vol. 3, no. 1, pp. 95–107, 2018, doi: 10.1007/s40964-018-0047-1.
- [282] B. Subeshan, Y. Baddam, and E. Asmatulu, "Current progress of 4D-printing technology," *Prog. Addit. Manuf.*, vol. 6, no. 3, pp. 495–516, 2021, doi: 10.1007/s40964-021-00182-6.
- [283] I. J. Macha *et al.*, "Journal of Drug Delivery Science and Technology Biocompatibility of a new biodegradable polymer-hydroxyapatite composite for biomedical applications," *J. Drug Deliv. Sci. Technol.*, vol. 38, pp. 72–77, 2017, doi: 10.1016/j.jddst.2017.01.008.
- [284] Y. F. Abbasi, P. Panda, S. Arora, B. Layek, and H. Bera, "Introduction to tailor-made biopolymers in drug delivery applications," *Tailor-Made Funct. Biopolym. Syst.*, pp. 1–31, Jan. 2021, doi: 10.1016/B978-0-12-821437-4.00015-3.
- [285] A. Zolfagharian, M. A. P. Mahmud, S. Gharaie, M. Bodaghi, A. Z. Kouzani, and A. Kaynak, "3D/4D-printed bending-type soft pneumatic actuators: fabrication, modelling, and control," *Virtual Phys. Prototyp.*, vol. 15, no. 4, pp. 373–402, Oct. 2020, doi: 10.1080/17452759.2020.1795209.
- [286] E. Yarali, R. Noroozi, A. Yousefi, M. Bodaghi, and M. Baghani, "Multi-Trigger Thermo-Electro-Mechanical Soft Actuators under Large Deformations," *Polymers*, vol. 12, no. 2. 2020, doi: 10.3390/polym12020489.
- [287] M. Jian, Y. Zhang, and Z. Liu, "Natural Biopolymers for Flexible Sensing and Energy Devices," *Chinese J. Polym. Sci.*, vol. 38, no. 5, pp. 459–490, 2020, doi: 10.1007/s10118-020-2379-9.
- [288] A. Zolfagharian, L. Durran, S. Gharaie, B. Rolfe, A. Kaynak, and M. Bodaghi, "4D printing soft robots guided by machine learning and finite element models," *Sensors Actuators A Phys.*, vol. 328, p. 112774, 2021, doi: <https://doi.org/10.1016/j.sna.2021.112774>.
- [289] E. Yarali *et al.*, "Magneto-/ electro-responsive polymers toward manufacturing, characterization, and biomedical/ soft robotic applications," *Appl. Mater. Today*, vol. 26, p. 101306, Mar. 2022, doi: 10.1016/J.APMT.2021.101306.
- [290] E. Colusso and A. Martucci, "An overview of biopolymer-based nanocomposites for optics and electronics," *J. Mater. Chem. C*, vol. 9, no. 17, pp. 5578–5593, 2021, doi: 10.1039/D1TC00607J.
- [291] C. Wang, T. Yokota, and T. Someya, "Natural Biopolymer-Based Biocompatible Conductors for Stretchable Bioelectronics," *Chem. Rev.*, vol. 121, no. 4, pp. 2109–2146, Feb. 2021, doi: 10.1021/acs.chemrev.0c00897.
- [292] L.-Q. Tao *et al.*, "Graphene-Paper Pressure Sensor for Detecting Human Motions," *ACS Nano*, vol. 11, no. 9, pp. 8790–8795, Sep. 2017, doi: 10.1021/acsnano.7b02826.
- [293] K. I. Fabijanic *et al.*, "Self-assembly of biopolymer films for UV protection of wood," *J. Mater. Res.*, vol. 37, no. 1, pp. 55–66, 2022, doi: 10.1557/s43578-021-00348-6.

- [294] C.-Y. Lee, S. Sayyar, P. J. Molino, and G. G. Wallace, “A robust 3D printed multilayer conductive graphene/polycaprolactone composite electrode,” *Mater. Chem. Front.*, vol. 4, no. 6, pp. 1664–1670, 2020, doi: 10.1039/C9QM00780F.
- [295] H. L. Corrêa and D. G. Corrêa, “Polymer Applications for Medical Care in the COVID-19 Pandemic Crisis: Will We Still Speak Ill of These Materials? ,” *Frontiers in Materials* , vol. 7. 2020.
- [296] P. Pandit *et al.*, “Potential biodegradable face mask to counter environmental impact of Covid-19,” *Clean. Eng. Technol.*, vol. 4, p. 100218, 2021, doi: <https://doi.org/10.1016/j.clet.2021.100218>.
- [297] O. Das *et al.*, “The need for fully bio-based facemasks to counter coronavirus outbreaks: A perspective,” *Sci. Total Environ.*, vol. 736, p. 139611, 2020, doi: <https://doi.org/10.1016/j.scitotenv.2020.139611>.
- [298] A. Jahandideh, M. Ashkani, and N. Moini, “Chapter 8 - Biopolymers in textile industries,” S. Thomas, S. Gopi, and A. B. T.-B. and their I. A. Amalraj, Eds. Elsevier, 2021, pp. 193–218.
- [299] A. Moeini, N. Germann, M. Malinconico, and G. Santagata, “Formulation of secondary compounds as additives of biopolymer-based food packaging: A review,” *Trends Food Sci. Technol.*, vol. 114, pp. 342–354, 2021, doi: <https://doi.org/10.1016/j.tifs.2021.05.040>.
- [300] N. Vrinceanu, S. Bucur, C. M. Rimbau, S. Neculai-Valeanu, S. Ferrandiz Bou, and M. P. Sucheai, “Nanoparticle/biopolymer-based coatings for functionalization of textiles: recent developments (a minireview),” *Text. Res. J.*, p. 00405175211070613, Jan. 2022, doi: 10.1177/00405175211070613.
- [301] T. E. Motaung and L. Z. Liganiso, “Critical review on agrowaste cellulose applications for biopolymers,” *Int. J. Plast. Technol.*, vol. 22, no. 2, pp. 185–216, 2018, doi: 10.1007/s12588-018-9219-6.
- [302] S. Bhaladhare and D. Das, “Cellulose: a fascinating biopolymer for hydrogel synthesis,” *J. Mater. Chem. B*, vol. 10, no. 12, pp. 1923–1945, 2022, doi: 10.1039/D1TB02848K.
- [303] A. Patti and D. Acierno, “Towards the Sustainability of the Plastic Industry through Biopolymers: Properties and Potential Applications to the Textiles World,” *Polymers* , vol. 14, no. 4. 2022, doi: 10.3390/polym14040692.
- [304] D. Massella, S. Giraud, J. Guan, A. Ferri, and F. Salaün, “Textiles for health: a review of textile fabrics treated with chitosan microcapsules,” *Environ. Chem. Lett.*, vol. 17, no. 4, pp. 1787–1800, 2019, doi: 10.1007/s10311-019-00913-w.
- [305] A. Rkhaila, T. Chtouki, H. Erguig, N. El Haloui, and K. Ounine, “Chemical Proprieties of Biopolymers (Chitin/Chitosan) and Their Synergic Effects with Endophytic Bacillus Species: Unlimited Applications in Agriculture,” *Molecules* , vol. 26, no. 4. 2021, doi: 10.3390/molecules26041117.
- [306] M. Klein and E. Poverenov, “Natural biopolymer-based hydrogels for use in food and agriculture,” *J. Sci. Food Agric.*, vol. 100, no. 6, pp. 2337–2347, Apr. 2020, doi: <https://doi.org/10.1002/jsfa.10274>.
- [307] E. G. Bahçegül, E. Bahçegül, and N. Özkan, “3D Printing of Hemicellulosic Biopolymers Extracted from Lignocellulosic Agricultural Wastes,” *ACS Appl. Polym. Mater.*, vol. 2, no. 7, pp. 2622–2632, Jul. 2020, doi: 10.1021/acsapm.0c00256.
- [308] “SurgiMend® PRS Thin Collagen Matrix.” <https://www.integralife.com/surgimend-prs-thin-collagen-matrix/product/surgical-reconstruction-plastic-reconstructive-surgery-hospital-or-surgimend-prs-thin-collagen-matrix> (accessed Dec. 13, 2021).
- [309] L. Pinto *et al.*, “Biopolymer hybrid materials: Development, characterization, and food packaging applications,” *Food Packag. Shelf Life*, vol. 28, p. 100676, 2021, doi: <https://doi.org/10.1016/j.fpsl.2021.100676>.
- [310] Y. Liu *et al.*, “A review of cellulose and its derivatives in biopolymer-based for food packaging application,” *Trends Food Sci. Technol.*, vol. 112, pp. 532–546, 2021, doi: <https://doi.org/10.1016/j.tifs.2021.04.016>.
- [311] A. Sobhan, K. Muthukumarappan, and L. Wei, “Biosensors and biopolymer-based nanocomposites for smart food packaging: Challenges and opportunities,” *Food Packag. Shelf Life*, vol. 30, p. 100745, 2021, doi: <https://doi.org/10.1016/j.fpsl.2021.100745>.
- [312] “A bounty of options for biopolymers | Packaging World.” <https://www.packworld.com/issues/sustainability/article/13373562/a-bounty-of-options-for-biopolymers#next-slide> (accessed Dec. 21, 2021).
- [313] “Berry Introduces 100% Recyclable BioPET Bottle - Beauty Packaging.” https://www.beutypackaging.com/contents/view_breaking-news/2019-05-20/berry-introduces-100-recyclable-biopet-bottle/ (accessed Dec. 21, 2021).
- [314] S. Torres-Giner, K. J. Figueroa-Lopez, B. Melendez-Rodriguez, C. Prieto, M. Pardo-Figuerez, and J. M. Lagaron, “Emerging Trends in Biopolymers for Food Packaging,” *Sustainable Food Packaging Technology*. pp. 1–33, Mar. 08, 2021, doi: <https://doi.org/10.1002/9783527820078.ch1>.
- [315] “Improving biopolymers for packaging applications using surface-tailored cellulose nanocrystals – Research Highlights - US Forest Service Research & Development.” https://www.fs.fed.us/research/highlights/highlights_display.php?in_high_id=1271 (accessed Dec. 21, 2021).

[316] “Commentary: Are biopolymers better for the environment? - Recycling Today.”
<https://www.recyclingtoday.com/article/biopolymers-compostable-plastic-recycling/> (accessed Dec. 21, 2021).