

Prioritization of Candidate Genes for the Effect of *Fob3b1* QTL on Chromosome 15 in Mouse Models for Polygenic Obesity and Leanness using Integrative Genomics

Key words

data integration;
gene expression;
gene prioritisation;
mouse models;
obesity;
QTL;
single nucleotide
polymorphism

Martin Šimon^{1**}, Tanja Kunej^{1#}, Nicholas M. Morton², Simon Horvat^{1*}

¹University of Ljubljana, Biotechnical Faculty, Department of Animal Science, Domžale 1230, Slovenija,

²Nottingham Trent University, Schools of Science and Technology, Department of Biosciences, Nottingham, United Kingdom, #Authors contributed equally to the study

*Corresponding authors: martin.simon@bf.uni-lj.si, simon.horvat@bf.uni-lj.si

Abstract: The accumulation of excess fat affects meat quality, fertility, productivity, and whole-body metabolism in farm animals. The mouse model presents an efficient tool for investigating these traits. Previous QTL analyses of the unique mouse selection lines for polygenic obesity (Fat line) and leanness (Lean line) have revealed four major obesity QTLs: *Fob1*, *Fob2*, *Fob3*, and *Fob4*. *Fob3*, located on chromosome 15, was later subdivided into *Fob3a* and *Fob3b*, which additionally split into *Fob3b1* and *Fob3b2*. Of the 158 genes annotated in *Fob3b1*, 16 candidate genes have been previously proposed for the QTL effects. However, genomic variability between the Fat and Lean lines at this locus has not been fully investigated. The present study aimed to validate previously identified candidates and to identify novel candidate genes potentially responsible for the *Fob3b1* effect. Data from whole-genome sequencing and transcriptome analyses of Fat and Lean mouse lines were integrated with obesity QTLs in cattle and pigs from Animal QTLdb and phenotypes obtained from the International Mouse Phenotyping Consortium (IMPC) and the Mouse Genome Database (MGD). Out of 158 genes located in the *Fob3b1* interval we prioritized 17 candidate genes, including six previously proposed (*Adgrb1*, *Col22a1*, *Cyp11b1*, *Dgat1*, *Gpihbp1* and *Ly6a*) and 11 novel candidates: *9030619P08Rik*, *Eppk1*, *Kcnk9*, *Ly6c1*, *Ly6d*, *Ly6h*, *Ly6i*, *Ly6m*, *Ptk2*, *Trappc9*, and a strong candidate *Ly6e* that deserve further functional analyses. Biological function and literature screening for candidate genes suggest that the *Fob3b1*'s impact on obesity may operate through triglyceride metabolism (*Dgat1* and *Gpihbp1*) and cytoskeletal and extracellular matrix remodelling (*Ly6a*, *Ly6e* and *Eppk1*). Further fine mapping, genetic and "omic" studies should clarify whether the *Fob3b1* effect is due to a causal genetic variant in one of the candidates or possibly due to an additive effect of a combination of these positional candidates. The applied bioinformatics approach in determining the priority of candidate genes for obesity can also serve as a model for other traits in veterinary and livestock sciences.

Received: 5 March 2024

Accepted: 16 May 2024

Introduction

Obesity, considered by many to be the epidemic of the 21st century, is broadly divided into two categories: the monogenic type and the more common polygenic type (1,2). Obesity leads to the development of metabolic disorders

such as diabetes mellitus, high blood pressure, cardiovascular diseases, and inflammation-related diseases (3). The accumulation of excess fat also affects meat quality, fertility, productivity, and whole-body metabolism in farm animals

and is also one of the most important health and welfare issues affecting companion animals (4). Rodent models such as mice and rats serve as invaluable tools for studying the complex biology of obesity, identifying new therapeutic targets, and evaluating the efficacy and safety of potential interventions (5). There are few mouse models for the polygenic type of obesity, but they have no lean counterparts derived from the same base population. Selective breeding for desired divergent phenotypes over an extended period creates novel, polygenic, and reproducible disease models (6,7). One such mouse model was developed by divergent selection on body fat percentage over more than 60 generations, resulting in the Fat and Lean lines, which differ in fatness by a factor of five (8).

Earlier genome-wide quantitative trait locus (QTL) analyses of the two selection lines revealed four major obesity QTLs (*Fob1*, *Fob2*, *Fob3*, and *Fob4*) (9). Further experimental data showed that the QTL interval with the highest LOD (logarithm of the odds) score, *Fob3* on chromosome 15, consists of two linked QTLs with smaller effects, *Fob3a* and *Fob3b* (10), which additionally split into *Fob3b1*, with a stronger effect, and *Fob3b2* (11). Sixteen candidate genes have been proposed for the *Fob3b1* effect based on previously identified obesity QTLs in mice and cattle, gene expression analyses obtained from the expression database, and based on their known biological functions (11). However, the genomic variability and differential gene expression between the Fat and Lean lines at this locus, which could significantly improve the prioritisation power for candidate genes, have not yet been fully investigated.

In the present study, integration of whole genome sequencing (WGS) focusing on single nucleotide polymorphisms (SNPs) and gene expression data of genes within *Fob3b1* in white adipose tissue of the Fat and Lean lines were performed to prioritise candidate genes responsible for the *Fob3b1* effect. In addition, candidates were complemented with their relatedness to obesity using gene and gene knock-out annotations and a comparative genomics approach between mouse, pigs, and cattle.

Material and methods

Whole genome sequencing (WGS) data of Fat and Lean mice were from our previous studies (12) (13). Differential gene expression data for three white adipose tissues (WAT) depots (epididymal WAT, subcutaneous WAT, mesenteric WAT) were from (14). The expression data from the three tissues were then joined and corrected for the batch effect using Empirical Bayes Analysis to obtain expression data in WAT. Gene expression was considered differential if expression differed between Fat and Lean mouse lines by at least 1.5-fold. Significance was checked at both $p < 0.05$ and adjusted $p < 0.05$ (differentially expressed genes; DEGs).

Two criteria were used for the candidate gene prioritisation for the *Fob3b1* effect: 1.) genes carried line-specific SNPs within coding regions (exons) according to the Ensembl Variant Effect Predictor (<https://www.ensembl.org/Tools/VEP>) (15) or 2.) genes were differentially expressed in WAT between the Fat and Lean mouse lines. The results were complemented with annotations related to obesity by the International Mouse Phenotyping Consortium (IMPC, www.mousephenotype.org) (16) using the search term "abnormal adipose tissue amount" and the Mouse Genome Database (MGD; <http://www.informatics.jax.org>) (17) using the search term "fat" in the terms for mammalian phenotypes. In addition, previous gene associations with obesity were extracted by literature screening using the Pubmed database and MeSH Terms *adipog**, *obes**, *fat*, *lipid droplet* and approved gene symbols and synonyms. The candidate genes were supplemented with orthologous genes within obesity-related QTLs in cattle and pigs, obtained using Animal QTLdb (<https://www.animalgenome.org/cgi-bin/QTLdb/>, Release 50, April 25, 2023) (18), Ensembl (19), and g:Profiler (20). First, the locations of QTLs related to subcutaneous fat/adipose thickness/amount were obtained from Animal QTLdb. Second, genes within these QTLs were identified using the Ensembl Biomart. Finally, orthologous genes in mice were obtained from g:Profiler. The genomic location of *Fob3b1* was obtained by converting genomic coordinates provided by (11) (71.38– 76.36 Mbp, mouse NCBI36 assembly) to genome assembly GRCm38 using UCSC Genome Browser tool liftOver (<https://genome.ucsc.edu/cgi-bin/hgLiftOver>). Location of regulatory elements (open chromatin, enhancer, promoter, promoter flanking region, or CTCF binding site) was obtained from Ensembl database.

Results

The present study aimed to prioritize genes responsible for the *Fob3b1* effect in mouse models for polygenic obesity and leanness. Two criteria were used for the candidate gene prioritisation: 1.) line-specific SNPs in coding regions or 2.) differential gene expression in WAT between the Fat and Lean mouse lines. The workflow with the main results is shown in Figure 1.

The *Fob3b1* interval, spanning from 15:71,550,331-76,532,745, contains 158 genes (GRCm38) of which 67 genes carry line-specific SNPs, and seven were differentially expressed in WAT between the Fat and Lean lines. By prioritization of 158 genes, we obtained 17 promising candidates: 10 genes with SNPs in coding regions (seven genes with missense variants, three genes with synonymous variants), six genes with differential expression, and *Ly6e* with both synonymous exonic variants in the Lean line and differential expression (Table 1). We were also interested if differential expression might be caused by potential regulatory variants (SNPs located within open chromatin, enhancer, promoter, promoter flanking region, or CTCF

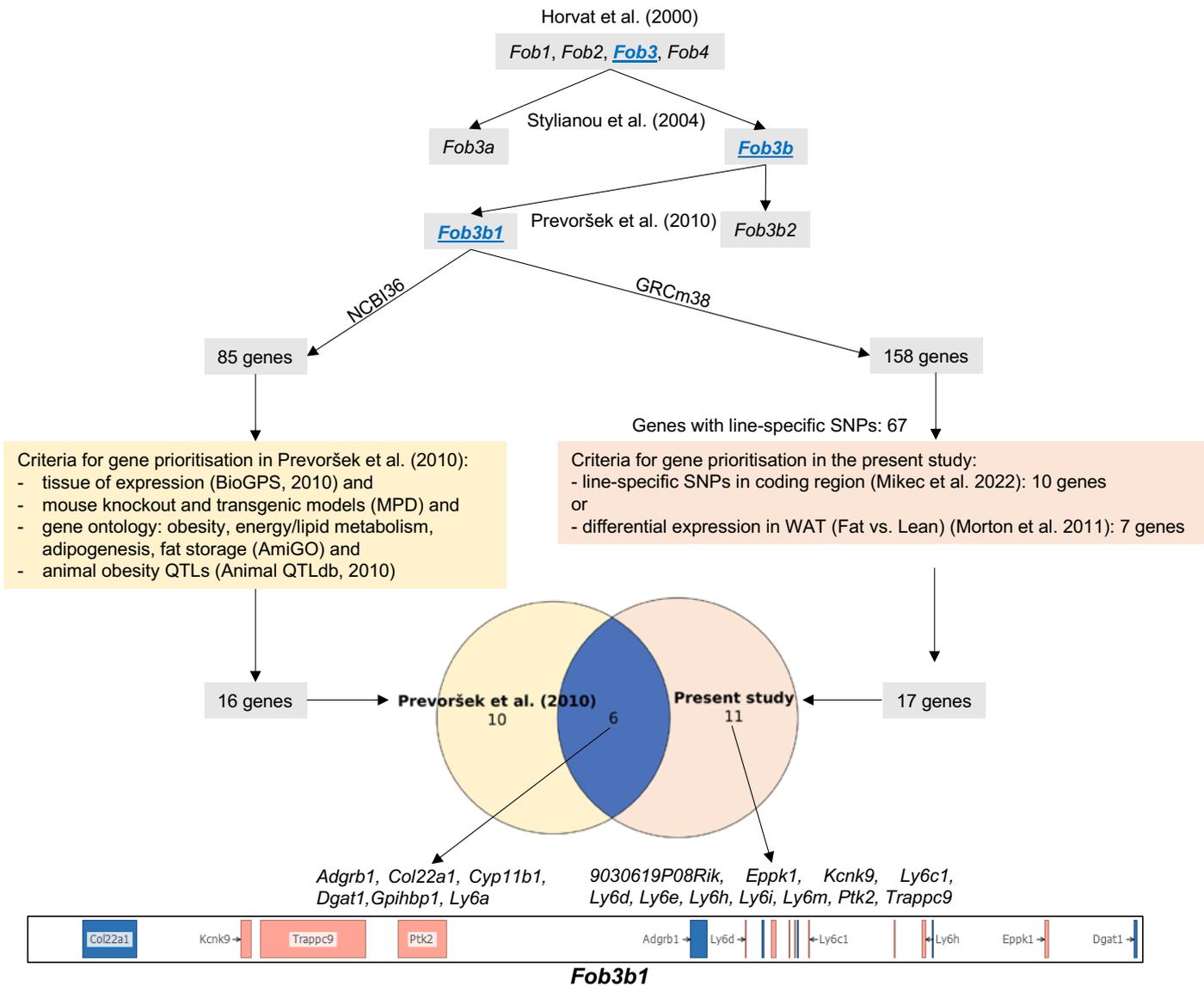


Figure 1: The workflow of the study for the prioritization of candidate genes responsible for the Fob3b1 effect

binding site). Among the DEGs, two genes, both in the Lean line, carry potentially regulatory variants that may affect their expression. In *9030619P08Rik*, the SNP rs31762288 is located within the open chromatin region and rs13482652 and rs32099107 are within promoter flanking region. As for *Ly6e*, all 42 potentially regulatory variants are within promoter flanking region. Out of 17 QTL prioritized candidates six were proposed previously (*Adgrb1, Col22a1, Cyp11b1, Dgat1, Gpihbp1, and Ly6a*) (11), while the *9030619P08Rik, Eppk1, Kcnk9, Ly6c1, Ly6d, Ly6e, Ly6i, Ly6h, Ly6m, Ptk2, and Trappc9* are newly proposed candidate genes. Some of the 17 candidate genes have been previously associated with obesity, but genes, such as *Col22a1, Eppk1, Ly6i, and Ly6m* have been proposed to be associated with obesity for the first time. The comparative genomics approach revealed 14 orthologous genes located within the obesity-related QTLs in cattle, however, none of them is located within the obesity QTLs in pigs (Table 1).

Discussion

In the present investigation, we undertook a comprehensive analysis by integrating whole genome sequencing (WGS) and transcriptomics data from the Fat and Lean mouse lines to systematically prioritize candidate genes responsible for the observed effects of *Fob3b1*. Our specific emphasis was directed toward SNPs in coding regions and the gene expression profiles of genes within the *Fob3b1* locus in WAT. For the SNPs in coding regions, we also included synonymous variants as accumulating experimental evidence has demonstrated that they may exert their impact on gene functions via splicing accuracy, mRNA stability, translation fidelity, protein folding, and expression (21).

As many as seven out of the candidate genes in the present study are part of the LY6 (lymphocyte antigen 6 complex) family of proteins involved in a variety of functions in cell proliferation, migration, cell-cell interaction, immune cell

Table 1: 27 positional candidate genes for *Fob3b1* effect; 16 from the study Prevoršek et al. (2010) (8), 17 from the present study (marked in bold), including six genes identified by both studies

Priority	Gene	SNPs ¹	Criteria 1: Line specific SNPs in coding region	SNP located within regulatory region	Criteria 2: DEG2	IMPC ³	MGI ³	Cattle QTL ⁴	Literature associated with obesity
high	Dgat1	B:17		B:2	↑		✓		✓
	Gpihbp1	/			↑			✓	✓
	<i>Rhpn1</i>	B:1						✓	
	Ly6a	L:56	L:3	L:9			✓		✓
moderate	Cyp11b1	L:27, F:1	L:1				✓	✓	✓
	<i>Cyp11b2</i>	L:47		L:2				✓	✓
	<i>Gpr20</i>	/						✓	✓
	Adgrb1	L:1, F:1	L:1	L:1				✓	
	<i>Tsta3</i>	/							
	<i>Arc</i>	/						✓	✓
low	<i>Psca</i>	/						✓	
	<i>Ly6g2</i>	L:104		L:13					
	<i>Gsdmd</i>	/						✓	✓
	<i>Naprt1</i>	/						✓	✓
	<i>Cyc1</i>	B:1							✓
	Col22a1	L:388, F:499, B:283	L:2, F:8, B:4	L:17, F:21, B:12					
Present study	Ly6e	L:106	L:2	L:42	↑			✓	
	Trappc9	L:1144, F:3, B:3	L:8	L:197, B:2		✓	✓	✓	✓
	9030619P08Rik	L:22		L:3	↑				
	Ly6d	/			↑			✓	
	Ly6h	/			↑			✓	
	Eppk1	/			↑				
	Kcnk9	L:13	L:1	L:4					✓
	Ly6c1	L:49	L:1						✓
	Ly6i	L:148	L:3	L:7					
	Ptk2	F:2	F:1	F:1					✓
Ly6m	L:65, B:2	L:3	L:7						

¹SNPs identified in both (B), Fat (F) or Lean (L) lines, ²Differentially expressed gene (Fat vs. Lean); ↑: upregulated, ³Associated with obesity-related traits in IMPC and MGI databases, ⁴Orthologous genes in obesity-related QTL in cattle obtained from Animal QTLdb

maturation, macrophage activation, and cytokine production, mainly by regulating acetylcholine signalling (22) that has been recently linked to insulin sensitivity, low-grade inflammation, adipose dysfunction and metabolic syndrome in obesity (23,24). While four of them (*Ly6a*, *Ly6c1*, *Ly6i*, *Ly6m*) carry exonic variants in the Lean line, the remaining three (*Ly6d*, *Ly6e*, and *Ly6h*) were found to be expressed to a higher level in WAT of the Fat line compared to the Lean line. In addition, higher expression of an uncharacterized *9030619P08Rik*, described as an LY6 pseudogene (25), and *Gpihbp1*, a member of the LY6 superfamily (26), was determined in the Fat line WAT. The expression of *Ly6d*, *Ly6h*, and *Gpihbp1* did not depend on regulatory SNPs in our study, suggesting that there may be genetic variations in the transcriptional regulators of these three genes located elsewhere in the genome. Meanwhile, *Ly6e* and *9030619P08Rik* in the Lean line carry potential regulatory variants that may explain their higher expression levels in the Fat line.

Among these genes, *Ly6ci*, *Ly6a*, and *Ly6e* are especially worth mentioning. While *Ly6ci* was linked to abnormal metabolic pathways in the early induction phase of autoimmune diabetes (27), altering T cell function (28), *LY6A* and *LY6E* were, in addition to their involvement in immunity (29,30) also linked to extracellular matrix remodelling (31,32). In adipose tissue of obese individuals, remodelling of the extracellular matrix, cytoskeletal reorganisation and increased cell proliferation enable the enlargement of obese adipocytes and WAT expansion (33,34). The *Ly6a* is not differentially expressed, however, only the Lean line carries SNPs, including two missense variants rs213983347 (V/A) and rs32279213 (D/G), located in the same exon and within the protein domain Ly-6 antigen/uPA receptor-like, suggesting their effect on protein function. In cattle, *LY6A* has been associated with fertility, potentially by affecting growth dynamics in the unborn calf (35), and *LY6D* is crucial for lipid accumulation and inflammation in nonalcoholic fatty liver disease (36). Meanwhile, *Eppk1*, a new candidate gene with higher expression in the Fat line, is involved in cytoskeleton reorganization and cell proliferation (37). *Col22a1*, which encodes an extracellular matrix protein, is not differentially expressed but has several exonic variants in the Lean and Fat lines. *COL22A1* has been shown to increase intramuscular fat in cattle (38) and polymorphisms in porcine *COL22A1* were associated with daily weight gain (39).

Moreover, an uncharacterized *9030619P08Rik* is thought to be translated into a stable circulating microprotein that may be involved in metabolic regulation and obesity (25), and *Gpihbp1* regulates the lipolytic processing of triglyceride-rich lipoproteins (26). Nucleotide substitutions in *GPIHBP1* cause lifelong chylomicronemia (40). Lipolysis of triglyceride-rich particles leads to lower protective HDL cholesterol levels (41), which was previously observed in the Fat compared to the Lean line (42). Furthermore, changes in (high basal/low stimulated) lipolysis rates are associated with insulin resistance, previously demonstrated in the Fat line (43), and future weight gain in humans (44). Some

polymorphisms in porcine *GPIHBP1* were proposed to be genetic risk factors affecting adipose traits (45).

Among the high-priority candidates from a previous study (11) the expression of *Gpihbp1* and *Dgat1* was found to be higher in WAT of the Fat line, although the sequences in the two lines were identical. *DGAT1* catalyses the final step of triglyceride synthesis (46), and *Dgat1*-deficient mice are lean and resistant to diet-induced obesity (47). In addition, *DGAT1* was associated with a backfat thickness (48), fat deposition (49), and intra-muscular fat in pigs (50), and beef marbling (51). It was also identified as one of very few causative genes for milk yield and composition - fat content in cattle (52)

Other potential candidates include *Cyp11b1*, *Adgrb1*, *Kcnk9*, *Trappc9*, and *Ptk2*. Twenty-six of 27 line-specific SNPs were identified in the Lean line *Cyp11b1*, including a synonymous rs31832746. *CYP11B1* is a rate-limiting enzyme in the synthesis of cortisol (53), an obesity-related steroid hormone (54) whose formation selectively increases within adipose tissue in obesity (55). Even more promising candidate for QTL effect is *Adgrb1*, with a potentially deleterious variant rs51566550 in the Lean line. *Adgrb1*^{-/-} mice exhibited increased susceptibility to seizures, delayed growth, and reduced brain weight (56). *ADGRB1* is involved in a membrane-initiated pathway to induce the expression of *Abca1* (ATP-binding cassette, sub-family A (ABC1), member 1) in apoptotic cells (57) whose specific knockout in adipocytes resulted in significantly lower body weight, epididymal fat pad weight and adipocyte size due to changes in lipogenesis and lipid accretion in mice (58). Additionally, it is noteworthy that *ADGRB1* may play a role in sensory food perception (59), which alone can cause metabolic changes (60,61). Similarly, *Kcnk9* encodes TWIK-related acid-sensitive K channel 3 (TASK3) protein that has been implicated in glucose sensing (62). *Kcnk9* transcript was significantly up-regulated in mice nodose neurons fed a high-fat diet. The authors proposed it as a therapeutic target for obesity treatment (63). Adipose-specific knockout of a closely related gene *Kcnk3* in mice resulted in an increased energy expenditure and resistance to obesity (64). A SNP rs2471083 near the potassium channel *KCNK9* has a parent-of-origin effect on body mass index (65) and was linked to abdominal visceral fat by GWAS (66). Another candidate is *Trappc9*, with eight synonymous variants in *Trappc9* of the Lean line. This gene plays a role in energy balance, and its deficiency leads to obesity (67). It has been linked to fat deposition-related traits in Hu sheep (68) and to body size traits in pigs (69). *PTK2* (also known as focal adhesion kinase FAK), best known for its involvement in integrin signalling, was shown to influence adipocyte differentiation and to influence obesity in mice (70). In addition to its role in leptin signal transduction (71), FAK signalling controls insulin sensitivity through the regulation of adipocyte survival (72), and FAK inhibition causes insulin resistance (73). A novel missense variant 15_73264244_G/T in the Fat line may therefore

Table 2: Candidate orthologous genes associated with milk traits in cattle and pigs.

Gene/Region	Effect on milk	Species/breed	Reference
<i>ADGRB1</i>	urea content	Holstein cattle	Ma et al. (2023) (79)
<i>ADGRB1</i>	lactose content	Fleckvieh cattle	Costa et al. (2019) (75)
	yield		
<i>CYP11B1</i>	yield	German Holstein cattle	Kaupe et al. (2007) (35)
<i>DGAT1</i>	protein content	Polish landrace pigs	Szyndler-Nędza and Piorkowska (2015) (74)
	lactose content		
<i>GPIHBP1</i>	fat content	Romanian Holstein cattle	Tăbăran et al. (2015) (81)
	fat content	cattle	Yang et al. (2017) (80)
	protein content		Dong et al. (2020) (76)
<i>LY6E</i>	yield	Holstein cattle	Jiang et al. (2018) (77)
<i>TRAPPC9</i>	protein content	Chinese Holstein cattle	Khan et al. (2022) (78)
	mastitis resistance		

influence various signalling pathways and contribute to the obese phenotype.

However, the prioritised candidate genes may play other roles in tissues not examined in the present study. Interestingly, *DGAT1*, *GPIHBP1*, *CYP11B1*, *ADGRB1*, *LY6E*, and *TRAPPC9* are also associated with milk production and milk composition traits in cattle and pigs, such as milk urea, lactose, protein, and fat contents and milk yield (35,74–81) (Table 2). Importantly, recent metabolomic and proteomic investigations revealed a correlation between infant obesity and milk composition from obese or non-obese mothers (82,83). Considering *Cyp11b1*, *Adgrb1*, *Ly6e*, and *Trappc9*, the Lean line carries exonic variants that may affect the protein function, these genes may also affect milk composition and yield and subsequently contribute to the lean or obese phenotype in our mouse models.

In summary, *Fob3b1* may influence energy balance, inflammation, various signalling pathways (acetylcholine, leptin, insulin), metabolism, and cell structure in WAT, however, it may also contribute to the obese/lean phenotype by influencing milk quantity and composition. For the *Fob3b1* effect on the adiposity of WAT, we propose genes involved in triglyceride metabolism (*Dgat1* and *Gpihbp1*), cytoskeleton, and extracellular matrix remodelling (*Ly6a*, *Ly6e*, and *Eppk1*) as the main contributors, calling for their future functional analyses.

The control of fat deposition, energy metabolism, and immune system functioning have high economic importance in farm animals. Excess fat accumulation affects meat

quality, fertility, productivity, and whole-body metabolism (84). Further functional studies of the proposed candidate genes are required to elucidate their involvement in fat deposition.

Conclusions

The present study identified 17 candidate genes potentially responsible for the *Fob3b1* QTL effect in mouse models for polygenic obesity and leanness. In particular, triglyceride metabolism, cytoskeleton and extracellular matrix remodelling may be the main contributors to the effect of *Fob3b1*. Of the 17 most promising candidate genes, four new obesity candidates were proposed: *Col22a1*, *Eppk1*, *Ly6i*, and *Ly6m*. Further work on fine mapping and functional analyses is required to determine whether the effect of *Fob3b1* is due to a causal genetic variant in one of these candidates or a combined effect of several of these positional candidates. The applied bioinformatics approach for prioritization of candidate genes for polygenic obesity in the present study can also be used to analyze other traits in veterinary medicine and livestock science. Obesity and its associated diseases pose a significant health risk, affecting not only physical well-being, but also reproductive health and overall animal welfare. These effects extend beyond farm animals to include companion animals, highlighting the interconnectedness of veterinary and human medicine in addressing obesity-related health problems in all species to improve animal health and welfare.

Acknowledgements

This work was financially supported by the Slovenian Research and Innovation Agency (ARIS) research program P4-0220 and research project J4-2548.

Conflict of interest statement. All authors declare that they have no competing interests.

Authors' contributions. MŠ: Formal analysis, writing - original draft preparation. NMM: writing – review & editing. SH and TK: conceptualization, writing – review & editing, supervision.

Ethics approval and consent to participate. The FLI (Fat) and FHI (Lean) selection lines have been maintained in our laboratory for more than 100 generations. All mice used in this study were maintained according to local ethical and EU regulatory guidelines under the Veterinary Administration of Republic of Slovenia permit No. U34401-23/2020/6.

References

- Loos RJF, Yeo GSH. The genetics of obesity: from discovery to biology. *Nat Rev Genet* 2022 ;23(2): 120–33. doi: 10.1038/s41576-021-00414-z
- González-Muniesa P, Martínez-González MA, Hu FB, et al. Obesity. *Nat Rev Dis Primers* 2017;3(1): 17034. doi: 10.1038/nrdp.2017.34
- Alsufiani H, Aldhaheeri GA, Omar UM, Bahdilal TM, et al. Antioxidant activity and inhibitory effect of 2,4,4'-trihydroxychalcone on digestive enzymes related to obesity. *Slov Vet Res* 2023; 60 (suppl. 25): 363-74. doi: 10.26873/SVR-1625-2022
- Haddad KK. How Successful are veterinary weight management plans for canine patients experiencing poor welfare due to being overweight and obese? *Animals (Basel)*. 2024; 14(5): 740. doi: 10.3390/ani14050740
- Ulker Ertugrul N, Yardimci A, Kaya Tektemur N, et al. Effects of irisin on the reproductive system of obese female rats induced by a high-fat diet. *Slov Vet Res (ahead of print)* doi: 10.26873/SVR-1754-2023
- Palma-Vera SE, Reyer H, Langhammer M, et al. Genomic characterization of the world's longest selection experiment in mouse reveals the complexity of polygenic traits. *BMC Biol* 2022; 20(1): 52. doi: 10.1186/s12915-022-01248-9
- Saul MC, Philip VM, Reinholdt LG, Chesler EJ. High-diversity mouse populations for complex traits. *Trends Genet* 2019; 35(7): 501–14. doi: 10.1016/j.tig.2019.04.003
- Sharp GL, Hill WG, Robertson A. Effects of selection on growth, body composition and food intake in mice I. Responses in selected traits. *Genet Res* 1984; 43(1): 75–92. doi: 10.1017/s0016672300025738
- Horvat S, Bünger L, Falconer VM, et al. Mapping of obesity QTLs in a cross between mouse lines divergently selected on fat content. *Mamm Genome* 2000; 11(1): 2–7. doi: 10.1007/s003350010002
- Stylianou IM, Christians JK, Keightley PD, et al. Genetic complexity of an obesity QTL (Fob3) revealed by detailed genetic mapping. *Mamm Genome* 2004; 15(6): 472–81. doi: 10.1007/s00335-004-3039-z
- Prevoršek Z, Gorjanc G, Paigen B, Horvat S. Congenic and bioinformatics analyses resolved a major-effect Fob3b QTL on mouse Chr 15 into two closely linked loci. *Mamm Genome* 2010; 21(3/4): 172–85. doi: 10.1007/s00335-010-9252-z
- Mikec Š, Šimon M, Morton NM, Atanur SS, Konc J, Dovč P, et al. Genetic variants of the hypoxia-inducible factor 3 alpha subunit (Hif3a) gene in the Fat and Lean mouse selection lines. *Mol Biol Rep* 2022; 49(6): 4619–31. doi: 10.1007/s11033-022-07309-0
- Šimon M, Mikec Š, Morton NM, et al. Whole genome sequencing of mouse lines divergently selected for fatness (FLI) and leanness (FHI) revealed several genetic variants as candidates for novel obesity genes. *Genes Genomics* 2024; 46(5): 557–75. doi: 10.1007/s13258-024-01507-9
- Morton NM, Nelson YB, Michailidou Z, et al. A Stratified transcriptomics analysis of polygenic fat and lean mouse adipose tissues identifies novel candidate obesity genes. *PLoS One*. 2011; 6(9): e23944. doi: 10.1371/journal.pone.0023944.g001
- McLaren W, Gil L, Hunt SE, et al. The ensembl variant effect predictor. *Genome Biol* 2016;17(1): 122. doi: 10.1186/s13059-016-0974-4
- Birling MC, Yoshiki A, Adams DJ, et al. A resource of targeted mutant mouse lines for 5,061 genes. *Nat Genet* 2021;53(4): 416–9. doi: 10.1038/s41588-021-00825-y
- Bult CJ, Blake JA, Smith CL, et al. Mouse genome database (MGD) 2019. *Nucleic Acids Res* 2019; 47(D1): D801–6. doi: 10.1093/nar/gky1056
- Hu ZL, Park CA, Reecy JM. Bringing the Animal QTLdb and CorrDB into the future: meeting new challenges and providing updated services. *Nucleic Acids Res* 2022; 50(D1): D956–61. doi: 10.1093/nar/gkab1116
- Martin FJ, Amode MR, Aneja A, et al. Ensembl 2023. *Nucleic Acids Res* 2023; 51(D1): D933–41. doi: 10.1093/nar/gkac958
- Raudvere U, Kolberg L, Kuzmin I, et al. g:Profiler: a web server for functional enrichment analysis and conversions of gene lists (2019 update). *Nucleic Acids Res* 2019; 47(W1): W191–8. doi: 10.1093/nar/gkz369.
- Wen P, Xiao P, Xia J. dbDSM: a manually curated database for deleterious synonymous mutations. *Bioinformatics* 2016; 32(12): 1914–6. doi: 10.1093/bioinformatics/btw086
- Loughner CL, Bruford EA, McAndrews MS, Delp EE, Swamynathan S, Swamynathan SK. Organization, evolution and functions of the human and mouse Ly6/uPAR family genes. *Hum Genomics* 2016; 10: 10. doi: 10.1186/s40246-016-0074-2
- Bai X, Ju H, Gao L, et al. Regulation of nicotinic acetylcholine receptor $\alpha 3$ subtype in adipose tissue dysfunction. *Prostaglandins Other Lipid Mediat* 2019; 142: 53–8. doi: 10.1016/j.prostaglandins.2019.04.001
- Cancello R, Zulian A, Maestrini, et al. The nicotinic acetylcholine receptor $\alpha 7$ in subcutaneous mature adipocytes: Downregulation in human obesity and modulation by diet-induced weight loss. *Int J Obes* 2012; 36(12): 1552–7. doi: 10.1038/ijo.2011.275
- Martinez TF, Lyons-Abbott S, Bookout AL, et al. Profiling mouse brown and white adipocytes to identify metabolically relevant small ORFs and functional microproteins. *Cell Metab* 2023; 35(1): 166–83. doi: 10.1016/j.cmet.2022.12.004
- Beigneux AP, Davies BSJ, Bensadoun A, Fong LG, Young SG. GPIHBP1, a GPI-anchored protein required for the lipolytic processing of triglyceride-rich lipoproteins. *J Lipid Res* 2009; 50(suppl.): S57–S62. doi: 10.1194/jlr.R800030-JLR200

27. Wu J, Kakoola DN, Lenchik NI, Desiderio DM, Marshall DR, Gerling IC. Molecular phenotyping of immune cells from young NOD mice reveals abnormal metabolic pathways in the early induction phase of autoimmune diabetes. *PLoS One* 2012; 7(10): e46941. doi: 10.1371/journal.pone.0046941
28. Yamanouchi S, Kuwahara K, Sakata A, et al. A T cell activation antigen, Ly6C, induced on CD4+ Th1 cells mediates an inhibitory signal for secretion of IL-2 and proliferation in peripheral immune responses. *Eur J Immunol* 1998; 28(2): 696–707. doi: 10.1002/(SICI)1521-4141(199802)28:02<696::AID-IMMU696>3.0.CO;2-N
29. Zhao X, Zheng S, Chen D, et al. LY6E restricts entry of human coronaviruses, including currently pandemic SARS-CoV-2. *J Virol* 2020; 94(18): e00562-20. doi: 10.1128/JVI.00562-20
30. Huang Q, Chan KY, Tobey IG, et al. Delivering genes across the blood-brain barrier: LY6A, a novel cellular receptor for AAV-PHP.B capsids. *PLoS One* 2019; 14(11): e0225206. doi: 10.1371/journal.pone.0225206
31. Tokunaga M, Inoue M, Jiang Y, Barnes RH, Buchner DA, Chun TH. Fat depot-specific gene signature and ECM remodeling of Sca1^{high} adipose-derived stem cells. *Matrix Biol* 2014; 36: 28–38. doi: 10.1016/j.matbio.2014.03.005.
32. Delgado-Chaves FM, Gómez-Vela F, Divina F, García-Torres M, Rodríguez-Baena DS. Computational analysis of the global effects of LY6E in the immune response to coronavirus infection using gene networks. *Genes (Basel)* 2020; 11(7): 831. doi: 10.3390/genes11070831
33. Pérez-Pérez R, García-Santos E, Ortega-Delgado FJ, et al. Attenuated metabolism is a hallmark of obesity as revealed by comparative proteomic analysis of human omental adipose tissue. *J Proteomics* 2012; 75(3): 783–95. doi: 10.1016/j.jprot.2011.09.016
34. Wagner G, Lindroos-Christensen J, Einwallner E, et al. HO-1 inhibits preadipocyte proliferation and differentiation at the onset of obesity via ROS dependent activation of Akt2. *Sci Rep* 2017; 7(1): 40881. doi: 10.1038/srep40881
35. Kaupe B, Brandt H, Prinzenberg EM, Erhardt G. Joint analysis of the influence of CYP11B1 and DGAT1 genetic variation on milk production, somatic cell score, conformation, reproduction, and productive lifespan in German Holstein cattle. *J Anim Sci*. 2007; 85(1): 11-21. doi: 10.2527/jas.2005-753.
36. Lee J, Kim H, Kang YW, et al. LY6D is crucial for lipid accumulation and inflammation in nonalcoholic fatty liver disease. *Exp Mol Med* 2023; 55(7): 1479–91. doi: 10.1038/s12276-023-01033-w
37. Ma D, Pan Z, Chang Q, et al. KLF5-mediated Eppk1 expression promotes cell proliferation in cervical cancer via the p38 signaling pathway. *BMC Cancer*. 2021; 21(1): 377. doi: 10.1186/s12885-021-08040-y
38. Chen D, Li W, Du M, Cao B. Adipogenesis, fibrogenesis and myogenesis related gene expression in longissimus muscle of high and low marbling beef cattle. *Livest Sci* 2019; 229: 188–93. doi:10.1016/j.livsci.2019.09.032
39. México V, Michelle Quiñones-Chayrez K, Luna-Nevárez G, et al. Molecular markers associated with response to vaccination against Porcine Reproductive and Respiratory Syndrome virus in a commercial swine farm from southern Sonora. *Vet México OA* 2023;10: 1–13. doi: 10.22201/fmvz.24486760e.2023.1164
40. Young SG, Song W, Yang Y, Birrane G, Jiang H, Beigneux AP, et al. A protein of capillary endothelial cells, GPIHBP1, is crucial for plasma triglyceride metabolism. *Proc Natl Acad Sci USA* 2022; 119(36): e2211136119. doi: 10.1073/pnas.2211136119
41. Hsia SH, Leiter LA. Obesity and dyslipidemia: Epidemiology, physiology, and effects of weight loss. *Endocrinologist*. 1995 Mar;5(2):118–31.
42. Simončič M, Režen T, Juvan P, et al. Obesity resistant mechanisms in the Lean polygenic mouse model as indicated by liver transcriptome and expression of selected genes in skeletal muscle. *BMC Genomics* 2011; 12(1): 96. doi: 10.1186/1471-2164-12-96
43. Morton NM, Densmore V, Wamil M, et al. A polygenic model of the metabolic syndrome with reduced circulating and intra-adipose glucocorticoid action. *Diabetes* 2005; 54(12): 3371–8. doi: 10.2337/diabetes.54.12.3371
44. Arner P, Andersson DP, Bäckdahl J, Dahlman I, Rydén M. Weight gain and impaired glucose metabolism in women are predicted by inefficient subcutaneous fat cell lipolysis. *Cell Metab* 2018; 28(1): 45–e3. doi: 10.1016/j.cmet.2018.05.004
45. Xu H, Tao X, Wei Y, et al. Cloning of porcine GPIHBP1 gene and its tissue expression pattern and genetic effect on adipose traits. *Gene* 2015; 557(2): 146–53. doi: 10.1016/j.gene.2014.12.017
46. Chen HC, Smith SJ, Ladha Z, et al. Increased insulin and leptin sensitivity in mice lacking acyl CoA:diacylglycerol acyltransferase 1. *J Clin Invest* 2002; 109(8): 1049–55. doi: 10.1172/JCI14672
47. Smith SJ, Cases S, Jensen DR, et al. Obesity resistance and multiple mechanisms of triglyceride synthesis in mice lacking Dgat. *Nat Genet* 2000; 25(1): 87–90. doi: 10.1038/75651
48. Cui JX, Zeng YQ, Wang H, Chen W, Du JF, Chen QM, et al. The effects of DGAT1 and DGAT2 mRNA expression on fat deposition in fatty and lean breeds of pig. *Livest Sci* 2011; 140(1/3): 292-6. doi: 10.1016/j.livsci.2011.04.007
49. Xing K, Liu H, Zhang F, Liu Y, Shi Y, Ding X, et al. Identification of key genes affecting porcine fat deposition based on co-expression network analysis of weighted genes. *J Anim Sci Biotechnol* 2021; 12(1): 100. doi: 10.1186/s40104-021-00616-9
50. Li T, Xu D, Zuo B, Lei M, Xiong Y, Chen H, et al. Ectopic overexpression of porcine DGAT1 increases intramuscular fat content in mouse skeletal muscle. *Transgenic Res* 2013; 22(1): 187–94. doi: 10.1007/s11248-012-9633-z
51. Li X, Ekerljung M, Lundström K, Lundén A. Association of polymorphisms at DGAT1, leptin, SCD1, CAPN1 and CAST genes with color, marbling and water holding capacity in meat from beef cattle populations in Sweden. *Meat Sci* 2013; 94(2): 153–8. doi: 10.1016/j.meatsci.2013.01.010
52. Grisart B, Coppieters W, Farnir F, et al. Positional candidate cloning of a QTL in dairy cattle: identification of a missense mutation in the bovine DGAT1 gene with major effect on milk yield and composition. *Genome Res* 2002; 12(2): 222–31. doi: 10.1101/gr.224202
53. Shimada H, Noro E, Suzuki S, et al. Effects of adipocyte-derived factors on the adrenal cortex. *Curr Mol Pharmacol* 2019; 13(1): 2–6. doi: 10.2174/1874467212666191015161334.
54. Lee MJ, Pramyothin P, Karastergiou K, Fried SK. Deconstructing the roles of glucocorticoids in adipose tissue biology and the development of central obesity. *Biochim Biophys Acta* 2014; 1842(3): 473–81. doi: 10.1016/j.bbdis.2013.05.029
55. Sandeep TC, Andrew R, Homer NZM, Andrews RC, Smith K, Walker BR. Increased in vivo regeneration of cortisol in adipose tissue in human obesity and effects of the 11 β -hydroxysteroid dehydrogenase type 1 inhibitor carbenoxolone. *Diabetes* 2005; 54(3): 872–9. doi: 10.2337/diabetes.54.3.872
56. Shiu FH, Wong JC, Yamamoto T, Lala T, Purcell RH, Owino S, et al. Mice lacking full length Adgrb1 (Bai1) exhibit social deficits, increased seizure susceptibility, and altered brain development. *Exp Neurol* 2022; 351: 113994. doi: 10.1016/j.expneurol.2022.113994

57. Fond AM, Lee CS, Schulman IG, Kiss RS, Ravichandran KS. Apoptotic cells trigger a membrane-initiated pathway to increase ABCA1. *J Clin Invest* 2015; 125(7): 2748–58. doi: 10.1172/JCI80300
58. Cuffe H, Liu M, Key CCC, et al. Targeted deletion of adipocyte abca1 (ATP-binding cassette transporter A1) impairs diet-induced obesity. *Arterioscler Thromb Vasc Biol* 2018; 38(4): 733–43. doi: 10.1161/ATVBAHA.117.309880
59. Ren W, Aihara E, Lei W, et al. Transcriptome analyses of taste organoids reveal multiple pathways involved in taste cell generation. *Sci Rep* 2017; 7(1): 4004. doi: 10.1038/s41598-017-04099-5
60. Kaplan REW, Webster AK, Chitrakar R, Dent JA, Baugh LR. Food perception without ingestion leads to metabolic changes and irreversible developmental arrest in *C. elegans*. *BMC Biol* 2018; 16(1): 112. doi: 10.1186/s12915-018-0579-3.
61. Brandt C, Nolte H, Henschke S, et al. Food perception rimes hepatic ER homeostasis via melanocortin-dependent control of mTOR activation. *Cell* 2018;175(5): 1321–e11. doi: 10.1016/j.cell.2018.10.015
62. Koekkoek LL, Mul JD, la Fleur SE. Glucose-sensing in the reward system. *Front Neurosci* 2017; 11: 716. doi: 10.3389/fnins.2017.00716
63. Park SJ, Yu Y, Wagner B, Valinsky WC, Lomax AE, Beyak MJ. Increased TASK channel-mediated currents underlie high-fat diet induced vagal afferent dysfunction. *Am J Physiol Gastrointest Liver Physiol* 2018; 315(4): G592–G601. doi: 10.1152/ajpgi.00335.2017
64. Chen Y, Zeng X, Huang X, et al. Crosstalk between KCNK3-mediated ion current and adrenergic signaling regulates adipose thermogenesis and obesity. *Cell* 2017; 171(4): 836–e13. doi: 10.1016/j.cell.2017.09.015
65. Hoggart CJ, Venturini G, Mangino M, et al. Novel approach identifies SNPs in SLC2A10 and KCNK9 with evidence for parent-of-origin effect on body mass index. *PLoS Genet* 2014; 10(7): e1004508. doi: 10.1371/journal.pgen.1004508.
66. Sung YJ, Pérusse L, Sarzynski MA, et al. Genome-wide association studies suggest sex-specific loci associated with abdominal and visceral fat. *Int J Obes (Lond)* 2016; 40(4): 662–74. doi: 10.1038/ijo.2015.217
67. Liang ZS, Cimino I, Yalcin B, et al. Trappc9 deficiency causes parent-of-origin dependent microcephaly and obesity. *PLoS Genet* 2020; 16(9): e1008916. doi: 10.1371/journal.pgen.1008916.
68. Cui P, Wang W, Zhang D, et al. Identification of TRAPPC9 and BAIAP2 gene polymorphisms and their association with fat deposition-related traits in Hu sheep. *Front Vet Sci.* 2022; 9: 928375. doi: 10.3389/fvets.2022.928375
69. Liu H, Song H, Jiang Y, et al. A single-step genome wide association study on body size traits using imputation-based whole-genome sequence data in Yorkshire pigs. *Front Genet* 2021; 12: 629049. doi: 10.3389/fgene.2021.629049
70. Chen H, Yan X, Sun A, Zhang L, Zhang J, Yan Y. High-fat-diet-induced extracellular matrix deposition regulates integrin–FAK signals in adipose tissue to promote obesity. *Mol Nutr Food Res* 2022; 66(7): e2101088. doi: 10.1002/mnfr.202101088
71. Hadley C, Cakir I, Cone RD. The role of the focal adhesion kinase family in leptin receptor signaling. *J Endocr Soc* 2020; 4(Suppl. 1): SAT-604. doi: 10.1210/jendso/bvaa046.666
72. Luk CT, Shi SY, Cai EP, et al. FAK signalling controls insulin sensitivity through regulation of adipocyte survival. *Nat Commun* 2017; 8(1): 14360. doi: 10.1038/ncomms14360
73. Bisht B, Srinivasan K, Dey CS. In vivo inhibition of focal adhesion kinase causes insulin resistance. *J Physiol* 2008; 586(16): 3825–37. doi: 10.1113/jphysiol.2008.157107
74. Szyndler-Nędza M, Piorkowska K. Effect of DGAT1 gene mutation in sows of dam-line on the composition of the produced milk and piglet rearing during 21-day lactation. *Afr J Biotechnol.* 2015; 14(31): 2478–83. doi: 10.5897/AJB2015.14817
75. Costa A, Schwarzenbacher H, Mészáros G, Fuerst-Waltl B, Fuerst C, Sölkner J, et al. On the genomic regions associated with milk lactose in Fleckvieh cattle. *J Dairy Sci* 2019; 102(11): 10088–99. doi: 10.3168/jds.2019-16663
76. Dong W, Yang J, Zhang Q, Jiang L. Role of GPIHBP1 in regulating milk protein traits in dairy cattle. *Anim Biotechnol* 2020; 31(1): 81–5. doi: 10.1080/10495398.2018.1536064
77. Jiang J, Prakapenka D, Ma L, Cole, et al. Extreme antagonistic pleiotropy effects of DGAT1 on fat, milk and protein yields. In: *Proceedings of the world congress on genetics applied to livestock production.* 2018: 142.
78. Khan MZ, Dari G, Khan A, Yu Y. Genetic polymorphisms of TRAPPC9 and CD4 genes and their association with milk production and mastitis resistance phenotypic traits in Chinese Holstein. *Front Vet Sci* 2022; 9: 1008497. doi: 10.3389/fvets.2022.1008497
79. Ma L, Luo H, Brito LF, Chang Y, Chen Z, Lou W, et al. Estimation of genetic parameters and single-step genome-wide association studies for milk urea nitrogen in Holstein cattle. *J Dairy Sci* 2023; 106(1): 352–63. doi: 10.3168/jds.2022-21857
80. Yang J, Liu X, Wang D, Ning C, Wang H, Zhang Q, et al. Functional validation of GPIHBP1 and identification of a functional mutation in GPIHBP1 for milk fat traits in dairy cattle. *Sci Rep* 2017; 7: 8546. doi: 10.1038/s41598-017-08668-6
81. Tăbăran A, Balteanu VA, Gal E, et al. Influence of DGAT1 K232A polymorphism on milk fat percentage and fatty acid profiles in romanian Holstein Cattle. *Anim Biotechnol* 2015; 26(2): 105–11. doi: 10.1080/10495398.2014.933740
82. Atanassov C, Viallemonteil E, Lucas C, et al. Proteomic pattern of breast milk discriminates obese mothers with infants of delayed weight gain from normal-weight mothers with infants of normal weight gain. *FEBS Open Bio* 2019; 9(4): 736–42. doi: 10.1002/2211-5463.12610
83. Isganaitis E, Venditti S, Matthews TJ, Lerin C, Demerath EW, Fields DA. Maternal obesity and the human milk metabolome: associations with infant body composition and postnatal weight gain. *Am J Clin Nutr* 2019; 110(1): 111–20. doi: 10.1093/ajcn/nqy334
84. Karabağ K, Alkan S, Karlı T, İkten C, Şahin İ, Mendeş M. Effects of selection in terms of meat yield traits on leptin receptor gene in japanese quail lines. *Slov Vet Res* 2022; 59(2): 89–98. doi: 10.26873/SVR-1316-202

Določanje prioritarnih kandidatnih genov znotraj intervala Fob3b1 QTL na kromosomu 15 pri mišjih modelih za poligeno debelost in vitkost z uporabo integrativne genomike

M. Šimon, T. Kunej, N. M. Morton, S. Horvat

Izvleček: Kopičenje odvečne maščobe vpliva na kakovost mesa, plodnost, proizvodnost in presnovo pri rejnih živalih. Mišji modeli predstavljajo učinkovito orodje za raziskovanje genetske osnove teh lastnosti. Predhodne analize QTL-ov edinstvenih mišjih selekcijskih linij za poligeno debelost (debela linija) in vitkost (vitka linija) so razkrile štiri glavne QTL-e za debelost: Fob1, Fob2, Fob3 in Fob4. Fob3, ki se nahaja na kromosomu 15, je bil kasneje razdeljen na Fob3a in Fob3b, zadnji pa se dodatno razdeli na Fob3b1 in Fob3b2. Od 158 genov, anotiranih v Fob3b1, je bilo v prejšnjih študijah predlaganih 16 kandidatnih genov. Vendar pa genomska variabilnost med debelo in vitko linijo na tem lokusu ni bila v celoti raziskana. Namen te študije je bil potrditi predhodno identificirane kandidate in identificirati nove kandidatne gene, ki bi lahko bili odgovorni za učinek Fob3b1. Podatki iz celotnega genoma sekvenciranja in transkriptomskih analiz debelih in vitkih mišjih linij so bili vključeni v primerjalno analizo s QTL-i za debelost pri govedu in prašičih iz Animal QTLdb ter fenotipi, pridobljenimi iz Mednarodnega konzorcija za fenotipizacijo miši (IMPC) in podatkovne zbirke mišjega genoma (MGD). Izmed 158 genov, lociranih v Fob3b1, smo prednostno obravnavali 17 kandidatnih genov, vključno s šestimi predhodno predlaganimi (Adgrb1, Col22a1, Cyp11b1, Dgat1, Gpihbp1 in Ly6a) in 11 novimi kandidati: 9030619P08Rik, Eppk1, Kcnk9, Ly6c1, Ly6d, Ly6h, Ly6i, Ly6m, Ptk2, Trappc9 in Ly6e. Biološka funkcija in pregled literature za kandidatne gene nakazuje, da lahko učinek Fob3b1 na debelost deluje preko metabolizma trigliceridov (Dgat1 in Gpihbp1) ter preoblikovanja citoskeleta in zunajceličnega matriksa (Ly6a, Ly6e in Eppk1). Nadaljnje natančno kartiranje, genetske in »omske« študije bodo pojasnili, ali je učinek Fob3b1 posledica vzročnega učinka ene same genetske različice ali morda aditivnega učinka kombinacije večjega števila teh pozicijskih kandidatov. Uporabljeni bioinformacijski pristop pri določanju prednostne liste kandidatnih genov za debelost lahko služi tudi kot model za preučevanje drugih lastnosti v veterinarskih in živilorejskih znanostih.

Ključne besede: povezovanje podatkov; izražanje genov; razvrstitev genov po pomembnosti; mišji modeli; debelost; QTL; posamezni nukleotid; polimorfizem