

# Carbon nanotubes assisted analytical detection – Sensing/delivery cues for environmental and biomedical monitoring

Tahir Rasheed<sup>a</sup>, Adeel Ahmad Hassan<sup>a</sup>, Fahmeeda Kausar<sup>a</sup>, Farooq Sher<sup>b</sup>, Muhamad Bilal<sup>c</sup> \* and Hafiz M. N. Iqbal<sup>d\*</sup>

<sup>a</sup>School of Chemistry and Chemical Engineering, Shanghai Jiao Tong University, Shanghai 200240, China.

<sup>b</sup>School of Mechanical, Aerospace and Automotive Engineering, Faculty of Engineering, environmental and computing, Coventry University, Coventry CV1 5FB, United Kingdom.

<sup>c</sup>School of Life Science and Food Engineering, Huaiyin Institute of Technology, Huaian 223003, China.

<sup>d</sup>Tecnologico de Monterrey, School of Engineering and Science, Monterrey, 64849, Mexico.

\*Corresponding authors emails: bilaluaf@hotmail.com (M. Bilal); hafiz.iqbal@tec.mx (H.M.N. Iqbal).

## Abstract

The architecture of carbon nanotubes (CNTs) demonstrate phenomenal electronic, mechanical, biological and thermal attributes for highly requisite real-time applications. For instance, electronic and biological features of CNTs are surprisingly striking to engineer robust sensing and/or delivery cues for environmental, analytical diagnostics, and biomedical settings. With CNTs enforcement, several types of pristine and hybrid nanomaterials have been fabricated, though using different support carriers and synthetic or biological materials and used as sensory items or exploited as drug delivery systems (DDSs). Regardless of intensive research and applied potentialities of CNTs, several concerns, such as biodegradability, biotoxicity, and biosafety remains challenging and should be dealt with care prior to design and fabrication. This is mainly because of the lacking standardized protocols and ramification of pristine CNTs or CNTs-based hybrid nano-constructs on the ecosystem and human body are not well-established. For the futuristic use of these remarkable materials in the environmental, analytical diagnostics, and biomedical settings, their biological attributes and multifunctional characteristics must be elucidated with state-of-the-art. Herein, we reviewed CNTs-assisted analytical

32 detection potentialities at large, and sensing/delivery potentialities of CNTs-based cues,  
33 in particular for environmental and biomedical monitoring. Several examples are given  
34 with particular emphasis to biosensors, DDSs, and implantations of CNTs-based cues to  
35 recognize viruses, cancerous cells, glucose, DNA, volatile organic compounds (VOCs)  
36 and various inorganic gases. The review is wrapped-up with concluding notes and brief  
37 outlook over the futuristic developments to further insight the CNTs-based robust cues  
38 and their perspectives for commercialization.

39 **Key words:** Carbon nanotubes; Nano-cues; Biosensors; Biomolecules; Drug release  
40 carriers; Analytical detection; Environmental and biomedical settings

41

## 42 **Introduction**

43 In the past few years, the fabrication of robust nano-structured material, so-called “smart  
44 biomaterials”, as sensing cues has gained great attention in various disciplines of science,  
45 as a booming field of contemporary research. For example, photoactive materials play  
46 important roles in the biosensing systems as transducer converting chemical information  
47 into detectable PEC signal. Photoactive material performs perform a leading role plays in  
48 biosensing of PEC signals through photoelectric conversion. High conversion efficiency  
49 and improved biocompatibility of photoactive materials contribute towards to high-  
50 performance PEC biosensors [1]. Vidya and Prabhat 2020, reported a fluorescence  
51 detection system for Adenosine tri-phosphate (ATP) based on the principle of dissociation  
52 of Thioflavin-T-sulphated- $\beta$ -cyclodextrin assembly dissociation by  $Zn^{2+}$  followed by  
53 reassociation. Such smart biomaterials based sensors provide a reliable, sensitive, and  
54 highly selective approach for ATP [2]. Among nano-structured material, carbon nanotubes  
55 (CNTs) based sensing cues for analytical detection purposes are of supreme interest  
56 [3,4]. Owing to the unique structural, functional, electronic and optical attributes of CNTs,  
57 there has been a significant number of applications from the environmental and  
58 biomedical monitoring perspective, which this review aims to cover to advance the  
59 existing literature with recent trends in this particular field of interest.

60

61 CNTs, particularly single-walled carbon nanotube (SWCNTs) and/or multiwall carbon  
62 nanotubes (MWCNTs), grasp noticeable position for cutting-edge applications in the

63 environment, and biomedicine [5-7]. Nevertheless, prior to such advance applications,  
64 the engineered cues must be subjected to rigorous quality controls to avoid system  
65 failure. However, much sadly, there are no commonly accepted standard regulations to  
66 maintain the quality assurance amongst the commercial suppliers of CNTs. In this  
67 context, the CNTs and CNTs-based constructs, as an analytical target, are a subject of  
68 immeasurable interest [8-10]. Furthermore, tuning the CNTs-based nano-structured cues  
69 facilitate the bio-sensing applications is of crucial importance.

70  
71 Herein, we comprehensively reviewed and discussed the applicability of numerous CNTs-  
72 based sensing/delivery cues with suitable examples. It is worth mentioning that CNTs are  
73 supreme candidates to engineer fascinating diagnostic/analytical devices/tools, for two  
74 reasons, i.e., (1) CNTs exhibit unique structural, functional, electronic and optical  
75 attributes that make them highly suitable to use for analytical detection, and (2) CNTs  
76 open-up innovative tactics to fully integrate and provide extraordinary potentials for further  
77 miniaturization. Following a brief introduction, we reviewed the recent advances in  
78 developing CNTs assisted biosensors, drug delivery systems (DDSs), and their suitable  
79 implantations to recognize viruses, cancerous cells, glucose, DNA, drug release carriers,  
80 volatile organic compounds and various inorganic gases.

81

## 82 **CNT-based biological sensors**

83 Detection of biological molecules and health monitoring are vital in the area of healthcare  
84 ranging from identification to immediate monitoring of the patient's condition. This claim  
85 the reduction of medical costs and mortality globally. For example, it provides early  
86 perceptions of a number of infections which in turn facilitate apposite preventative  
87 measures and treatment. Presently, expensive equipments and dedicated laboratories  
88 are required for a complete medical testing protocol. This necessitates the development  
89 of an efficient approach to deal with the problem. In a study a group of researchers  
90 devised a latest NaYF<sub>4</sub>:Yb,Tm@ ZnO-based biosensor for the detection of CEA linked  
91 with a 3D printed device, which was applied in an equipment used for the detection of  
92 cancer biomarker. NaYF<sub>4</sub>:Yb,Tm@ZnO-based PEC biosensor was used for highly  
93 selective sensing of CEA with the detection limit of 0.032 ng mL<sup>-1</sup> [11]. Similarly,

94 Zhenzhong et al. (2018) introduced a digitalized paper electrode-based sensor  
95 associated with multiwalled carbon nanotube, for the detection of carcinoembryonic  
96 antigen (CEA). The efficiency of sensor toward targeted CEA was observed in the range  
97 of 0.5-60 ng/mL with 167 ng/mL as a limit of detection under optimal conditions [12]. A  
98 near-IR activated-based non-enzymatic PEC sensor was designed by Zhongbin et al.  
99 (2019) [13] for the detection of  $\alpha$ -fetoprotein (AFP) within a dynamic linear range of 10  $\mu$ g  
100 mL<sup>-1</sup> to 50  $\mu$ g with 1.2  $\mu$ g mL<sup>-1</sup> as a limit of detection accompanied decreasing  
101 photocurrent with the rise in AFP concentration. Herein, the CNTs based sensors dealing  
102 with the biological molecules are presented for the benefit of researchers working in the  
103 field of biomolecular sensors [14,15].

104

### 105 **CNTs and volatile organic compounds (VOCs)**

106 Biological volatile organic compounds are not only product and sub product of the cell  
107 metabolism/stress triggered by reactive oxidative species (ROS) [16-18]. They are  
108 categorized on the basis of their functionalities: aliphatic and aromatic hydrocarbons,  
109 aldehydes, ketones, nitriles, alcohols and esters [18]. VOCs concentration varies in  
110 breath and in body excreted fluids with the change in diet, environmental contact, and  
111 diseased state of body [19]. Typically, VOCs started by the cellular activity in human body  
112 include aldehydes, hydrocarbons, and ketones [15]. VOCs higher concentration can find  
113 in normal breath samples; sensor arrays are constructed to differentiate among chemical  
114 species. Arrays of sensors can build “fingerprints” for a class or provided compound.  
115 Haick and coworkers demonstrated the first CNT-based detection of VOCs with an  
116 objective to differentiate between the humanly subjects of cancer of lungs and renal  
117 failure [20]. They designed semiconducting SWCNTs based sensing arrays having  
118 coating of organic materials then, examined their operation to differentiate between  
119 healthy breathing and breath of diseased subjects, **Figure 1** [21]. The obtained signals’  
120 PCA demonstrated the difference between healthy and cancerous breath, was merged  
121 due to the effect from humidity, for an effective discrimination was reduced on the relative  
122 humidity from 80% to below 10% [21]. Later in their study, chemiresistors were replaced  
123 with FET devices having minimal effects from humidity [20]. Recently, they further  
124 reported functionalized SWCNTs arrays together with modified gold NPs on molecular

125 basis to identify seventeen different disease conditions with 86 % accuracy from 1404  
126 subjects [22]. Different groups worked on CNT-based sensing arrays to differentiate  
127 between different VOCs. They further fabricated PCA plots from chemiresistors signals  
128 containing SWCNTs and 8-metalloporphyrins to make difference among different types  
129 of VOCs, such as (a) hydrocarbons, (b) aromatic hydrocarbons, (c) ketones, (d) amines  
130 and (e) alcohols. They differentiated amines from other VOCs by using charge-transfer  
131 competence and considerable differentiation among the rest 4 types based their swelling  
132 effect and intermolecular interaction [23]. Shirsat et al. reported hybrids of SWCNTs and  
133 metalloporphyrins to distinguish methanol, ethanol, acetone, and methyl ethyl ketone [24].  
134 The responses of sensory arrays are dependent on the surfactant-analyte interactions  
135 and assembly with MWCNTs [25]. Every sensor discriminates up to a limit; however, e, g  
136 PCA established separation between water, toluene, ethanol, methanol, acetone, and  
137 chloroform [25]. Selective sensing arrays are successfully produced by noncovalent  
138 functionalization; they are unable to produce significant sensing strength toward the  
139 severe conditions. Sarkar et al. covalently linked poly(tetraphenylporphyrin) on SWCNTs  
140 for the detection of acetone, and the sensing stability exceeds over 180 days period [26].  
141 Wang and Swager used cross-sensitive recognition groups for the functionalization of  
142 MWCNTs following two steps synthesis procedure **Figure 2** [27]. Each selector is used to  
143 amplify the targeted analyte interactions. Allyl- and Propargyl-MWCNTs are polar,  
144 hydrogen-bond acceptors and favorable to strongly interact with large dipoles vapors.  
145 Long alkyl chains (3 and 4) selectors with favorable dispersive interactions designed to  
146 sense aliphatic compounds. Calix(4)arenes [28] used to adsorb the vapors of chlorinated  
147 and aromatic hydrocarbons because of highly polarized pocket; however, crown ether  
148 [29] offered basicity due to H-bonding interactions with alcohols and acids. Therefore, the  
149 sensing arrays classified the tested VOCs into 5 different categories without having the  
150 humidity interference. Furthermore, the responses distinct patterns, when subjected to  
151 LDA precisely recognized all twenty VOCs. Furthermore, the chosen selectors  
152 differentiated chemical spaces in an adequate way. Therefore, the precise chemical  
153 designs, instead of randomly collected selectors, are ideal at precise characterization of  
154 VOCs complex. Sensory arrays are advantageous by covering multiple units is the  
155 “fingerprints” library what can be easily modernized for new class analyte detection. One

156 targeted biomarker is sufficient proof for the presence of a disease; hence the single  
157 analyte detection could be a strong diagnostic tool [30]. Therefore, sensory arrays will  
158 beneficial when applied in parallel with sensors for single analyte detection. Wang et al.  
159 designed CNTs with vertical alignment having conductive coating of polymer to observe  
160 n-pentane with 50 ppm of LOD and appreciable precision over chemicals like methanol  
161 and toluene [31]. Poly(3,4-ethylenedioxythiophene) (PEDOT) used as coating material  
162 for vertically aligned CNTs through chemical vapor oxidative deposition subsequently  
163 nonconducting polystyrene (PS) coating. Pentane adsorption on PEDOT surface  
164 interrupts the pathways' conductivity, and selectivity is observed by PS layer what  
165 eliminates VOCs with certain polarity [31]. Calix(4)arenes have an interacting potential  
166 with aromatic hydrocarbons. **Figure 3** shows quartz crystal microbalance (QCM) analysis  
167 and NMR binding analysis, the authors confirmed the selectivity resulted from the  
168 advantageous p-xylene binding inside the cavity of calixarene over 2 other isomers [32].  
169 Ding et al. designed chemFET sensors using SWCNTs/TiO<sub>2</sub> composites that showed  
170 response at 400 ppb to acetone vapor. The suggested sensory mechanism depends on  
171 the electron/hole pairs generation in TiO<sub>2</sub> layer through UV photoinduction and acetone  
172 adsorption that prevents significant conductance drop [33]. This sensor used for the  
173 detection of acetone up to 20 ppm in humidity and O<sub>2</sub> despite disturbances caused by  
174 water vapor and air. Yoon et al. have incorporated soft Lewis acid Pd<sup>2+</sup> cations enfolded  
175 around SWCNTs. Wherein, selective sensor was produced by the coordination of PdCl<sub>2</sub>  
176 with P4VP-wrapped SWCNTs toward thioethers vapors [34].

177

### 178 **Harnessing the power of CNTs for glucose detection**

179 The detection of glucose level in blood is a commonly used medical test. This  
180 concentration indicates how to manage diabetes, resulting an increase in the glucose  
181 sensors demand. Both chemFET and electrochemical sensors are being used with  
182 selector such as glucose oxidase (GOx). The reduction potential in H<sub>2</sub>O<sub>2</sub> production and  
183 transfer of electron from GOx to CNT electrodes beneficial for electrochemical sensing  
184 by composites of GOx/CNT [35]. However, the enzymatic adsorption onto CNT electrodes  
185 reported several problems, such as denaturation and enzyme leaching (Balasubramanian  
186 et al. 2006). For this purpose, metal and metal oxide are used to solve this problem.

187 Suitable examples were mentioned for MWCNTs by Wang et al. (2009) [32] coated with  
188 ZnO NPs and by Chen et al. (2012) [36] coated with Pt–Pd bimetallic NPs. A polymeric  
189 layer was applied to remove commonly used interferents. Different groups reported  
190 adsorption of GOx on CNTs using electropolymerization of conductive polymers. Herein,  
191 GOx is combined with a monomer and then electropolymerization at CNT electrode. Gao  
192 et al. reported a large range of 2.5– 20 mM and resulted that the aligned Fe particles and  
193 MWCNTs were crucial to lower down H<sub>2</sub>O<sub>2</sub> oxidation potential, in this way veroxidation of  
194 PPy can be prevented. Firstly, SWCNTs were attached to the gold electrodes surface  
195 with a mixed single layer of cystamine and thioethanol together with EDC. On the other  
196 hand, nonenzymatic CNT glucose sensors mainly dependent on the characteristics of  
197 metal-based NPs coated on CNTs to build glucose response electrochemically. Lin et al.  
198 applied copper and nickel NP-coated MWCNTs to show an activity toward the oxidation  
199 of glucose [37], while Gougis et al. deposited AU nanoparticles onto CNT electrodes [38].  
200 Currently, Baghayeri et al. described the electrodeposition of silver nanoparticles onto  
201 MWCNTs functionalized with metformin, used as selective glucose electrochemical  
202 sensors, **Figure 4** [39]. Those sensors showed a minimal detection limit at 0.3 nM without  
203 interference from biological entities found inside blood serum and urine samples. Lerner  
204 et al. detected glucose by means of complexes formed by boronic acid moieties, **Figure**  
205 **5** [40].

206

### 207 **Harnessing the power of CNTs for DNA detection**

208 DNA detection has prime importance toward analysis of genetic disorders, detection of  
209 pathogens, anticipation biowarfare agents, and drug discovery [35]. Sensors comprising  
210 optical, piezoelectric, and electrochemical transductions occur due to the selective base-  
211 pair interactions within the strands of DNA [41]. CNT-based DNA sensors were initially  
212 studied because of high selectively, precision, and reproducibility. In these sensors, a  
213 single-stranded DNA (ssDNA) was immobilized on the electrode and electrical current  
214 changes were activated due to complementary sequence hybridization [42]. Several  
215 researchers stated that chemFETs can be used to sense resistance change of CNT or  
216 groups of CNTs in DNA presence. To monitor FET transfer properties of pristine SWCNTs  
217 upon the addition of ssDNA oligonucleotides. Subsequently, DNA hybridization with

218 selected DNA caused the reduced conductance at the voltage of gate. Such scheme  
219 successfully differentiated between mutant and HFE gene alleles (wild-type), resulted in  
220 the hereditary hemochromatosis. In this way, the sensitivity can be improved of FET-  
221 based devices for the detection of DNA complementary strands [43]. Upon the addition  
222 of intercalator, the hybridized sensors having DNA presented noticeably reduced  
223 conductivity than those of the samples with incompatible DNA. Dong et al. introduced the  
224 usage of Au NPs, functionalized with DNA, to improve FET sensors having the LOF of  
225 100 fM, **Figure 6** [44]. Herein, every targeted DNA bind to the Au NPs functionalized with  
226 DNA wherein SWCNT with immobilized DNA. Remarkably, the authors have described  
227 that Ta electrodes-based devices possessed larger enhancement than those of the AU  
228 electrode-based devices. The selectivity promoted through the DNA–CNT composites-  
229 based sensors is emphasized through the base-pair mismatches detection ability. The  
230 charge transfer can transpire through the aromatic DNA base-pairs over significant  
231 distances, it has a prime level of sensitivity toward the base pairing integrity. The authors  
232 reported an original conductivity measurements of single DNA duplex supported through  
233 covalent bonds between a single SWCNT, **Figure 7a** [45]. They cut individual SWCNTs  
234 through an electron beam to fabricate the devices and then ensured the carboxylic acid  
235 functional groups presence on the gap both sides through oxygen plasma. The gaps were  
236 connected by ssDNA anchored through amide linkages at both ends. This device clearly  
237 observed the conductivity difference between duplexes. **Figure 7b** displays that a single  
238 mismatch (both CA and GT) causes current reduction through the strand of DNA.  
239 Weizmann et al. demonstrated the ssDNA-bridged CNTs networks for complementary  
240 DNA chemiresistive detection, **Figure 8** [46]. The ssDNA gaps furnished the insulation of  
241 materials. Double-stranded DNA assemblies were formed through the selective binding  
242 of ssDNA analyte; however, extended sequence of DNA transport cannot provide enough  
243 sensing conductivity.

244

### 245 **Detection of substance in humane body**

246 Numerous studies have applied biosensors of different types to detect various substances  
247 such as drugs (acetaminophen), H<sub>2</sub>O<sub>2</sub>, neurotransmitters (dopamine), urea and glucose,  
248 among others. The material constituent's modification for every biosensor and

249 functionalized CNTs are observed to enhance the response and device properties.  
250 Bioanalytical studies are mainly based on the conductive polymers, to their charge carry  
251 ability, sensitivity, and biocompatibility up to negligible perturbations. The CNTs  
252 fabrication in electrodes prepared through conductive polymers constructs p-p  
253 hydrophobic and electronic interactions with the aromatic compounds. Microorganisms  
254 having multiple enzymes are involved in biosensing of conductive polymers. Enzymes  
255 may be simply applied in numerous substrates under particular conditions of culture.  
256 However, a minimal distance between microbial cells and transducer is recommended for  
257 efficient biochemical signals what may be attained through immobilization of surface cells  
258 of transducer [47]. Paracetamol is widely used for the pain relief, an analgesic and  
259 antipyretic drug., it is essential to know about its mechanistic toxicity. The paracetamol  
260 can be sensed using different techniques: liquid chromatography;  
261 electrospectrophotometry. The electrochemical methods have the following  
262 characteristics including fast response, low cost, high sensitivity, simple instrumentation,  
263 and low required power. The electrochemical methods have been applied to analyze the  
264 electroactive compounds [47]. Bayram and Akyilmaz (2016) [47] designed *Bacillus*  
265 species based microbial biosensor to determine the sensitivity of paracetamol. This  
266 biosensor comprised of MWCNTs modified gold electrode, glutaraldehyde crosslinking  
267 agent and polyaniline (PANI). They introduced a microbial system used for the  
268 quantitative detection of paracetamol. Additionally, the connections between PANI and  
269 MWCNTs offer improved stability and biosensor responses' conductivity. Dopamine as  
270 neurotransmitter is accountable for transmitting motor, cognitive, and behavioral  
271 functions. Parkinson's disease is sensed by the neurotransmitter disorder. The dopamine  
272 detection is a big challenge because of its lower concentration than those of uric and  
273 ascorbic acids.<sup>56</sup> The biosensors selectivity can be improved to modify the electrode  
274 materials through platinum, glassy carbon or gold with CNTs. Palomäki et al. (2018) [48]  
275 applied films of tetrahedral amorphous carbon (ta-C) to detect dopamine with uric and  
276 ascorbic acids at physiological concentrations. Comparing electroanalytical performance  
277 and biocompatibility, MWCNTs were grown by CVD on top of ta-Cy. Such direct growth  
278 of MWCNTs on ta-C surface ensures reproducibility and appropriate mechanical strength.  
279 They determined that the detection limits did not improve by the modification of ta-C films

280 with MWCNTs but provided the selectivity for dopamine detection and dopamine  
281 interferences, ascorbic and uric acids, simultaneously, at physiological concentrations.  
282 Considering the electroanalytical behavior, ta-C and ta-C+ MWCNTs have great tendency  
283 to use for the in vivo sensing. The CNTs proportion of length/diameter offers high  
284 proportion of surface/volume and promote their rapid electron transfer ability. Chen et al.  
285 applied BCNTs to produce an amperometric enzymatic sensor. A poly(o-aminophenol)  
286 film (POAP), was used for the electrode protection from contaminating in H<sub>2</sub>O<sub>2</sub>, AU,  
287 glucose biosensors, was used to immobilize GOD. Thus, the glucose biosensor  
288 performance dependent on the glassy carbon electrode, examined in detail, high  
289 precision, short response time, low LOD, and good stability were found. Thus, BCNTs  
290 have potential applications in amperometric biosensors [49]. Enzymes specific catalytic  
291 activity for certain analytes together with their viable accessibility proves them desirable  
292 for the biosensors' fabrication. However, the leading challenge is the enzymes stability  
293 on transducers' surface as the enzyme's contamination directed toward the poor  
294 analytical performance. The enzymatic biosensors production may involve different  
295 treatments for immobilization [50]. H<sub>2</sub>O<sub>2</sub> is widely applicable for pathogen elimination, cell  
296 signaling, disinfection, bleaching and control of odors. H<sub>2</sub>O<sub>2</sub> concentration monitoring is  
297 practiced observing the enzymatic reactions progress. The biomolecules and transition  
298 metal complexes such as myoglobin, hemoglobin (HB), and Prussian blue are used to  
299 develop the H<sub>2</sub>O<sub>2</sub> sensors. HB is an economical and stable protein molecule what can  
300 interact with oxygen, H<sub>2</sub>O<sub>2</sub>, and carbon dioxide in the biocatalyst. PANI/polysaccharide  
301 composites are the potential applicants for numerous applications, and natural  
302 polysaccharide (starch) are abundantly available, used for manufacturing of  
303 biocompatible materials. The urea evaluation can be made using techniques such as  
304 chromatographic, spectrophotometric, calorimetric, electrochemical and fluorimetry  
305 techniques. However, electrochemical technique, have advantages of lower cost, high  
306 selectivity and sensitivity of analyte detection and less operating time. The urease  
307 application coupled with MWCNTs-PAMAM was utilized to determine the urea level in  
308 human body. The optimized pH, operating potential, temperature, and storage  
309 characteristics decide the analytical performance of the G5 electrode. It is resulted the  
310 G5 of MWCNTs-PAMAM demonstrated a considerably improved performance of

311 biosensor compared to PAMAM of other generations, associated with the enzyme  
312 immobilization efficiency. Additionally, the urea biosensor showed high sensitivity, shorter  
313 response time, low detection limit, high sensitivity, stability, and reproducibility. This  
314 biosensor can be used to quantify human blood urea different samples [51].

315

### 316 **Harnessing the power of CNTs for virus's detection**

317 Viruses are non-cellular infectious agents. Viruses promote different diseases in human  
318 body and in animals. Thus, virus's detection efficiency has an extreme importance and  
319 alternatives are searched in this regard. For this purpose, CNTs are being increasingly  
320 employed, because of electrical properties and high surface area [14]. Earlier, several  
321 detection studies were applied, for example, in the detection of dengue [52-55]. Dengue  
322 is a viral disease which is primarily spread by mosquitoes, Wasik et al. (2017) [52] have  
323 reported heparin functionalized SWCNTs functionalized to design electronic  
324 chemiresistive biosensor. The authors introduced functionalization of SWCNTs with 1-  
325 pyrenemethylamine through non-covalent modification. Three different constituents of  
326 ethyl-3-dimethyl aminopropyl carbodiimide, heparin and N-hydroxy succinimide and were  
327 used to prepare the biosensor. It was observed that an increase in the electrical  
328 resistance occurs during functionalization and dengue virus interaction to heparin  
329 receptor. Tran et al. (2017) [53] reported CNT field effect transistor (CNTFET) in which  
330 functionalization of CNTs based on nitric acid and then, influenza virus DNA was  
331 immobilized on the CNTs network through physisorption. This was done through dipping  
332 of DNA sensor in a targeted DNA solution, dissolved in PBS. Some factors such as the  
333 concentration of the probe, thermal impact, immobilization time, reproducibility, range of  
334 detection and response time, are assessed. The results showed DNA sensor exhibited a  
335 short time to response together with high duplicability Fu et al. (2017) [54] reported a fast  
336 and highly efficient influenza virus (AIV) detection subtype H5N1 DNA sequences.  
337 SWCNTs (sc-SWCNTs) and nitrogen-doped MWCNTs (N-MWCNTs) were used in the  
338 biosensor as sensing elements. SWCNTs have no horizontal alignment. So, 5 to 20 mm  
339 length of sc-SWCNTs were achieve through short fragments CVD epitaxial elongation on  
340 the substrates made of quartz. After CNTs preparation, they founded target substrate  
341 arrays. Then, a standard microfabrication procedure was used to fabricate chemiresistor-

342 type sensors. Generally, CNT-based DNA sensors showed remarkable characteristics,  
343 such as flexibility, small size, easy to use, and overly sensitive, what make them favorable  
344 in portable applications and clinical diagnostics [54].

345

### 346 **Harnessing the power of CNTs for cancer detection**

347 Cancer disrupts the immune system and defense mechanism of the host [56]. This  
348 disease has become caused of countless death around the globe therefore several efforts  
349 were put to reduce the effect of this disease in the recent times. In this connection CNTs  
350 have been reported for the development of fast and effective detection as well as  
351 treatment tool of several cancer diseases [57-60]. Zanganeh et al. (2016), [57] developed  
352 an electrical biosensor for detection of cancer cells. The CNTs were conjugated to folic  
353 acid (FA-VACNTs) molecules. The direct-current plasma enhanced CVD was applied to  
354 produce CNTs for biosensor. The CNTs were then functionalized to form NH<sub>2</sub>-VACNTs  
355 by double barrier discharge plasma system. Human lung cells QUDB and MRC-5 were  
356 isolated, which are carcinoma and normal tumors, respectively. The biosensors were  
357 tested by taking cells of same concentrations from culture flask and applying on VACNT  
358 and FA-VACNT using microfluidic pump. The FA-VACNT electrodes showed better  
359 electrical response of the sensors. This device could be applied to detect and monitor  
360 cancer [57]. The nanohybrid microelectrode has shown good detection performance for  
361 H<sub>2</sub>O<sub>2</sub> with efficient structural mechanics integrated with electrochemical properties.  
362 Therefore, this biosensor has demonstrated high sensitivity and selectivity over a wide  
363 linear range with excellent mechanical stability and better biocompatibility and could be  
364 useful in chemotherapy or radiotherapy treatments [58]. CNTs based biosensors were  
365 also helpful in cancer treatment by detection of the concentration of substances. In this  
366 regard, H. Zhou et al. (2018) [61] produced an analytical device which could detect  
367 anticancer drug in the whole blood, using as a model system to detect methotrexate  
368 (MTX). MTX is used to cure several cancers; though, disproportionate use of it can cause  
369 intoxication. Controlling the MTX concentration in blood by optimized dosage is important  
370 to reduce its side effects. Uniformly distributed tungsten phosphide and nitrogen-doped  
371 CNTs produced synergistic effect to transmit electronic channel which ensured excellent

372 performance of the proposed sensor with a reproducibility for detection of MTX,  
373 satisfactory selectivity, short response time, wide and detection range in whole blood.

374

### 375 **Harnessing the power of CNTs as drug release carriers**

376 Controlled drug release has become the furthestmost challenging as it requires to  
377 overcome both biological as well as physicochemical barriers [62]. SWCNTs and  
378 MWCNTs have been potentially used in vaccines, immunopotential and gene transfer  
379 fields of medicinal chemistry [63,64]. Since 2004, CNTs were reported for drug delivery,  
380 [65] and after that several methods have been manifested to enhance the use of nano-  
381 materials for DDSs. In this field the recent CNTs approaches are functionalization, coating  
382 with polymers, encapsulation, liposomes, and fabrication of structures for example,  
383 buckypapers, hydrogels, irregular meshes, and membranes, as shown in Fig. 5. The  
384 purpose of these modifications was to improve CNTs' dispersion in biological fluids, water  
385 solubility and simultaneously decrease their cytotoxicity [66]. More than 60% CNTs as  
386 drug delivery are applied for cancer treatment which is now considered a fatal disease  
387 worldwide [67]. Death rate in the recent years has increased due to various cancer  
388 diseases as per Annual Report [68], where, pancreas, brain and liver are among other  
389 types of cancers. Due to this reason the scientists have given much attention to this area.  
390 As reported by Wang and coworkers (2015) [69], that compared to nonfunctionalized  
391 CNTs, functionalized ones are less toxic which can be excreted from the human system.  
392 Added modifications such as, encapsulation or functionalization, can also be carried out  
393 on CNTs after loading a model drug. To overcome the challenges for triggered drug  
394 delivery, efforts are being carried out by using several alternative options. Some of them  
395 will be presented here. A dual drug delivery system which is pH responsive have been  
396 developed by Yang et al. (2017) [70]. This system can be used for cancer chemotherapy.  
397 The group utilizes the bigger inner-diameter MWCNTs. Doxorubicin (Dox) and Cisplatin  
398 (CDDP) anti-tumorous drugs were taken as non-covalent functionalization agents. The  
399 inner-side of MWCNTs were encapsulated by CDDP, and folic acid (FA). Different pH  
400 conditions were employed to manifest a biphasic release profile of FA bonded DDS and  
401 DOX was noticed with the pH-sensitive release pattern. DOX and CDDP share a collegial  
402 antitumor activity by hindering cell growth and ultimately leading the cell to its death. In

2016, a bio-based scheme for Michael addition reaction and surface PEGylation of CNTs together with mussel-based chemistry was introduced by Xu et al. The later was a new strategy motivated by the efficiency of mussel to adhere to various material surfaces. The mussel proteins have amino acid 3,4- dihydroxyphenyl-L-alanine as the vital constituent which is responsible for its strong adhesive ability [71]. For the intracellular delivery of Dox, Xu et al. (2016) [71] demonstrated that PEGylated CNTs had biological potential. These PEGylated CNTs with enhanced biocompatibility toward cancer cells and are well dispersed in organic solutions and water, are fairly proved by results. PEGylated CNTs was efficiently used as substrate to load Dox and introduced into cancer cells. Because of the design-ability of polymerization and effectively strong adhesion of PDA, this technique could also be implied for surface modification to subsume other polymer materials. Based on photothermal effects, a thermos-sensitive hydrogel to control drug release was proposed by Dong et al. in 2017 [72]. Chitosan (CH) and MWCNTs combined with DDS had built on a thermosensitive hydrogel concept. Firstly, Dox was loaded on MWCNTs with non-covalent bonds and followed by addition of CH, the stirring mixture was resulted in CH-MWCNTs. Free Dox and Dox-loaded CH-MWCNTs having anti-tumor cell proliferation effect demonstrated that at any concentration, CH-MWCNTs loaded with Dox had decreased toxicity in comparison to free Dox, and ultimately the Dox is brought properly by CH-MWCNTs to the tumor cells and release it in the cytoplasm. So, decreased toxicity avocats the prolonged therapeutic effects. The encapsulation of hydrophilic drugs is proficient with MWCNTs/hydrogel as a drug carrier for in vitro drug release, it accelerates the drug release rate after initiation of photothermal conversion which is stimulated with an 808 nm laser irradiation. Sciortino et al. (2017) [73] used Dox as an anticancer drug and mentioned the efficiency of using MWCNTs by evaluating length and incubation time. Long (500 222 nm) and short (130 85 nm) MWCNTs were loaded with Dox and embellished with biotin. The non-internalized long MWCNTs release Dox impromptu which lessen the differences as indicated by the afterwards testing [73]. Dox is being extensively studied as an anti-tumor drug just like CNTs, and it could be seen from previously mentioned examples that many endeavors have been done for introducing numerous alternatives to overcome its leftover cytotoxicity. SWCNTs were modified by applying a simple non-covalent method with an asparagine–glycine–arginine

434 (NGR) peptide. In addition, they were loaded with Gd–DTPA by n–p interactions and Dox  
435 by  $\pi$ – $\pi$  stacking interactions. This system facilitates the diagnosis of the tumor and  
436 chemotherapy in a unique system by accumulating in and entering tumor cells. The  
437 developed material is capable of increasing drug intracellular accumulation by efficiently  
438 promoting cellular uptake, and they are appropriate carriers for DDS because having no  
439 cytotoxicity against MCF-7 cells. Moreover, scientists also used subcutaneous injection  
440 to prepare tumor-bearing mice model after the tumor and performed intracellular studies  
441 which spread to a volume of 100 mm<sup>3</sup>. The tail vein intravenous injection was implied for  
442 treating tumor-bearing mice in order to evaluate targeting efficiency and bio-distribution  
443 of DOX/NGR-SWCNTs/Gd–DTPA. Tumor-bearing mice were also used as model to  
444 examine the antitumor potency, and coronal and axial MR images of mice were achieved.  
445 Excellent tumor targeting properties were manifested by the results. The developed  
446 system is noticed to be out of harm’s way for tumor therapy at the treatment dosage and  
447 is indicated by systemic toxicity. DOX/NGR-SWCNTs/Gd–DTPA injection was used for  
448 the treatment of group resulted with increased signal intensity, furnishing outstanding  
449 image contrast in *in-vivo* MRI. The toxicity of SWCNTs is reduced by modification with  
450 hydroxypropyl-b-cyclodextrin (HP-b-CD) and the delivery of formononetin (FMN), which  
451 is an anticancer drug, were beautifully demonstrated by Liu et al. in (2018) [74]. SWCNTs  
452 were detected with relatively high entrapment efficiency on account of covalently  
453 functionalized and formerly carboxylated with cyclodextrin (CD-SWCNTs) and, FMN was  
454 loaded posteriorly which in-turn could be positioned on HP-b-CD (CD “side”) of SWCNTs.  
455 Their results exhibited that FMN-loaded SWCNTs could be appropriate carriers of the  
456 anticancer drug for in vivo delivery, and the antitumor activity FMN is greatly enhanced  
457 as CD-SWCNTs-FMN. The previously cited SWCNTs were also used by Razzazan et al.  
458 (2016) [75] as a drug carrier for cancer treatment; instead gemcitabine (GEM) covalently  
459 bonded to the surface of the tubes was chosen as an anticancer agent. After a wide  
460 application of SWCNTs in successive acylation, carboxylation, PEGylation and amination  
461 reactions, SWCNTs and SWCNTs-PEG was covalently bonded to GEM. The drug  
462 entrapment efficiency of both SWCNTs–GEM and SWCNTs– PEG–GEM was observed  
463 to be alike, fairly enough to endorse intracellular cytotoxic and concentration effects.  
464 SWCNTs provide a pH-responsive release profile because they release GEM quickly at

465 lower pH values following cleavage of ester bonds. The cytotoxicity of SWCNTs PEG–  
466 GEM was reduced by PEGylation, MTT assay was used for evaluating in vitro cytotoxicity  
467 against MIA PaCa-2 and A549 cells. Instead of pure GEM, cell uptake of SWCNTs was  
468 improved when conjugated with GEM and PEG. Evaluation of the in vivo tumor volume  
469 in mice and perceived the enhanced antitumor activity of GEM owing to its delayed  
470 degradation while attached to SWCNTs. It was noticed that SWCNTs–PEG–GEM  
471 decreased the tumor growth, compared to SWCNTs–GEM, after 23 days. Consequently,  
472 the tumor efficacy of GEM increases when conjugated with PEGylated SWCNTs,  
473 advocating this novel platform to be a proficient for delivering hydrophilic anticancer drugs  
474 efficiently. This report is regarded as the solely introduction of the anti-cancer drug which  
475 is covalently attached to the surface of the CNTs. Currently, Schwengber and coworkers  
476 (2017) [76] have studied buckypapers of CNTs which are regarded as electromodulated  
477 transdermal DDS. Nylon membranes involving SWCNTs–COOH were used to produce  
478 buckypapers, and other model drugs were used like clonidine hydrochloride (CHC),  
479 ketorolac tromethamine (KT), flurbiprofen (FB) and selegiline hydrochloride (SHC).  
480 SWCNTs–COOH individually adsorbed the drugs into it, and an elevated level of drug  
481 loading (89.4%) was attained. The lowest value for FB was spotted for 8 h of release,  
482 followed by SHC, CHC and KT. Grounding on the drug usage, the bucky papers  
483 demonstrated the important attributes for passive drug release developed for controlled  
484 drug release.

485

#### 486 **Conclusion notes and future outlook**

487 In conclusion, though the incorporating CNTs into analytical detection tools, i.e.,  
488 biosensors have resulted a great deal of study in the past few years. However, the full  
489 potential of CNTs or CNTs-assisted multifunctional materials is yet to be realized. So far,  
490 the most widely employed applications of CNTs have been the engineering of numerous  
491 detection tools and/or as a sorption material for various polluting agents. In addition to the  
492 above-mentioned applications of CNTs and considering the extensive attention in CNTs,  
493 it is not astonishing that one can spotlight different analytical potentialities of CNTs.  
494 The ongoing current pace in the development and deployment of these applications  
495 designates prompt application of CNTs in fabricating contemporary analytical

496 instrumentation and in developing new analytical tools. From the future applications  
497 perspective, the studies should be designed by imagining broader application of CNTs in  
498 nano-electronic tools for multi-industrial sectors. In addition, the strategy of individually  
499 addressed matrices of nano-electrodes may lead to the construction of robust multi-  
500 component biosensors. Moreover, considering the surface, chemical, or biological  
501 functionalization of CNTs or CNTs-assisted nanostructured cues can be exploited in  
502 developing new micro-separation techniques. In the near future, all above-mentioned  
503 analytical features of the growing environmental and biomedical monitoring field of  
504 nanotechnology should lead to momentous progress in analytics and related areas.

505

#### 506 **Acknowledgments**

507 The listed author(s) are highly obliged to their institutes and universities for the literature  
508 services.

509

#### 510 **Declaration of interests**

511 The listed author(s) declare no conflict of interest.

512

513 **References**

- 514 [1] Zhou, Q., & Tang, D. (2020). Recent advances in photoelectrochemical biosensors for  
515 analysis of mycotoxins in food. *TrAC Trends in Analytical Chemistry*, 124, 115814.
- 516 [2] Singh, V. R., & Singh, P. K. (2020). A supramolecule based fluorescence turn-on and  
517 ratiometric sensor for ATP in aqueous solution. *Journal of Materials Chemistry B*, 8(6),  
518 1182-1190.
- 519 [3] Arribas, A. S., Moreno, M., González, L., Blázquez, N., Bermejo, E., Zapardiel, A., &  
520 Chicharro, M. (2020). A comparative study of carbon nanotube dispersions assisted  
521 by cationic reagents as electrode modifiers: Preparation, characterization and  
522 electrochemical performance for gallic acid detection. *Journal of Electroanalytical  
523 Chemistry*, 857, 113750.
- 524 [4] Trojanowicz, M. (2006). Analytical applications of carbon nanotubes: a review. *TrAC  
525 trends in analytical chemistry*, 25(5), 480-489.
- 526
- 527 [5] Bilal, M., Rasheed, T., Mehmood, S., Tang, H., Ferreira, L. F. R., Bharagava, R. N., &  
528 Iqbal, H. M. (2020). Mitigation of environmentally-related hazardous pollutants from  
529 water matrices using nanostructured materials—A review. *Chemosphere*, 126770.
- 530 [6] Raphey, V. R., Henna, T. K., Nivitha, K. P., Mufeedha, P., Sabu, C., & Pramod, K.  
531 (2019). Advanced biomedical applications of carbon nanotube. *Materials Science  
532 and Engineering: C*, 100, 616-630.
- 533 [7] Rasheed, T., Nabeel, F., Adeel, M., Rizwan, K., Bilal, M., & Iqbal, H. M. (2019). Carbon  
534 nanotubes-based cues: a pathway to future sensing and detection of hazardous  
535 pollutants. *Journal of Molecular Liquids*, 292, 111425.
- 536 [8] Hatefi-Mehrjardi, A., Karimi, M. A., Soleymanzadeh, M., & Barani, A. (2020). Highly  
537 Sensitive Detection of Dopamine, Ascorbic and Uric Acids using Dianix Yellow/Multi-  
538 walled Carbon Nanotubes Modified Electrode. *Journal of Analytical Chemistry*, 75,  
539 366-377.
- 540 [9] Stefano, J. S., Lima, A. P., Nascentes, C. C., Krzyzaniak, S. R., Mello, P. A.,  
541 Gonçalves, J. M., ... & Munoz, R. A. (2020). Electrochemical detection of 2, 4, 6-

542 trinitrotoluene on carbon nanotube modified electrode: Effect of acid  
543 functionalization. *Journal of Solid State Electrochemistry*, 24(1), 121-129.

544 [10] Valcarcel, M., Simonet, B. M., Cardenas, S., & Suarez, B. (2005). Present and future  
545 applications of carbon nanotubes to analytical science. *Analytical and bioanalytical*  
546 *chemistry*, 382(8), 1783-1790.

547 [11] Lv, S., Zhang, K., Zhu, L., & Tang, D. (2019). ZIF-8-assisted NaYF<sub>4</sub>: Yb, Tm@ ZnO  
548 converter with exonuclease III-powered DNA walker for near-infrared light responsive  
549 biosensor. *Analytical Chemistry*, 92(1), 1470-1476.

550 [12] Yu, Z., Tang, Y., Cai, G., Ren, R., & Tang, D. (2018). Paper electrode-based flexible  
551 pressure sensor for point-of-care immunoassay with digital multimeter. *Analytical*  
552 *chemistry*, 91(2), 1222-1226.

553 [13] Luo, Z., Qi, Q., Zhang, L., Zeng, R., Su, L., & Tang, D. (2019). Branched  
554 polyethylenimine-modified upconversion nanohybrid-mediated photoelectrochemical  
555 immunoassay with synergistic effect of dual-purpose copper ions. *Analytical*  
556 *chemistry*, 91(6), 4149-4156.

557 [14] , N., Chen, X., Ren, T., Zhang, P., & Yang, D. (2015). Carbon nanotube based  
558 biosensors. *Sensors and Actuators B: Chemical*, 207, 690-715.

559 [15] Ellis, J. E., & Star, A. (2016). Carbon nanotube based gas sensors toward breath  
560 analysis. *ChemPlusChem*, 81(12), 1248.

561 [16] Amal, H., Leja, M., Funka, K., Skapars, R., Sivins, A., Ancans, G., ... & Haick, H.  
562 (2016). Detection of precancerous gastric lesions and gastric cancer through exhaled  
563 breath. *Gut*, 65(3), 400-407.

564 [17] Broza, Y. Y., Mochalski, P., Ruzsanyi, V., Amann, A., & Haick, H. (2015). Hybrid  
565 volatolomics and disease detection. *Angewandte Chemie International*  
566 *Edition*, 54(38), 11036-11048.

567 [18] Hakim, M., Broza, Y. Y., Barash, O., Peled, N., Phillips, M., Amann, A., & Haick, H.  
568 (2012). Volatile organic compounds of lung cancer and possible biochemical  
569 pathways. *Chemical reviews*, 112(11), 5949-5966.

- 570 [19] Bajtarevic, A., Ager, C., Pienz, M., Klieber, M., Schwarz, K., Ligor, M., ... & Hilbe, W.  
571 (2009). Noninvasive detection of lung cancer by analysis of exhaled breath. *BMC*  
572 *cancer*, 9(1), 348.
- 573 [20] Peng, G., Tisch, U., & Haick, H. (2009). Detection of nonpolar molecules by means  
574 of carrier scattering in random networks of carbon nanotubes: toward diagnosis of  
575 diseases via breath samples. *Nano letters*, 9(4), 1362-1368.
- 576 [21] Peng, G., Trock, E., & Haick, H. (2008). Detecting simulated patterns of lung cancer  
577 biomarkers by random network of single-walled carbon nanotubes coated with  
578 nonpolymeric organic materials. *Nano letters*, 8(11), 3631-3635.
- 579 [22] Nakhleh, M. K., Amal, H., Jeries, R., Broza, Y. Y., Aboud, M., Gharra, A., ... & Glass-  
580 Marmor, L. (2017). Diagnosis and classification of 17 diseases from 1404 subjects  
581 via pattern analysis of exhaled molecules. *ACS nano*, 11(1), 112-125.
- 582 [23] Liu, S. F., Moh, L. C., & Swager, T. M. (2015). Single-walled carbon nanotube–  
583 metalloporphyrin chemiresistive gas sensor arrays for volatile organic  
584 compounds. *Chemistry of Materials*, 27(10), 3560-3563.
- 585 [24] Shirsat, M. D., Sarkar, T., Kakoullis Jr, J., Myung, N. V., Konnanath, B., Spanias, A.,  
586 & Mulchandani, A. (2012). Porphyrin-functionalized single-walled carbon nanotube  
587 chemiresistive sensor arrays for VOCs. *The Journal of Physical Chemistry C*, 116(5),  
588 3845-3850.
- 589 [25] Chatterjee, S., Castro, M., & Feller, J. F. (2015). Tailoring selectivity of sprayed  
590 carbon nanotube sensors (CNT) towards volatile organic compounds (VOC) with  
591 surfactants. *Sensors and Actuators B: Chemical*, 220, 840-849.
- 592 [26] Sarkar, T., Srinives, S., Sarkar, S., Haddon, R. C., & Mulchandani, A. (2014). Single-  
593 walled carbon nanotube–poly (porphyrin) hybrid for volatile organic compounds  
594 detection. *The Journal of Physical Chemistry C*, 118(3), 1602-1610.
- 595 [27] Wang, F., & Swager, T. M. (2011). Diverse chemiresistors based upon covalently  
596 modified multiwalled carbon nanotubes. *Journal of the American Chemical*  
597 *Society*, 133(29), 11181-11193.
- 598 [28] Hong, G., Diao, S., Antaris, A. L., & Dai, H. (2015). Carbon nanomaterials for  
599 biological imaging and nanomedicinal therapy. *Chemical reviews*, 115(19), 10816-  
600 10906.

- 601 [29] Wang, J., & Musameh, M. (2005). Carbon-nanotubes doped polypyrrole glucose  
602 biosensor. *Analytica Chimica Acta*, 539(1-2), 209-213.
- 603 [30] Peveler, W. J., Yazdani, M., & Rotello, V. M. (2016). Selectivity and specificity: pros  
604 and cons in sensing. *ACS sensors*, 1(11), 1282-1285.
- 605 [31] Wang, X., Ugur, A., Goktas, H., Chen, N., Wang, M., Lachman, N., ... & Gleason, K.  
606 K. (2016). Room temperature resistive volatile organic compound sensing materials  
607 based on a hybrid structure of vertically aligned carbon nanotubes and conformal  
608 oCVD/iCVD polymer coatings. *Acs Sensors*, 1(4), 374-383.
- 609 [32] Wang, Y. T., Yu, L., Zhu, Z. Q., Zhang, J., Zhu, J. Z., & Fan, C. H. (2009). Improved  
610 enzyme immobilization for enhanced bioelectrocatalytic activity of glucose  
611 sensor. *Sensors and Actuators B: Chemical*, 136(2), 332-337.
- 612 [33] Ding, M., Sorescu, D. C., & Star, A. (2013). Photoinduced charge transfer and  
613 acetone sensitivity of single-walled carbon nanotube–titanium dioxide  
614 hybrids. *Journal of the American Chemical Society*, 135(24), 9015-9022.
- 615 [34] Yoon, B., Liu, S. F., & Swager, T. M. (2016). Surface-anchored poly (4-vinylpyridine)–  
616 single-walled carbon nanotube–metal composites for gas detection. *Chemistry of*  
617 *Materials*, 28(16), 5916-5924.
- 618 [35] Wang, J. (2005). Carbon-nanotube based electrochemical biosensors: A  
619 review. *Electroanalysis: An International Journal Devoted to Fundamental and*  
620 *Practical Aspects of Electroanalysis*, 17(1), 7-14.
- 621 [36] Chen, K. J., Lee, C. F., Rick, J., Wang, S. H., Liu, C. C., & Hwang, B. J. (2012).  
622 Fabrication and application of amperometric glucose biosensor based on a novel  
623 PtPd bimetallic nanoparticle decorated multi-walled carbon nanotube  
624 catalyst. *Biosensors and Bioelectronics*, 33(1), 75-81.
- 625 [37] Lin, K. C., Lin, Y. C., & Chen, S. M. (2013). A highly sensitive nonenzymatic glucose  
626 sensor based on multi-walled carbon nanotubes decorated with nickel and copper  
627 nanoparticles. *Electrochimica Acta*, 96, 164-172.
- 628 [38] Gougis, M., Tabet-Aoul, A., Ma, D., & Mohamedi, M. (2014). Laser synthesis and  
629 tailor-design of nanosized gold onto carbon nanotubes for non-enzymatic  
630 electrochemical glucose sensor. *Sensors and Actuators B: Chemical*, 193, 363-369.

- 631 [39] Baghayeri, M., Amiri, A., & Farhadi, S. (2016). Development of non-enzymatic  
632 glucose sensor based on efficient loading Ag nanoparticles on functionalized carbon  
633 nanotubes. *Sensors and Actuators B: Chemical*, 225, 354-362.
- 634 [40] Lerner, M. B., Kybert, N., Mendoza, R., Villechenon, R., Bonilla Lopez, M. A., &  
635 Charlie Johnson, A. T. (2013). Scalable, non-invasive glucose sensor based on  
636 boronic acid functionalized carbon nanotube transistors. *Applied Physics  
637 Letters*, 102(18), 183113.
- 638 [41] Fu, D., & Li, L. J. (2010). Label-free electrical detection of DNA hybridization using  
639 carbon nanotubes and graphene. *Nano Reviews*, 1(1), 5354.
- 640 [42] Balasubramanian, K., & Burghard, M. (2006). Biosensors based on carbon  
641 nanotubes. *Analytical and bioanalytical chemistry*, 385(3), 452-468.
- 642 [43] Star, A., Tu, E., Niemann, J., Gabriel, J. C. P., Joiner, C. S., & Valcke, C. (2006).  
643 Label-free detection of DNA hybridization using carbon nanotube network field-effect  
644 transistors. *Proceedings of the National Academy of Sciences*, 103(4), 921-926.
- 645 [44] Dong, X., Lau, C. M., Lohani, A., Mhaisalkar, S. G., Kasim, J., Shen, Z., ... & Li, L. J.  
646 (2008). Electrical Detection of Femtomolar DNA via Gold-Nanoparticle Enhancement  
647 in Carbon-Nanotube-Network Field-Effect Transistors. *Advanced Materials*, 20(12),  
648 2389-2393.
- 649 [45] Guo, X., Gorodetsky, A. A., Hone, J., Barton, J. K., & Nuckolls, C. (2008).  
650 Conductivity of a single DNA duplex bridging a carbon nanotube gap. *Nature  
651 nanotechnology*, 3(3), 163-167.
- 652 [46] Weizmann, Y., Chenoweth, D. M., & Swager, T. M. (2011). DNA- cnt nanowire  
653 networks for DNA detection. *Journal of the American Chemical Society*, 133(10),  
654 3238-3241.
- 655 [47] Bayram, E., & Akyilmaz, E. (2016). Development of a new microbial biosensor based  
656 on conductive polymer/multiwalled carbon nanotube and its application to  
657 paracetamol determination. *Sensors and Actuators B: Chemical*, 233, 409-418.
- 658 [48] Palomäki, T., Peltola, E., Sainio, S., Wester, N., Pitkänen, O., Kordas, K., ... & Laurila,  
659 T. (2018). Unmodified and multi-walled carbon nanotube modified tetrahedral  
660 amorphous carbon (ta-C) films as in vivo sensor materials for sensitive and selective  
661 detection of dopamine. *Biosensors and Bioelectronics*, 118, 23-30.

- 662 [49] Chen, X., Chen, J., Deng, C., Xiao, C., Yang, Y., Nie, Z., & Yao, S. (2008).  
663 Amperometric glucose biosensor based on boron-doped carbon nanotubes modified  
664 electrode. *Talanta*, 76(4), 763-767.
- 665 [50] Gautam, V., Singh, K. P., & Yadav, V. L. (2018). Polyaniline/multiwall carbon  
666 nanotubes/starch nanocomposite material and hemoglobin modified carbon paste  
667 electrode for hydrogen peroxide and glucose biosensing. *International journal of*  
668 *biological macromolecules*, 111, 1124-1132.
- 669 [51] Dervisevic, M., Dervisevic, E., & Şenel, M. (2018). Design of amperometric urea  
670 biosensor based on self-assembled monolayer of cystamine/PAMAM-grafted  
671 MWCNT/Urease. *Sensors and Actuators B: Chemical*, 254, 93-101.
- 672 [52] Wasik, D., Mulchandani, A., & Yates, M. V. (2017). A heparin-functionalized carbon  
673 nanotube-based affinity biosensor for dengue virus. *Biosensors and*  
674 *Bioelectronics*, 91, 811-816.
- 675 [53] Tran, T. L., Nguyen, T. T., Tran, T. T. H., Tran, Q. T., & Mai, A. T. (2017). Detection  
676 of influenza A virus using carbon nanotubes field effect transistor based DNA  
677 sensor. *Physica E: Low-dimensional Systems and Nanostructures*, 93, 83-86.
- 678 [54] Fu, Y., Romay, V., Liu, Y., Ibarlucea, B., Baraban, L., Khavrus, V., ... & Gemming, T.  
679 (2017). Chemiresistive biosensors based on carbon nanotubes for label-free  
680 detection of DNA sequences derived from avian influenza virus H5N1. *Sensors and*  
681 *Actuators B: Chemical*, 249, 691-699.
- 682 [55] Dias, A. C. M., Gomes-Filho, S. L., Silva, M. M., & Dutra, R. F. (2013). A sensor tip  
683 based on carbon nanotube-ink printed electrode for the dengue virus NS1  
684 protein. *Biosensors and Bioelectronics*, 44, 216-221.
- 685 [56] Radogna, F., & Diederich, M. (2018). Stress-induced cellular responses in  
686 immunogenic cell death: implications for cancer immunotherapy. *Biochemical*  
687 *pharmacology*, 153, 12-23.
- 688 [57] Zanganeh, S., Khodadadei, F., Tafti, S. R., & Abdolahad, M. (2016). Folic acid  
689 functionalized vertically aligned carbon nanotube (FA-VACNT) electrodes for cancer  
690 sensing applications. *Journal of Materials Science & Technology*, 32(7), 617-625.

- 691 [58] Zhang, L. P., Tan, X. X., Huang, Y. P., & Liu, Z. S. (2018). Floating liquid crystalline  
692 molecularly imprinted polymer coated carbon nanotubes for levofloxacin  
693 delivery. *European Journal of Pharmaceutics and Biopharmaceutics*, 127, 150-158.
- 694 [59] Ji, S., Lee, M., & Kim, D. (2018). Detection of early stage prostate cancer by using a  
695 simple carbon nanotube@ paper biosensor. *Biosensors and Bioelectronics*, 102,  
696 345-350.
- 697 [60] Ding, N., Dou, C., Wang, Y., Liu, F., Guan, G., Huo, D., ... & Tan, J. (2018). Antishear  
698 Stress Bionic Carbon Nanotube Mesh Coating with Intracellular Controlled Drug  
699 Delivery Constructing Small-Diameter Tissue-Engineered Vascular  
700 Grafts. *Advanced Healthcare Materials*, 7(11), 1800026.
- 701 [61] Zhou, H., Ran, G., Masson, J. F., Wang, C., Zhao, Y., & Song, Q. (2018). Novel  
702 tungsten phosphide embedded nitrogen-doped carbon nanotubes: A portable and  
703 renewable monitoring platform for anticancer drug in whole blood. *Biosensors and*  
704 *Bioelectronics*, 105, 226-235.
- 705 [62] Yun, Y. H., Lee, B. K., & Park, K. (2015). Controlled Drug Delivery: Historical  
706 perspective for the next generation. *Journal of Controlled Release*, 219, 2-7.
- 707 [63] Kostarelos, K., Lacerda, L., Pastorin, G., & Wu, W. (2007). WieckowskiSebastien, J.  
708 Luangsivilay, S. Godefroy, D. Pantarotto, J.-P. Briand, S. Muller, M. Prato and A.  
709 Bianco. *Nat. Nanotechnol*, 2, 108-113.
- 710 [64] Pippa, N., Chronopoulos, D. D., Stellas, D., Fernández-Pacheco, R., Arenal, R.,  
711 Demetzos, C., & Tagmatarchis, N. (2017). Design and development of multi-walled  
712 carbon nanotube-liposome drug delivery platforms. *International Journal of*  
713 *Pharmaceutics*, 528(1-2), 429-439.
- 714 [65] Liu, Z., Robinson, J. T., Tabakman, S. M., Yang, K., & Dai, H. (2011). Carbon  
715 materials for drug delivery & cancer therapy. *Materials today*, 14(7-8), 316-323.
- 716 [66] Taghavi, S., Nia, A. H., Abnous, K., & Ramezani, M. (2017). Polyethylenimine-  
717 functionalized carbon nanotubes tagged with AS1411 aptamer for combination gene  
718 and drug delivery into human gastric cancer cells. *International journal of*  
719 *pharmaceutics*, 516(1-2), 301-312.
- 720 [67] Jung, K. W., Won, Y. J., Kong, H. J., Oh, C. M., Cho, H., Lee, D. H., & Lee, K. H.  
721 (2015). Cancer statistics in Korea: incidence, mortality, survival, and prevalence in

722 2012. *Cancer research and treatment: official journal of Korean Cancer*  
723 *Association*, 47(2), 127.

724 [68] Cronin, K. A., Lake, A. J., Scott, S., Sherman, R. L., Noone, A. M., Howlader, N., ...  
725 & Kohler, B. A. (2018). Annual Report to the Nation on the Status of Cancer, part I:  
726 National cancer statistics. *Cancer*, 124(13), 2785-2800.

727 [69] Wang, N., Feng, Y., Zeng, L., Zhao, Z., & Chen, T. (2015). Functionalized multiwalled  
728 carbon nanotubes as carriers of ruthenium complexes to antagonize cancer multidrug  
729 resistance and radioresistance. *ACS applied materials & interfaces*, 7(27), 14933-  
730 14945.

731 [70] Wei, C., Dong, X., Zhang, Y., Liang, J., Yang, A., Zhu, D., ... & Lv, F. (2018).  
732 Simultaneous fluorescence imaging monitoring of the programmed release of dual  
733 drugs from a hydrogel-carbon nanotube delivery system. *Sensors and Actuators B:*  
734 *Chemical*, 273, 264-275.

735 [71] Xu, H., Liu, M., Lan, M., Yuan, H., Yu, W., Tian, J., ... & Wei, Y. (2016). Mussel-  
736 inspired PEGylated carbon nanotubes: biocompatibility evaluation and drug delivery  
737 applications. *Toxicology research*, 5(5), 1371-1379.

738 [72] Dong, X., Wei, C., Liang, J., Liu, T., Kong, D., & Lv, F. (2017). Thermosensitive  
739 hydrogel loaded with chitosan-carbon nanotubes for near infrared light triggered drug  
740 delivery. *Colloids and Surfaces B: Biointerfaces*, 154, 253-262.

741 [73] Sciortino, N., Fedeli, S., Paoli, P., Brandi, A., Chiarugi, P., Severi, M., & Cicchi, S.  
742 (2017). Multiwalled carbon nanotubes for drug delivery: Efficiency related to length  
743 and incubation time. *International journal of pharmaceutics*, 521(1-2), 69-72.

744 [74] Liu, X., Xu, D., Liao, C., Fang, Y., & Guo, B. (2018). Development of a promising  
745 drug delivery for formononetin: Cyclodextrin-modified single-walled carbon  
746 nanotubes. *Journal of Drug Delivery Science and Technology*, 43, 461-468.

747 [75] Razzazan, A., Atyabi, F., Kazemi, B., & Dinarvand, R. (2016). In vivo drug delivery  
748 of gemcitabine with PEGylated single-walled carbon nanotubes. *Materials Science*  
749 *and Engineering: C*, 62, 614-625.

750 [76] Schwengber, A., Prado, H. J., Bonelli, P. R., & Cukierman, A. L. (2017). Development  
751 and in vitro evaluation of potential electromodulated transdermal drug delivery

752 systems based on carbon nanotube buckypapers. *Materials Science and*  
753 *Engineering: C*, 76, 431-438.

754

755 **Figure captions**

756

757 **Figure 1** SWCNTs based sensing arrays having coating of organic materials as lung  
758 cancer biomarkers. a) patterns of responses of chemiresistive networks of SWCNTs; b)  
759 PCA score plots of the 10 sensing arrays. [21].

760 **Figure 2** MWCNTs for discrimination of 20 representative VOCs. a) Chemical structures,  
761 b) Patterns of changes, c) Principal component score plots. [27].

762 **Figure 3** a) Chemical structures of xylene isomers and poly(3-hexylthiophene) (P3HT),  
763 b) SWCNTs, c) SWCNT and P3HT to p-xylene, o-xylene, and m-xylene at 400 ppm. [32].

764 **Figure 4** Nonenzymatic glucose sensor based on Ag nanoparticles (NPs) on  
765 functionalized CNTs. [39].

766 **Figure 5** Boronic acid-functionalized CNT-based FET sensor for the detection of glucose.  
767 [40].

768 **Figure 6** Detection of femtomolar DNA using Au NPs to enhance SWCNT-based FET  
769 sensors. [44].

770 **Figure 7** Conductivity of a single DNA duplex bridged between SWCNT. [45].

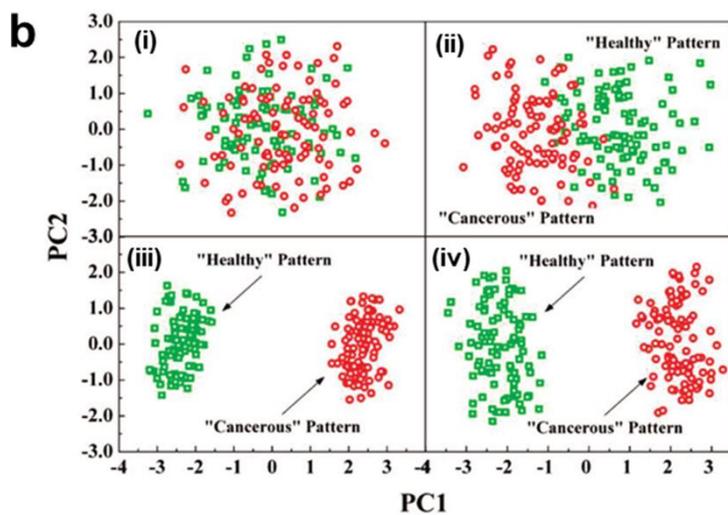
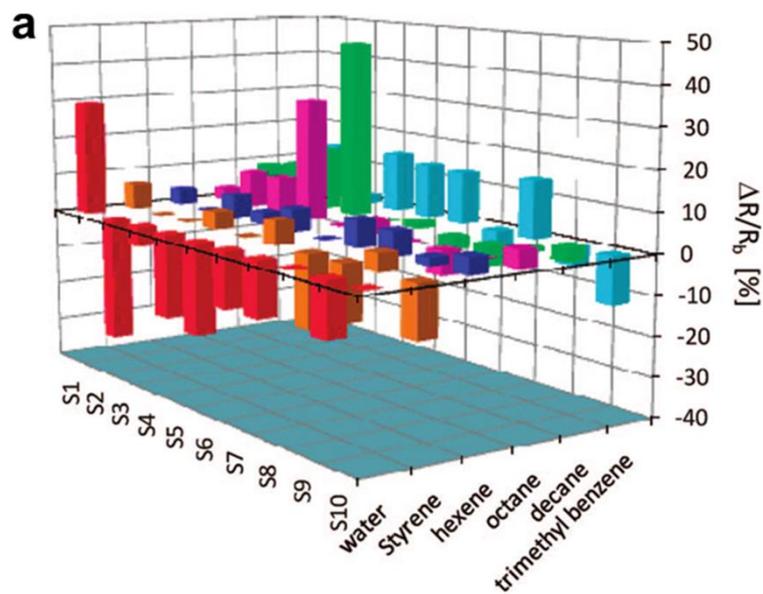
771 **Figure 8** CNT-network-based DNA detection scheme. [46].

772

773 List of Figures

774

775



776

777

778

779

780

781

782

783

Figure 1

784

785

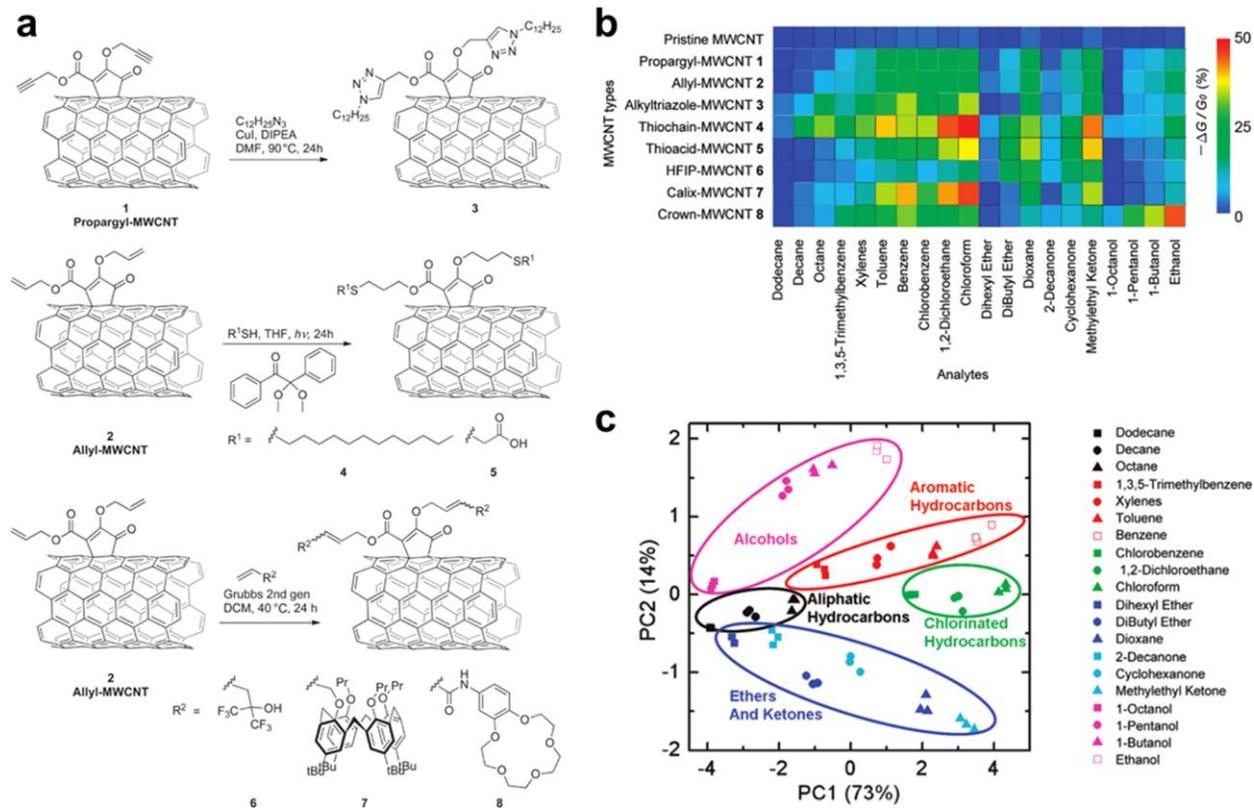


Figure 2

786

787

788

789

790

791

792

793

794

795

796

797

798

799

800

801  
802

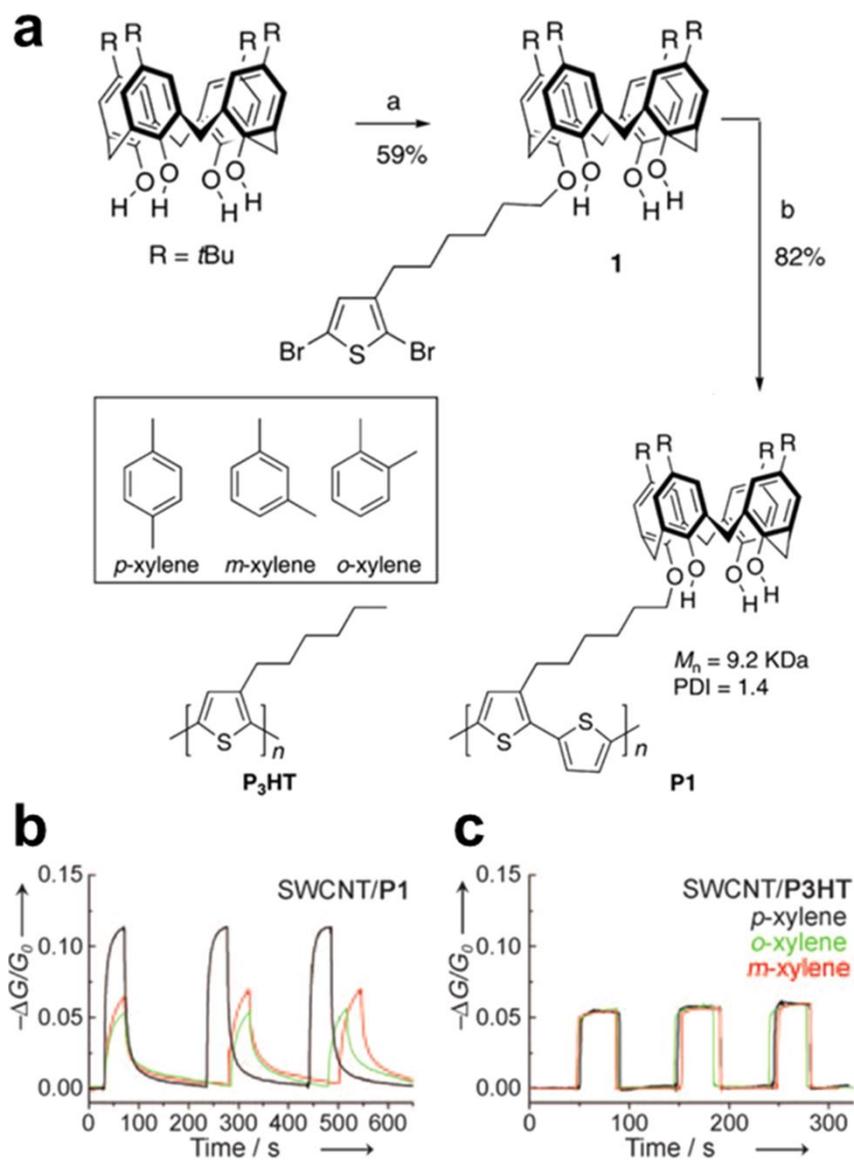


Figure 3

803  
804  
805  
806  
807  
808  
809  
810  
811

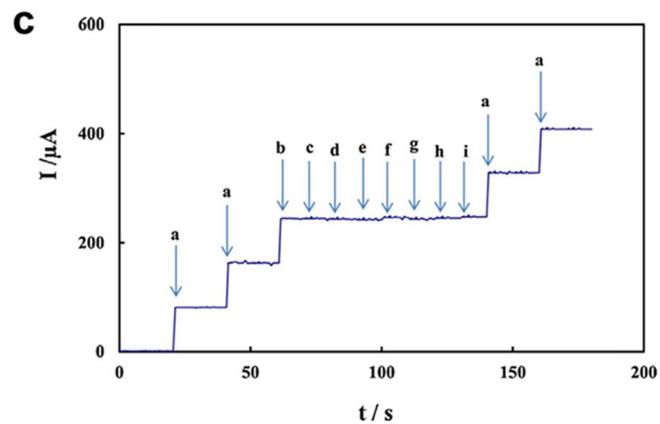
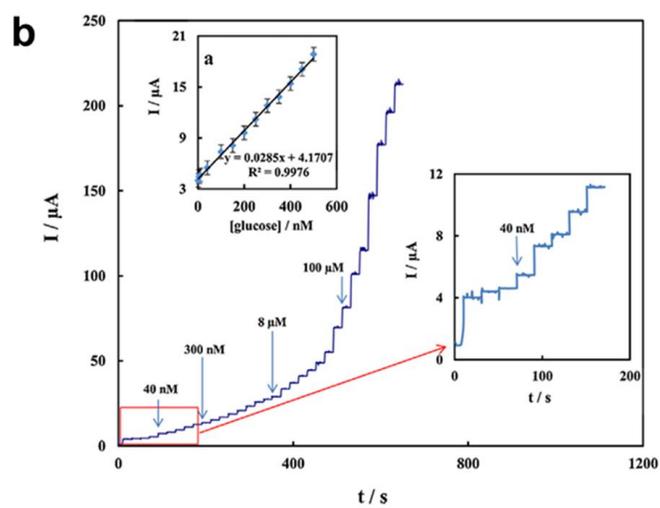
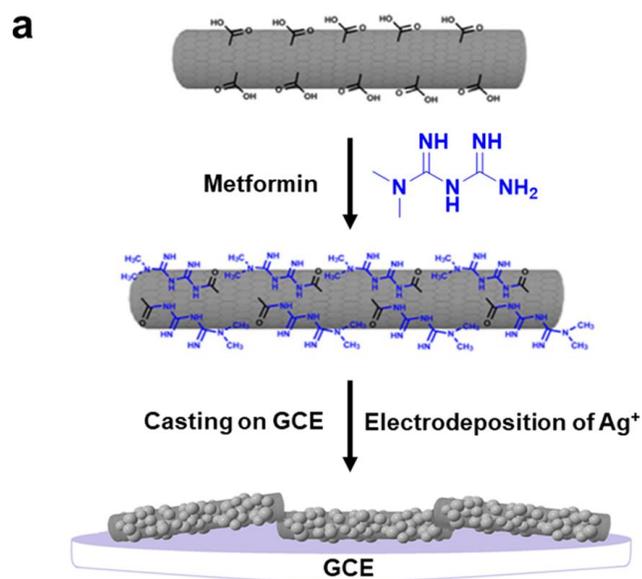


Figure 4

816

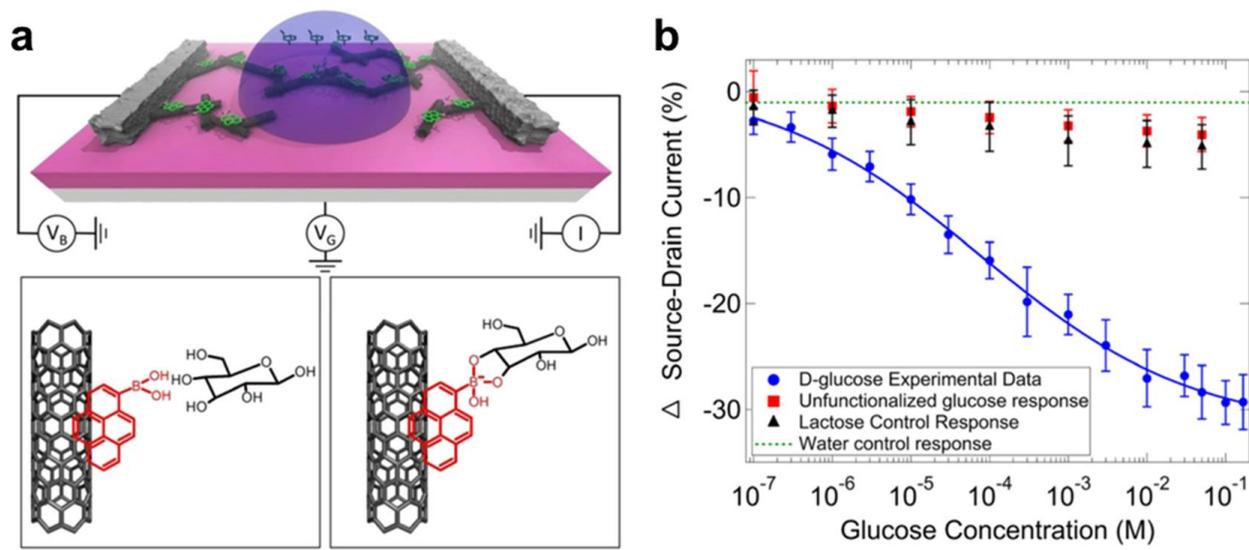


Figure 5

817

818

819

820

821

822

823

824

825

826

827

828

829

830

831

832

833

834

835

836

837

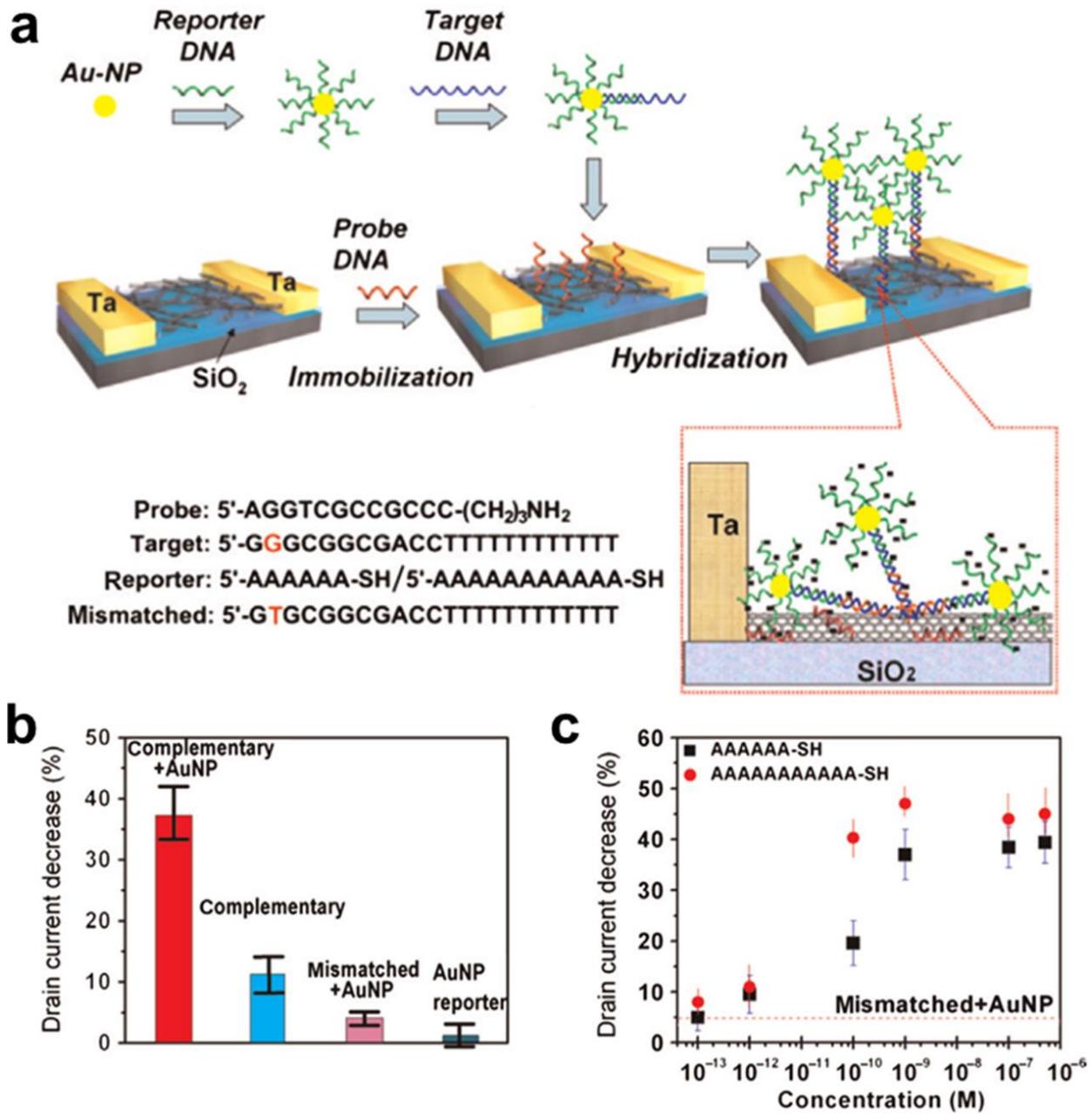


Figure 6

839

840

841

842

843

844

845

846

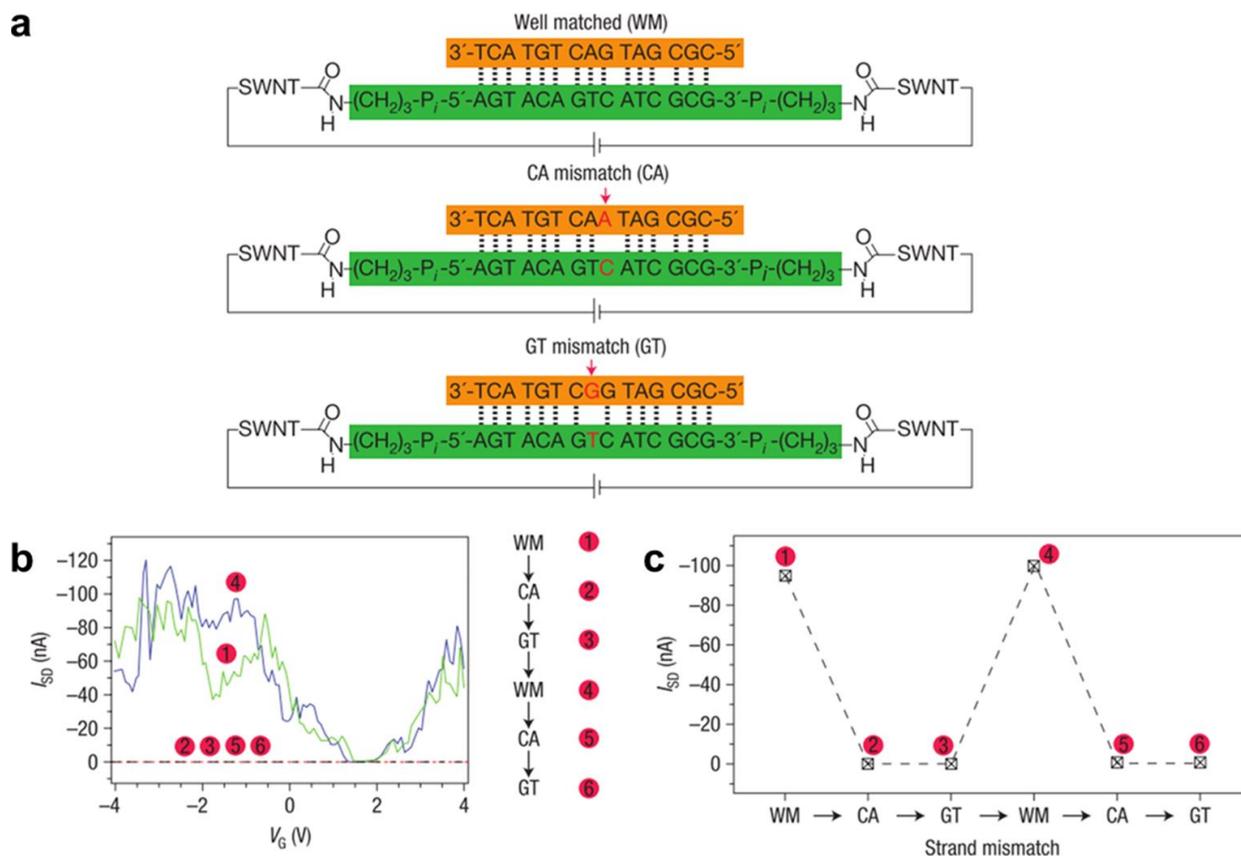


Figure 7

848

849

850

851

852

853

854

855

856

857

858

859

860

861

862

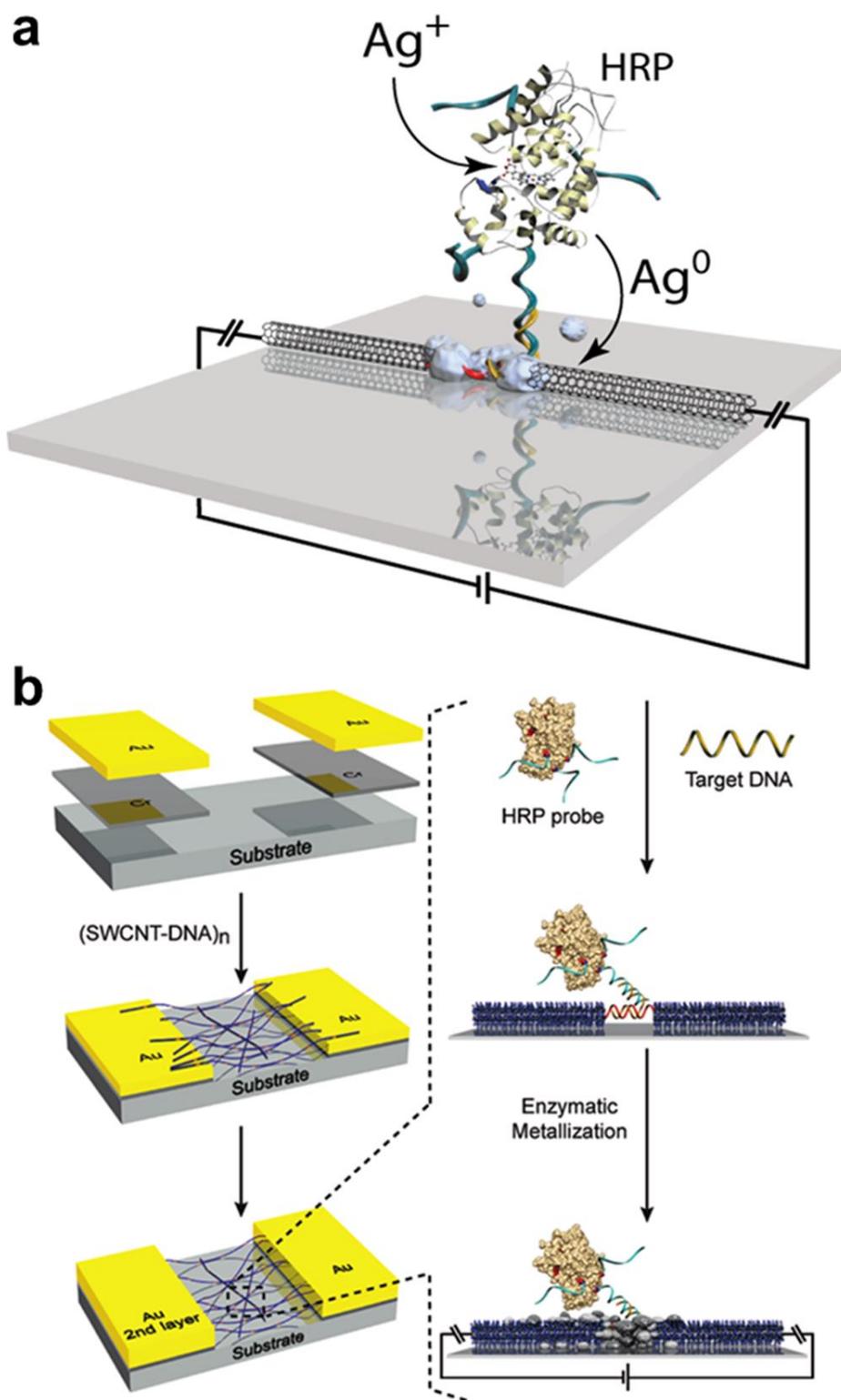


Figure 8