

**RESEARCH ARTICLE**

## Sex-related hypothalamic-pituitary-gonadal and hypothalamic-pituitary-adrenal axis adaptation during military training

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### Abstract

Reproductive endocrine function adapts to psychological, environmental, and energy-associated stressors. Multistressor environments upregulate hypothalamic-pituitary-adrenal (HPA) axis, causing suppression of the hypothalamic-pituitary-gonadal (HPG) axis, but it is not known if this pattern or its magnitude is sex biased. We compared HPG and HPA axis activity in 9 men and 34 women undergoing Army training. One-hour low-dose gonadorelin and Synacthen tests were conducted at 1 and 29 wk, measuring gonadotrophins and cortisol. Cortisol was measured from hair every 3 mo. Morning and evening salivary cortisol and psychometric questionnaires were measured at six timepoints. Sexes were compared over time by two-way ANOVA. Gonadotrophin responses were significantly higher in women than men in week 1, but no sex difference was seen at week 29 (no significant sex  $\times$  time interaction). Week 1 cortisol response was higher among men, but week 29 cortisol response was higher among women (sex  $\times$  time  $F_{(1,44)} = 18.0$ ,  $P < 0.001$ ). Hair cortisol was higher among women than men beforehand, not different between sexes during the first 3 mo, and significantly higher among women during training months 5–11 ( $F_{(3,15)} = 3.25$ ,  $P = 0.024$ ). Morning salivary cortisol was higher among women in weeks 8 and 14, but higher among men in week 29 ( $F_{(4,76)} = 4.0$ ,  $P = 0.005$ ). No differences were seen in evening salivary cortisol. Psychometrics did not change or differ between sexes. HPA axis responses to military training were greater among women than men. HPG axis responses suggest greater downregulation among women. These findings will enable equitable and individualized management of people undergoing periods of intensive physical stress.

**NEW & NOTEWORTHY** We conducted a comprehensive comparison of adrenal and reproductive function in men and women undergoing 11-mo military training. We found progressively elevated cortisol levels and dynamic cortisol response to stress among women, but not men, and suppression of reproductive function among women. The physiological impact of stressful military training was greater among women than men; this could not be explained by energy balance, and sex-specific effects of sleep, socio-ethnographic, or other stressors may be responsible.

cortisol; endocrinology; military; reproductive hormones; sex

### INTRODUCTION

Regulation of reproductive function is dynamic and adaptive. External factors such as psychological stress, environmental challenges, and availability of energy can be considered “stressors,” which activate the hypothalamic-pituitary-adrenal (HPA) axis, increasing the production of the primary human glucocorticoid, cortisol. Activation of the HPA axis suppresses the hypothalamic-pituitary-gonadal (HPG) axis (1–4).

The HPA axis demonstrates sex-biased activity. Gonadal steroids are known to modulate HPA axis reactivity; circulating estrogens enhance the glucocorticoid response to stress, increasing adrenal sensitivity to adrenocorticotrophin (ACTH) (5, 6), whereas androgens have the opposite effect, attenuating the HPA axis response to a stressor (7) by dampening adrenal responsiveness (8). Adolescent girls demonstrate greater sensitivity to ACTH-(1–24) than boys (9). Sexual dimorphism in stress processing has also been observed in

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Submitted 10 September 2024 / Revised 20 November 2024 / Accepted 20 November 2024



the absence of gonadal steroids. Following pharmacologically induced HPG axis suppression, enhanced ACTH and cortisol responses to corticotrophin-releasing hormone (CRH) and exercise were observed among men compared with women (10). However, the sex difference in adrenal responsiveness was attenuated, suggesting that neural regulation of the HPA axis and central feedback mechanisms may be sexually dimorphic (10), perhaps reflecting differing energetic requirements (11). HPA axis function is also moderated by cortisol-binding globulin (CBG) (12), the production of which is governed by estrogens, and arginine vasopressin (AVP), both sexually dimorphic in their abundance.

Clinical studies seeking to understand the effect of exercise and its associated stressors on reproductive function demonstrate that insufficient caloric intake relative to exercise energy expenditure is a primary stressor that can lead to suppression of the HPG axis, manifesting clinically as hypothalamic amenorrhea (13). Although clinical manifestations of HPG axis suppression are likely to be more apparent in women than men, a recent review suggests men may be less susceptible to these effects (13).

We have previously reported that women undertaking arduous military training demonstrated anovulation and significant suppression of gonadotroph function (14) and activation of the HPA axis (15). Men undertaking the same training demonstrated significantly greater energy deficits than women, due to higher energy expenditure (16). We aimed to compare dynamic HPG and HPA axis activity between men and women from these studies. We hypothesized that exposure to military training would produce a sex-biased HPG and HPA axis change, with enhanced responsiveness and basal levels of cortisol, with attendant suppression of HPG axis function, among women compared with men.

## METHODS

### Participants

Participants were training to enter the British Army as Officers at the Royal Military Academy, Sandhurst, UK. This course provides comprehensive physical, mental, and environmental challenges over three, 14-wk terms separated by 2 or 3 wk of leave, as described previously (14, 17). Officer Cadets undergo competitive selection for roles within the army during week 20. Training is undertaken in mixed-sex groups; the proportion of female Officer Cadets is typically around 8%–12%.

The Female Endocrinology in Arduous Training cohort study examined endocrine, metabolic, and bone health in female Officer Cadets over three successive Commissioning Course intakes. Findings from female participants demonstrated high levels of physical activity (17), activation of the HPA axis (15), and suppression of the HPG axis with maladaptive metabolic changes (14) over 44 wk of training, with temporary uncoupling of bone turnover (18). A cohort of men was studied during one intake. We have previously demonstrated more negative energy balance among these men than female contemporaries (16). Here we present a sex comparison of HPA and HPG axis changes. Inclusion criteria were: medically fit to commence the course and aged 18–30 yr. Participant health status was confirmed by entry medical

examination before enrolment, including history, examination, and ECG, completed according to entry requirements for UK Defence (19). Exclusion criteria were pregnancy, known history of adrenal, gonadal, or gonadotrophin-releasing hormone (GnRH) insufficiency, pituitary disease, thyroid disease in the past year, diabetes, hyperparathyroidism, osteopenia, oral, inhaled, or topical glucocorticoid use or ongoing musculoskeletal injury. Due to the known effects of synthetic estrogens on CBG, total cortisol, hair cortisol, luteinizing hormone (LH), and follicle-stimulating hormone (FSH), combined oral contraceptive pill (COCP) users were excluded from this analysis (9, 14, 20). Participants who withdrew from or did not commence training ( $n = 10$  and 7, respectively) or from the study measures (0 participants) were excluded from the analysis. All participants provided informed consent. Ethical approval was obtained from the Ministry of Defence Research Ethics Committee.

### Experimental design.

Participant body mass and height were measured and a questionnaire was completed, which ascertained age, contraceptive use, the occurrence of any stressful life events in the past month, levels of financial and work stress, Impact of Events Scale-Revised (IES-R) (21) with relation to any stressful events, Patient Health Questionnaire 9 (PHQ-9) (22), Beck Anxiety Inventory (BAI) (23), and Connor Davidson Resilience Scale 10 (CDRISC 10) (24). In weeks 1, 14, 29, and 44 of training, PHQ-9, BAI, CDRISC 10, significant personal stressors and IES-R, and levels of work and financial stress were recorded.

A simultaneous, low-dose gonadotrophin-releasing hormone (GnRH) and ACTH test were used to detect differences in cortisol, LH, and FSH responsiveness (25) at weeks 1 and 28 of training. Due to constraints imposed by the training schedule, dynamic testing was completed in the late afternoon, and testing was not synchronized to the menstrual cycle phase. Participants were allowed to relax before a 20G cannula was inserted into an antecubital fossa vein. A sample of blood was taken from the cannula in EDTA-containing tubes. After 10–15 min, 10 µg gonadorelin hydrochloride (Intrapharm, Maidenhead, UK) followed by 1.0 µg ACTH-(1–24) (Synacthen, Mallinckrodt, Dublin, Ireland) was injected followed by a 10 mL saline flush. Sampling was repeated after 20, 30, 40, and 60 min. Venous blood was also sampled after a 10 h fast in weeks 1, 14, and 29. Blood was centrifuged and stored at  $-80^{\circ}\text{C}$  until analysis. Saliva was sampled using a synthetic swab (Salivette, Sarstedt, Leicester, UK), during weeks 1, 8, 14, 16, 20, and 29, before bed in the evening followed by first thing the following morning. Saliva was sampled before brushing teeth and participants were given verbal, written, and video instructions on the technique. Saliva was stored at  $5^{\circ}\text{C}$  for up to 3 days before being stored at  $-80^{\circ}\text{C}$  until analysis. A 5 mm diameter section of hair was sampled from the scalp at the posterior vertex region at the start of the study and at weeks 14 and 29. Hair was stored in aluminum foil at room temperature until analysis.

### Laboratory Methods

LH and FSH were assayed by Abbott Architect (Abbot, Longford, Ireland). Plasma cortisol was extracted by supported liquid extraction using the Biotage Extrahera robot (Biotage AB, Stockholm, Sweden) and measured using

tandem liquid chromatography-mass spectrometry (LC-MS/MS), as described previously (15). Total CBG was assayed from plasma using immunoassay according to the method of Lewis and Elder (26). Cortisol was assayed from saliva using a commercial immunoassay according to the manufacturer's instructions (Salimetrics, State College, PA). Hair cortisol was assayed in 1 cm segments, assuming an average growth rate of 1 cm per month (27), by Dresden Lab Service GmbH (Dresden, Germany) using LC-MS/MS, as described previously (28), providing average 1-mo cortisol exposure. Coefficient variations were <4% for Architect assays and <10% for immunoassays.

### Statistical Analysis

Data were assessed for normality using the Shapiro-Wilk test. Normally distributed data are presented as means  $\pm$  SD and non-normal data as median [interquartile range (IQR)]. Peak fold-wise increases in LH and FSH from before GnRH administration, and 1-h fold-wise increase area under the curve (AUC, calculated by the trapezoidal rule) were calculated. Data from female progestrone-only contraceptive users and noncontraceptive users were pooled for analyses since no differences were seen between these groups in fold-wise LH or FSH responses (Table 3). Missing data were excluded [97 saliva samples (18.8%) due to insufficient volume for analysis]. No values were reported below the level of quantification and no data were imputed. The peak and 1-h AUC of cortisol response to ACTH-(1-24) administration were calculated. Hair cortisol concentrations were analyzed using individual mean concentrations over four consecutive 3–4-mo periods, to account for differing hair length (4 mo pre, and three subsequent 3–4-mo periods of training). Peak and AUC fold-wise LH and FSH, peak and AUC plasma cortisol, hair cortisol, PHQ9, and BAI data were transformed by base  $e$  (Ln).

Male and female physical and psychological characteristics at baseline were compared using independent samples *t* tests.

Sex differences in CDRISC-10, IES-R, and Ln-transformed PHQ9 and BAI were assessed over time using mixed, repeated measures ANOVA. Ln-transformed fold-wise peak and AUC of LH and FSH response to GnRH, peak and AUC of plasma cortisol response to ACTH-(1-24), CBG, and fasted plasma cortisol were compared across groups from weeks 1 to 29 by two-way ANOVA (sex  $\times$  time). Post hoc comparisons compared sexes at each timepoint (independent samples *t* test). Male and female Ln-transformed average hair cortisol and morning and evening salivary cortisol concentrations were compared over time by mixed repeated measures ANOVA.

Statistical analyses were conducted in SPSS for Mac, version 29.0 (IBM, New York). Significance was set at  $P < 0.05$ .

## RESULTS

A total of 78 Officer Cadets volunteered for the study (68 female, 10 male), of whom 61 completed the study [7 did not commence training (all female), 10 withdrew from training (9 female, 1 male)] and 18 were excluded from this analysis due to COCP use. A breakdown of contraceptive use among included participants is shown in Table 1. A complete dataset is presented for 43 participants (34 female and 9 male).

### Physical and Psychological Characteristics

Physical and psychological characteristics are shown in Table 1. Men were significantly taller and heavier than women, resilience levels were robust, and scores of anxiety and depression were low, with no differences between sexes. Women had experienced numerically more adverse events in the month before training than men. Levels of financial and work-related stress were similar between the sexes. During training, scores of depression (PHQ-9) increased from weeks 1 to 14 and 29, and anxiety (BAI) increased marginally at week 14, whereas resilience (CDRISC-19) did not significantly change. No sex-by-time interactions were seen (Table 2). The number of adverse events increased from weeks 1 to 29 in

**Table 1.** Participant characteristics 2–4 wk before the study

	Women ( <i>n</i> = 34)	Men ( <i>n</i> = 9)
Age	24.9 $\pm$ 2.9	24.8 $\pm$ 3.0
Height, m	1.67 $\pm$ 0.06	1.83 $\pm$ 0.07*
Weight, kg	66.7 $\pm$ 8.2	85.3 $\pm$ 7.2*
Hormonal contraception used throughout study		Not applicable
Oral progestogen	6	
Depot progestogen	1	
Progestogen implant	9	
Intrauterine progestogen-eluting device	4	
None	14	
CD RISC 10	32.7 $\pm$ 0.8	33.5 $\pm$ 1.8
PHQ-9	2 (1, 4.5)	2 (0.5, 2)
BAI	7 (2, 11)	3 (2, 8)
Adverse events and IES-R	13 events in 13 participants (38%)	2 events in 2 participants (22%)
IES-R 2 and 0		
High or severe financial stress, <i>n</i> (%)	23.3 $\pm$ 14.3	0 (0%)
How often do you experience stress due to work?	1 (3%)	
Never	1 (3%)	0 (0%)
Some periods	24 (71%)	7 (78%)
Several periods	8 (24%)	1 (11%)
Permanently	0 (0%)	0 (0%)
Not working	0 (0%)	1 (11%)

Means  $\pm$  SD or median (interquartile range). BAI, Beck Anxiety Inventory; CDRISC 10, Connor Davidson Resilience Scale 10-point version; IES-R, Impact of Events Scale-Revised; PHQ-9, Patient Health Questionnaire 9-point version. \* $P < 0.001$  male vs. female.

**Table 2.** Resilience, anxiety, low mood, adverse life events, work stress, and financial stress

	Week 1	Week 14	Week 29	Week 44	Time	Sex × Time
CDRISC 10 (means ± SD)						
Women	30.9 ± 0.9	29.3 ± 0.9	29.3 ± 1.0	29.1 ± 0.9	$P = 0.2$	$P = 0.6$
Men	32.4 ± 1.3	30.2 ± 1.95	33.1 ± 1.1	31.7 ± 1.5	$F(3,39) 1.55$	$F(1,113) 0.70$
BAI (median, IQR)						
Women	5 (2, 8)	6 (1, 11)	3 (2, 8)	2 (1, 5)	$P < 0.001$	$P = 0.3$
Men	5 (1, 7)	2 (0, 10)	1 (0, 8)	1 (0, 2)	$F(3,37) 15.1$	$F(1,117) 1.00$
PHQ-9						
Women	2 (1, 6)	5 (2, 7)	5 (1, 7)	3 (1, 6)	$P < 0.001$	$P = 0.6$
Men	2 (1, 4)	5 (3, 7)	4 (1, 6)	2 (2, 3)	$F(3,39) 8.14$	$F(1,123) 0.63$
Adverse events, IES-R (means ± SD)						
1						
Women	5 in 4 participants 8.0 ± 7.3	8 in 8 participants 10.0 ± 5.2	10 in 10 participants 29.0 ± 15.0	5 in 5 participants 25.0 ± 15.1		
Men	4 in 4 participants 9.7 ± 5.0	4 in 4 participants 8 ± 7.5	4 in 4 participants 29.7 ± 24.0	1 in 1 participant (51)		
How often do you experience stress due to work? n (%)						
Never						
Women	5 (15%)	0 (0%)	0 (0%)	0 (0%)		
Men	1 (11%)	0 (0%)	0 (0%)	0 (0%)		
Some periods						
Women	19 (56%)	13 (38%)	13 (38%)	20 (59%)		
Men	5 (56%)	1 (11%)	2 (22%)	3 (33%)		
Several periods						
Women	7 (21%)	19 (56%)	17 (5%)	11 (32%)		
Men	2 (22%)	5 (56%)	4 (44%)	5 (56%)		
Permanently						
Women	1 (3%)	2 (6%)	1 (3%)	2 (6%)		
Men	0 (0%)	1 (11%)	2 (22%)	0 (0%)		
Not working						
Women	4 (12%)	0 (0%)	0 (0%)	0 (0%)		
Men	2 (22%)	0 (0%)	0 (0%)	0 (0%)		
High or severe financial stress n (%)						
Women	1 (3)	1 (3)	2 (6)	3 (9)		
Men	0	0	0	0		

BAI, Beck Anxiety Inventory; CDRISC 10, Connor Davidson Resilience Scale 10-point version; IES-R, Impact of Events Scale-Revised; IQR, interquartile range; PHQ-9, Patient Health Questionnaire 9-point version; SD, standard deviation.

women but not in men; the stressor impact score (IES-R) associated with these events increased from weeks 1 to 29 but did not differ between men and women. Work-related and financial stresses were similar between the sexes.

## LH and FSH

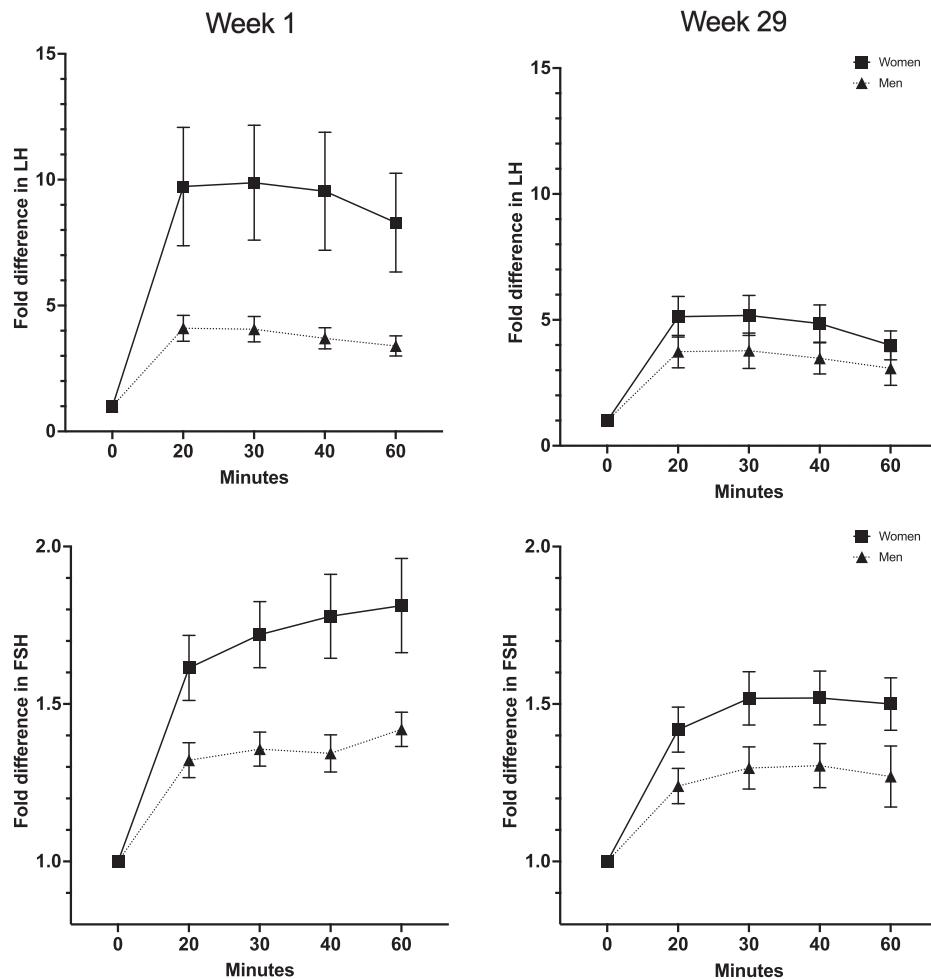
The median (IQR) duration from the first day of the last menstrual period to the GnRH test was 16 (7, 20) and 16 (8, 20.5) days, for the weeks 1 and 29 tests, respectively ( $P = 0.78$ ). Gonadotroph responses to GnRH are shown in Fig. 1. Although visual trends suggest a greater decrease in female responses after 29 wk than male, no statistically significant sex-by-time interaction was seen for FSH (AUC  $F_{(1,60)} = 0.6$ ,  $P = 0.40$ ; Peak:  $F_{(1,60)} = 0.81$ ,  $P = 0.30$ ) or LH (AUC  $F_{(1,60)} = 1.0$ ,  $P = 0.30$ ; Peak  $F_{(1,60)} = 2.0$ ,  $P = 0.15$ ). There was also no overall change over time in FSH response (AUC  $F_{(1,60)} = 3.27$ ,  $P = 0.07$ ; Peak:  $F_{(1,60)} = 1.65$ ,  $P = 0.20$ ) although peak LH response decreased (AUC:  $F_{(1,60)} = 3.52$ ,  $P = 0.06$ ; Peak:  $F_{(1,60)} = 0.8$ ,  $P = 0.03$ ). In week 1, the response to GnRH was higher among women than men, both FSH (AUC  $t = -2.92$ ,  $P = 0.01$ ; Peak:  $t = -3.04$ ,  $P = 0.002$ ) and LH (AUC  $t = -2.48$ ,  $P = 0.009$ ; Peak  $t = -2.6$ ,  $P = 0.007$ ) but there was no significant difference in week 29 (AUC FSH:  $t = -1.68$ ,  $P = 0.06$ ; Peak FSH:  $t = -1.52$ ,  $P = 0.07$ ; AUC LH:  $t = -0.67$ ,  $P = 0.20$ ; Peak LH:  $t = -0.64$ ,  $P = 0.30$ ). A subgroup analysis also showed no differences in FSH or LH responses between

progesterone-containing contraceptive users and nonusers (Table 3).

## Plasma Cortisol Exposure in Male and Female Cohorts during Training

Cortisol responses to ACTH are shown in Fig. 2. At week 1, cortisol response was greater in men than women but was higher at week 29 among women than men (sex × time interaction; Peak:  $F_{(1,44)} = 17.8$ ,  $P < 0.001$ ; AUC:  $F_{(1,44)} = 18.0$ ,  $P < 0.001$ ). CBG did not differ significantly between women and men at week 1 ( $686 \pm 123$  nmol/L vs.  $753 \pm 173$  nmol/L,  $P = 0.40$ ) or week 29 ( $594 \pm 311$  nmol/L vs.  $385 \pm 172$  nmol/L,  $P = 0.06$ ); there was no interaction of sex × time ( $F_{(1,42)} = 1.15$ ,  $P = 0.30$ ). Total fasted plasma cortisol did not differ between women and men at week 1 or 29 ( $686 \pm 123$  nmol/L vs.  $753 \pm 173$  nmol/L,  $P = 0.20$ , and  $662 \pm 164$  nmol/L vs.  $670 \pm 99$  mmol/L, respectively,  $P = 0.90$ ) with no interaction of sex × time ( $F_{(1,44)} = 0.24$ ,  $P = 0.60$ ).

Hair cortisol concentration varied according to sex (sex × time  $F_{(3,15)} = 3.25$ ,  $P = 0.024$ ) being higher among women in the months before training, with no significant sex difference seen during the first 3 mo, and higher concentrations among women during months 5–7 and 9–11 of training (Fig. 3A). Incomplete salivary cortisol samples were obtained from male participants in week 14 so this timepoint was excluded from data analysis. Morning salivary cortisol varied during



**Figure 1.** Male and female gonadotrophin responses to 10  $\mu$ L gonadorelin during weeks 1 and 29 of military training. A two-way ANOVA showed no significant sex  $\times$  time interaction. Data are means  $\pm$  SE.

training according to sex (sex  $\times$  time  $F_{(4, 76)} = 4.00, P = 0.005$ ) being higher among women in week 8, but higher among men in week 20 (a week when significant psychological pressure was induced), with no differences at other times (Fig. 3B). No differences were seen in evening salivary cortisol during training or by sex (sex  $\times$  time  $F_{(4, 80)} = 1.1, P = 0.30$ ; Fig. 3C).

## DISCUSSION

To the best of our knowledge, this is the first study to compare dynamic endocrine function in men and women during arduous military training. Although both men and women experienced elevated cortisol levels during training, there was a notable sex-biased impact on the magnitude and pattern of HPA axis activation. The initial response of cortisol to ACTH-(1-24) was greater among men, commensurate with expected sex differences in stress responses (29), but after 29 wk of physical challenge, the response among women was enhanced, whereas the response in men was decreased. Average hair cortisol concentrations increased in women throughout the study and were significantly higher than in men before the study and during the latter 4–11 mo of training. Hair cortisol concentrations are expected to be higher among men in comparison with women on average (30). Morning salivary-free cortisol was higher among women during the first 14 wk of training suggesting greater

anticipatory stress (31). However, contrary to our hypothesis, the female gonadotroph response was not significantly suppressed compared with men after 29 wk (assessed as the interaction between sexes over time), although post hoc tests demonstrated a reduction in gonadotroph response over time in women, but not men.

Arduous military training imposes significant mental and physiological stress on individuals, driving an adaptive HPA axis response to mobilize energy (15, 32). In studies of military training, HPA axis upregulation and attendant HPG axis suppression (usually manifested as a decreased serum testosterone among male participants) are generally associated with an energy deficit (33–35). Previously, we conducted a detailed assessment of energy balance in this cohort using doubly labeled water and weighted dietary analysis, finding that men had significantly higher energy expenditure and energy intake, and a more negative energy balance overall, compared with women (16). There were no significant sex interactions in basal serum androgen or cortisol levels over time. In the present cohort of women, we also measured energy availability (energy intake minus exercise energy expenditure) (17), and while energy availability was low (range  $-10 \pm 11$  to  $23 \pm 15$  kcal kg FFM day $^{-1}$ , depending on the time of training and measure used), sex differences in energy deficit cannot plausibly account for the difference in cortisol levels we observe herein.

**Table 3.** Comparison of 1-h fold-wise LH and FSH responses to gonadorelin between progestogen-containing contraceptive users and nonusers

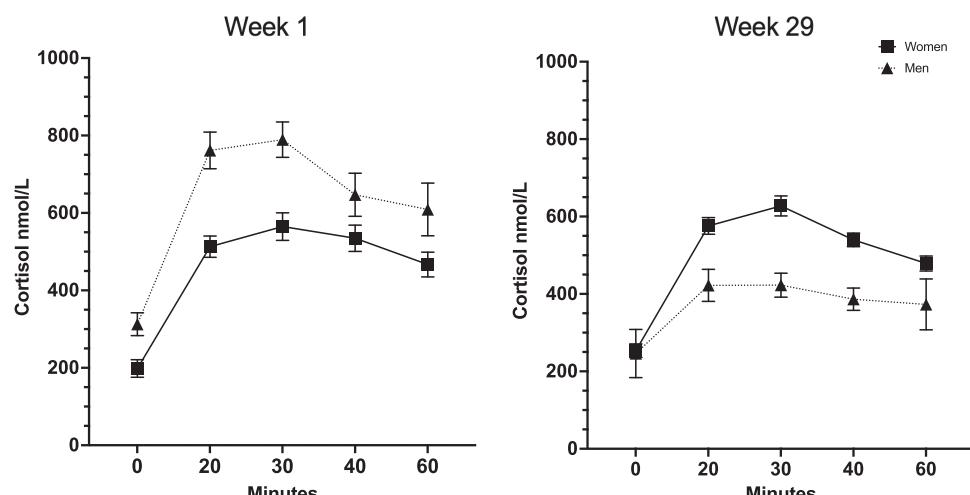
Oral Injected or Implanted Progesterone Contraceptive (n = 16)	No Contraceptive (n = 14)	Intrauterine Contraceptive System (n = 4)	F(2, 34)*	P*	F(2, 32)**	P**
Fold-wise change in FSH, median (IQR)						
Peak						
Week 1	1.44 (1.33, 1.58)	1.47 (1.28, 2.54)	2.15 (1.36, 2.53)	1.55	0.23	0.93
Week 29	1.39 (1.29, 1.63)	1.36 (1.2, 1.73)	1.82 (1.17, 2.43)	1.46	0.25	
AUC						
Week 1	69.9 (65.4, 78.2)	70.4 (63.9, 122)	104.6 (64.0, 115.4)	1.36	0.27	0.85
Week 29	66.0 (62.8, 76.7)	67.3 (58.8, 82.4)	86.2 (57.0, 115.5)	1.66	0.21	
Fold-wise change in LH, median (IQR)						
Peak						
Week 1	4.99 (3.00, 7.90)	4.67 (3.17, 7.08)	11.96 (3.04, 18.84)	0.52	0.60	0.29
Week 29	3.51 (2.49, 6.18)	3.44 (2.49, 5.85)	5.08 (2.02, 8.06)	0.019	0.98	
AUC						
Week 1	227 (138, 350)	219 (147, 322)	545 (141, 793)	0.54	0.59	0.27
Week 29	163 (112, 265)	161 (114, 264)	236.23 (91, 367)	0.024	0.98	

Data were transformed by natural logarithm before analysis. AUC, area under the curve; IQR, interquartile range. \*One-way ANOVA of difference between groups at each timepoint. \*\*Mixed repeat-measures ANOVA between groups over time (main effect of contraceptive group  $\times$  time).

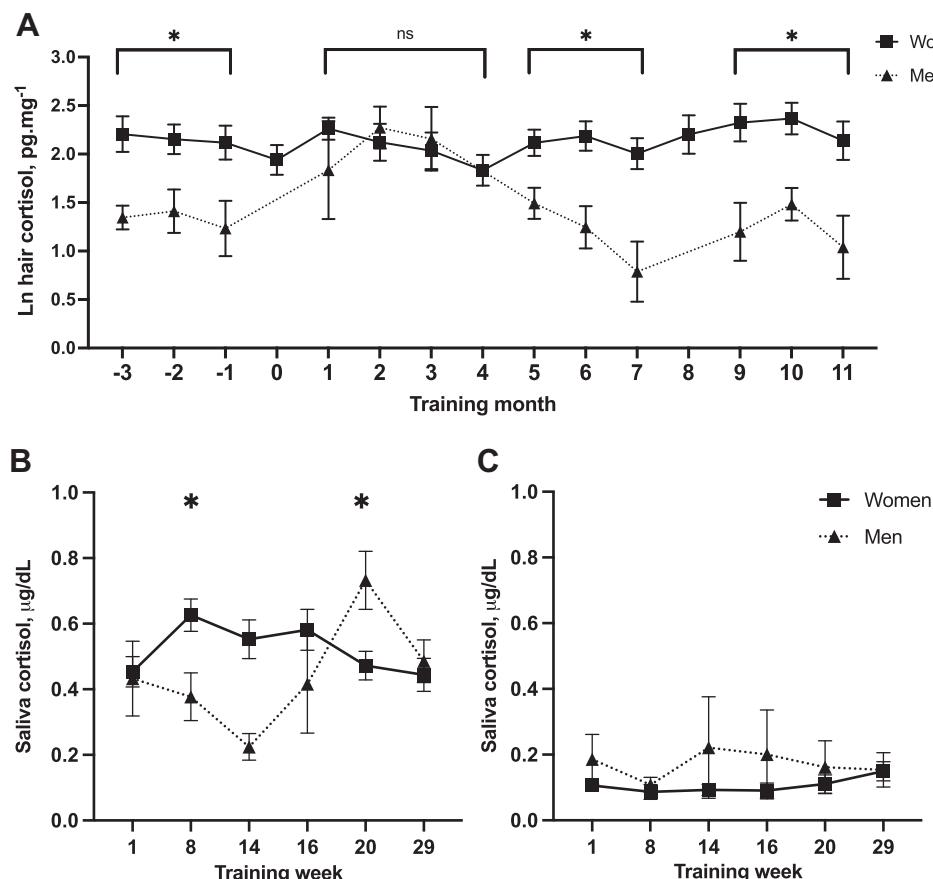
These data suggest that there are complex influences on the HPG axis function and point to factors other than the HPA axis function. There was a lack of clear sex-biased effect on LH and FSH during intensive military training. These findings support previous work in humans, demonstrating intact endocrine functionality in females exposed to significant, sustained environmental and psychological stressors (36). Although no statistical interaction was observed between the sexes over time, the sex difference in gonadotroph function was numerically smaller at week 29 than week 1. We previously reported suppressed gonadotroph responsiveness during training, among the larger cohort of women (also including COCP users) (14). These data indicate that physiological stress, likely associated with CRH and/or AVP stimulation, was greater among women, manifesting as relatively enhanced adrenal responsiveness after 29 wk. A study with greater statistical power would be required to determine if a significant sex difference in gonadotroph function is seen over time.

Pituitary sensitivity to GnRH is influenced by upstream peptides including kisspeptin, and varies according to sex and across the phases of the menstrual cycle (37). Previously published urinary hormone data suggest that noncontraceptive users remained in the follicular phase throughout the study (14). There was also no difference in gonadotroph response between women using progestogen-only contraception and noncontraceptive users. The greater gonadotroph response among women than men may reflect physiological differences in gonadotroph sensitivity. At the time of writing, we are not aware of any published data that give sex-adjusted normative values for fold-wise response to a low-dose GnRH test.

A possible explanation for the heightened HPA axis response among women lies in the neuroendocrine differences between the sexes. Estrogens have been implicated in enhancing HPA axis sensitivity (5, 6). Estrogen receptors are abundantly expressed in the paraventricular nucleus which is central to HPA axis regulation and is closely associated with the suprachiasmatic nucleus—the circadian clock (38).



**Figure 2.** Male and female cortisol response to 1  $\mu$ L adrenocorticotrophin (ACTH)-1-24 over 1 h during weeks 1 and 29 of military training. Significant sex  $\times$  time interaction, for the area under the curve  $F_{1, 44} = 18.0$ ,  $P < 0.001$ . Data are means  $\pm$  SE.



**Figure 3.** Hair and salivary cortisol. Natural logarithm of hair cortisol concentrations (A), morning salivary cortisol concentrations (B), and evening salivary cortisol concentrations (C). Significant sex  $\times$  time interactions were observed for hair cortisol ( $P = 0.024$ ) and morning salivary cortisol ( $P = 0.005$ ), but not evening salivary cortisol ( $P = 0.3$ ). Week 14 saliva cortisol was excluded from statistical analysis due to missing data. Data are means  $\pm$  SE. Ln, natural logarithm. \*Significant sex difference observed, independent samples  $t$  test,  $P < 0.05$ ; ns, not significant.

Sleep deprivation is commonly reported during military training (39), and was marked during the study (40). Altered sleep has a potentiating effect on HPA axis reactivity [reviewed by Dalsen and Markus (41)] and sex differences have been observed in HPA axis vulnerability to sleep disturbance. An enhanced HPA axis response to CRH was seen among women with poor sleep compared with a smaller cohort of men with poor sleep (42), whereas 8-yr old (43) and adolescent girls (44) demonstrated an enhanced response to a standardized stress test following sleep deprivation compared with adolescent boys. The underlying reason for a greater response in women is not clear but could relate to increased estrogen receptor-mediated sensitivity amplifying crosstalk between the paraventricular and suprachiasmatic nuclei (38), or given findings from a range of ages and hormonal milieus, it seems plausible that some sex-associated effect of sleep disturbance on the HPA axis may exist independently of sex steroids.

Arginine vasopressin is an important hypothalamic activator of the HPA axis and demonstrates a striking sex dimorphism in rodents, with females showing greater expression of AVP following stress (38). Coactivation of AVP and the HPA axis was observed in women but not in men during an insulin stress test (45). Impaired hydration and heat stress are encountered frequently during military training and may have potentiated the stress response in women, although hydration status was not directly measured. Future studies should address the importance of sex dimorphisms in the co-activation of AVP and HPA axis in response to environmental stress.

Military training is complex and entails multifaceted psychological and social challenges as well as physical stressors, for example, immersion in a nuanced sociocultural environment, reduced volition, simulated threats, and continuous assessment. We observed slight overall increases in scores of depression and anxiety, with no change in resilience. There was no effect of sex observed over time; however, we did observe more adverse events in women than men. A larger study measuring ethnographic stress, coping, and psychological well-being may elucidate sex differences in responses to multistressor training.

Discordant morning salivary and hair cortisol during training highlights the importance of sampling protocols in studies examining cortisol abundance in humans. Although hair cortisol was similar between men and women during the first 3 mo, it was higher among women thereafter; for morning salivary cortisol, the opposite was seen. Greater anticipatory stress drives morning cortisol (31), which may have been more frequent in women during the early stages of training, and in men during week 20 (the week of competitive regimental selection). However, overall cortisol exposure was similar in the early weeks of training, driven perhaps by the multiple, frequent stressors experienced at this stage—the “shock of capture.”

Strengths of this study include the combination of dynamic and basal hormone markers over a long duration. We believe this is the first time that a low-dose pituitary function test has been applied or combined with basal cortisol markers during multistressor training. The multistressor exposure is relevant

to the increasing diversity of gender in arduous employment roles. Participants were well matched in terms of age and demographics and these findings are supported by detailed measurements of energy balance reported elsewhere (16).

Limitations of this work include the sample size of men, caused by ethical constraints on recruitment, and relatively low numbers of women commencing the Commissioning Course. The training requirements meant that we were unable to synchronize tests to the menstrual cycle (there was a 1-wk window in which to arrange testing). There was significant variability in cycle timing during both tests, contributing to high variability in LH; however, by using fold-wise responses, we were able to detect trends in pituitary responsiveness and make meaningful comparisons across groups. Moreover, due to course requirements, it was not possible to perform dynamic function testing in the early morning.

In conclusion, military training enhanced the HPA axis response and increased average cortisol exposure among women, but not men. This HPA axis response was associated with limited evidence suggesting a greater suppression of HPG axis response in women than men. These findings suggest that there are complex influences on HPG axis function, which extend beyond HPA axis activation. It is important to consider sex-biased differences in strategies to address stressors, such as sleep deprivation, hydration status, or psychological coping strategies.

## DATA AVAILABILITY

Data will be made available upon reasonable request.

## GRANTS

This work was supported by the Ministry of Defence, UK (ASC Task 0108).

## DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

## AUTHOR CONTRIBUTIONS

R.M.G., S.L.W., R.A.A., J.P.G., R.M.R., and D.R.W. conceived and designed research; R.M.G., T.J.O., R.L.K., and S.L.W. performed experiments; R.M.G., T.J.O., S.L.W., and D.R.W. analyzed data; R.M.G., T.J.O., C.L.D., R.A.A., J.P.G., R.M.R., and D.R.W. interpreted results of experiments; R.M.G. prepared figures; R.M.G. and T.J.O. drafted manuscript; R.M.G., T.J.O., S.L.W., C.L.D., R.A.A., J.P.G., and D.R.W. edited and revised manuscript; R.M.G., T.J.O., R.L.K., S.L.W., C.L.D., R.A.A., J.P.G., R.M.R., and D.R.W. approved final version of manuscript.

## REFERENCES

1. **Warren MP.** The effects of exercise on pubertal progression and reproductive function in girls. *J Clin Endocrinol Metab* 51: 1150–1157, 1980. doi:10.1210/jcem-51-5-1150.
2. **Schneider MB, Fisher M, Friedman SB, Bijur PE, Toffler AP.** Menstrual and premenstrual issues in female military cadets: a unique population with significant concerns. *J Pediatr Adolesc Gynecol* 12: 195–201, 1999. doi:10.1016/s1083-3188(99)00025-x.
3. **Lauder TD, Williams MV, Campbell CS, Davis G, Sherman R, Pulos E.** The female athlete triad: prevalence in military women. *Mil Med* 164: 630–635, 1999.
4. **Loucks AB.** Exercise training in the normal female: effects of low energy availability on reproductive function. In: *Endocrinology of Physical Activity and Sport*, edited by Constantini NH, Hackney AC. New York: Humana Press, Springer, 2013. doi:10.1007/978-1-62703-314-5.
5. **Figueiredo HF, Ulrich-Lai YM, Choi DC, Herman JP.** Estrogen potentiates adrenocortical responses to stress in female rats. *Am J Physiol Endocrinol Physiol* 292: E1173–E1182, 2007. doi:10.1152/ajpendo.00102.2006.
6. **Kirschbaum C, Kudielka BM, Gaab J, Schommer NC, Hellhammer DH.** Impact of gender, menstrual cycle phase, and oral contraceptives on the activity of the hypothalamus-pituitary-adrenal axis. *Psychosom Med* 61: 154–162, 1999. doi:10.1097/00006842-199903000-00006.
7. **Viau V, Meaney MJ.** The inhibitory effect of testosterone on hypothalamic-pituitary-adrenal responses to stress is mediated by the medial preoptic area. *J Neurosci* 16: 1866–1876, 1996. doi:10.1523/JNEUROSCI.16-05-01866.1996.
8. **Rubinow DR, Roca CA, Schmidt PJ, Danaceau MA, Putnam K, Cizza G, Chrousos G, Nieman L.** Testosterone suppression of CRH-stimulated cortisol in men. *Neuropsychopharmacology* 30: 1906–1912, 2005. doi:10.1038/sj.npp.1300742.
9. **Reynolds RM, Hii HL, Pennell CE, McKeague IW, de Kloet ER, Lye S, Stanley FJ, Mattes E, Foster JK.** Analysis of baseline hypothalamic-pituitary-adrenal activity in late adolescence reveals gender specific sensitivity of the stress axis. *Psychoneuroendocrinology* 38: 1271–1280, 2013. doi:10.1016/j.psyneuen.2012.11.010.
10. **Roca CA, Schmidt PJ, Deuster PA, Danaceau MA, Altemus M, Putnam K, Chrousos GP, Nieman LK, Rubinow DR.** Sex-related differences in stimulated hypothalamic-pituitary-adrenal axis during induced gonadal suppression. *J Clin Endocrinol Metab* 90: 4224–4231, 2005 [Erratum in *J Clin Endocrinol Metab* 90: 5522, 2005]. doi:10.1210/jc.2004-2525.
11. **Solomon MB, Furay AR, Jones K, Packard AE, Packard BA, Wulsin AC, Herman JP.** Deletion of forebrain glucocorticoid receptors impairs neuroendocrine stress responses and induces depression-like behavior in males but not females. *Neurosci* 203: 135–143, 2012. doi:10.1016/j.neuroscience.2011.12.014.
12. **Kajantie E, Phillips DL.** The effects of sex and hormonal status on the physiological response to acute psychosocial stress. *Psychoneuroendocrinology* 31: 151–178, 2006. doi:10.1016/j.psyneuen.2005.07.002.
13. **Areta JL, Taylor HL, Koehler K.** Low energy availability: history, definition and evidence of its endocrine, metabolic and physiological effects in prospective studies in females and males. *Eur J Appl Physiol* 121: 1–21, 2021. doi:10.1007/s00421-020-04516-0.
14. **Gifford RM, O'Leary TJ, Wardle SL, Double RL, Homer NZM, Howie AF, Greeves JP, Anderson RA, Woods DR, Reynolds RM.** Reproductive and metabolic adaptation to multistressor training in women. *Am J Physiol Endocrinol Physiol* 321: E281–E291, 2021. doi:10.1152/ajpendo.00019.2021.
15. **Gifford RM, O'Leary TJ, Double RL, Wardle SL, Wilson K, Boyle LD, Homer NZM, Kirschbaum C, Greeves JP, Woods DR, Reynolds RM.** Positive adaptation of HPA axis function in women during 44 weeks of infantry-based military training. *Psychoneuroendocrinology* 110: 104432, 2019. doi:10.1016/j.psyneuen.2019.104432.
16. **O'Leary TJ, Gifford RM, Knight RL, Wright J, Handford S, Venables MC, Reynolds RM, Woods D, Wardle SL, Greeves JP.** Sex differences in energy balance, body composition, and metabolic and endocrine markers during prolonged arduous military training. *J Appl Physiol* (1985) 136: 938–948, 2024. doi:10.1152/japplphysiol.00864.2023.
17. **Gifford RM, Greeves JP, Wardle SL, O'Leary TJ, Double RL, Venables M, Boos C, Langford J, Woods DR, Reynolds RM.** Measuring the exercise component of energy availability during arduous training in women. *Med Sci Sports Exerc* 53: 860–868, 2021. doi:10.1249/MSS.0000000000002527.
18. **O'Leary TJ, Wardle SL, Gifford RM, Double RL, Reynolds RM, Woods DR, Greeves JP.** Tibial macrostructure and microarchitecture adaptations in women during 44 weeks of arduous military training. *J Bone Miner Res* 36: 1300–1315, 2020. doi:10.1002/jbm2.4290.
19. **Ministry of Defence.** Joint Service Manual of Medical Fitness (Online). Ministry of Defence. [https://data.parliament.uk/DepositedPapers/Files/DEP2019-0604/Joint\\_Service\\_Manual\\_of\\_Medical\\_Fitness.pdf](https://data.parliament.uk/DepositedPapers/Files/DEP2019-0604/Joint_Service_Manual_of_Medical_Fitness.pdf) [2024 Dec 2].

20. **Meulenberg PM, Ross HA, Swinkels LM, Benraad TJ.** The effect of oral contraceptives on plasma-free and salivary cortisol and cortisone. *Clin Chim Acta* 165: 379–385, 1987. doi:10.1016/0009-8981(87)90183-5.
21. **Weiss DS.** The impact of event scale-revised. In: *Assessing Psychological Trauma and PTSD: A Practitioner's Handbook*, edited by Wilson JP, Keane TM. New York: Guilford Press, 1997, p. 168–189.
22. **Kroenke K, Spitzer RL, Williams JB.** The PHQ-9: validity of a brief depression severity measure. *J Gen Intern Med* 16: 606–613, 2001. doi:10.1046/j.1525-1497.2001.016009606.x.
23. **Beck AT, Epstein N, Brown G, Steer RA.** An inventory for measuring clinical anxiety: psychometric properties. *J Consult Clin Psychol* 56: 893–897, 1988. doi:10.1037/0022-006x.56.6.893.
24. **Campbell-Sills L, Stein MB.** Psychometric analysis and refinement of the Connor–Davidson resilience scale (CD-RISC): validation of a 10-item measure of resilience. *J Trauma Stress* 20: 1019–1028, 2007. doi:10.1002/jts.20271.
25. **Morosini PP, Sarzani R, Arnaldi G, Taccaletti A.** Hypothalamic amenorrhea. Different patterns in the pulsatile secretion of LH during 24 hours and different responses to the stimulation test with GnRH. *Minerva Endocrinol* 14: 153–158, 1989.
26. **Lewis JG, Elder PA.** Corticosteroid-binding globulin reactive centre loop antibodies recognise only the intact natured protein: elastase cleaved and uncleaved CBG may coexist in circulation. *J Steroid Biochem Mol Biol* 127: 289–294, 2011. doi:10.1016/j.jsbmb.2011.08.006.
27. **Pragst F, Balikova MA.** State of the art in hair analysis for detection of drug and alcohol abuse. *Clin Chim Acta* 370: 17–49, 2006. doi:10.1016/j.cca.2006.02.019.
28. **Iob E, Kirschbaum C, Steptoe A.** Positive and negative social support and HPA-axis hyperactivity: evidence from glucocorticoids in human hair. *Psychoneuroendocrinology* 96: 100–108, 2018. doi:10.1016/j.psyneuen.2018.06.008.
29. **Kudielka BM, Kirschbaum C.** Sex differences in HPA axis responses to stress: a review. *Biol Psychol* 69: 113–132, 2005. doi:10.1016/j.biopsych.2004.11.009.
30. **Stalder T, Steudt-Schmidgen S, Alexander N, Klucken T, Vater A, Wichmann S, Kirschbaum C, Miller R.** Stress-related and basic determinants of hair cortisol in humans: a meta-analysis. *Psychoneuroendocrinology* 77: 261–274, 2017. doi:10.1016/j.psyneuen.2016.12.017.
31. **Adam EK, Quinn ME, Tavernier R, McQuillan MT, Dahlke KA, Gilbert KE.** Diurnal cortisol slopes and mental and physical health outcomes: a systematic review and meta-analysis. *Psychoneuroendocrinology* 83: 25–41, 2017. doi:10.1016/j.psyneuen.2017.05.018.
32. **Clow A, Edwards S, Owen G, Evans G, Evans P, Hucklebridge F, Casey A.** Post-awakening cortisol secretion during basic military training. *Int J Psychophysiol* 60: 88–94, 2006. doi:10.1016/j.ijpsycho.2005.05.007.
33. **Henning PC, Scofield DE, Spiering BA, Staab JS, Matheny RW Jr, Smith MA, Bhasin S, Nindl BC.** Recovery of endocrine and inflammatory mediators following an extended energy deficit. *J Clin Endocrinol Metab* 99: 956–964, 2014. doi:10.1210/jc.2013-3046.
34. **Friedl KE, Moore RJ, Hoyt RW, Marchitelli LJ, Martinez-Lopez LE, Askew EW.** Endocrine markers of semistarvation in healthy lean men in a multistressor environment. *J Appl Physiol* (1985) 88: 1820–1830, 2000. doi:10.1152/jappl.2000.88.5.1820.
35. **Ponce T, Mainenti MRM, Barros T, Cahuê FLC, Fernanda Martins dos Santos C, Piazera BKL, Salerno VP, Vaisman M.** Biochemical and hormones markers in firefighters: effects of “Search, rescue, and survival training” and its recovery. *J Strength Cond Res* 38: e189–e201, 2024. doi:10.1519/JSC.00000000000004695.
36. **Gifford RM, O'Leary T, Cobb R, Blackadder-Weinstein J, Double R, Wardle SL, Anderson RA, Thake CD, Hattersley J, Imray CHE, Wilson A, Greeves JP, Reynolds RM, Woods DR.** Female reproductive, adrenal, and metabolic changes during an Antarctic traverse. *Med Sci Sports Exerc* 51: 556–567, 2019. doi:10.1249/MSS.0000000000001803.
37. **Skorupskaitė K, George JT, Anderson RA.** The kisspeptin-GnRH pathway in human reproductive health and disease. *Hum Reprod Update* 20: 485–500, 2014. doi:10.1093/humupd/dmu009.
38. **Heck AL, Handa RJ.** Sex differences in the hypothalamic-pituitary-adrenal axis' response to stress: an important role for gonadal hormones. *Neuropharmacology* 44: 45–58, 2019. doi:10.1038/s41386-018-0167-9.
39. **Crowley SK, Wilkinson LL, Burroughs EL, Muraca ST, Wigfall LT, Louis-Nance T, Williams EM, Glover SH, Youngstedt SD.** Sleep during basic combat training: a qualitative study. *Mil Med* 177: 823–828, 2012. doi:10.7205/milmed-d-12-00022.
40. **Koivula F, Wardle SL, Double R, Gifford RM, Woods DR, Reynolds RM, Handford S, Wright J, O'Leary TJ, Greeves JP.** Sleep patterns during arduous military training in men and women. *Med Sci Sports Exerc* 51: 277–278, 2019. doi:10.1249/01.mss.0000561335.43488.5b.
41. **van Dalzen JH, Markus CR.** The influence of sleep on human hypothalamic–pituitary–adrenal (HPA) axis reactivity: a systematic review. *Sleep Med Rev* 39: 187–194, 2018. doi:10.1016/j.smrv.2017.10.002.
42. **Hori H, Teraishi T, Sasayama D, Ozeki Y, Matsuo J, Kawamoto Y, Kinoshita Y, Hattori K, Higuchi T, Kunugi H.** Poor sleep is associated with exaggerated cortisol response to the combined dexamethasone/CRH test in a non-clinical population. *J Psychiatr Res* 45: 1257–1263, 2011. doi:10.1016/j.jpsychires.2011.04.001.
43. **Pesonen AK, Kajantie E, Heinonen K, Pyhälä R, Lahti J, Jones A, Matthews KA, Eriksson JG, Strandberg T, Räikkönen K.** Sex-specific associations between sleep problems and hypothalamic–pituitary–adrenocortical axis activity in children. *Psychoneuroendocrinology* 37: 238–248, 2012. doi:10.1016/j.psyneuen.2011.06.008.
44. **Mrug S, Tyson A, Turan B, Granger DA.** Sleep problems predict cortisol reactivity to stress in urban adolescents. *Physiol Behav* 155: 95–101, 2016. doi:10.1016/j.physbeh.2015.12.003.
45. **Kacheva S, Kolk K, Morgensthaler NG, Brabant G, Karges W.** Gender-specific co-activation of arginine vasopressin and the hypothalamic–pituitary–adrenal axis during stress. *Clin Endocrinol (Oxf)* 82: 570–576, 2015. doi:10.1111/cen.12608.